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A Multiplexing Activity-Based Protein-Profiling Platform for Dissection of a Native Bacterial Xyloglucan-Degrading System

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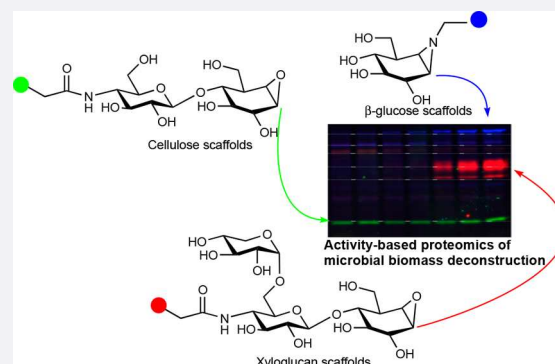


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ABSTRACT: Bacteria and yeasts grow on biomass polysaccharides by expressing and excreting a complex array of glycoside hydrolase (GH) enzymes. Identification and annotation of such GH pools, which are valuable commodities for sustainable energy and chemistries, by conventional means (genomics, proteomics) are complicated, as primary sequence or secondary structure alignment with known active enzymes is not always predictive for new ones. Here we report a “low-tech”, easy-to-use, and sensitive multiplexing activity-based protein-profiling platform to characterize the xyloglucan-degrading GH system excreted by the soil saprophyte, *Cellvibrio japonicus*, when grown on xyloglucan. A suite of activity-based probes bearing orthogonal fluorophores allows for the visualization of accessory *exo*-acting glycosidases, which are then identified using biotin-bearing probes. Substrate specificity of xyloglucanases is directly revealed by imbuing xyloglucan structural elements into bespoke activity-based probes. Our ABPP platform provides a highly useful tool to dissect xyloglucan-degrading systems from various sources and to rapidly select potentially useful ones. The observed specificity of the probes moreover bodes well for the study of other biomass polysaccharide-degrading systems, by modeling probe structures to those of desired substrates.



INTRODUCTION

Natural biodiversity presents a wealth of strategies to enhance fitness by solving complex biological problems. However, when it comes to breaking down biomass, these solutions are not fully understood due to the intricacies of deciphering the roles and behaviors of complex multienzyme systems. Significant effort is currently devoted to obtaining such information as a means to improve functional inference,^{1–3} facilitate biomolecule characterization,⁴ clarify microbial niches,^{5,6} and facilitate the reconstitution of enzyme systems in fermentative workhorse organisms.^{7,8} Comprehensive methods that allow the sensitive and specific detection of several active enzymes simultaneously within a native proteomic background will pave the way to the efficient screening of microbes for biomass-degrading potential. Such methods would, in turn, spotlight vital enzyme components that act on different parts of substrates, aiding in the conversion of biomass polysaccharides into sustainable energy and resources for the chemical sector.

One such biomass polysaccharide with major, yet largely unrealized, biotechnological potential is xyloglucan (Figure 1A). Xyloglucan is a ubiquitous cellulose-binding β -(1,4)-glucan with diversified α -(1,6)-xylose branches extending from two to three of each set of four glucose residues in a semiregular species-specific pattern.^{9,10} Xyloglucan branches have been found to contain β -(1,2)-galactopyranose, α -L-(1,2)-

fucopyranose, α -L-(1,2)-arabinofuranose, and, in rare circumstances, β -(1,2)-xylopyranose residues,¹¹ among others.⁹

Xyloglucan is a predominant hemicellulose in the primary cell walls of many plants.¹² As a result, it is an important source of dietary fiber that sustains key gut microbiota.¹³ Xyloglucan is important for plant development. Its structure and modifications can dictate the mechanical properties of plant cell walls, playing a crucial role in plant growth and response to environmental stresses. Alterations to the xyloglucan structure in the plant cell wall can render plants less susceptible to pathogenic attack.¹⁴ Xyloglucan in soil can support the growth of myriad microbial species, sustaining microbial diversity and, by extension, soil health.¹⁵ The diversity of soil microbes that degrade xyloglucan hints at its importance in soil communities.¹⁶ Understanding the behaviors of xyloglucan-degrading microbes may yet provide insights into novel biological control strategies for plant diseases.

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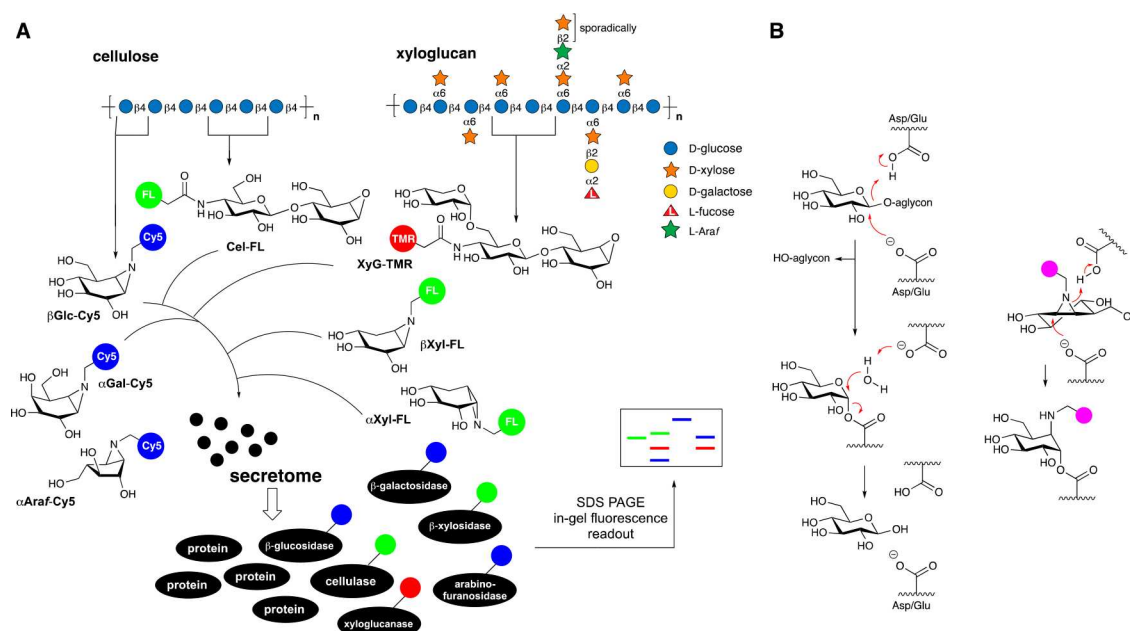


Figure 1. (A) Strategy of the multiplexing activity-based protein profiling (ABPP) platform subject of the here-presented study. Activity-based probes (ABPs) bearing orthogonal fluorophores are designed to emulate xyloglucan/cellulose structural elements. Treatment of secretomes of microbes from various sources grown on xyloglucan followed by SDS-PAGE resolution will yield color-coded fingerprints of both *exo*-acting and *endo*-acting GHs. (B) Mode of action of a retaining β -*exo*-glucosidase and its mechanism-based, covalent, and irreversible inhibition, thereby labeling, by cyclophellitol aziridine ABPs.

Effective xyloglucan-degrading GH systems (recently reviewed by Attia and Brumer¹⁷) contain *exo*-acting glycosidases acting as debranching accessory enzymes and *endo*-acting enzymes (xyloglucanases) that generate short, branched β -(1,4)-glucan oligomers (xyloglucan oligosaccharides).^{18–23} *Exo*-acting enzymes are expected to address each of the glycosidic linkages reported in xyloglucan branches, though the reported enzyme diversity falls short of reported branch diversity.^{9,24} *Endo*-acting enzymes, taking on both linear and branched β -(1,4)-glucans, are needed to effectively degrade xyloglucan, yet discrimination between their substrate specificities using contemporary methods (primary sequence alignment, structure alignment) is unreliable, rendering comparisons of xyloglucan-degrading systems from different organisms challenging.

