SUPPLEMENTARY MATERIAL

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Research	Population	Intervention	Comparison	Outcome					
questions		(Exposure)							
-									
What is the evidence that interferon measurement is useful in the diagnosis of									
RMDs?									
	Deerste								
	People	IFIN pathway	RIVID VS CONTROL	Association					
	presenting	assay	population	between					
	with any RMD	measurement		measurement of					
	Control			IFN pathway					
	control			activation and					
	population			RMD diagnosis					
	lidence that inte	erteron measure	ment reflects di	sease activity in					
RMDS?									
	People	IFN pathway	Disease	Association					
	presenting	assav	activity	between					
	with any RMD	measurement	measurement	measurement of					
		modeuromont	modouromont	IFN pathway					
				activation and					
				discosso activity					
				(difference					
				between groups,					
				association,					
				correlation)					
What is the ev	vidence that inte	erferon measure	ment is useful f	or the prognosis					
(natural history	v) of clinical stat	tus in RMDs?							
	y) of official stat								
	People	IFN pathway	Disease	Association					
	presenting	assay	exacerbation /	between					
	with any RMD,	measurement	flare vs no	measurement of					
	'at-risk' of		disease	IFN pathway					
	RMDs or in		exacerbation /	activation and					
	preclinical		no flare	disease					
				exacerbation,					
1	1	1	1	- ,					

Supplementary Text 1: Research questions (PICO framework)

	stages			Progressi	on to	progression	to
				clinical	RMD	clinical RMD	or
				(fulfillmen	t of	severity	
				classificat	ion		
				criteria)			
				Severity of	of the		
				clinical c	ourse		
				(occurren	ce of		
				comorbidi	ty or		
				organ			
				involveme	ent or		
				damage)			
What is the ev	idence that inte	erferon	measure	ment is us	seful fo	or the progno	sis
(response to tr	eatment) in RMI	Ds?					
	People	IFN	pathway	Clinical		Association	
	People presenting	IFN assav	pathway	Clinical	to	Association	
	People presenting with any RMD	IFN assay measi	pathway	Clinical response treatment	to	Association between	t of
	People presenting with any RMD starting a new	IFN assay measu	pathway urement	Clinical response treatment	to	Association between measuremen	t of
	People presenting with any RMD starting a new	IFN assay measi	pathway urement	Clinical response treatment	to	Association between measuremen IFN pathy	t of way
	People presenting with any RMD starting a new therapy	IFN assay measi	pathway urement	Clinical response treatment	to	Association between measuremen IFN pathy activation a	t of way and
	People presenting with any RMD starting a new therapy	IFN assay measi	pathway urement	Clinical response treatment	to	Association between measuremen IFN pathy activation a clinical respon	t of way and nse
What is the ev	People presenting with any RMD starting a new therapy	IFN assay measu	pathway urement measure	Clinical response treatment	to spons	Association between measuremen IFN pathy activation a clinical respon	t of way and nse ges
What is the ev with changing	People presenting with any RMD starting a new therapy idence that inte disease status o	IFN assay measu erferon or treat	pathway urement measure ment)?	Clinical response treatment	to spons	Association between measuremen IFN pathy activation a clinical respon	t of way and nse ges
What is the ev with changing	People presenting with any RMD starting a new therapy	IFN assay measu erferon or treat	pathway urement measure ment)?	Clinical response treatment	to spons	Association between measuremen IFN pathy activation a clinical respon	t of way and nse ges
What is the ev with changing	People presenting with any RMD starting a new therapy idence that inte disease status of People	IFN assay measu erferon or treat	pathway urement measure ment)?	Clinical response treatment ment is res Change	to spons in	Association between measuremen IFN pathy activation a clinical respon ive (i.e. chang Responsiven	t of way and nse ges ess
What is the ev with changing	People presenting with any RMD starting a new therapy idence that inte disease status of People presenting	IFN assay measu erferon or treat IFN assay	pathway urement measure ment)? pathway	Clinical response treatment ment is res Change serial	to spons in IFN	Association between measuremen IFN pathy activation a clinical respon ive (i.e. chang Responsiven (change)	t of way and nse ges ess of
What is the ev with changing	People presenting with any RMD starting a new therapy idence that inte disease status of People presenting with any RMD	IFN assay measu erferon or treat IFN assay measu	pathway urement measure ment)? pathway urement	Clinical response treatment ment is res Change serial measuren	to spons in IFN nents	Association between measuremen IFN pathy activation a clinical respon ive (i.e. chang Responsivent (change) serial	t of way and nse ges ess of
What is the ev with changing	People presenting with any RMD starting a new therapy idence that inte disease status of People presenting with any RMD	IFN assay measu erferon or treat IFN assay measu	pathway urement measure ment)? pathway urement	Clinical response treatment ment is res Change serial measuren	to spons in IFN nents	Association between measuremen IFN pathy activation a clinical respon ive (i.e. chang (change) serial measuremen	t of way and nse ges of ts
What is the ev with changing	People presenting with any RMD starting a new therapy idence that inte disease status of People presenting with any RMD	IFN assay measu rferon or treat IFN assay measu	pathway urement measure ment)? pathway urement	Clinical response treatment ment is res Change serial measuren	to spons in IFN nents	Association between measuremen IFN pathy activation a clinical respon ive (i.e. chang (change) serial measuremen of IFN pathy	t of way and nse ges of ts way
What is the ev with changing	People presenting with any RMD starting a new therapy idence that inte disease status of People presenting with any RMD	IFN assay measu rferon or treat IFN assay measu	pathway urement measure ment)? pathway urement	Clinical response treatment ment is res Change serial measuren	to spons in IFN nents	Association between measuremen IFN pathy activation a clinical respon ive (i.e. chang (change) serial measuremen of IFN pathy activation	t of way and nse ges of ts way

Supplementary Text 2: Search strategy for Ovid MEDLINE

- 1 interferon/ (21958)
- 2 (interferon* adj2 (biomarker* or sign*)).ti. (992)
- 3 (interferon* adj2 (biomarker* or sign*)).ab. /freq=2 (246)
- 4 exp interferon type i/ (48691)
- 5 "type 1 IFN".ti. (41)
- 6 "type 1 IFN".ab. /freq=2 (90)
- 7 (type 1 adj3 interferon*).ti. (205)
- 8 (type 1 adj3 interferon*).ab. /freq=2 (98)
- 9 (interferon* adj1 (alpha or alfa)).ti. (12678)
- 10 (interferon* adj1 (alpha or alfa)).ab. /freq=2 (5014)
- 11 (interferon* adj1 beta).ti. (4226)
- 12 (interferon* adj1 beta).ab. /freq=2 (1557)
- 13 or/1-12 [inteferons] (71976)
- 14 Lupus Erythematosus, Systemic/ (53310)
- 15 (systemic adj2 lupus).ti. (28721)
- 16 (systemic adj2 lupus).ab. /freq=2 (4269)
- 17 exp Arthritis, Rheumatoid/ (110425)

18 ((reumat* or rheumat* or psoriatic or juvenile or inflammatory or idiopathic) adj3 (arthrit* or artrit*)).ti. (71763)

19 ((reumat* or rheumat* or psoriatic or juvenile or inflammatory or idiopathic) adj3 (arthrit* or artrit*)).ab. /freq=2 (21982)

- 20 Arthralgia/ (7938)
- 21 arthralgia.ti. (631)
- 22 arthralgia.ab. /freq=2 (888)
- 23 Connective Tissue Diseases/ (6410)
- 24 connective tissue disease*.ti. (3395)
- 25 connective tissue disease*.ab. /freq=2 (1408)
- 26 exp Scleroderma, Systemic/ (20043)
- 27 (scleroderma or systemic sclerosis).ti. (17190)
- 28 (scleroderma or systemic sclerosis).ab. /freq=2 (6774)
- 29 Sjogren's Syndrome/ (12463)
- 30 ((sjogren* or sjoegren or sicca) adj2 syndrome).ti. (9830)
- 31 ((sjogren* or sjoegren or sicca) adj2 syndrome).ab. /freq=2 (3362)
- 32 (spondyloarthropath* or spondylarthropath*).ti. (1514)
- 33 (spondyloarthropath* or spondylarthropath*).ab. /freq=2 (748)
- 34 Spondylitis, Ankylosing/ (14342)
- 35 ankylosing spondylitis.ti. (8361)
- 36 ankylosing spondylitis.ab. /freq=2 (2758)
- 37 Vasculitis/ (12667)
- 38 vasculitis.ti. (11783)
- 39 vasculitis.ab. /freq=2 (9137)
- 40 Antiphospholipid Syndrome/ (7889)
- 41 (antiphospholipid syndrome or APS or APLS).ti. (5112)
- 42 (antiphospholipid syndrome or APS or APLS).ab. /freq=2 (9483)

- 43 Still's Disease, Adult-Onset/ (1280)
- 44 Still's Disease.ti. (1536)
- 45 Still's Disease.ab. /freq=2 (311)
- 46 exp Myositis/ (19181)
- 47 myositis.ti. (4738)
- 48 myostitis.ab. /freq=2 (0)
- 49 (Behcet* adj (disease or syndrome)).ti. (7844)
- 50 (Behcet* adj (disease or syndrome)).ab. /freq=2 (2829)
- 51 ((IgG4* or Immunoglobulin G4*) adj2 (syndrome or disease)).ti. (1246)
- 52 ((IgG4* or Immunoglobulin G4*) adj2 (syndrome or disease)).ab. /freq=2 (1188)
- 53 or/14-52 [RMD] (291413)
- 54 13 and 53 (1855)
- 55 exp animals/ not humans.sh. (4641021)
- 56 54 not 55 (1754)
- 57 remove duplicates from 56 (1744)
- 58 limit 57 to english language (1563)

Supplementary Text 3: Search strategy for Embase

- 1 interferon/ (78376)
- 2 (interferon* adj2 (biomarker* or sign*)).ti. (1447)
- 3 (interferon* adj2 (biomarker* or sign*)).ab. /freq=2 (518)
- 4 alpha interferon/ (54088)
- 5 alpha interferon A/ (289)
- 6 beta interferon/ (24399)
- 7 (interferon* adj1 (alpha or alfa)).ti. (15287)
- 8 (interferon* adj1 (alpha or alfa)).ab. /freq=2 (6077)
- 9 (interferon* adj1 beta).ti. (5808)
- 10 (interferon* adj1 beta).ab. /freq=2 (2295)
- 11 (type 1 adj3 interferon*).ti. (350)
- 12 "type 1 INF*".ab. /freq=2 (86)
- 13 or/1-12 [type 1 interferon] (151212)
- 14 systemic lupus erythematosus/ (96577)
- 15 (systemic adj2 lupus).ti. (40830)
- 16 (systemic adj2 lupus).ab. /freq=2 (6638)
- 17 exp Arthritis, Rheumatoid/ (219451)

18 ((reumat* or rheumat* or psoriatic or juvenile or inflammatory or idiopathic) adj3 (arthrit* or artrit*)).ti. (114993)

19 ((reumat* or rheumat* or psoriatic or juvenile or inflammatory or idiopathic) adj3 (arthrit* or artrit*)).ab. /freq=2 (41573)

- 20 Arthralgia/ (60181)
- 21 arthralgia.ti. (911)
- 22 arthralgia.ab. /freq=2 (1866)
- 23 Connective Tissue Diseases/ (11630)
- 24 connective tissue disease*.ti. (4727)
- 25 connective tissue disease*.ab. /freq=2 (2429)
- 26 Scleroderma, Systemic/ (17314)
- 27 (scleroderma or systemic sclerosis).ti. (26554)
- 28 (scleroderma or systemic sclerosis).ab. /freq=2 (12470)
- 29 Sjogren's Syndrome/ (12089)
- 30 ((sjogren* or sjoegren or sicca) adj2 syndrome).ti. (13295)
- 31 (spondyloarthropath* or spondylarthropath*).ab. /freq=2 (1029)
- 32 Spondylitis, Ankylosing/ (15590)
- ankylosing spondylitis.ti. (13053)
- 34 ankylosing spondylitis.ab. /freq=2 (5036)
- 35 Vasculitis/ (40212)
- 36 vasculitis.ti. (16629)
- 37 vasculitis.ab. /freq=2 (15699)
- 38 Antiphospholipid Syndrome/ (16255)
- 39 (antiphospholipid syndrome or APS or APLS).ti. (7161)
- 40 (antiphospholipid syndrome or APS or APLS).ab. /freq=2 (14445)
- 41 Still's Disease, Adult-Onset/ (1317)
- 42 still's disease.ti. (2021)

- 43 still's disease.ab. /freq=2 (519)
- 44 Myositis/ (16388)
- 45 myositis.ti. (77)
- 46 myositis.ab. /freq=2 (37)
- 47 Behcet Syndrome/ (8519)
- 48 (Behcet* adj2 (syndrome or disease)).ti. (11087)
- 49 (Behcet* adj2 (syndrome or disease)).ab. /freq=2 (4765)
- 50 ((IgG4* or "Immunoglobulin G") adj2 (sydrome or disease)).ti. (1569)
- 51 ((IgG4* or "Immunoglobulin G") adj2 (sydrome or disease)).ab. /freq=2 (1713)
- 52 or/14-51 [RMD] (523688)
- 53 13 and 52 [interferon type 1 and RMD] (9125)
- 54 (exp animal/ or nonhuman/) not exp human/ (7037850)
- 55 53 not 54 (8634)

Supplementary Text 4: Search strategy for Web of Science

Indexes=SCI-EXPANDED, CPCI-S, ESCI Timespan=1900-2019

1 (TS=((interferon* Near/1 (alpha or alfa)))) AND LANGUAGE: (English)

2 (TS=((interferon* Near/1 beta))) AND LANGUAGE: (English)

3 (TS=((type 1 Near/3 interferon*))) AND LANGUAGE: (English)

4 TS=("type 1 INF*") or TS=(interferon* Near/2 biomarker*) or TS=(interferon* Near/2 sign*)

- # 5 #4 OR #3 OR #2 OR #1
- # 6 TS=((systemic Near/2 lupus))

7 TS=(((reumat* or rheumat* or psoriatic or juvenile or inflammatory or idiopathic)Near/3 (arthrit* or artrit*)))

8 TS=(Arthralgia)

9 TS=("connective tissue disease*")

10 TS=((scleroderma or "systemic sclerosis"))

- # 11 TS=(((sjogren* or sjoegren or Sjogren's or sicca) Near/2 syndrome))
- # 12 TS=((spondyloarthropath* or spondylarthropath*))
- # 13 TS=("ankylosing spondylitis")

14 TS=(vasculitis)

- # 15 TS=(("antiphospholipid syndrome" or APS or APLS))
- # 16 TS=("still's disease")

17 TS=(Myositis)

18 TS=((Behcet* Near/2 (syndrome or disease)))

19 TS=(((IgG4* or "Immunoglobulin G") Near/2 (sydrome or disease)))

20 #19 OR #18 OR #17 OR #16 OR #15 OR #14 OR #13 OR #12 OR #11 OR #10 OR #9 OR #8 OR #7 OR #6

21 (#20 AND #5) AND LANGUAGE: (English)

22 (TS=("type 1 INF*") or TS=(interferon* Near/2 biomarker*) or TS=(interferon* Near/2 sign*)) AND LANGUAGE: (English)

- # 24 (#23 AND #20) AND LANGUAGE: (English)
- # 23 #22 OR #3 OR #2 OR #1

Supplementary Text 5: Inclusion and exclusion criteria (first step)

Inclusion criteria

- Subjects: human patients with RMDs
- RMDs: systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), arthritis, juvenile idiopathic arthritis (JIA), arthralgia, Sjögren's syndrome (pSS), Sicca syndrome, dermatomyositis (DM), polymyositis (PM), myositis, connective tissue disease (CTD), scleroderma, systemic sclerosis (SSc), psoriatic arthritis (PsA), ankylosing spondylitis (AS), spondyloarthropathies (SpA), vasculitis, NCA-associated vasculitis (AAV), giant cell arteritis (GCA), antiphospholipid syndrome (APS), Still disease, adult-onset Still disease (AOSD), Behçet disease (BD), IgG4-related disease (IGRD)
- Language: English only
- Study design: longitudinal studies, cross-sectional studies, randomized controlled trials, case-control studies, cohort studies, non-controlled trials, intervention studies
- Samples: blood, serum, plasma studies

Exclusion criteria

- Subjects: animal studies, pre-clinical studies, genetic studies
- Study design: case studies, letters, non-original articles (reviews, editorials, opinion pieces, etc)
- Samples: peripheral tissues or fluids other than blood, serum or plasma
- Papers that do not specify the type of interferon that the assay measures

Supplementary Text 6: Eligibility criteria for clinical research questions

RQ1: What is the evidence that interferon measurement is useful in the diagnosis of RMDs?

