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# RESEARCH ARTICLE



# Spatial extent predicts Andean epiphyte biodiversity responses to habitat loss and fragmentation across human-modified landscapes

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# Abstract

**Aim:** Human-driven landscape processes such as habitat loss and fragmentation act on biodiversity, but their effects are mediated by the spatial scale at which they are observed. We aim to analyse the scale-of-effects (direction and spatial extent) of landscape-scale processes that best explain species richness and abundance across epiphyte communities.

Location: Neotropics, Northern Andes, Colombia, Eastern cordillera.

Taxon: Vascular epiphytes, Orchidaceae.

**Methods:** We used field data to unravel the scale-of-effect of three landscape processes—habitat loss (forest cover), fragmentation (number of patches) and edge effects (edge density)—on epiphyte biodiversity. Vascular epiphytes were sampled in the understorey across 141 plots within 23 Andean forests in the eastern Colombian cordillera We focused on the community-level responses (species richness and total abundance) of the hyperdiverse vascular epiphyte communities using generalized linear mixed models to quantify the direction and the spatial extent of the scale-of-effects.

**Results:** Habitat loss and edge effects act at fine spatial extents (scale-ofeffects = 200 m), predicting low species richness and abundance across groups. Likewise, fragmentation negatively impacts communities, but operates at larger spatial extents (scale-of-effects = 2000-2400m radius). The detection of these effects is contingent upon the spatial extent and specific landscape processes involved. Models of habitat loss within a spatial extent of 800-1500m (large confidence intervals), fragmentation below 300m, and edge effects above 800m show weak statistical support (marginal  $r^2$  = 0.02-0.1). Thus, the impacts of these landscape processes may be overlooked if studied at inadequate spatial extents.

**Main Conclusions:** We showed that habitat loss, fragmentation and edge effects all play a negative role on understorey epiphytic communities, but their detectability is scale dependant. The scale-of-effects can assist landscape designs that are beneficial for epiphytic communities, by preserving forest cover, and reducing fragmentation and exposure to edge effects at small scales (200–300 m). Conversely, landscape scale actions directed at reducing habitat loss and fragmentation function at larger

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spatial extents (>2000 m). Selecting a priori or inadequate spatial extents of analysis can obscure the detectability of landscape processes.

KEYWORDS

community ecology, fragmentation, habitat loss, landscape ecology, orchids, tropical Andes, understorey

# 1 | INTRODUCTION

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Ecological patterns and processes are inherently scale dependent (Levin, 1992; Wiens, 1989). Failing to quantify the spatial scale at which biodiversity patterns and processes are shaped can lead to incorrect quantification of their impacts and potentially ineffective conservation strategies in a human-dominated world (Arroyo-Rodríguez et al., 2020). Detectability of this spatial scale is important because populations, species and communities use, interpret and respond to their habitats and surroundings in different ways. For instance, the average spatial extents at which species have the strongest relationship with landscape drivers is 1000m, but its extent ranged from 40m to 5500m, making it highly taxa and context dependent (Saura, 2020). Although the importance of spatial scale and the role of the landscape on shaping biodiversity is well-acknowledged (Arroyo-Rodríguez et al., 2020; Jackson & Fahrig, 2015; Turner et al., 1989), our understanding of their effects and consequences on plant biodiversity is still poor across mountainous landscapes.

Mountain ecosystems are one of the richest biodiversity hotspots worldwide, but they have been severely transformed by human activities (Rahbek et al., 2019; Vancutsem et al., 2021). For instance, the Northern Andes ranks first in diversity of birds, butterflies and plants (Myers et al., 2000; Pérez-Escobar et al., 2022), but less than 40% of its natural landscape remains (Rodríguez Eraso et al., 2013). In general, once the natural landscape is transformed by human activities, three main landscape processes appear: (1) habitat loss or the decline of suitable habitat in terms of area (Ewers & Didham, 2008; Fahrig, 2003); (2) fragmentation as the physical partitioning of the landscape into smaller pieces (Fahrig, 2003, 2017); and (3) edge effects from the creation of new edges between two or more habitats (Malcolm, 1994; Ries et al., 2017). The effects of these landscape processes are highly context-dependent because evidence has shown negative, positive, neutral and interactive effects.

Habitat loss is consistently the strongest landscape process on species richness, with negative effects across taxa, scales and response metrics (Arroyo-Rodríguez et al., 2020; Fahrig, 2017; Saura, 2020), to the point of threatening species' populations with extinction (Brooks et al., 2002). In contrast, fragmentation studies have shown positive, neutral and negative effects on biodiversity (Arroyo-Rodríguez & Dias, 2010; Fletcher et al., 2018; Gestich et al., 2019). A multiscale study across the Brazilian Atlantic Forest shows that fragmentation negatively affects species richness (flora and fauna), but the strength of impact is modulated by forest cover in the landscape (Püttker et al., 2020). Edge effects also have several negative impacts on plant diversity including the reduction of forestdependent species (Parra-Sánchez et al., 2016; Püttker et al., 2020). However, the detectability of the direction and extension of these landscape processes relies on the spatial extent ('scale-of-effects').

The scale-of-effects is a concept that seeks to unravel the most informative spatial extent at which the effect of a particular landscape process is detectable and relevant for biodiversity (Fahrig, 2013; Jackson & Fahrig, 2015). Investigating the scale-ofeffects requires understanding of the spatial grain and spatial extent. The spatial grain determines the smallest unit of analysis, which in scale-of-effects studies is the sampling plot where the targeted taxa and metrics are collected (Turner et al., 1989; McGarigal et al., 2009). The spatial extent refers to the overall geographic scope of the study area, encompassing the boundaries within which data are analysed (McGarigal et al., 2009; Turner et al., 2001). In scale-ofeffects studies, the spatial extent progressively varies in area to find the most relevant scale, using concentric extents centred at the grain (i.e. sample plot) to quantify the effects of landscape process on biodiversity (Jackson & Fahrig, 2015). By further expanding the concept, scale-of-effects can also inform the magnitude and direction of the correlation, which provides insights into the impact (positive, negative, neutral or not detected) that the landscape process has on biodiversity.

The quantification of the scale-of-effects moves beyond a purely ecological-methodological concern, towards a pragmatic tool that could improve the successful implementation of conservation strategies such as land-sharing/land-sparing actions (Fischer et al., 2014) and restoration initiatives (Crouzeilles & Curran, 2016), by detecting the spatial extent and direction of the effects of human activity on biodiversity. For instance, a global meta-analysis found that successful biodiversity outcomes following forest restoration are best predicted by the amount of forest cover present within a 5–10-km radius (Crouzeilles & Curran, 2016). Therefore, understanding of this scale-dependent response can help target effective restoration (Brancalion et al., 2019).

Scale-of-effects studies on plant species richness point to negative effects of habitat loss within 1500-2000m in the Brazilian Atlantic Forest (Püttker et al., 2020), but no effect at fine spatial extents within 50-650m in Costa Rica (Holl et al., 2017). While fragmentation shows a reduction of species richness at spatial extents of 1800-2000m, its effects are associated with edge effects (Püttker et al., 2020). In Mexico, edge density negatively affects seedling richness within 50-1000m, but fragmentation exerts no effects (San-José et al., 2020). These studies demonstrate the need to empirically quantify the scale-of-effects (Arroyo-Rodríguez et al., 2020; Jackson & Fahrig, 2015). As we quantify the scale-of-effects on different taxa and ecosystems, new patterns might emerge, especially because data collection is often systematically, geographically or taxonomically biased, which could bias predictions, especially for rare taxa in human-modified landscapes (Fardila et al., 2017; Trimble & van Aarde, 2012).

We focus our study on understorey vascular epiphytes, that is, non-soil rooted vascular plants that live on a host without parasitizing them (Zotz, 2013). This group represents 10% of the global vascular flora (Taylor et al., 2021; Zotz, 2013), yet epiphytes are usually underrepresented in multiscale studies. Vascular epiphytes have several attributes that in combination make them an interesting group for quantifying the impacts of habitat loss and fragmentation and the spatial extent at which they best predict biodiversity. These attributes include: high taxonomical diversity; species with high specialization; high mortality rate (four times higher than soilrooted plants; Zuleta et al., 2016); potential dispersal limitation (Janzen et al., 2020); large numbers of threatened species (Calderón-Saenz, 2007); high interspecies-ecological equivalence (Catchpole & Kirkpatrick, 2010); and low-to-negligible interspecific competition (Zotz, 2016). At local scale, large trees support higher epiphyte diversity than smaller trees, and trees with complex crowns support more epiphytes than trees with simple crown structure (Woods et al., 2015; Zuleta et al., 2016). In contrast, remnant trees often host vascular epiphytes in their crowns (Einzmann & Zotz, 2017a, 2017b). However, the scale-of-effects of habitat loss, fragmentation, and edge effects have not been quantified on vascular epiphytes leaving a big gap in our understanding of the appropriate scale for landscape management of this speciose group (Flores-Palacios & García-Franco, 2007; Hernandez-Perez & Solano, 2015; Köster et al., 2009; Larrea et al., 2010; Parra-Sanchez & Banks-Leite, 2020; Ruete et al., 2018; Zuleta et al., 2016).