Cellvibrio japonicus is a saprotrophic bacterium which possesses the remarkable ability to grow in isolation using a variety of hemicellulosic polysaccharides as sole carbon sources.^{25–27} It produces a diverse collection of GHs, assembling enzyme systems that can degrade cellulose, xylans, mannans, and xyloglucans, among others.^{28,29} In contrast to polysaccharide utilization loci found in many gut bacteria,⁵ *C. japonicus* polysaccharide-degrading systems are not tightly organized into complete substrate-specific gene clusters, complicating the enumeration of the components of a complete enzyme system. Recent transcriptomic work identified a cluster of four genes within the *C. japonicus* genome, three of which are essential for xyloglucan oligosaccharide (XyGO) saccharification.²⁸ However, this gene cluster does not include any apparent xyloglucanase or *exo*- β -glucosidase or any other possible accessory activities. Recombinant production and characterization of homology-selected putative xyloglucanases in the *C. japonicus* genome identified Cel5D, Cel5E, Cel5F, and CjGH74A as specific xyloglucanases, but the roles of each of these remain

unclear.^{30,31} Among *exo*- β -glucosidases, Cel3D was found to be xyloglucan oligosaccharide-specific.²⁷ Yet, knocking out these genes only generated a mild growth phenotype, suggesting the presence of additional compensating enzymes.

With the aim to annotate these compensating *exo*-glycosidases and to allow for rapid discrimination between cellulose- and xyloglucan-acting *endo*-glycosidases, we developed a multiplexing activity-based protein-profiling (ABPP) assay, the results of which are presented here. ABPP allows for the rapid and sensitive functional annotation of active enzymes in complex biological samples.^{32–35} Key to the predictive value of an activity-based probe (ABP) is its enzyme selectivity, and we have found in the past that fluorescent, configurational, and functional isosteres of the natural retaining β -glucosidase inhibitor, cyclophellitol, are eminently suited to forecast the substrate preference of both *exo*- and *endo*-acting glycosidases.^{36–45} More so than other GH-directed probe designs, which are often limited in activity and/or GH selectivity, cyclophellitol-based ABPs are viable tools to assess polysaccharide-induced microbial secretomes for desirable activities, which can then be selected for further annotation. Besides targeting a single GH within a biological system, ABPP assays can be executed in a multiplexing format,^{44,45} allowing dissection of complex enzyme systems such as that of the xyloglucan degradome of *C. japonicus* studied here.

The work presented here comprises the design and validation of trisaccharidic xyloglucan (“XyG”) type cyclophellitol probes and their validation as *bona fide*, predictive tools for the identification of xyloglucanase activities and their discrimination from cellulase activities within a xyloglucan-elicited *C. japonicus* degradome. The XyG probes complement our previously described^{36–45} suite of *exo*- and *endo*-GH probes, which we combined to investigate the time-dynamic and substrate concentration-dependent expression of xyloglucanases, cellulases, and retaining β -*exo*-glucosidases in

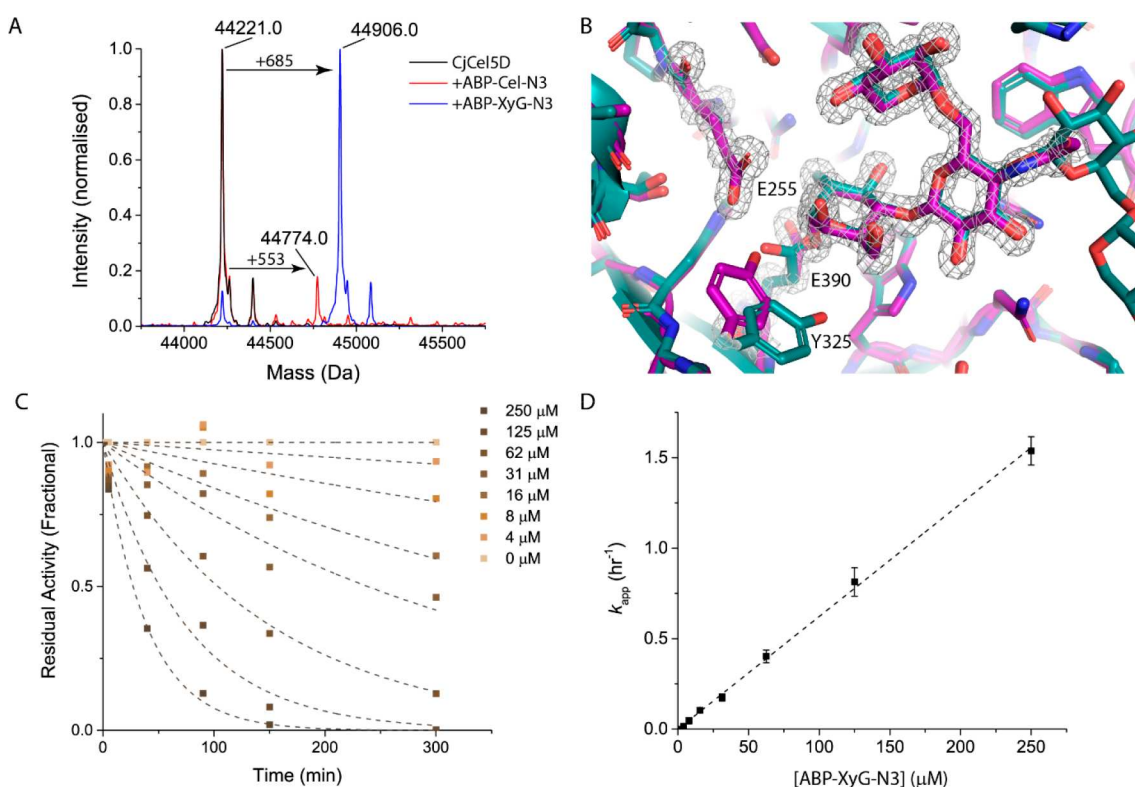


Figure 2. Labeling of xyloglucanases with inhibitors and probes. (A) Intact MS of CjGH5D xyloglucanase treated with 100 μM ABP-Cel-N3, 100 μM ABP-XyG-N3, or vehicle control for 1 h. (B) Crystal structure of CjCel5D labeled with ABP-XyG-N3 (purple). $2F_o - F_c$ density is shown for the ligand and catalytic residues as a gray mesh contoured at 2σ . The complex between CjCel5D and 2-fluoro-XXXG (PDB 6HAA) is superimposed in teal. (C) Residual activity kinetics of CjCel5D inhibited by different concentrations of ABP-XyG-N3. The model fits are shown as dashed lines. 6C4MU-XXXG was used as substrate. (D) k_{app} vs inhibitor concentration for CjCel5D interacting with ABP-XyG-N3. The model fit is shown as a dashed line.

secretomes obtained from *C. japonicus* grown on xyloglucan, xyloglucan oligomers, and other polysaccharide food sources. In this way and using both in-gel detection (with fluorescent probes) and proteomics annotation (with biotinylated probes), we revealed Cel5D and Cel5F to be the exclusive specific retaining xyloglucanases (inverting glycosidases cannot be detected with cyclophellitols), filling distinct functional niches, with Cel5C being a cellulase and Cel3A, Cel3B, and Cel3D acting as β -exo-glucosidases. Utilization of α -L-arabinofuranose, β -galactopyranose, β -xylopyranose, and α -xylopyranose configured cyclophellitol probes allowed further in-depth dissection of *C. japonicus* secretomes and lysates. Our results provide a blueprint for designing multiplexing ABPP assays for the rapid profiling of secretomes of microorganisms grown on specific polysaccharide materials and in which the probes are designed to represent structural elements of the carbohydrate source and, therefore, activities of the corresponding retaining GHs.

