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THEMED ISSUE REVIEW

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The role of platelet P2Y₁₂ receptors in inflammation

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Abstract

Inflammation is a complex pathophysiological process underlying many clinical conditions. Platelets contribute to the thrombo-inflammatory response. Platelet $P2Y_{12}$ receptors amplify platelet activation, potentiating platelet aggregation, degranulation and shape change. The contents of platelet alpha granules, in particular, act directly on leucocytes, including mediating platelet–leucocyte aggregation and activation via platelet P-selectin. Much evidence for the role of platelet $P2Y_{12}$ receptors in inflammation comes from studies using antagonists of these receptors, such as the thienopyridines clopidogrel and prasugrel, and the cyclopentyltriazolopyrimidine ticagrelor, in animal and human experimental models. These suggest that antagonism of $P2Y_{12}$ receptors decreases markers of inflammation with some evidence that this reduces incidence of adverse clinical sequelae during inflammatory conditions. Interpretation is complicated by pleiotropic effects such as those of the thienopyridines on circulating leucocyte numbers and of ticagrelor on adenosine reuptake. The available evidence suggests that $P2Y_{12}$ receptors are prominent mediators of inflammation and $P2Y_{12}$ receptor antagonism as a potentially powerful strategy in a broad range of inflammatory conditions.

KEYWORDS

inflammation, leucocytes, P2Y₁₂ receptors, platelets

1 | INTRODUCTION

Inflammation is a double-edged entity, important in the repair of tissue damage and fighting infection, but harmful when excessively or inappropriately stimulated. Inflammation is central to a broad range of diseases, including cardiovascular, infective, autoimmune, neoplastic, degenerative and metabolic conditions (Furman et al., 2019). For example, at the core of atherosclerotic cardiovascular disease (ASCVD) is inappropriate inflammation in response to endothelial injury and infiltration of the vessel wall by low-density and very-low-density lipoproteins, leading to

Abbreviations: ACS, acute coronary syndrome; ASCVD, atherosclerotic cardiovascular disease; CCS, chronic coronary syndrome; CLP, caecal ligation and puncture; COVID-19, coronavirus disease 2019; GPIIb/IIIa, glycoprotein IIb/IIIa; MACEs, major adverse cardiovascular events; MI, myocardial infarction; PCI, percutaneous coronary intervention; RCTs, randomised controlled trials; VASP, vasodilator-associated phosphoprotein.

plaque formation (Libby et al., 2019). In particular, transmigration of blood monocytes to become vessel wall macrophages that unsuccessfully attempt to clear lipid-rich debris is a key feature of atherogenesis. Furthermore, reducing inflammation in patients with ASCVD can reduce the risk of major adverse cardiovascular events (MACEs) (Ridker et al., 2017; Tardif et al., 2019). Likewise, though, in infections, inflammation can be an appropriate and useful host response, overstimulation of inflammation is responsible for severe consequences such as septic shock (Delano & Ward, 2016).

Platelets are small anucleate cells circulating in very high numbers, derived from megakaryocytes (Chang et al., 2007). Their primary physiological role is in haemostasis, but they can become inappropriately activated, for example, upon atherosclerotic plaque rupture or erosion, leading to thrombosis (Parker & Storey, 2018). Upon activation, platelets undergo shape change, aggregate and release further

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HARMACOLOGICA

prothrombotic mediators that propagate the thrombotic response. However, platelets also play a significant role in inflammation, including in settings such as atherogenesis, acute thrombosis or sepsis (Giustozzi et al., 2021).

The platelet $P2Y_{12}$ receptor is key to the amplification of platelet activation response and is central in governing interactions between platelets and other inflammatory cells (Storey et al., 2000). In this review, we discuss the biology and pharmacology of the $P2Y_{12}$ receptor before focussing on its role in inflammation and how $P2Y_{12}$ receptor antagonists (' $P2Y_{12}$ inhibitors') can influence the inflammatory response. Though we focus on studies from the cardiovascular domain, this reflects the current clinical utility of $P2Y_{12}$ inhibitors. Other articles have considered this topic (Mansour et al., 2020; Thomas & Storey, 2015), but this fresh summary takes advantage of more recent findings from, for example, mechanistic studies of $P2Y_{12}$ inhibitors and clinical trial findings from the coronavirus disease 2019 (COVID-19) pandemic.

2 | PLATELET P2Y₁₂ RECEPTOR STRUCTURE AND FUNCTION

In humans, P2Y₁₂ receptors are encoded by the P2RY12 gene (rs2046934), located on chromosome 3 (3q25.1), containing 47 kb with a product of 342 amino acids (Rudež et al., 2008). Molecular identification of P2Y₁₂ receptors was achieved in 2001 (Hollopeter et al., 2001). The greatest expression of these receptors occurs in platelets and areas of the brain (mostly midbrain) but has also been detected in vascular smooth muscle cells, lung and leucocytes (Gachet, 2012).

The endogenous ligand of the $P2Y_{12}$ receptor is ADP (Zhang et al., 2001). Although ADP is found in all cell types containing mitochondria and at low levels in the plasma, it is present at high concentrations in the dense granules in platelets and is released upon platelet activation (Storey, 2006).

The P2Y₁₂ receptor is a seven-transmembrane domain G proteincoupled receptor, linked to G_i, which is activated upon ADP binding, mediated by conformational change (Zhang et al., 2014). Though P2Y₁₂ does not play a major role in the initiation of platelet activation or aggregation, it plays a central role in the amplification of these processes (Storey et al., 2000). The α subunit of G_i mediates inhibition of the formation of cAMP by adenylate cyclase, reducing the activity of PKA (Knighton et al., 1991). In platelets, PKA is responsible for the phosphorylation of vasodilator-associated phosphoprotein (VASP) (Butt et al., 1994). Unphosphorylated VASP stimulates the externalisation of the glycoprotein IIb/IIIa (GPIIb/IIIa) complex, mediating the final common pathway of platelet aggregation (Horstrup et al., 1994). Activation of P2Y₁₂ receptors therefore increases the availability of unphosphorylated VASP, promoting platelet aggregation. The β - γ subunit of P2Y₁₂-coupled G_i further contributes to GPIIb/IIIa activation by stimulating PI3K activity, in turn increasing activity of PKB (Akt) (Cosemans et al., 2006). Importantly, PI3K also mediates platelet degranulation (Borst et al., 2012). This occurs via a number of downstream pathways, shown in Figure 1, leading to changes in actin dynamics, in turn resulting in migration and fusion of granules with the cell surface membrane and ejection of their contents (Aloui et al., 2014; Stegner & Nieswandt, 2011).

3 | ROLE OF P2Y₁₂-MEDIATED PLATELET ACTIVATION IN INFLAMMATION

Platelet granules released upon activation include those of the α and dense types, as well as lysosomes (Heijnen & van der Sluijs, 2015). Each of these types contributes to the proinflammatory effects of P2Y₁₂ receptor activation. The alpha granules contain high levels of platelet P-selectin (CD62P), which is externally expressed upon fusion of the granule with the cell surface membrane (Harrison & Cramer, 1993). P-selectin binds to P-selectin glycoprotein ligand-1 (PSGL-1) on leucocytes, leading to formation of platelet-leucocyte aggregates. Specific actions depend on the leucocyte type. Platelet P-selectin interacting with monocyte PSGL-1, for example, promotes release of proinflammatory cytokines such as TNF- α , IL-1 β , IL-8 and the chemokine CCL2, via activation of the transcription factor NFkB pathway (Weyrich et al., 1995), additionally promoting surface expression of adhesion molecules (Martins et al., 2006). These cytokines induce the liver to release higher levels of C-reactive protein (CRP) (Yeh, 2004). Platelet-neutrophil aggregation promotes release of neutrophil extracellular traps (NETosis), which physically interact with pathogens and also contribute to many pathological inflammatory conditions, including atherosclerosis, thrombosis and sepsis (Etulain et al., 2015; Mozzini et al., 2017; O'Brien et al., 2017). Externalisation of GPIIb/IIIa induced by platelet activation also contributes to platelet-leucocyte aggregation through binding to the integrin CD11b, via bridging with fibrinogen (Schwarz et al., 2002).

As well as platelet P-selectin, the alpha granules contain a variety of soluble proinflammatory factors, released into the plasma upon fusion with the cell surface membrane. The range and effects of these are summarised in Table 1. These may directly modulate the activity of inflammatory cells or stimulate acellular coagulation. Stimulating the coagulation cascade results in an increase in the generation of factors such as thrombin or activated protein C that can activate specific cell receptors on mononuclear or endothelial cells and may affect, for example, cytokine production or inflammatory cell apoptosis (Levi et al., 2004). P2Y₁₂ receptor-mediated platelet activation also contributes further to thrombin generation via increasing scramblase activity leading to externalisation of phosphatidylserine on the platelet cell membrane, stabilising the prothrombinase complex (Bevers et al., 1982).

Dense granules contain high concentrations of ADP, ATP and 5-HT (McNicol & Israels, 1999). Release of dense granules into the extracellular space leads to autocrine and paracrine positive feedback on platelet activation. As well acting on $P2Y_{12}$ receptors, ADP stimulates platelet $P2Y_1$ receptors, which in contrast to $P2Y_{12}$ receptors play a significant role in the initiation of platelet activation, including shape change (Hechler et al., 1998). Stimulation of $P2Y_{12}$ receptors, however, does not lead to sustained platelet activation. As well as effects on platelets, extracellular ADP contributes to inflammation through mechanisms such as activation of other members of the **P2Y receptor family** found on leucocytes and

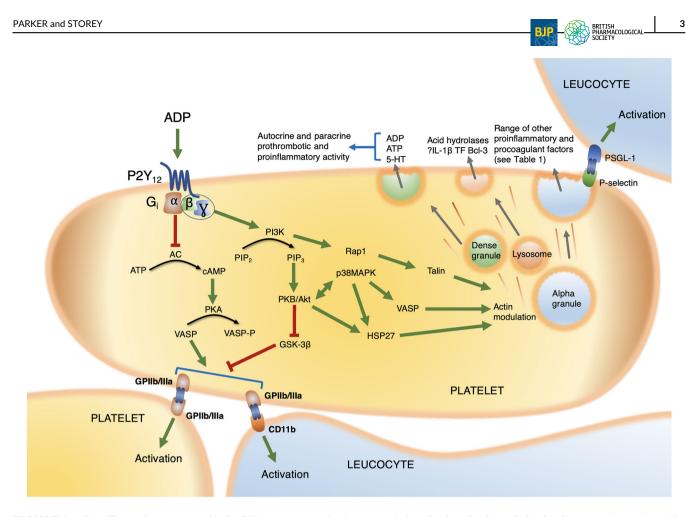


FIGURE 1 Signalling pathways upon platelet P2Y₁₂ receptor activation, potentiating platelet–platelet and platelet–leucocyte interaction. AC, adenylate cyclase; CD11b, cluster of differentiation 11b; GPIIb/IIIa, glycoprotein Ilb/IIIa; GSK-3β, glycogen synthase kinase-3β; HSP27, heat shock protein 27; MAPK, mitogen-activated protein kinase; PIP, phosphatidylinositol bisphosphate; PKA, protein kinase A; PKB, protein kinase B; PSGL-1, P-selectin glycoprotein ligand 1; TF, tissue factor; VASP, vasodilator-associated phosphoprotein; VASP-P, phosphorylated vasodilator-associated phosphoprotein.

endothelium, and promotion of fibroblast proliferation (Borges et al., 2021). ATP is also an agonist at platelet P2X₁ receptors, contributing to increased cytosolic calcium and resultant shape change and aggregation response, particularly in the presence of high shear stress (Mahaut-Smith et al., 2011). Extracellular ATP can also drive inflammasomal activation in monocytes (Cauwels et al., 2014). 5-HT contributes to platelet activation via the G_q -coupled 5-HT_{2A} receptor and to a wide range of other inflammatory processes in other cell types (Wu et al., 2019).

Platelet lysosomes contain high concentrations of acid hydrolases, able to break down glycoproteins, glycolipids and glycosaminoglycans (Ciferri et al., 2000). Their role, once released into the extracellular space during platelet activation, has not been fully elucidated but appears to include thrombus remodelling and receptor cleavage. It has also been hypothesised that platelet lysosomal release may facilitate release of the proinflammatory mediators, IL-1 β , tissue factor and Bcl-3 (Heijnen & van der Sluijs, 2015).

As well as degranulation, activated platelets release extracellular vesicles (EVs) (van der Pol et al., 2012). Their cargo includes not only procoagulant factors but also a broad range of inflammatory modulators, and it is now appreciated that EVs can induce inflammatory

changes in endothelium and leukocytes (Vajen et al., 2015). As EVs are able to cross lipid membranes, they can therefore additionally influence the actions of extravascular cell types (Puhm et al., 2021). Clinical studies of inhibitors suggest that $P2Y_{12}$ receptor signalling plays a prominent role in EV release (Gasecka et al., 2020).

4 | CLINICALLY USED PLATELET P2Y₁₂ INHIBITORS

Antagonists of platelet P2Y₁₂ receptors are widely used in clinical practice for the treatment and secondary prevention of ASCVDs, including coronary artery, cerebrovascular and peripheral artery diseases. **Ticlopidine, clopidogrel** and **prasugrel** are orally administered thienopyridine prodrugs whose active metabolites bind selectively and irreversibly to P2Y₁₂ receptors (Abbracchio et al., 2019; Hollopeter et al., 2001). Though ticlopidine is generally no longer available, clopidogrel is still extensively prescribed. Clopidogrel has a relatively slow onset time (around 2 h) when compared to newer agents, and offset of significant antiplatelet effect takes around 5 days (Gurbel et al., 2009). Furthermore, a significant proportion

PARKER and STOREY

TABLE 1 Effects of key factors contained within platelet alpha granules.

