

Have Therapeutics Enhanced Our Knowledge of Axial Spondyloarthritis?

S. R. Harrison^{1,2} · H. Marzo-Ortega¹

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Abstract

Purpose of Review An overview of how the treatment landscape of axial spondyloarthritis (axSpA) has shaped our understanding of the disease.

Recent Findings Prior to the millennium, non-steroidal anti-inflammatory drugs (NSAIDs) were the only treatment for axSpA, yet only 30% of patients responded and many developed side effects. In 2003, the first biological disease-modifying drug (bDMARD) was licensed for axSpA which substantially improved outcomes in comparison to NSAIDs. In 2022, there are now several bDMARDs for axSpA; however, they too are not universally efficacious in treating axial inflammation and may have deleterious effects on extramusculoskeletal manifestations. Nevertheless, successful or not, each bDMARD gives invaluable insight into axSpA immunobiology.

Summary This review discusses how much we have learned from the use of bDMARDs in axSpA, how this has redefined our understanding of the disease, and how we might use this knowledge to develop new and better treatments for axSpA in the future.

Keywords Axial spondyloarthritis · Ankylosing spondylitis · Biological disease-modifying antirheumatic drugs (bDMARDs) · Therapeutics

Introduction

Axial spondyloarthritis [axSpA; formerly ankylosing spondylitis(AS)] is an inflammatory arthritis affecting primarily the sacroiliac joints (SIJs) and spine, and is considered to be the prototype of a group of clinically and genetically related diseases called the seronegative spondyloarthropathies (SpA). The wider SpA family now includes radiographic-axSpA (r-axSpA/AS), non-radiographic(nr)-axSpA, psoriatic arthritis (PsA), reactive arthritis and enteropathic arthritis [1]. Clinical features include inflammatory back pain with or without peripheral manifestations (arthritis, enthesitis and dactylitis), and extra-musculoskeletal manifestations [uveitis, psoriasis and

inflammatory bowel disease (IBD)]. Historically, the diagnosis depended on the presence of characteristic bone changes of the sacroiliac joints and/or spine on plain film X-rays [2]; however, in recent decades MRI has enabled the detection of spinal inflammation without bony disease. Accordingly, in 2011, The Ankylosing Spondylitis Association (ASAS) published new classification criteria distinguishing between those with prototypical plain film changes (r-axSpA) and those with only clinical and/or MRI features of the disease (nr-axSpA) [3].

The first-line treatment for axSpA is non-steroidal antiinflammatory drugs (NSAIDs) or cyclooxygenase 2(COX-2) inhibitors [4]; however, approximately a third of patients fail to respond or are intolerant to these agents [5, 6]. In spite of this, NSAIDs/ COX-2 inhibitors remained the only pharmacological options for patients for decades, until 2003, when the first biological disease-modifying anti-rheumatic drug (bDMARD), the TNF α inhibitor (TNFi) etanercept, received marketing authorisation for r-axSpA [7]. Since then, several biologics have been added to the therapeutic arsenal, which now includes five TNFis (etanercept, infliximab, adalimumab, golimumab, certolizumab and their biosimilars), two IL-17A inhibitors (secukinumab and ixekizumab), and two Janus Kinase inhibitors (JAKi) (tofacitinib

H. Marzo-Ortega h.marzo-ortega@leeds.ac.uk

¹ The University of Leeds, Leeds Institute for Rheumatic and Musculoskeletal Medicine (LIRMM), NIHR Leeds Biomedical Research Centre, Leeds Teaching Hospitals Trust, Leeds, UK

² The University of Leeds, Leeds Institute of Cardiovascular and Metabolic Medicine, the LIGHT building, Clarendon Way, Leeds, UK

and upadacitinib) [8–13]. Other agents in development and/ or close to receiving market authorisation include brodalumab (IL-17 receptor blocker) [14], bimekizumab (IL-17A + IL-17F inhibitor) [15–18], namilumab ([granulocytemacrophage colony-stimulating factor (GM-CSF)] inhibitor) [19] and filgotinib (JAKi) [20]. Contrastingly, IL-23 inhibitors were unsuccessful in Phase III trials, despite the positive result in early open-label studies [21–24].

Before reading this review, it is useful for the reader to appreciate some important caveats about drug development and discovery in axSpA. Most early bDMARDs and their molecular targets were conceptualised in other immunemediated inflammatory diseases (IMIDs) and extrapolated to axSpA. The TNFis were first trialled in axSpA on the premise that axSpA might share common inflammatory pathways with rheumatoid arthritis (RA) given the emerging evidence of inflammatory lesions on the SIJ [25]. Molecular studies then followed identifying other cytokines including IL-1 and IL-6 in the serum of axSpA patients [26–28]. Yet, evidence for IL-1 inhibition in axSpA is limited with only two small open-label studies performed in ankylosing spondylitis (r-axSpA) in the mid-2000s [29, 30]. Later, Phase II/III trials of tocilizumab and sarilumab (IL-6is) failed to reach their endpoints [31, 32], despite real-world evidence suggesting that IL-6i may have a role in a subset of synovial-driven refractory spondyloarthritis [33].

Detailed studies of the molecular biology of axSpA came a little later leading to large Phase III trials with agents such as IL-17is, IL-23is and JAKis, opening up new and exciting drug development opportunities specific to this disease [34]. However, it is important to note that the majority of drug development is based on studies of established r-axSpA/AS, derived from pre-clinical models or from studies in other IMIDs; with more work needed to explore differences between these and real-world clinical models [35]. Finally, cellular immunity (the target for all bDMARDs) is only a small piece of the puzzle, since the pathogenesis of axSpA is also shaped by a number of other mechanical, environmental and genetic factors that could not possibly be covered in this focused review on therapeutics [36-38]. Figure 1 provides an overview of the current understanding of the molecular mechanisms of axSpA in relation to existing bDMARDs. The rest of this review details exactly how our understanding of molecular mechanisms in axSpA has been shaped by bDMARDs.

TNF Inhibitors

TNF α is a pleiotropic cytokine with a plethora of direct and indirect effects on both innate and adaptive immunity [39]. The main cellular sources of TNF α in axSpA are monocytes/macrophages, but it may also be produced by natural killer (NK) cells, T cells, neutrophils and tissue-resident non-immune cells such as fibroblasts [40, 41]. In axSpA, TNFα contributes to pathology primarily via modulation of innate immune responses and, to a lesser extent, Th1 and Th17 signalling (Fig. 1) [42-45]. The net effect of these actions is increased production of IL-1, IL-6 and other pro-inflammatory mediators, recruitment of adaptive immune cells (T and B cells) and macrophages, Th1 polarisation of CD4+T cells, tissue inflammation and ultimately propagation of the immune response [44]. In addition, the majority of axSpA patients are positive for the HLA-B27 allele (up to 90% in some studies) [46, 47]. In vitro, axSpA patients have high levels of NK and CD4+T cells expressing KIR3DL2 which is capable of recognising HLA-B27 homodimers expressed on the surface of cells triggering activation and release of IFNy, TNFa and IL-17 [48, 49]. IL-17 works synergistically with TNF α to promote the release of downstream inflammatory mediators and modulate bone metabolism [44, 50], as well as contributing to the pathogenesis of axSpA via other TNF-independent mechanisms (discussed later in this review).

Given the above, it is unsurprising that TNFis are effective for axSpA. However, the response is not ubiquitous, and over a third of patients experience non-response (NR) to their first TNFi [51, 52]. Possible explanations include noncompliance with treatment, individual differences in drug metabolism, pharmacodynamics and pharmacokinetics or the development of anti-drug antibodies [53, 54]. Several studies from a decade ago linked HLA-B27 misfolding with downstream TNF α production [55]; an observation that was then confirmed in a recent meta-analysis showing HLA-B27 positivity was associated with improved BASDAI50 response to TNFis compared with HLA-B27 negative disease [56]. Furthermore, a smaller study showed a significantly improved response in HLA-B27 allele homozygous individuals compared with heterozygous [57]. On the other hand, there have been no clinically significant differences reported for bDMARD response rates between r-axSpA and nr-axSpA for any of the different biologic agents to date (Table 1) [58].

Following TNFi NR, the National Institute of Clinical Excellence (NICE) allows the use of a second TNFi or an IL-17i in axSpA [8]. Whilst some axSpA patients will respond to a second TNFi, overall response rates to a second bDMARD (TNFi or IL-17i) are lower [59], pointing to a possible change in an individual's immune environment, and those rates of NR may be worse for those with primary NR compared with secondary NR. Manica et al. showed no difference in Ankylosing Spondylitis Disease Activity Score Clinically Important Improvement (ASDAS-CII) for the second TNFi between those with primary NR versus secondary NR to their first TNFi; however, there was a difference when the more stringent ASDAS inactive disease (ASDAS-ID) outcome measure was used [60]. The underlying molecular basis for bDMARD NR in axSpA remains poorly characterised.



