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1 **IMPACTS OF HEAT STRESS ON MEAT QUALITY AND**
2 **STRATEGIES FOR AMELIORATION; A REVIEW**

3

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20 **ABSTRACT**

21 During the summer, high ambient temperature and humidity cause economic loss
22 to the global livestock industry via reduced livestock productivity and increased
23 mortality. The problem of heat stress (HS) is likely to be exacerbated by global warming
24 and climate change. Recent research has shown that HS not only leads to physiological
25 and metabolic perturbations in live animals, but can also affect carcass and meat quality
26 characteristics plausibly by altering the rate and extent of postmortem muscle glycolysis
27 and resultant pH. However, these impacts of HS are not consistent across species.
28 Higher incidence of pale soft and exudative (PSE) meat has been reported in poultry.
29 On the contrary, higher incidence of high ultimate pH and dark firm and dry (DFD)
30 meat or no impacts of HS have been reported in sheep and cattle. With the limited data
31 on HS impacts on meat quality of ruminants, it is difficult to explain the exact
32 mechanisms driving these variable impacts. However, it is hypothesized that the
33 severity and duration of HS may lead to variable impacts due to lack of opportunity to
34 adapt to acute heat exposure. Longer HS exposure may allow ruminants to adapt
35 to heat and may not record any negative impacts on meat quality. This paper reviews
36 the recent research on impacts of HS on meat quality characteristics and identify the
37 key areas of further research required to better understand these negative impacts to
38 develop strategies for amelioration. In addition, some mitigation strategies of HS have
39 also been discussed which include both managerial and nutritional interventions.

40

41 **Key words:** heat stress, meat quality, ruminants, monogastrics, feed additives.

42

43

44 **ABBREVIATIONS**

45	HS	Heat stress
46	TN	Thermoneutral
47	PSE	Pale, soft, exudative
48	DFD	Dark firm and dry meat
49	THI	Temperature–humidity index
50	RH	Relative humidity
51	RT	Rectal temperature
52	RR	Respiration rate
53	WHC	Water holding capacity
54	WBSF	Warner–Bratzler shear force
55	pHu	Ultimate pH
56	IMF	Intramuscular fat
57	LTL	Longissimus thoracis et lumborum
58	LL	Longissimus lumborum

59 **1. INTRODUCTION**

60 High ambient temperature compromises animal welfare and productivity due to
61 heat stress (HS) experienced over summer months. Heat stress has become one of the
62 major challenges facing global livestock production, especially in the warmer parts of
63 the world (Bernabucci et al. 2010; Rhoads et al. 2013). Recent deaths of 2400 Australian
64 sheep due to HS on a ship to the Middle East
65 ([https://www.theguardian.com/world/2018/apr/05/disgusting-death-of-2900-](https://www.theguardian.com/world/2018/apr/05/disgusting-death-of-2900-australian-sheep-on-ship-to-middle-east-sparks-investigation)
66 [australian-sheep-on-ship-to-middle-east-sparks-investigation](https://www.theguardian.com/world/2018/apr/05/disgusting-death-of-2900-australian-sheep-on-ship-to-middle-east-sparks-investigation)) has highlighted animal
67 welfare issues and severe consequences of HS in sheep. It has also increased the

68 vulnerability of Australian live livestock export business, which could lead to millions
69 of dollar economic losses. According to meteorological data, Australia's yearly
70 temperature was on average higher 1 to 1.5 °C from 1910 to 2018, (State of the Climate
71 2018, Australian Government Bureau of Meteorology). In Australia, during the years
72 2013-2017, there was a 27 per cent increase in the number of hot days above 35°C
73 (Osei-Amponsah et al. 2019), and year 2019 was warmest year on record, with the
74 annual national mean temperature 1.52 °C above average (BOM, 2019). The CMIP5
75 (Coupled Model Intercomparison Project phase 5) climate modelling projections
76 indicated that the heat waves in Australia will become longer, more frequent and hotter
77 by the end of 21st century (Cowan et al. 2014). It is predicted that the impacts of HS
78 will be further exacerbated by climate change. As reviewed by others (Gregory 2010;
79 Johnson 2018; Gonzalez et al. 2020), extremes in summertime temperatures impose
80 negative effects on animal growth performance, alter carcass composition and increase
81 the risk of pale, soft, exudative (PSE) or PSE-like meat in pigs, poultry and beef cattle,
82 and dark firm and dry (DFD) meat in ruminants.

83 Therefore, a better understanding of HS impacts on animal welfare, production and
84 product quality is required to develop suitable strategies to mitigate HS to promote
85 animal welfare and sustainable production. This paper reviews the current knowledge
86 and research progress of HS impacts on meat quality and identifies some of the potential
87 nutritional interventions to alleviate the negative consequences of HS to improve meat
88 quality.

89 **2. HEAT STRESS**

90 "Stress" is defined as the body's response to external forces, which tends to
91 displace its system from the resting or basal state (Hancock 2006). Heat stress occurs

92 when the ambient temperature is higher than the animal's thermoneutral zone (TNZ,
93 the range of environmental temperature within which the animals uses no additional
94 energy to maintain their body temperature). The TNZ is influenced by many
95 environmental factors which would affect heat gain and loss from an animal's body
96 such as wind velocity, air temperature, humidity, precipitation, solar radiation, animal
97 breed and availability of shade or shelter (Bernabucci et al. 2010; Aggarwal and
98 Upadhyay 2013).

99 **2.1 ASSESSMENT OF HEAT STRESS SEVERITY**

100 The temperature–humidity index (THI) is used to quantify HS and is calculated
101 based on the environmental temperature combined with relative humidity (high
102 humidity could reduce an animal's evaporative cooling efficiency, thus affecting animal
103 TNZ). The THI offers an objective method to assess the animal's response to HS and
104 the influence of different climate elements.