- Clinical outcome: RMD diagnosis (validated classification criteria or diagnosis confirmed by a physician)
- Control population
- Study type: cross-sectional studies, diagnostic accuracy studies, case-control studies, cohort studies

RQ2: What is the evidence that interferon measurement reflects disease activity in RMDs?

- Clinical outcome: any disease activity measured by a validated instrument, including CRP or ESR levels, expressed as continuous or dichotomous variable
- Study type: longitudinal studies, randomized control trials, non-controlled trials, cross-sectional studies, case-control studies, cohort studies

RQ3: What is the evidence that interferon measurement is useful for the prognosis (natural history) of clinical status in RMDs?

- Clinical outcomes:
 - Disease exacerbation (pre-defined increase in disease activity)
 - Flare occurrence (by validated flare instruments)
 - Progression to clinical RMD (fulfillment of classification criteria)
 - Severity of the clinical course (occurrence of comorbidity or organ involvement or damage)
- Study type: longitudinal studies, randomized control trials, non-controlled trials, cohort studies

RQ4: What is the evidence that interferon measurement is useful for the prognosis (response to treatment) in RMDs?

- Clinical outcome: clinical response (validated response criteria or pre-defined change in disease active instrument) upon new treatment
- Study type: longitudinal studies, randomized control trials, non-controlled trials

RQ5: What is the evidence that interferon measurement is responsive (i.e. changes with changing disease status or treatment)?

- Clinical outcomes:
 - Change in measurements of IFN pathway activation before and after any treatment
 - Change in measurements of IFN pathway activation before and after disease exacerbation or flare
- Study type: longitudinal studies, randomized control trials, non-controlled trials, cohort studies

Supplementary Text 7: Risk of bias instruments

Research question	Instrument
RQ1	QUADAS-2 [1]
RQ2	JBI analytical cross sectional studies critical appraisal tool [2]
RQ3	QUIPS [3]
RQ4	Cochrane RoB2 [4] QUIPS [3]
RQ5	Cochrane RoB2 [4] JBI case series critical appraisal tool [5]

Supplementary Text 8: Summary and characteristics of assays measuring IFN pathway activation.

Category	Sub-category	Target	Sample source	Readout
Immunoassays	Immunoassay	Soluble protein(s) (IFNa/b proteins, IFN-stimulated proteins, chemokines, etc)	Serum, plasma, or whole blood lysates	Protein(s) concentration (individual or grouped as protein scores)
	DELFIA	Soluble protein(s) (IFNa/b proteins, IFN-stimulated proteins, chemokines, etc)	Serum or plasma	Protein(s) concentration (individual or grouped as protein scores)
	Simoa	Soluble protein(s) (IFNa/b proteins, IFN-stimulated proteins, chemokines, etc)	Serum or plasma	Protein(s) concentration (individual or grouped as protein scores)
	Acid-precipitation immunoassay	Soluble protein(s) (IFNa/b proteins, IFN-stimulated proteins, chemokines, etc)	Serum or plasma	Protein(s) concentration (individual or grouped as protein scores)
	Radioinmmunoassay	Soluble protein(s) (IFNa/b proteins, IFN-stimulated proteins, chemokines, etc)	Serum or plasma	Protein(s) concentration (individual or grouped as protein scores)
Flow cytometry	Flow cytometry to detect IFN-induced proteins	Membrane-bound IFN-stimulated proteins	Whole blood or isolated peripheral blood mononuclear cells (PBMCs)	Protein expression levels
Microarrays	Microarray modules	RNA from subsets of IFN-stimulated genes (usually pre-defined and sharing expression patterns)	RNA isolated from whole blood or PBMCs	Quantitative gene expression (fold enrichment/change) or categorical (number of genes up- and down- regulated, modular expression of gene

				signatures)
	Microarray scores and clustering	RNA from subsets of IFN-stimulated genes (usually pre-defined and sharing expression patterns)	RNA isolated from whole blood or PBMCs	Quantitative gene expression (gene scores)
qPCR	<i>qPCR</i>	RNA levels (IFN-a/b gene(s), IFN-stimulated gene(s), IFN-induced chemokine(s), etc)	RNA isolated from whole blood or PBMCs	Quantitative gene expression (fold enrichment/changes) either individual or composite scores (set of genes)
RNA Seq	RNA Seq	RNA from IFN-gene expression in whole transcriptome expression analysis	RNA isolated from whole blood or PBMCs	Quantitative gene expression or categorical (transcriptomic signature)
Nanostring	Nanostring	RNA from IFN-stimulated genes (pre-defined)	RNA isolated from whole blood or PBMCs	Quantitative gene expression (gene scores)
DNA methylation	DNA methylation arrays	Analysis of DNA methylation of IFN-stimulated genes in whole genome arrays	DNA from whole blood, isolated PBMCs or tissues	Number of genes with differential methylation, either individually or in gene sets
	Bisulfite sequencing	Analysis of DNA methylation in pre-defined IFN- stimulated genes sets	DNA from whole blood, isolated PBMCs or tissues	Number of genes with differential methylation, either individually or in pre-defined sets)
Reporter cell assays	Reporter cell assays by qPCR	IFN-stimulated gene(s) expression by qPCR in an experimental system upon stimulation with biological samples	Serum or plasma	Quantitative gene expression (fold

				enrichment/changes)
	Reporter cell assays (other)	IFN-stimulated gene(s) expression by luminometric or colorimetric in an experimental system upon stimulation with biological samples	Serum or plasma	Quantitative gene expression (fold enrichment/changes)
Cytopathic effect assay	Cytopathic effect assay	Functional (antiviral) effects of IFN in protecting cells from viral infections by analyzing cytopathic features	Serum or plasma	Antiviral activity measured in titres, arbitrary units, etc (quantitative)
Plaque-reducing assay	Plaque-reducing assay	Functional (antiviral) effects of IFN in protecting cells from viral infections by analyzing plaques of cultured cells	Serum or plasma	Measure of plaques of cultured cells (semi- quantitative)
Immunohistochemistry (IHQ)	Immunohistochemistry (IHQ)	IFN-stimulated proteins expression in tissues	Whole blood or other tissues	Protein levels (semi- quantitative)

Supplementary Table 1: Summary of the studies reporting assays to evaluate the potential use of IFN assays in the diagnosis of SLE.

Category	Sub-category	n	Results	Diagnostic	Risk of bias	References
Immunoassays	Immunoassays detecting IFNa protein	21	17/21 studies found higher IFNa levels in SLE vs control populations 4 studies reported frequency of positive patients (from 40 to 72%) using different cut-offs	1 (Zecevic, 2018) AUC 0.517 (0.382-0.653), p=0.807; S: 7.3%, Sp: 96.0%, PPV: 80.0%, NPV: 32.0%	High	[6–26]
	Immunoassays detecting IFNb protein	1	4 SLE patients exhibited high IFNb serum levels, whereas 2 exhibited similar levels to controls	0	High	[9]
	Immunoassays detecting IFN- induced proteins	6	Targets: Mx1 (2), Galectin 3 – binding protein (1), galectin 9 (1), MIP1a (2) Main findings: Increased Mx1 levels in whole blood (lysed) in SLE (2) Increased Galectin 3 serum levels in SLE (1) Increased Galectin 9 – binding protein serum levels in SLE (1) Increased MIP1a serum levels in SLE (1)	0	High	[27–32]
	DELFIA for IFNa	2	<i>Main findings:</i> 2/2 reported IFNa was detected more often in SLE (9/20 and 6/11) samples than controls (0 and 0 HC)	0	High	[33,34]

	DELFIA for IFN-induced proteins	1	Targets sSIGLEC-1 (1) Main findings: sSIGLEC-1 serum levels increased in SLE vs HC	0	High	[28]
	Simoa for IFNa	2	<i>Main findings:</i> 2/2 reported IFNa serum levels higher in SLE than controls	0	High	[35,36]
	Other assays (Acid- precipitation immunoassay (1), RIA (2))	3		0	Unclear/high	[34,37,38]
Flow cytometry	Flow cytometry to detect IFN- induced proteins	6	Targets: SIGLEC1 (3), CD64 (1), ITIM1 (1), Mx1 (1) Main findings: SIGLEC1 expression was higher in monocytes (2) and pDC (1) in SLE vs controls Increased expression of CD64 (1) and Mx1 (1) in monocytes and ITIM1 in platelets (1) in SLE vs controls	0	High	[20,28,31,39–41]
Microarrays	Microarray modules	5	Sources: WB (3), PBMC (1), sorted cell populations (CD4+ and CD8+ T cells, monocytes and neutrophils) (1) Main findings: 5/5 reported IFN-related modules among the most upregulated genes in SLE vs HC M1.2, M3.4, and M5.12 reported in 3/5 Modular IFN signature observed in SLE ranging from 83% to 100% of the patients	0	High	[42–46]

	Microarray	20	Sources:	0	High	[20,39,41,47–63]
	scores and		WB (10), PBMC (4), sorted cell			
	Clustering		monocytes or platelets) (6)			
			Main findings:			
			15/20 reported IFN score and/or ISG			
			expression significantly higher in SLE			
			VS controls			
			ranged from 50 to 81% (reported in			
			5/20)			
			1/20 reported IFN score follow a			
			bimodal distribution (1)			
qPCR	qPCR on	4	Sources:	1 (Yuan, 2018):	High	[6,64–66]
	Individual ISG		VVB (2), PBMC (2)	15G15: AUC		
			I = I = I = I = I = I = I = I = I = I =	0.820 (0.715–		
			Main findings:	p=0.000015), S:		
			Increased expression of IFNA (1),	89.3%, Sp:		
			Mx1 (2) or ISG15 (1) in SLE vs	68.8%		
			controls	4 (5		
	qPCR on	38	Sources: M/R (10) loukoovtos (2) RMC (11)	1 (Feng, 2015):	Unclear/nign	[18,20,29,31,47,56,62,67–
	Several 13G		sorted cell populations (monocytes or	$0.805 \text{ p}=1.4 \cdot 10^{-10}$		96]
			platelets) (5)	5		
			Targets:			
			ISG scores calculated from 2 (1), 3			
			(7), 4 (5), 5 (9), 6 (3), 8 (1), 9 (2), 10			
			(1) of 23 (1) genes; A studies computed 2 scores (GC-			
			A/B. score A/B. type I/II)			
			Main findings:			
			38/38 reported IFN score and/or ISG			
			expression significantly higher in SLE			
			vs controls			
			Differences across ISGs reported in			

			4/38 IFN positive patients ranged from 57 to 64%			
	qPCR on chemokines	1	Sources: Leukocytes (1) Targets: CCL2 and IP-10 (1) Main findings: Increased expression of CCL2 and IP-10 in SLE vs controls	0	High	[72]
Nanostring		1	Sources: WB Targets: 5 ISG Main findings: IFN score higher in SLE vs controls	0	High	[97]
DNA D methylation m ai	DNA methylation arrays	9	Sources: WB (2), PBMC (2), sorted cell populations (CD4+ T-cells, CD19+ B- cells, CD14+ monocytes, neutrophils and low density granulocytes) (5) Main findings: 9/9 reported SLE-specific differentially methylated genes (mostly hypomethylated) related to ISG 6/9 reported ISG-related DMG among those with the largest size effect	1 (Imgenberg- Kreuz, 2019) Hypomethylation (DMG): AUC 0.940	High	[98–106]
	Bisulfite sequencing	3	Sources: WB (2), sorted cell populations (neutrophils) (1) Main findings: 3/3 confirmed SLE-specific hypomethylated sites	1 (Zhao, 2016) Hypomethylation IFI44L: Site 1: AUC 0.968 (0.954-0.981), S: 93.6%; Sp: 96.8%. Site 2: AUC 0.982 (0.972-0.992), S: 94.1%; Sp:	High	[100,102,107]

				98.2%		
Reporter cell assays	Reporter cell assays by qPCR	8	Main findings: 7/8 reported high IFN activity in SLE vs controls 1/8 reported no differences 1/8 reported important differences across genes	0	High	[86,108–114]
	Reporter cell assays (other)	4	Main findings: 4/4 reported higher IFN bioactivity in SLE vs controls 1 reported high IFN activity in 42% of SLE patients	0	High	[115–117]
Cytopathic effect assay		3	Main findings: 3/3 reported higher IFN activity in SLE vs controls	0	High	[118–120]
Plaque- reducing assay		4	Main findings: 4/4 reported higher IFN activity in SLE compared to controls	0	High	[121–124]
IHQ		1	<i>Target:</i> MxA <i>Main findings:</i> Granulocytes and monocytes from SLE patients also stained positively for MxA protein	0	High	[32]

Supplementary Table 2: Summary of the studies reporting assays to evaluate the potential use of IFN assays in the diagnosis of RA.

Category	Sub-category	n assays	Results	Diagnostic statistics	Risk of bias	References
Immunoassays	Immunoassays detecting IFNa protein	2	2/2 studies found higher IFNa levels in RA vs control populations	0	High	[125,126]
	Immunoassays detecting IFNb protein	2	 1/2 reported IFNb marginally detected in juvenile chronic arthritis patients 1/2 reported IFNb serum levels higher in juvenile chronic arthritis patients compared to controls 	0	High	[125,127]
	DELFIA for IFNa	1	<i>Main findings:</i> IFNa was detected more often in RA (5/19) samples than in controls (0)	0	High	[128]
	Other assays (RIA (2))	2	<i>Main findings:</i> 2/2 reported no differences in IFNa serum levels in RA patients vs controls	0	High	[129,130]
Microarrays	Microarray scores and clustering	7	Sources: WB (5), PBMC (1), sorted cell populations (monocytes) (1) Targets: ISG scores ranging from 5 to 43 ISG Main findings: 7/7 reported IFN score and/or ISG expression significantly higher in RA vs HC IFN signature positive/high patients ranged from 22% to 45% (reported in 2/7) IFN signature qualitatively differs between RA and SLE (reported in 2/7)	0	High	[52,57,61,131– 134]
qPCR	qPCR on individual ISG	3	Sources: WB (3) Targets: IFIT1, IFIT2, IFI44L, ISG15, MXA, MXB, EPSTRLI1, RSAD, HERC5, Ly6E, IFI6, IFI35 (1), IFI44, IFI44L, IFI6, and MX1 (2) or IFI35,	0	High	[84,125,135]

			IFIT3, IFI44, IFI44L, OAS1, SIGLEC1 (1) <i>Main findings:</i> 1 reported MX1, MXB, IFIT1 and IFIT2, HERC5, ISG15, LY6E, RSAD2, IFI25 increased in established RA but not in early RA vs controls. No differences for IFI44, IFI6 1 reported IFI44L, IFI6, MX1 and IRF4 increased in RA. No differences in IFI44 1 reported IFI35 increased in RA. No differences in IFI44 (1)			
	qPCR on several ISG	6	Sources: WB (5), PBMC (1) Targets: ISG scores calculated from 3 (1), 4 (1), 5 (2), 7 (1) or 23 (1) genes; 1 study computed 2 scores (GC-A/B) Main findings: 6/6 studies reported IFN score increased in RA (any subgroup) vs controls 1/6 reported IFN score higher in early RA but not in established 1/6 reported IFN score higher in established vs very early IFN positive patients ranged from 13% to 40% (reported in 3/6)	0	Unclear/high	[70,77,109,136– 138]
	qPCR on chemokines	1	Sources: WB Targets: RANTES, MCP-1, CCL19, MIG, IP-10, CXCL11, and IL-8 (1) Main findings: Increased expression of IFN-induced chemokine score in RA vs controls	0	High	[77]
RNA-seq		1	Sources: sorted cell populations (neutrophils) Main findings: Most differently regulated signaling pathway in RA neutrophils was IFN signaling	0	High	[139]

Reporter cell assays	Reporter cell assays by qPCR	2	<i>Main findings:</i> 1/2 reported high IFN activity in 29.7% of RA patients vs 6.3% of controls	0	High	[109,140]
	Reporter cell assays (other)	1	<i>Main findings:</i> 1/1 detected decreased IFN-like substance in RA vs controls (below sensitivity in 39/49 of RA patients)	0	High	[130]
Cytopathic effect assay		1	<i>Main findings:</i> Higher OAS activity in RA inactive vs active patients and vs controls	0	Unclear/high	[141]
Plaque- reducing assay		3	Main findings: 3/3 reported higher IFN activity in RA vs controls	0	High	[121,122,142]

Supplementary Table 3: Summary of the studies reporting assays to evaluate the potential use of IFN assays in the diagnosis of pSS.