Factor	Effects		
β-Thromboglobulin (CXCL7) (Majumdar et al., 1991)	emoattraction of neutrophils and fibroblasts, maturation of megakaryocytes, increased tissue plasminogen ctivator synthesis by fibroblasts		
CCL5 (Cognasse et al., 2022)	Interacts with P-selectin to mediate monocyte tissue migration, effector of memory T cells		
CD40 ligand (Elgueta et al., 2009)	Binds to CD40 on B cells/dendritic cells, acts as helper T-cell signal, induces germinal centre formation, isotype class switching and antibody production		
Defensins Thrombocidins Kinocidins (Leeten et al., 2021)	Antimicrobial activity		
Factor V (Harrison & Cramer, 1993)	When activated, forms a component of the prothrombinase complex		
Fibrinogen (Blair & Flaumenhaft, 2009)	Facilitates platelet-platelet adhesion, converted to fibrin by thrombin		
Fibronectin (Zucker et al., 1979)	Mediates thrombus formation and cell adhesion		
Insulin-like growth factor 1 (Hers, 2007)	Stimulates PKB/Akt in a wide range of cell types leading to cell growth and inhibition of apoptosis, as well as potentiating platelet activation		
Interleukin-8 (CXCL8) (Chen et al., 2020)	Neutrophil chemotaxis, angiogenesis		
P-selectin (CD62P) (Vandendries et al., 2004)	Binds to PSGL-1 to facilitate platelet-leucocyte aggregation and activation		
Platelet factor 4 (CXCL4) (Nevzorova et al., 2019)	Neutralises heparin-like molecules, therefore reducing anti-thrombin III activity, contributes to platelet activation, induces platelet death, promotes chemotaxis of leucocytes and fibroblasts		
Platelet-derived growth factors (Kardas et al., 2020)	Proliferation of smooth muscle cells, fibroblasts and glial cells		
Thrombospondin (Isenberg et al., 2008)Stimulates platelet aggregation by blocking the antiplatelet effect of nitric oxide, blocks growth induces apoptosis of endothelial cells			
TNF-α (Pokrovskaya et al., <mark>2020)</mark>	Stimulation of inflammatory processes, primarily through the NF- κB and MAPK pathways		
TGF- β (Grainger et al., 1995)	Wide range of effects, including on inflammation such as stimulation of resting macrophages, inhibition of B cells and differentiation of T cells		
VEGF (Battinelli et al., 2011)	Promotes angiogenesis		
Von Willebrand factor (Blair & Flaumenhaft, 2009)	Facilitates platelet-platelet and platelet-endothelium adhesion, stabilises factor VIII		

Abbreviations: MAPK, mitogen-activated protein kinase; PKB, protein kinase B; PSGL-1, P-selectin glycoprotein ligand 1.

of the population (approximately one third) are 'resistant' to its effect, partly related to impairment of its enzymatic conversion to the active metabolite by the cytochrome P450, CYP 2C19 (Matetzky et al., 2004). Prasugrel is a newer thienopyridine with the advantage of reduced variability in levels of platelet inhibition and shorter onset time (30 min) than clopidogrel, but a longer off-set time of around 7 days (Dobesh, 2009). Though still a prodrug, prasugrel's metabolism is not CYP2C19-dependent and does not vary significantly between individuals, there being no evidence of a resistant human phenotype despite some inter-individual variability in response to licensed maintenance doses (Jakubowski et al., 2007).

Reversibly binding, orally administered P2Y₁₂ receptor antagonists are also available. **Ticagrelor** is an orally active cyclopentyltriazolopyrimidine with an onset time of around 30 min in stable patients and offset of action between 2 and 5 days (Gurbel et al., 2009). Unlike thienopyridines, ticagrelor is not a prodrug and so is active on P2Y₁₂ receptors as soon as it is absorbed. Ticagrelor additionally weakly inhibits **equilibrative nucleoside transporter 1 (ENT1)**, responsible for **adenosine** uptake by erythrocytes and platelets (Armstrong et al., 2014; Aungraheeta et al., 2016).

Cangrelor and selatogrel are reversibly binding, parenterally administered P2Y₁₂ receptor antagonists with rapid onset (around 3 and 15 min, respectively), quick offset (recovery of pre-dose levels of platelet inhibition by 60–120 min and 24 h after discontinuation, respectively) and high reliability of effect (Parker et al., 2017; Parker & Storey, 2020). Like ticagrelor, they are not prodrugs. Cangrelor is an ATP analogue that is administered intravenously, and selatogrel is an analogue of 2-phenyl-pyrimidine-4-carboxamide that is administered subcutaneously. The main metabolite of cangrelor weakly inhibits adenosine reuptake, but to a lesser degree than ticagrelor (Armstrong et al., 2014). Neither selatogrel nor its metabolites have any known effects on adenosine kinetics. Selatogrel is not yet licensed nor commercially available but is undergoing a phase III trial of prehospital administration in patients with suspected acute myocardial infarction (MI) (NCT04957719).

5 | EFFECTS OF P2Y₁₂ RECPTOR ANTAGONISTS ON INFLAMMATION

5.1 | Pharmacodynamic studies

As P2Y₁₂ receptors are central to the amplification of platelet activation, inhibition of the receptor in vitro leads to profound reduction in platelet reactivity in response to a range of agonists (Figure 2). This effect is additive to that of **aspirin** whether tested in vitro or in human subjects (Hennigan et al., 2022). In contrast to aspirin, however, P2Y₁₂ receptor antagonists and/or their active metabolites have a significant effect in reducing platelet P-selectin expression both in the unstimulated blood samples and when stimulated with agonists such as ADP (Frelinger et al., 2007; Judge et al., 2008; Storey et al., 2002). This leads to a significant reduction in platelet-monocyte and platelet-neutrophil conjugate formation. Increasing the potency and reliability of the antagonism of P2Y₁₂ receptors with a newer agent increases the strength of effect on platelet P-selectin expression when compared to clopidogrel (Parker, Gorog, et al., 2020).

Numerous studies have measured circulating or expressed levels of α -granule-derived factors in participants receiving oral P2Y₁₂ receptor antagonists. Levels of P-selectin expression or solubilisation

and expression of CD40L have been demonstrated to be lowered by, for example, loading with clopidogrel in patients with acute coronary syndrome (ACS) and/or undergoing percutaneous coronary intervention (PCI) (Vivekananthan et al., 2004; Xiao & Théroux, 2004). Another study found lower levels of soluble CD40L (sCD40L) and the chemokine CCL5 (RANTES) in patients with chronic coronary syndrome (CCS) receiving clopidogrel and aspirin when compared to aspirin alone (Heitzer et al., 2006). However, another study failed to demonstrate a significant effect of clopidogrel alone, when compared to aspirin alone, on levels of soluble P-selectin, sCD40L, transforming growth factor- β or MCP-1 in a similar population (Solheim et al., 2006). Patients receiving prasugrel had greater inhibition of CD40L and/or P-selectin expression than when receiving clopidogrel in settings such as CCS and type 2 diabetes (Braun et al., 2008; Parker, Gorog, et al., 2020). When compared to clopidogrel, ticagrelor reduced the release of platelet- and leucocyte-derived EVs, including those expressing P-selectin, during treatment for acute myocardial infarction (Gasecka et al., 2020).

Studies have also measured more general markers of inflammation in patients with coronary syndromes receiving antagonists of $P2Y_{12}$ receptors, in particular CRP. Some studies have shown that clopidogrel reduces CRP, either when assessed at baseline versus on-

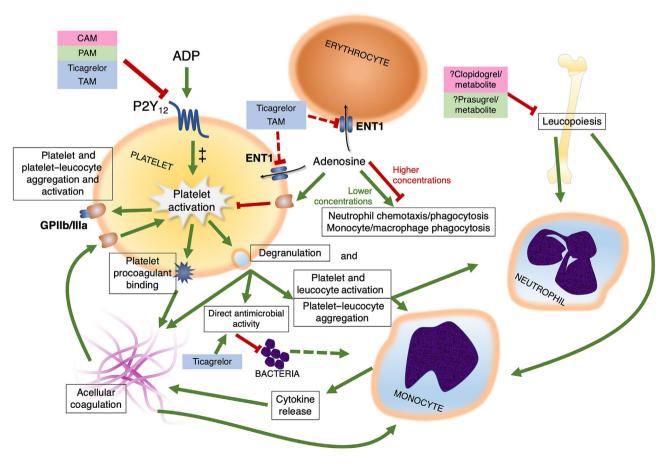


FIGURE 2 On-target and potential off-target effects of commonly used oral platelet P2Y₁₂ receptor antagonists on inflammation and related processes. CAM, clopidogrel active metabolite; ENT1, equilibrative nucleoside transporter 1; GPIIb/IIIa, glycoprotein IIb/IIIa; PAM, prasugrel active metabolite; TAM, ticagrelor active metabolite.

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treatment or in comparison with aspirin, including those with ACS, CCS and/or undergoing PCI (Chen et al., 2006; Heitzer et al., 2006; Vivekananthan et al., 2004). However, other studies have failed to show a significant effect (Azar et al., 2006; Hajsadeghi et al., 2016; Solheim et al., 2006; Woodward et al., 2004). There is also evidence that TNF- α is significantly reduced by clopidogrel treatment (Chen et al., 2006) and that the effect may be dose-dependent (Gurbel et al., 2006).

In a study of 120 patients undergoing PCI, prasugrel was associated with a greater reduction in high-sensitivity CRP (hs-CRP) compared to clopidogrel, but in another study, levels of hs-CRP were similar between the two drugs, although a lower-than-standard dose of prasugrel was used (Hajsadeghi et al., 2016; Masuyama et al., 2021).

The effect of ticagrelor on CRP has been assessed in comparison to clopidogrel. Studies have shown that in patients with coronary syndromes, ticagrelor treatment is associated with a higher CRP or hs-CRP, accompanied by higher levels of other inflammatory markers such as **IL-6** or endocan, compared to when receiving clopidogrel (Storey et al., 2014; Wei et al., 2017); though another study showed no significant difference, there were trends supporting this effect (Husted et al., 2010). Potential reasons for higher inflammatory markers when receiving ticagrelor than clopidogrel despite more potent antagonism of P2Y₁₂ receptors are discussed below.

During primary PCI for myocardial infarction with ST-elevation, in a small study, Abo-Aly et al., (2021) showed that adding cangrelor to aspirin and ticagrelor induced greater inhibition of platelets and reduced circulating levels of proinflammatory factors such as sCD40L and CCL5.

Data concerning effects of $P2Y_{12}$ receptor antagonists on inflammatory markers during other conditions are also available. In a trial of ticagrelor in patients with pneumonia, when compared to placebo, ticagrelor inhibited platelet-leucocyte aggregation and was associated with a significant reduction in circulating IL-6 levels and a shorter requirement for supplemental oxygen (Sexton et al., 2018). There were no significant effects on other inflammatory markers, though, in contrast to IL-6, these were not elevated above the reference range.

5.2 | Effects during in vivo experimental inflammation

Important data on the effects of P2Y₁₂ receptor antagonists on acute inflammation have also been obtained using in vivo experimental models. These allow controlled and comprehensive study of the inflammatory response. In particular, the use of experimental endotoxaemia has greatly informed our understanding of the interaction between platelet P2Y₁₂ receptors and inflammation (Suffredini & Noveck, 2014). Endotoxin stimulates monocyte activation via **TLR4**. As well as the obvious relevance to gram-negative sepsis, TLR4 acts as a damage-associated molecular pattern receptor, and so this pathway is applicable to other forms of tissue damage, including atherogenesis and atherothrombosis. In mouse or rat models of endotoxaemia, antagonism of $P2Y_{12}$ receptors with clopidogrel reduced detectable levels of the proinflammatory cytokines IL-6 and TNF- α , associated with less severe liver and lung injury (Hagiwara et al., 2011; Winning et al., 2009), though this effect was not seen in a pig model (Lipcsey et al., 2005). In another study, prasugrel significantly inhibited the release of proinflammatory cytokines during murine endotoxaemia (Totani et al., 2012).

Experimental human endotoxaemia has also been used to study the effects of antagonists of $P2Y_{12}$ receptors on acute inflammation, summarised in Table 2. Taken together, these suggest that antagonism of platelet $P2Y_{12}$ receptors leads to reduced acute inflammatory response when stimulated via TLR4 but that the ability of $P2Y_{12}$ receptor antagonists to reduce acute inflammation may be affected by concurrent treatment with aspirin, as is common in many patients with ASCVD. This is being studied further in the WILLOW TREE trial (NCT03869268), expected to report its results this year.

The murine caecal ligation and puncture (CLP) model of intrabdominal polymicrobial sepsis has likewise been used to study the role of P2Y₁₂ receptors in inflammation. Clopidogrel in wild-type (WT) mice mimicked the effect of P2Y12 knockout (KO) in preventing CLP-induced increases in P-selectin expression and platelet-leucocyte aggregation, which was associated with protection from acute lung and liver injury (Liverani et al., 2016). Levels of TNF- α and IL-6 after CLP were significantly lower in P2Y12-KO, than in WT mice. Notably, however, clopidogrel significantly reduced leucocyte counts after CLP in both WT and KO mice, suggesting a $P2Y_{12}$ -independent mechanism. On the other hand, another study of the murine CLP model failed to show benefits of clopidogrel treatment or deletion of platelet-specific P2Y₁₂ receptors (Rabouel et al., 2021). Potential explanations for the discrepancy in findings between the two groups include that the models were not necessarily equivalent given some differences in circulating cell count responses to sepsis. The effect of clopidogrel has also been studied in murine models of chest sepsis caused by Streptococcus pneumoniae or Klebsiella pneumoniae, with no significant influence seen on host response to infection (Claushuis et al., 2019; van den Boogaard et al., 2015).

Murine CLP has also been used to study the effects of ticagrelor (Rahman et al., 2013). During polymicrobial sepsis, ticagrelor significantly inhibited the formation of platelet-neutrophil aggregates and acute lung injury, when assessed by histological scoring, bronchoalveolar fluid neutrophil count or weight of lung oedema. Ticagrelor had no significant effect on neutrophil CD11b expression or formation of lung chemokines.