We aim to analyse the scale-of-effects (direction and spatial extent) of landscape-scale processes that best explain understorey species richness and abundance across epiphyte communities by specifically asking: (1) what is the direction and the spatial extent at which habitat loss best explain epiphyte communities; and (2) what is the direction and the spatial extent at which fragmentation and edge effects best explain epiphyte communities? Our first expectation focuses on the positive correlation of epiphyte diversity with forest cover, regardless of the spatial extent, because forest cover promotes resource availability and landscape connectivity (Arroyo-Rodríguez et al., 2020). Our second expectation lies in the consistent negative effects of habitat fragmentation and edge effects on epiphyte diversity irrespective of the spatial extent, due to the high habitat specialization, potential dispersal limitation in epiphyte communities and sensitivity to environmental changes caused by habitat loss and fragmentation (Janzen et al., 2020; Werner, 2011; Zotz, 2005; Zuleta et al., 2016). Our third expectation is that effects might be different for orchids acting at low spatial extents versus other epiphytes, due to the sensitivity of orchids to local disturbance

reflected in altered species densities, high compositional turnover in the Andes, and high proportion of orchid species threatened with extinction relative to any other epiphytic family (Carmona-Higuita et al., 2023; Parra-Sanchez et al., 2023; Parra-Sánchez et al., 2016).

# 2 | MATERIALS AND METHODS

The study was located in the departments of Cundinamarca, Boyacá, Meta and Santander, in the Colombian eastern cordillera (Figures 1 and 2). We sampled 23 forests across a wide elevational range (1163-3415m) belonging to the Andean mountains (Pérez-Escobar et al., 2022), with mean cloud cover of 82% (25-90%; Wilson & Jetz, 2016), temperature ranging 7-28°C and annual precipitation of 891-3711 mm per year. Andean forests were randomly selected based on forest availability in the area, logistic constraints (accessibility, permit from landowners, topography), and safety and security considerations. We randomized forest selection because forests at both extremes of the gradient are scarce either due to deforestation (<1400m; Aide et al., 2019), or natural absence above the treeline (>3000m; Bader et al., 2007; Lüttge, 1997). Forest belonged to two types: Andino (21 forests) and Altoandino (2) (Etter et al., 2021; Table S1). Forests had discrete boundaries surrounded by a predominantly pastureland matrix. Although all sampled sites belonged to the network of protected areas in Colombia (Sistema nacional de areas protegidas, SINAP), we considered them as having undergone low-intensity human-induced modification, given the easy accessibility by roads or footpaths, selective logging for local uses and knowledge from local field assistants.

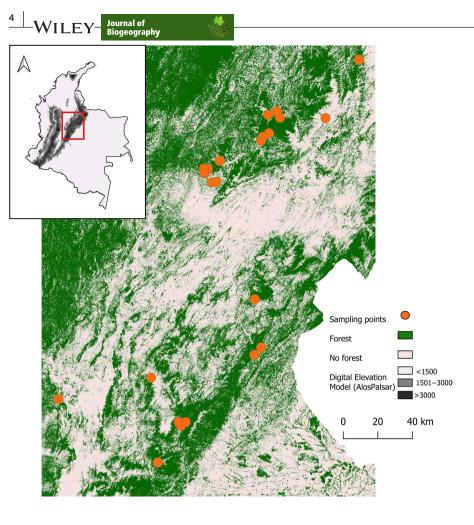
# 2.1 | Sampling

Surveys were conducted from January to November 2019 following Parra-Sanchez et al. (2023). We sampled between 2 and 18 sampling plots in the understorey of  $10 \times 30$  m in each forest (n=141). Larger forest areas had more plots to ensure wider coverage. Sampling plots were randomly placed in sections of the forest at least 30 m from edges to reduce the effects of cattle movement and grazing (*pers. obser.* and Parra-Sánchez et al., 2016). Sampling plots were located between 150 and 600 m apart inside each forest. This design allows us to capture a large variability of species richness and high turnover across communities (Parra-Sanchez et al., 2023).

We recorded all individuals on standing tree trunks, fallen tree trunks and branches, vines, lianas, leaves on standing trees or herbaceous plants, palm trees, tree ferns and cycads without soil contact in the understorey. The operational concept of understorey we used was the section of the forest between 50 and 200cm aboveground, where it is expected to have lower light levels, reduced wind exposure and greater humidity compared to the upper canopy (Coenders-Gerrits et al., 2020; Ewers & Banks-Leite, 2013). The number of species and adult individuals (i.e. ramets) were recorded, while younger stages were not included due to a large uncertainty

**FIGURE 1** Sampling points of epiphytic communities across the eastern cordillera in Colombia (WGS84). Centroid of the focal Andean Forest fragments (in orange, n=23), their respective surrounding forest (black) and non-forest (grey), and AlosPalsar Digital Elevation model

(Tadono et al., 2014).



in species identification. Ramets usually comprised various individual stems, but for our purpose, a ramet only counts as one single individual spatially separated from another set of stems of the same or another species, or an empty space (Sanford, 1968). Adult plants were defined as all large individuals (within the observed variation in the local population) with evidence of developed floral or reproductive structure, large ramets, many stems per stand and a prominent root system.

Identification to species or morphospecies was conducted following specialized literature and expertise of local experts at the Herbarium VALLE. Orchid species with no flowers were taken to nurseries in the vicinity of the sampled plots, and later visited for identification. All dubious individuals and juveniles were ignored due to uncertainty in their taxonomical rank.

# 2.2 | Response variables

We used observed values of species richness and total abundance per plot for the two groups: orchids and non-orchid epiphytes (hereafter NOE; including Bromeliaceae, Gesneriaceae, Araceae and ferns). We focused on orchids as the most species-rich epiphytic family (Zotz 2021), dominant Andean plant family (Pérez-Escobar et al., 2022), its high sensitivity to forest disturbance and edge effects (Parra-Sanchez & Banks-Leite, 2020) and higher proportion of species threatened with extinction than other epiphytic families (Carmona-Higuita et al., 2023). We identified 82.2% of orchid plants to species level (273 species) and the remaining 17.8% plants to morphotypes (59 spp). For all NOE, we morphotyped each species at the plot level due to the impossibility of obtaining enough diagnostic characters and high levels of uncertainty in taxonomic identity across plots, resulting in a species richness count per plot. Therefore, analyses used the total number of species and morphospecies for orchids, and only morphospecies at plot level of NOE.

# 2.3 | Environmental predictors

We calculated landscape metrics at twelve arbitrary selected spatial extents centred on the sampling plot, thus: buffers of 100, 200, 300, 500, 800, 1000, 1200, 1500, 1800, 2000, 2200 and 2400m (Figure 2). The landscape metrics capture high variability across the range of spatial extents (Figure S1). We included a wide range of spatial extents because the dispersal of epiphyte species varies from a few meters to long distance. Epiphytic plants have large dispersal kernels that allow them to cross fragmented habitats (Einzmann & Zotz, 2017a, 2017b; Mondragón et al., 2015), although other wind-dispersed seeds from epiphytic bromeliads and orchid species land very closely to their source <10m (Mondragón et al., 2015; Mondragon & Calvo-Irabien, 2006). The grain of the

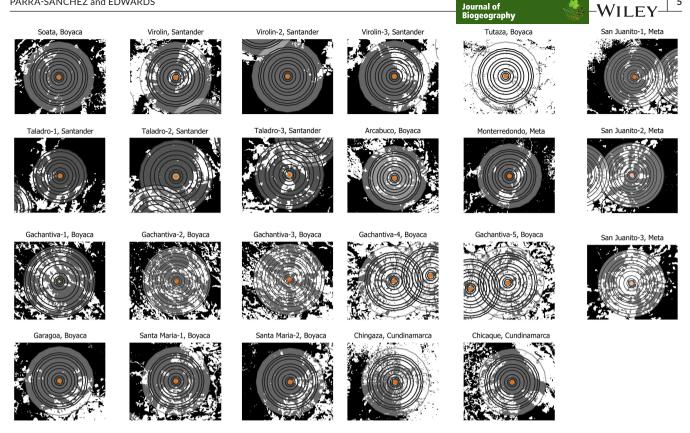


FIGURE 2 Location of studied forests (n = 23) along the eastern cordillera, Colombia. Maps depict forest cover (black), matrix (white), points represent (orange dots) the centroid of each forest, and buffers around each point represent the spatial extents analysed in our study (100-2400 m).

forest/non-forest map does not offer sufficient resolution to guantify scales below  $100 \text{ m} (30 \times 30 \text{ m map resolution})$ .