105 A number of equations have been developed to calculate THI, using dry bulb
106 temperature (db°C) and relative humidity (RH, %). For example in sheep, $THI = db^{\circ}C -$
107 $\{(0.31 - 0.31 \times RH) \times (db^{\circ}C - 14.4)\}$ (Marai et al. 2007). Using this equation, a THI lower
108 than 22.2 is classified as thermoneutral, a THI from 22.2 to 23.3 is moderate HS, and a
109 THI of 23.3 to 25.6 it is referred to severe HS , and extreme severe HS occurs when
110 THI exceeds 25.6 (St Pierre 2003; Marai et al. 2007). Recently, another index, the heat
111 load index (HLI), has been developed. This HLI is more sensitive especially for animals
112 living outdoors as it incorporates the effects of wind movement and solar radiation. The
113 HLI is calculated based on THI and combine with heat radiation heat and wind velocity,
114 such as: $HLI = 8.62 + (0.38 \times RH) + (1.55 \times BG) - (0.5 \times WS) + e^{(2.4 - WS)}$ when black
115 globe temperature above 25 °C (BG= black globe temperature, ° C; WS = wind speed,

116 m/s; e = base of the natural logarithm) (Gaughan et al., 2008). As a new index, HLI has
117 been used for sheep (Mengistu et al., 2017), beef (Gaughan et al., 2010) and dairy cattle
118 (Lee et al., 2018) research.

119 Whenever livestock are exposed to environmental temperatures outside their TNZ,
120 thermoregulatory mechanisms, such as increased respiration rate (RR), are activated to
121 achieve homeothermy. However, if the ambient temperature and humidity continue to
122 increase, body temperature regulation becomes difficult, and core body temperature
123 starts to increase, eventually resulting in HS (Chauhan et al. 2014). Under HS
124 conditions, the animal is forced to make drastic changes in its physiological and
125 behavioral responses such as increased RR and reduced feed intake, to cope with the
126 building heat load and increased core temperature (Aggarwal and Upadhyay 2013).
127 Therefore, core body temperature and RR are often measured along with THI to assess
128 an animal's HS responses. Rectal temperature (RT) and RR are the most sensitive
129 physiological responses that are elevated in response to HS. Even a 1°C increase in RT
130 is enough to impact most animal's growth performance and health parameters
131 (McDowell et al. 1996; Berman 2011; Chauhan et al. 2015). For example, acute HS for
132 3 hrs can increase broiler chickens' core temperature from 42.0°C, to 42.9°C (Lin et
133 al. 2006). In sheep, chronic HS (28 to 40°C, 30 to 40% RH; 1 week) increased lamb RR,
134 RT, skin temperature and decreased feed intake as compared to lambs in the
135 thermoneutral group (18 to 21°C, 40 to 50% RH) (Chauhan et al. 2015, 2016).

136 **2.2 HEAT STRESS AND MEAT QUALITY**

137 In the review by Gregory (2010), he postulated that HS will potentially affect
138 broiler meat quality via the influence of HS on postmortem muscle pH decline rate and
139 ultimate pH (pHu). Post-mortem muscle pH decline is affected by glycogen levels and

140 the rate of glycolysis, which in turn can be affected by preslaughter stress and muscle
141 temperature, respectively (Chauhan and England 2018). However, the impacts of HS
142 on meat quality among different species or breeds of animal are not consistent, possibly
143 because of the variation in thermotolerance. In general, the acceptable temperature
144 range for an animal depends on genetics and environment such as feeding level, housing,
145 air speed and stocking rate (Randall 1993). Recent studies have reported variable meat
146 quality performance during summer in different species and breeds of animals. Meat
147 from ruminants, such as mutton and beef, had lower lightness and higher ultimate
148 muscle pH values, similar to dark firm and dry meat (DFD) during summer (Kim et al.
149 2003; Mach et al. 2008; Weglarz 2010), while poultry and pig meat had greater
150 lightness, and lower pHu similar to PSE meat during summer (Langer et al. 2010;
151 McCurdy et al. 1996; Gregory 2010).

152 Most controlled HS experiments have focused on poultry (broiler or turkey)
153 meat quality, and the results of HS impacts on meat quality parameters are variable
154 even within these species. As summarized in Figure 1, out of the 23 publications
155 reviewed, 9 publications reported a negative effect of HS on the meat quality and 3
156 studies reported no significant effect (Figure 1). Interestingly, recent research (section
157 3.2) has reported some new phenomenon and mechanisms for meat quality changes
158 under HS conditions which may be responsible for the variable effects on carcass
159 quality characteristics and meat quality attributes.

160 **2.2.1 HEAT STRESS AND CARCASS CHARACTERISTICS**

161 As a result of adaptive responses to chronic HS, animals generally reduce feed
162 intake in an attempt to decrease metabolic heat production (Marai et al. 2007), which
163 has implications for carcass fat deposition, carcass yield and intramuscular fat content