Experimental models of infective endocarditis have similarly been utilised. In a rat model of *Enterococcus faecalis* or *Streptococcus gallolyticus* infective endocarditis, neither aspirin nor ticlopidine when given alone influenced vegetation weight, but combining the two drugs significantly reduced the chance of developing endocarditis and infective burden when it did develop (Veloso et al., 2015). A similar finding was made in a rabbit model (Nicolau et al., 1999). Whether the observed effect is due to effects on thrombosis, inflammation, infection or a combination of these remains unclear. It has been hypothesised that in *Staphylococcus aureus* bacteraemia, for example, potent antagonism of P2Y₁₂ receptors may prevent infection-induced platelet death and TABLE 2 Studies of P2Y₁₂ receptor antagonists during human experimental endotoxaemia.

Paper	Sample population	Endotoxin regimen	Treatment allocation	Key findings
Spiel et al. (2012)	20 healthy males	2-ng·kg ⁻¹ IV bolus	60-mg loading dose of prasugrel or placebo 2 h before endotoxin in a crossover design	Prasugrel inhibited platelet activation responses during endotoxaemia. Release of vWF during endotoxaemia appeared to antagonise the ability of prasugrel to inhibit platelet plug formation in high shear stress conditions
Thomas et al. (2015)	30 healthy males	2-ng·kg ⁻¹ IV bolus	No drug (n = 10), clopidogrel 75 mg OD (n = 10) or ticagrelor 90 mg BD (n = 10) for 1 week before endotoxin in a parallel-group design	During endotoxaemia, both clopidogrel and ticagrelor significantly reduced peak levels of TNF- α and IL-6. Ticagrelor additionally significantly inhibited CCL2 (MCP-1) and G-CSF and increased peak levels of IL-8. Both drugs reduced platelet P-selectin expression, PMA and PNA formation. Reduced levels of inflammation when receiving a P2Y ₁₂ receptor antagonist were associated with less prothrombotic change in fibrin clot dynamics
Kiers et al. (2017)	40 healthy males	1-ng·kg ⁻¹ IV bolus, then 1-ng·kg ⁻¹ IV infusion over 1 h	Ticagrelor (180-mg loading dose then 90 mg BD) plus aspirin (160-mg loading dose then 80 mg OD) (n = 10); clopidogrel (300-mg loading dose then 75 mg OD) plus aspirin (n = 10); aspirin alone (n = 10) or placebo (n = 10) for 7 days prior to endotoxin in a parallel-group design	Neither ticagrelor nor clopidogrel (both given alongside aspirin) significantly affected cytokine release during endotoxaemia. Aspirin, when given alone, increased release of TNF- α when compared to no drug or aspirin plus ticagrelor. Mechanistic studies suggested that the proinflammatory effect of aspirin was due to inhibition of platelet PGE ₂ release

Abbreviations: BD, twice daily; G-CSF, granulocyte colony-stimulating factor; IV, intravenous; OD, once daily; PMA, platelet–monocyte aggregate; PNA, platelet–neutrophil aggregate; vWF, von Willebrand factor.

therefore preserve the antimicrobial actions of platelets provided by release of factors such as defensins (Leeten et al., 2021), though given that this is α -degranulation mediated, it is not clear what would be the optimal balance of inhibition and permittance of α -degranulation.

Myocardial infarction is also an inflammatory stimulus, and the effects of $P2Y_{12}$ inhibitors on inflammatory processes in animal models of myocardial infarction have been studied. In a murine model of myocardial infarction, antagonism of $P2Y_{12}$ receptors with prasugrel reduced proinflammatory emergency haematopoiesis, but this appeared to be mediated by a non-platelet mechanism (Seung et al., 2022). P2Y₁₂ KO also reduced myocardial platelet and neutrophil accumulation during reperfusion after experimental myocardial infarction (von Elverfeldt et al., 2014), a similar effect being seen when receiving the P2Y₁₂ receptor antagonist prasugrel, in a murine model of autoimmune myocarditis (Schmidt et al., 2021). Neutrophil CD11b was reduced by ticagrelor in a murine model of myocardial infarction and reperfusion (Mauler et al., 2019). Though not yet entirely clear and excluding non-platelet-dependent mechanisms, the effects of P2Y₁₂ receptor antagonists in these settings are probably a

combination of anti-inflammatory action limiting tissue damage and antithrombotic actions reducing microthrombosis with resulting reductions in ongoing ischaemia or progressive infarction. Effects of ticagrelor on ischaemia-reperfusion injury have been linked to its pleiotropic effect on adenosine reuptake, which is discussed later.

5.3 | Insights from studies of clinical outcomes

It is well established from large randomised clinical trials that adding an oral antagonist of $P2Y_{12}$ receptors to aspirin treatment reduces the risk of MACE in ACS and high-risk CCS (Parker & Storey, 2016a). Similarly, $P2Y_{12}$ receptor antagonists are used to reduce MACE in cerebrovascular and peripheral artery disease (Parker, Schulte, et al., 2020). It seems clear therefore that $P2Y_{12}$ receptor antagonists can reduce the frequency and/or the effects of atherothrombosis. Atherothrombosis is a thrombo-inflammatory phenomenon, and though most focus has been given to effects on the thrombotic component, it is plausible that anti-inflammatory effects also contribute.

BRITISH PHARMACOLOGICAL

Large randomised controlled trials (RCTs) of clopidogrel and placebo or prasugrel and clopidogrel in ACS showed no significant difference in all-cause or non-cardiovascular mortality between treatments (Chen et al., 2005; CURE Trial Investigators, 2001; Wiviott et al., 2007). However, in the PLATelet inhibition and patient Outcomes (PLATO) trial, which studied ticagrelor and clopidogrel in patients with ACS, all-cause mortality was significantly lower in the group receiving ticagrelor, with numerically lower non-cardiovascular death (Wallentin et al., 2009). This was contributed to by significantly fewer fatal pulmonary infections, and there was also a reduction in the number of non-fatal infections (Storey et al., 2014).

Observational studies have suggested that treatment with clopidogrel may increase the risk of developing community-acquired pneumonia but conversely may reduce the severity of conditions such as pneumonia or *S. aureus* bacteraemia when these do occur (Caffrey et al., 2022; Gross et al., 2013; Winning et al., 2009, 2010). Another study in patients with sepsis requiring intensive care unit admission suggested that there may be a benefit of clopidogrel when given alone but not in combination with aspirin (Otto et al., 2013). Observational data have also shown a significantly reduced risk of gram-positive infections in ACS patients receiving ticagrelor compared to clopidogrel (Lupu et al., 2020).

Few RCTs have studied the effect of P2Y₁₂ receptor antagonists on inflammation in non-cardiovascular conditions but nevertheless have demonstrated some potentially promising findings. The Prasugrel IN Asthma (PRINA) trial randomised 26 patients with chronic asthma to receive prasugrel or placebo in a crossover study. Using mannitol challenge to assess bronchial responsiveness, prasugrel significantly reduced bronchial hyperreactivity, though it had no influence on exhaled levels of **NO** (Lussana et al., 2015). More recently, an RCT of ticagrelor or placebo in 60 patients with pneumonia found that, compared to placebo, ticagrelor reduced levels of IL-6 and plateletleucocyte aggregates (Sexton et al., 2018). Promisingly, there was also some evidence of modest beneficial effects on lung function.

Most recently, $P2Y_{12}$ receptor antagonists have been trialled in the setting of COVID-19. COVID-19 infection can lead to an intense thrombo-inflammatory response, and detectable markers of platelet activation, including those of α -degranulation, can be profoundly raised (Gąsecka et al., 2021; Gorog et al., 2022; Parker & Storey, 2021b). Despite this, trials of P2Y₁₂ receptor antagonists in critically or non-critically ill hospitalised patients with COVID-19 have been disappointing, showing no significant improvements in outcomes such as the requirement for organ support (Berger et al., 2022; Bradbury et al., 2022).

5.4 | Limitations in the interpretation of evidence from clinical studies

Several factors may complicate the interpretation of data from observational or interventional clinical studies when considering how these demonstrate the role of the platelet P2Y₁₂ receptors in inflammation.

In studies of $P2Y_{12}$ receptor antagonists in settings such as ACS or PCI, it may be difficult to disentangle direct effects on inflammation from indirect effects on the burden of proinflammatory infarction, which is known to be less with increasing potency of $P2Y_{12}$ receptor antagonism (Khan et al., 2016), though peri-procedural myocardial infarction was not reduced by increasing potency, after PCI for CCS (Orme et al., 2018). Conceivably, even studies in conditions such as pneumonia may be affected by differential effects on type 1 and type 2 myocardial infarctions that often accompany severe infection, though study in this area is very much lacking (DeFilippis et al., 2019).

Observational studies may be particularly difficult to interpret given that patients receiving or not receiving $P2Y_{12}$ receptor antagonists as part of their usual clinical care will inherently have differing risk profiles, which, despite statistical techniques, are challenging to completely adjust for. Similarly, the choice of $P2Y_{12}$ receptor antagonist is likely be influenced by clinical characteristics. This underlines the importance of obtaining robust data from RCTs.

As well as platelet P2Y₁₂ receptors, non-platelet P2Y₁₂ receptors may also play a role in inflammation that may contribute to the effects seen in studies of P2Y₁₂ receptor antagonists. Dendritic cell P2Y₁₂ receptors plays a role in endocytosis and **IL-12** production that may affect T-cell responses (Schnurr et al., 2005). P2Y₁₂ receptors on vascular smooth muscle cells are involved in expression of proinflammatory factors regulating monocyte chemotaxis and have a role in inflammatory processes such as early atherogenesis and allograft vasculopathy (Harada et al., 2011; Satonaka et al., 2015; West et al., 2014). Likewise, monocyte P2Y₁₂ receptors may play a role in modulation of function (Micklewright et al., 2018). P2Y₁₂ receptors of undetermined location also mediates **leukotriene E₄-induced** pulmonary inflammation (Paruchuri et al., 2009).

Furthermore, as well as antagonism of P2Y₁₂ receptors, pleiotropic effects of drugs may play a role in determining the effects on inflammation and infection seen during clinical studies. The thienopyridines have long been associated with cytopaenias, in particular leucopaenia. In the case of ticlopidine, this may occur to a clinically significant extent in around 5% of patients during prolonged administration and has led to its near abandonment (Janzon, 1996). Though clinically meaningful leucopaenia occurs much less commonly with clopidogrel, there is evidence from animal and human studies that clopidogrel still significantly reduces leucocyte count (Liverani et al., 2016; Nelson et al., 2022; Storey et al., 2014). Effects of prasugrel and other P2Y₁₂ receptor antagonists on leucocyte count have not been well studied, though in vitro studies have suggested that prasugrel metabolites may inhibit neutrophil function via a non-P2Y₁₂ receptor-mediated pathway (Liverani et al., 2012).

There is no evidence that ticagrelor reduces leucocyte count during maintenance therapy (Storey et al., 2014), but its additional property of weak inhibition of ENT1 may also contribute to observed effects on inflammation. Though early evidence suggested that ticagrelor significantly increased plasma levels of adenosine, multiple subsequent clinical studies attempting to demonstrate this effect during therapeutic administration have shown no significant difference, compared to clopidogrel, no drug or during offset of ticagrelor's antiplatelet effect (Bonello et al., 2014; Kiers et al., 2017; Orme et al., 2018; Ow et al., 2020). Therefore, although in vitro studies demonstrate that ticagrelor certainly inhibits adenosine reuptake by erythrocytes and platelets (Nylander et al., 2013; Parker & Storey, 2016b), the clinical significance of this effect at therapeutic plasma concentrations of ticagrelor is not completely clear. As well as augmenting the antiplatelet effect of P2Y₁₂ receptor antagonists (Nylander et al., 2013), adenosine has dimorphic effects on neutrophil and monocyte function, which are potentiated by ticagrelor in vitro (Alsharif et al., 2015). At lower concentrations, adenosine primarily stimulates high-affinity A1 receptors, with downstream effects that are more proinflammatory than anti-inflammatory, including promotion of neutrophil chemotaxis and phagocytosis and macrophage phagocytosis (Barletta et al., 2012). At higher concentrations, adenosine primarily acts on low-affinity A2A and A2B receptors, with downstream effects that are more anti-inflammatory than proinflammatory, including down-regulation of cytokine release (Haskó & Pacher, 2012). Ticagrelor-related increase in adenosine exposure has also been associated with other effects relevant to inflammation such as up-regulation of cyclooxygenase 2 and nitric oxide synthase in animal models (Nanhwan et al., 2014; Ye et al., 2015). Finally, there is some evidence that ticagrelor, but not thienopyridines, may have direct antimicrobial properties, including against organisms that are antibiotic-resistant (Lancellotti et al., 2019). This appears to be an effect particularly against gram-positive organisms. For example, in a murine model of S. aureus infective endocarditis, treatment with ticagrelor, but not clopidogrel, reduced vegetation formation compared to drug-free controls, which appeared to be related to effects on bacterial toxin production and endothelial adherence (Oury et al., 2023).

6 | FUTURE WORK AND SCOPE FOR CLINICAL UTILITY OF P2Y₁₂ RECEPTOR ANTAGONISTS IN INFLAMMATION

Given that antagonists of P2Y₁₂ receptors are already utilised for the treatment and prevention of ASCVD, the potential for optimising and expanding their use in this population is seemingly most feasible. For example, current evidence suggests that prolonged antagonism of P2Y₁₂ receptors in patients with high-risk CCS can offer net benefits in clinical outcomes (Parker & Storey, 2021a), though determining whether any anti-inflammatory and antithrombotic effects are contributory is difficult to assess. Whether P2Y₁₂ receptor antagonists find a significant role in non-ASCVD inflammatory conditions in addition to or instead of existing drug treatments remains to be seen. This will primarily depend on whether clinical benefits that outweigh any risks can be robustly demonstrated in appropriately powered RCTs. So far, the most powered RCTs to study hard clinical outcomes in patients receiving P2Y₁₂ receptor antagonists for a non-cardiovascular condition have been in the setting of COVID-19. Although trials have demonstrated that, in severe COVID-19, other forms of antiinflammatory therapy such as corticosteroids and monoclonal antibodies against the IL-6 receptor may significantly improve clinical



outcomes (Zeraatkar et al., 2022), those of P2Y₁₂ receptor antagonists have been disappointing, as we have discussed. Nevertheless, both animal and human studies have some conflicting results that may point to the benefit of P2Y12 receptor antagonism in inflammatory conditions being situation-dependent. Notably, in the setting of ASCVD, not all anti-inflammatory therapies appear to be equally efficacious; for example, colchicine and canakinumab reduced cardiovascular risk, but methotrexate had no significant effect (Ridker et al., 2017, 2019; Tardif et al., 2019). In the area of sepsis, there is still debate as to whether anti-inflammatory therapies, in general, can be beneficial due to the careful balance of preserving the physiological mechanisms of fighting infection whilst avoiding inappropriate overstimulation of inflammation (Nedeva et al., 2019), and clinical trials of any anti-inflammatory therapies in sepsis have overall been disappointing. Whether additional properties beyond antithrombotic and anti-inflammatory effects are relevant remains to be explored: Ticagrelor's apparent direct antimicrobial properties are certainly intriguing, but if and how they complement existing antibiotic pharmacotherapy is not yet clear. However, these properties may also hypothetically mitigate any loss of defence against infection caused by the anti-inflammatory effect of P2Y₁₂ receptor antagonism.