Category	Sub-category	n assays	Results	Diagnostic statistics	Risk of bias	References
Immunoassays	Immunoassays detecting IFNa protein	4	4/4 reported higher IFNa serum levels in pSS vs controls	0	High	[97,143–145]
	Immunoassays detecting IFNb protein	1	IFNb detected in most samples, increased in pSS but not significant	0	High	[146]
	Immunoassays detecting IFN-induced proteins	4	Targets: MCP-1, MIP-1α, MIP-1β, MIG and IP-10 (1), Mx1 (1), IFI16 (1), IP-10 (1) Main findings: 4/4 increased IFN-induced protein serum levels in pSS vs controls	0	Unclear/high	[143,147–149]
	DELFIA for IFNa	1	IFNa was detected more often in pSS (3/38) samples than HC (0 and 0 HC), but low levels in all three patients	0	High	[128]
	Other assays (RIA (1))	1	Circulating IFNa lower in pSS vs controls	0	High	[150]
Flow cytometry	Flow cytometry to detect IFN-induced proteins	2	Targets: SIGLEC1 (1), Mx1 (1) Main findings: SIGLEC1 (1/1) and Mx1 (1/1) expression were higher in monocytes in pSS vs controls Mx1 expression was correlated to IFN score (1/1)	0	High	[148,149]
Microarrays	Microarray scores and clustering	3	Sources: WB (1), PBMC (1), sorted cell populations (CD14+/low monocyte subsets) (1) Targets: ISG scores ranging from 9 to 128 ISG Main findings: 3/3 found IFN-related genes among the most differentially expressed genes	0	High	[146,151,152]

qPCR	qPCR on individual	3	Sources:		High	[144,146,152]
	ISG		(1) WB (2), sorted cell populations (monocytes)			
			Targets:			
			IFNĂ (2) or IFI27 (1)			
			Main findings:			
			Increased expression of IFNA (2/3) or IFI27			
			(1/3) in pSS vs controls			
			Increased expression of IFNA in a fraction of			
	a DCD on soveral ISC	7	pSS patients (5/23 patients)	0	High	[07.00.440.450
	GPCR on several ISG	1	WB(4) $BBMC(2)$ sorted cell populations	0	піgn	[07,89,148,153-
			(monocytes) (1)			156]
			Targets:			
			ISG scores calculated from $3(1), 5(5), 6(1)$;			
			1 study computed 2 scores (type I/II)			
			Main findings:			
			7/7 reported IFN score significantly higher in			
			pSS vs controls			
			IFN positive patients ranged from 51 to 70%			
			(reported in 5/8)		Llinh	[00.445]
RNA-seq		2	Sources:		High	[83,145]
			Colles)			
			Main findings:			
			2/2 reported several IFN-related genes among			
			those differentially expressed between pSS			
			and controls			
Nanostring		3	Sources:	0	High	[83,97,145]
			WB (2), sorted cell populations (CD19+ B-			
			cells)			
			largets: several ISG			
			2/2 confirmed IEN related gappe increased in			
			nSS ve controle			
			1/3 reported IFN score (5 genes) did not			

			differ between pSS and controls			
DNA methylation	DNA methylation arrays	2	Sources: WB (1), sorted cell populations (CD4+ T-cells) Main findings: 1/2 differentially methylated sites related to IFN pathway 1/2 extensive hypomethylation in IFN-related genes (STAT1, IFI44L, IFITM1 and USP18)	1 (Imgenberg- Kreuz, 2018) AUC=0.910	High	[103,157]
Reporter cell assays	Reporter cell assays by qPCR	3	<i>Main findings:</i> 3/3 reported high IFN activity in pSS patients vs controls	0	High	[111,146,158]
	Reporter cell assays (other)	1	<i>Main findings:</i> 1/1 increased IFN activity in pSS vs controls	0	High	[117]
Plaque- reducing assay		2	<i>Main findings:</i> 2/2 reported higher IFN activity in pSS vs controls	0	Unclear/high	[121,122]

Supplementary Table 4: Summary of the studies reporting assays to evaluate the potential use of IFN assays in the diagnosis of SSc.

Category	Sub-category	n assays	Results	Diagnostic statistics	Risk of bias	References
induced protein	Immunoassays detecting IFN- induced proteins	4	<i>Targets:</i> 93 proteins (1), CXCL10, CXCL11, CCL10 and CCL8 (1), IP-10 and I-TAC (1), IP-10, MCP-1, MIP1a and RANTES (1) <i>Main findings:</i> 1/4 reported increased IP10 and I-TAC in SSc vs controls 1/4 reported CXCL10 and CXCL11 increased in SSc vs controls 1/4 reported increased IP-10, MCP-1 and MIP1a in SSc vs controls 1/4 reported 37 proteins higher in SSc vs controls (including MCP1, MIP1b, RANTES, MIP3b, MIG, IP10, I-TAC, and MIF) IFN chemokine score: higher in SSc vs controls (39.2% SSc positive in IFN inducible score)	0	High	[159–162]
	DELFIA for IFNa	1	<i>Main findings:</i> 33 and 23 out of 79 SSc sera induced IFNa when combined with necrotic or apoptotic material, higher compared to controls	0	High	[159]
Flow cytometry	Flow cytometry to detect IFN- induced proteins	1	Targets: SIGLEC1 (1) Main findings: SIGLEC1 expression was higher in monocytes from SSc vs controls (almost absent) SIGLEC1 was highly expressed in SSc patients with high IFN score	0	High	[163]

Microarrays	Microarray modules	1	Sources: WB Main findings: IFN-related (M1.2 and M3.4) and one neutrophil module (M5.15) were the only statistically significant upregulated modules in SSc vs controls	0	High	[164]
	Microarray scores and clustering	9	Sources: WB (6), PBMC (2), sorted cell populations (monocytes) (1) Targets: ISG scores ranging from 9 to 129 ISG Main findings: 4/9 reported a variable number of IFN- related genes among those genes differentially expressed between SSc vs controls 4/9 reported higher IFN scores in SSc vs controls 1/9 reported some IFN-related genes among those genes differentially expressed between IcSSc vs controls 67% IFN positive patients in SSc (reported in 1/10) IFN positive prevalence varied across disease phenotypes (RP/nc/lc/dc) from 33.3% to 100% (reported in 1/9)	1 (Brkic, 2015) IFN score AUC: 0.823, S: 0.667, Sp: 0.881	Unclear/high	[41,47,52,57,160,163,165– 167]
qPCR	qPCR on individual ISG	5	Sources: WB (2), PBMC (3) Targets: MxA (1), SIGLEC (2), IRF7, G1P3 and S100A8 (1), or IFNA, IFNB, IRF7 OAS, MxA and 6-16 (1) Main findings: 1/5 reported increased expression of MxA in 9/50 SSc vs controls	0	High	[163,166–169]

			 2/5 reported increased expression of SIGLEC1 in SSc vs controls 1/5 reported increased expression of IRF7, G1P3 and S100A8 in SSc vs controls 1/5 reported increased expression of IFNA, IFNb and OAS in SSc vs controls 			
	qPCR on several ISG	7	Sources: WB (6), sorted cell populations (monocytes) (1) Targets: ISG scores calculated from 3 (1), 5 (4), 6 (1), or 11 (1) ISGs Main findings: 6/7 reported IFN score significantly higher in SSc vs controls 1/7 reported IFN score elevated in 2/13 SSc patients	0	High	[47,83,89,165,170,171]
Nanostring		2	Sources: WB (2) Main findings: 1/2 IFN score in 2/13 SSc patients 1/2 IFN score (5 genes) did not exhibit differences between SSc and controls	0	High	[83,97]
RNA-seq		1	Sources: sorted cell populations (monocytes) (1) Main findings: 4/99 IFN-related InRNA were upregulated in SSc vs controls		High	[161]
DNA methylation	DNA methylation arrays	1	Sources: Sorted cell populations (CD4+ and CD8+ T-cells) (1) <i>Main findings:</i> Two IFN pathways were the most significantly enriched among	0	High	[172]

			hypomethylated regions			
Reporter cell assays	Reporter cell assays by qPCR	1	Main findings: High IFN score in SSc (9/13) vs controls (0/13)	0	High	[173]
	Reporter cell assays (other)	2	Main findings: 2/2 IFN bioactivity did not differ between SSc vs controls	0	High	[117]
Plaque- reducing assay		3	Main findings: 2/3 IFN bioactivity increased in SSc vs controls 1/3 IFN bioactivity did not differ in SSc vs controls	0	Unclear/high	[121,122,169]

Supplementary Table 5: Summary of the studies reporting assays to evaluate the potential use of IFN assays in the diagnosis of PM/DM.

Category	Sub-category	n assavs	Results	Diagnostic statistics	Risk of bias	References
Immunoassays	Immunoassays detecting IFNa protein	4	<i>Main findings:</i> 2/4 reported higher IFNa serum levels in PM/DM vs controls 1/4 reported higher IFNa serum levels in a subset of patients (anti-MDA5 positive) 1/4 IFNa serum levels did not differ between PM/DM vs controls	0	High	[97,174–176]
	Immunoassays detecting IFNb protein	3	Main findings: 2/3 IFNb serum levels elevated in PM/DM vs controls 1/3 IFNb serum levels did not differ between PM/DM and controls	0	High	[175,176]
	Simoa for IFNa	1	<i>Main findings:</i> IFNa was increased in PM/DM vs controls	0	High	[36]
Microarrays	Microarray scores and clustering	6	Sources: WB (5), PBMCs (1) Targets: ISG scores ranging from 5 to 43 ISGs Main findings: 6/6 reported IFN-related genes among the most differentially expressed genes / largest fold change 1/6 reported signature in 10/12 PM/DM patients	0	High	[48,52,57,175,177,178]
qPCR	qPCR on individual ISG	3	Sources: WB (2), PBMCs (1) Targets: MXA (1), IRF7, ISG15 and MXA (1) or IFIT1 and MXA (1) Main findings:	0	High	[174,178,179]

			 1/3 reported increased MXA expression in PM/DM 1/3 reported increased expression of IRF7, ISG15, MXA in PM/DM 1/3 reported increased expression of IFIT1 and MXA in PM/DM vs controls 			
	qPCR on several ISG	7	Sources: WB (6), PBMC (1) Targets: ISG scores calculated from 3 (1), 6 (2), 8 (1), 13 (1), 23 (1); 1 study computed 2 scores (GC-A/B) Main findings: 6/7 reported IFN score elevated in PM/DM vs controls 1/7 reported IFN positive patients in 2/8 PM/DM patients 1/7 IFN score in PM/DM is predominant GC-A (SLE-like)	1 (Feng, 2015): IFN score: AUC 0.805, p=1.4·10 ⁻⁵	High	[36,73,83,136,177,180]
RNA-seq		2	Sources: WB (1), sorted cell populations (B-cells) Main findings: 1/2 reported IFN-related genes among the most significantly expressed genes between PM/DM vs controls 1/2 validated microarrays results	0	High	[83,181]
Nanostring		2	Sources: WB (2) Main findings: 1/2 IFN score increased in PM/DM patients (100%) vs controls 1/2 IFN score in 2/8 PM/DM patients	0	High	[83,97]
Reporter cell assays	Reporter cell assays by qPCR	3	Main findings: 1/3 reported high IFN activity in PM/DM patients 1/3 reported no increased IFN activity in	0	Unclear/high	[182–184]
		PM/DM (most did not induce up- regulation of the ISG, 2 induced high IFNa activity, and 5 induced low activity (+)) vs controls 1/3 high IFN bioactivity was dependent on IFN score status in PM/DM patients				
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Reporter cell assays (other)	1	<i>Main findings:</i> Increased IFN activity present in 54% (13/24) of DM, 67% (8/12) of PM and 5% (1/22) of myositis samples	0	Unclear/high	[175]	

Supplementary Table 6: Summary of the studies reporting assays analyzing associations between IFN assays and disease activity in SLE.

Category	Sub-category	Reference	n	Disease activity measure	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa	Abdel Galil, 2018 [6]	126	SLEDAI-2K	Positive correlation (r=0.640, p<0.001)	High
	protein	Baechler, 2003 [49]	48	Number of SLE disease criteria	Positive correlation (p<0.0002)	High
		Becker- Merok, 2013 [7]	87	SLEDAI	No association (p=0.070)	High
		Bengtsson, 2000 [185]	30	SLEDAI	Positive correlation (r=0.479, p=0.007)	High
		Fernández- Matilla, 2000 [19]	142	SELENA- SLEDAI CRP ESR	No association	Unclear
		Fragoso- Loyo, 2012 [8]	34	SLEDAI-2K	Positive correlation (r=0.330, p=0.050)	High
		Jonsen, 2003 [186]	57	SLEDAI	No association	High
		Kim, 1987 [38]	30	Clinical activity score	Positive correlation (r=0.600, p<0.010)	High
		Mathian, 2019 [187]	68	SELENA- SLEDAI	Positive association (high SELENA-SLEDAI score: AUC 0.830; Sensitivity: 68.7 (57.6–78.4); specificity: 85.1 (74.3–92.6); PPV: 85.1; NPV 68.7)	High
		Mathian, 2019b [35]	254	Remission (SELENA- SLEDAI)	Positive association (abnormal IFN levels associated with no remission (OR 11.4, p<0.0001))	High
		Oke, 2017 [11]	261	SLEDAI	No association	Unclear

	Postal, 2012 [12]	57	SLEDAI	Positive correlation (r=0.430, p=0.012)	High
	Rodero, 2017 [36]	72	SLEDAI ESR CRP	SLEDAI: positive association (higher SLEDAI in patients with high IFN (p<0.001) ESR: positive association (higher ESR in patients with high IFN (p<0.010) CRP: no association (ns)	Low
	Rose, 2013 [188]	79	mSLEDAI BILAG-2004	mSLEDAI: positive correlation (r=0.4359, p<0.0001) BILAG-2004: positive correlation (r=0.4476, p<0.0001)	Low
	Rose, 2017 [189]	26	BILAG-2004	Positive correlation (r=0.420, p=0.0022)	High
	Schneider, 2015 [190]	172	SLEDAI SLICC	SLEDAI: positive correlation (r=0.219, p=0.004) SLICC: no association (r=0.063, p=ns)	Low
	Shi, 1987 [37]	47	Disease activity stages	Positive association (81% samples of patients with high disease activity were positive compared to 10% samples positive during quiescent stages; p<0.005)	High
	Willis, 2012 [14]	35	SLAM-R	Positive correlation (r=0.314, p=0.054)	High
	Yin, 2014 [16]	79	SLEDAI CRP	SLEDAI: no correlation (r=0.129, p=0.264) CRP: no correlation (r=0.183, p=0.126)	Unclear
	Hashad, 2012 [21]	52	SLEDAI	Positive correlation (p=0.030)	High
	Ma, 2012 [23]	37	SLEDAI	No association (active SLE vs inactive SLE: p=0.690; active SLE vs HC: p=0.098; inactive SLE vs HC: p=0.077)	High
	Mandal, 2014 [24]	129	SLEDAI	Positive correlation (r=0.260, p=0.002)	High
	Robak, 2004 [26]	36	SLAM	No association	High
	Oke, 2019 [191]	497	SLEDAI SDI SLAM	SLEDAI: no association (high IFNa: OR 0.9 (0.5-1.9), p=ns) SDI: no association (high IFNa: OR 0.8 (0.4-1.6), p=ns) SLAM: no association (high IFNa: OR 0.8 (0.4-1.6), p=ns)	Unclear
Immunoassays detecting IFNb protein	Munroe, 2017 [192]	13	SELENA- SLEDAI	Positive correlation (p<0.0001)	High