Fig. 1 The two major inflammatory cell types thought to be implicated in axSpA are Th1 and Th17 CD4+T cells. Naïve T cells are polarised to Th17 cells in the presence of IL-23, IL-6 and TGFβ. IL-23 may be activated in a number of ways including gut microbiota-host dendritic cell interactions, activation of tissue-resident cells due to entheseal stress, unfolded protein response (UPR) in triggered by misfolded HLA-B27, or via IL-36 though the mechanisms of the latter are poorly understood. IL-23R activation triggers downstream JAK/STAT pathway signalling and gene transcription, polarising naïve CD4+T cells to Th17-cells. Th17 cells produce cytokines including IL-17 and IL-22 which drive inflammation. Th1 polarisation is driven by IL-12 which also signals via the JAK/STAT pathway. Th1 cells produce TNFα, IL-6 and IFNγ which promote inflammation.

Another indication supporting that an individual's immunobiology can change with treatment, and/or with the duration of disease, is the observation of NR after rechallenge with the same bDMARD following an interruption to treatment, e.g. for surgery or whilst receiving treatment for a concurrent infection [61]. Infections themselves may prime the immune system and can be a trigger for new autoimmune disease or change in the manifestations of an existing one; the classic example being reactive arthritis [62]. The infectious challenge to the immune system can result in loss of efficacy even in patients with years of good response.

matory pathways involved in axSpA. As discussed above, IL-6 is also required for Th17 cell polarisation. All bDMARDs used in axSpA target one or more of these inflammatory pathways, resulting in direct and indirect inhibition of inflammatory pathways associated with axSpA. Key: ADA, adalimumab; BIM, bimekizumab; BROD, brodalumab; CER, certolizumab; ETA, etanercept; GM-CSF, granulocyte–macrophage colony-stimulating factor; GOL, golimumab; IFNy, interferon-gamma; ILC, innate lymphoid cells; IFX, infliximab; IL, interleukin; IXE, ixekizumab; JAK, janus kinase; p, phosphorylation; SEC, secukinumab; STAT, signal transducers and activators of transcription; TNF α , tumour necrosis factor- alpha; TOF, tofacitinib; UPA, upadacitinib; UPR, unfolded protein response; $\gamma\delta$ T, gammadelta T cells

Given the above, it was suggested that combining bDMARD therapies might be more efficacious in some patients; a theory tested by Hammoura et al. who used a murine model to test whether dual inhibition of IL-17 and TNFi would be superior to TNFi and/or IL-17i. Unfortunately, efficacy was similar across all 3 treatment arms suggesting no additive or synergistic effects of combining different classes of bDMARD treatment [63]. However, all mice were treatment-naïve at the time of study entry, and so the efficacy of dual-blockade in the setting of primary or secondary NR to one or more bDMARDs remains unexplored. Similar results were found in humans where dual inhibition
 Table 1
 Summary of licensed

 bDMARDs in axSpA and their
 efficacy on different disease

 features
 features

bDMARD	Year licensed by NICE	License (r- and nr- axSpA)	Efficacy				
			Axial disease	Peripheral disease	Skin psoriasis	Uveitis	IBD
Etanercept	2003	r-	х	x	x		x
Infliximab	2004	r- and nr-	х	Х	х	x	х
Adalimumab	2008	r- and nr-	х	х	х	х	х
Golimumab	2009	r-	х	х	х	х	х
Certolizumab	2013	r- and nr-	х	х	х	х	х
Secukinumab	2016	r- and nr-	х	х	х		
Ixekizumab	2021	r- and nr-	х	х	х		x**
Upadacitinib		r- and nr-	х	х	х		x**
Tofacitinib		r- and nr-	х	х	х		
Bimekizumab	2022*	r- and nr-					

Key: axSpA axial spondylarthritis, bDMARD biological disease-modifying anti-rheumatic drug, nr non-radiographic, r radiographic

*Pre-authorisation granted full document to be published Aug 2022

**For treatment of ulcerative colitis only

of TNF α and IL-17A with ABT-122 was shown to have a similar efficacy and safety profile to that of the TNFi alone in trials of rheumatoid arthritis and psoriatic arthritis [64] with limited real-life reports of highlighted an enhanced side effect risk profile in resistant SpA [65].

TNFis are not universally efficacious across all EMMs, and in some cases may even exacerbate them. Notably, etanercept has been linked with an increased risk of uveitis [66] as well as other ocular complications including intermediate uveitis, posterior uveitis, scleritis and very rarely orbital myositis [67]. Current recommendations are to avoid etanercept in patients who have or who go on to develop uveitis [66]. Yet paradoxically, other TNFis are the preferred choice of bDMARD for uveitis [68]. The explanation for this differential effect could lie in the unique mode of action of etanercept when compared with other TNFis. Etanercept, a soluble receptor blocker, blocks the receptor but does not remove circulating TNFa like the other TNFi agents on the market [68] and there are some data to suggest its larger molecular weight results in poorer intraocular permeability, reducing efficacy [68, 69]. Indeed, two RCTs failed to show the superiority of etanercept over placebo in the treatment of ocular pathologies [70, 71]. However, this would not account for the paradoxically increased incidence of uveitis that has also been observed in some studies [66]. Perhaps one reason for the difference is that etanercept also blocks TNFβ, however in murine models of uveitis TNFβ levels are also increased, therefore one would expect this to help treat uveitis and not cause it [72]. Another study showed that TNF receptor (TNF-R) and TNFa levels were elevated in ocular fluids from patients with active uveitis, and blocking the TNF-R resulted in an increase in TNFα production by T cell populations. The authors therefore postulate that the TNF-R might have a regulatory role in uveitis over and above just mopping up TNF α [73]. Clearly further research is needed to fully understand the reasons for etanercept failure in the treatment of uveitis [74].

IL-17 Inhibitorsfailure in the treatment of

IL-17 plays an important role in the defence against fungal and certain bacterial pathogens. It was first linked to human disease through a model of mouse autoimmune encephalitis in 2005 [75], heralding the discovery of IL-17 pathway dysregulation in several autoimmune and autoinflammatory diseases including axSpA [76]. The IL-17 family includes 6 known cytokines (IL-17A-IL17F) [77]. The most important in axSpA are IL-17A and IL-17F which form heterodimers capable of activating the IL-17RA and IL-17RC complex on target cells [78]. Several cells are capable of IL-17 including CD4 + T cells, CD8+T cells, mucosal-associated invariant T cells (MAIT cells), innate lymphoid cells (ILCs), gamma delta T cells $(\gamma \delta T$ -cells) and invariant natural killer T cells (iNKTs), neutrophils, mast cells and eosinophils [78]. Although a number of these cells may play a role in IL-17 production in axSpA, most IL-17 is thought to be produced by Th17 cells [79–81]. IL-17 production, in turn, stimulated IL-1 β , TNF α , IL-6 and IL-23 by synovial fibroblasts, monocytes and macrophages generating a positive feedback loop for further Th17 cell differentiation [82]. In addition to IL-17, Th17 polarised cells produce other cytokines (GM-CSF), chemokines (CXCL1, XCL2, CXCL8, CCL20, etc.), antimicrobial peptidases, matrix metalloproteinases, complement and other acute phase reactants [83, 84]. This pro-inflammatory storm ultimately results in dysregulated bone metabolism, axial and peripheral joint inflammation and enthesitis in both mouse models and in-vitro studies of peripheral blood and tissues from patients with axSpA [78, 85].

Presently, two IL-17A inhibitors, secukinumab and ixekizumab, have been licensed by NICE, the Medicines and Healthcare products Regulatory Agency (MHRA) and Food and Drug Administration (FDA) for use in axSpA either as 1st or 2nd line treatment [9, 10, 86-89]. Bimekizumab (an IL-17A and IL-17F inhibitor) has also now received preauthorisation [16] with results from Phase III trials showing similar efficacy to TNFi (Table 1) [90]. Although to date, there are no head-to-head comparator studies in axSpA, adalimumab was used as a comparator in the COAST-V trial, and ixekizumab demonstrated a greater absolute improvement in ASAS-40 scores between week 0 and week 16 compared to adalimumab, although the study was not powered to detect significance [91]. Despite largely similar efficacy on disease control overall, there are some important differences with regards to EMMs. IL-17 inhibition showed superior efficacy on skin psoriasis compared with TNFis, and therefore axSpA patients with difficult-to-treat skin psoriasis may warrant earlier used IL-17is as opposed to a TNFis [92]. Similarly, psoriatic arthritis with a predominantly axial pattern of joint involvement may benefit from the early introduction of IL-17i because conventional non-biological DMARDs, although effective for skin psoriasis and peripheral joint inflammation, may not effectively treat axial disease [93–95].