There are numerous other non-cardiovascular conditions in which the anti-inflammatory properties of P2Y₁₂ receptor antagonists may be hypothetically able to play a useful role, for example, chronic inflammatory diseases such as rheumatoid arthritis (RA). There is some limited evidence that treatment with ticagrelor may be of benefit in patients with methotrexate-resistant RA (Garshick et al., 2021), though it remains unclear if the effect seen was mediated via antagonism of P2Y₁₂ receptors or by potentiating effects of methotrexate on adenosine accumulation, and the effects of P2Y₁₂ receptor antagonists in animal models of RA are conflicting (Entsie et al., 2023). Though a range of other existing anti-inflammatory therapies exist for these conditions, a chronic inflammatory state is strongly linked to cardiovascular risk (Hansildaar et al., 2021), and it is plausible, but not yet determined, that the additional antiinflammatory and antithrombotic effects may contribute to potential long-term net benefits.

The main drawback of P2Y₁₂ receptor antagonists as antiinflammatory agents, similar to their use as antithrombotic drugs, is likely to be an increase in bleeding risk, though the magnitude of this risk when used outside of combination antithrombotic therapy appears to be similar to that of aspirin (CAPRIE Steering Committee, 1996; Johnston et al., 2016) and has to be weighed against potential benefits from the antithrombotic and antiinflammatory effects to determine net clinical benefit in the particular population. The availability of rapid reversal strategies for P2Y₁₂ receptor antagonists may help to address some concerns. For example, bentracimab, a novel monoclonal antibody fragment against ticagrelor and its active metabolite, may be one such strategy (Bhatt et al., 2019).

Mechanistically, the effect of inflammation, mediated by platelet $P2Y_{12}$ receptors, on leucocytes has focussed on monocytes and neutrophils, and more work is needed to determine if other cell types play

BRITISH PHARMACOLOGIC

a role. Mast cells, for example, can both activate and be activated by platelets (Boyce, 2007; Karhausen et al., 2020), though it has not been well determined if P2Y₁₂ receptors are involved in mediating these effects. Platelets also play a role in adaptive and innate immunity (Marcoux et al., 2021). Platelets can process and present antigens to CD8⁺ T cells via proteasomal activity, primarily medicated by major histocompatibility complex 1, which is contained within alpha granules (Zufferey et al., 2014). Factors such as CD40L also stimulate T- and B-cell function (Blumberg et al., 2009). Furthermore, platelets respond to antibodies, expressing a range of Fc receptors. Most notably, this includes the Fc receptor for immunoglobulin G IIA (FcyRIIA), which binds immune complexes and microorganisms and is known to play a role in a wide range of inflammatory, autoimmune and infective conditions (Marcoux et al., 2021). That P2Y12 receptors are directly involved in the function and regulation of these processes is not fully established, but, given that aspects of platelet activation such as degranulation and EV release appear important, its signalling would be predicted to enhance them. As discussed earlier, the further exploration of the role of non-platelet P2Y12 receptors in inflammation is additionally needed.

7 | CONCLUSIONS

The P2Y₁₂ receptor and its associated downstream pathways play a central role in the amplification of platelet activation, potentiating effects such as platelet aggregation, degranulation, shape change and platelet procoagulant activity. Degranulation in particular mediates proinflammatory effects, including the formation of platelet-leucocyte aggregates via platelet P-selectin binding to leucocyte PSGL-1. Studies of P2Y₁₂ receptor antagonists have shown strong inhibitory effects on degranulation and platelet–leucocyte aggregation. Interpreting data from clinical studies of P2Y₁₂ receptor antagonists is complicated by pleiotropic effects of thienopyridines and ticagrelor, but overall, these support a proinflammatory role of platelet let P2Y₁₂ receptors and suggest that inhibition of the receptor, alongside effects on thrombus formation, is potentially useful in a range of inflammatory conditions from atherosclerosis to sepsis and beyond.

7.1 | Nomenclature of targets and ligands

Key protein targets and ligands in this article are hyperlinked to corresponding entries in http://www.guidetopharmacology.org and are permanently archived in the Concise Guide to PHARMACOLOGY 2021/22 (Alexander, Christopoulos et al., 2021; Alexander, Fabbro et al., 2021a, b; Alexander, Kelly et al., 2021a, b).

AUTHOR CONTRIBUTIONS

William A. E. Parker: Writing—original draft (lead); writing—review and editing (equal). Robert F. Storey: Conceptualization (lead); writing—review and editing (equal).

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DATA AVAILABILITY STATEMENT

No original data are associated with this manuscript.

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REFERENCES

- Abbracchio, M.-P., Boeynaems, J.-M., Boyer, J. L., Burnstock, G., Ceruti, S., Fumagalli, M., Gachet, C., Hills, R., Humphries, R. G., Inoue, K., Jacobson, K. A., Kennedy, C., King, B. F., Lecca, D., Müller, C. E., Miras-Portugal, M. T., Ralevic, V., & Weisman, G. A. (2019). P2Y receptors: P2Y₁₂ receptors (version 2019.3). *IUPHAR/BPS Guide to Pharmacology*, 2021(3). https://doi.org/10.2218/gtopdb/F52/2021.3
- Abo-Aly, M., George, B., Shokri, E., Chelvarajan, L., el-Helw, M., Smyth, S. S., Abdel-Latif, A., & Ziada, K. (2021). Cangrelor in addition to standard therapy reduces cardiac damage and inflammatory markers in patients with ST-segment elevation myocardial infarction. *Journal of Thrombosis and Thrombolysis*, 52(3), 934–940. https://doi. org/10.1007/s11239-020-02345-8
- Alexander, S. P., Christopoulos, A., Davenport, A. P., Kelly, E., Mathie, A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Pawson, A. J., Southan, C., Davies, J. A., Abbracchio, M. P., Alexander, W., al-Hosaini, K., Bäck, M., Barnes, N. M., Bathgate, R., ... Ye, R. D. (2021). The Concise Guide to PHARMACOLOGY 2021/22: G protein-coupled receptors. *British Journal of Pharmacology*, 178(S1), S27–S156. https://doi.org/10.1111/bph.15538
- Alexander, S. P., Fabbro, D., Kelly, E., Mathie, A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Pawson, A. J., Southan, C., Davies, J. A., Beuve, A., Brouckaert, P., Bryant, C., Burnett, J. C., Farndale, R. W., Friebe, A., Garthwaite, J., ... Waldman, S. A. (2021a). THE CONCISE GUIDE TO PHARMACOLOGY 2021/22: Catalytic receptors. *British Journal of Pharmacology*, 178(S1), S264–S312. https://doi.org/10.1111/bph.15541
- Alexander, S. P., Fabbro, D., Kelly, E., Mathie, A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Pawson, A. J., Southan, C., Davies, J. A., Boison, D., Burns, K. E., Dessauer, C., Gertsch, J., Helsby, N. A., Izzo, A. A., Koesling, D., ... Wong, S. S. (2021b). THE CONCISE GUIDE TO PHARMACOLOGY 2021/22: Enzymes. *British Journal of Pharmacology*, 178(S1), S313–S411. https://doi.org/10.1111/bph.15542
- Alexander, S. P., Kelly, E., Mathie, A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Pawson, A. J., Southan, C., Buneman, O. P., Cidlowski, J. A., Christopoulos, A., Davenport, A. P., Fabbro, D., Spedding, M., Striessnig, J., Davies, J. A., Ahlers-Dannen, K. E., ... Zolghadri, Y. (2021a). THE CONCISE GUIDE TO PHARMACOLOGY 2021/22: Other Protein Targets. *British Journal* of *Pharmacology*, 178(S1), S1–S26. https://doi.org/10.1111/bph. 15537
- Alexander, S. P., Kelly, E., Mathie, A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Pawson, A. J., Southan, C., Davies, J. A., Amarosi, L., Anderson, C. M. H., Beart, P. M.,

Broer, S., Dawson, P. A., Hagenbuch, B., Hammond, J. R., Inui, K., ... Verri, T. (2021b). THE CONCISE GUIDE TO PHARMACOLOGY 2021/22: Transporters. *British Journal of Pharmacology*, *178*(S1), S412–S513. https://doi.org/10.1111/bph.15543

- Aloui, C., Prigent, A., Sut, C., Tariket, S., Hamzeh-Cognasse, H., Pozzetto, B., Richard, Y., Cognasse, F., Laradi, S., & Garraud, O. (2014). The signaling role of CD40 ligand in platelet biology and in platelet component transfusion. *International Journal of Molecular Sciences*, 15(12), 22342–22364. https://doi.org/10.3390/ijms1512222342
- Alsharif, K. F., Thomas, M. R., Judge, H. M., Khan, H., Prince, L. R., Sabroe, I., Ridger, V. C., & Storey, R. F. (2015). Ticagrelor potentiates adenosine-induced stimulation of neutrophil chemotaxis and phagocytosis. *Vascular Pharmacology*, 71, 201–207. https://doi.org/10.1016/j. vph.2015.02.006
- Armstrong, D., Summers, C., Ewart, L., Nylander, S., Sidaway, J. E., & van Giezen, J. J. (2014). Characterization of the adenosine pharmacology of ticagrelor reveals therapeutically relevant inhibition of equilibrative nucleoside transporter 1. *Journal of Cardiovascular Pharmacology and Therapeutics*, 19(2), 209–219. https://doi.org/10.1177/ 1074248413511693
- Aungraheeta, R., Conibear, A., Butler, M., Kelly, E., Nylander, S., Mumford, A., & Mundell, S. J. (2016). Inverse agonism at the P2Y₁₂ receptor and ENT1 transporter blockade contribute to platelet inhibition by ticagrelor. *Blood*, 128(23), 2717–2728. https://doi.org/10. 1182/blood-2016-03-707844
- Azar, R. R., Kassab, R., Zoghbi, A., Aboujaoudé, S., el-Osta, H., Ghorra, P., Germanos, M., & Salamé, E. (2006). Effects of clopidogrel on soluble CD40 ligand and on high-sensitivity C-reactive protein in patients with stable coronary artery disease. *American Heart Journal*, 151(2), 521. e521-521.e524. https://doi.org/10.1016/j.ahj.2005.10.021
- Barletta, K. E., Ley, K., & Mehrad, B. (2012). Regulation of neutrophil function by adenosine. Arteriosclerosis, Thrombosis, and Vascular Biology, 32(4), 856–864. https://doi.org/10.1161/atvbaha.111.226845
- Battinelli, E. M., Markens, B. A., & Italiano, J. E. Jr. (2011). Release of angiogenesis regulatory proteins from platelet alpha granules: Modulation of physiologic and pathologic angiogenesis. *Blood*, 118(5), 1359–1369. https://doi.org/10.1182/blood-2011-02-334524
- Berger, J. S., Kornblith, L. Z., Gong, M. N., Reynolds, H. R., Cushman, M., Cheng, Y., McVerry, B. J., Kim, K. S., Lopes, R. D., Atassi, B., Berry, S., Bochicchio, G., de Oliveira Antunes, M., Farkouh, M. E., Greenstein, Y., Hade, E. M., Hudock, K., Hyzy, R., Khatri, P., ... Whitson, M. (2022). Effect of P2Y12 inhibitors on survival free of organ support among non-critically ill hospitalized patients with COVID-19: A randomized clinical trial. *Journal of the American Medical Association*, 327(3), 227– 236. https://doi.org/10.1001/jama.2021.23605
- Bevers, E. M., Comfurius, P., VanRijn, J. L. M. L., Hemker, H. C., & Zwall, R. F. A. (1982). Generation of prothrombin-converting activity and the exposure of phosphatidylserine at the outer surface of platelets. *European Journal of Biochemistry*, 122, 429–436. https://doi.org/ 10.1111/j.1432-1033.1982.tb05898.x
- Bhatt, D. L., Pollack, C. V., Weitz, J. I., Jennings, L. K., Xu, S., Arnold, S. E., Umstead, B. R., Mays, M. C., & Lee, J. S. (2019). Antibody-based ticagrelor reversal agent in healthy volunteers. *The New England Journal of Medicine*, 380(19), 1825–1833. https://doi.org/10.1056/ NEJMoa1901778
- Blair, P., & Flaumenhaft, R. (2009). Platelet α-granules: Basic biology and clinical correlates. *Blood Reviews*, 23(4), 177–189. https://doi.org/10. 1016/j.blre.2009.04.001
- Blumberg, N., Spinelli, S. L., Francis, C. W., Taubman, M. B., & Phipps, R. P. (2009). The platelet as an immune cell–CD40 ligand and transfusion immunomodulation. *Immunologic Research*, 45(2–3), 251–260. https:// doi.org/10.1007/s12026-009-8106-9
- Bonello, L., Laine, M., Kipson, N., Mancini, J., Helal, O., Fromonot, J., Gariboldi, V., Condo, J., Thuny, F., Frere, C., Camoin-Jau, L., Paganelli, F., Dignat-George, F., & Guieu, R. (2014). Ticagrelor

increases adenosine plasma concentration in patients with an acute coronary syndrome. *Journal of the American College of Cardiology*, 63(9), 872–877. https://doi.org/10.1016/j.jacc.2013.09.067