At each spatial extent, we quantified three landscape metrics linked to different ecological processes. First, habitat loss as the percent of forest cover (hereafter, forest cover) as a metric of habitat amount available in the landscape as a proxy of resource availability (Fahrig, 2013). Second, fragmentation as the number of forest patches present in the local landscape (also termed as fragmentation per se; Fahrig, 2013). The number of patches guantifies the process of habitat partitioning into small, unconnected fragments affecting landscape configuration (spatial arrangement of habitats). Third, edge density, calculated as the total length of forest edge divided by each spatial extent (associated with edge effects, Crouzeilles & Curran, 2016). Edge density quantifies the intensity of edge effects with higher values being characteristic of more fragmented forests. The premise was that the absence of forest leads to the depletion of hosts for epiphytic individuals. While other land covers, such as sparse trees in the pasture matrix and plantations, provide habitat for some epiphyte species, they have far lower species richness than later secondary or old-growth forests (Einzmann & Zotz, 2017; H. Einzmann & Zotz, 2016).

Landscape metrics were measured using the 30-m resolution "global change forest map" from 2018 (hereafter termed GCF; Hansen et al., 2013). We defined a threshold of 50% forest cover (non-forest <=50, and >50% as forest), after validating 334 field

points within and outside forest fragments to five forest cover thresholds (40%, 50%, 60%, 70%, 90%). To validate our threshold of 50% forest cover, we transformed all pixels into binary forest/ non-forest maps for each threshold percentage, then we extracted the cover type of each of our 334 validation points, thus each point would be either forest or non-forest. Finally, we compared the proportion of points per threshold that matches our validation points and the threshold with the highest proportion of points was selected (Table S2). From this binary map, we calculated the proportion of forest cover in the landscape, number of patches of habitat (forest) and edge density exposed to the conditions of the non-forest habitat. We constrained our analyses to forest fragments with patch sizes >9 ha due to the resolution of GCF. Pixels were classified based on the nearest eight pixels connected in any direction allowing us to classify based upon the category of the majority of the surrounded pixels (McGarigal et al., 2012).

#### Data analysis 2.4

We focused on two aspects of scale-of-effects: the direction and the spatial extent. We used a state-of-the-art modelling approach with linear mixed models (GLMM) with Poisson distribution errors to quantify the direction (positive, negative, or not significant correlation) and spatial extent of each landscape metric (forest

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cover, fragmentation and edge density) on each response variable (species richness and total abundance) separately for orchids and NOE. Thus, each diversity metric at plot level was modelled independently with their own set of landscape metrics per spatial extent

In each model, we included a nested spatial random effect (sampling points nested within forests) and an offset parameter (offset = elevation) to account for the expected correlation between epiphyte species richness and elevation (Cardelus et al., 2006; Parra-Sanchez et al., 2023). All explanatory variables were standardized to have means equal to zero and two standard deviations to adjust for the differences in scales (z-transformation). We further explore a not significant effect of the predictors by using a null model, including only the intercept with the same structure of the models (i.e. error family, offset and response variable). Model assumptions were verified with marginal r-squared and residuals for each model were visually checked for normality and homoscedasticity (Nakagawa et al., 2017).

Although four spatial extents overlapped at 1500m extents (Figure 2), we found no spatial autocorrelations across these four models (Durbin-Watson-Test p: <0.05). Several studies have shown no evidence that overlapping landscapes compromised either spatial autocorrelation or the scale-of-effects (Zuckerberg et al., 2012, 2020). We, therefore, considered that our results are robust and conservative.

The direction of the scale-of-effects was obtained from each model (as the sign of each predictor's estimate) and illustrated by plotting the GLMM's estimates with confidence intervals (95%) to convey uncertainty. The spatial extent of the scale-of-effects was obtained as the difference between the model with the lowest Akaike-Information Criterion AIC value against the AIC of each individual model ( $\Delta$ AIC) (Jackson & Fahrig, 2015). The range of spatial extents with  $\Delta AIC < 2$  provide a comparable fit to the observed data. Thus, if all models fall into  $\triangle AIC < 2$  or  $\triangle AIC > 2$ , there is not a detectable spatial extent. However, if, for example, a particular landscape metric from 100 to 1000m has an  $\triangle$ AIC <2 but from 1000 to 1500m shows  $\triangle AIC > 2$ , then our spatial extent would be 100-1000 m. We plotted the  $\triangle$ AIC of each model as a dependent variable against its respective spatial extent to visually assess the spatial extent. Previous studies have used similar statistical approaches with Akaike's Information Criterion (Amiot et al., 2021; Gestich et al., 2019) and parameter estimates (Galán-Acedo et al., 2018), although the use of AIC has been criticized (Arnold, 2010; Galipaud et al., 2017).

Models considering fragmentation metrics and forest cover at the same time had multicollinearity issues >5 VIF (variation inflation factor) due to the high correlation among predictors (>0.80 in the fixed effects matrix correlation). The nature of this collinearity lies in the intrinsic link between habitat loss and fragmentation (Didham & Ewers, 2012; Fletcher et al., 2018; Püttker et al., 2020). Therefore, models with multiple predictors were excluded from the study.

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#### 2.5 Software

All analyses were performed in R Statistical Software (R Core Team, 2021). Data manipulation was done with 'tidyverse' (Wickham et al., 2019) and 'jtools' (Long, 2019). Landscape metrics were computed with 'landscapemetrics' (Hesselbarth et al., 2019), and generalized linear-mixed effect models were fitted using 'Ime4' and 'gImmTMB' (Bates et al., 2015; Magnusson et al., 2017). Plotting was done using 'ggplot2' (Wickham & Winston, 2015) and 'sjPlot' (Lüdecke, 2023).

#### RESULTS 3

#### 3.1 **Epiphyte surveys**

In total, we recorded 16,802 orchid individuals belonging to 332 species (273 species and 59 morphotypes), while 8782 individuals of NOE were found (non-orchids), with a range of 1 to 12 morphospecies in a single plot. Orchids were found in 88% of plots (n = 125), while 96% of plots had NOE such as ferns (136 of 141 plots). We found the highest species richness in a forest plot with 28 species (bosque Andino, 18 orchids, 8 NOE) and 899 individuals (357 orchids, 542 NOE).

#### Landscape context 3.2

Our results show that our forests have, in general, high levels of forest cover in the landscape. Considering the largest spatial extent (2400 m). 69% of the sample forests had ~70% of forest cover (mean: 67%, range 5.9-100%), while between 61 and 78% of the sampled forests had a low number of patches (mean: 37.7, range 4-59), and low to middle levels of edge density (mean: 48.22 range: 10.64-77.62). Across sites, forest cover showed a consistent pattern of high cover at 100-200m (mean: 94.6%), decreasing as the spatial extent increases (mean: 72.2% at radii >800m; Figure S1). The number of patches constantly increased with spatial extent without reaching a plateau (Figure S1). Edge density was higher at the spatial extent between 100 and 800m, gradually flattening with the spatial extent (radii >800m; Figure S1).

## Spatial extent and impact of forest cover on 3.3 community structure

The effect of forest cover on orchids and NOE was consistently positive across community-level metrics (species richness and total abundance), with the scale-of-effects detected at extents of 100-300 m (GLMM forest cover, p < 0.05, Figures 3 and 4; Tables S3 and S4). Likewise, the scale-of-effect of total abundance remains consistent at 200 m (Figure S3; Tables S3 and S4), indicating that the significant relationship between forest cover and community-level metrics are

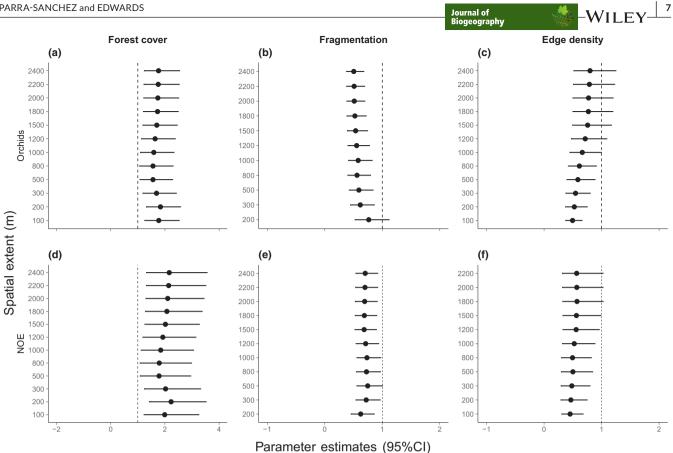


FIGURE 3 Direction of the effects of forest cover (%), fragmentation (number of patches) and edge density on orchids (a-c) and NOE communities (d-f) across 12 radii (100-2400m) in the eastern cordillera of Colombia. Parameter estimates from generalized linear mixed effects models (GLMM) fitted to estimate the effects of each landscape predictor on species richness. Points indicate parameter estimates, error bars indicate 95% confidence intervals, and dashed lines show no effect.

detected at the same spatial scales. However, there was large uncertainty in confidence intervals and limited explanatory power of the models within the spatial extents of 500–1200m across groups, but larger for orchids' total abundance (>300m). This uncertainty suggests that the effect of forest cover might not be reliably detected within these spatial extents (Figure 3, Figure S4; Tables S3 and S4). The total explanatory power of the models is moderate (orchids' conditional pseudo- $r^2$ =0.77-0.99; NOE' conditional pseudo $r^2 = 0.84 - 0.98$ ) and marginal pseudo- $r^2$  ranges between 0.07 and 0.26 across metrics (Tables S3 and S4).