Immunoassa detecting IFN induced proteins	Immunoassays detecting IFN- induced proteins	Oliveira, 2018 [28]	34	SLEDAI	Positive correlation (sSIGLEC: r=0.100, p=0.070)	High
	,	Zhuang, 2005 [18]	66	SLEDAI	No association (Mx1: ns)	High
		Bauer, 2006 [193]	30	SLEDAI SLAM-R ESR	SLEDAI: positive correlation (MCP-2: r=0.520, p<0.010; MCP-1: r=0.350, p<0.050; I-TAC: r=0.420, p<0.050; IP-10: r=0.370, p<0.050; MIP-3B: p<0.001; MIP-1A: r=0.590, p<0.010); no association (MIG: p=ns) SLAM-R: positive correlation (MCP-2; r=0.600, p<0.010; MCP-1: r=0.390, p<0.050; MIG: r=0.480, p<0.010; I-TAC: r=0.670, p<0.0001; IP-10: r=0.500, p<0.010; MIP-3B: r=0.57, p<0.001; MIP-1A: r=0.680, p<0.001) ESR: positive correlation (MCP-2: r=0.620, p<0.010; MCP-1: r=0.540, p<0.050; MIG: r=0.560, p<0.010; I-TAC r=0.700, p<0.0001; IP-10: r=0.560, p<0.001; MIP-3B: r=0.330, p<0.050; MIP-1A: r=0.600, p<0.001)	Unclear
		Bauer, 2009 [194]	267	SLEDAI	Positive association (chemokine score: SLEDAI was elevated in chemokine-high patients compared to chemokine-intermediate ($p=3.8\times10^{-5}$) and chemokine-low patients ($p=1.0\times10^{-7}$).	High
		Casey, 2018 [90]	304	SLEDAI-2K CLASI	SLEDAI-2K: positive correlation (27 proteins were significantly correlated with SLEDAI-2K and significantly higher in high SLEDAI-2K vs low SLEDAI-2K) CLASI: positive correlation (7 proteins were significantly correlated with CLASI and showed higher concentrations in patients with high CLASI)	Unclear
		Connelly, 2016 [195]	151	Adjusted mean SLEDAI	Positive association (high IFN chemokine score scores were significantly associated with high disease activity (OR [95% CI]: 2.56 [1.03, 6.37]; p=0.040)	High
		Connelly, 2018 [196]	109	SLEDAI-2K	Positive correlation (IFN chemokine score: r=0.730, (95% CI: 0.12, 1.43), p=0.020)	High
		Lee, 2016 [197]	30	SLEDAI	Positive association (IFN chemokine score high patients exhibited higher SLEDAI compared to IFN low group, p=0.0006)	High

		Munroe, 2017 [192]	13	SELENA- SLEDAI	Positive correlation (MCP1/CCL2: p=0.0203; MCP3/ccl7: p=0.0022; MIP1ab/CCL4: p=0.0388; CXCVL8/IL8: p=0.0034; MIG/CXCL: p=0.0159; IP10/CXCL10: p=0.0298)	High
		Nielsen, 2014 [27]	70 SLE- DK 68 SLE- SE 26 SLE- IFNa	SLEDAI SLICC	SLEDAI: positive association (Galectin 3-binding protein: SLE-DK: r=0.410 p=0.0004; SLE-IFNa: r=0.420 p=0.030); no association (Galectin 3-binding protein: SLE-SE: r=-0.0025 p=0.980) SLICC: no association (Galectin 3-binding protein: SLE- DK: r=-0.08 p=0.530; SLE-SE: r=0.036 p=0.770)	Low
		Rose, 2013 [188]	79	mSLEDAI-2K BILAG-2004	mSLEDAI-2K: positive correlation (IP-10: r=0.335, p=0.001; SIGLEC1: r=0.434, p<0.001) BILAG-2004: positive correlation (IP-10: r=0.385, p<0.001; SIGLEC1: r=0.540, p<0.001)	Low
		Rose, 2017 [189]	26	BILAG-2004	Positive correlation (IP-10: r=0.500, p=0.002)	High
		Van den Hoogen, 2018 [87]	50	SLEDAI	Positive correlation (Galectin-9: r=0.320, p=0.003)	High
		Wahadat, 2018 [31]	23	SELENA- SLEDAI	No association (MxA: ns)	High
Flow Fride Cytometry to in price for the second sec	Flow cytometry to detect IFN- induced proteins	Biesen, 2008 [41]	25	SLEDAI	Target (source): Siglec1 (monocytes) Main findings: positive correlation: Siglec1–positive inflammatory monocytes and Siglec1–positive resident monocytes both correlated with SLEDAI (p=0.006 and p=0.005, respectively)	High
		Li, 2010 [39]	108	SLEDAI	<i>Target (source):</i> CD64 (monocytes) <i>Main findings:</i> positive correlation (r=0.3023, p=0.0017)	High
		Oliveira, 2018 [28]	34	SLEDAI	<i>Target (source):</i> Siglec1 (monocytes) <i>Main findings:</i> positive correlation (r=0.100, p=0.070)	High
		Rose, 2013 [188]	79	mSLEDAI-2K BILAG-2004	Target (source): Siglec1 (monocytes) Main findings: mSLEDAI-2K: positive correlation (r=0.434, p<0.0001); BILAG-2004: positive correlation	Low

					(r=0.5409, p<0.0001)	
		Wilhem, 2016 [40]	22	SLEDAI	<i>Target (source):</i> Siglec1 (pDCs), Siglec1 (monocytes) <i>Main findings:</i> positive correlation (Siglec1 (pDCs): r=0.550, p=0.008; Siglec1 (monocytes): r=0.620, p=0.003)	High
Microarrays	Microarray modules	Chiche, 2014 [46]	62	SELENA- SLEDAI	Source: WB Main findings: significant correlation was observed between expression of M3.4 and M5.12 modules and the presence of a flare	High
		Mackay, 2016 [44]	43	SLEDAI	Source: WB Main findings: no association between SLEDAI and IFN modules	High
	Microarray scores and clustering	Assassi, 2010 [47]	74	SLAM-R	Source: WB Main findings: positive correlation (IFN score: p=0.010)	High
		Baechler, 2003 [49]	48	Number of SLE criteria	Source: PBMC Main findings: positive association (IFN-high group showed higher number of SLE criteria compared to IFN- low group, p=0.004)	High
		Baechler, 2007 [48]	12	SLEDAI	Source: WB Main findings: positive association (SLEDAI was higher in IFNhigh compared to IFNIow, p=0.050)	High
		Bauer, 2006 [193]	81	SLEDAI SLAM-R ESR	Source: WB Main findings: SLEDAI: positive correlation (IFN score: r=0.430, p<0.050) SLAM-R: positive correlation (IFN score: r=0.680, p<0.0001) ESR: positive correlation (IFN score: r=0.520, p<0.010)	Low
		Higgs, 2011 [52]	261	SLEDAI BILAG-2004	Source: WB Main findings: SLEDAI: positive association (IFN score higher in SLEDAI>8 compared to SLEDAI<8, p<0.0001) BILAG-2004: positive association (IFN score higher in BILAG>8 compared to BILAG<8, p=0.0006)	Unclear
		Kawasaki, 2011 [198]	12	SLEDAI (active vs inactive)	Source: sorted cell populations (T-cells) Main findings: positive association (IFN-related genes were the most frequently annotated among genes differing	High

					between active and inactive SLE patients)	
		Kennedy, 2015 [63]	61 60 135 80 238	BILAG SELENA- SLEDAI CLASI	Source: PBMC Main findings: BILAG, SELENA-SLEDAI: no associations (ns) CLASI damage score: positive association (IFN score high patients exhibited higher CLASI score compared to IFN score low patients, p=0.0209) CLASI activity score: no associations (ns)	High
		Lauwerys, 2013 [58]	27	SLEDAI-2K	Source: WB Main findings: no associations (ns)	High
		Petri, 2009 [199]	66	SLEDAI	Source: WB Main findings: positive association (high IFNr group patients exhibited greater SLEDAI scores, p=2.3.10 ⁻⁴)	High
		Becker, 2013 [50]	15	SLEDAI	Source: sorted cell populations Main findings: positive associations (a number of IFN- related differentially expressed genes was observed between active or inactive disease in B, T and myeloid cells)	High
		Bennet, 2003 [131]	30	SLEDAI	Source: WB Main findings: positive association (10 IFN-related genes correlated with SLEDAI, all p<0.050)	High
		Biessen, 2008 [41]	47	SLEDAI	Source: sorted cell populations (monocytes) Main findings: positive association (IFN-positive patients had higher SLEDAI than IFN-negative patients, p<0.010)	High
		Nikpour, 2008 [200]	269	SLEDAI-2K	Source: WB Main findings: positive association (IFN-positive patients had higher SLEDAI-2K than IFN-negative patients, p<0.001)	High
		Sharma, 2015 [201]	42	SLEDAI	Source: sorted cell populations (CD4+ T-cells, CD8+ T- cells, CD14+ monocytes and CD20+ B-cells) Main findings: no association (ns)	High
qPCR q ii	qPCR on individual ISG	Abdel Galil, 2018 [6]	126	SLEDAI	Source: WB Number of ISG: 1 Main findings: IFNA expression higher in the highly active group than in the mild and moderately active groups	High
		Li, 2009	39	SLEDAI	Source: PBMC	High

	[65]			Number of ISG: 1 Main findings: positive correlation (MXA: r=0.4814, p=0.0019)	
	Yuan, 2018 [66]	28	SLEDAI	Source: WB Number of ISG: 1 Main findings: positive correlation (ISG15: r=0.520, p=0.0059)	High
	Rodríguez- Carrio, 2019 [84]	75	SLEDAI	Source: WB Number of ISG: 4 Main findings: no associations (any ISG)	High
	Tang, 2008 [85]	144	SLEDAI-2K	Source: PBMC Number of ISG: 1 Main findings: positive correlation (LY6E: r=0.300, p<0.010)	High
	Zhuang, 2005 [18]	88	SLEDAI	Source: WB Number of ISG: 1 Main findings: no association (Mx1)	High
	Kawasaki, 2011 [198]	12	SLEDAI	Source: sorted cell populations (T-cells) Number of ISG: 6 Main findings: positive association (IFI35, JAK1, STAT1, IFITM1, JAK2, STAT2 expression was higher in active phase of SLE vs non active phase, all p<0.05)	High
qPCR for IFN scores	Assassi, 2010 [47]	17	SLAM-R	Source: WB Number of ISG: 3 Main findings: positive correlation (IFN score: p=0.010)	High
	Braunstein, 2012 [68]	30	CLASI	Source: PBMC Number of ISG: 5 Main findings: positive correlation (IFN score: r=0.550, p=0.001)	Low
	Dominguez- Gutierrez, 2014 [71]	103	SLEDAI: active (SLEDAI >4) or inactive (SLEDAI ≤4)	Source: leukocytes Number of ISG: 3 Main findings: no associations	High
	El-Sherbiny, 2018 [74]	114	BILAG Overall, BILAG Skin, BILAG MSK.	Source: WB Number of ISG: 31 (2 scores) Main findings: positive associations depended on organ	Unclear

			BILAG Haem	involvement: IFN score A/IFN score B: OR (90% CI), p Mucocutaneous: 1.71 (1.02-2.86), p=0.042 / 1.41 (1.08- 1.84), p=0.012 Musculoskeletal: 0.79 (0.46-1.34), p=0.381 / 0.82 (0.63- 1.08), p=0.165 Haematological: 1.90 (1.11-3.24), p=0.020 / 1.44 (1.09- 1.90), p=0.012	
	Feng, 2017 [75]	35	SLEDAI SLICC	Source: WB Number of ISG: 5 Main findings: SLEDAI: positive correlation (IFN score: r=0.690, p=0.001) SLICC: no associations	Low
	Feng, 2015 [76]	69	SLEDAI	Source: WB Number of ISG: 5 Main findings: positive correlation (IFN score: r=0.310, p=0.014)	Low
	Feng, 2006 [92]	48	SELENA- SLEDAI	Source: WB Number of ISG: 5 Main findings: positive association (IFN score: higher in active disease (SLEDAI 5–12) or severe disease activity (SLEDAI >12) vs inactive or mild disease activity (SLEDAI 0-4): p=0.009 and p=0.002)	High
	Fu, 2008 [77]	68	SLEDAI	Source: WB Number of ISG: 5 Main findings: positive correlation (IFN score: p=0.023)	High
	Kennedy, 2015 [63]		SELENA- SLEDAI CLASI	Source: WB Number of ISG: 3 Main findings: SELENA-SLEDAI: no associations CLASI damage score: positive association (higher in IFNhigh vs IFNIow patients, p=0.020) CLASI activity score: no associations	Low
	Kirou, 2005 [93]	73	SLEDAI-2K score SLEDAI-2K	Source: PBMC Number of ISG: 3 Main findings:	Unclear

				truncated	SLEDAI-2K: positive association (higher in IFNhigh vs	
				ESR	IFNIow, p=0.003)	
				Number of SLE	SDI: positive association (higher in IFNhigh vs IFNlow,	
				criteria	p=0.0009)	
				SDI	ESR: positive association (higher in IFNhigh vs IFNlow,	
					p=0.0105)	
					Number of ACR criteria: positive association (higher in	
					IFNhigh vs IFNIow, p=0.001)	
		Landolt-	94	SLEDAI-2K	Source: PBMC	High
		Maricortena.			Number of ISG: 5	
		2009 [80]			Main findings: positive correlation (r=0.253, p=0.0318)	
		Liu 2018	44	SI EDAI	Source: PBMC	High
		[81]		OLLD/ (Number of ISG: 4	i ngri
					Main findings: positive association (IENbigh patients	
					showed higher SI EDAL vs IENIow, $p<0.050$)	
		Wahadat	23	SELENA-	Source: sorted populations (monocytes)	High
		2018 [31]	20	SLEDAL	Number of ISG: 6	light
		2010[31]		OLLDAN	Main findings: no association	
		Merrill 2017	98		Source: WB	Unclear
		[202]	50		Number of ISG: 1 score	Uncical
				DILAO	Main findings:	
					SI EDAI: positive association (IENhigh patients showed	
					higher SI EDAL vs IENIow p=0.009)	
					BILAG: (IENhigh patients showed higher SLEDAL vs	
					PENION p=0.040	
		Sharma	42	SI EDAI	Source: sorted populations (monocytes)	Hiah
		2015 [201]	12	OLLD/ (Number of ISG: 46	i ngri
					Main findings: no association	
	aPCR for IFN-	Fu. 2008	68	SLEDAI	Source: WB	Hiah
	induced	[77]			Number of ISG: 12	
	chemokines				Main findings: positive correlation (IFN chemokine score	
					r=0.340, p<0.005)	
DNA	DNA	Coit, 2013	18	SLEDAI	Source: sorted populations (CD4+ T-cells)	High
methylation	methylation	[98]			Main findings: no association	
	arrays	Joseph.	57	SLEDAI	Source: PBMC	Hiah
		2019 [99]	-		Main findings: positive association (higher	
			1	1		

					hypomethylation in several ISG in active SLE (SLEDAI>6) vs controls, but no differences between inactive SLE (SLEDAI<6) and controls	
		Absher, 2013 [101]	49	SLEDAI	Source: sorted populations (CD4+ T-cells) Main findings: no association	High
	Pyrosequencing	Zhao, 2016 [107]	30	SLEDAI	Source: WB Main findings: positive association (higher methylation in IFI44L promoter in remission vs patients with active disease, p<0.001 and p=0.036)	High
Reporter cell assays	Reporter cell assays by	Andrade, 2015 [108]	28	SLEDAI	Assay: Mx1 expression Main findings: no association	High
	qPCR	Nielsen, 2014 [27]	70 SLE- DK 68 SLE- SE 26 SLE- IFNa	SLEDAI SLICC	Assay: Mx1 expression Main findings: no association	High
Reas		Oke, 2019 [191]	497	SLEDAI SDI SLAM	Assay: MX1, EIF2AK2 and IFIT1 expression Main findings: SLEDAI: positive correlation (r=0.3, p<0.0001) SDI: negative correlation (r=-0.13, p<0.0001) SLAM: positive correlation (r=0.3, p<0.0001)	Low
	Reporter cell assays (other)	Dall'era, 2005 [116]	64	SLEDAI ESR	Assay: luciferase/ chemoluminiscence Main findings: SLEDAI: positive correlation (IFN activity: r=0.451, p<0.001) ESR: positive correlation (IFN activity: r=0.481, p<0.001)	Low
		Kato, 2018 [117]	54	SLEDAI	Assay: luciferase/ chemoluminiscence Main findings: positive correlation (IFN bioactivity: r=0.3034, p=0.0185; ISG-inducing activity: r=0.407, p=0.0009)	High
Cytopathic effect assay		Hervier, 2011 [120]	54	SLEDAI	Positive association (increased IFN activity in patients with active SLE vs inactive, p=0.004)	High