- Borges, P. A., Waclawiak, I., Georgii, J. L., Fraga-Junior, V. S., Barros, J. F., Lemos, F. S., Russo-Abrahão, T., Saraiva, E. M., Takiya, C. M., Coutinho-Silva, R., Penido, C., Mermelstein, C., Meyer-Fernandes, J. R., Canto, F. B., Neves, J. S., Melo, P. A., Canetti, C., & Benjamim, C. F. (2021). Adenosine diphosphate improves wound healing in diabetic mice through P2Y₁₂ receptor activation. *Frontiers in Immunology*, *12*, 651740. https://doi.org/10.3389/fimmu.2021.651740
- Borst, O., Münzer, P., Gatidis, S., Schmidt, E.-M., Schönberger, T., Schmid, E., Towhid, S. T., Stellos, K., Seizer, P., May, A. E., Lang, F., & Gawaz, M. (2012). The inflammatory chemokine CXC motif ligand 16 triggers platelet activation and adhesion via CXC motif receptor 6-dependent phosphatidylinositide 3-kinase/Akt signaling. *Circulation Research*, 111(10), 1297–1307. https://doi.org/10.1161/ CIRCRESAHA.112.276444
- Boyce, J. A. (2007). Mast cells and eicosanoid mediators: A system of reciprocal paracrine and autocrine regulation. *Immunological Reviews*, 217, 168–185. https://doi.org/10.1111/j.1600-065X.2007. 00512.x
- Bradbury, C. A., Lawler, P. R., Stanworth, S. J., McVerry, B., McQuilten, Z., Higgins, A. M., Mouncey, P. R., al-Beidh, F., Rowan, K. M., Berry, L. R., Lorenzi, E., Zarychanski, R., Arabi, Y. M., Annane, D., Beane, A., van Bentum-Puijk, W., Bhimani, Z., Bihari, S., Bonten, M. J. M., ... Gordon, A. C. (2022). Effect of antiplatelet therapy on survival and organ support-free days in critically ill patients with COVID-19: A randomized clinical trial. *Journal of the American Medical Association*, 327(13), 1247–1259. https://doi.org/10.1001/jama.2022.2910
- Braun, O. Ö., Johnell, M., arenhorst, C., James, S., Brandt, J. T., Jakubowski, J. A., Winters, K. J., Wallentin, L., Erlinge, D., & Siegbahn, A. (2008). Greater reduction of platelet activation markers and platelet-monocyte aggregates by prasugrel compared to clopidogrel in stable coronary artery disease. *Thrombosis & Haemostasis*, 100(10), 626–633. https://doi.org/10.1160/TH08-05-0313
- Butt, E., Abel, K., Krieger, M., Palm, D., Hoppe, V., Hoppe, J., & Walter, U. (1994). cAMP- and cGMP-dependent protein kinase phosphorylation sites of the focal adhesion vasodilator-stimulated phosphoprotein (VASP) in vitro and in intact human platelets. *Journal of Biological Chemistry*, 269(20), 14509–14517. https://doi.org/10.1016/S0021-9258(17)36652-8
- Caffrey, A. R., Appaneal, H. J., LaPlante, K. L., Lopes, V. V., Ulloa, E. R., Nizet, V., & Sakoulas, G. (2022). Impact of clopidogrel on clinical outcomes in patients with *Staphylococcus aureus* bacteremia: A national retrospective cohort study. *Antimicrobial Agents and Chemotherapy*, 66(6), e0211721. https://doi.org/10.1128/aac.02117-21
- CAPRIE Steering Committee. (1996). A randomised, blinded, trial of clopidogrel versus aspirin in patients at risk of ischaemic events (CAPRIE). *Lancet*, 348(9038), 1329–1339. https://doi.org/10.1016/S0140-6736 (96)09457-3
- Cauwels, A., Rogge, E., Vandendriessche, B., Shiva, S., & Brouckaert, P. (2014). Extracellular ATP drives systemic inflammation, tissue damage and mortality. *Cell Death & Disease*, 5(3), e1102. https://doi.org/10. 1038/cddis.2014.70
- Chang, Y., Bluteau, D., Deb II, I. N., & Vainchenker, W. (2007). From hematopoietic stem cells to platelets. *Journal of Thrombosis and Haemostasis*, 5(Suppl. 1), 318–327. https://doi.org/10.1111/j.1538-7836.2007. 02472.x
- Chen, Y., Zhong, H., Zhao, Y., Luo, X., & Gao, W. (2020). Role of platelet biomarkers in inflammatory response. *Biomarker Research*, 8(1), 28. https://doi.org/10.1186/s40364-020-00207-2
- Chen, Y. G., Xu, F., Zhang, Y., Ji, Q. S., Sun, Y., Lü, R. J., & Li, R. J. (2006). Effect of aspirin plus clopidogrel on inflammatory markers in patients with non-ST-segment elevation acute coronary syndrome. *Chinese Medical Journal*, 119(1), 32–36.

BRITISH PHARMACOLOGICA 12

- BJP BJP SOCIETY
- Chen, Z. M., Jiang, L. X., Chen, Y. P., Xie, J. X., Pan, H. C., Peto, R., Collins, R., Liu, L. S., & COMMIT (ClOpidogrel and Metoprolol in Myocardial Infarction Trial) collaborative group. (2005). Addition of clopidogrel to aspirin in 45 852 patients with acute myocardial infarction: Randomised placebo-controlled trial. *Lancet*, 366(9497), 1607–1621. https://doi.org/10.1016/S0140-6736(05)67660-X
- Ciferri, S., Emiliani, C., Guglielmini, G., Orlacchio, A., Nenci, G. G., & Gresele, P. (2000). Platelets release their lysosomal content in vivo in humans upon activation. *Thrombosis & Haemostasis*, 83(1), 157–164. https://doi.org/10.1055/s-0037-1613772
- Claushuis, T. A. M., de Vos, A. F., Roelofs, J., de Boer, O. J., van't Veer, C., & van der Poll, T. (2019). Platelet-dense granules worsen preinfection thrombocytopenia during gram-negative pneumonia-derived sepsis. *Journal of Innate Immunity*, 11(2), 168–180. https://doi.org/10. 1159/000494147
- Cognasse, F., Duchez, A. C., Audoux, E., Ebermeyer, T., Arthaud, C. A., Prier, A., Eyraud, M. A., Mismetti, P., Garraud, O., Bertoletti, L., & Hamzeh-Cognasse, H. (2022). Platelets as key factors in inflammation: Focus on CD40L/CD40. Frontiers in Immunology, 13, 825892. https:// doi.org/10.3389/fimmu.2022.825892
- Cosemans, J. M., Munnix, I. C., Wetzker, R., Heller, R., Jackson, S. P., & Heemskerk, J. W. (2006). Continuous signaling via PI3K isoforms β and γ is required for platelet ADP receptor function in dynamic thrombus stabilization. *Blood*, 108(9), 3045–3052. https://doi.org/10.1182/ blood-2006-03-006338
- CURE Trial Investigators. (2001). Effects of clopidogrel in addition to aspirin in patients with acute coronary syndromes without ST-segment elevation. New England Journal of Medicine, 345, 494–502. https://doi. org/10.1056/NEJMoa010746
- DeFilippis, A. P., Chapman, A. R., Mills, N. L., de Lemos, J. A., Arbab-Zadeh, A., Newby, L. K., & Morrow, D. A. (2019). Assessment and treatment of patients with type 2 myocardial infarction and acute nonischemic myocardial injury. *Circulation*, 140(20), 1661–1678. https:// doi.org/10.1161/circulationaha.119.040631
- Delano, M. J., & Ward, P. A. (2016). The immune system's role in sepsis progression, resolution, and long-term outcome. *Immunological Reviews*, 274(1), 330–353. https://doi.org/10.1111/imr.12499
- Dobesh, P. P. (2009). Pharmacokinetics and pharmacodynamics of prasugrel, a thienopyridine P2Y12 inhibitor. *Pharmacotherapy*, 29(9), 1089– 1102. https://doi.org/10.1592/phco.29.9.1089
- Elgueta, R., Benson, M. J., de Vries, V. C., Wasiuk, A., Guo, Y., & Noelle, R. J. (2009). Molecular mechanism and function of CD40/CD40L engagement in the immune system. *Immunological Reviews*, 229(1), 152–172. https://doi.org/10.1111/j.1600-065X. 2009.00782.x
- Entsie, P., Kang, Y., Amoafo, E. B., Schöneberg, T., & Liverani, E. (2023). The signaling pathway of the ADP receptor P2Y₁₂ in the immune system: Recent discoveries and new challenges. *International Journal of Molecular Sciences*, 24(7), 6709. https://doi.org/10.3390/ ijms24076709
- Etulain, J., Martinod, K., Wong, S. L., Cifuni, S. M., Schattner, M., & Wagner, D. D. (2015). P-selectin promotes neutrophil extracellular trap formation in mice. *Blood*, 126(2), 242–246. https://doi.org/10. 1182/blood-2015-01-624023
- Frelinger, A., Jakubowski, J., FuLi, Y., Barnard, M., Fox, M., Linden, M., Sugidachi, A., Winters, K., Furman, M., & Michelson, A. (2007). The active metabolite of prasugrel inhibits ADP-stimulated thromboinflammatory markers of platelet activation: Influence of other blood cells, calcium, and aspirin. *Thrombosis & Haemostasis*, 98, 192–200. https://doi.org/10.1160/TH07-01-0010
- Furman, D., Campisi, J., Verdin, E., Carrera-Bastos, P., Targ, S., Franceschi, C., Ferrucci, L., Gilroy, D. W., Fasano, A., Miller, G. W., Miller, A. H., Mantovani, A., Weyand, C. M., Barzilai, N., Goronzy, J. J., Rando, T. A., Effros, R. B., Lucia, A., Kleinstreuer, N., & Slavich, G. M. (2019). Chronic inflammation in the etiology of disease across the life

span. Nature Medicine, 25(12), 1822-1832. https://doi.org/10.1038/ s41591-019-0675-0

- Gachet, C. (2012). P2Y₁₂ receptors in platelets and other hematopoietic and non-hematopoietic cells. *Purinergic Signal*, 8(3), 609–619. https:// doi.org/10.1007/s11302-012-9303-x
- Garshick, M. S., Rosenthal, P. B., Luttrell-Williams, E., Cronstein, B. N., & Berger, J. S. (2021). Ticagrelor added to methotrexate improves rheumatoid arthritis disease severity. *Rheumatology (Oxford)*, 60(11), 5473– 5475. https://doi.org/10.1093/rheumatology/keab481
- Gąsecka, A., Borovac, J. A., Guerreiro, R. A., Giustozzi, M., Parker, W., Caldeira, D., & Chiva-Blanch, G. (2021). Thrombotic complications in patients with COVID-19: Pathophysiological mechanisms, diagnosis, and treatment. *Cardiovascular Drugs and Therapy*, *35*(2), 215–229. https://doi.org/10.1007/s10557-020-07084-9
- Gasecka, A., Nieuwland, R., Budnik, M., Dignat-George, F., Eyileten, C., Harrison, P., Lacroix, R., Leroyer, A., Opolski, G., Pluta, K., van der Pol, E., Postuła, M., Siljander, P., Siller-Matula, J. M., & Filipiak, K. J. (2020). Ticagrelor attenuates the increase of extracellular vesicle concentrations in plasma after acute myocardial infarction compared to clopidogrel. *Journal of Thrombosis and Haemostasis*, 18(3), 609–623. https://doi.org/10.1111/jth.14689
- Giustozzi, M., Ehrlinder, H., Bongiovanni, D., Borovac, J. A., Guerreiro, R. A., Gąsecka, A., Papakonstantinou, P. E., & Parker, W. A. E. (2021). Coagulopathy and sepsis: Pathophysiology, clinical manifestations and treatment. *Blood Reviews*, 50, 100864. https://doi.org/10.1016/j.blre.2021.100864
- Gorog, D. A., Storey, R. F., Gurbel, P. A., Tantry, U. S., Berger, J. S., Chan, M. Y., Duerschmied, D., Smyth, S. S., Parker, W. A. E., Ajjan, R. A., Vilahur, G., Badimon, L., Berg, J. M., Cate, H., Peyvandi, F., Wang, T. T., & Becker, R. C. (2022). Current and novel biomarkers of thrombotic risk in COVID-19: A consensus statement from the International COVID-19 Thrombosis Biomarkers Colloquium. *Nature Reviews Cardiology*, 19(7), 475–495. https://doi.org/10.1038/s41569-021-00665-7
- Grainger, D. J., Wakefield, L., Bethell, H. W., Farndale, R. W., & Metcalfe, J. C. (1995). Release and activation of platelet latent TGF-β in blood clots during dissolution with plasmin. *Nature Medicine*, 1(9), 932–937. https://doi.org/10.1038/nm0995-932
- Gross, A. K., Dunn, S. P., Feola, D. J., Martin, C. A., Charnigo, R., Li, Z., Abdel-Latif, A., & Smyth, S. S. (2013). Clopidogrel treatment and the incidence and severity of community acquired pneumonia in a cohort study and meta-analysis of antiplatelet therapy in pneumonia and critical illness. *Journal of Thrombosis and Thrombolysis*, 35, 147–154. https://doi.org/10.1007/s11239-012-0833-4
- Gurbel, P. A., Bliden, K. P., Butler, K., Tantry, U. S., Gesheff, T., Wei, C., Teng, R., Antonino, M. J., Patil, S. B., Karunakaran, A., Kereiakes, D. J., Parris, C., Purdy, D., Wilson, V., Ledley, G. S., & Storey, R. F. (2009). Randomized double-blind assessment of the ONSET and OFFSET of the antiplatelet effects of ticagrelor versus clopidogrel in patients with stable coronary artery disease: The ONSET/OFFSET study. *Circulation*, 120(25), 2577–2585. https://doi.org/10.1161/circulationaha.109. 912550
- Gurbel, P. A., Bliden, K. P., & Tantry, U. S. (2006). Effect of clopidogrel with and without eptifibatide on tumor necrosis factor-alpha and C-reactive protein release after elective stenting: Results from the CLEAR PLATELETS 1b study. *Journal of the American College of Cardiol*ogy, 48(11), 2186–2191. https://doi.org/10.1016/j.jacc.2005.12.084
- Hagiwara, S., Iwasaka, H., Hasegawa, A., Oyama, M., Imatomi, R., Uchida, T., & Noguchi, T. (2011). Adenosine diphosphate receptor antagonist clopidogrel sulfate attenuates LPS-induced systemic inflammation in a rat model. *Shock*, 35(3), 289–292. https://doi.org/10. 1097/SHK.0b013e3181f48987
- Hajsadeghi, S., Chitsazan, M., Chitsazan, M., Salehi, N., Amin, A., Bidokhti, A. A., Babaali, N., Bordbar, A., Hejrati, M., & Moghadami, S. (2016). Prasugrel results in higher decrease in high-sensitivity