### Spatial extent and impact of fragmentation on 3.4 | community structure

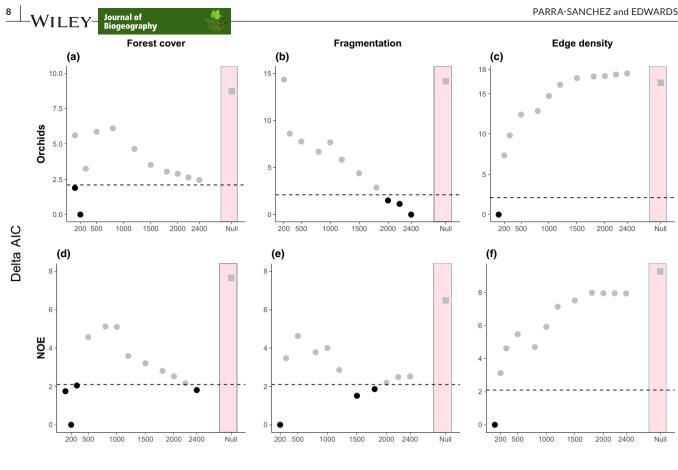
Overall, the scale-of-effects of fragmentation is negative with species richness and total abundance, and detectable between 2000 and 2400m on orchids and 200m for NOEs (GLMM fragmentation, p < 0.05; Tables S5 and S6). Fragmentation consistently exerts negative effects on orchid and NOE across spatial extents, but it is undetectable at 200m in orchids and between 300 and 800m in NOEs (models' performances were similar to null models, delta AIC=3.5-8.6; Figure 3; Tables S5 and S6). The explanatory power

of our models was moderate (orchids conditional pseudo- $r^2 = 0.79$ -0.98; NOE conditional pseudo- $r^2 = 0.54 - 0.74$ ) and marginal pseudo $r^2$  ranged between 0.04 and 0.32.

## Spatial extent and impact of edge effects on 3.5 community structure

The edge density correlates negatively with our metrics and the scale-of-effects was detected between 100 and 200m (Figure 3; Tables S7 and S8), but undetectable at spatial extents larger than 800m (delta AIC=2.2-28.7; Figure 4). At extents ≥1000m, models were not significant and showed  $\triangle$ AIC close to the Null model (GLMM edge density, p > 0.05;  $\Delta AIC < 5$ ). In contrast, NOE' total abundance consistently showed a negative pattern with edge density and were significant across all spatial extents without affecting the scale-of-effects. The total explanatory powers of edge density models' are moderate (NOE conditional pseudo-r<sup>2</sup>=0.75-0.98; orchids conditional pseudo- $r^2 = 0.82 - 0.98$ ) and the component related to the fixed effects alone (marginal pseudo- $r^2$ ) ranged between 0.04 and 0.34.

Edge density was the landscape variable with the highest variance explained across models (Figures S2-S4; Tables S7 and S8).



Spatial extent (m)

**FIGURE 4** Scale of effect of forest cover (%), fragmentation (number of patches) and edge density orchids (a–c) and NOE communities (d–f) in the eastern cordillera of Colombia. Y-axis depicts the difference in Akaike's Information Criterion (Delta AIC) in linear mixed models of each landscape predictor on species richness (q=0). The null model includes only the intercept (pink shade). The dashed line represents the  $\Delta$ AIC threshold ( $\Delta$ AIC=2) were models have equivalent statistical support. Models with  $\Delta$ AIC <2 are indicated with black symbols.

However, the null model was the least plausible model across response variables ( $\Delta_i$  < 5; Tables S3–S8). The scale-of-effects for forest cover, fragmentation and edge density overlapped at different spatial extents. This pattern demonstrates the connection between landscape processes and the scale at which they could interact (Figure 4; Figure S3).

# 4 | DISCUSSION

Unravelling the scale-of-effects (direction and spatial extent) at which landscape drivers shape biodiversity is key for theoretical and practical use. Our empirical study provides evidence that: (1) forest cover has positive effects on orchid and NOE communities across spatial extents, supporting our first expectation; (2) fragmentation and edge effects exert negative impacts on epiphyte diversity, supporting our second expectation, but these impacts are irrespective of the focal group and its scale dependence, rejecting our third expectation; and (3) choosing an inappropriate spatial extent might mask the effects of landscape processes on epiphytic communities. Preserving or improving forest cover, reducing edge effects at low spatial extents (<500 m radius) and reducing fragmentation at larger spatial extents (>2000 m radius) are landscape strategies that might exert positive effects on epiphytic plant diversity.

# 4.1 | The effects of habitat loss and fragmentation on epiphytic communities

We found a consistent positive correlation between epiphytic communities and forest cover, especially between 100 and 200 m (scaleof-effect), while the detectability was obscure across spatial extents <500 and >1200 m. A high proportion of forest cover provides suitable conditions for a greater number of species to coexist and thrive (Arroyo-Rodríguez et al., 2020; Fahrig, 2013). We speculate this reflects sufficient availability of habitat in the landscape allowing species and individuals to reach suitable areas, while the opposite might occur in areas with lower forest cover (Ahuatzin et al., 2022; Arroyo-Rodríguez et al., 2020). However, the ecological processes and mechanisms involved in this response requires further study.

Fragmentation's effect on epiphyte communities is negative but its detection depends on the spatial extent. Fragmentation might affect understorey epiphytes by moderating large-scale processes at 2000-2400m such as frequent dispersal due to lower connectivity across landscapes (Ewers & Didham, 2005). Furthermore, some reviews and empirical studies suggest that fragmentation is not important in landscapes with high forest cover in the landscape (e.g. >60%), because of the low variability in landscape configuration and high structural connectivity (Püttker et al., 2020; Villard & Metzger, 2014). Yet, fragmentation consistently impacts our understorey epiphytic communities in landscapes with high proportion of forest cover (69% of our landscapes have on average 70% of forest cover). Thus, the partitioning of the landscape might be enough to affect dispersal of Andean epiphytic communities (Mccormick & Jacquemyn, 2014; Mondragon et al., 2014). Further studies expanding the spatial extents might enhance our understanding on the effects of fragmentation on epiphytes (as the number of fragments increased constantly as the spatial extent increased; Figure S2).

We found that high edge density predicts lower species richness and abundance. We detected the scale-of-effects for total abundance of NOE (100-500 m), but not for orchids or NOE species richness (Jackson & Fahrig, 2015), which may suggest that epiphytic communities respond to edge effects below our spatial extents, as seen in other studies in the Colombian Andes (ca. 30m, Parra-Sánchez et al., 2016). Epiphytic plants in the Brazilian Atlantic Forest show edge effects can penetrate at least 500 m inside forests and the abundance of 89% of epiphyte species in the canopy and understorey is negatively affected by edge effects reducing the conditions of the core area (Hernandez-Perez & Solano, 2015; Parra-Sanchez & Banks-Leite, 2020). Our results thus highlight the need to quantify edge effects and to create strategies to reduce their effects on biodiversity when designing or managing landscapes for conservation. This is especially important because much of the remaining tropical forest (>70%) globally is now near human-modified habitats (Haddad et al., 2015), pushing forest species to interact with edge conditions that might provide them with less optimal conditions (Ewers & Didham, 2005; Haddad et al., 2015; Tabarelli et al., 2012).