Changes of axSpA are present in 10–36% of IBD patients, therefore IBD is of major clinical significance in axSpA [96]. TNF α is are effective treatment for IBD [97]. However, trials have demonstrated an increased incidence of IBD, as well as exacerbation of IBD, in axSpA patients treated with IL-17i [98]. The same observations were made in studies of IL-17 is in non-axSpA IBD, with two clinical trials of the IL-17 is secukinumab and brodalumab stopped early due to worsening of IBD symptoms in Crohn's disease patients [99, 100]. These results perplexed experts initially as earlier studies preclinical and genetic studies had demonstrated dysregulation of the IL-17/23 axis in IBD [98]. However, studies have since also shown that IL-17 inhibition may interfere with dysregulation of the gut epithelial barrier, predisposing to infection and inflammation which can, in turn, exacerbate or trigger IBD [98]. Additionally, in a murine model of autoimmune uveitis, authors demonstrated IL-17A may be important for negative feedback of pro-inflammatory Th17 cell responses. They showed that IL-17A, through NFκβ, induces IL-24 production by Th17 cells, thereby downregulating IL-17F and GM-CSF to suppress Th17 cell activity [101]. On the other hand, there is evidence that the effects of IL-24 are dosedependent, with low doses suppressing Th1 cells and higher doses promoting Th1 and Th17 cell activity in the study of colorectal cancer [102]. IL-24 also acts on the IL-20 receptors on epithelial cells in the colonic mucosa where it plays a regulatory role [103] and high levels of IL-24 have also been observed in patients with skin psoriasis and in RA synovial fluid [103]. Altogether, these studies suggest a differential role for IL-17A, IL-17F and their downstream cytokines in the inflammatory process which may be cell- and tissue-specific, and may account for the lack of efficacy of IL-17is in IBD. It also suggests that dual IL-17A/IL-17F inhibition may have other effects, and therefore that agents like bime-kizumab may behave differently to agents that block IL-17A only, however more work is needed in this area [104].

IL-23 Inhibitors

IL-23 is another cytokine which works in conjunction with IL-17 in the pathogenesis of axSpA [105]. Structurally, IL-23 is a heterodimer complex of two subunits, p40 and p19, which when combined interact with the IL-23R on target cells triggering downstream activation of JAK/STAT signalling pathways and transcription of several pro-inflammatory mediators including IL-17, IL-22 and TNFα [106, 107]. Several animal studies demonstrate the importance of IL-23 in the pathogenesis of SpA [108–113]. Moreover, IL-23R and STAT2/3 risk alleles were reported amongst the non-HLA-B27 associations for axSpA in a recent meta-analysis of ankylosing spondylitis genome-wide association studies [114, 115]. Although individual risk effect estimates were small, this is true for all non-HLA-B27 risk alleles and does not necessarily mean they are not relevant to disease in at least some patients, particularly given that functional studies of the role of IL-23 in axSpA suggest otherwise. IL-23 and IL-17 levels are elevated in peripheral blood from AS patients [116-118], and IL-23 can stimulate IL-17 production by a number of cells including CD8 + T cells, $\gamma\delta$ T-cells as well as other lymphoid cell lines [83-85].

Given these findings, and the success of four IL-23 inhibitors (the p19IL-23 inhibitor risankizumab, the p40IL-12/23 inhibitor ustekinumab, the p19IL-23 inhibitor IL-23 inhibitor guselkumab and the p19IL-23 inhibitor tildrakizumab) in treating patients with skin psoriasis and/or PsA [119–129], many thought IL-23 inhibitors would be a viable treatment option in axSpA. Yet despite early promising results from a German open-label study of IL-23 inhibitors, Phase III trials failed to demonstrate efficacy [21–24]. These trials used comparable patient groups and similar endpoints to other bDMARD trials in axSpA, and therefore, lack of efficacy cannot be accounted for simply on the basis of trial design [105].

To understand the possible cause for the lack of efficacy of IL-23 inhibitors in axSpA requires a deeper understanding of the IL-17/IL-23 signalling pathway and differences that might exist in different tissue compartments and at different time points in the disease trajectory [130]. Recent studies on the spinal human enthesis have shown evidence of IL-17A production independent of IL-23R expression [131]. Furthermore, there are myeloid cells resident in the entheses of the spine capable of producing IL-23 in response to mechanical stress factors [132]. Other potential sources of IL-23 in axSpA that have been proposed include, cellular endoplasmic reticulum stress responses in relation to misfolded HLA-B27 proteins, aberrant host response to a microbial insult (as observed in reactive arthritis patients) and IL-36 production, although there is very little research on the latter [133]. Yet robust evidence that any of these pathways are clinically important and are active in-vivo, is yet to be obtained. Moreover, whilst these theories offer an explanation to the original source of IL-23 production, in established disease, data suggest that these may no longer be important, since Th17 cells can produce IL-17 independent of IL-23 as discussed above. Furthermore, it raises the question of whether IL-23 inhibition may only be of value in the very early or pre-clinical disease state, which has yet to be defined in axSpA [134].

Alternatively, it is possible that there is a subset of patients who continue to show a IL-23-driven disease who might benefit from IL-23is, or that certain EMMs are more driven by IL-23 than others owing to tissue-specific factors. Of even more significant interest is perhaps the fact that IL-23i, somewhat counterintuitively may be efficacious in axial psoriatic arthritis. Post hoc analysis of the axial PsA patient subsets from larger PsA trials of IL-23 inhibitors (PSUM-MIT, DISCOVER1 and DISCOVER2) showed patients still felt joint symptoms had improved [105, 120, 124, 125]. The caveat to this is that all primary outcome measures used in the trials measured disease activity as a whole, which includes peripheral disease, skin psoriasis and axial disease. Furthermore, no MRI imaging was available to correlate symptomatic improvement with objective evidence of a reduction in spine/SIJ inflammation on spinal MRI. Nevertheless, this observation is of great interest and there is a need for dedicated trials of IL-23i in axial PsA focusing on their impact on axial disease specifically.

JAK Inhibitors

Cytokines signal via numerous downstream pathways, one of which is the Janus Kinase/ signal transducers and activators of transcription (JAK/STAT) [135]. The JAK/STAT family includes JAK 1,2,3 and tyrosine kinase 2 (TYK2), which is associated with the intracellular aspect of type I/II cytokine receptors on the cell surface, including the IL-23R, IL-6R, type 1 and type 2 IFN receptors, IL-7R and GM-CSF receptors. When activated, JAKs phosphorylate themselves and their receptors, then dephosphorylate after 15-30 min to prevent permanent receptor activation. STAT molecules bind during the active state and are phosphorylated, whereupon they migrate to the nucleus and bind to target genes promoting transcription. JAKis in commercial use have variable selectivity for particular JAK family members, allowing for some degree of differential receptor modulation [136]. The two JAKis currently licensed for the treatment of axSpA; upadacitinib and tofacitinib; selectively target JAK1, but also may interact with other JAKs, therefore have a broad spectrum of activity on the receptor subtypes above. The downstream effect is the modulation of several innate and adaptive immune processes contributing to axSpA including Th1 and Th17 differentiation, growth/maturation of lymphoid cells, and tissue inflammation. A detailed review of the specific downstream effects of JAKis can be found here [137].

JAKis demonstrated clear efficacy for the treatment of axSpA in trials meeting their primary and secondary endpoints with similar outcomes as reported in trials of TNFi and IL-17is [138–140]. However, as JAKis have only just been authorised for the treatment of axSpA, more data are needed to establish their efficacy against EMM such as IBD and AU (Table 1) and to fully understand their mechanism(s) of effect in axSpA. JAKis target several inflammatory pathways, but not all are directly involved in axSpA, therefore the mechanism of their effect on disease is likely indirect. With this comes the risk of further unwanted and/or unanticipated side effects. Indeed, this has already been observed in RA, with emerging real-world clinical data demonstrating an increased risk of cardiovascular and thrombotic events in the RA population. Accordingly, JAKis should not be used in those at high risk for thrombosis or those over the age of 65 [141, 142]. On the other hand, JAKis have been in use in RA for a number of years with no apparent increase in incidence of IBD, psoriasis or uveitis [143]. Furthermore, tofacitinib and upadacitinib are actually approved by the FDA, MHRA and NICE for the treatment of ulcerative colitis [141, 142, 144–147]. Upadacitinib but not tofacitinib, has shown favourable results in Crohn's disease, although regulatory approval for use in Crohn's disease has yet to be granted [148, 149]. Nevertheless, these studies suggest that, like TNFis, JAKi might have beneficial effects on both the joints and gut of patients with axSpA and IBD [150]. JAKis have been trialled in psoriatic arthritis, but not skin psoriasis specifically. Despite this, trial data suggested that the improvement in the psoriatic arthritis severity index (PASI) reflected the beneficial effect of JAKis on both arthritis and skin psoriasis [136].