C-reactive protein level in patients undergoing percutaneous coronary intervention comparing to clopidogrel. *Clinical Medicine Insights: Cardiology*, 10, 149–155. https://doi.org/10.4137/cmc.S32804

- Hansildaar, R., Vedder, D., Baniaamam, M., Tausche, A.-K., Gerritsen, M., & Nurmohamed, M. T. (2021). Cardiovascular risk in inflammatory arthritis: Rheumatoid arthritis and gout. *The Lancet Rheumatology*, 3(1), e58–e70. https://doi.org/10.1016/S2665-9913(20)30221-6
- Harada, K., Matsumoto, Y., & Umemura, K. (2011). Adenosine diphosphate receptor P2Y₁₂-mediated migration of host smooth muscle-like cells and leukocytes in the development of transplant arteriosclerosis. *Transplantation*, 92(2), 148–154. https://doi.org/10.1097/TP. 0b013e318221d407
- Harrison, P., & Cramer, E. M. (1993). Platelet α-granules. *Blood Reviews*, 7, 52–62. https://doi.org/10.1016/0268-960x(93)90024-x
- Haskó, G., & Pacher, P. (2012). Regulation of macrophage function by adenosine. Arteriosclerosis, Thrombosis, and Vascular Biology, 32(4), 865–869. https://doi.org/10.1161/atvbaha.111.226852
- Hechler, B., Eckly, A., Ohlmann, P., Cazenave, J.-P., & Gachet, C. (1998). The P2Y₁ receptor, necessary but not sufficient to support full ADPinduced platelet aggregation, is not the target of the drug clopidogrel. *British Journal of Haematology*, 103, 858–866. https://doi.org/10. 1046/j.1365-2141.1998.01056.x
- Heijnen, H., & van der Sluijs, P. (2015). Platelet secretory behaviour: As diverse as the granules ... or not? *Journal of Thrombosis and Haemosta*sis, 13(12), 2141–2151. https://doi.org/10.1111/jth.13147
- Heitzer, T., Rudolph, V., Schwedhelm, E., Karstens, M., Sydow, K., Ortak, M., Tschentscher, P., Meinertz, T., Böger, R., & Baldus, S. (2006). Clopidogrel improves systemic endothelial nitric oxide bioavailability in patients with coronary artery disease: Evidence for antioxidant and antiinflammatory effects. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 26(7), 1648–1652. https://doi.org/10.1161/01.ATV. 0000225288.74170.dc
- Hennigan, B. W., Good, R., Adamson, C., Parker, W. A. E., Martin, L., Anderson, L., Campbell, M., Serruys, P. W., Storey, R. F., & Oldroyd, K. G. (2022). Recovery of platelet reactivity following cessation of either aspirin or ticagrelor in patients treated with dual antiplatelet therapy following percutaneous coronary intervention: A GLOBAL LEADERS substudy. *Platelets*, 33(1), 141–146. https://doi. org/10.1080/09537104.2020.1863937
- Hers, I. (2007). Insulin-like growth factor-1 potentiates platelet activation via the IRS/PI3Kα pathway. *Blood*, 110(13), 4243–4252. https://doi. org/10.1182/blood-2006-10-050633
- Hollopeter, G., Jantzen, H. M., Vincent, D., Li, G., England, L., Ramakrishnan, V., Yang, R. B., Nurden, P., Nurden, A., Julius, D., & Conley, P. B. (2001). Identification of the platelet ADP receptor targeted by antithrombotic drugs. *Nature*, 409(6817), 202–207. https:// doi.org/10.1038/35051599
- Horstrup, K., Jablonka, B., Hönig-Liedl, P., Just, M., Kochsiek, K., & Walter, U. (1994). Phosphorylation of focal adhesion vasodilatorstimulated phosphoprotein at Ser157 in intact human platelets correlates with fibrinogen receptor inhibition. *European Journal of Biochemistry*, 225(1), 21–27. https://doi.org/10.1111/j.1432-1033.1994. 00021.x
- Husted, S., Storey, R. F., Harrington, R. A., Emanuelsson, H., & Cannon, C. P. (2010). Changes in inflammatory biomarkers in patients treated with ticagrelor or clopidogrel. *Clinical Cardiology*, 33(4), 206– 212. https://doi.org/10.1002/clc.20732
- Isenberg, J. S., Romeo, M. J., Yu, C., Yu, C. K., Nghiem, K., Monsale, J., Rick, M. E., Wink, D. A., Frazier, W. A., & Roberts, D. D. (2008). Thrombospondin-1 stimulates platelet aggregation by blocking the antithrombotic activity of nitric oxide/cGMP signaling. *Blood*, 111(2), 613–623. https://doi.org/10.1182/blood-2007-06-098392
- Jakubowski, J. A., Matsushima, N., Asai, F., Naganuma, H., Brandt, J. T., Hirota, T., Freestone, S., & Winters, K. J. (2007). A multiple dose study of prasugrel (CS-747), a novel thienopyridine P2Y₁₂ inhibitor,

compared with clopidogrel in healthy humans. *British Journal of Clinical Pharmacology*, 63(4), 421–430. https://doi.org/10.1111/j.1365-2125. 2006.02792.x

- Janzon, L. (1996). The STIMS trial: The ticlopidine experience and its clinical applications. Vascular Medicine, 1(2), 141–143. https://doi.org/10. 1177/1358863x9600100210
- Johnston, S. C., Amarenco, P., Albers, G. W., Denison, H., Easton, J. D., Evans, S. R., Held, P., Jonasson, J., Minematsu, K., Molina, C. A., Wang, Y., & Wong, K. S. L. (2016). Ticagrelor versus aspirin in acute stroke or transient ischemic attack. *The New England Journal of Medicine*, 375(1), 35–43. https://doi.org/10.1056/NEJMoa1603060
- Judge, H. M., Buckland, R., Sugidachi, A., Jakubowski, J. A., & Storey, R. F. (2008). The active metabolite of prasugrel effectively blocks the platelet P2Y₁₂ receptor and inhibits procoagulant and pro-inflammatory platelet responses. *Platelets*, 19, 125–133. https://doi.org/10.1080/ 09537100701694144
- Kardas, G., Daszyńska-Kardas, A., Marynowski, M., Brząkalska, O., Kuna, P., & Panek, M. (2020). Role of platelet-derived growth factor (PDGF) in asthma as an immunoregulatory factor mediating airway remodeling and possible pharmacological target. *Frontiers in Pharmacology*, 11, 47. https://doi.org/10.3389/fphar.2020.00047
- Karhausen, J., Choi, H. W., Maddipati, K. R., Mathew, J. P., Ma, Q., Boulaftali, Y., Lee, R. H., Bergmeier, W., & Abraham, S. N. (2020). Platelets trigger perivascular mast cell degranulation to cause inflammatory responses and tissue injury. *Science Advances*, 6(12), eaay6314. https://doi.org/10.1126/sciadv.aay6314
- Khan, J. N., Greenwood, J. P., Nazir, S. A., Lai, F. Y., Dalby, M., Curzen, N., Hetherington, S., Kelly, D. J., Blackman, D., Peebles, C., Wong, J., Flather, M., Swanton, H., Gershlick, A. H., & McCann, G. P. (2016). Infarct size following treatment with second- versus third-generation P2Y₁₂ antagonists in patients with multivessel coronary disease at STsegment elevation myocardial infarction in the CvLPRIT study. *Journal of the American Heart Association*, *5*(6), e003403. https://doi.org/10. 1161/jaha.116.003403
- Kiers, D., van der Heijden, W. A., van Ede, L., Gerretsen, J., de Mast, Q., van der Ven, A. J., el Messaoudi, S., Rongen, G. A., Gomes, M., Kox, M., Pickkers, P., & Riksen, N. P. (2017). A randomised trial on the effect of anti-platelet therapy on the systemic inflammatory response in human endotoxaemia. *Thrombosis & Haemostasis*, 117(9), 1798–1807. https://doi.org/10.1160/th16-10-0799
- Knighton, D. R., Zheng, J., Ten Eyck, L. F., Xuong, N.-H., Taylor, S. S., & Sowadski, J. M. (1991). Structure of a peptide inhibitor bound to the catalytic subunit of cyclic adenosine monophosphate-dependent protein kinase. *Science*, 253, 414–420. https://doi.org/10.1126/science. 1862343
- Lancellotti, P., Musumeci, L., Jacques, N., Servais, L., Goffin, E., Pirotte, B., & Oury, C. (2019). Antibacterial activity of ticagrelor in conventional antiplatelet dosages against antibiotic-resistant grampositive bacteria. JAMA Cardiology, 4(6), 596–599. https://doi.org/10. 1001/jamacardio.2019.1189
- Leeten, K., Jacques, N., Lancellotti, P., & Oury, C. (2021). Aspirin or ticagrelor in Staphylococcus aureus infective endocarditis: Where do we stand? Frontiers in Cell and Development Biology, 9, 716302. https:// doi.org/10.3389/fcell.2021.716302
- Levi, M., van der Poll, T., & Büller, H. R. (2004). Bidirectional relation between inflammation and coagulation. *Circulation*, 109(22), 2698– 2704. https://doi.org/10.1161/01.CIR.0000131660.51520.9A
- Libby, P., Buring, J. E., Badimon, L., Hansson, G. K., Deanfield, J., Bittencourt, M. S., Tokgözoğlu, L., & Lewis, E. F. (2019). Atherosclerosis. Nature Reviews. Disease Primers, 5(1), 56. https://doi.org/10.1038/ s41572-019-0106-z
- Lipcsey, M., Larsson, A., Olovsson, M., Sjölin, J., & Eriksson, M. B. (2005). Early endotoxin-mediated haemostatic and inflammatory responses in the clopidogrel-treated pig. *Platelets*, 16(7), 408–414. https://doi.org/ 10.1080/09537100500163168

13

- Liverani, E., Rico, M. C., Garcia, A. E., Kilpatrick, L. E., & Kunapuli, S. P. (2012). Prasugrel metabolites inhibit neutrophil functions. *Journal of Pharmacology and Experimental Therapeutics*, 344(1), 231–243. https:// doi.org/10.1124/jpet.112.195883
- Liverani, E., Rico, M. C., Tsygankov, A. Y., Kilpatrick, L. E., & Kunapuli, S. P. (2016). P2Y₁₂ receptor modulates sepsis-induced inflammation. Arteriosclerosis, Thrombosis, and Vascular Biology, 36(5), 961–971. https:// doi.org/10.1161/atvbaha.116.307401
- Lupu, L., Shepshelovich, D., Banai, S., Hershkoviz, R., & Isakov, O. (2020). Effect of ticagrelor on reducing the risk of gram-positive infections in patients with acute coronary syndrome. *American Journal of Cardiology*, 130, 56–63. https://doi.org/10.1016/j.amjcard.2020.06.016
- Lussana, F., di Marco, F., Terraneo, S., Parati, M., Razzari, C., Scavone, M., Femia, E. A., Moro, A., Centanni, S., & Cattaneo, M. (2015). Effect of prasugrel in patients with asthma: Results of PRINA, a randomized, double-blind, placebo-controlled, cross-over study. *Journal of Thrombosis* and Haemostasis, 13(1), 136–141. https://doi.org/10.1111/jth.12779
- Mahaut-Smith, M. P., Jones, S., & Evans, R. J. (2011). The P2X1 receptor and platelet function. *Purinergic Signalling*, 7(3), 341–356. https://doi. org/10.1007/s11302-011-9224-0
- Majumdar, S., Gonder, D., Koutsis, B., & Poncz, M. (1991). Characterization of the human beta-thromboglobulin gene. Comparison with the gene for platelet factor 4. *Journal of Biological Chemistry*, 266(9), 5785– 5789. https://doi.org/10.1016/S0021-9258(19)67665-9
- Mansour, A., Bachelot-Loza, C., Nesseler, N., Gaussem, P., & Gouin-Thibault, I. (2020). P2Y₁₂ inhibition beyond thrombosis: Effects on inflammation. *International Journal of Molecular Sciences*, 21(4), 1391. https://doi.org/10.3390/ijms21041391
- Marcoux, G., Laroche, A., Espinoza Romero, J., & Boilard, E. (2021). Role of platelets and megakaryocytes in adaptive immunity. *Platelets*, 32(3), 340–351. https://doi.org/10.1080/09537104.2020.1786043
- Martins, P. A. D. C., van Gils, J. M., Mol, A., Hordijk, P. L., & Zwaginga, J. J. (2006). Platelet binding to monocytes increases the adhesive properties of monocytes by up-regulating the expression and functionality of β_1 and β_2 integrins. *Journal of Leukocyte Biology*, *79*(3), 499–507. https://doi.org/10.1189/jlb.0605318
- Masuyama, T., Sakuma, M., Waku, R., Hirose, S., Kitahara, K., Naganuma, J., Yazawa, H., Toyoda, S., Abe, S., Nakajima, T., & Inoue, T. (2021). Effects of switching from clopidogrel to prasugrel at the chronic phase after coronary stenting on antiplatelet action and vascular endothelial function: Switch-Pras study. *Heart and Vessels*, 36(4), 442–451. https://doi.org/10.1007/s00380-020-01714-w
- Matetzky, S., Shenkman, B., Guetta, V., Shechter, M., Beinart, R., Goldenberg, I., Novikov, I., Pres, H., Savion, N., Varon, D., & Hod, H. (2004). Clopidogrel resistance is associated with increased risk of recurrent atherothrombotic events in patients with acute myocardial infarction. *Circulation*, 109(25), 3171–3175. https://doi.org/10.1161/ 01.cir.0000130846.46168.03
- Mauler, M., Herr, N., Schoenichen, C., Witsch, T., Marchini, T., Härdtner, C., Koentges, C., Kienle, K., Ollivier, V., Schell, M., Dorner, L., Wippel, C., Stallmann, D., Normann, C., Bugger, H., Walther, P., Wolf, D., Ahrens, I., Lämmermann, T., ... Duerschmied, D. (2019). Platelet serotonin aggravates myocardial ischemia/reperfusion injury via neutrophil degranulation. *Circulation*, 139(7), 918–931. https://doi. org/10.1161/circulationaha.118.033942
- McNicol, A., & Israels, S. J. (1999). Platelet dense granules: Structure, function and implications for haemostasis. *Thrombosis Research*, 95, 1–18. https://doi.org/10.1016/s0049-3848(99)00015-8
- Micklewright, J. J., Layhadi, J. A., & Fountain, S. J. (2018). P2Y₁₂ receptor modulation of ADP-evoked intracellular Ca²⁺ signalling in THP-1 human monocytic cells. *British Journal of Pharmacology*, 175(12), 2483–2491. https://doi.org/10.1111/bph.14218
- Mozzini, C., Garbin, U., Fratta Pasini, A. M., & Cominacini, L. (2017). An exploratory look at NETosis in atherosclerosis. *Internal and Emergency Medicine*, 12(1), 13–22. https://doi.org/10.1007/s11739-016-1543-2