Our findings that fragmentation and edge effects negatively influenced both orchids and NOE might be counterintuitive for two reasons. First, most epiphyte species and all orchids have winddispersed propagules (Zotz, 2016), which should not restrict species dispersal (Einzmann et al., 2017). Second, our studied forests have a relatively high proportion of forest cover, which should decrease inter-patch isolation distances, favouring dispersal (Fahrig, 2013, 2017). Yet our results support the idea that epiphytes might be dispersal limited. Dispersal limitation is invoked in epiphytes because many species' diaspores land very closely to their mother-plants (Acevedo et al., 2020; Cascante-Marín et al., 2009), and epiphyte communities present signalling of recruitment limitation and positive density dependence (Janzen et al., 2020) that might be involved in slow colonization events (Einzmann et al., 2021). Likewise, dispersal limitation might surge from tracking suitable conditions within forests due to the intrinsic dependency of some orchids to mycorrhizae colonies for germination that occur in clusters (McCormick

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et al., 2018; Romero-Salazar et al., 2022). Hence, fragmentation and edge density might further exacerbate epiphytes' intrinsic dispersal limitation by increasing seed mortality of juveniles and seedlings due to dry conditions (Benzing, 1998; Migenis & Ackerman, 1993; Mondragón et al., 2015; Winkler et al., 2005), and forest-dependent species experiencing high mortality in human-modified habitats (Scheffknecht et al., 2010).

Detectability of the scale-of-effects depends on the spatial extent. Thus, landscape processes can be undetected if the spatial extent is chosen a priori or not explored at all. For instance, we found a consistent detrimental trend of habitat loss impacting both groups, but the high uncertainty in the estimates suggests that the effect of forest cover might not be reliably detected within 800-1200-m spatial extents (Figure 2). Low detectability may be a product of the interaction with other landscape drivers (edge density plateaus at 800m) impacting the ability of epiphytes to disperse and utilize alternative habitats. Likewise, the role of fragmentation was undetected at extents of 200m and edge effects at spatial extents >800m. This might be a product of low landscape heterogeneity because fragmentation had the lowest values at 200m (Figure S1), while edge density plateaus at extents >800 m. Thus, the observed pattern might be highly related to landscape variables and regional context, which suggests that landscape management strategies might provide positive impacts for both groups at the same spatial scale (Galán-Acedo et al., 2018; Smith et al., 2011).

The fact that scale-of-effects did not differ, in most cases, between orchids and NOE communities suggests that both groups are shaped by similar patterns and processes such as resource availability or community dynamics in the landscape (e.g. migrations, source-sink dynamics, adaptation to the same understorey microclimate; Ewers & Banks-Leite, 2013; Jackson & Fahrig, 2015; Smith et al., 2011). This might be the product of communities composed of highly ecologically equivalent species (Catchpole & Kirkpatrick, 2010). For instance, long-term monitoring of epiphytic communities has shown that short-distance dispersal and population growth are the dominant process shaping the community dynamics (Cascante-Marín et al., 2009; Einzmann et al., 2021; Einzmann & Zotz, 2017a, 2017b).

Preserving forest cover and reducing the impacts of fragmentation and edge effects have a direct and positive impact on epiphytes diversity at the adequate spatial extent. Hypothetically, if the direction of 'forest cover' is sustained across larger spatial extents than in our study, preserving epiphytes could be compatible with forest restoration strategies, in which the scale-of-effects of restoration actions is positive at buffers of 5–10km (Crouzeilles & Curran, 2016). Indeed, epiphytes transplanted into forests under restoration had high survival rates in human-modified forests from radii <100m (Reid et al., 2016).

# 4.2 | Conservation implications and conclusions

Our study suggests that preserving or increasing the amount of habitat at any spatial extent should promote epiphyte diversity, °⊥w

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although its positive impact might improve if combined with the reduction of fragmentation at coarse spatial (2000–2400 m) and edge effects at fine spatial (200 m) extents. The inclusion of strategies to reduce fragmentation and to maintain or increase forest cover at the same spatial extent would be positive for epiphytic diversity and a good starting point for designing biodiversity-rich landscapes in the tropics.

The constitution in Brazil and Colombia includes legislation to protect forests within private properties in rural areas. Private landowners are responsible for preserving at least 80% of native vegetation in the Brazilian Amazon, 30% in the Brazilian Cerrado and 20% in some Andean municipalities in Colombia (LEI No 12.651, de 25 de maio, 2012; Ley 388 de 1997, 1997). However, the landscape configuration of set-aside areas has not been regulated. Thus, the same percent of forest might be found in different small fragments or even fragmented corridors across private land, increasing edge effects. In terms of landscape management, including combined actions to preserve or encourage low fragmented landscapes with increasing forest cover at the same spatial extent (either at 100–300m or 1500–2400m) should have important benefits for conservation of these epiphytic plants. This scale is probably more suitable for bottom-up actions with stakeholders.

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## CONFLICT OF INTEREST STATEMENT

The authors have no relevant financial or non-financial interests to disclose.

# DATA AVAILABILITY STATEMENT

The data sets generated during and/or analysed during the current study are available at figshare:10.6084/m9.figshare.22223893.v1.

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# REFERENCES

Acevedo, M. A., Beaudrot, L., Meléndez-Ackerman, E. J., & Tremblay, R. L. (2020). Local extinction risk under climate change in a neotropical asymmetrically dispersed epiphyte. *Journal of Ecology*, 108(4), 1553–1564. https://doi.org/10.1111/1365-2745.13361

- Ahuatzin, D. A., González-Tokman, D., Silva, R. R., González, J. E. V., Escobar, F., Ribeiro, M. C., Acosta, J. C. L., & Dáttilo, W. (2022). Forest cover modulates diversity and morphological traits of ants in highly fragmented tropical forest landscapes. *Biodiversity and Conservation*, 31(8–9), 2097–2117. https://doi.org/10.1007/s1053 1-022-02428-3
- Aide, T. M., Grau, H. R., Graesser, J., Andrade-Nuñez, M. J., Aráoz, E., Barros, A. P., Campos-Cerqueira, M., Chacon-Moreno, E., Cuesta, F., Espinoza, R., Peralvo, M., Polk, M. H., Rueda, X., Sanchez, A., Young, K. R., Zarbá, L., & Zimmerer, K. S. (2019). Woody vegetation dynamics in the tropical and subtropical Andes from 2001 to 2014: Satellite image interpretation and expert validation. *Global Change Biology*, 25(6), 2112–2126. https://doi.org/10.1111/gcb.14618
- Amiot, C., Santos, C. C., Arvor, D., Bellón, B., Fritz, H., Harmange, C., Holland, J. D., Melo, I., Metzger, J.-P., Renaud, P.-C., de Roque, F. O., Souza, F. L., & Pays, O. (2021). The scale of effect depends on operational definition of forest cover—Evidence from terrestrial mammals of the Brazilian savanna. *Landscape Ecology*, *36*(4), 973–987. https://doi.org/10.1007/s10980-021-01196-9
- Arnold, T. W. (2010). Uninformative parameters and model selection using Akaike's information criterion. *Journal of Wildlife Management*, 74, 1175-1178. https://doi.org/10.1111/j.1937-2817.2010.tb012 36.x
- Arroyo-Rodríguez, V., & Dias, P. A. D. (2010). Effects of habitat fragmentation and disturbance on howler monkeys: A review. *American Journal* of *Primatology*, 72(1), 1–16. https://doi.org/10.1002/ajp.20753
- Arroyo-Rodríguez, V., Fahrig, L., Tabarelli, M., Watling, J. I., Tischendorf, L., Benchimol, M., Cazetta, E., Faria, D., Leal, I. R., Melo, F. P. L., Morante-Filho, J. C., Santos, B. A., Arasa-Gisbert, R., Arce-Peña, N., Cervantes-López, M. J., Cudney-Valenzuela, S., Galán-Acedo, C., San-José, M., Vieira, I. C. G., ... Tscharntke, T. (2020). Designing optimal human-modified landscapes for forest biodiversity conservation. *Ecology Letters*, 23(9), 1404–1420. https://doi.org/10.1111/ ele.13535
- Bader, M. Y., Rietkerk, M., & Bregt, A. K. (2007). Vegetation structure and temperature regimes of tropical alpine treelines. Arctic, Antarctic, and Alpine Research, 39(3), 353–364. https://doi.org/10.1657/ 1523-0430(06-055)[BADER]2.0.CO;2
- Bates, D., Maechler, M., Bolker, B., & Walker, S. (2015). Fitting linear mixed-effects models using lme4. *Journal of Statistical Software*, 67, 1–48.
- Benzing, D. H. (1998). Vulnerabilities of tropical forests to climate change: The significance of resident epiphytes. In *Potential Impacts* of *Climate Change on Tropical Forest* (pp. 519–540). SpringerLink.
- Brancalion, P. H. S., Niamir, A., Broadbent, E., Crouzeilles, R., Barros, F.
  S. M., Almeyda Zambrano, A. M., Baccini, A., Aronson, J., Goetz,
  S., Leighton Reid, J., Strassburg, B. B. N., Wilson, S., & Chazdon,
  R. L. (2019). Global restoration opportunities in tropical rainforest
  landscapes. *Science Advances*, *5*(7), 1–12. https://doi.org/10.1126/
  sciadv.aav3223
- Brooks, T. M., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A.
  B., Rylands, A. B., Konstant, W. R., Flick, P., Pilgrim, J., Oldfield, S.,
  Magin, G., & Hilton-Taylor, C. (2002). Habitat loss and extinction in the hotspots of biodiversity. *Conservation Biology*, 16(4), 909–923. https://doi.org/10.1046/j.1523-1739.2002.00530.x
- Calderón-Saenz, E. (2007). *Libro rojo de plantas de Colombia*: Volumen 6, Orqídeas, primera parte. Instituto Alexander von Humboldt-Ministerio de Ambiente, Vivienda y Desarrollo Territorial.
- Cardelus, C. L., Colwell, R. K., & Watkins, J. E. (2006). Vascular epiphyte distribution patterns: Explaining the mid-elevation richness peak. *Journal of Ecology*, 94(1), 144–156. https://doi.org/10.1111/j.1365-2745.2005.01052.x
- Carmona-Higuita, M. J., Mendieta-Leiva, G., Gómez-Díaz, J. A., Villalobos, F., Ramos, F. N., Elias, J. P. C., Jiménez-López, D. A., Zuluaga, A., Holst, B., Kessler, M., Mathieu, G., Zizka, A., Zotz, G.,