Potential Future bDMARDs

As described previously, new bDMARDs are in late-phase development, including a novel IL-17R blocker brodalumab, with a different mode of action of IL-17 direct cytokine inhibition [14]. Similarly, other JAKi may soon be licensed, and it will be interesting to see if/how their different receptor selectivity impact on the efficacy of treatment. Finally, there is also a new class of agent in early trial phases, nanilumab, a GM-CSF inhibitor (19). The role of GM-CSF was first suggested as working upstream of the IL-17/23 inhibitor axis in triggering the disease, and early-phase trials have shown some success in axSpA and RA [151]. If trials are successful, this could one day become another class of bDMARD for axSpA.

Summary

There are now a significant array of biologic and targeted synthetic drugs available for the treatment of axSpA, each with somewhat divergent effects on axial/peripheral joint symptoms and EMMs. TNFis show efficacy for the treatment of axial/peripheral disease, enthesitis, IBD and uveitis (except for etanercept). IL-17 inhibitors treat axial/peripheral joint symptoms and are particularly useful in the presence of concomitant skin psoriasis. On the other hand, they may trigger or exacerbate flares of IBD and are second to TNFis for the treatment of uveitis. JAKis have established efficacy on peripheral and axial symptoms in trials but effects on EMMs are as yet unknown. How and why these differences might exist between drugs, and how this divergence relates to the underlying molecular pathogenesis, remains unclear. Specific studies in pre-clinical and early disease may now be possible with increased awareness of the disease, advancing imaging methods and the growing use of predictive genomics technologies across the whole spectrum of IMIDs, and may reveal new drug targets that can tell us more about the disease in its early stages. At the same time, results of ongoing pharmacovigilance studies of existing drugs over the coming years may reveal new insights into how these treatments affect the pathogenesis of axSpA in the short- and long-term, which may further change our understanding of the biology of this disease throughout its course. Even more valuable will be the data on how they impact particular subsets of patients (nr- vs r-, HLA-B27 allele positive versus negative, males vs. females, etc.) which may account for some differences in the observed treatment efficacy. Twenty years of biologics for axSpA have already taught us a lot about this disease. With well-planned drug development and post-market surveillance, the next 20 years are likely to teach us even more.

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Author Contribution SRH performed the literature search and wrote the manuscript and associated tables/figures. HMO provided expert support and critical review during the manuscript preparation and writing and also edited all sections of the manuscript.

Data Availability Not applicable

Declarations

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References

- McAllister K, Goodson N, Warburton L, Rogers G. Spondyloarthritis: diagnosis and management: summary of NICE guidance. Br Med J. 2017;356:j839.
- Khmenlinskii N, Regel A, Baraliakos X. The role of imaging in diagnosing axial spondyloarthritis. Front Med. 2018;5:106.
- Rudwaleit M, van der Heijde D, Landewé R, Akkoc N, Brandt J, Chou CT, et al. The Assessment of SpondyloArthritis International Society classification criteria for peripheral spondyloarthritis and for spondyloarthritis in general. Ann Rheum Dis. 2011;70(1):25–31.
- van der Heijde D, Ramiro S, Landewé R, Baraliakos X, Van den Bosch F, Sepriano A, et al. 2016 update of the ASAS-EULAR management recommendations for axial spondyloarthritis. Ann Rheum Dis. 2017;76(6):987–91.
- Baraliakos X, Kiltz U, Peters S, Appel H, Dybowski F, Igelmann M, et al. Efficiency of treatment with non-steroidal anti-inflammatory drugs according to current recommendations in patients

with radiographic and non-radiographic axial spondyloarthritis. Rheumatol. 2017;56(1):95–102.

- Baraliakos X, Kiltz U, Peters S, Appel H, Dybowski F, Igelmann M, et al. Efficiency of treatment with non-steroidal anti-inflammatory drugs according to current recommendations in patients with radiographic and non-radiographic axial spondyloarthritis. Rheumatol. 2017;56(1):95–102.
- Gorman JD, Sack KE, Davis JC Jr. Treatment of ankylosing spondylitis by inhibition of tumor necrosis factor alpha. N Engl J Med. 2002;346(18):1349–56.
- National Institute of Clinical Excellence. TNF-inhibitors for ankylosing spondylitis and non-radiographic axial spondyloarthritis [Internet]. London. 2016 [cited 2021 May 1]. Available from: https://www.nice.org.uk/guidance/ta383/resources/tnfal pha-inhibitors-for-ankylosing-spondylitis-and-nonradiographicaxial-spondyloarthritis-pdf-2984675907013
- National Institute of Clinical Excellence (NICE). Secukinumab for treating non-radiographic axial spondyloarthritis [TA719] [Internet]. London. 2021 [cited 2022 Aug 10]. Available from: https://www.nice.org.uk/guidance/ta719
- National Institute of Clinical Excellence (NICE). Ixekizumab for treating axial spondyloarthritis [TA718] [Internet]. London. 2021 [cited 2022 Aug 10]. Available from: https://www.nice. org.uk/guidance/ta718
- National Institute of Clinical Excellence (NICE). Upadacitinib for treating active ankylosing spondylitis [ID3848] [Internet]. London. 2022 [cited 2022 Aug 10]. Available from: https:// www.nice.org.uk/guidance/indevelopment/gid-ta10735
- National Institute of Clinical Excellence (NICE). Tofacitinib for treating active ankylosing spondylitis [ID3865] [Internet]. London. 2022 [cited 2022 Aug 10]. Available from: https:// www.nice.org.uk/guidance/indevelopment/gid-ta10771
- Braun J, Brandt J, Listings J, Zink A, Alten R, Golder W, et al. Treatment of active ankylosing spondylitis with infliximab: a randomised controlled multicentre trial. Lancet. 2002;359(9313):1187–93.
- 14. Wei JC, Kim T, Kishimoto M, Ogusu N, Jeong H, Kobayashi S, et al. Efficacy and safety of brodalumab, an anti-IL17RA monoclonal antibody, in patients with axial spondyloarthritis: 16-week results from a randomised, placebo-controlled, phase 3 trial. Ann Rheum Dis. 2021;80(8):1014–21.
- UCB Biopharma SRL. A study to evaluate the efficacy and safety of bimekizumab in subjects with active nonradiographic axial spondyloarthritis (BE MOBILE 1) [Internet]. Bethsada. 2019 [cited 2022 Aug 10]. Available from: https://www.clini caltrials.gov/ct2/show/NCT03928704
- UCB Biopharma SRL. A study to evaluate the efficacy and safety of bimekizumab in subjects with active ankylosing spondylitis (BE MOBILE 2) [Internet]. Bethsada. 2019 [cited 2022 Aug 10]. Available from: https://www.clinicaltrials.gov/ct2/ show/NCT03928743
- 17. van der Heijde D, Genslet LS, Doedhar A, Baraliakos X, Poddubnyy D, Kivitz A, et al. Dual neutralisation of interleukin-17A and interleukin-17F with bimekizumab in patients with active ankylosing spondylitis: results from a 48-week phase IIb, randomised, double-blind, placebo-controlled, dose-ranging study. Ann Rheum Dis. 2020;79(5):595–604.
- UCB Biopharma S.P.R.L. A study to evaluate the long-term safety, tolerability and efficacy of bimekizumab in subjects with active axial spondyloarthritis including ankylosing spondylitis and nonradiographic axial spondyloarthritis (BE MOV-ING) [Internet]. Bethesda. 2020 [cited 2021 Apr 21]. Available from: https://clinicaltrials.gov/ct2/show/NCT04436640
- Izana Bioscience Ltd. Efficacy and safety of namilumab for moderate-to-severe axial spondyloarthritis (NAMASTE)

[Internet]. Bethsada. 2022 [cited 2022 Aug 10]. Available from: https://clinicaltrials.gov/ct2/show/results/NCT03622658