	Mathian, 2019 [187]	96	SELENA- SLEDAI	Positive association (IFN bioassay identified high disease activity: AUC 0.790; Sensitivity: 62.7 (51.3–73.0); Specificity: 94.0 (85.4–98.4); PPV: 92.9; NPV: 67.0)	High	
		Rich, 1986 [203]	11	Disease activity	Positive association (IFN activity positivity associated with disease activity)	High
		Preble, 1982 [119]	86	Flare/remission	Positive association (52.3% samples during flare contained IFN (8-128 IU/ml) vs 25% samples during remission)	High
Plaque- reducing assay	Hooks, 1982 [123]	14	Clinical disease activity grades	Positive association (IFN titre positively associated with high disease activity)	High	
	Hooks, 1979 [121]	28	Active/inactive disease groups	Positive association (IFN was found in the serum of 71% patients with active disease but only in 21% of those with inactive disease)	High	
	Ytterberg, 1982 [124]	23	Disease activity stages (absent, possible, definite mild, definite moderate, definite severe)	Positive correlation (IFN levels: r=0.620, p<0.010)	High	

Supplementary Table 7: Summary of the studies reporting assays analyzing associations between IFN assays and disease activity in RA.

Category	Sub-category	Reference	n	Disease activity measure	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa protein	Rodríguez- Carrio, 2014 [126]	120	DAS28 CRP ESR	DAS28: positive association (higher in IFNhigh vs IFNlow patients, p<0.050) CRP: no association ESR: positive association (higher in IFNhigh vs IFNlow patients, p<0.050)	High
		Weix, 2013 [125]	7	DAS28-CRP	No association	High
	Immunoassays detecting IFNb protein	Weix, 2013 [125]	7	DAS28-CRP	No association	High
Microarrays	Microarray scores and clustering	Cantaert, 2010 [204]	21	DAS28 ESR CRP	Source: WB Main findings: no association	Unclear
		Higgs, 2011 [52]	45	Moderate (n=29) vs severe (n=16) stages	Source: WB Main findings: no association	High
		Reynier, 2011 [132]	81	DAS28	Source: WB Main findings: no association	Unclear
qPCR	qPCR on individual ISG	Weix, 2013 [125]	10	DAS28-CRP	Source: PBMC Number of ISG: 6 Main findings: no association (no differences in any of the genes between active or inactive disease)	High
	qPCR for IFN scores	Rodríguez- Carrio, 2019 [84]	98	DAS28 ESR	Source: WB Number of ISG: 4 (cluster analysis) Main findings: DAS28: positive association (higher DAS28 in cluster I, low	Unclear

					expression of ISG, p<0.001) ESR: positive association (higher ESR in cluster III, high expression of ISG, p=0.025)	
		Cooles, 2018 [138]	75	DAS28	Source: WB Number of ISG: 6 Main findings: positive correlation in early RA (IFN score: r=0.193, p=0.014) and established RA (IFN score: r=0.554, p=0.036)	Unclear
		De Jong, 2016 [70]	182	DAS28 ESR CRP	Source: WB Number of ISG: 19 Main findings: no associations	Unclear
		Rodríguez- Carrio, 2017 [137]	98	DAS28	Source: WB Number of ISG: 4 Main findings: no association in very early RA (IFN score: r=-0.055, p=0.014), or bDMARD-naïve RA (IFN score: p=ns), and negative association in bDMARD patients (IFN score: r=- 0.358, p=0.032)	High
		Thurlings, 2010 [205]	20 31	DAS28 ESR CRP	Source: PBMC Number of ISG: 3 Main findings: no association	Unclear
Reporter cell assays	Reporter cell assays (other)	Shiozawa, 1986 [130]	40	Disease activity groups (active if arthritis w/pain and/or swelling in >4 joints by physician assessment + fatigue and >11 morning stiffness + increased	Assay: luciferase/ chemoluminiscence Main findings: no association	High

			ESR and/or CRP; inactive if criteria for clinical remission)		
Cytopathic effect assay	Hertzog, 1988 [141]	12	Disease activity groups: inactive (MS<15min, no fatigue, no joint pain, no joint tenderness, no joint/tendon swelling, ESR<20) or active (no inactive criteria)	Negative association (OAS activity: higher in inactive RA vs controls, p<0.001)	High
Plaque- reducing assay	Arvin, 1984 [142]	65	Active vs inactive disease	Positive association (IFN activity: present in 10% inactive samples vs 47% active samples)	High

Supplementary Table 8: Summary of the studies reporting assays analyzing associations between IFN assays and disease activity in pSS.

Category	Sub-category	Reference	n	Disease activity measure	Results	Risk of bias
Immunoassays	Immunoassays detecting IFN-	James, 2019 [206]	47	ESSDAI	Positive association (IFN-induced protein clusters positively associated with ESSDAI)	High
	induced proteins	Maria, 2014 [148]	24	ESSDAI	Positive correlation (MxA: r=0.650, p<0.010)	High
		Rose, 2016 [149]	25	ESSDAI	No association (IP-10: p=0.580)	High
Flow cytometry	Flow cytometry to detect IFN-	Maria, 2014 [148]	28	ESSDAI	<i>Target (source):</i> MxA (monocytes) <i>Main findings:</i> positive correlation (MxA: r=0.450, p=0.020)	High
	induced proteins	Rose, 2016 [149]	25	ESSDAI	<i>Target (source):</i> Siglec1 (monocytes) <i>Main findings:</i> positive correlation (Siglec1: r=0.540, p=0.005)	High
Microarrays	Microarray scores and clustering	James, 2019 [206]	47	ESSDAI	Source: WB Main findings: no associations between clusters and ESSDAI (p=0.190)	High
qPCR	qPCR on individual ISG	Kimoto, 2011 [152]	37	ESR Schirmer test (mm/5 min)	Source: WB Number of ISG: 1 Main findings: ESR: positive correlation (IFI27: r=0.333, p<0.050) Schirmer R: no association (IFI27: r=0.076, p=0.709) Schirmer L: no association (IFI27: r=-0.085, p=0.677)	Low
	qPCR for IFN scores	Bodewes, 2018 [89]	86	ESSDAI ClinESSDAI CRP ESR Schirmer test Saliva flow	Source: WB Number of ISG: 3 scores Main findings: ESSDAI: no association (p=0.472) ClinESSDAI: no association (p=0.929) CRP: no association (p=0.567) ESR: positive association (higher ESR in IFN-I/II high patients, p<0.001)	Low

			Schirmer test: negative association (decreased in IFN-I/II	
			high patients, p=0.028)	
			Saliva flow: no association (p=0.274)	
Bodewes,	77	ESSPRI	Source: WB	Unclear
2019 [153]			Number of ISG: 7	
			Main findings: negative association (IFN signature	
			associated with reduced scores of the pain and fatigue	
			domain of the ESSPRI)	
Brkic, 2013	38	ESSDAI	Source: monocytes	Low
[154]		ESSPRI	Number of ISG: 5	
			Main findings: positive correlation (IFN signature: r=0.458,	
			p=0.003)	
Maria,	114	ESSDAI	Source: monocytes	Unclear
2014 [148]			Number of ISG: 5	
			Main findings: positive association (IFNhigh patients	
			showed higher ESSDAI than IFNIow patients, p<0.050)	
Olsson,	90	ESSDAI	Source: WB	Low
2019 [207]		ESSPRI	Number of ISG: 5	
			Main findings:	
			ESSDAI articular domain: positive association (p<0.010)	
			ESSPRI total score: positive association (p=0.040)	
			ESSPRI sicca score: positive association (p=0.030)	
			ESSPRI pain score: positive association (p=0.020)	

Supplementary Table 9: Summary of the studies reporting assays analyzing associations between IFN assays and disease activity in SSc.

Category	Sub-category	Reference	n	Disease activity measure	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa protein	Eloranta, 2010 [159]	70	ESR Digital ulcers	ESR: positive correlation (r=0.390, p<0.001) Digital ulcers: positive association (p=0.029)	High
	Immunoassays detecting IFN-	munoassays Eloranta, 70 ESR Positive correlation (IP-10 r=0.380, p=0.002; MC p=0.045; MIP-1A: r=0.460, p<0.001)	Positive correlation (IP-10 r=0.380, p=0.002; MCP-1: r=0.250, p=0.045; MIP-1A: r=0.460, p<0.001)	High		
	induced proteins	Liu, 2013 [162]	266	mRSS DLC FVC	mRSS: positive association (chemokine score: r=0.210, p=0.002), no association (IP-10: r=0.16, p=0.014; I-TAC: r=0.04, p=0.205) DLC: negative association (chemokine score: r=-0.18, p=0.008; I-TAC: r=-0.21, p=0.002), no association (IP-10: r=- 0.10, p=0.160) FVC: negative association (chemokine score: r=-0.17, p=0.012; I-TAC: r=-0.21, p=0.003), no association (IP-10: r=- 0.08, p=0.272)	Low
Microarrays	Microarray scores and clustering	Assassi, 2010 [47]	74	mRSS FVC DLCO	Source: WB Main findings: no association	Unclear
		Higgs, 2011 [52]	45	mRSS	Source: WB Main findings: positive association (mRSS higher in gene signature positive vs negative groups, p=0.030)	High
		Bos, 2009 [165]	43	Digital ulcers	Source: WB Main findings: positive association (greater number of digital ulcers in IFNhigh patients, p=0.050)	High
		Tan, 2006 [166]	18	mRSS	Source: WB Main findings: no association	High
qPCR	qPCR on individual ISG	Airo, 2008 [168]	50	FVC DLCO Skin score	Source: WB Number of ISG: 1 Main findings:	Unclear

				Digital ulcers	FVC: negative association (lower FVC in patients with high MxA levels, p=0.020) DLCO: no association (p=0.070) Skin score: no association (p=0.140) Digital ulcers: positive association (higher number of digital	
					ulcers in patients with high MxA levels, p=0.002)	
Reporter cell	Reporter cell	Eloranta,	70	ESR	Assay: IFNa-inducing capacity of sera	Unclear
assays	assays (other)	2010 [159]		Digital	Main findings:	
				ulcers	ESR: positive association (higher ESR in patients with high	
					IFNa-inducing capacity, p=0.022)	
					Digital ulcers: positive association (higher ESR in patients with	
					high IFNa-inducing capacity, p=0.025)	

Supplementary Table 10: Summary of the studies reporting assays analyzing associations between IFN assays and disease activity in PM/DM.

Category	Sub-category	Reference	n	Disease activity measure	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa	Krol, 2011 [208]	19	MYOACT MRI-VAS	MYOACT: no association MRI-VAS: negative correlation (r=-0.580, p=0.0095)	High
	protein	Sun, 2012 [174]	16	Skin lesion activity score ESR	Skin lesion activity score: positive correlation (r=0.600, p=0.0147) ESR: positive correlation (r=0.530, p=0.0329)	High
		Huard, 2017 [209]	42	CDASI	CDASI: no association (r=0.290, p=0.090)	Low
	Immunoassays detecting IFNb protein	Huard, 2017 [209]	42	CDASI	CDASI: positive correlation (r=0.540, p=0.0003)	Low
Immune detectir	Immunoassays detecting IFN-	Huard, 2017 [209]	42	CDASI	Positive correlation (CXCL10: r=0.630, p<0.0001)	Low
	induced proteins	Baechler, 2007 [48]	12	Disease activity score (muscle strength testing, muscle enzyme elevation, ulcerative skin disease and patient's report of functional assessment)	Positive association (IP-10: elevated in active patients (no statistical analyses reported; MCP-1: r=0.550, p<0.050; MCP- 2: r=0.600, p<0.010)	High
Microarrays	Microarray scores and	Baechler, 2007 [49]	12	Disease activity score	Source: WB Main findings: positive correlation (IFN score: r=0.440,	High

	clustering			(muscle strength testing, muscle enzyme elevation, ulcerative skin disease and patient's report of functional assessment)	p=0.060; higher in active disease vs inactive disease, p=0.050)	
		Greenberg, 2012 [177]	21	MITAX	Source: WB Main findings: positive association (IFN score higher in patients with high disease activity vs low or moderate disease activity, p<0.010)	High
		Higgs, 2011 [52]	45	Low / high disease activity	Source: WB Main findings: positive association (IFN score higher in high disease activity vs low, p=0.004)	Unclear
		Walsh, 2007 [178]	36	Active disease / improving disease	Source: WB Main findings: positive association (genes most upregulated in patients vs controls were highly down-regulated in improving disease group: IFI27, IFI44L, RSAD2, IFI44, OAS1, BIRC4BP)	Unclear
qPCR	qPCR on individual ISG	O'Connor, 2006 [179]	14	DAS muscle DAS skin	Source: PBMC Number of ISG: 1 Main findings: DAS muscle: positive correlation (MxA: r=0.800, p<0.001) DAS skin: no association (MxA: r=-0.208, p=0.476)	Low
	qPCR for IFN scores	Bilgic, 2009 [180]	37	Physician global VAS MMT8 score	Source: WB Number of ISG: 3 Main findings: Physician global VAS: positive correlation (IFN signature: r=0.410, p=0.007) MMT8 score: negative correlation (IFN signature: r=-0.48, p=0.002)	Unclear

		Ekholm, 2016 [184]	92	Physician global disease activity assessment Patient global disease activity assessment MMT8	Source: WB Number of ISG: 8 Main findings: Physician global disease activity assessment: no association (p=0.442) Patient global disease activity assessment: no association (p=0.443) MMT8: no association (p=0.250)	Unclear
		Huard, 2017 [209]	42	CDASI	Source: WB Number of ISG: 10 Main findings: positive correlation (IFN signature: r=0.610, p<0.001)	Low
	qPCR for IFN- induced chemokines	Bilgic, 2009 [180]	37	Physician global VAS Muscle VAS score MMT8 score	Source: WB Number of ISG: 3 Main findings: VAS score: positive correlation (r=0.61, p<0.0001) Muscle VAS score: positive correlation (r=0.470, p<0.001) MTT8 score: negative correlation (r=-0.440, p=0.002)	Low
RNA-seq	RNA-seq	Huard, 2017 [209]	42	CDASI	Source: WB Main findings: positive association (K-means: optimal grouping of patients with a CDASI=12 (accuracy=0.952); cluster analysis: type I IFN-induced genes correlated with CDASI, patients with mild CDASI similar to controls)	Unclear
Reporter cell assays	Reporter cell assays by qPCR	Ekholm, 2016 [184]	40	Physician global disease activity assessment Patient global disease activity assessment	Assay: RSAD2, IFI44L and MX1 expression Main findings: no association	Unclear

		MMT8		
Niewold, 2009 [182]	30	Muscle enzymes DAS skin	Assay: IFIT1, MX1, PRKR expression Main findings: Active patients: positive correlation (serum CPK: r=0.525, p=0.025, AST: r=0.705, p=0.002, and aldolase: r=0.447, p=0.036) Inactive patients: negative correlation (DAS skin: r=-0.781, p=0.002)	Unclear

Supplementary Table 11: Summary of the studies reporting assays analyzing associations between IFN assays and prognosis (natural history) in SLE.