- Nanhwan, M. K., Ling, S., Kodakandla, M., Nylander, S., Ye, Y., & Birnbaum, Y. (2014). Chronic treatment with ticagrelor limits myocardial infarct size: An adenosine and cyclooxygenase-2-dependent effect. Arteriosclerosis, Thrombosis, and Vascular Biology, 34(9), 2078– 2085. https://doi.org/10.1161/atvbaha.114.304002
- Nedeva, C., Menassa, J., & Puthalakath, H. (2019). Sepsis: Inflammation is a necessary evil. *Frontiers in Cell and Development Biology*, 7, 108. https://doi.org/10.3389/fcell.2019.00108
- Nelson, T. A., Parker, W. A. E., Ghukasyan Lakic, T., Westerbergh, J., James, S. K., Siegbahn, A., Becker, R. C., Himmelmann, A., Wallentin, L., & Storey, R. F. (2022). Differential effect of clopidogrel and ticagrelor on leukocyte count in relation to patient characteristics, biomarkers and genotype: A PLATO substudy. *Platelets*, 33(3), 425– 431. https://doi.org/10.1080/09537104.2021.1934667
- Nevzorova, T. A., Mordakhanova, E. R., Daminova, A. G., Ponomareva, A. A., Andrianova, I. A., Le Minh, G., & Weisel, J. W. (2019). Platelet factor 4-containing immune complexes induce platelet activation followed by calpain-dependent platelet death. *Cell Death Discovery*, 5(1), 106. https://doi.org/10.1038/s41420-019-0188-0
- Nicolau, D. P., Tessier, P. R., & Nightingale, C. H. (1999). Beneficial effect of combination antiplatelet therapy on the development of experimental *Staphylococcus aureus* endocarditis. *International Journal of Antimicrobial Agents*, 11(2), 159–161. https://doi.org/10.1016/S0924-8579 (98)00092-2
- Nylander, S., Femia, E. A., Scavone, M., Berntsson, P., Asztély, A. K., Nelander, K., Löfgren, L., Nilsson, R. G., & Cattaneo, M. (2013). Ticagrelor inhibits human platelet aggregation via adenosine in addition to P2Y₁₂ antagonism. *Journal of Thrombosis and Haemostasis*, 11(10), 1867–1876. https://doi.org/10.1111/jth.12360
- O'Brien, X. M., Biron, B. M., & Reichner, J. S. (2017). Consequences of extracellular trap formation in sepsis. *Current Opinion in Hematology*, 24(1), 66–71. https://doi.org/10.1097/moh.00000000000303
- Orme, R. C., Parker, W. A. E., Thomas, M. R., Judge, H. M., Baster, K., Sumaya, W., Morgan, K. P., McMellon, H. C., Richardson, J. D., Grech, E. D., Wheeldon, N. M., Hall, I. R., Iqbal, J., Barmby, D., Gunn, J. P., & Storey, R. F. (2018). Study of two dose regimens of ticagrelor compared with clopidogrel in patients undergoing percutaneous coronary intervention for stable coronary artery disease. *Circulation*, 138, 1290–1300. https://doi.org/10.1161/CIRCULATIONAHA.118. 034790
- Otto, G. P., Sossdorf, M., Boettel, J., Kabisch, B., Breuel, H., Winning, J., & Lösche, W. (2013). Effects of low-dose acetylsalicylic acid and atherosclerotic vascular diseases on the outcome in patients with severe sepsis or septic shock. *Platelets*, 24(6), 480–485. https://doi.org/10.3109/ 09537104.2012.724482
- Oury, C., Meyers, S., Jacques, N., Leeten, K., Jiang, Z., Musumeci, L., Lox, M., Debuisson, M., Goffin, E., Pirotte, B., Delvenne, P., Nchimi, A., Hubert, C., Heptia, M., Hubert, P., Kuijpers, M. J. E., Vanassche, T., Martinod, K., Verhamme, P., & Lancellotti, P. (2023). Protective effect of ticagrelor against infective endocarditis induced by virulent *Staphylococcus aureus* in mice. *JACC: Basic to Translational Science*. https:// doi.org/10.1016/j.jacbts.2023.02.003
- Ow, K. W., Parker, W. A. E., Porter, M. M., Hanson, J., Judge, H. M., Briffa, N. P., Thomas, M. R., & Storey, R. F. (2020). Offset of ticagrelor prior to coronary artery bypass graft surgery for acute coronary syndromes: Effects on platelet function and cellular adenosine uptake. *Platelets*, 31(7), 945–951. https://doi.org/10.1080/09537104.2019. 1709631
- Parker, W. A., Bhatt, D. L., Prats, J., Day, J. R. S., Steg, P. G., Stone, G. W., Hamm, C. W., Mahaffey, K. W., Price, M. J., Michael Gibson, C., White, H. D., Storey, R. F., & the CHAMPION PHOENIX Investigators. (2017). Characteristics of dyspnoea and associated clinical outcomes in the CHAMPION PHOENIX study. *Thrombosis & Haemostasis*, 117(6), 1093–1100. https://doi.org/10.1160/th16-12-0958

- Parker, W. A., & Storey, R. F. (2016a). Long-term antiplatelet therapy following myocardial infarction: Implications of PEGASUS-TIMI 54. *Heart*, 102(10), 783–789. https://doi.org/10.1136/heartjnl-2015-307858
- Parker, W. A. E., Gorog, D. A., Geisler, T., Vilahur, G., Sibbing, D., Rocca, B., & Storey, R. F. (2020). Prevention of stroke in patients with chronic coronary syndromes or peripheral arterial disease. *European Heart Journal Supplements*, 22(Suppl M), M26-m34. https://doi.org/ 10.1093/eurheartj/suaa165
- Parker, W. A. E., Schulte, C., Barwari, T., Phoenix, F., Pearson, S. M., Mayr, M., Grant, P. J., Storey, R. F., & Ajjan, R. A. (2020). Aspirin, clopidogrel and prasugrel monotherapy in patients with type 2 diabetes mellitus: A double-blind randomised controlled trial of the effects on thrombotic markers and microRNA levels. *Cardiovascular Diabetology*, 19(1), 3. https://doi.org/10.1186/s12933-019-0981-3
- Parker, W. A. E., & Storey, R. F. (2016b). Ticagrelor: Agonising over its mechanisms of action. Blood, 128(23), 2595–2597. https://doi.org/10. 1182/blood-2016-10-743930
- Parker, W. A. E., & Storey, R. F. (2018). Thrombotic response. In The ESC textbook of cardiovascular medicine (3rd ed.). Oxford University Press.
- Parker, W. A. E., & Storey, R. F. (2020). Pharmacology and potential role of selatogrel, a subcutaneous platelet P2Y₁₂ receptor antagonist. *Expert Opinion on Emerging Drugs*, 25(1), 1–6. https://doi.org/10.1080/ 14728214.2020.1729121
- Parker, W. A. E., & Storey, R. F. (2021a). Antithrombotic therapy for patients with chronic coronary syndromes. *Heart*, 107(11), 925. https://doi.org/10.1136/heartjnl-2020-316914
- Parker, W. A. E., & Storey, R. F. (2021b). Platelets and the endothelium: Active participants in severe COVID-19 infection. JACC Basic to Translational Science, 6(3), 219–221. https://doi.org/10.1016/j.jacbts.2021. 01.009
- Paruchuri, S., Tashimo, H., Feng, C., Maekawa, A., Xing, W., Jiang, Y., Kanaoka, Y., Conley, P., & Boyce, J. A. (2009). Leukotriene E₄-induced pulmonary inflammation is mediated by the P2Y₁₂ receptor. *Journal of Experimental Medicine*, 206(11), 2543–2555. https://doi.org/10.1084/ jem.20091240
- Pokrovskaya, I. D., Yadav, S., Rao, A., McBride, E., Kamykowski, J. A., Zhang, G., Aronova, M. A., Leapman, R. D., & Storrie, B. (2020). 3D ultrastructural analysis of α-granule, dense granule, mitochondria, and canalicular system arrangement in resting human platelets. *Research and Practice in Thrombosis and Haemostasis*, 4(1), 72–85. https://doi. org/10.1002/rth2.12260
- Puhm, F., Boilard, E., & Machlus, K. R. (2021). Platelet extracellular vesicles: Beyond the blood. Arteriosclerosis, Thrombosis, and Vascular Biology, 41(1), 87–96. https://doi.org/10.1161/ATVBAHA.120.314644
- Rabouel, Y., Magnenat, S., Delabranche, X., Gachet, C., & Hechler, B. (2021). Platelet P2Y₁₂ receptor deletion or pharmacological inhibition does not protect mice from sepsis or septic shock. *TH Open*, 5(3), e343-e352. https://doi.org/10.1055/s-0041-1733857
- Rahman, M., Gustafsson, D., Wang, Y., Thorlacius, H., & Braun, O. O. (2013). Ticagrelor reduces neutrophil recruitment and lung damage in abdominal sepsis. *Platelets*, 25, 257–263. https://doi.org/10.3109/ 09537104.2013.809520
- Ridker, P. M., Everett, B. M., Pradhan, A., MacFadyen, J. G., Solomon, D. H., Zaharris, E., Mam, V., Hasan, A., Rosenberg, Y., Iturriaga, E., Gupta, M., Tsigoulis, M., Verma, S., Clearfield, M., Libby, P., Goldhaber, S. Z., Seagle, R., Ofori, C., Saklayen, M., ... CIRT Investigators. (2019). Low-dose methotrexate for the prevention of atherosclerotic events. *The New England Journal of Medicine*, 380(8), 752–762. https://doi.org/10.1056/NEJMoa1809798
- Ridker, P. M., Everett, B. M., Thuren, T., MacFadyen, J. G., Chang, W. H., Ballantyne, C., Fonseca, F., Nicolau, J., Koenig, W., Anker, S. D., Kastelein, J. J. P., Cornel, J. H., Pais, P., Pella, D., Genest, J., Cifkova, R., Lorenzatti, A., Forster, T., Kobalava, Z., ... Glynn, R. J. (2017). Antiinflammatory therapy with canakinumab for atherosclerotic disease.

New England Journal of Medicine, 377(12), 1119-1131. https://doi. org/10.1056/NEJMoa1707914

- Rudež, G., Pons, D., Leebeek, F., Monraats, P., Schrevel, M., Zwinderman, A., de Winter, R., Tio, R., Doevendans, P., Jukema, W., & de Maat, M. (2008). Platelet receptor P2RY12 haplotypes predict restenosis after percutaneous coronary interventions. *Human Mutation*, 29(3), 375–380. https://doi.org/10.1002/humu.20641
- Satonaka, H., Nagata, D., Takahashi, M., Kiyosue, A., Myojo, M., Fujita, D., Ishimitsu, T., Nagano, T., Nagai, R., & Hirata, Y. (2015). Involvement of P2Y₁₂ receptor in vascular smooth muscle inflammatory changes via MCP-1 upregulation and monocyte adhesion. *American Journal of Physiology. Heart and Circulatory Physiology*, 308(8), H853–H861. https://doi.org/10.1152/ajpheart.00862.2013
- Schmidt, S. N., Reichardt, W., Kaufmann, B. A., Wadle, C., von Elverfeldt, D., Stachon, P., Hilgendorf, I., Wolf, D., Heidt, T., Duerschmied, D., Peter, K., Bode, C., von zur Mühlen, C., & Maier, A. (2021). P2Y₁₂ inhibition in murine myocarditis results in reduced platelet infiltration and preserved ejection fraction. *Cell*, 10(12), 3414. https://doi.org/10.3390/cells10123414
- Schnurr, M., Toy, T., Shin, A., Wagner, M., Cebon, J., & Maraskovsky, E. (2005). Extracellular nucleotide signaling by P2 receptors inhibits IL-12 and enhances IL-23 expression in human dendritic cells: A novel role for the cAMP pathway. *Blood*, 105(4), 1582–1589. https://doi.org/10. 1182/blood-2004-05-1718
- Schwarz, M., Nordt, T., Bode, C., & Peter, K. (2002). The GP IIb/IIIa inhibitor abciximab (c7E3) inhibits the binding of various ligands to the leukocyte integrin Mac-1 (CD11b/CD18, α_Mβ₂). Thrombosis Research, 107(3-4), 121-128. https://doi.org/10.1016/s0049-3848(02) 00207-4
- Seung, H., Wrobel, J., Wadle, C., Bühler, T., Chiang, D., Rettkowski, J., Cabezas-Wallscheid, N., Hechler, B., Stachon, P., Maier, A., Weber, C., Wolf, D., Duerschmied, D., Idzko, M., Bode, C., von zur Mühlen, C., Hilgendorf, I., & Heidt, T. (2022). P2Y₁₂-dependent activation of hematopoietic stem and progenitor cells promotes emergency hematopoiesis after myocardial infarction. *Basic Research in Cardiology*, 117(1), 16. https://doi.org/10.1007/s00395-022-00927-6
- Sexton, T. R., Zhang, G., Macaulay, T. E., Callahan, L. A., Charnigo, R., Vsevolozhskaya, O. A., Li, Z., & Smyth, S. (2018). Ticagrelor reduces thromboinflammatory markers in patients with pneumonia. *JACC Basic Transl Sci*, 3(4), 435–449. https://doi.org/10.1016/j.jacbts.2018. 05.005
- Solheim, S., Pettersen, A. A., Arnesen, H., & Seljeflot, I. (2006). No difference in the effects of clopidogrel and aspirin on inflammatory markers in patients with coronary heart disease. *Thrombosis and Haemostasis*, 96(5), 660–664. https://doi.org/10. 1160/TH06-06-0337
- Spiel, A. O., Derhaschnig, U., Schwameis, M., Bartko, J., Siller-Matula, J. M., & Jilma, B. (2012). Effects of prasugrel on platelet inhibition during systemic endotoxaemia: A randomized controlled trial. *Clinical Science*, 123(10), 591–600. https://doi.org/10.1042/CS20120194
- Stegner, D., & Nieswandt, B. (2011). Platelet receptor signaling in thrombus formation. *Journal of Molecular Medicine*, 89(2), 109–121. https:// doi.org/10.1007/s00109-010-0691-5
- Storey, R. F. (2006). Biology and pharmacology of the platelet P2Y12 receptor. Current Pharmaceutical Design, 12(10), 1255–1259. https:// doi.org/10.2174/138161206776361318
- Storey, R. F., James, S. K., Siegbahn, A., Varenhorst, C., Held, C., Ycas, J., Husted, S. E., Cannon, C. P., Becker, R. C., Steg, P. G., Åsenblad, N., & Wallentin, L. (2014). Lower mortality following pulmonary adverse events and sepsis with ticagrelor compared to clopidogrel in the PLATO study. *Platelets*, 25, 517–525. https://doi.org/10.1016/s0735-1097(12)60483-5
- Storey, R. F., Judge, H. M., Wilcox, R. G., & Heptinstall, S. (2002). Inhibition of ADP-induced P-selectin expression and platelet-leukocyte conjugate formation by clopidogrel and the P2Y₁₂ receptor antagonist AR-