164 (IMF). These impacts on carcass characteristics have been reported both in
165 monogastrics and ruminants, and HS has been well known to reduce feed intake and
166 carcass yield in poultry (Lu et al. 2007; Quinteiro-Filho et al. 2010; Zhang et al. 2012;
167 Imik et al. 2012), pigs (Colditz and Kellaway 1972; Spencer et al. 2003; Pearce et al.
168 2013), goats (Sivakumar et al. 2010; Lu 1989); cattle (O'Brien et al. 2010; Maloiy et al.
169 2008) and sheep (Maloiy et al. 2008). The magnitude of changes in carcass
170 characteristics may vary between different species, while overall, HS-induced carcass
171 yield loss has been estimated to cause extensive economic losses to livestock industries
172 (St Pierre 2003). Long term HS also results in decreased subcutaneous fat and lower
173 IMF levels, which may assist the animal with better heat dissipation rates (Ma et al.
174 2015). For example Lu et al. (2007) demonstrated that Arbor Acres (AA) broilers'
175 breast IMF decreased when birds were exposed to 34°C HS conditions for 3 weeks.
176 Combining with the reduction of fat content, lower acetyl coenzyme A carboxylase
177 enzyme and L (+) P-hydroxyacyl CoA dehydrogenase activity were also reported in a
178 pig HS experiment (Ma et al. 2015). Pearce et al. (2011) reported lower lipolytic
179 enzymes activity in a pig HS experiment, which was independent of the HS-induced
180 reduction in feed intake. Conversely, no significant impact of HS on broiler leg
181 intramuscular fat was reported when birds were exposed to 32°C or 22°C for 3 weeks
182 with ad libitum feeding (Baziz et al. 1996). Similarly, in goats there was no significant
183 effect of one month of out-shed feeding (exposed to THI 73.5-86.5) on IMF contents
184 compared with in-shed feeding (exposed to THI 69.9-74.9) (Archana et al., 2018). The
185 absence of any effects on goats could be because of the better ability of both goat breeds
186 to cope with HS as both breeds were indigenous Indian breeds that have better
187 thermotolerance. Mader et al. (1999) and Ponnampalam et al. (2016) also did not
188 observe any difference in subcutaneous fat thickness over the longissimus lumborum

189 in cattle, and carcass fat score for lambs, respectively, between HS (28 to 40°C, 30 to
190 40 RH, 1 week) and TN groups. Interestingly, the carcass weight is significantly
191 impacted by HS due to the reduction in feed intake, but the impacts on carcass
192 composition are equivocal and need further research under controlled conditions.
193 However, some the variation in carcass characteristic under HS conditions may be
194 attributed to the inherent differences in heat dissipation capacity of different
195 species/breeds and the severity and length of the HS conditions imposed. Lu et al. (2007)
196 demonstrated that IMF of a better heat adapted chicken breed (Beijing You) was less
197 impacted by HS compared with a high production breed (Arbor Acres). Similarly, in
198 ruminants, Archana et al. (2018) suggested that higher adaptability of indigenous goat
199 breeds to HS could explain why HS had no impact on majority of carcass characteristics.

200 **2.2.2 HEAT STRESS AND POSTMORTEM MUSCLE METABOLISM**

201 After an animal is harvested for meat, two major changes occur to skeletal muscle;
202 physical structural changes, and biochemical changes due to cessation of blood flow
203 and oxygen supply, and lack of glucose supply. However, despite these constraints,
204 skeletal muscle endeavors to achieve premortem homeostatic balance and continues to
205 synthesize and utilize ATP through the metabolism of stored glycogen (England et al.
206 2013), which is the main source of energy used to generate glycolytic substrates in
207 postmortem muscle. This futile effort of muscle to maintain homeostatic control
208 culminates in accumulation of lactate and hydrogen ions, as pyruvate can no longer
209 enter the mitochondria, as the electron chain has stopped working due to a lack of
210 oxygen (England et al. 2013; Cheah and Cheah 1971). Consequently, there is a decline
211 in muscle pH as lactate and hydrogen ions accumulate (Tarrant 1981). The regulation
212 of these biochemical processes is critical in determining the quality of meat. Any

213 deviation from the normal rate and extent of postmortem metabolism or glycolysis may
214 lead to poor meat quality development, such as faster pH decline and lower pHu which
215 results in PSE meat (Briskey et al. 1964); and meat with a high pHu called DFD, and
216 dark-cutting in ruminants (Tarrant 1989), both of which are inferior in quality and more
217 prone to bacterial spoilage.

218 In terms of metabolic perturbations, HS leads to elevated plasma glucocorticoid
219 (cortisol) concentrations due to an activation of the hypothalamic–pituitary–
220 adrenal axis. The secretion of cortisol stimulates physiological adjustments that enable
221 an animal to tolerate the stress caused by the hot environment (Christison and Johnson
222 1972). Generally, animal blood cortisol concentrations will rise as the ambient
223 temperature and exposure time increases (Kadim et al. 2014; Quinteiro-Filho et al.
224 2010). However, these metabolic changes reported in livestock have not been
225 consistently observed in poultry (Lin et al. 2006). Based on the variation in the
226 metabolic rate antemortem, postmortem muscle physiological and biochemical
227 parameters may vary in heat stressed animals as compared to animals reared under
228 thermoneutral conditions. Heat stress has been implicated in higher muscle glycolysis
229 rate postmortem in broilers transported during summer (Xing et al. 2016), and in
230 broilers exposed to chronic heat stress conditions (Aksit et al. 2006). For example, 0.5
231 hr summer transportation (32-42°C) increased the AMP/ATP ratio of AA broiler breast,
232 leading to increased AMPK (AMP-activated protein kinase) activity and ultimately
233 resulting in a lower pHu value (Xing et al. 2015). Similarly, Liang et al. (2018) reported
234 that broilers under short term high ambient temperature condition (36°C; 1 hr) had
235 higher AMPK activity than thermoneutral group (25 ± 1°C) at 1 hr. postmortem. There
236 are number of HS studies in poultry and pigs that have reported lower pHu or rapid pH
237 decline in postmortem muscle (Zhang et al. 2012; Tang et al. 2013; Wang et al. 2016;

238 Downing et al. 2017) However, it is intriguing that other studies did not observe any
239 significant effect of HS on muscle pHu (Lu et al. 2007; Owens et al. 2000; Costa et al.
240 2006; Shi et al. 2017).