Category	Sub-category	Reference	n	Endpoint	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa protein	Rose, 2013 [188]	31	Flare (new BILAG A or B score at 180 days)	Positive association (High IFN serum levels (>20 pg/ml) predicted flare, p=0.0004)	Unclear
	Simoa for IFNa	Mathian, 2019 [187]	74	SELENA-SLEDAI flare instrument	Positive association (IFNa serum levels detect flare: AUC 0.840, sensitivity: 73.0 (61.4–82.7), specificity: 82.9 (72.5–90.6); PPV: 80.6; NPV 75.9)	Low
		Mathian, 2019b [35]	254	SELENA-SLEDAI flare instrument (1 year)	Positive association (elevated IFNa levels at baseline predicted higher risk of relapse, HR 5.5 (2.4-2.5), p<0.0001; isolated elevated IFNa levels predicted flare, HR 5.5 (1.7-18.1), p=0.005)	Low
	DELFIA for IFNa	Rose, 2017 [189]	26	Change in mSLEDAI-2K Change in BILAG- 2004 Flare (new BILAG A or B score at 180 days) Remitting disease (improving of A or B in BILAG)	Change in mSLEDAI-2K: positive correlation (change in IFNa serum levels: r=0.447, p=0.001) Change in BILAG-2004: positive correlation (change in IFNa serum levels: r=0.420, p=0.002) Flare: positive association (AUC 0.56 (0.31-0.81), p=0.600, sensitivity: 41.67, specificity: 100, PPV: 100) Remitting disease: positive association (AUC 0.84 (0.52-0.97), p=0.005, sensitivity: 62.50 specificity: 90.48, PPV: 76.00)	High
	Immunoassays detecting IFNb protein	Munroe, 2017 [192]	26	SELENA-SLEDAI flare instrument (6- 12 weeks) SELENA-SLEDAI at follow-up	SELENA-SLEDAI flare instrument: positive association (IFNb serum levels lower at baseline in patients without flare vs those with flare, p<0.010) SELENA-SLEDAI at follow-up: positive correlation (IFNb: r=0.426, p=0.0691)	High
	Immunoassays detecting IFN- induced proteins	Rose, 2017 [189]	26	Change in mSLEDAI-2K Change in BILAG- 2004 Flare (new BILAG A or B score at 180	Change in mSLEDAI-2K: positive correlation (r=0.759, p<0.0001) Change in BILAG-2004: positive correlation (r=0.504, p=0.0002) Flare: positive association (AUC 0.75 (0.55-0.91), p=0.017, sensitivity: 50.00, specificity: 95.24, PPV:	High

		Munroe, 2017 [192]	26	days) Remitting disease (improving of A or B in BILAG) SELENA-SLEDAI flare instrument (6- 12 weeks) SELENA-SLEDAI at follow-up	75.00) Remitting disease: positive association (AUC 0.75 (0.49-0.97), p=0.040, sensitivity: 62.50, specificity: 95.24, PPV: 71.43) SELENA-SLEDAI flare instrument: MCP1: positive association (lower at baseline in patients non-flare vs those with flare, p<0.001) MCP3: positive association (lower at baseline in patients non-flare vs those with flare, p<0.001) MIG: positive association (MIG lower at baseline in patients non-flare vs those with flare, p<0.001) IP10: positive association (lower at baseline in patients non-flare vs those with flare, p<0.001)	High
					MIP-1b: positive association (lower at baseline in patients non-flare vs those with flare, p<0.001) SELENA-SLEDAI at follow-up MCP1: positive correlation (r=0.452, p=0.0547) MCP3: positive correlation (r=0.573, p=0.0178) MIG: positive correlation (r=0.468, p=0.0468) IP10: no association (r=0.426, p=0.0691) MIP-1b: no association (r=0.407, p=0.0697)	
		Bauer, 2009 [210]	267	Flare (1 year)	Positive association (IFN-chemokine score rise at the time of flare (p<0.001) and decreased as disease remitted (p<0.001); IFN-chemokine score predicted flare in patients with SLEDAI<4 at baseline, HR 2.52 (1.63-4.09), p<0.0001)	Low
Flow cytometry	Flow cytometry to detect IFN- induced proteins	Rose, 2017 [189]	26	Change in mSLEDAI-2K Change in BILAG- 2004 Flare (new BILAG A or B score at 180 days) Remitting disease (improving of A or B	Target (source): Siglec1 (monocytes)Main findings:Change in mSLEDAI-2K: positive correlation (r=0.463, p<0.001)Change in BILAG-2004: positive correlation (r=0.448, p<0.001)Flare: positive association (AUC 0.78 (0.57-0.96), p=0.008, sensitivity: 83.33, specificity: 90.48, PPV: 71.43)	High

				in BILAG)	Remitting disease: positive association (AUC 0.75	
					(0.42-1.00), p=0.040, sensitivity: 75.00, specificity:	
					95.24, PPV: 85.71)	
Microarrays	Microarray	Hoffman,	1760	SELENA-SLEDAI	Source: WB	Low
	scores and	2017 [53]		flare instrument, 1	Number of ISG: 34	
	clustering			year	Main findings: positive association (IFN score: RR 5.6,	
	Ŭ			-	p=0.0015 (ILLUMINATE1 cohort); RR 5.9, p=0.0002	
					(ILLUMINATE2 cohort))	
		Mackay,	27	SELENA-SLEDAI	Source: WB	Unclear
		2016 [44]		flare instrument (1	Number of ISG: 2 IFN scores (A/B)	
				vear)	Main findings: positive association (IFN score higher	
				<i>,</i>	at baseline in patients with flare, $p=0.006$)	
qPCR	qPCR for IFN	El-Sherbiny,	60	Flare occurrence	Source: PBMC	High
	scores	2018 [74]		Organ involvement	Number of ISG: 1	-
				-	Main findings:	
					Flare occurrence: positive association (high IFN score	
					A, p=0.042), no association (IFN score B, p=0.343)	
					Organ involvement: no association (IFN score A,	
					p=0.124), positive association (high IFN score B,	
					p=0.037)	
		Feng, 2006	48	SELENA-SLEDAI	Source: WB	High
		[92]		flare instrument	Number of ISG: 6	-
		···			Main findings:	
					IFN score: positive association (higher in patients with	
					severe flare vs those mild/moderate (p=0.020) or stable	
					(p=0.020))	
					LY6E: positive association (higher in patients with	
					severe flare vs stable, p=0.020)	
		Landolt-	27	Change in disease	Source: WB	Unclear
		Maricortena,		activity (1 year)	Number of ISG: 5	
		2009 [80]			Main findings: no association (IFN score: r=-0.022,	
					p=0.910)	
		Md Yusof,	105	CTD development	Source: PBMC	Low
		2018 [82]		(1 year)	Number of ISG: 2 IFN scores (A/B)	
					Main findings: positive association (IFN scores higher	
					in at-risk progressors vs non-progressors, p=0.018 and	

					p<0.001)	
		Steiman, 2015 [211]	102	Clinical quiescence: SACQ (2-year without clinical activity but persistent serologic activity), SQCQ (inactive controls, 2-year without clinical or serologic activity) and SACA (clinical activity requiring use of GC and/or immunosuppressive	Source: WB Number of ISG: 5 Main findings: positive association (IFIT1, ISG15, LY6E, MX1 and OAS1 expression were higher in SACA vs SACQ, p=0.0034, p=0.044, p=0.0014, p=0.027 and p=0.0047; IFN score was higher in SACA vs SACQ, p=0.0025)	Unclear
Nanostring	Nanostring	Wither, 2017 [97]	23 UCTD 19 ANA+	SARD development (1 year)	Source: WB Number of ISG: 5 Main findings: no association	High
DNA methylation	DNA methylation arrays	Ulf-Moller, 2018 [104]	15 twin pairs	Flare (2 years)	Source: sorted populations (CD4+ T-cells, monocytes, granulocytes, B cells) Main findings: positive association (Several ISG exhibit differential methylation, mostly hypomethylation, in twins with flare vs those in remission in CD4+ T-cells (8), monocytes (8), granulocytes (9), and B-cells (8))	High
Reporter cell assays	Reporter cell assays by qPCR	Andrade, 2015 [108]	28	Poor outcomes of pregnancy	Assay: Mx1 expression Main findings: positive association (MX1 expression was higher in patients who developed preeclampsia vs those with other (p<0.006) or without outcomes (p<0.040))	High
Cytopathic effect assay		Mathian, 2019 [187]	74	SELENA-SLEDAI flare instrument	Positive association (increased IFN bioactivity predicted flare: AUC 0.780, sensitivity: 63.5 (51.5– 74.4), specificity: 88.2 (78.7–94.4), PPV: 83.9, NPV: 71.3)	Low

Supplementary Table 12: Summary of the studies reporting assays analyzing associations between IFN assays and prognosis (natural history) in RA.

Category	Sub-	Reference	n	Endpoint	Results	Risk of bias
	category			-		
Microarrays	Microarray scores and clustering	van Baarsen, 2010 [212]	109	RA development (1 year)	Source: WB Number of ISG: 52 gene sets (cluster analysis) Main findings: positive association (IFN signature associated with RA development, OR 21 (95% CI: 2.8–156.1), p=0.003)	High
qPCR	qPCR for IFN scores	Cooles, 2018 [138]	632	Disease activity (DAS28) at 6 months	Source: WB Number of ISG: 5 Main findings: positive correlation (r=0.319, p=0.002)	High
		Lubbers, 2013 [213]	115 73	RA development	Source: WB Number of ISG: 7 Main findings: positive association (cohort 1: 15/25 IFNhigh patients developed RA vs 29/90 IFNIow, p=0.001, AUC 0.602 (95% CI 0.491- 0.714, p=0.066; cohort 2: IFN score higher in presymptomatic individuals who developed RA vs controls, p=0.002; IFNhigh status: 14/23 RA, 15/25 presymptomatic, 10/45 HC, p=0.004)	High

Supplementary Table 13: Summary of the studies reporting assays analyzing associations between IFN assays and prognosis (natural history) in SSc.

Category	Sub-	Reference	n	Endpoint	Results	Risk of bias
	category					
Microarrays	Microarray	Assassi,	62	Increase in	Source: WB	High
-	modules	2019		FVC%, 26	Main findings:	-
		[164]		months	Increase in FVC%: negative correlation (M1.2: r=-0.430, p=0.009; M3.4:	
				Change in	r=-0.450, p=0.007)	
				mRSŠ, 26	Change in mRSS: no associations	
				months		

Supplementary Table 14: Summary of the studies reporting assays analyzing associations between IFN assays and prognosis (response to treatments) in SLE.

Category	Sub-	Reference	n	Treatment [target]	Endpoint	Results	Risk of bias
Microarrays	Microarray scores and clustering	Hoffman, 2017 [53]	1760	Tabalumab [B cell activating factor]	SRI (52 weeks)	Source: WB Number of ISG: 34 Main findings: no association	Unclear
qPCR	qPCR for IFN scores	Furie, 2017 [214]	304	Anifrolumab (300 mg, n=99 and 1000 mg, n=103) [IFN alpha receptor subunit 1]	SRI(4) including GC taper (52 weeks) SRI(4) excluding GC taper (52 weeks) SRI(7) (52 weeks) BICLA (52 weeks)	Source: WB Number of ISG: 4 Main findings: positive association (greater effect size in IFNhigh patients) Anifrolumab 300 mg SRI(4) including GC taper: IFNhigh OR 4.30 (2.34-7.91), p<0.001; IFNlow 1.47 (0.55-3.93), $p=0.514SRI(4) excluding GC taper: IFNhigh OR 2.98 (1.69-5.24),p<0.001$; IFNlow 2.07 (0.77-5.53), $p=0.225SRI(7): IFNhigh OR 4.59 (2.26-9.33), p<0.001; IFNlow0.94 (0.29-3.04), p=0.930BICLA: IFNhigh OR 3.65 (2.02-6.60), p<0.001; IFNlow3.19 (1.16-8.73), p=0.059Anifrolumab 1000 mgSRI(4) including GC taper: IFNhigh OR 2.52 (1.37-4.64),p=0.013$; IFNlow 0.89 (0.34-2.35), $p=0.849SRI(4) excluding GC taper: IFNhigh OR 2.33 (1.34-4.04),p=0.012$; IFNlow 0.85 (0.34-2.12), $p=0.763SRI(7): IFNhigh OR 2.65 (1.29-5.34), p=0.26; IFNlow0.81 (0.25-2.61), p=0.763BICLA: IFNhigh OR 2.41 (1.34-4.35), p=0.014; IFNlow1.38 (0.51-3.70), p=0.596$	Low
		Kalunian, 2016 [215]	238	Rontalizumab [IFN alpha protein]	BILAG index response (24 weeks) SRI4 (24	Source: WB Number of ISG: 3 Main findings: positive association (SRI response was higher and steroid use was lower in the IFNIow	Unclear

			weeks)	rontalizumab treated patients)	
				BILAG response index: IFNhigh: placebo 21/55 (38.2%) vs rontalizumab 53/123 (43.1%), 5.2% (-7.6 - 18.1%), p=0.510 IFNIow: placebo 12/24 (50%) vs rontalizumab 18/33 (54.5%), 4.5% (-17.7 - 26.5%), p=0.790 SRI4: IFNhigh: placebo 26/55 (47.3%) vs rontalizumab 55/123 (44.7%), -2.3% (-15.5 - 10.9%), p=0.780 IFNIow: placebo 10/24 (41.7%) vs rontalizumab 24/33 (72.7%), 31.1% (8.9 - 51.0%) p=0.030	
Merrill, 2018 [216]	201	Anifrolumab 300 mg [IFNA receptor subunit 1]	SLEDAI-2K- defined resolution of rash BILAG- defined improvement in rash Improvement in CLASI Resolution in SLEDAI-2K- defined arthritis Improvement in BILAG- defined arthritis Mean change in swollen and tender joint counts	Source: WB Number of ISG: 4 Main findings: positive association (greater effect in IFNhigh patients) SLEDAI-2K-defined resolution of rash: IFNhigh: placebo 7/65 (10.8) vs anifrolumab 33/67 (49.3), OR 8.08 (3.72-17.52), p<0.001 IFNlow: placebo 6/23 (26.1) vs anifrolumab 6/21 (28.6), OR 1.40 (0.43-4.53), p=0.639 BILAG-defined improvement in rash: IFNhigh: placebo 17/64 (26.6) vs anifrolumab 35/61 (57.4), OR 3.78 (2.00-7.14), p<0.001 IFNlow: placebo 7/21 (33.3) vs anifrolumab 13/21 (61.9), OR 3.93 (1.28-12.04), p=0.044 Improvement in CLASI: IFNhigh: placebo 21/67 (31.3) vs anifrolumab 43/70 (61.4), OR 3.67 (2.01-6.71), p<0.001 IFNlow: placebo 9/22 (40.9) vs anifrolumab 14/22 (63.3), OR 2.70 (0.94-7.80), p=0.123 Resolution in SLEDAI-2K-defined arthritis: IFNhigh: placebo 29/73 (39.7) vs anifrolumab 41/73 (56.2), OR 2.11 (1.20-3.71), p=0.030 IFNlow: placebo 13/26 (50.5) vs anifrolumab 14/24 (58.3), OR 1.41 (0.55-3.64), p=0.547	Unclear

					Improvement in BILAG-defined arthritis: IFNhigh: placebo $34/72$ (47.2) vs anifrolumab $47/71$ (66.2), OR 2.39 (1.34-4.27), p=0.013 IFNlow: placebo $13/23$ (56.5) vs anifrolumab $18/23$ (78.3), OR (2.91 (0.97-8.72), p=0.110 Mean change in swollen and tender joint counts: IFNhigh: placebo $-3.0(5.8)$ vs anifrolumab $-4.9(6.1)$, mean difference (SE) $-1.9(0.8)$, p=0.014 IFNlow: placebo $-4.5(6.1)$ vs anifrolumab $-7.4(6.3)$, mean difference (SE), $-2.1(1.4)$, p=0.140	
	Petri, 2013 [217]	213	Sifalimumab (0.3-10 mg)	Mean reduction in	Source: WB Number of ISG: 21	Unclear
	[-··]		[IFN alpha	SELENA-	Main findings: no association (greater but non-	
			proteinij	OLEDAI	change: -2.6 vs -2.4)	

Supplementary Table 15: Summary of the studies reporting assays analyzing associations between IFN assays and prognosis (response to treatments) in RA.