- 20. van der Heijde D, Baraliakos X, Gensler LS, Maksyowych WP, Tseluyko V, Nadashkevich O, et al. Efficacy and safety of filgotinib, a selective Janus kinase 1 inhibitor, in patients with active ankylosing spondylitis (TORTUGA): results from a randomised, placebo-controlled, phase 2 trial. Lancet. 2018;392(10162):2378–87.
- Deodhar A, Gensler LS, Sieper J, Clark M, Calderon C, Wang Y, et al. Three Multicenter, Randomized, Double-Blind, Placebo-Controlled Studies Evaluating the Efficacy and Safety of Ustekinumab in Axial Spondyloarthritis. Arthritis Rheumatol. 2019;71(2):258–70.
- Baeten D, Østergaard M, Wei JC, Sieper J, Järvinen P, Tan L, et al. Risankizumab, an IL-23 inhibitor, for ankylosing spondylitis: results of a randomised, double-blind, placebo-controlled, proof-of-concept, dose-finding phase 2 study. Ann Rheum Dis. 2018;77(9):1295–302.
- 23. Mease PJ, Chohan S, Fructuoso FJG, Luggen ME, Rahman P, Raychaudhuri SP, et al. Efficacy and safety of tildrakizumab in patients with active psoriatic arthritis: results of a randomised, double-blind, placebo-controlled, multiple-dose, 52-week phase IIb study. Ann Rheum Dis. 2021;80(9):1147–57.
- Poddubnyy D, Hermann KA, Callhoff J, Listings J, Sieper J. Ustekinumab for the treatment of patients with active ankylosing spondylitis: results of a 28-week, prospective, open-label, proofof-concept study (TOPAS). Ann Rheum Dis. 2014;73(5):817–23.
- Braun J, Bollow M, Eggens U, König H, Distler A, Sieper J. Use of dynamic magnetic resonance imaging with fast imaging in the detection of early and advanced sacroiliitis in spondylarthropathy patients. Arthritis rheum. 1994;37(7):1039–45.
- 26. Bal A, Unlu E, Bahar G, Aydog E, Elsioglu E, Yorgancioglu R. Comparison of serum IL-1 beta, sIL-2R, IL-6, and TNF-alpha levels with disease activity parameters in ankylosing spondylitis. Clin Rheumatol. 2007;26(2):211–5.
- Liu W, Wu Y, Zhang L, Xue B, Wang YBL, et al. Elevated serum levels of IL-6 and IL-17 may associate with the development of ankylosing spondylitis. Int J Clin Exp Med. 2015;8(10):17362–76.
- Pedersen SJ, Hetland ML, Sørensen IJ, Ostergaard M, Nielsen HJ, Johansen JS. Circulating levels of interleukin-6, vascular endothelial growth factor, YKL-40, matrix metalloproteinase-3, and total aggrecan in spondyloarthritis patients during 3 years of treatment with TNFα inhibitors. Clin Rheumatol. 2010;29(11):1301–9.
- Tan AL, Marzo-Ortega H, O'Connor P, Fraser A, Emery P, McGonagle D. Efficacy of anakinra in active ankylosing spondylitis: a clinical and magnetic resonance imaging study. Ann Rheum Dis. 2004;63(9):1041–5.
- Haibel H, Rudwaleit M, Listing J, Sieper J. Open label trial of anakinra in active ankylosing spondylitis over 24 weeks. Ann Rheum Dis. 2005;64(2):296–8.
- Sieper J, Braun J, Kay J, Badalamenti S, Radin AR, Jiao L, et al. Sarilumab for the treatment of ankylosing spondylitis: results of a phase II, randomised, double-blind, placebo-controlled study (ALIGN). Ann Rheum Dis. 2015;74(6):1051–7.
- 32. Sieper J, Porter-Brown B, Thompson L, Harari O, Dougados M. Assessment of short-term symptomatic efficacy of tocilizumab in ankylosing spondylitis: results of randomised, placebo-controlled trials. Ann Rheum Dis. 2014;73:1051–7.
- Meralshi M, De Marco G, Podgorski M, McGonagle D, Marzo-Ortega H. Evidence of response to IL-6 inhibition in some cases of refractory spondyloarthritis-associated peripheral synovitis. Ann Rheum Dis. 2016;75(7):209275.

- Baeten DL, Kuchroo VK. How Cytokine Networks Fuel Inflammation: Interleukin-17 and a tale of two autoimmune diseases. Nat Med. 2013;19:824–5.
- Marzo-Ortega H. Axial spondyloarthritis: coming of age. Rheumatol. 2020;59(suppl4):iv1-5.
- Brown MA, Li Z, Cao KL. Biomarker development for axial spondyloarthritis. Nat Rev Rheumatol. 2020;16(8):448–63.
- 37. Gracey E, Vereecke L, McGovern D, Frohling M, Schett G, Danese S, et al. Revisiting the gut–joint axis: links between gut inflammation and spondyloarthritis. Nat Rev Rheumatol. 2020;16:415–33.
- Gracey E, Burssens A, Cambre I, Scheet G, Lories R, McInnes IB, et al. Tendon and ligament mehanical loading in the pathogenesis of inflammatory arthritis. Nat Rev Rheumatol. 2020;16:193–207.
- Holbrook J, Lara-Reyna S, Jarosz-Griffiths H, McDermott M. Tumour necrosis factor signalling in health and disease. F1000Res. 2019;8:111.
- Garriets V, Goyal A, Khaddour K. Tumour Necrosis Factor Inhibitors. Florida: StatPearls Publishing; 2021. p. 1–14.
- 41. Tracey D, Klareskog L, Sasso EH, Salfeld JG, Tak PP. Tumor necrosis factor antagonist mechanisms of action: a comprehensive review. Parmacology Ther. 2008;117(2):244–79.
- 42. Menegatti S, Guillemot V, Latis E, Yahia-Cherbal H, Mittermüller D, Rouilly V, et al. Immune response profiling of patients with spondyloarthritis reveals signalling networks mediating TNF-blocker function in vivo. Ann Rheum Dis. 2021;80(4):475–86.
- 43. Bautista-Caro M, Arroyo-Villa I, Castillo-Gallego C, Miguel E, Peiteado D, Puig-Kröger A, et al. Decreased Th17 and Th1 cells in the peripheral blood of patients with early non-radiographic axial spondyloarthritis: a marker of disease activity in HLA-B27⁺ patients. Rheumatol (Oxford). 2013;52(2):352–62.
- Garcia-Montoya L, Emery P. Disease modification in ankylosing spondylitis with TNF inhibitors: spotlight on early phase clinical trials. Expert Opin Investig Drugs. 2021;30(11):1109–24.
- Lata M, Hettinghouse AS, Liu C. Targeting tumor necrosis factor receptors in ankylosing spondylitis. Ann N Y Acad Sci. 2019;1442(1):5–16.
- 46. Kishimoto M, Ono K, Fukui S, Kawaai S, Deshpande GA, Yoshida K, et al. Clinical characteristics of non-radiographic versus radiographic axial spondyloarthritis in Asia and nonradiographic axial spondyloarthritis in other regions: results of the cross-sectional ASAS-COMOSPA study. RMD Open. 2021;7(3):e001752.
- 47. Onna MV, Jurik AG, van der Heijde D, van Tubergen A, Heuft-Dorenbosch L, Landewé R. HLA-B27 and gender independently determine the likelihood of a positive MRI of the sacroiliac joints in patients with early inflammatory back pain: a 2-year MRI follow-up study. Ann Rheum Dis. 2011;70(11):1981–5.
- 48. Wong-Baeza I, Rodley A, Shaw J, Hatano H, Rysnik O, McHugh K, et al. KIR3DL2 binds to HLA-B27 dimers and free H chains more strongly than other HLA class I and promotes the expansion of T cells in ankylosing spondylitis. J Immunol. 2013;190(7):3216–24.
- 49. Bowness P, Ridley A, Shaw J, Chan AT, Wong-Baeza I, Fleming M, et al. Th17 cells expressing KIR3DL2+ and responsive to HLA-B27 homodimers are increased in ankylosing spondylitis. J Immunol. 2011;186(4):2672–80.
- 50. Gravallese EM, Schett G. Effects of the IL-23–IL-17 pathway on bone in spondyloarthritis. Nat Rev Rheumatol. 2018;14(11):631–40.
- Moral E, Plasencia C, Navarro-Compán C, Pascual Salcedo D, Jurado T, Tornero C, et al. AB0657 discontinuation of anti-TNF therapy in patients with axial spondyloarthritis in

clinical practice: Prevalence and Causes. Ann Rheum Dis. 2016;75(Suppl2):1129.