241 Contrary to responses in pig and poultry, the meat of ruminants generally has
242 higher pHu values under both chronic and acute HS conditions. For example, chronic
243 HS caused by seasonal high temperatures was reported to increase the pHu of *psoas*
244 *major* and *psoas minor* muscles in goats (Omani and Somali) and sheep (Somali and
245 Merino) (Kadim et al. 2008). As discussed previously, these effects of HS could be
246 explained by glycogen depletion before slaughter and higher glycolytic enzyme activity
247 postmortem, since normal pHu (<5.8) muscle has lower glycogen phosphorylase
248 enzyme activity and glycolytic potential at 0.5 hr postmortem compared to muscle with
249 a high pHu (≥ 5.8) (Apaoblaza et al. 2015). Antemortem glycogen content and
250 glycolytic enzyme activity postmortem would explain the difference between pig,
251 poultry and ruminant meat under HS conditions. For example, ruminant longissimus
252 muscle glycogen concentration of 44.5mmol/kg is required to achieve normal pH
253 decline to <5.8 (Apaoblaza et al. 2015), however, HS would result in the depletion of
254 muscle glycogen (Jentjens et al., 2002) leading to high ultimate pH. On the other hand,
255 in chickens, Wang et al. (2016) reported that short term HS (40°C, 1h) had no impact
256 on chicken breast initial glycogen content and HS increased the glycolytic enzyme
257 activity leading to faster rate and greater extent of pH decline.

258 It is worth noticing that longer heat exposure or higher ambient temperature may
259 not necessarily cause more negative impact on muscle pH and meat quality (section
260 2.2.3). For example, Xing et al. (2015) observed that the negative impact on broilers'
261 breast gradually weakened as summer transportation time extend from 1 to 4 hrs (Figure
262 2). Similarly, Tang et al. (2013) demonstrated that 10 hrs heat exposure (40°C) had no

263 impact on broilers' breast pHu, which was contrary to shorter heat exposure time (1 to
264 5 hrs ; Figure 3). It appears that exposure to extreme HS conditions for a short duration
265 leads to acute stress response accelerating the rate of postmortem muscle glycolysis and
266 greater decline in muscle pHu. However, prolonged exposure to such conditions may
267 lead to adaptation of animals to HS and therefore no adverse effects are observed on
268 postmortem muscle pH decline. Nevertheless, long-term exposure to HS is well known
269 to have implications for animal welfare and may influence consumers decision to buy
270 meat, though it may not necessarily have impacts on meat quality.

271 **2.2.3 HEAT STRESS AND MEAT COLOUR, WATER HOLDING CAPACITY AND** 272 **WBSF**

273 Heat stress may result in higher pHu, which leads to less shrinkage of the
274 myofilament lattice and results in a darker meat colour (higher light absorption, less
275 light scattering, higher oxygen consumption) (Gregory 2010; Hughes et al. 2019).
276 Though there are limited studies with controlled temperature conditions focusing on the
277 effect of HS on ruminant meat quality, some studies have demonstrated that HS has
278 negative effect on meat colour, water holding capacity and results in higher pHu value.
279 For example, studies by Kadim et al. (2004, 2007, 2008, 2014) reported that acute and
280 chronic HS decreased goat and sheep meat lightness, with lower WHC and increased
281 toughness. Similarly, Liu et al. (2015) also reported that grazing sheep without access
282 to shade produced meat with a higher pHu and poor overall quality (as summarized in
283 table 1). Although the negative effects of HS on ruminant meat can be similar to those
284 responses observed in DFD meat, which has higher WHC compared to normal meat,
285 the responses noted in heat stressed (seasonal comparisons and summer transportation)
286 ruminant meat had lower WHC (Kadim et al. 2008, 2014), which is usually associated
287 with PSE-like meat. On the other hand, HS increases the rate of pH decline in

288 postmortem muscle in broilers and pigs, which results in higher lightness and lower
289 WHC. For example, acute HS (32°C, 2h) significantly accelerated the pH_{15min} decline
290 in broiler breast muscle and degraded meat quality attributes such as higher drip loss
291 (72h postmortem), higher breast meat colour and lower sensory score (Sandercock et
292 al. 2001). Similar trends were also observed in chronically heat stressed broiler and
293 turkey meat that was paler in colour and had a lower WHC under seasonal HS
294 conditions (Lu et al. 2007). In pigs, chronic HS (35°C, 78% RH; 30 days) increased
295 LTL WBSF and decreased intramuscular fat content (Shi et al. 2017; Table 2).

296 Interestingly, although the majority of publications have reported negative impacts
297 of HS on meat quality, recent studies suggest that the negative impacts of HS (both
298 under acute and chronic conditions) on meat quality are not consistent and are even
299 contradictory. Some of these variable effects can be attributed to differences in ambient
300 temperature, relative humidity, and exposure time to high environmental temperature
301 between individual studies. Ponnampalam et al. (2016) reported that 7 days of cyclic
302 HS (28-40 °C, 30-40% RH) did not negatively influence lamb muscle pH_u and TBARS
303 levels. Northcutt et al. (1994) observed that broilers reared at an ambient temperature
304 of 40°C had better WHC of *pectoralis major muscle* compared to control (25 °C).
305 Moreover, it appears that meat quality is also affected by the duration of exposure to
306 heat stress conditions which may result in variable meat quality parameters. For
307 example, Tang et al. (2013) measured the meat quality of the *pectoralis major* of
308 broilers under different heat (37±1 °C) exposure time before slaughter and found that
309 the muscle lightness, cooking loss, and WBSF values were maximum after 3-5 hrs of
310 heat exposure. After 10 hrs. of heat exposure, muscle quality parameters showed a trend
311 towards recovery and better meat quality was observed (Figure 3). Similar trends were
312 also reported by Xing et al (2015), where 0.5 hrs of transportation during summer

313 resulted in lower chicken meat quality parameters, and the birds transported for 4 hrs
314 showed similar breast lightness, WHC and pHu relative to the non-transportation group.
315 Zhang et al. (2019) also demonstrated that broilers' breast meat quality was affected
316 mostly at 36 °C heat exposure. As the ambient temperature increased from 38 °C to
317 40 °C, certain meat quality parameters showed a reverse trend and improved in quality,
318 and the plasma cortisol level had no increase between 38 °C and 40 °C (Figure 4).
319 Therefore, acute HS is more likely to affect broilers' meat quality as extreme high
320 temperatures or acute heat exposure does not allow the adaptive responses to HS to
321 occur, whereas chronic HS may provide opportunities for animals to adapt thus having
322 less impact on meat quality (Sandercock et al. 2001). However, this does not undermine
323 the implications of heat stress on animal welfare irrespective of impacts on meat quality.
324