RESULTS AND DISCUSSION

Assembly and Validation of the Suite of Activity-Based Probes. Xyloglucan (XyG), the primary carbohydrate source used in this study, contains α -(1,6)-xylose branches at the +2 and/or +3 glucose residues of linear tetraglucose stretches. Other branching sugars include β -(1,2)-galactopyranose, α -L-(1,2)-fucopyranose, α -L-(1,2)-arabinofuranose, and, in rare circumstances, β -(1,2)-xylopyranose.¹¹ These structures (with the exception of fucose) are captured in the set of mono-, di-, and trisaccharidic cyclophellitol probes as depicted

in Figure 1A. These ABPs react with their target GH in a mechanism-based fashion to form a covalent and irreversible enzyme–inhibitor adduct as depicted for inactivation and tagging of retaining β -exo-glucosidases in Figure 1B. Variation in configuration and substitution pattern yields probes designed to target xyloglucanases (denoted as ABP-XyG), cellulases (ABP-Cel),⁴⁵ retaining β -exo-glucosidases (ABP- β Glc),³⁷ retaining β -exo-galactosidases (ABP- β Gal),⁴⁶ retaining β -exo-xylosidases (ABP- β Xyl),⁴⁴ retaining α -exo-xylosidases (ABP- α Xyl), and retaining α -L-exo-arabinofuranosidases (ABP- α Araf).³⁸ The probes were prepared in fluorescent form bearing either a Cy5 dye (denoted with the extension Cy5), a Cy3 dye (Cy3), or a Bodipy-FL dye (FL) to allow for in-gel multiplexing ABPP detection. For the purpose of kinetic measurements, probes were prepared with simple azide (N3) tags. For the purpose of target GH identification by pull-down/mass spectrometry proteomics, all probes were also prepared in biotinylated form (Bio). With the exception of the ABP-XyG and ABP- α Xyl probes, the synthesis and labeling efficacy of all probes on recombinant and/or isolated GHs, as well as detection of these in complex biological samples, have been reported previously.^{37,38,44–46} The full structures of all probes and the synthesis of the ABP-XyG and ABP- α Xyl probes are given in the Supporting Information.

In order to validate the ABP-XyG probes for profiling xyloglucanases in complex biological samples, we first established the potency and mode of action of ABP-XyG-N3 as inhibitor of various previously characterized recombinant xyloglucanases. Incubation of pure recombinant *Bacteroides*

Table 1. Kinetic Parameters for Covalent Inhibition of *endo*-Glucanases by ABP-Cel-N3 and ABP-XyG-N3^a

enzyme	compound	K_I (μ M)	k_{inact} (min^{-1})	k_{inact}/K_I ($\text{M}^{-1} \text{s}^{-1}$)	specificity
PpXG5	ABP-Cel-N3	ND	ND	<0.1	>7.8
	ABP-XyG-N3	>500	>3	0.78 ± 0.08	
BoGH5	ABP-Cel-N3	ND	ND	<0.1	ND
	ABP-XyG-N3	ND	ND	<0.1	
CjCel5D	ABP-Cel-N3	ND	ND	<0.1	>17
	ABP-XyG-N3	>500	>1.4	1.7 ± 0.1	
BaCel5A	ABP-Cel-N3	>200	>0.5	39 ± 3	<0.0026
	ABP-XyG-N3	ND	ND	<0.1	
HiCel7B	ABP-Cel-N3	3.9 ± 0.3	0.50 ± 0.04	2100	0.13
	ABP-XyG-N3	22 ± 2	0.36 ± 0.03	270	
CjCel5B	ABP-Cel-N3	>250	>1	41 ± 3	0.14
	ABP-XyG-N3	>250	>0.2	5.6 ± 0.8	
CjCel5C	ABP-Cel-N3	11 ± 1	0.116 ± 0.005	97	<0.0010
	ABP-XyG-N3	ND	ND	<0.1	

^aWhere it was not possible to obtain distinct k_{inact} and K_I parameters at the inhibitor concentrations tested, the combined k_{inact}/K_I parameter is shown for these cases. ND: not determined. Specificity as determined from the $((k_{\text{inact,XyG-N3}}/K_{I,\text{ABP-XyG-N3}})/(k_{\text{inact,Cel-N3}}/K_{I,\text{Cel-N3}}))$ values.

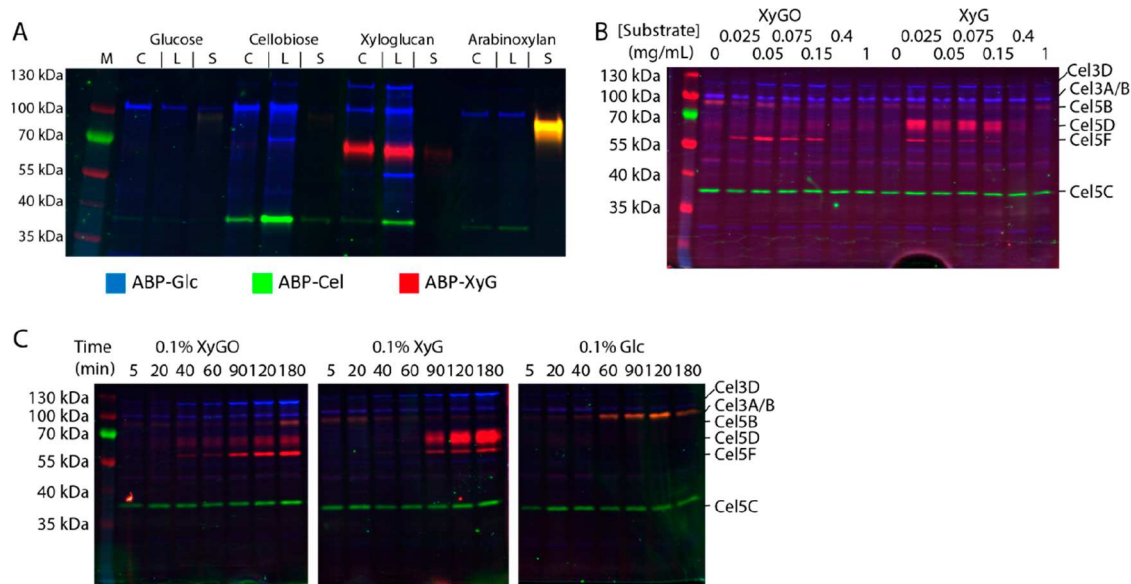


Figure 3. ABPP analysis of *C. japonicus* cultures. (A) Laser-scanning fluorescence image of a 4–20% SDS-PAGE separation of *C. japonicus* proteins following treatment of intact cells (C), lysate (L), or supernatant (S) with a mixture of ABP- β Glc-FL, ABP-Cel-Cy3, and ABP-XyG-Cy5. The carbon source on which the cells were grown is noted above each set of three lanes. (B) Representative gel image from ABPP analysis of *C. japonicus* lysate following 2 h of growth in the presence of increasing concentrations of xyloglucan or xyloglucan oligosaccharides. (C) Representative gel image from ABPP analysis of *C. japonicus* lysate collected over time from growth in the presence of 0.1% xyloglucan oligosaccharides, xyloglucan, or glucose (Glc).

ovatus BoGH5A,¹³ *Paenibacillus pabuli* PpXG5,¹⁹ and *C. japonicus* CjCel5D³⁰ with 100 μ M ABP-XyG-N3 for 1 h under optimal activity conditions gave near-quantitative labeling as assessed by intact protein mass spectrometry analysis, while identical treatment with ABP-Cel-N3 gave minimal labeling (Figure 2A, Supplemental Figures 2–3). Pretreatment of ABP-XyG-N3 and ABP-XyG-Cy5 with 0.1 mg/mL BoGH31 α -xylosidase for 1 h at 37 °C caused no significant change in labeling behavior and no detectable formation of a dextrosylated species by LC-MS, indicating that ABP-XyG-Cy5 and ABP-XyG-N3 are resistant to *exo*-hydrolyase activity (Supplemental Figure 4). This is consistent with the known recognition mode of xyloglucan-specific α -xylosidases, requiring an unsubstituted nonreducing chain terminus.^{47,48}

X-ray diffraction data of the complex between CjCel5D and ABP-XyG-N3 shows near-perfect mimicry of the known glycosyl enzyme intermediate state in the –1 and –2 subsites (Figure 2B). Irreversible inhibition kinetics, measured using bespoke 4-methylumbelliferyl (4MU) and 6-chloro-4-methylumbelliferyl (6C4MU) XXXG fluorogenic substrates (Figure 2C and D and Supplemental Figures 5–25; see the Supporting Information for synthetic details and Tuomivaara et al. for oligosaccharide nomenclature),⁹ showed probe selectivity values $((k_{i,\text{XyG-N3}}/K_{I,\text{XyG-N3}})/(k_{i,\text{Cel-N3}}/K_{I,\text{Cel-N3}}))$ ranging from >17 for CjCel5D to 0.13 and <0.0026 for HiCel7B⁴⁹ and BaCel5A,⁵⁰ two well-known cellulases, respectively (Table 1).