& Krömer, T. (2023). Conservation status of vascular epiphytes in the neotropics. *Biodiversity and Conservation*, *33*, 51–71. https://doi.org/10.1007/s10531-023-02730-8

- Cascante-Marín, A., von Meijenfeldt, N., de Leeuw, H. M. H., Wolf, J. H. D., Oostermeijer, J. G. B., & den Nijs, J. C. M. (2009). Dispersal limitation in epiphytic bromeliad communities in a costa Rican fragmented montane landscape. *Journal of Tropical Ecology*, 25(1), 63–73. https://doi.org/10.1017/S0266467408005622
- Catchpole, D. J., & Kirkpatrick, J. B. (2010). The outstandingly speciose epiphytic flora of a single strangle fig (*Ficus crassiuscula*) in a Peruvian cloud forest. In L. A. Bruijnzeel, F. N. Scatena, & L. S. Hamilto (Eds.), *Tropical montane cloud forests: Science for conservation and management* (pp. 142–146). Cambridge University Press.
- Coenders-Gerrits, M., Schilperoort, B., & Jiménez-Rodríguez, C. (2020).
   Evaporative processes on vegetation: An inside look. In *Precipitation Partitioning by Vegetation* (pp. 35–48). Springer International Publishing. https://doi.org/10.1007/978-3-030-29702-2\_3
- Crouzeilles, R., & Curran, M. (2016). Which landscape size best predicts the influence of forest cover on restoration success? A global metaanalysis on the scale of effect. *Journal of Applied Ecology*, *53*(2), 440–448. https://doi.org/10.1111/1365-2664.12590
- Didham, R. K., & Ewers, R. M. (2012). Predicting the impacts of edge effects in fragmented habitats: Laurance and Yensen's core area model revisited. *Biological Conservation*, 155, 104–110. https://doi. org/10.1016/j.biocon.2012.06.019
- Einzmann, H., & Zotz, G. (2016). How diverse are epiphyte assemblages in plantations and secondary forests in tropical lowlands? *Tropical Conservation Science*, 9(2), 629–647.
- Einzmann, H. J. R., Weichgrebe, L., & Zotz, G. (2021). Long-term community dynamics in vascular epiphytes on Annona glabra along the shoreline of Barro Colorado Island, Panama. *Journal of Ecology*, 109(4), 1931–1946. https://doi.org/10.1111/1365-2745.13618
- Einzmann, H. J. R., & Zotz, G. (2017a). Dispersal and establishment of vascular epiphytes in human-modified landscapes. *AoB Plants*, 9(6), 1–13. https://doi.org/10.1093/aobpla/plx052
- Einzmann, H. J. R., & Zotz, G. (2017b). "No signs of saturation": Long-term dynamics of vascular epiphyte communities in a human-modified landscape. *Biodiversity and Conservation*, 26(6), 1393–1410. https:// doi.org/10.1007/s10531-017-1306-z
- Etter, A., Andrade, A., Saavedra, K., Amaya, P., Cortés, J., & Arévalo, P. (2021). Ecosistemas colombianos: amenazas y riesgos. Una aplicación de la Lista Roja de Ecosistemas a los ecosistemas terrestres continentales. Pontificia Universidad Javeriana y Conservación Internacional-Colombia.
- Ewers, R. M., & Banks-Leite, C. (2013). Fragmentation impairs the microclimate buffering effect of tropical forests. *PLoS One*, 8(3), e58093. https://doi.org/10.1371/journal.pone.0058093
- Ewers, R. M., & Didham, R. K. (2005). Confounding factors in the detection of species responses to habitat fragmentation. *Biological Reviews*, 81(1), 117–142. https://doi.org/10.1017/S146479310 5006949
- Ewers, R. M., & Didham, R. K. (2008). Pervasive impact of large-scale edge effects on a beetle community. Proceedings of the National Academy of Sciences of the United States of America, 105(14), 5426– 5429. https://doi.org/10.1073/pnas.0800460105
- Fahrig, L. (2003). Effects of habitat fragmentation on biodiversity. Annual Review of Ecology, Evolution, and Systematics, 34(1), 487–515. https://doi.org/10.1146/annurev.ecolsys.34.011802.132419
- Fahrig, L. (2013). Rethinking patch size and isolation effects: The habitat amount hypothesis. *Journal of Biogeography*, 40(9), 1649–1663. https://doi.org/10.1111/jbi.12130
- Fahrig, L. (2017). Ecological responses to habitat fragmentation per Se. Annual Review of Ecology, Evolution, and Systematics, 48, 1–23. https://doi.org/10.1146/annurev-ecolsys-110316-022612
- Fardila, D., Kelly, L. T., Moore, J. L., & McCarthy, M. A. (2017). A systematic review reveals changes in where and how we have studied habitat

loss and fragmentation over 20 years. *Biological Conservation*, 212, 130–138. https://doi.org/10.1016/j.biocon.2017.04.031

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Fischer, J., Abson, D. J., Butsic, V., Chappell, M. J., Ekroos, J., Hanspach, J., Kuemmerle, T., Smith, H. G., & von Wehrden, H. (2014). Land sparing versus land sharing: Moving forward. *Conservation Letters*, 7(3), 149–157. https://doi.org/10.1111/conl.12084