325 **2.2.4 HEAT STRESS AND MEAT SAFETY**

326 There are a limited number of studies that have investigated the effect of HS on
327 meat safety, but it is generally accepted that high ambient temperature and humidity
328 present favorable conditions for pathogen colonization which may lead to safety risks
329 for meat and by-products. Chronic stress may influence the course of animal infection
330 and/or the susceptibility to a microorganism (Elenkov and Chrousos 1999); alter the
331 hosts intestinal barrier susceptibility to pathogenic bacteria; and increase intestinal
332 permeability and luminal attachment for bacteria (Bailey et al. 2004). Higher
333 catecholamine and glucocorticoid (stress hormone) secretion may affect the animals
334 intestinal barrier function and microbial environment (Verbrugghe et al. 2012).
335 Secretion of norepinephrine also accelerates intestinal motility, colonic transit and
336 transepithelial ion transport, which may also influence gut microbial populations

337 (Mizuta et al. 2006). Apart from the direct effects of stress, higher pHu caused by HS
338 offers a conducive environment for microbial growth. Faucitano et al. (2010) found that
339 DFD meat had highest total aerobic mesophilic and presumptive lactic acid bacteria
340 counts when stored for 35 days at 4 °C following vacuum packaging. Similar results
341 have also been reported in broiler meat (Allen et al. 1997).

342 **3. MANAGEMENT STRATEGIES FOR ALLEVIATION OF** 343 **HEAT STRESS ANTEMORTEM**

344 In general, HS is the result of an imbalance between heat production and heat loss
345 mechanisms. Therefore, in addition to physical modifications of the animal's
346 environment to minimize the exposure to heat, various mitigation strategies need to be
347 targeted to either reduce heat production in the body or to enhance heat loss from the
348 body, to achieve homeothermy. The strategies to reduce HS have been broadly
349 classified as environmental modifications, genetic selection for improving
350 thermotolerance, and nutritional strategies to improve feed intake and decrease
351 metabolic heat production (Beede and Collier 1986; Chauhan et al. 2015).

352 **3.1 SHADE AND SHELTER MANAGEMENT**

353 Provision of shade is likely the most direct way to protect animals from direct and
354 indirect exposure to solar radiation while not necessarily reducing the air temperature.
355 For example, man-made shelters can significantly reduce the risk of HS in cattle
356 compared with lack of shade availability (Van laer et al. 2014; Eigenberg et al. 2005).
357 The provision of shade not only ameliorates the heat load of cattle (Gaughan et al. 2010;
358 Mader et al. 1999) but also reduces the mortality in extreme weather conditions (Darrell
359 Busby 1997). A limited number of studies have investigated the effects of shade on

360 meat quality, and it has been observed that provision of shade produced sheep meat
361 with less high pHu and better WHC (Liu et al. 2012). However, there were no
362 differences in growth performance, carcass fat percentage (Mader et al., 1999), or meat
363 quality of shade feedlot cattle as compared with conditions of no shade (DiGiacomo et
364 al., 2014). While shade can be effective in amelioration of some of the negative impacts
365 of HS under hot and dry conditions, but it may not be as effective under hot and humid
366 conditions (Renaudeau et al. 2012; Gaughan et al. 2010). In general, provision of shade
367 or shelter is recommended in the areas where ambient temperature exceeds 24°C, to
368 promote the welfare of farm animals and prevent production losses due to HS
369 (Silanikove 2000). While provision of shade can be an effective method to reduce heat
370 load in dairy cows even in temperate climate zones such as under New Zealand summer
371 conditions (Fisher et al. 2008; Kendall et al. 2006; Schütz et al. 2008), the beneficial
372 effects of shade for meat quality during summer in temperate zones requires further
373 research.

374 **3.2 EVAPORATIVE COOLING AND CONVECTION**

375 Evaporative and convective cooling are effective means of heat loss from the
376 animals body and can be utilized to cool animals both on farm and during lairage at the
377 processing plant. It involves the provision of air movement and water-cooling systems.
378 Recent research in broilers and turkeys has shown that better growth performance can
379 be obtained with provision of optimal ventilation in general (Yahav et al. 2005;
380 Renaudeau et al. 2012). Combining a well-oriented, semi-open building, with a high
381 and well isolated roof will provide natural ventilation to the animals. Among other
382 strategies, fan systems with shower like sprinkler-misters can be used to reduce the
383 ambient temperature in low RH areas, thus reducing the heat load on animals. The

384 sprinkler-fan system could improve feed intake of dairy cows by 7% to 10% as a result
385 of lower body temperature and respiration rates (Aggarwal and Upadhyay 2013).
386 During summer, water cooling could be an effective strategy to alleviate the negative
387 effects of HS on meat quality. Long and Tarrant (1990) found showering of pigs with
388 water brought about a 2°C temperature drop in pig LTL muscle after slaughter in
389 summer, which was sufficient to reduce paleness and drip loss in loin chops. Xing et al.
390 (2016) pointed out that 10 min of water spray after transportation on a hot day produced
391 a significant reduction in broiler HS and improved chicken meat quality. However,
392 these responses to water cooling are less pronounced or absent in humid regions
393 (Renaudeau et al. 2012).