Dissection of the *C. japonicus* Xyloglucanase Degradomes by Multiplexing ABPP. To dissect the native xyloglucan-degrading system of *C. japonicus*, we prepared “primed” cells by growth on glucose to carbon-limited

saturation in MOPS minimal medium (see [Supporting Information](#) for details). Subsequent dilution into medium containing glucose, cellobiose, tamarind xyloglucan, or wheat arabinoxylan was hypothesized to reveal substrate-specific responses. Secretome, intact cell, and lysate samples from each culture were treated with a triplex probe mixture containing ABP- β Glc-FL, ABP-Cel-Cy3, and ABP-XyG-Cy5 ([Figure 3A](#), and [Supplemental Figure 26](#) for Coomassie stain). We observed strong and uniquely xyloglucan-induced production of a ~ 65 kDa ABP-XyG-Cy5-selective outer membrane-associated enzyme. ABP- β Glc-FL treatment revealed two xyloglucan upregulated glucosidase bands at ~ 120 and ~ 58 kDa, and ABP-Cel-Cy3 treatment shows a cellulase band at ~ 40 kDa also observed in the cellobiose culture. Surprisingly, a major ~ 75 kDa secreted band from growth on arabinoxylan reacted with both ABP-Cel-Cy3 and ABP-XyG-Cy5 (major yellow band). This band was observed at a much lower intensity in samples from growth on glucose and was not observed in samples from growth on xyloglucan.

Pulldowns from saturation cultures using ABP- β Glc-Bio, ABP-Cel-Bio, and ABP-XyG-Bio unambiguously identified Cel5D as the only exclusively ABP-XyG-Bio-reactive band, Cel5C as the exclusively ABP-Cel-Bio-reactive band, and Cel3A, Cel3B, and Cel3D as the ABP- β Glc-reactive bands ([Supplemental File 1](#)). The secreted ABP-Cel- and ABP-XyG-reactive band in the arabinoxylan secretome (the intense band in [Figure 3A](#), last lane) was identified as Cel5B. Cel5B and Cel5D both ran ~ 15 kDa heavier on SDS-PAGE than would be expected from their amino acid sequences. To investigate the origin of this discrepancy, ~ 50 μ g of native Cel5B was partially purified from 200 mL of secretome collected from growth of *C. japonicus* on arabinoxylan to carbon-limited saturation via ultrafiltration and anion-exchange chromatography ([Supplemental Figure 27](#)). SDS-PAGE of the purified protein followed by staining with the Pro-Q Emerald glycoprotein gel stain kit (Invitrogen) yielded a strong glycoprotein signal at the band position of Cel5B ([Supplemental Figure 28](#)). Extending this analysis to xyloglucan-grown cell lysate yielded a complex pattern of apparent glycoproteins, including a band at the position of Cel5D. Intact mass of the purified Cel5B measured via denaturing LC-ESI-MS gave a protein peak with a series of deconvoluted mass values from 71 to 75 kDa with spacing of 162 Da, indicating heavy glycosylation with variable hexose content ([Supplemental Figure 29](#)). Acid hydrolysis of the Cel5B sample followed by HPAEC-PAD analysis of the resulting monosaccharides revealed a complex mixture, including peaks that match D-mannose, D-glucose, D-galactose, and L-arabinose standards ([Supplemental Figure 30](#)). L-Arabinose and D-xylose may be derived from the arabinoxylan substrate, but glucose, galactose, and mannose must have been synthesized by *C. japonicus*, underscoring the versatility of this species to grow—and derive the building blocks it needs—from such well-defined, single food stocks as used here. Considering the intact MS and monosaccharide composition, we propose that the underlying glycan structure is a galactoglucomannan O-glycan. Peptide LC-MS/MS analysis of native Cel5B digested with ProAlanaase (Promega) yielded no detectable peptides from the serine-rich linker between the N-terminal catalytic domain and the C-terminal domain ([Supplemental Figure 31](#)).

Having identified the core components of the *C. japonicus* xyloglucan-degrading system, we investigated the time-dynamics and substrate concentration-dependence of xyloglu-

canase expression. We diluted primed *C. japonicus* cells 10-fold into medium containing either xyloglucan or xyloglucan oligosaccharides. ABPP using ABP-XyG-Cy5 on cells harvested during the early induction with xyloglucan revealed two bands, the lower, sharper band running at the expected molecular weight of Cel5E or Cel5F and a higher, more diffuse Cel5D band ([Figure 3B](#), [Supplemental Figure 32](#)). Notably, the lower band was primarily induced by xyloglucan oligosaccharides while Cel5D was primarily induced during growth on xyloglucan. To identify the putative xyloglucanase, primed cells were collected by centrifugation and resuspended in 100 mL of fresh medium containing 150 μ g/mL of xyloglucan oligosaccharides. After 2 h of incubation, cells and secretome were separated by centrifugation and tested for ABP-XyG-Cy5-reactive bands. The band of interest was found exclusively in the secretome while Cel5D was found in the cell fraction, so the secretome was collected and concentrated 50-fold by ultrafiltration prior to pulldown using ABP-XyG-Bio. This identified Cel5D, Cel5E, Cel5F, and Cel5B ([Supplemental Figure 33](#), [Supplemental File 1](#)) as probe-reactive components.

To our surprise, both xyloglucan and xyloglucan oligosaccharides induced xyloglucanase expression more efficiently at low (0.05–0.15 mg/mL) concentrations ([Figure 3B](#), [Supplemental Figure 32](#)). This may suggest that *C. japonicus* can adapt to grow on persistently low levels of xyloglucan, such as those reported in soil samples near root tips.⁵¹ Sampling cultures grown in 0.1% xyloglucan or xyloglucan oligosaccharides over 3 h showed that induction by xyloglucan oligosaccharides occurs rapidly, with a xyloglucanase band detectable after only 30 min ([Figure 3C](#), [Supplemental Figure 32](#)). We also observed that growth on glucose resulted in low-level expression of Cel5B, while growth on xyloglucan resulted in expression of Cel5D correlating with a decrease in observed Cel5B activity, suggesting that Cel5B is acting as a “sensing” enzyme that is repressed by the detection of xyloglucan. Cel3A/B and Cel5C showed no change in expression under any condition tested, indicating that these are constitutively expressed. Interestingly, Cel5D was more strongly expressed in the presence of xyloglucan than xyloglucan oligosaccharides, but expression of Cel5D in the presence of xyloglucan occurred with a lag. This may be explained by a period of time required to generate small, inducing fragments from large xyloglucan molecules. Cel5F/Cel5E and Cel3D expression appeared to be driven primarily by xyloglucan oligosaccharides, indicating that they are differentially regulated from Cel5D. Thus, in spite of being secreted, Cel5F/Cel5E do not appear to be “sensing” enzymes since their expression is dependent on induction by xyloglucan fragments. We speculate that it is instead acting as a “booster” enzyme, aiding the solubilization of xyloglucan.