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- Fletcher, R. J., Didham, R. K., Banks-Leite, C., Barlow, J., Ewers, R. M., Rosindell, J., Holt, R. D., Gonzalez, A., Pardini, R., Damschen, E. I., Melo, F. P. L., Ries, L., Prevedello, J. A., Tscharntke, T., Laurance, W. F., Lovejoy, T., & Haddad, N. M. (2018). Is habitat fragmentation good for biodiversity? *Biological Conservation*, 226, 9–15. https:// doi.org/10.1016/j.biocon.2018.07.022
- Flores-Palacios, A., & García-Franco, J. G. (2007). Habitat isolation changes the beta diversity of the vascular epiphyte community in lower montane forest, Veracruz, Mexico. *Biodiversity* and Conservation, 17(1), 191–207. https://doi.org/10.1007/s1053 1-007-9239-6
- Galán-Acedo, C., Arroyo-Rodríguez, V., Estrada, A., & Ramos-Fernández, G. (2018). Drivers of the spatial scale that best predict primate responses to landscape structure. *Ecography*, 41(12), 2027–2037. https://doi.org/10.1111/ecog.03632
- Galipaud, M., Gillingham, M. A. F., & Dechaume-Moncharmont, F.-X. (2017). A farewell to the sum of Akaike weights: The benefits of alternative metrics for variable importance estimations in model selection. *Methods in Ecology and Evolution*, *8*, 1668–1678. https:// doi.org/10.1111/2041-210X.12835
- Gestich, C. C., Arroyo-Rodríguez, V., Ribeiro, M. C., da Cunha, R. G. T., & Setz, E. Z. F. (2019). Unraveling the scales of effect of landscape structure on primate species richness and density of titi monkeys (Callicebus nigrifrons). *Ecological Research*, 34(1), 150–159. https:// doi.org/10.1111/1440-1703.1009
- Haddad, N. M., Brudvig, L. A., Clobert, J., Davies, K. F., Gonzalez, A., Holt, R. D., Lovejoy, T. E., Sexton, J. O., Austin, M. P., Collins, C. D., Cook, W. M., Damschen, E. I., Ewers, R. M., Foster, B. L., Jenkins, C. N., King, A. J., Laurance, W. F., Levey, D. J., Margules, C. R., ... Townshend, J. R. (2015). Habitat fragmentation and its lasting impact on Earth's ecosystems. *Science Advances*, 1(2), e1500052. https://doi.org/10.1126/sciadv.1500052
- Hansen, M. C. C., Potapov, P. V., Moore, R., Hancher, M., Turubanova, S. A. A., Tyukavina, A., Thau, D., Stehman, S. V. V., Goetz, S. J. J., Loveland, T. R. R., Kommareddy, A., Egorov, A., Chini, L., Justice, C. O. O., & Townshend, J. R. G. R. G. (2013). High-resolution global maps of 21st-century forest cover change. *Science*, 342(6160), 850–853. https://doi.org/10.1126/science.1244693
- Hernandez-Perez, E., & Solano, E. (2015). Effects of habitat fragmentation on the diversity of epiphytic orchids from a montane forest of southern Mexico. *Journal of Tropical Ecology*, 31(2), 103–115. https://doi.org/10.1017/S0266467414000662
- Hesselbarth, M. H. K., Sciaini, M., With, K. A., Wiegand, K., & Nowosad, J. (2019). Landscapemetrics: An open-source R tool to calculate landscape metrics. *Ecography*, 42(10), 1648–1657. https://doi.org/ 10.1111/ecog.04617
- Holl, K. D., Reid, J. L., Chaves-Fallas, J. M., Oviedo-Brenes, F., & Zahawi,
  R. A. (2017). Local tropical forest restoration strategies affect tree recruitment more strongly than does landscape forest cover. *Journal of Applied Ecology*, *54*(4), 1091–1099. https://doi.org/10. 1111/1365-2664.12814
- Jackson, H. B., & Fahrig, L. (2015). Are ecologists conducting research at the optimal scale? *Global Ecology and Biogeography*, 24(1), 52–63. https://doi.org/10.1111/geb.12233
- Janzen, T., Zotz, G., & Etienne, R. S. (2020). Community structure of vascular epiphytes: A neutral perspective. Oikos, 129(6), 853–867. https://doi.org/10.1111/oik.06537
- Köster, N., Friedrich, K., Nieder, J., & Barthlott, W. (2009). Conservation of epiphyte diversity in an Andean landscape transformed by human land use. *Conservation Biology: The Journal of the Society*

WILEY- Journal of Biogeogra

for Conservation Biology, 23(4), 911–919. https://doi.org/10.1111/j. 1523-1739.2008.01164.x

- Lüdecke, D. (2023). sjPlot: Data Visualization for Statistics in Social Science (R package version 2.8.14). https://CRAN.R-project.org/package= sjPlot
- Lüttge, U. (1997). Páramos. In Physiological Ecology of Tropical Plants (pp. 349-371). Springer. https://doi.org/10.1007/978-3-662-03340-1 9
- Larrea, M. L., & Werner, F. A. (2010). Response of vascular epiphyte diversity to different land-use intensities in a neotropical montane wet forest. Forest Ecology and Management, 260(11), 1950–1955. https://doi.org/10.1016/j.foreco.2010.08.029
- LEI No 12651, DE 25 DE MAIO. (2012). Código forestal. Presidência da República do Brasil.
- Levin, S. A. (1992). The problem of pattern and scale in ecology. *Ecology*, 73(6), 1943–1967. https://doi.org/10.2307/1941447
- Ley 388 de 1997. (1997). Congreso de la República de Colombia.
- Long, J. (2019). jtools: Analysis and Presentation of Social Scientific Data. R package version 2.0.1. R Core Team.
- Magnusson, A., Skaug, H. J., Nielsen, A., Berg, C. W., Kristensen, K., Maechler, M., van Bentham, K. J., Bolker, B. M., & Brooks, M. E. (2017). glmmTMB: Generalized linear mixed models using template model builder. R package version 0.1.3.
- Malcolm, J. A. Y. R. (1994). Edge effects in central Amazonian Forest fragments. Ecology, 75(8), 2438–2445.
- McCormick, M. K., & Jacquemyn, H. (2014). What constrains the distribution of orchid populations? *New Phytologist*, 202(2), 392–400. https://doi.org/10.1111/nph.12639
- McCormick, M. K., Whigham, D. F., & Canchani-Viruet, A. (2018). Mycorrhizal fungi affect orchid distribution and population dynamics. In New Phytologist (Vol. 219, 4, pp. 1207–1215). Blackwell Publishing Ltd. https://doi.org/10.1111/nph.15223
- McGarigal, K., Cushman, S., & Ene, E. (2012). FRAGSTATS v4: Spatial pattern analysis program for categorical and continuous maps. University of Massachusetts, Amherst.
- McGarigal, K., Tagil, S., & Cushman, S. A. (2009). Surface metrics: an alternative to patch metrics for the quantification of landscape structure. *Landscape Ecology*, 24, 433–450. https://doi.org/10.1007/ s10980-009-9327-y
- Migenis, L. E., & Ackerman, J. D. (1993). Orchid—Phorophyte relationships in a forest watershed in Puerto Rico. *Journal of Tropical Ecology*, 9(2), 231–240. https://doi.org/10.1017/S0266467400007227
- Mondragón, D., Valverde, T., Hernández-Apolinar, M., Mondragon, D., Valverde, T., & Hernandez-Apolinar, M. (2015). Population ecology of epiphytic angiosperms: A review. *Tropical Ecology*, 56(1), 1–39. https://doi.org/10.13140/2.1.4043.5849
- Mondragon, D., & Calvo-Irabien, L. M. (2006). Seed dispersal and germination of the epiphyte tillandsia *Brachycaulos* (Bromeliaceae) in a tropical dry Forest, Mexico. *The Southwestern Naturalist*, 51, 462– 470. https://doi.org/10.1894/0038-4909(2006)51[462:SDAGOT] 2.0.CO;2
- Mondragon, D., Valverde, T., & Hernandez, M. (2014). Population ecology of epiphytic angiosperms: A review. *Tropical Ecology*, 56(1), 1–39. https://doi.org/10.13140/2.1.4043.5849
- Myers, N., Mittermeier, R. A., Mittermeier, C. G., da Fonseca, G. A. B., & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403(6772), 853–858. https://doi.org/10.1038/35002501
- Nakagawa, S., Johnson, P. C. D., & Schielzeth, H. (2017). The coefficient of determination R<sup>2</sup> and intra-class correlation coefficient from generalized linear mixed-effects models revisited and expanded. *Journal of the Royal Society Interface*, 14(134), 20170213. https:// doi.org/10.1098/rsif.2017.0213
- Pérez-Escobar, O. A., Zizka, A., Bermúdez, M. A., Meseguer, A. S., Condamine, F. L., Hoorn, C., Hooghiemstra, H., Pu, Y., Bogarín, D., Boschman, L. M., Pennington, R. T., Antonelli, A., & Chomicki, G. (2022). The Andes through time: Evolution and distribution of

Andean floras. In *Trends in Plant Science* (Vol. 27, 4, pp. 364–378). Elsevier Ltd. https://doi.org/10.1016/j.tplants.2021.09.010