394 **3.3 NUTRITIONAL STRATEGIES**

395 One of the largest negative effects of HS on animal production is the reduction in
396 feed intake, therefore dietary interventions targeting improved energy metabolism may
397 alleviate some of the negative effects of HS on animal growth and meat quality
398 characteristics. Similarly, HS is known to cause oxidative stress in poultry (Mujahid et
399 al. 2009), pigs (Liu et al. 2018), sheep (Chauhan et al. 2015), and cattle (Bernabucci et
400 al. 2002; Bernabucci et al. 2010; Garner et al. 2017). Therefore, supplementation with
401 dietary antioxidants provides an opportunity to alleviate some of the negative impacts
402 of HS. Antioxidants play an important role in preventing oxidative damage at the
403 cellular level. Cellular defense against oxidative damage exists at four different levels:
404 scavenging free radicals, chelation of metal ions, regeneration of antioxidants, and
405 repair of oxidized molecules (Chauhan et al., 2014). Antioxidants, such as phenolic
406 compounds and alkaloids from plant extracts, react with radicals to form more stable
407 compounds via hydrogen proton or directly donate electrons to react with radicals,

408 which could inhibit or delay oxidative chain reactions (Prakash et al. 2013). Some
409 antioxidants not only play a role in scavenging free radicals directly, but also play an
410 important role in enzyme antioxidant systems. For example, quercetin inhibits inducible
411 nitric oxide synthase (iNOS) activity and improves antioxidant enzyme activity which
412 may improve raw meat colour stability and flavor via oxidation inhibition (Goliomytis
413 et al. 2014).

414 Additionally, dietary manipulation to alter systemic insulin sensitivity may be an
415 effective strategy to reduce the HS impacts, as appropriate insulin action is one of the
416 key components of successfully adapting to and surviving a heat load (Rhoads et al.
417 2013). Recent research has demonstrated that the negative impacts of HS are not only
418 associated with reduction of feed intake, but that there are direct impacts of HS on
419 energy metabolism, with a shift towards increased carbohydrate use and reduced lipid
420 oxidation (see review by Rhoads et al. (2013)). Therefore, diets or nutritional
421 supplements promoting glucose use may be beneficial to reduce the negative impacts
422 of HS. Other additives to the diet could either improve body water balance or improve
423 body condition preslaughter, each of which may improve meat quality attributes. Some
424 of the potential nutritional supplements known to improve meat quality are briefly
425 reviewed in this part.

426 **3.3.1 ELECTROLYTES**

427 As discussed previously, when body temperature increases during HS conditions,
428 evaporative heat loss plays the main role of heat dissipation, which often results in
429 respiratory alkalosis due to hyperventilation, and dehydration due to excessive
430 salivation (West 2003). This ultimately leads to increased blood pH as carbon dioxide
431 partial pressure and total concentration decreases. This adverse respiratory alkalosis

432 and dehydration situation could be relieved by electrolyte supplementation (Teeter et
433 al. 1985; Sawka and Montain 2000). Aqueous electrolyte solution has been shown to
434 improve broiler growth performance under stressful conditions and also subsequent
435 meat quality (Whiting et al. 1991; Deyhim and Teeter 1991; Belay and Teeter 1993;
436 Ait-Boulahsen et al. 1995). Kumar et al. (2010) showed that the supplementation of
437 ascorbate in the diet in addition to electrolytes alleviated oxidative stress in buffaloes
438 and helped to boost the cell mediated immunity under HS conditions. In cattle,
439 electrolyte supplementation alleviated carcass yield loss during transportation stress
440 and reduced DFD meat frequency (Schaefer et al. 1990, 1997). However, it should be
441 noted that antemortem electrolyte therapy in cattle had no significant effect on the beef
442 flavor, juiciness, WBSF and overall palatability rating of beef (Jeremiah et al. 1992).

443 **3.3.2 VITAMIN E**

444 Vitamin E (Vit E) is the most important lipid soluble chain breaking antioxidant
445 present in cell membranes and prevents the oxidative damage to lipids and proteins by
446 directly scavenging free radicals and reducing lipid peroxide formation. Pervious
447 research has demonstrated that Vit E supplementation leads to a significant reduction
448 in meat product oxidation and improvement in colour stability (Mitsumoto et al. 1991;
449 Guerra-Rivas et al. 2016). Several studies have shown that Vit E supplementation with
450 or without selenium (Se), may improve physiological responses and animal
451 performance under HS conditions. For example, Sivakumar et al. (2010) found that Vit
452 E and Vitamin C supplementation improved goat RR, feed intake and cortisol levels
453 under HS conditions (40C, 30% RH; 21days). In a series of experiments in sheep and
454 pigs, our lab has demonstrated that the combination of supranutritional Vit E and Se
455 supplementation can be used to reduce the negative impacts of HS (Chauhan et al. 2015;

456 Liu et al. 2016). High levels of Vit E and Se supplementation significantly alleviated
457 HS effects in sheep via improving feed intake and antioxidant enzyme activity
458 (Chauhan et al. 2015). Similarly, in another study in pigs, Liu et al. (2016) demonstrated
459 that high levels of dietary Se and Vit E can reduce both oxidative stress and intestinal
460 leakiness. However, the effect of Vit E and Se supplementation on meat quality
461 performance under HS conditions requires further research as the limited research so
462 far has indicated equivocal results. Ponnampalam et al. (2016) did not observe any
463 significant effect of Vit E and Se supplementation on the meat quality of lambs finished
464 under hot summer conditions; but recent research by the same group reported that
465 supranutritional Vit E supplementation during finishing under HS conditions improved
466 aged lamb meat colour stability and reduced the brownness formation (Baldi et al.
467 2019). Furthermore, lambs supplemented with supranutritional Vit E during the
468 finishing phase had higher muscle Vit E content as compared to lambs finished on
469 Lucerne diet, however, there was no difference in lipid oxidation between the two
470 groups.