Not having been previously functionally or structurally characterized in detail, we produced and purified Cel5B and Cel5C recombinantly in *E. coli* to assess further the correlation between probe reactivity and enzyme specificity. We found that Cel5B and Cel5C were both cellulases, efficiently degrading carboxymethylcellulose and mixed-linkage β -glucan ([Supplemental Table 3](#)). Cel5B showed only weak activity toward tamarind xyloglucan while Cel5C had weak activity toward carob galactomannan and no detectable xyloglucanase activity. Measurements of irreversible inhibition kinetics showed strong selectivity of Cel5C for ABP-Cel-N3 over ABP-XyG-N3 and only weak selectivity of Cel5B for ABP-Cel-

N3 over ABP-XyG-N3, matching in-gel fluorescence results (Table 1).

To determine the molecular basis for the reactivity of ABP-XyG-Cy5 with CjCel5B but not CjCel5C, we crystallized both enzymes and solved their structures by molecular replacement in both unliganded and ABP-bound forms (Supplemental Figure 34). Cel inhibitor bound to CjCel5C displaying torsion angles (Φ, Ψ) of ($-83^\circ, 94^\circ$) between the nonreducing β -D-glucose and cyclophellitol moiety in ABP-Cel (Supplemental Figure 34F). O6' is recognized in the -2 position by both H87 and Y137, and O2' is recognized by the backbone carbonyl of S311. In contrast, CjCel5B recognizes ABP-XyG-N3 with ($-83^\circ, 133^\circ$) torsion angles (Supplemental Figure 34C). The consequent twist in the glucose backbone positions the α -(1,6)-xylose residue above W28 and W33, forming a hydrogen bond between O4 and D64. Notably, the active site cleft of CjCel5B is significantly more open beyond the -2 subsite, so we hypothesized that cellulase-specificity in Cel5B is dictated primarily by an inability to accommodate α -(1,6)-xylose residues in the positive subsites. To test this, we synthesized 4MU-XXXG and 6C4MU-XXXG as fluorogenic xyloglucanase substrates (see the Supporting Information for synthetic details). Kinetics for the hydrolysis of 4MU-XXXG and commercially available 4MU-celotetraose were measured to isolate contributions to specificity from the negative subsites. Cel5B showed a ~ 14 -fold preference for 4MU-GGGG over 4MU-XXXG (Supplemental Table 2), roughly in line with its sevenfold preference for ABP-Cel-N3 over ABP-XyG-N3 (Table 1) but highly divergent from its 3000-fold specificity toward carboxymethyl cellulose (CMC) over xyloglucan (Supplemental Table 3), supporting our hypothesis.

The detection of the putative β -xylosidase Xyl39A using ABP- β Glc-FL (the band around 55 kDa in Figure 3A) was particularly interesting, since this enzyme, having 45% identity to the *Xanthomonas citri* XynB,⁵² is adjacent to Cel5D in the genome. Staining *C. japonicus* lysates with the beta-xylose configured probe, ABP- β Xyl-FL and ABP- β Glc-Cy5, confirmed the xyloglucan-dependent expression of Xyl39A and also revealed it has specificity toward ABP- β Xyl-FL (Supplemental Figure 36). Chemical proteomics confirmed that Xyl39A was found in xyloglucan-grown cells and could be pulled down with ABP- β Xyl-Bio and ABP- β Glc-Bio (Supporting Information, Supplemental File 1). To investigate the specificity of CjXyl39A further, we produced the enzyme recombinantly in *E. coli*. Activity measurements against a variety of 4-methylumbelliferyl (4MU) glycosides showed specific recognition of β -D-xylose over other glycosides (Supplemental Table 4). Functionalization of α -(1,6)-xylose branches with β -(1,2)-xylose has been reported in xyloglucan extracted from the leaves¹¹ (but not fruit⁵³) of argan trees, and a recent report has identified xyloglucan β -xylopyranosyl-transferase from *Vaccinium corymbosum*.⁵⁴ We speculate that the coexpression of Xyl39A and Cel5D during growth on xyloglucan indicates an evolved adaptation of *C. japonicus* toward degradation of β -xylosylated xyloglucan; however, we were not able to obtain a suitable sample of β -xylosylated xyloglucan for testing.

Having dissected the *endo*- β -glucanase, *exo*- β -glucosidase, and *exo*- β -xylosidase components of the native *C. japonicus* xyloglucan-degrading system, we turned to the essential *exo*- α -xylosidase, *exo*- α -L-arabinofuranosidase, and *exo*- β -galactosidase activities as potential handles for characterizing xyloglucan-degrading systems using ABPP. Staining with

ABP- α Araf-Cy5 showed the presence of Abf51A in *C. japonicus* under all growth conditions. Abf51A staining was less intense in samples grown on glucose and more intense in samples grown on arabinoxylan (Supplemental Figure 35). We conclude from this that Abf51A displays similar regulatory logic to the *E. coli* *araBAD* operon⁵⁵ and is not coregulated with xyloglucan-degrading machinery.

Using ABP- α Xyl-Cy5 to detect CjXyl31A in the xyloglucan-grown *C. japonicus* lysate during induction by xyloglucan oligosaccharides revealed the emergence of a band at the expected ~ 115 kDa, but identification of the band was hindered by weak reactivity and significant nonspecific labeling (Supplemental Figure 37). We attribute the poor potency and selectivity of this probe to a lack of binding in the positive subsites known to be important for substrate recognition in this enzyme class.⁴⁸ Indeed, the ABP- α Xyl-Cy5 probe performs well on purified recombinant CjXyl31A but is less effective in doped lysates, consistent with its poor performance on *C. japonicus* lysates (Supplemental Figure 38). Finally, comparing the reactivity of ABP- β Gal-Cy5 and ABP- β Glc-Cy5 in *C. japonicus* cell lysates showed strong, and clearly orthogonal, labeling of putative β -glucosidases and β -galactosidases (Supplemental Figure 39). A pulldown from the xyloglucan-grown lysate using ABP- β Gal-Bio revealed the presence of Bgl35A, the known xyloglucan oligosaccharide-specific β -galactosidase, and Bgl2A, an uncharacterized putative β -galactosidase which was previously reported not to be upregulated in response to growth on xyloglucan (Supporting Information, Supplemental File 1).

CONCLUSIONS

We have developed a platform with which native bacterial xyloglucan-degrading systems can be sensitively detected and functionally interrogated. Dissection of native proteomes derived from *C. japonicus* grown on various polysaccharide food sources using these tools reveals features not previously observed, including the production of β -xylosidase during growth on xyloglucan, low-level secretion of Cel5B which we conclude to be a cellulase during growth on glucose, the occurrence of significant enzyme glycosylation, and the different expression and secretion behaviors of the vanguard xyloglucanases, Cel5D, Cel5E, and Cel5F.

The ability to detect xyloglucanases with such high sensitivity and throughput enabled us to measure the concentration-dependence and time-dependence of xyloglucanase activity traced back to specific enzymes in response to different inducers. This revealed surprisingly sensitive xyloglucan detection by *C. japonicus*. This sensitivity may reflect a low-xyloglucan ecological niche where *C. japonicus* thrives.

Building on these developing capabilities, we envision the assembly of different polysaccharide-specific toolkits to enable the characterization of native component enzymes from diverse microbial polysaccharide-degrading systems. These toolkits will provide a significant boost in speed, data richness, and robustness compared to state-of-the-art carbohydrate zymography techniques.⁵⁶ One significant advantage of cyclophellitol-based probe designs is their specificity, leading to clear, interpretable results as demonstrated here. Indeed, a developing toolkit for the analysis of xylanases and cellulases was recently applied to enzyme discovery from diverse fungal secretomes.⁵⁷ ABPP methods will continue to facilitate the characterization of native component enzymes from a plethora

of microbial species, shedding light on the nuanced strategies that microbes employ to degrade and assimilate complex polysaccharides.

Importantly, known, or putative, glycosidase products, which are the result of enzyme recognition and processing of specific polysaccharide substructural stretches, will continue to be imbued in mechanism-based probe designs, extending what has been reported here for linear and branched hemicellulose structures (cellulose versus xyloglucan). This allows the rapid empirical establishment of enzyme specificities in situations where such preferences cannot be gleaned from genomic data alone.