- 52. Mease PJ, van der Heijde D, Karki CML, Park Y, Greenberg JD. Tumor necrosis factor inhibitor discontinuation in patients with ankylosing spondylitis: an observational study from the US-based Corrona registry. Rheumatol Ther. 2018;5(2):537–50.
- 53. Atiqi S, Hooljberg F, Loeff FC, Rispens T, Wolbink GJ. Immunogenicity of TNF-inhibitors. Front Immunol. 2020;11:312.
- Menegatti S, Bianchi E, Rogge L. Anti-TNF therapy in spondyloarthritis and related diseases, impact on the immune system and prediction of treatment responses. Front Immunol. 2019;10:382.
- 55. Glintborg B, Ostergaard M, Krogh NS, Dreyer L, Kristensen HL, Hetland ML. Predictors of treatment response and drug continuation in 842 patients with ankylosing spondylitis treated with anti-tumour necrosis factor: results from 8 years' surveillance in the Danish nationwide DANBIO registry. Ann Rheum Dis. 2010;69(11):2002–8.
- 56. Maneiro JR, Souto A, Salgado E, Mera A, Gomez-Reino JJ. Predictors of response to TNF antagonists in patients with ankylosing spondylitis and psoriatic arthritis: systematic review and meta-analysis. RMD Open. 2015;1(1):e000017.
- 57. Marzo-Ortega H, Poddubnyy D, Pournara E, Schulz B, Deodhar A, Baraliakos X. OA35 Efficacy of secukinumab and HLA-B27 subtypes: results from a Phase 3b randomised controlled trial in axial SpA. Rheumatol. 2022;61(suppl1):keac132.035.
- 58. Ciurea A, Kissling S, Bürki K, Baraliakos X, de Hooge M, Hebeisen M, et al. Current differentiation between radiographic and non-radiographic axial spondyloarthritis is of limited benefit for prediction of important clinical outcomes: data from a large, prospective, observational cohort. RMD Open. 2021;8(1):e002067.
- Navarro-Compán V, Plasencia-Rodríguez C, Miguel E, Del Campo PD, Balsa A, Gratacós J. Switching biological diseasemodifying antirheumatic drugs in patients with axial spondyloarthritis: results from a systematic literature review. RMD Open. 2017;3(2):e000524.
- 60. Manica SR, Sepriano A, Pimentel-Santos F, Gouveia N, Barcelos A, Branco JC, et al. Effectiveness of switching between TNF inhibitors in patients with axial spondyloarthritis: is the reason to switch relevant? Arthritis Res Ther. 2020;22(195)
- 61. Lamberg T, Sipponen T, Valtanen S, Eklund KK, Mälkönen T, Aalto K, et al. Short interruptions of TNF-inhibitor treatment can be associated with treatment failure in patients with immunemediated diseases. Autoimmunity. 2022;55(5):275–84.
- 62. Inman RD. Mechanisms of Disease: infection and spondyloarthritis. Nat Rev Rheumatol. 2006;2(3):163–9.
- 63. Hammoura I, Fiechter RH, Bryant SH, Westmoreland S, Kingsbury G, Waegell W, et al. Dual blockade of TNF and IL-17A inhibits inflammation and structural damage in a rat model of spondyloarthritis. Int J Mol Sci. 2022;23(2):859.
- 64. Mease P, Genovese M, Weinblatt M, Peloso P, Chen K, Li Y. Safety and efficacy of ABT-122, a TNF and IL-17–targeted dual variable domain (DVD)–IgTM, in psoriatic arthritis patients with inadequate response to methotrexate: results from a phase 2 trial. Arthritis Rheumatol. 2018;70(11):1710–20.
- 65. De Marco G, McGonagle D, Matheison HR, Meralshi M, Magee C, FitzGerald O, et al. Combined inhibition of tumour necrosis factor-alpha and interleukin-12/23 for long-standing, refractory psoriatic disease: a differential role for cytokine pathways? Rheumatol. 2018;57(11):2053–5.
- 66. Lie E, Lindström U, Zverkova-Sandström T, Olsen IC, Forsbladd'Elia H, Askling J, et al. Tumour necrosis factor inhibitor treatment and occurrence of anterior uveitis in ankylosing spondylitis: results from the Swedish biologics register. Ann Rheum Dis. 2017;76(9):1515–21.

- 67. Susanna FN, Pavesio C. A review of ocular adverse events of biological anti-TNF drugs. J Ophthalmic Inflamm Infect. 2020;10(1):11.
- 68. Jiang Q, Li Z, Tao T, Duan R, Wang X, Su W. TNF-α in uveitis: from bench to clinic. Front Pharmacol. 2021;12:740057.
- 69. Dick AD, Rosenbaum JT, Al-Dhibi HA, Belfort R Jr, Brézin AP, Chee SP, et al. Guidance on noncorticosteroid systemic immunomodulatory therapy in noninfectious uveitis: Fundamentals Of Care for UveitiS (FOCUS) Initiative. Ophthalmology. 2018;125(5):757–73.
- Baughman RP, Lower EE, Bradley DA, Raymond LA, Kaufman A. Etanercept for refractory ocular sarcoidosis: results of a double-blind randomized trial. Chest. 2005;128(2):1062–1047.
- Foster CS, Tufail F, Waheed NK, Chu D, Miserocchi E, Baltatzis S, et al. Efficacy of etanercept in preventing relapse of uveitis controlled by methotrexate. Arch Ophthalmol (Chigago, III 1960). 2003;121(4):437–40.
- Savion S, Oddo S, Grover S, Caspi RR. Uveitogenic T lymphocytes in the rat: pathogenicity vs lymphokine production, adhesion molecules and surface antigen expression. J Neuroimmunol. 1994;55(1):35–44.
- Sugita S, Takase H, Taguchi C, Mochizuki M. Role of soluble TNF receptors for TNF-a in uveitis. Investig Ophthalmol Vis Sci. 2007;48(7):3246–52.
- Fabiani C, Vitale A, Lopalco G, Iannone F, Frediani B, Cantarini L. Different roles of TNF inhibitors in acute anterior uveitis associated with ankylosing spondylitis: state of the art. Clin Rheumatol. 2016;35(11):2631–8.
- Langrish CL, Chen Y, Blumenschein M, Mattson J, Basham B, Sedgwick JD, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. J Exp Med. 2005;201(2):233–40.
- Ţiburcă L, Bembea M, Zaha DC, Jurca AD, Vesa CM, Rațiu IA, et al. The treatment with interleukin 17 inhibitors and immune-mediated inflammatory diseases. Curr Issues Mol Biol. 2022;44(5):1851–66.
- McGeachy MJ, Cua DJ, Gaffen SL. The IL-17 family of cytokines in health and disease. Immunity. 2019;50(4):892–906.
- Rosine N, Miceli-Richard C. Innate Cells: The alternative source of IL-17 in axial and peripheral spondyloarthritis? Front Immunol. 2021;11:553742.
- Jandus C, Bioley G, Pivals J, Dudler J, Speiser D, Romero P. Increased numbers of circulating polyfunctional Th17 memory cells in patients with seronegative spondylarthritides. Arthritis Rheum. 2008;58(8):2307–17.
- Shen H, Goodall JC, Hill Gaston JS. Frequency and phenotype of peripheral blood Th17 cells in ankylosing spondylitis and rheumatoid arthritis. Arthritis Rheum. 2009;60(6):1647–56.
- Zhang L, Li Y, Li Y, Lei Q, Liu X, Yuan C, et al. Increased frequencies of Th22 cells as well as Th17 cells in the peripheral blood of patients with ankylosing spondylitis and rheumatoid arthritis. PLoS ONE. 2012;7(4):e31000.
- Tesmer LA, Lundy SK, Sarkar S, Fox DA. Th17 cells in human disease. Immunol Rev. 2008;223(1):87–113.
- Gaffen SL. An overview of IL-17 function and signalling. Cytokine. 2008;43(3):402–7.
- Onishi RM, Gaffen SL. Interleukin-17 and its target genes: mechanisms of interleukin-17 function in disease. Immunology. 2010;129(3):311–21.
- Groen SS, Sinkeviciute D, Bay-Jensen A, Thudium CS, Karsdal MA, Thomsen SF, et al. Exploring IL-17 in spondyloarthritis for development of novel treatments and biomarkers. Autoimmune Rev. 2021;20(3):102760.
- Wexler M. FDA approves Taltz for non-radiographic axial spondyloarthritis [Internet]. Axial Spondyloarthritis News. 2020 [cited 2021 May 5]. Available from: https://creakyjoints.org/treat

ment/taltz-ixekizumab-approved-non-radiographic-axial-spond yloarthritis/#:~:text=Lilly's Taltz (ixekizumab) is the,June 1%2C 2020.