BRITISH PHARMACOLOGICAI C69931MX but not aspirin. *Thrombosis & Haemostasis, 88,* 488–494. https://doi.org/10.1055/s-0037-1613242

- Storey, R. F., Sanderson, H. M., White, A. E., May, J. A., Cameron, K. E., & Heptinstall, S. (2000). The central role of the P_{2T} receptor in amplification of human platelet activation, aggregation, secretion and procoagulant activity. *British Journal of Haematology*, 110(4), 925–934. https://doi.org/10.1046/j.1365-2141.2000.02208.x
- Suffredini, A. F., & Noveck, R. J. (2014). Human endotoxin administration as an experimental model in drug development. *Clinical Pharmacol*ogy & Therapeutics, 96(4), 418–422. https://doi.org/10.1038/clpt. 2014.146
- Tardif, J. C., Kouz, S., Waters, D. D., Bertrand, O. F., Diaz, R., Maggioni, A. P., Pinto, F. J., Ibrahim, R., Gamra, H., Kiwan, G. S., Berry, C., López-Sendón, J., Ostadal, P., Koenig, W., Angoulvant, D., Grégoire, J. C., Lavoie, M. A., Dubé, M. P., Rhainds, D., ... Roubille, F. (2019). Efficacy and safety of low-dose colchicine after myocardial infarction. New England Journal of Medicine, 381(26), 2497–2505. https://doi.org/10.1056/NEJMoa1912388
- Thomas, M. R., Outteridge, S. N., Ajjan, R. A., Phoenix, F., Sangha, G. K., Faulkner, R. E., Ecob, R., Judge, H. M., Khan, H., West, L. E., Dockrell, D. H., Sabroe, I., & Storey, R. F. (2015). Platelet P2Y₁₂ inhibitors reduce systemic inflammation and its prothrombotic effects in an experimental human model. *Arteriosclerosis, Thrombosis, and Vascular Biology*, *35*(12), 2562–2570. https://doi.org/10.1161/atvbaha.115. 306528
- Thomas, M. R., & Storey, R. F. (2015). Effect of P2Y₁₂ inhibitors on inflammation and immunity. *Thrombosis and Haemostasis*, 114(3), 490–497. https://doi.org/10.1160/TH14-12-1068
- Totani, L., Dell'Elba, G., Martelli, N., Di Santo, A., Piccoli, A., Amore, C., & Evangelista, V. (2012). Prasugrel inhibits platelet-leukocyte interaction and reduces inflammatory markers in a model of endotoxic shock in the mouse. *Thrombosis & Haemostasis*, 107(6), 1130–1140. https:// doi.org/10.1160/th11-12-0867
- Vajen, T., Mause, S. F., & Koenen, R. R. (2015). Microvesicles from platelets: Novel drivers of vascular inflammation. *Thrombosis and Haemosta*sis, 114(8), 228–236. https://doi.org/10.1160/TH14-11-0962
- van den Boogaard, F. E., Schouten, M., de Stoppelaar, S. F., Roelofs, J. J., Brands, X., Schultz, M. J., van't Veer, C., & van der Poll, T. (2015). Thrombocytopenia impairs host defense during murine *Streptococcus pneumoniae* pneumonia. *Critical Care Medicine*, 43(3), e75-e83. https://doi.org/10.1097/ccm.00000000000853
- van der Pol, E., Böing, A. N., Harrison, P., Sturk, A., & Nieuwland, R. (2012). Classification, functions, and clinical relevance of extracellular vesicles. *Pharmacological Reviews*, 64(3), 676–705. https://doi.org/10.1124/pr. 112.005983
- Vandendries, E. R., Furie, B. C., & Furie, B. (2004). Role of P-selectin and PSGL-1 in coagulation and thrombosis. *Thrombosis & Haemostasis*, 92, 459–466. https://doi.org/10.1160/TH04-05-0306
- Veloso, T. R., Oechslin, F., Que, Y.-A., Moreillon, P., Entenza, J. M., & Mancini, S. (2015). Aspirin plus ticlopidine prevented experimental endocarditis due to *Enterococcus faecalis* and *Streptococcus gallolyticus*. *Pathogens and Disease*, 73(8), ftv060. https://doi.org/10.1093/ femspd/ftv060
- Vivekananthan, D. P., Bhatt, D. L., Chew, D. P., Zidar, F. J., Chan, A. W., Moliterno, D. J., Ellis, S. G., & Topol, E. J. (2004). Effect of clopidogrel pretreatment on periprocedural rise in C-reactive protein after percutaneous coronary intervention. *Amercian Journal of Cardiology*, 94(3), 358–360. https://doi.org/10.1016/j.amjcard.2004.04.035
- von Elverfeldt, D., Maier, A., Duerschmied, D., Braig, M., Witsch, T., Wang, X., Mauler, M., Neudorfer, I., Menza, M., Idzko, M., Zirlik, A., Heidt, T., Bronsert, P., Bode, C., Peter, K., & von zur Muhlen, C. (2014). Dual-contrast molecular imaging allows noninvasive characterization of myocardial ischemia/reperfusion injury after coronary vessel occlusion in mice by magnetic resonance imaging. *Circulation*, 130(8), 676–687. https://doi.org/10.1161/CIRCULATIONAHA.113.008157

- Wallentin, L., Becker, R. C., Budaj, A., Cannon, C. P., Emanuelsson, H., Held, C., Horrow, J., Husted, S., James, S., Katus, H., Mahaffey, K. W., Scirica, B. M., Skene, A., Steg, P. G., Storey, R. F., & Harrington, R. A. (2009). Ticagrelor versus clopidogrel in patients with acute coronary syndromes. *New England Journal of Medicine*, 361(11), 1045–1057. https://doi.org/10.1056/NEJMoa0904327
- Wei, P., Han, B., Zhang, W. J., Bai, J., Jiang, C. Y., Qiu, C. R., Zhang, Q., Fu, Q., & Yang, X. J. (2017). Effect of ticagrelor on the serum level of hs-CRP, ESM-1 and short-term prognosis of patients with acute STEMI. Experimental and Therapeutic Medicine, 13(2), 604–608. https://doi.org/10.3892/etm.2016.3987
- West, L. E., Steiner, T., Judge, H. M., Francis, S. E., & Storey, R. F. (2014). Vessel wall, not platelet, P2Y₁₂ potentiates early atherogenesis. *Car-diovascular Research*, 102(3), 429–435. https://doi.org/10.1093/cvr/cvu028
- Weyrich, A. S., McIntyre, T. M., McEver, R. P., Prescott, S. M., & Zimmerman, G. A. (1995). Monocyte tethering by P-selectin regulates monocyte chemotactic protein-1 and tumor necrosis factor-alpha secretion. Signal integration and NF-kappa B translocation. *Journal of Clinical Investigation*, 95(5), 2297–2303. https://doi.org/10.1172/ jci117921
- Winning, J., Neumann, J., Kohl, M., Claus, R. A., Reinhart, K., Bauer, M., & Lösche, W. (2010). Antiplatelet drugs and outcome in mixed admissions to an intensive care unit. *Critical Care Medicine*, 38(1), 32–37. https://doi.org/10.1097/CCM.0b013e3181b4275c
- Winning, J., Reichel, J., Eisenhut, Y., Hamacher, J., Kohl, M., Deigner, H. P., Claus, R. A., Bauer, M., & Lösche, W. (2009). Anti-platelet drugs and outcome in severe infection: Clinical impact and underlying mechanisms. *Platelets*, 20(1), 50–57. https://doi.org/10.1080/ 09537100802503368
- Wiviott, S. D., Braunwald, E., McCabe, C. H., Montalescot, G., Ruzyllo, W., Gottlieb, S., Neumann, F. J., Ardissino, D., de Servi, S., Murphy, S. A., Riesmeyer, J., Weerakkody, G., Gibson, C. M., & Antman, E. M. (2007).
 Prasugrel versus clopidogrel in patients with acute coronary syndromes. New England Journal of Medicine, 357(20), 2001–2015. https://doi.org/10.1056/NEJMoa0706482
- Woodward, M., Lowe, G. D., Francis, L. M., Rumley, A., & Cobbe, S. M. (2004). A randomized comparison of the effects of aspirin and clopidogrel on thrombotic risk factors and C-reactive protein following myocardial infarction: The CADET trial. *Journal of Thrombosis and Haemostasis*, 2(11), 1934–1940. https://doi.org/10.1111/j.1538-7836.2004.01017.x
- Wu, H., Denna, T. H., Storkersen, J. N., & Gerriets, V. A. (2019). Beyond a neurotransmitter: The role of serotonin in inflammation and immunity. *Pharmacological Research*, 140, 100–114. https://doi.org/10.1016/j. phrs.2018.06.015
- Xiao, Z., & Théroux, P. (2004). Clopidogrel inhibits platelet-leukocyte interactions and thrombin receptor agonist peptide-induced platelet activation in patients with an acute coronary syndrome. *Journal of the American College of Cardiology*, 43(11), 1982–1988. https://doi.org/10. 1016/j.jacc.2003.10.071
- Ye, Y., Birnbaum, G. D., Perez-Polo, J. R., Nanhwan, M. K., Nylander, S., & Birnbaum, Y. (2015). Ticagrelor protects the heart against reperfusion injury and improves remodeling after myocardial infarction. Arteriosclerosis, Thrombosis, and Vascular Biology, 35(8), 1805–1814. https://doi. org/10.1161/atvbaha.115.305655
- Yeh, E. T. (2004). CRP as a mediator of disease. *Circulation*, 109(21 Suppl 1), li11–li14. https://doi.org/10.1161/01.Cir.0000129507.12719.80
- Zeraatkar, D., Cusano, E., Martínez, J. P. D., Qasim, A., Mangala, S., Kum, E., Bartoszko, J. J., Devji, T., Agoritsas, T., Guyatt, G., Izcovich, A., Khamis, A. M., Lamontagne, F., Rochwerg, B., Vandvik, P., Brignardello-Petersen, R., & Siemieniuk, R. A. C. (2022). Use of tocilizumab and sarilumab alone or in combination with corticosteroids for covid-19: Systematic review and network meta-analysis. *BMJ Medicine*, 1(1), e000036. https://doi.org/10.1136/bmjmed-2021-000036

- Zhang, F. L., Luo, L., Gustafson, E., Lachowicz, J., Smith, M., Qiao, X., Liu, Y. H., Chen, G., Pramanik, B., Laz, T. M., Palmer, K., Bayne, M., & Monsma, F. J. Jr. (2001). ADP is the cognate ligand for the orphan G protein-coupled receptor SP1999. *Journal of Biological Chemistry*, 276, 8608–8615. https://doi.org/10.1074/jbc.M009718200
- Zhang, J., Zhang, K., Gao, Z. G., Paoletta, S., Zhang, D., Han, G. W., Li, T., Ma, L., Zhang, W., Müller, C. E., Yang, H., Jiang, H., Cherezov, V., Katritch, V., Jacobson, K. A., Stevens, R. C., Wu, B., & Zhao, Q. (2014). Agonist-bound structure of the human P2Y₁₂ receptor. *Nature*, 509(7498), 119–122. https://doi.org/10.1038/nature13288
- Zucker, M. B., Mosesson, M. W., Broekman, M. J., & Kaplan, K. L. (1979). Release of platelet fibronectin (cold-insoluble globulin) from alpha granules induced by thrombin or collagen; lack of requirement for plasma fibronectin in ADP-induced platelet aggregation. *Blood*, 54(1), 8–12. https://doi.org/10.1182/blood.V54.1.8.8
- Zufferey, A., Schvartz, D., Nolli, S., Reny, J. L., Sanchez, J. C., & Fontana, P. (2014). Characterization of the platelet granule proteome: Evidence of the presence of MHC1 in alpha-granules. *Journal of Proteomics*, 101, 130–140. https://doi.org/10.1016/j.jprot.2014.02.008

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