The exquisite specificity of our cyclophellitol-based probe designs compares well to alternative probe designs,^{58,59} leading to relatively simple gel images, with fluorescent bands pointing to probe-reactive proteins that, in all likelihood, feature substrate specificities correlating with that of the probe structure. It should be noted that the suite of probes presented here—indeed probes based on the cyclophellitol scaffold—are reactive toward retaining glycosidases only, excluding inverting glycosidases for identification using our platform. This caveat aside, designing probes targeting *exo*- and *endo*-glycosidases produced to digest different biomass polysaccharides is expected to shed light also in other microbial digestive systems. As well, and as was demonstrated recently, bespoke probes can also be used in machine-learning-assisted, *de novo* glycosidase design.⁶⁰

■ ASSOCIATED CONTENT

Data Availability Statement

The data that support the findings of this study are openly available in the protein databank at <https://www.rcsb.org/>, reference numbers 8BQA, 8BQB, 8BQC, 8BN7, and 8OZ1.

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acscentsci.3c00831>.

Excel sheet of proteomic data (XLSX)

Detailed experimental conditions and methods: enzyme production, structure solution, characterization, biological assays, and organic synthesis (Figures S1–S39 and Tables S1–S4) (PDF)

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Author Contributions

N.G.S.M. carried out the biochemical and structural biology experiments with assistance from W.A.O. Synthesis of ABPs, inhibitors and enzyme substrates was performed by Q.P.O.F. C.B., T.B., L.I.W., and C.d.B. N.G.S.M., J.D.C.C., L.I.W., H.S.O., and G.J.D. wrote the manuscript. H.S.O. and G.J.D. conceived and supervised the research and obtained the funding required.

Notes

The authors declare no competing financial interest.

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■ REFERENCES

- (1) Helbert, W.; Poulet, L.; Drouillard, S.; Mathieu, S.; Loiodice, M.; Couturier, M.; Lombard, V.; Terrapon, N.; Turchetto, J.; Vincentelli, R.; et al. Discovery of Novel Carbohydrate-Active Enzymes through the Rational Exploration of the Protein Sequences Space. *Proc. Natl. Acad. Sci. U. S. A.* **2019**, *116* (13), 6063–6068.
- (2) Hess, M.; Sczyrba, A.; Egan, R.; Kim, T. W.; Chokhawala, H.; Schroth, G.; Luo, S.; Clark, D. S.; Chen, F.; Zhang, T.; et al. Metagenomic Discovery of Biomass-Degrading Genes and Genomes from Cow Rumen. *Science* (1979) **2011**, *331* (6016), 463–467.
- (3) Piao, H.; Froula, J.; Du, C.; Kim, T. W.; Hawley, E. R.; Bauer, S.; Wang, Z.; Ivanova, N.; Clark, D. S.; Klenk, H. P.; et al. Identification of Novel Biomass-Degrading Enzymes from Genomic Dark Matter:

Populating Genomic Sequence Space with Functional Annotation. *Biotechnol. Bioeng.* **2014**, *111* (8), 1550–1565.

(4) Ndeh, D.; Rogowski, A.; Cartmell, A.; Luis, A. S.; Baslé, A.; Gray, J.; Venditto, I.; Briggs, J.; Zhang, X.; Labourel, A.; et al. Complex Pectin Metabolism by Gut Bacteria Reveals Novel Catalytic Functions. *Nature* **2017**, *544* (7648), 65–70.

(5) Terrapon, N.; Lombard, V.; Drula, É.; Lapébie, P.; Al-Masaudi, S.; Gilbert, H. J.; Henrissat, B. PULDB: The Expanded Database of Polysaccharide Utilization Loci. *Nucleic Acids Res.* **2018**, *46* (D1), D677–D683.

(6) Louca, S.; Polz, M. F.; Mazel, F.; Albright, M. B. N.; Huber, J. A.; O'Connor, M. I.; Ackermann, M.; Hahn, A. S.; Srivastava, D. S.; Crowe, S. A.; et al. Function and Functional Redundancy in Microbial Systems. *Nature Ecology & Evolution* **2018**, *2* (6), 936–943.

(7) Yang, D.; Park, S. Y.; Park, Y. S.; Eun, H.; Lee, S. Y. Metabolic Engineering of *Escherichia Coli* for Natural Product Biosynthesis. *Trends Biotechnol.* **2020**, *38* (7), 745–765.

(8) Carbonell, P.; Koch, M.; Duigou, T.; Faulon, J. L. Enzyme Discovery: Enzyme Selection and Pathway Design. In *Methods in Enzymology*; Academic Press Inc.: Cambridge, MA, 2018; Vol. 608, pp 3–27.

(9) Tuomivaara, S. T.; Yaoi, K.; O'Neill, M. A.; York, W. S. Generation and Structural Validation of a Library of Diverse Xyloglucan-Derived Oligosaccharides, Including an Update on Xyloglucan Nomenclature. *Carbohydr. Res.* **2015**, *402*, 56–66.

(10) Pauly, M.; Keegstra, K. Biosynthesis of the Plant Cell Wall Matrix Polysaccharide Xyloglucan. *Annu. Rev. Plant Biol.* **2016**, *67*, 235–259.

(11) Ray, B.; Loutelier-Bourhis, C.; Lange, C.; Condamine, E.; Driouch, A.; Lerouge, P. Structural Investigation of Hemicellulosic Polysaccharides from *Argania Spinosa*: Characterisation of a Novel Xyloglucan Motif. *Carbohydr. Res.* **2004**, *339* (2), 201–208.

(12) Park, Y. B.; Cosgrove, D. J. Xyloglucan and Its Interactions with Other Components of the Growing Cell Wall. *Plant Cell Physiol* **2015**, *56* (2), 180–194.

(13) Larsbrink, J.; Rogers, T. E.; Hemsworth, G. R.; McKee, L. S.; Tauzin, A. S.; Spadiut, O.; Klintner, S.; Pudlo, N. A.; Urs, K.; Koropatkin, N. M.; et al. A Discrete Genetic Locus Confers Xyloglucan Metabolism in Select Human Gut Bacteroidetes. *Nature* **2014**, *506* (7489), 498–502.

(14) Tedman-Jones, J. D.; Lei, R.; Jay, F.; Fabro, G.; Li, X.; Reiter, W. D.; Brearley, C.; Jones, J. D. G. Characterization of Arabidopsis Mur3 Mutations That Result in Constitutive Activation of Defence in Petioles, but Not Leaves. *Plant Journal* **2008**, *56* (5), 691–703.

(15) Zhang, J.; Liu, W.; Bu, J.; Lin, Y.; Bai, Y. Host Genetics Regulate the Plant Microbiome. *Curr. Opin Microbiol.* **2023**, *72*, 102268.

(16) Drula, E.; Garron, M. L.; Dogan, S.; Lombard, V.; Henrissat, B.; Terrapon, N. The Carbohydrate-Active Enzyme Database: Functions and Literature. *Nucleic Acids Res.* **2022**, *50* (D1), D571–D577.

(17) Attia, M. A.; Brumer, H. Recent Structural Insights into the Enzymology of the Ubiquitous Plant Cell Wall Glycan Xyloglucan. *Curr. Opin Struct. Biol.* **2016**, *40*, 43–53.

(18) McGregor, N.; Morar, M.; Fenger, T. H.; Stogios, P.; Lenfant, N.; Yin, V.; Xu, X.; Evdokimova, E.; Cui, H.; Henrissat, B.; et al. Structure-Function Analysis of a Mixed-Linkage β -Glucanase/Xyloglucanase from the Key Ruminant Bacteroidetes *Prevotella bryantii* B14. *J. Biol. Chem.* **2016**, *291* (3), 1175–1197.

(19) Gloster, T. M.; Ibatullin, F. M.; Macauley, K.; Eklöf, J. M.; Roberts, S.; Turkenburg, J. P.; Bjørnvad, M. E.; Jørgensen, P. L.; Danielsen, S.; Johansen, K. S.; et al. Characterization and Three-Dimensional Structures of Two Distinct Bacterial Xyloglucanases from Families GH5 and GH12. *J. Biol. Chem.* **2007**, *282* (26), 19177–19189.