Category	Sub-category	Reference	n	Treatment	Endpoint	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa protein	Rodríguez- Carrio, 2014 [126]	26	Anti-TNF (various agents) [TNF protein]	EULAR clinical response (12 weeks)	Positive association (lower effect in IFNhigh patients (0% good response, 41.6% moderate response, 58.3% no response) compared to IFNlow (35% good response, 35% moderate response, 0% no response).	High
Microarrays	Microarray scores and clustering	Cantaert, 2010 [204]	21	Infliximab [TNF protein]	EULAR clinical response (24 weeks)	Source: WB Number of ISG: 43 Main findings: no association	High
		Reynier, 2011 [132]	22	Anti-TNF (various agents) [TNF protein]	EULAR clinical response (24 weeks)	Source: WB Number of ISG: 35 Main findings: no association	High
		Sanayama, 2014 [218]	40 20	Tocilizumab [anti-IL-6R]	Clinical response by physician global assessment	Source: PBMC Number of ISG: number of differentially expressed genes (DEG) Main findings: positive association (409 probes fulfilled criteria for DEG (>1.5 FC): IFI6 (higher in responders, 0.006, AUC 0.693), MX2 (higher in responders, p=0.004, AUC 0.920), OASL (higher in responders, p=0.024, AUC 0.627; IFI6, MX2 and OASL score: AUC 0.853, sensitivity 80.0, specificity 80.0, PPV 92.0, NPV 57.0)	High
		van Baarsen, 2010 [212]	15	Infliximab [TNF protein]	EULAR clinical response (24 weeks)	Source: WB Number of ISG: 15 Main findings: positive association (increased IFN score at 4 weeks was associated with poor clinical response, p=0.022)	High
		Vosslamber,	13	Rituximab	Change in	Source: WB	High

		2011 [219]		[CD20]	DAS:	Number of ISG: number of differentially expressed	
					>1.2/<1.2	genes, gene clusters	
					(24 weeks)	Main findings:	
					EULAR	Change in DAS: positive association (increased IFN	
					clinical	response during treatment was associated with good	
					response	or moderate response, p=0.018)	
					(24 weeks)	EULAR clinical response: positive association	
						(equivalent results)	
qPCR	qPCR for IFN	Raterman,	27	Rituximab	Responders	Source: WB	Low
	scores	2012 [220]		[CD20]	(dDAS>1.2)	Number of ISG: 8	
					vs non-	Main findings: positive association (IFN score at	
					responders	baseline was negatively correlated with dDAS<1.2,	
					(dDAS<1.2)	OR: 0.25 (0.09-0.70), p=0.008; IFN score at baseline	
					(24 weeks)	was lower in responders vs non-responders, AUC	
						0.820, p=0.0074)	
		Cooles,	32	MTX, HCQ	EULAR	Source: WB	Low
		2018 [138]			clinical	Number of ISG: 5	
					response	Main findings:	
					(24 weeks)	EULAR clinical response: positive association (IFN	
					Need of	score at baseline was negatively associated with	
					additional	good response, p=0.044)	
					GC doses	Need of additional GC doses: positive association	
					(24 weeks)	(IFN score at baseline was negatively associated	
						with additional GC administration, p=0.0003)	
		De Jong,	40	Rituximab	Change	Source: WB	Unclear
		2015 [221]		[CD20]	dDAS>1.2	Number of ISG: 8	
					(24 weeks)	Main findings: positive association (higher IFN	
						score at baseline was associated with non-response,	
						AUC 0.828 (0.699-0.957), p<0.001)	
		Rodríguez-	18	GC+MTX	EULAR	Source: WB	Unclear
		Carrio, 2017	13	Anti-TNF	clinical	Number of ISG: 4	
		[137]		(various	response	Main findings:	
		_		agents)	(12 and 24	VERA cohort (n=18)	
				[TNF	weeks)	EULAR clinical response: positive association	
				protein]	DAS28 (24	(higher IFN score was associated with poor clinical	
					and 52	response, p=0.006, AUC 0.917 (0.782 – 1.000),	

					weeks)	p=0.004) DAS28: positive correlation (24 weeks: r=0.620, p=0.008; 52 weeks: r=0.552, p=0.041) Anti-TNF (n=13) DAS28: positive association (IFN score at baseline predicted higher DAS28 at 12 weeks after adjusting for confounders, B (95% CI): 0.577 (0.052-1.102), p=0.035)	
		Thurlings, 2010 [205]	20 31	Rituximab [CD20]	EULAR clinical response, (12 and 24 weeks) DAS28 decrease (12 and 24 weeks)	Source: PBMC Number of ISG: 3 Main findings: Cohort 1 (n=20) EULAR clinical response: no association DAS28 decrease: no association Cohort 2 (n=31) EULAR clinical response: positive association (IFNhigh patients less likely to achieve a good response, 12 weeks p=0.043, 24 weeks: p=0.059) DAS28 decrease: positive association (lower reduction in IFNhigh patients, 12 weeks: p=0.008, 24 weeks: p=0.027) Pooled analysis: positive association (high IFN signature associated with poor clinical response and DAS28 decrease)	Unclear
RNA-seq	RNA-seq	Wright, 2015 [139]	30	Anti-TNF (various agents) [TNF protein]	EULAR clinical response (12 weeks) Change DAS28>1.2 (12 weeks) Change in DAS28 (12	Source: neutrophils Number of ISG: 128 Main findings: EULAR clinical response: positive association (high IFN score at baseline predicted good clinical response, AUC 0.760) Change DAS28>1.2: positive association (high IFN score more likely to have a change DAS28<1.2, AUC 0.640)	High

					weeks)	Change in DAS28: positive correlation (r=0.210, p=0.020)	
Reporter cell assays	Reporter cell assays by qPCR	Thurlings, 2010 [205]	20 31	Rituximab [CD20]	EULAR clinical response (12 and 24 weeks) DAS28 decrease (12 and 24 weeks)	Assay: serum IFN bioactivity Main findings: EULAR clinical response: no association DAS28 decrease: no association	High
		Mavragani, 2010 [140]	47	Anti-TNF (various agents) [TNF protein]	EULAR clinical response (14 weeks)	Assay: serum IFN bioactivity Main findings: positive association (high IFN bioactivity associated with a good clinical response, OR 1.36 (1.05-1.76), p=0.027)	High
		Muskardin, 2016 [222]	32 92	Anti-TNF (various agents) [TNF protein]	EULAR clinical response (12 weeks)	Assay: serum IFN bioactivity Main findings: Test cohort (n=32) EULAR clinical response: positive association (IFNb/a ratio >1.3 associated with a lack of EULAR clinical response, p=0.010) Validation cohort (n=92) EULAR clinical response: positive association (IFNb/a ratio >1.3 associated with no response at 12 weeks, OR 6.67 (1.37-32.55), p=0.018)	High
Supplementary Table 16: Summary of the studies reporting assays analyzing associations between IFN assays and prognosis (response to treatments) in PM/DM.

Category	Sub-category	Reference	n	Treatment [target]	Endpoint	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa protein	Reed, 2012 [223]	51	Immunomodulatory treatment (AZA, MTX, MMF, HCQ and GC)	Change in global VAS Change in muscle VAS Change in extra- skeletal VAS	Change in global VAS: no association (IFNa: r=- 0.140, p=0.460) Change in muscle VAS: no association (IFNa: r=-0.17, p=0.380) Change in extra-skeletal VAS: no association (IFNa: r=-0.220, p=0.260)	High
	Immunoassays for IFN- induced proteins	Reed, 2012 [223]	51	Immunomodulatory treatment (AZA, MTX, MMF, HCQ and GC)	Change in global VAS Change in muscle VAS Change in extra- skeletal VAS	Change in global VAS: negative correlation (MIP1a: r=-0.190, p=0.320, IP-10: r=-0.610, p<0.001; IFN-chemokine score: r=-0.590, p<0.001) Change in muscle VAS: no association (MIP1a: r=-0.190, p=0.330); negative correlation (IP-10: r=-0.510, p<0.001; IFN-chemokine score: r=- 0.500, p<0.0001) Change in extra-skeletal VAS: no association (MIP1a: r=-0.090, p=0.670), negative correlation (IP-10: r=-0.640, p<0.001; IFN-chemokine score: r=-0.620, p<0.001)	High
		Reed, 2015 [224]	177	Rituximab [CD20]	Changes in muscle VAS (16 weeks) Physician global VAS scores (16	Changes in muscle VAS: positive association (IFN-chemokine score at baseline predicted larger improvements in muscle VAS, depending on the autoantibody profile) Physician global VAS scores: no association (p=0.090)	High

					weeks)		
qPCR	qPCR for IFN scores	Reed, 2012 [223]	51	Immunomodulatory treatment (AZA, MTX, MMF, HCQ and GC)	Change in global VAS Change in muscle VAS Change in extra- skeletal VAS	Source: WB Number of ISG: 3 Main findings: Change in global VAS: negative correlation (IFN score: r=-0.430, p=0.003) Change in muscle VAS: negative correlation (IFN score: r=-0.560, p<0.0001) Change in extra-skeletal VAS: negative correlation (IFN score: r=-0.300, p=0.048)	High

Supplementary Table 17: Summary of the studies reporting assays analyzing associations between IFN assays and prognosis (response to treatments) in pSS.

Category	Sub-	Reference	n	Treatment	Endpoint	Results	Risk of bias
a DCD		Padawaa	27		Change in	Source: W/P	High
YPCK		DOUEWES,	31			Number of ISC: 5	підп
	IFN SCORES	2019[155]			24 and 48	Main findings:	
	300/63				24 and 40	Change in ESP: no association	
					Change in	Change in LON. No association	
						Change in ESSDAL and ESSDEL: no association	
					Igo anu IgM levels	Change in coular or oral dryness: no association Change in SE-	
					$(12 \ 24 \text{ and})$	36 or HAD scales: no association	
					(12, 24 anu 48 weeks)		
					Change in		
					ESSDAI		
					and		
					ESSPRI		
					(12, 24 and		
					48 weeks)		
					Change in		
					ocular or		
					oral		
					dryness		
					(12, 24 and		
					48 weeks)		
					Change in		
					SF-36 or		
					HAD		
					scales (12,		
					24 and 48		
					weeks)		
		Quartuccio,	12	Belimumab	Change in	Source: PBMC	High
		2017 [225]		[B	ESSDAI	Number of ISG: 3	
				lymphocyte	(28 and 52	Main findings: no association	

		stimulator]	weeks)	

	Supplementary	/ Table 18: Summar	y of the studies repor	rting assays analy	zing responsiveness t	to change of IFN assay	s in SLE.
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Category	Sub-category	Reference	n	Treatment	Timepoints	Results	Risk of bias
-				[target]			
Immunoassays	Immunoassays detecting IFNa protein	Bengtsson, 2000 [185]	30	Usual care	Pre-flare / flare / post- flare visits Intervals: Pre-flare vs flare: median 4 months (1- 21) Post-flare 1 vs flare: 2 months (0.5-9) Post-flare 2 vs post-flare 1: 2.5 months (1	Changes observed: IFNa levels increased at flare vs pre-flare (p=0.005), post-flare 1 (p=0.0008) and post-flare 2 (p=0.002)	Unclear
					5)		
		Fragoso- Loyo, 2012 [8]	20	Usual care	6 months	No change (IFNa: T0 52.4 (3.2–1074.4) vs T6 77.6 (3.2–2321.4), p=0.200)	High
		Willis, 2012 [14]	35	HCQ	Consecutive visits (not specified)	No change (IFNa decreased (non-significant) by 33.5%: 573.06 to 381.03, p=0.2507)	Low
	Immunoassays for IFN- induced proteins	Bauer, 2009 [210]	73	Usual care	Consecutive visits from patients who exhibited a reduction in disease	<i>Target:</i> 3 chemokines (CXCL10, CCL19, CCL2) <i>Main findings:</i> changes observed (IFN-chemokine score drop at disease activity reduction, p<0.0010; Chemokine score increased at interim visits (42% increase) and at flare visits (53% increase))	Low

		Connelly, 2018 [196]	109	Usual care	activity (decrease in SLEDAI ≥3) along 1 year Consecutive visits (median length follow up = 3.2 years, median clinical visits = 7)	Target: 3 chemokines (CXCL10, CCL19, CCL2) Main findings: changes observed in association with disease activity (increase of one unit in IFN-CK score was significantly associated with an increase in SLEDAI-2K of 0.7 (RC = 0.73, (95% CI: 0.12, 1.43) p = 0.02)) Target: 134 chemokines (CXCL10, CCL19)	Unclear
		2018 [90]	304	(3, 300 or 1000 mg) [IFNA receptor subunit 1]	i yeai	CCL2) Main findings: changes observed (Anifrolumab suppressed 11/27 proteins elevated in patients with SLEDAI-2K>10 and 5/7 proteins elevated in patients with CLASI>10)	Unclear
Flow cytometry		Alexander, 2015 [226]	12	Bortezomib (21-day cycles) [proteasome]	2 (mean) 21-day cycles	<i>Target (source):</i> Siglec1 (monocytes) <i>Main findings:</i> changes observed (Siglec1 expression decreased upon treatment, baseline: 42.3 vs post-treatment: 18.8, p<0.001)	Low
		Li, 2010 [39]	4	High dose IV prednisolone (1 g/day)	3 days	<i>Target (source):</i> CD64 (monocytes) <i>Main findings:</i> changes observed (CD64 decreased at day 3, p=0.006)	High
Microarrays	Microarray modules	Chiche, 2014 [46]	29	Usual care	At least three visits: median follow-up: 8.3 (2–28) months, median interval	Source: WB Main findings: changes observed (module M1.2 very stable over time within individual patients (coefficient of variation=0.05); module M3.4: greater variation (CV=0.39); module M5.12: much greater variation (CV=0.91))	Low

				between visits:3.2 (0.5–19)		
	Petri, 2019 [227]	243	Usual care	2 or more clinical visits (not specified)	Source: WB Main findings: no changes (IFN modules highly stable over time: M1.2 (ICC 0.88), M3.4 (ICC 0.79), M5.12 (ICC 0.75))	Unclear
Microarray scores and clustering	Petri, 2009 [199]	11	Usual care	2-3 longitudinal visits (not specified)	Source: WB Number of ISG: 4 Main findings: no changes (IFN scores remained constant, despite significant changes in disease activity)	Unclear
	Bennet, 2003 [131]	3	IV glucocorticoids 30 mg/kg/day	3 days	Source: PBMC Number of ISG: several ISG Main findings: changes observed (significant downregulation of several ISG, overall p<0.001; interferon signature decreased, but granulocyte-related signature not affected)	High
	Hoffman, 2017 [53]	1760	Tabalumab [B cell activating factor]	1 year	Source: WB Number of ISG: 34 ISG Main findings: no change (no change in IFN score over the 52-week duration in the placebo or tabalumab group; IFN signature was not down-modulated in patients who showed improvement in disease activity)	Unclear
	Kawasaki, 2011 [198]	12	Prednisolone initiation or increase in doses	Before (active phase) and after (inactive phase) of steroid treatment	Source: CD3+ T-cells Number of ISG: signaling pathways and differentially expressed genes (DEG) Main findings: changes observed (711 DEG detected; interferon pathway ranked the top, with higher levels during the active phase)	Low
	Lauwerys, 2013 [58]	27	IFNa-kinoid	168 days	Source: WB Number of ISG: 21	Unclear