- European Medicines Agency. Cosentyx [Internet]. Amsterdam. 2021 [cited 2021 Jun 29]. Available from: https://www.ema. europa.eu/en/medicines/human/EPAR/cosentyx
- European Medicines Agency. Taltz [Internet]. Amsterdam. 2021 [cited 2021 Jun 29]. Available from: https://www.ema.europa.eu/ en/medicines/human/EPAR/taltz#assessment-history-section
- Food and Drug Administration. Taltz highlights of prescribing information [Internet]. Maryland. 2020 [cited 2022 Aug 19]. Available from: https://www.accessdata.fda.gov/drugsatfda_ docs/label/2021/125521s014lbl.pdf
- Harrison SR, Marzo-Ortega H. Ixekizumab: an IL-17A inhibitor for the treatment of axial spondylarthritis. Expert Rev Clin Immunol. 2021;17(10):1059–71.
- 91. van der Heijde D, Cheng-Chung Wei J, Dougados M, Mease P, Deodhar A, Maksymowych WP, et al. Ixekizumab, an interleukin-17A antagonist in the treatment of ankylosing spondylitis or radiographic axial spondyloarthritis in patients previously untreated with biological disease-modifying anti-rheumatic drugs (COAST-V): 16 week results of a phase 3 ra. Lancet. 2018;392(10163):2441–51.
- Vegas LP, Penso L, Claudepierre P, Sbidian E. Long-term Persistence of First-line Biologics for Patients With Psoriasis and Psoriatic Arthritis in the French Health Insurance Database. JAMA Dermatol. 2022;158(5):513–22.
- Coates LC, Soriano ER, Corp N, Bertheussen H, Duffin KC, Campanholo CB, et al. Group for Research and Assessment of Psoriasis and Psoriatic Arthritis (GRAPPA): updated treatment recommendations for psoriatic arthritis 2021. Nat Rev Rheumatol. 2022;18(8):465–79.
- 94. Gossec L, Baraliakos X, Kerschbaumer A, Wit M, McInnes I, Dougados M, et al. EULAR recommendations for the management of psoriatic arthritis with pharmacological therapies: 2019 update. Ann Rheum Dis. 2020;7(6):700–12.
- 95. Singh JA, Guyatt G, Ogdie A, Gladman DD, Deal C, Deodhar A, et al. Special Article: 2018 American College of Rheumatology/ National Psoriasis Foundation Guideline for the Treatment of Psoriatic Arthritis. Arthritis Rheumatol. 2019;71(1):5–32.
- Gionchetti P, Calabrese C, Rizzello F. Inflammatory Bowel Disease and Spondyloarthropathies. J Rheumatol. 2015;93(Suppl):21–3.
- National Institute of Clinical Excellence (NICE). Crohn's disease: management [Internet]. London. 2019 [cited 2022 Aug 10]. Available from: https://www.nice.org.uk/guidance/ng129
- Fauny M, Moulin D, D'Amico F, Netter P, Petitpain N, Arnone D, et al. Paradoxical gastrointestinal effects of interleukin-17 blockers. Ann Rheum Dis. 2020;79(9):1132–8.
- 99. Hueber W, Sands BE, Lewitzky S, Vandemeulebroecke M, Reinisch W, Higgins PDR, et al. Secukinumab, a human anti-IL-17A monoclonal antibody, for moderate to severe Crohn's disease: unexpected results of a randomised, double-blind placebo-controlled trial. Gut. 2012;61(12):1693–700.
- 100. Targan SR, Feagan B, Vermeire S, Panaccoine R, Melmed G, Landers C, et al. A randomized, double-blind, placebo-controlled phase 2 study of brodalumab in patients with moderate-to-severe Crohn's disease. Am J Gastroenterol. 2016;111(11):1599–607.
- Chong WP, Mattapallil MJ, Raychaudhuri K, Bing SJ, Wu S, Zhong Y, et al. The cytokine IL-17A limits Th17 pathogenicity via a negative feedback loop driven by autocrine induction of IL-24. Immunity. 2020;53(2):384–97.
- Zhang Y, Liu Y, Xu Y. Interleukin-24 regulates T cell activity in patients with colorectal adenocarcinoma. Front Oncol. 2019;9:1401.

- 103. Persaud L, De Jesus D, Brannigan O, Richiez-Paredes M, Huaman J, Alvarado G, et al. Mechanism of action and applications of interleukin 24 in immunotherapy. Int J Mol Sci. 2016;17(6):869.
- Mehta P, Lawrence A, Aggarwal A. Paradoxical gastrointestinal effects of interleukin-17 blockers. Ann Rheum Dis. 2020;ePub ahead of print.
- 105. Mease P, van den Bosch F. IL-23 and axial disease: do they come together ? Rheumatol. 2021;60(Supp14):iv28-33.
- 106. Schinocca C, Rizzo C, Fasano S, Grasso G, Barbera LL, Ciccia F, et al. Role of the IL-23/IL-17 pathway in rheumatic diseases: an overview. Front Immunol. 2021;12:637829.
- 107. Hasegawa H, Mizoguchi I, Chiba Y, Ohashi M, Xu M, Yoshimoto T. Expanding diversity in molecular structures and functions of the IL-6/IL-12 heterodimeric cytokine family. Front Immunol. 2016;7:479.
- Sherlock JP, Joyce-Shailk B, Turner SP, Chao C, Sathe M, Grein J, et al. IL-23 induces spondyloarthropathy by acting on ROR-γt+ CD3+CD4-CD8- entheseal resident T cells. Nat Med. 2012;18(7):1069–76.
- Sakaguchi S. Naturally arising CD4+ regulatory t cells for immunologic self-tolerance and negative control of immune responses. Annu Rev Immunol. 2004;22:531–62.
- 110. Yoshitomi H, Sakaguchi N, Kobayashi K, Brown GD, Tagami T, Sakihama T, et al. A role for fungal {beta}-glucans and their receptor Dectin-1 in the induction of autoimmune arthritis in genetically susceptible mice. J Exp Med. 2005;201(6):949–60.
- 111. Ruutu M, Thomas G, Steck R, Degli-Esposti MA, Zinkernegel MS, Alexander K, et al. β-glucan triggers spondylarthritis and Crohn's disease-like ileitis in SKG mice. Arthritis Rheum. 2012;4(7):2211–22.
- 112. Dibra D, Xia X, Gagea M, Lozano G, Li S. A spontaneous model of spondyloarthropathies that develops bone loss and pathological bone formation: A process regulated by IL27RA-/- and mutant-p53. PLoS ONE. 2018;13(3):e0193485.
- Nakae S, Nambu A, Sudo K, Iwakuru Y. Suppression of immune induction of collagen-induced arthritis in IL-17-deficient mice. J Immunol. 2003;171(11):6137–77.
- 114. Ruyssen-Witrand A, Luxembourger C, Antagrel A, Nigon D, Claudepierre P, Degboe Y, et al. Association between IL23R and ERAP1 polymorphisms and sacroiliac or spinal MRI inflammation in spondyloarthritis: DESIR cohort data. Arthritis Res Ther. 2019;21(1):22.
- Li Z, Brown MA. Progress of genome-wide association studies of ankylosing spondylitis. Clin Transl Immunol. 2017;6(12):e163.
- 116. Mei Y, Pan F, Gao J, Ge R, Duan Z, Seng Z, et al. Increased serum IL-17 and IL-23 in the patient with ankylosing spondylitis. Clin Rheumatol. 2011;30(2):269–73.
- 117. Wendling D, Cedoz J, Racadot E, Dumoulin G. Serum IL-17, BMP-7, and bone turnover markers in patients with ankylosing spondylitis. Jt bone spine. 2007;74(3):304–5.
- 118. Chen W, Chang Y, Lin K, Lai C, Wang S, Hsiao K, et al. Association of serum interleukin-17 and interleukin-23 levels with disease activity in Chinese patients with ankylosing spondylitis. J Chinese Med Assoc. 2012;75(7):303–5.
- Gottlieb A, Menter A, Mendelsohn A, Shen Y, Li S, Guzzo C, et al. Ustekinumab, a human interleukin 12/23 monoclonal antibody, for psoriatic arthritis: randomised, double-blind, placebocontrolled, crossover trial. Lancet. 2009;373(9664):633–40.
- 120. McInnes IB, Kavanaugh A, Gottlieb AB, Puig L, Rahman P, Ritchlin C, et al. Efficacy and safety of ustekinumab in patients with active psoriatic arthritis: 1 year results of the phase 3, multicentre, double-blind, placebo-controlled PSUMMIT 1 trial. Lancet. 2013;382(9894):780–9.
- 121. Reich K, Warren RB, Iversen L, Puig L, Pau-Charles I, Igarashi A, et al. Long-term efficacy and safety of tildrakizumab

for moderate-to-severe psoriasis: pooled analyses of two randomized phase III clinical trials (reSURFACE 1 and reSURFACE 2) through 148 weeks. Br J Dermatol. 2020;182(3):605–17.