(20) Calzado, F.; Prates, E. T.; Gonçalves, T. A.; Rubio, M. V.; Zubieta, M. P.; Squina, F. M.; Skaf, M. S.; Damásio, A. R. L. Molecular Basis of Substrate Recognition and Specificity Revealed in Family 12 Glycoside Hydrolases. *Biotechnol. Bioeng.* **2016**, *113* (12), 2577–2586.

(21) Simmons, T. J.; Fry, S. C. Bonds Broken and Formed during the Mixed-Linkage Glucan: Xyloglucan Endotransglucosylase Reaction Catalysed by Equisetum Hetero-Trans- β -Glucanase. *Biochem. J.* **2017**, *474* (7), 1055–1070.

(22) Ariza, A.; Eklöf, J. M.; Spadiut, O.; Offen, W. A.; Roberts, S. M.; Besenmatter, W.; Friis, E. P.; Skjot, M.; Wilson, K. S.; Brumer, H. Structure and Activity of *Paenibacillus Polymyxa* Xyloglucanase from Glycoside Hydrolase Family 44. *J. Biol. Chem.* **2011**, *286*, 33890–33900.

(23) Stratilová, B.; Firáková, Z.; Klaudivy, J.; Šesták, S.; Kozmon, S.; Strouhalová, D.; Garajová, S.; Ait-Mohand, F.; Horváthová, Á.; Farkaš, V.; et al. Engineering the Acceptor Substrate Specificity in the Xyloglucan Endotransglycosylase TmXET6.3 from *Nasturtium* Seeds (*Tropaeolum Majus* L.). *Plant Mol. Biol.* **2019**, *100* (1), 181–197.

(24) Hsieh, Y. S. Y.; Harris, P. J. Xyloglucans of Monocotyledons Have Diverse Structures. *Mol. Plant* **2009**, *2*, 943.

(25) Gardner, J. G.; Keating, D. H. Genetic and Functional Genomic Approaches for the Study of Plant Cell Wall Degradation in *Cellvibrio Japonicus*. In *Methods in Enzymology*; Academic Press Inc.: Cambridge, MA, 2012; Vol. 510, pp 331–347.

(26) Ueda, K.; Ishikawa, S.; Itami, T.; Asai, T. Studies on the Aerobic Mesophilic Cellulose-Decomposing Bacteria. *J. Agric. Chem. Soc. Japan* **1952**, *26* (1), 35–41.

(27) Nelson, C. E.; Attia, M. A.; Rogowski, A.; Morland, C.; Brumer, H.; Gardner, J. G. Comprehensive Functional Characterization of the Glycoside Hydrolase Family 3 Enzymes from *Cellvibrio Japonicus* Reveals Unique Metabolic Roles in Biomass Saccharification. *Environ. Microbiol.* **2017**, *19* (12), S025–S039.

(28) Larsbrink, J.; Thompson, A. J.; Lundqvist, M.; Gardner, J. G.; Davies, G. J.; Brumer, H. A Complex Gene Locus Enables Xyloglucan Utilization in the Model Saprophyte *Cellvibrio Japonicus*. *Mol. Microbiol.* **2014**, *94*, 418.

(29) DeBoy, R. T.; Mongodin, E. F.; Fouts, D. E.; Tailford, L. E.; Khouri, H.; Emerson, J. B.; Mohamoud, Y.; Watkins, K.; Henrissat, B.; Gilbert, H. J. Insights into Plant Cell Wall Degradation from the Genome Sequence of the Soil Bacterium *Cellvibrio Japonicus*. *J. Bacteriol.* **2008**, *190*, S455.

(30) Attia, M. A.; Nelson, C. E.; Offen, W. A.; Jain, N.; Davies, G. J.; Gardner, J. G.; Brumer, H. In Vitro and in Vivo Characterization of Three *Cellvibrio Japonicus* Glycoside Hydrolase Family 5 Members Reveals Potent Xyloglucan Backbone-Cleaving Functions. *Biotechnol. Biofuels* **2018**, *11* (1), 45.

(31) Attia, M.; Stepper, J.; Davies, G. J.; Brumer, H. Functional and Structural Characterization of a Potent GH74 Endo-Xyloglucanase from the Soil Saprophyte *Cellvibrio Japonicus* Unravels the First Step of Xyloglucan Degradation. *FEBS Journal* **2016**, *283*, 1701.

(32) Liu, Y.; Patricelli, M. P.; Cravatt, B. F. Activity-Based Protein Profiling: The Serine Hydrolases. *Proc. Natl. Acad. Sci. U. S. A.* **1999**, *96* (26), 14694–14699.

(33) Leung, D.; Hardouin, C.; Boger, D. L.; Cravatt, B. F. Discovering Potent and Selective Reversible Inhibitors of Enzymes in Complex Proteomes. *Nat. Biotechnol.* **2003**, *21* (6), 687–691.

(34) Willems, L. I.; Overkleeft, H. S.; Van Kasteren, S. I. Current Developments in Activity-Based Protein Profiling. *Bioconjug Chem.* **2014**, *25* (7), 1181–1191.

(35) Wu, L.; Armstrong, Z.; Schröder, S. P.; de Boer, C.; Artola, M.; Aerts, J. M.; Overkleeft, H. S.; Davies, G. J. An Overview of Activity-Based Probes for Glycosidases. *Curr. Opin Chem. Biol.* **2019**, *53*, 25–36.

(36) Witte, M. D.; Kallemeyn, W. W.; Aten, J.; Li, K. Y.; Strijland, A.; Donker-Koopman, W. E.; Van Den Nieuwendijk, A. M. C. H.; Bleijlevens, B.; Kramer, G.; Florea, B. I.; et al. Ultrasensitive In Situ Visualization of Active Glucocerebrosidase Molecules. *Nat. Chem. Biol.* **2010**, *6* (12), 907–913.

(37) Kallemeyn, W. W.; Li, K. Y.; Witte, M. D.; Marques, A. R. A.; Aten, J.; Scheij, S.; Jiang, J.; Willems, L. I.; Voorn-Brouwer, T. M.; Van Roomen, C. P. A. A.; et al. Novel Activity-Based Probes for Broad-Spectrum Profiling of Retaining β -Exoglucosidases in Situ and

in Vivo. *Angewandte Chemie - International Edition* **2012**, *51* (50), 12529–12533.

(38) McGregor, N. G. S.; Artola, M.; Nin-Hill, A.; Linzel, D.; Haon, M.; Reijngoud, J.; Ram, A.; Rosso, M. N.; Van Der Marel, G. A.; Codee, J. D. C.; et al. Rational Design of Mechanism-Based Inhibitors and Activity-Based Probes for the Identification of Retaining α -l-Arabinofuranosidases. *J. Am. Chem. Soc.* **2020**, *142* (10), 4648–4662.

(39) Wu, L.; Jiang, J.; Jin, Y.; Kallemeyn, W. W.; Kuo, C. L.; Artola, M.; Dai, W.; Van Elk, C.; Van Eijk, M.; van der Marel, G. A.; et al. Activity-Based Probes for Functional Interrogation of Retaining β -Glucuronidases. *Nat. Chem. Biol.* **2017**, *13* (8), 867–873.

(40) Armstrong, Z.; Kuo, C. L.; Lahav, D.; Liu, B.; Johnson, R.; Beenakker, T. J. M.; De Boer, C.; Wong, C. S.; Van Rijssel, E. R.; Debets, M. F.; et al. Manno- Epi-Cyclophellitols Enable Activity-Based Protein Profiling of Human α -Mannosidases and Discovery of New Golgi Mannosidase II Inhibitors. *J. Am. Chem. Soc.* **2020**, *142* (30), 13021–13029.