471 **3.3.3 FLAVONOIDS**

472 Flavonoids exist widely in plant tissues and have the most variety compared to
473 other polyphenol compounds. According to their basic structure, flavonoids generally
474 can be classified into flavones, flavanones, anthocyanidins and flavanols (Lotito and
475 Frei 2006). Among the various flavonoids, catechins have been the most commonly
476 used in recent research on animal diets (Perumalla and Hettiarachchy 2011). In addition
477 to their chain breaking antioxidant function, catechins are also involved in the
478 regeneration of endogenous antioxidants like Vitamin E. For example, dietary
479 supplementation of 2mg/kg green tea (catechins) for 4 weeks in rats, increased the

480 plasma and tissue alpha-tocopherol concentration (Frank et al. 2006). In vitro, catechins
481 spared alpha-tocopherol both by inhibiting its free radical mediated oxidation and by
482 regenerating it from its a-tocopheroxyl radical (Frank et al. 2006). In human research,
483 Vitamin E-sparing and regenerating effects were observed for green tea extracts (Zhu
484 et al. 1999). In animal diets, supplementation with catechins has been shown to enhance
485 meat antioxidant capacity and improve meat quality. For example, catechins
486 supplementation (3000–4000 mg tea catechin (tea catechin/kg fed for 40-60 days)
487 improved goat meat (fresh) colour, oxidation stability and WHC (Zhong et al. 2009).
488 Tang et al. (2001) further found that 200 and 300 mg/kg tea catechin supplementation
489 was effective in significantly delaying lipid oxidation as compared to the control in
490 chicken heart, liver, thigh meat and breast, and 300mg/kg TC had antioxidant capacity
491 equal to 200 mg/kg of Vitamin E diets. However, 10-100 mg/kg feeding of green tea
492 polyphenols had no significant effects on pork quality, plasma antioxidant capacity and
493 tissue Vitamin E level (Augustin et al. 2008).

494 In HS studies, dietary genistein (one of the isoflavones) supplementation (400 and
495 800 mg/kg) was reported to reduce the negative effects of cyclic heat (34°C) stress on
496 Japanese quail carcass yield and serum TBARS level (Onderci et al. 2004). Another
497 study by Kamboh et al. (2013; 2014) reported that 5 mg/kg genistein feeding, and 10
498 or 20 mg/kg genistein-hesperidin feeding in broiler groups lowered breast HSP70
499 mRNA expression under summer stress and had higher WHC (%), but 20 mg/kg
500 genistein-hesperidin diets increased broiler breast meat lightness values. Further
501 research is required to demonstrate the potential benefits of flavonoids supplementation
502 in ruminant diets to improve meat quality during summer.

503 **3.3.4 BETAINE**

504 Betaine is a quaternary ammonium compound and present in vertebrates' viscera
505 and amniotic fluid. It can be extracted from sugar beets or synthesized by chemical
506 reactions. Betaine was adapted as a replacement for methionine and choline to play a
507 role of methyl donor and osmoprotectant in poultry and fish diets as early as the 1970s
508 (Fernandez-Figares et al. 2002; Kidd et al. 1997). Dietary betaine supplementation has
509 been shown to increase carcass leanness (muscle accretion) and decrease carcass fat
510 thickness in pigs and poultry, as reviewed by Eklund et al. (2005). However, recent
511 studies showed that betaine supplementation for 2 days pre-transport did not improve
512 broiler meat quality and growth performance (Wray Cahen et al. 2004; Huang et al.
513 2006; Downing et al. 2017; Park and Park 2017).

514 During summer, dietary betaine enhanced rabbits growth performance during high
515 ambient temperatures and decreased RT and RR (Hassan et al. 2011). Betaine had HS
516 ameliorative effect in a dose dependent manner such that a lower dose of betaine
517 successfully ameliorated the increased skin and core temperatures, heart rates and RR,
518 however, higher betaine doses had negative impacts on these parameters (DiGiacomo
519 et al. 2014, 2016). Research by He et al. (2015) found that supplemental dietary betaine
520 reduced the effect of chronic HS on broilers body weight and fat deposition. In our lab,
521 Liu et al. (2016) found that the combination of Se, Vitamin E, Cr and betaine (based on
522 standard diets) could reduce the loss of body reserve of lactating sows during summer.
523 And Shakeri et al. (2019) demonstrated that 1g/kg betaine significantly improved the
524 growth performance of Ross-308 chicken and under cyclic heat stress condition (33 °C,
525 45-60% RH; 35 days). Dietary betaine also could reduce cadmium-induced oxidative
526 stress (rats and broilers) via increasing CAT, GPx and SOD activity (Alirezai et al.
527 2011, 2012a, 2012b). On the contrary, some studies reported that betaine
528 supplementation did not change pig lipid metabolism (Fernandez-Figares et al. 2002;

529 Wray Cahen et al. 2004). As dietary betaine may contribute to an animal's antioxidant
530 capacity, it therefore could affect animal meat quality characteristics such as colour,
531 WHC and lipid oxidation. The study by Liu et al. (2015) suggested that betaine and
532 guanidinoacetic acid supplementation could increase the pork pH_{24h} and improve meat
533 WHC and WBSF values. In broilers, dietary betaine improved meat pH_{24h} and WHC
534 (Fu et al. 2016). However, further studies are required to demonstrate similar effects of
535 betaine on meat quality of ruminants finished during summer.

536 **3.3.5 OTHER POTENTIAL FEED ADDITIVES FOR THE PREVENTION OF HEAT**

537 **STRESS DAMAGE**

538 In addition to the existing feed additives, other antioxidant compounds or growth
539 promotion additives might also play an important role in prevention of HS and
540 improvements in meat quality. These additives have been shown to improve antioxidant
541 capacity in animal feeding experiments but require further experimentation in animals
542 finished under HS conditions. For example, addition of 50-200 mg/kg lycopene (red
543 carotenoid) to diets significantly decreased broiler muscle TBARS, CAT, GSH-Px
544 levels and growth performance (Marzoni et al. 2014; Jiang et al. 2015). Other studies
545 involving meat sensory tests showed that supplementation with 15mg/kg β -carotene
546 significantly increased cooked chicken meat juiciness as compared with 200 mg/kg
547 Vitamin E supplementation, and WBSF as compared with control group, however, on
548 the other hand, β -carotene diets had negative effect on cooked meat odor and flavor
549 (Ruiz et al. 2001). Carnosine, a dipeptide natural antioxidant, significantly improved
550 pig plasma GSH-Px, SOD activity and muscle glycogen content when supplemented in
551 the diet at 100 mg/kg (Ma et al. 2010). Looking at the improvements in animal
552 antioxidant capacity following supplementation of these additives, it is tempting to

553 hypothesize that these feed additives may have potential benefits on meat quality of
554 animals finished during HS conditions and therefore warrant further investigation.