- 122. Mease PJ, Kellner H, Morita A, Kivitz AJ, Papp KA, Aslanyan S, et al. OP0307 Efficacy and safety of risankizumab, a selective il-23p19 inhibitor, in patients with active psoriatic arthritis over 24 weeks: results from a phase 2 trial. Ann Rheum Dis. 2018;77:200–1.
- 123. Gordon KB, Strober S, Lebwohl M, Augustin M, BLauvelt A, Poulin Y, et al. Efficacy and safety of risankizumab in moderate-to-severe plaque psoriasis (UltIMMa-1 and UltIMMa-2): results from two double-blind, randomised, placebo-controlled and ustekinumab-controlled phase 3 trials. Lancet. 2018;392(10148):650–61.
- 124. Deodhar A, Helliwell PS, Boehncke W, Kollmeier AP, Hsia EC, Subramanian RA, et al. Guselkumab in patients with active psoriatic arthritis who were biologic-naive or had previously received TNF α inhibitor treatment (DISCOVER-1): a double-blind, randomised, placebo-controlled phase 3 trial. Lancet. 2020;395(10230):1115–25.
- 125. Mease PJ, Rahman P, Gottlieb AB, Kollmeier AP, Hsia EC, Xu XL, et al. Guselkumab in biologic-naive patients with active psoriatic arthritis (DISCOVER-2): a doubleblind, randomised, placebo-controlled phase 3 trial. Lancet. 2020;395(10230):1126–36.
- 126. Coates LC, Gossec L, Theander E, Bergmans P, Neuhold M, Karyekar CS, et al. Efficacy and safety of guselkumab in patients with active psoriatic arthritis who are inadequate responders to tumour necrosis factor inhibitors: results through one year of a phase IIIb, randomised, controlled study (COS-MOS). Ann Rheum Dis. 2022;81(3):359–69.
- 127. Blauvelt A, Papp KA, Griffiths CEM, Randazzo B, Wasfi Y, Shen Y, et al. Efficacy and safety of guselkumab, an anti-interleukin-23 monoclonal antibody, compared with adalimumab for the continuous treatment of patients with moderate to severe psoriasis: Results from the phase III, double-blinded, placebo- and active comparator–. J Am Acad Dermatol. 2017;76(3):405–17.
- 128. Leonardi CL, Kimball AB, Papp KA, Yeilding N, Guzzo C, Wang Y, et al. Efficacy and safety of ustekinumab, a human interleukin-12/23 monoclonal antibody, in patients with psoriasis: 76-week results from a randomised, double-blind, placebocontrolled trial (PHOENIX 1). Lancet. 2008;371(9625):1665–74.
- 129. Papp KA, Langley RG, Lebwohl M, Krueger GG, Szapary P, Yeilding N, et al. Efficacy and safety of ustekinumab, a human interleukin-12/23 monoclonal antibody, in patients with psoriasis: 52-week results from a randomised, double-blind, placebocontrolled trial (PHOENIX 2). Lancet. 2008;371(9625):1675–84.
- McGonagle D, Watad A, Sharif K, Bridgewood C. Why inhibition of IL-23 lacked efficacy in ankylosing spondylitis. Front Immunol. 2021;12:614255.
- 131. Cuthbert RJ, Watad A, Fragkakis EM, Dunsmuir R, Loughenbury P, Khan A, et al. Evidence that tissue resident human enthesis γδT-cells can produce IL-17A independently of IL-23R transcript expression. Ann Rheum Dis. 2019;78(11):1559–65.
- 132. Bridgewood C, Watad A, Russell T, Palmer TM, Marzo-Ortega H, Khan A, et al. Identification of myeloid cells in the human enthesis as the main source of local IL-23 production. Ann Rheum Dis. 2019;78(7):929–33.
- 133. Bridgewood C, Sharif K, Sherlock J, Watad A, McGonagle D. Interleukin-23 pathway at the enthesis: The emerging story of enthesitis in spondyloarthropathy. Immunol Rev. 2020;294(1):27–47.
- 134. National Ankylosing Spondylitis Society (NASS). What are the issues in axial SpA (AS)? [Internet]. London, UK. 2021 [cited 2021 Jun 14]. Available from: https://nass.co.uk/about-as/as-facts-and-figures/

- 135. Hu X, Fu M, Zhao X, Wang W. The JAK/STAT signaling pathway: from bench to clinic. Signal Transduct Target Ther. 2021;6(402).
- 136. Traves PG, Murray B, Campigotto F, Galien R, Meng A, Di Paolo JA. JAK selectivity and the implications for clinical inhibition of pharmacodynamic cytokine signalling by filgotinib, upadacitinib, tofacitinib and baricitinib. Ann Rheum Dis. 2021;80(7):865–75.
- 137. McInnes IB, Szekanecz Z, McGonagle D, Maksymowych WP, Pfeil A, Lippe R, et al. A review of JAK-STAT signalling in the pathogenesis of spondyloarthritis and the role of JAK inhibition. Rheumatol. 2022;61(5):1783–94.
- 138. Doedhar A, Sliwinska-Stanczyk P, Xu H, Baraliakos X, Gensler LS, Fleishaker D, et al. Tofacitinib for the treatment of ankylosing spondylitis: a phase III, randomised, double-blind, placebocontrolled study. Ann Rheum Dis. 2021;80(8):1004–13.
- Deodhar A, van den Bosch F, Poddubnyy D, Maksymowych WP, van der Heijde D, Kim T, et al. Upadacitinib for the treatment of active non-radiographic axial spondyloarthritis (SELECT-AXIS 2): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet. 2022;400(10349):369–79.
- 140. van der Heijde D, Song I, Pangan AL, Deodhar A, Van den Bosch F, Maksymowych WP, et al. Efficacy and safety of upadacitinib in patients with active ankylosing spondylitis (SELECT-AXIS 1): a multicentre, randomised, double-blind, placebo-controlled, phase 2/3 trial. Lancet. 2019;394(10214):2108–17.
- 141. Food and Drug Administration (FDA). Xeljanz, Xeljanz XR (tofacitinib): Drug safety communication - initial safety trial results find increased risk of serious heart-related problems and cancer with arthritis and ulcerative colitis medicine [Internet]. Maryland. 2021 [cited 2022 Aug 11]. Available from: https:// www.fda.gov/safety/medical-product-safety-information/xeljanzxeljanz-xr-tofacitinib-drug-safety-communication-initial-safetytrial-results-find-increased#:~:text=In 2018%2C FDA approved the,%2C PsA%2C and ulcerative colitis.
- 142. Geriatric Medicine Journal. MHRA approves new treatment for ulcerative colitis [Internet]. Shoreham-by-Sea. 2022 [cited 2022 Aug 11]. Available from: https://www.gmjournal.co.uk/mhraapproves-new-treatment-for-ulcerative-colitis
- 143. Harrington R, Nokhata ASS, Conway R. JAK inhibitors in rheumatoid arthritis: an evidence-based review on the emerging clinical data. J Inflamm Res. 2020;13:519–31.

- 144. National Institute of Clinical Excellence (NICE). Upadacitinib for treating moderately to severely active ulcerative colitis [ID3953] [Internet]. London. 2022 [cited 2022 Aug 11]. Available from: https://www.nice.org.uk/guidance/indevelopment/ gid-ta10866
- National Institute of Clinical Excellence (NICE). Tofacitinib for moderately to severely active ulcerative colitis [Internet]. London. 2018 [cited 2022 Aug 11]. Available from: https://www. nice.org.uk/guidance/ta547
- 146. Medicines and Health Regulatory Authority (MHRA). MHRA Drug Safety Update [Internet]. London. 2021 [cited 2022 Aug 11]. Available from: https://assets.publishing.service.gov.uk/ government/uploads/system/uploads/attachment_data/file/10235 78/Oct-2021-DSU-PDF.pdf
- 147. Abbvie. RINVOQ® (upadacitinib) Receives FDA Approval for the Treatment of Adults with Moderately to Severely Active Ulcerative Colitis [Internet]. Dublin. 2022 [cited 2022 Aug 11]. Available from: https://news.abbvie.com/news/press-releases/ rinvoq-upadacitinib-receives-fda-approval-for-treatment-adultswith-moderately-to-severely-active-ulcerative-colitis.htm
- Sandboen WJ, Feagan BG, Loftus EV Jr, Peyrin-Biroulet L, Assche GV, D'Haens G, et al. Efficacy and safety of upadacitinib in a randomized trial of patients with Crohn's disease. Gastroenterology. 2020;158(8):2123–38.
- 149. Panés J, Sandboen WJ, Schreiber S, Sands BE, Vermeire S, D'Haens G, et al. Tofacitinib for induction and maintenance therapy of Crohn's disease: results of two phase IIb randomised placebo-controlled trials. Gut. 2017;66(7):1049–59.
- Ananthakrishnan AN. Upadacitinib for ulcerative colitis. Lancet. 2022;399(10341):2077–8.
- 151. Taylor PC, Saurigny D, Vencovsky J, Takeuchi T, Nakamura T, Matsievskaia G, et al. Efficacy and safety of namilumab, a human monoclonal antibody against granulocyte-macrophage colonystimulating factor (GM-CSF) ligand in patients with rheumatoid arthritis (RA) with either an inadequate response to background methotrexate therapy or an i. Arthritis Res Ther. 2019;21(1):101.

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