(41) McGregor, N. G. S.; Coines, J.; Borlandelli, V.; Amaki, S.; Artola, M.; Nin-Hill, A.; Linzel, D.; Yamada, C.; Arakawa, T.; Ishiwata, A.; et al. Cysteine Nucleophiles in Glycosidase Catalysis: Application of a Covalent β -l-Arabinofuranosidase Inhibitor. *Angewandte Chemie - International Edition* **2021**, *60* (11), 5754–5758.

(42) Marques, A. R. A.; Willems, L. I.; Herrera Moro, D.; Florea, B. I.; Scheij, S.; Ottenhoff, R.; van Roomen, C. P. A. A.; Verhoek, M.; Nelson, J. K.; Kallemeyn, W. W.; et al. A Specific Activity-Based Probe to Monitor Family GH59 Galactosylceramidase, the Enzyme Deficient in Krabbe Disease. *ChemBioChem.* **2017**, *18* (4), 402–412.

(43) Chen, Y.; Armstrong, Z.; Artola, M.; Florea, B. I.; Kuo, C.-L.; de Boer, C.; Rasmussen, M. S.; Abou Hachem, M.; van der Marel, G. A.; Codee, J. D. C.; et al. Activity-Based Protein Profiling of Retaining α -Amylases in Complex Biological Samples. *J. Am. Chem. Soc.* **2021**, *143* (5), 2423–2432.

(44) Schröder, S. P.; De Boer, C.; McGregor, N. G. S.; Rowland, R. J.; Moroz, O.; Blagova, E.; Reijngoud, J.; Arentshorst, M.; Osborn, D.; Morant, M. D.; et al. Dynamic and Functional Profiling of Xylan-Degrading Enzymes in *Aspergillus* Secretomes Using Activity-Based Probes. *ACS Cent Sci.* **2019**, *5* (6), 1067–1078.

(45) de Boer, C.; McGregor, N. G. S.; Peterse, E.; Schröder, S. P.; Florea, B. I.; Jiang, J.; Reijngoud, J.; Ram, A. F. J.; van Wezel, G. P.; van der Marel, G. A.; et al. Glycosylated Cyclophellitol-Derived Activity-Based Probes and Inhibitors for Cellulases. *RSC Chem. Biol.* **2020**, *1* (3), 148–155.

(46) Kuo, C.-L.; Su, Q.; van den Nieuwendijk, A. M. C. H.; Beenakker, T. J. M.; Offen, W. A.; Willems, L. I.; Boot, R. G.; Sarris, A. J.; Marques, A. R. A.; Codee, J. D. C. The Development of a Broad-Spectrum Retaining β -Exo-Galactosidase Activity-Based Probe. *Organ. Biomol. Chem.* **2023**, *21*, 7813.

(47) Silipo, A.; Larsbrink, J.; Marchetti, R.; Lanzetta, R.; Brumer, H.; Molinaro, A. NMR Spectroscopic Analysis Reveals Extensive Binding Interactions of Complex Xyloglucan Oligosaccharides with the *Cellvibrio Japonicus* Glycoside Hydrolase Family 31 α -Xylosidase. *Chem.—Eur. J.* **2012**, *18* (42), 13395–13404.

(48) Larsbrink, J.; Izumi, A.; Ibatullin, F. M.; Nakhai, A.; Gilbert, H. J.; Davies, G. J.; Brumer, H. Structural and Enzymatic Characterization of a Glycoside Hydrolase Family 31 α -Xylosidase from *Cellvibrio Japonicus* Involved in Xyloglucan Saccharification. *Biochem. J.* **2011**, *436* (3), 567–580.

(49) Mackenzie, L. F.; Sulzenbacher, G.; Divne, C.; Jones, T. A.; Wöldike, H. F.; Schülein, M.; Withers, S. G.; Davies, G. J. Crystal Structure of the Family 7 Endoglucanase I (Cel7B) from *Humicola Insolens* at 2.2 Å Resolution and Identification of the Catalytic Nucleophile by Trapping of the Covalent Glycosyl-Enzyme Intermediate. *Biochem. J.* **1998**, *335* (2), 409–416.

(50) Varrot, A.; Davies, G. J. Direct Experimental Observation of the Hydrogen-Bonding Network of a Glycosidase along Its Reaction Coordinate Revealed by Atomic Resolution Analyses of Endoglucanase Cel5A. *Acta Crystallogr. D Biol. Crystallogr.* **2003**, *59* (3), 447–452.

(51) Galloway, A. F.; Pedersen, M. J.; Merry, B.; Marcus, S. E.; Blacker, J.; Benning, L. G.; Field, K. J.; Knox, J. P. Xyloglucan Is Released by Plants and Promotes Soil Particle Aggregation. *New Phytol* **2018**, *217* (3), 1128–1136.

(52) Morais, M. A. B. de; Polo, C. C.; Domingues, M. N.; Persinoti, G. F.; Pirolla, R. A. S.; de Souza, F. H. M.; Correa, J. B. de L.; dos Santos, C. R.; Murakami, M. T. Exploring the Molecular Basis for Substrate Affinity and Structural Stability in Bacterial GH39 β -Xylosidases. *Front. Bioeng. Biotechnol.* **2020**, *8*, 419.

(53) Aboughe-Angone, S.; Nguema-Ona, E.; Ghosh, P.; Lerouge, P.; Ishii, T.; Ray, B.; Driouich, A. Cell Wall Carbohydrates from Fruit Pulp of *Argania Spinosa*: Structural Analysis of Pectin and Xyloglucan Polysaccharides. *Carbohydr. Res.* **2008**, *343* (1), 67–72.

(54) Immelmann, R.; Gawenda, N.; Ramírez, V.; Pauly, M. Identification of a Xyloglucan Beta-xylopyranosyltransferase from *Vaccinium Corymbosum*. *Plant Direct* **2023**, *7* (7), e514.

(55) Schleif, R. AraC Protein, Regulation of the l-Arabinose Operon in *Escherichia Coli*, and the Light Switch Mechanism of AraC Action. *FEMS Microbiol. Rev.* **2010**, *34* (5), 779–796.

(56) Lin, M. C.; Kuo, H. W.; Kao, M. R.; Lin, W. D.; Li, C. W.; Hung, K. S.; Yang, S. C.; Yu, S. M.; Ho, T. H. D. From Simple and Specific Zymographic Detections to the Annotation of a Fungus *Daldinia Caldarium* D263 That Encodes a Wide Range of Highly Bioactive Cellulolytic Enzymes. *Biotechnol. Biofuels* **2021**, *14* (1), 1–15.

(57) McGregor, N. G. S.; de Boer, C.; Santos, M.; Haon, M.; Navarro, D.; Schroder, S.; Berrin, J. G.; Overkleeft, H. S.; Davies, G. J. Activity-Based Protein Profiling Reveals Dynamic Substrate-Specific Cellulase Secretion by Saprotrophic Basidiomycetes. *Biotechnol. Biofuels Bioprod.* **2022**, *15* (1), 1–14.

(58) Chauvigné-Hines, L. M.; Anderson, L. N.; Weaver, H. M.; Brown, J. N.; Koech, P. K.; Nicora, C. D.; Hofstad, B. A.; Smith, R. D.; Wilkins, M. J.; Callister, S. J.; et al. Suite of Activity-Based Probes for Cellulose-Degrading Enzymes. *J. Am. Chem. Soc.* **2012**, *134* (50), 20521–20532.

(59) Jain, N.; Tamura, K.; Déjean, G.; Van Petegem, F.; Brumer, H. Orthogonal Active-Site Labels for Mixed-Linkage Endo- β -Glucanases. *ACS Chem. Biol.* **2021**, *16* (10), 1968–1984.

(60) Lipsh-Sokolik, R.; Khersonsky, O.; Schröder, S. P.; de Boer, C.; Hoch, S. Y.; Davies, G. J.; Overkleeft, H. S.; Fleishman, S. J. Combinatorial Assembly and Design of Enzymes. *Science* (1979) **2023**, *379* (6628), 195–201.