- Püttker, T., Crouzeilles, R., Almeida-Gomes, M., Schmoeller, M., Maurenza, D., Alves-Pinto, H., Pardini, R., Vieira, M. V., Banks-Leite, C., Fonseca, C. R., Metzger, J. P., Accacio, G. M., Alexandrino, E. R., Barros, C. S., Bogoni, J. A., Boscolo, D., Brancalion, P. H. S., Bueno, A. A., Cambui, E. C. B., ... Prevedello, J. A. (2020). Indirect effects of habitat loss via habitat fragmentation: A cross-taxa analysis of forest-dependent species. *Biological Conservation*, 241, 108368. https://doi.org/10.1016/j.biocon.2019.108368
- Parra-Sánchez, E., Armenteras, D., & Retana, J. (2016). Edge influence on diversity of orchids in Andean cloud forests. *Forests*, 7(3), 63. https://doi.org/10.3390/f7030063
- Parra-Sanchez, E., & Banks-Leite, C. (2020). The magnitude and extent of edge effects on vascular epiphytes across the Brazilian Atlantic Forest. Scientific Reports, 10(1), 18847. https://doi.org/10.1038/ s41598-020-75970-1
- Parra-Sanchez, E., Pérez-Escobar, O. A., & Edwards, D. P. (2023). Neutralbased processes overrule niche-based processes in shaping tropical montane orchid communities across spatial scales. *Journal of Ecology*, 111, 1614–1628. https://doi.org/10.1111/1365-2745. 14140
- R Core Team. (2021). R: A language and environment for statistical computing. R Foundation for Statistical Computing.
- Rahbek, C., Borregaard, M. K., Colwell, R. K., Dalsgaard, B., Holt, B. G., Morueta-Holme, N., Nogues-Bravo, D., Whittaker, R. J., & Fjeldså, J. (2019). Humboldt's enigma: What causes global patterns of mountain biodiversity? *Science*, *365*(6458), 1108–1113. https://doi. org/10.1126/science.aax0149
- Reid, J. L., Chaves-Fallas, J. M., Holl, K. D., Zahawi, R. A., & Wulf, M. (2016). Tropical forest restoration enriches vascular epiphyte recovery. *Applied Vegetation Science*, 19(3), 508–517. https://doi.org/ 10.1111/avsc.12234
- Ries, L., Murphy, S. M., Wimp, G. M., & Fletcher, R. J. (2017). Closing persistent gaps in knowledge about edge ecology. *Current Landscape Ecology Reports*, 2(1), 30–41. https://doi.org/10.1007/s4082 3-017-0022-4
- Rodríguez Eraso, N., Armenteras-Pascual, D., & Alumbreros, J. R. (2013). Land use and land cover change in the Colombian Andes: Dynamics and future scenarios. *Journal of Land Use Science*, 8(2), 154–174. https://doi.org/10.1080/1747423X.2011.650228
- Romero-Salazar, N. C., Galvis-Gratz, J. M., & Moreno-López, J. P. (2022). Hongos formadores de micorrizas aislados a partir de raíces de la orquídea Rodriguezia granadensis (LINDL.) RCHB. F. Revista U.D.C.A Actualidad & Divulgación Científica, 25(1). https://doi.org/ 10.31910/rudca.v25.n1.2022.2086
- Ruete, A., Jönsson, M. T., & Snäll, T. (2018). Conservation benefits of international Aichi protection and restoration targets for future epiphyte metapopulations. *Journal of Applied Ecology*, 55(1), 118–128. https://doi.org/10.1111/1365-2664.12964
- San-José, M., Arroyo-Rodríguez, V., & Meave, J. A. (2020). Regional context and dispersal mode drive the impact of landscape structure on seed dispersal. *Ecological Applications*, 30(2), 1–12. https://doi.org/ 10.1002/eap.2033
- Sanford, W. (1968). Distribution of epiphytic orchids in semi-deciduous tropical forest in southern Nigeria. *Journal of Ecology*, *56*(3), 697–705.
- Saura, S. (2020). The habitat amount hypothesis implies negative effects of habitat fragmentation on species richness. Journal of Biogeography, 2019, 1–12. https://doi.org/10.1111/jbi.13958
- Scheffknecht, S., Winkler, M., Hlber, K., Rosas, M. M., & Hietz, P. (2010). Seedling establishment of epiphytic orchids in forests and coffee plantations in Central Veracruz, Mexico. *Journal of Tropical Ecology*, 26(1), 93–102. https://doi.org/10.1017/S0266467409990332
- Smith, A. C., Fahrig, L., & Francis, C. M. (2011). Landscape size affects the relative importance of habitat amount, habitat fragmentation, and

matrix quality on forest birds. *Ecography*, 34(1), 103–113. https://doi.org/10.1111/j.1600-0587.2010.06201.x

- Tabarelli, M., Peres, C. A., & Melo, F. P. L. (2012). The 'few winners and many losers' paradigm revisited: Emerging prospects for tropical forest biodiversity. *Biological Conservation*, 155, 136–140. https:// doi.org/10.1016/j.biocon.2012.06.020
- Tadono, T., Ishida, H., Oda, F., Naito, S., Minakawa, K., & Iwamoto, H. (2014). Precise global DEM generation by ALOS PRISM. ISPRS Annals of the Photogrammetry, Remote Sensing and Spatial Information Sciences, II-4, 71-76. https://doi.org/10.5194/isprs annals-II-4-71-2014
- Taylor, A., Zotz, G., Weigelt, P., Cai, L., Karger, D. N., König, C., & Kreft, H. (2021). Vascular epiphytes contribute disproportionately to global centres of plant diversity. *Global Ecology and Biogeography*, 31(1), 62–74. https://doi.org/10.1111/geb.13411
- Trimble, M. J., & van Aarde, R. J. (2012). Geographical and taxonomic biases in research on biodiversity in human-modified landscapes. *Ecosphere*, 3(12), 16. https://doi.org/10.1890/es12-00299.1
- Turner, M. G., O'Neill, R. V., Gardner, R. H., & Milne, B. T. (1989). Effects of changing spatial extent on landscape pattern analysis. *Landscape Ecology*, 3, 153–162.
- Turner, M. G., Gardner, R. H., & O'neill, R. V., & O'Neill, R. V. (2001). Landscape ecology in theory and practice (Vol. 401). Springer.
- Vancutsem, C., Achard, F., Pekel, J.-F., Vieilledent, G., Carboni, S., Simonetti, D., Gallego, J., Aragao, L. E., & Nasi, R. (2021). Long-term (1990–2019) monitoring of forest cover changes in the humid tropics. Science. *Advances*, 7(10), eabe1603.
- Villard, M. A., & Metzger, J. P. (2014). Beyond the fragmentation debate: A conceptual model to predict when habitat configuration really matters. *Journal of Applied Ecology*, 51(2), 309–318. https://doi.org/ 10.1111/1365-2664.12190
- Werner, F. A. (2011). Reduced growth and survival of vascular epiphytes on isolated remnant trees in a recent tropical montane forest clearcut. Basic and Applied Ecology, 12(2), 172–181. https://doi.org/10. 1016/j.baae.2010.11.002
- Wickham, H., Averick, M., Bryan, J., Chang, W., D'Agostino-McGowan, L., François, R., Grolemund, G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T. L., Miller, E., Bache, S. M., Müller, K., Ooms, J., Robinson, D., Seidel, D. P., Spinu, V., ... Yutani, H. (2019).
  Welcome to the tidyverse. *Journal of Open Source Software*, 4(43), 1686. https://doi.org/10.21105/joss.01686
- Wickham, H., & Winston, C. (2015). Ggplot2: An Implementation of the Grammar of Graphics. The Comprehensive R Archive Network.
- Wiens, J. A. (1989). Spatial scaling in ecology published by: British Ecological Society Stable Spatial scaling in ecology1. Functional Ecology, 3(4), 385–397.
- Wilson, A. M., & Jetz, W. (2016). Remotely sensed high-resolution global cloud dynamics for predicting ecosystem and biodiversity distributions. PLoS Biology, 14(3), 1–20. https://doi.org/10.1371/journal. pbio.1002415
- Winkler, M., Hülber, K., & Hietz, P. (2005). Effect of canopy position on germination and seedling survival of epiphytic bromeliads in a Mexican humid montane forest. *Annals of Botany*, 95(1997), 1039– 1047. https://doi.org/10.1093/aob/mci115
- Woods, C. L., Cardelús, C. L., & Dewalt, S. J. (2015). Microhabitat associations of vascular epiphytes in a wet tropical forest canopy. *Journal* of Ecology, 103(2), 421–430. https://doi.org/10.1111/1365-2745. 12357
- Zotz, G. (2005). Differences in vital demographic rates in three populations of the epiphytic bromeliad, Werauhia sanguinolenta. *Acta Oecologica*, 28(3), 306–312. https://doi.org/10.1016/j.actao.2005. 05.009

Zotz, G. (2013). The systematic distribution of vascular epiphytes-A critical update. *Botanical Journal of the Linnean Society*, 1888, 453–481. https://doi.org/10.1111/boj.12010/full

ournal of iogeography

- Zotz, G. (2016). Plants on Plants–The biology of vascular epiphytes. Springer International Publishing.
- Zotz, G., Weigelt, P., Kessler, M., Kreft, H., & Taylor, A. (2021). EpiList 1.0: a global checklist of vascular epiphytes. *Ecology*, 102(6), e03326. https://doi.org/10.1002/ecy.3326
- Zuckerberg, B., Cohen, J. M., Nunes, L. A., Bernath-plaisted, J., Clare, J. D. J., Gilbert, N. A., Kozidis, S. S., Nelson, S. B. M., Shipley, A. A., & Thompson, K. L. (2020). A Review of Overlapping Landscapes: Pseudoreplication or a Red Herring in Landscape Ecology?
- Zuckerberg, B., Desrochers, A., Hochachka, W. M., Fink, D., Koenig, W. D., & Dickinson, J. L. (2012). Overlapping landscapes: A persistent, but misdirected concern when collecting and analyzing ecological data. *The Journal of Wildlife Management*, 76(5), 1072–1080. https:// doi.org/10.1002/jwmg.326
- Zuleta, D., Benavides, A. M., López-Rios, V., & Duque, A. (2016). Local and regional determinants of vascular epiphyte mortality in the Andean mountains of Colombia. *Journal of Ecology*, 104(3), 841– 849. https://doi.org/10.1111/1365-2745.12563

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# SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

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