						<i>Main findings:</i> changes observed (21 ISG changed from day 0 to day 122 and 168; IFNa- kinoid reduced the expression of ISG in IFNpositive patients)	
		Petri, 2013 [217]	161	Sifalimumab (0.3-10 mg/kg) every other 2 weeks [IFNa protein]	26 weeks under treatment + 24 weeks follow up	Source: WB Number of ISG: 21 Main findings: changes observed (sustained inhibition with sifalimumab in IFNhigh patients, maximum average inhibition of 38.7% in the 1 mg/kg group)	Low
qPCR	qPCR for IFN scores	Kennedy, 2015 [63]	61	Rontalizumab [IFNa protein]	36 weeks	Source: PBMC Number of ISG: 3 Main findings: changes observed (rontalizumab decreased IFN score over time vs placebo, IFNIow patients exhibited a greater effect but downregulation in IFNhigh did not reach IFNIow levels)	Unclear
		Dominguez- Gutierrez, 2014 [71]	60	Usual care	Follow-up of patients with at least 2 visits (not specified)	Source: PBMC Number of ISG: 3 Main findings: changes observed (patients can be classified into increasing and decreasing IFN score groups)	High
		Liu, 2018 [81]	7	Not specified	Not specified	Source: PBMC Number of ISG: 4 Main findings: changes observed (IFN-I score decrease (p<0.050) upon treatment or follow up (duration and treatment not disclosed))	High
		Aranow, 2015 [228]	48	Vitamin D (0, 2000 IU or 4000 IU)	12 weeks	Source: WB Number of ISG: 3 Main findings: no change (IFN score was unchanged upon vitamin D treatment)	Low
		Fu, 2008 [77]	4	Treatment for active nephritis (various agents)	12 weeks	Source: WB Number of ISG: 7 (IFN-induced chemokines) Main findings: changes observed (3/4 had their chemokine-induced score reduced and improved nephritis; 1/4 had their score	High

					increased and progressed to renal failure)	
	Furie, 2019 [79]	12	BIIB059 (20 mg/kg) [BDCA2]	1 day	Source: WB Number of ISG: 9 Main findings: changes observed (a single BIIB059 dose lead to a rapid and partial neutralization of ISG expression compared to placebo)	Unclear
	Hasni, 2019 [229]	10	Omalizumab [IgE]	16 weeks	Source: WB Number of ISG: 4 Main findings: no change (IFN score: all group p=0.110, IFNhigh p=0.052)	Unclear
	Kawasaki, 2011 [198]	12	Prednisolone initiation or increase in doses	Before (active phase) and after (inactive phase) of steroid treatment	Source: CD3+ T-cells Number of ISG: 6 Main findings: changes observed (higher ISG expression in active vs non-active phase, all p<0.050)	Low
	Li, 2010 [39]	4	High dose IV prednisolone (1 g/day)	3 days	Source: WB Number of ISG: 3 Main findings: changes observed (Mx1 expression decreased at day 3, p<0.050)	High
	McBride, 2012 [230]	32	Rontalizumab (single-dose or repeat dose) [IFN alpha protein]	6 months	Source: WB Number of ISG: 7 Main findings: changes observed (rontalizumab induced a substantial decline (>50%) in relative gene expression at 3 mg/kg (31.3% of baseline) and 10 mg/kg (23.1% of baseline), different doses lead to different length of the sustained suppression)	Unclear
	Merrill, 2011 [231]	34	Sifalimumab (doses from 0 to 30 mg/kg) [IFN alpha protein]	84 days	Source: WB Number of ISG: 21 Main findings: changes observed (doses 0, 3 and 10 led to changes in IFN score at day 14 (p<0.050, p=0.010 and p=0.010); no changes	Unclear

						with doses 1 or 3, p=0.090 and p=0.060)	
Reporter cell assays	Reporter cell assays by qPCR	Hua, 2006 [109]	2	Usual care	5 or 7 months	Number of ISG: 5 Main findings: changes observed (IFN score varied over time, but fluctuations may be confounded by GC pulses or HCQ)	High
		Niewold, 2008 [111]	28	Usual care	Not specified	Number of ISG: 3 Main findings: changes observed (patients could be classified into stable or unstable IFN signature)	High
Cytopathic cell assay		Lackovic, 1984 [118]	4	Usual care	Not specified	No change (no consistent pattern)	High
		von Wussow, 1988 [232]	61	Usual care	Not specified	No change (no consistent pattern)	High
Plaque assay		Ytterberg, 1982 [124]	14	Usual care	Not specified	Changes observed (IFN bioactivity patterns found in association with disease activity, p<0.050)	High

Category	Sub-category	Reference	n	Treatment [target]	Timepoints	Results	Risk of bias
Immunoassays	Immunoassays for IFN- induced proteins	Quartier, 2011 [233]	24	Anakinra [IL-1a]	6 months	<i>Target:</i> 2 chemokines (IP-10 and TRAIL) <i>Main findings:</i> changes observed (IP-10 and TRAIL significantly increased in anakinra-treated patients at 6 months, p=0.0316 and p=0.0003)	Unclear
Microarrays	Microarray modules	Quartier, 2011 [233]	24	Anakinra [IL-1a]	6 months	Source: WB Main findings: changes observed (coordinated upregulation of type I IFN-inducible transcripts (module M3.1) in the anakinra but not in the placebo-treated group, regardless of the clinical response)	Unclear
	Microarray scores and clustering	van Baarsen, 2010 [212]	15	Anti-TNF (infliximab) [TNF protein]	1 month	Source: WB Number of ISG: 34 Main findings: no changes (no variation in ISG (mean 34 genes or mean 3 IFN genes) after 1 month upon anti-TNF)	Low
qPCR	qPCR for IFN scores	Bienkowska, 2014 [234]	340	Baminercept (700 or 200 mg) [lymphotoxin β]	14 weeks	Source: WB Number of ISG: 15/3 Main findings: changes observed (IFN reduced in IFNhigh patients upon treatment in two cohorts)	Unclear
		Cantaert, 2010 [204]	18	Anti-TNF [TNF protein]	1 month	Source: WB Number of ISG: 43 Main findings: no change (TNF blockade did not modulate the mean expression of ISG, increasing in 3 patients, decrease in 11 patients)	Low
		Cooles, 2018 [138]	11	Initiation of GC, MTX, HCQ	6 months	Source: WB Number of ISG: 5 Main findings: changes observed (sustained decrease in IGS after 6 months of treatment, 1 month: p<0.010; 3 months: p<0.010 and 6 months: p<0.050)	Low

Supplementary Table 19: Summary of the studies reporting assays analyzing responsiveness to change of IFN assays in RA.

		Rodríguez- Carrio, 2017 [137]	13	Anti-TNF (various agents) [TNF protein]	3 months	Source: WB Number of ISG: 4 Main findings: no change (no changes in ISG or IFN score upon TNF-blockade, all p>0.050)	Low
		Rodríguez- Carrio, 2019 [84]	13	Anti-TNF (various agents) [TNF protein]	3 months	Source: WB Number of ISG: 5 Main findings: no change (no changes in ISG or IFN score upon TNF-blockade regardless of the response status, all p>0.050)	Low
		Vosslamber, 2011 [219]	22	Rituximab [CD20]	3 months	Source: WB Number of ISG: 6 Main findings: changes observed (IFN score increased at 3 months compared to baseline in responders but not in non-responders, p=0.0040)	Low
		Weix, 2013 [125]	10	Not specified (pregnant RA patients)	Pre- pregnancy to postpartum	Source: PBMC Number of ISG: 6 Main findings: changes observed (IFI35 and IFI44 expression changed along pregnancy and postpartum; no differences in IFI44L, IFIT3 and OAS1)	Unclear
Reporter cell assays	Reporter cell assays by qPCR	Muskardin, 2015 [222]	32 92	Anti-TNF (various agents) [TNF protein]	4-6 weeks	<i>Number of ISG:</i> 5 <i>Main findings:</i> changes observed (decrease in total IFN activity, IFNb activity and IFNb/a ratio was associated with no response, all p<0.050)	Unclear

Supplementary	/ Table 20: Summar	v of the studies repo	orting assav	/s analvzing	responsiveness	to change of	IFN assavs in SSc.

Category	Sub-category	Reference	n	Treatment	Timepoints	Results	Risk of bias
Immunoassays	Immunoassays for IFN- induced proteins	Liu, 2013 [162]	63	Usual care	Consecutive visits (mean time in study: 3.1±1.2 years)	<i>Target:</i> IP-10 and I-TAC <i>Main findings:</i> no changes observed (IP-10 p=0.977; I-TAC p=0.512; IFN-induced chemokine score p=0.621)	Unclear
		Assassi, 2019 [164]	62	Cyclophosphamide (n=35) Hematopoietic cell transplant (n=27)	6 and 12 months	<i>Target:</i> 15 chemokines <i>Main findings:</i> changes observed (chemokine protein score significantly decreased in the hematopoietic cell transplant arm; no significant change in the cyclophosphamide group)	Unclear
Microarrays	Microarray modules	Assassi, 2019 [164]	62	Cyclophosphamide (n=35) Hematopoietic cell transplant (n=27)	6 and 12 months	Source: WB Main findings: changes observed (decline in IFN modules (M1.2 and M3.4) post treatment, greater effect in the hematopoietic cell transplant group)	Unclear

Category	Sub-category	Reference	n	Treatment [target]	Timepoints	Results	Risk of bias
Immunoassays	Immunoassays detecting IFNa protein	Pollard, 2013 [143]	28	Rituximab (1000 mg, day 1 and 15) [CD20]	Pre- and post- treatment (5, 12, 24, 36 and 48 weeks)	Changes observed (IFNa decreased levels after treatment at 5 and 12 weeks, p<0.050)	High
	Immunoassays for IFN- induced proteins	Pollard, 2013 [143]	28	Rituximab (1000 mg, day 1 and 15) [CD20]	Pre- and post- treatment (5, 12, 24, 36 and 48 weeks)	<i>Target:</i> MIP-1b and MIG <i>Main findings:</i> changes observed (MIP1b decreased levels after treatment at 12 weeks, p<0.050; MIG decreased levels after treatment at 5, 12 and 36 weeks, all p<0.050)	High
Flow cytometry		Rose, 2016 [149]	11	GC (n=5) HCQ (n=6)	Pre- and post- treatment (GC: 5.5 months, HCQ: 5 months)	<i>Target (source):</i> Siglec1 (monocytes) <i>Main findings:</i> changes observed (Siglec1 expression reduced upon GC and HCQ, p=0.028 and p=0.046)	Unclear
qPCR	qPCR for IFN scores	Brkic, 2013 [154]	69	Usual care	Two measurements (3.6±2.5 years)	Source: sorted cell populations (monocytes) Number of ISG: 5 Main findings: no change	High
		Bodewes, 2019 [153]	77	HCQ	Pre- and post- treatment (24 weeks)	Source: WB Number of ISG: 5 Main findings: changes observed (IFN-I score reduced upon HCQ treatment at 24 weeks, p<0.050)	Unclear
Reporter cell assays	Reporter cell assays by qPCR	Mavragani, 2007 [158]	10	Etanercept (25 mg, twice weekly) [TNF protein]	Pre- and post- treatment (12 weeks)	Number of ISG: 2 Main findings: changes observed (IFN bioactivity increased upon treatment, baseline 4.12±1.77 vs post-treatment 7.46±5.34, p=0.040)	Unclear

Supplementary Table 21: Summary of the studies reporting assays analyzing responsiveness to change of IFN assays in pSS.

Supplementary	/ Table 22: Summar	y of the studies re	porting assay	s analyzing re	esponsiveness to	o change of IFN ass	avs in PM/DM.

Category	Sub-category	Reference	n	Treatment [target]	Timepoints	Results	Risk of bias
Immunoassays	Immunoassays for IFN- induced proteins	Reed, 2015 [224]	177	Rituximab [CD20]	Pre- and post- treatment (16 weeks)	<i>Target:</i> IP-10, I-TAC and MCP-1 <i>Main findings:</i> changes observed (IFN chemokine scores fluctuated upon treatment depending on autoantibody profiles)	Unclear
		Reed, 2012 [223]	51	Immunosuppressive regimens (various agents)	Two consecutive visits (interval not specified)	<i>Target:</i> IP-10, I-TAC and MCP-1 <i>Main findings:</i> changes observed (IFN chemokine scores correlated with changes in global (r=0.530, p<0.001), muscle (r=0.500, p<0.001) and extra-skeletal VAS (r=0.550, p<0.001); changes in IP-10 correlated changes in global (r=0.530, p<0.001), muscle (r=0.440, p=0.002) and extra-skeletal VAS (r=0.520, p<0.001)	Low
		López de Padilla, 2015 [235]	200	Rituximab [CD20]	Pre- and post- treatment (8 and 16 weeks)	<i>Target:</i> MCP1, IP10, I-TAC <i>Main findings:</i> no changes	Unclear
Microarrays	Microarray scores and clustering	Higgs, 2014 [236]	39	Sifalimumab [IFN alpha protein]	Pre- and post- treatment (28, 56 and 98 days)	Source: WB Number of ISG: 13 Main findings: changes observed (IFN score neutralized (from 53% to 66%) across the three time points vs placebo, p=0.019)	Unclear
qPCR	qPCR for IFN scores	Greenberg, 2012 [177]	24	Usual care	Over 80 visits (at least 2/patient)	Source: sorted cell populations (monocytes) Number of ISG: 13 Main findings: changes observed (21/24 patients exhibited an elevation of the type I IFN score IFN score, IFI44L and RSAD2 expression	Unclear

						changed in parallel to disease activity over time; 3/24 patients experienced no change in disease activity and exhibited no change in IFN score, IFI44L or RSAD2)	
		Huard, 2017 [209]	13	Usual care	2 or more longitudinal visits/patient	Source: WB Number of ISG: 10 Main findings: changes observed (IFN score changed from baseline correlated changes in CDASI, r=0.280, p=0.060)	High
		Walsh, 2007 [178]	9	Usual care	2 paired samples (active vs improving disease)	Source: WB Number of ISG: 25 Main findings: changes observed (MxA, RSAD2, IFI44L, HERC5, ISG15 and OASL expression decreased with disease improvement; little changes observed in all genes in refractory patients)	High
		O'Connor, 2006 [179]	14	Immunosuppressive therapy (>1 dose of IV methyprednisolone 30 mg/kg, parenteral MTX, folic acid and others)	1 year	Source: WB Number of ISG: 1 Main findings: changes observed (change in MxA expression correlated change in DAS muscle (r=0.630, p=0.040) but not DAS skin (r=0.000, p>0.990))	High
		Reed, 2012 [223]	51	Immunosuppressive regimens (various agents)	Two consecutive visits (interval not specified)	Source: WB Number of ISG: 3 Main findings: changes observed (changes in IFN gene score correlated changes in global (r=0.330, p=0.023) and muscle VAS (r=0.440, p=0.002)	Low
Reporter cell assays	Reporter cell assays by qPCR	Dastmalchi, 2008 [237]	10	Infliximab [TNF protein]	Not specified	Number of ISG: 3 Main findings: changes observed (IFN bioactivity increased upon treatment, regardless of disease improvement, baseline: 1.54 (2.41) vs post-treatment: 3.88 (4.03), p=0.037)	Low

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