555 **4. CONCLUSION**

556 HS is one of the greatest challenges facing livestock production and is likely to be
557 exacerbated by rising ambient temperatures due to climate change. There has been vast
558 interest from an animal production and welfare point of view in the development of HS
559 mitigation strategies to reduce the economic losses incurred by the global livestock
560 industry during summer. Significant progress has been achieved in understanding the
561 impacts of HS on physiological and metabolic status of animals affecting animal
562 performance. Research in recent years has been focused on HS impacts on animal
563 product quality and suitable interventions to mitigate these negative impacts. There is
564 some evidence that HS preslaughter may lead to higher incidence of PSE in pigs and
565 poultry, and DFD in ruminants. However, these influences are not consistent across
566 different studies and vary depending on the severity and duration of HS exposure. There
567 is a dearth of well-designed controlled environmental studies comparing impacts of
568 high ambient temperature on meat quality of ruminants. These research gaps need to be
569 addressed and would be a step forward before developing suitable strategies to alleviate
570 HS impacts on meat quality. Nevertheless, there are some nutritional strategies that
571 have been found effective to ameliorate some of the negative effects of HS on animal
572 physiology and metabolic responses, such as electrolytes, flavonoids, Vitamin E and
573 betaine.

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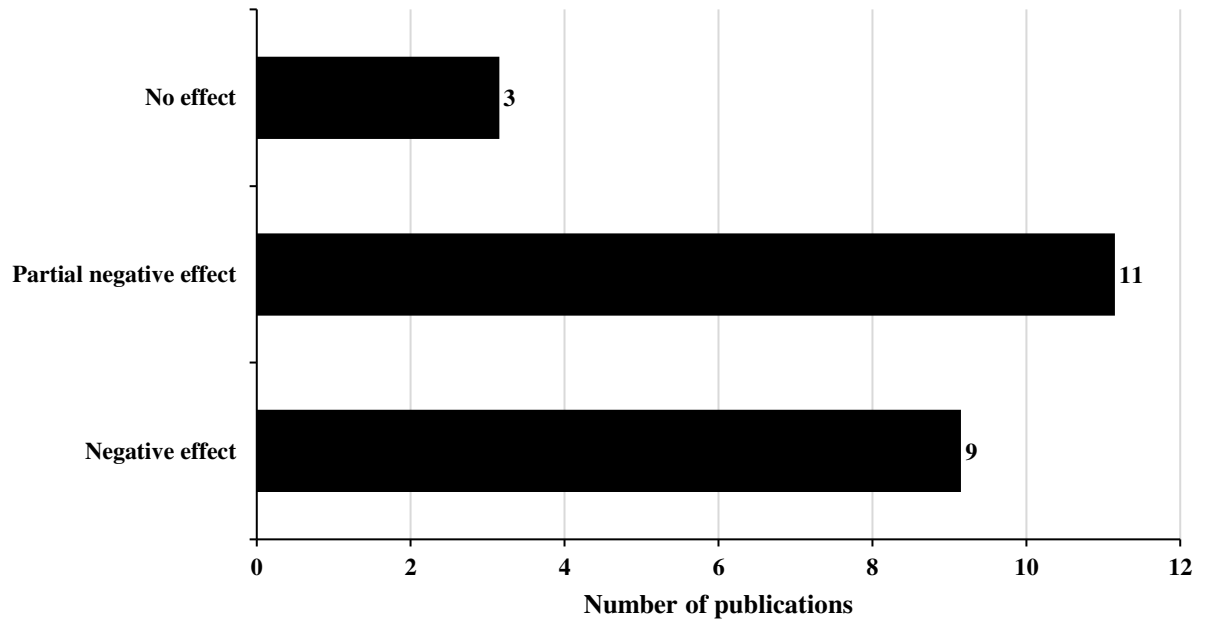
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1077 **Figure 1.** Summary of effects of heat stress HS in animals on subsequent meat quality
 1078 across 23 studies.

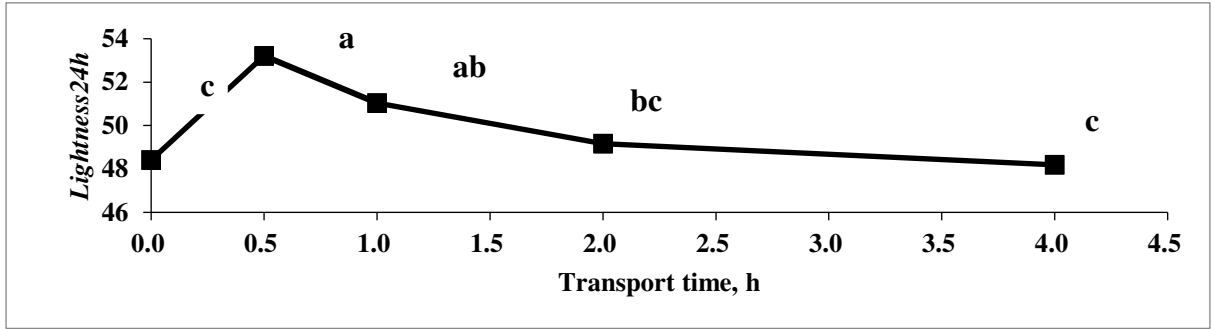
1079 Negative effect= (Sams and McKee 1997, Kadim et al. 2004, Akşit 2006, Kadim
 1080 2007, Feng et al. 2008, Kadim et al. 2008, Dai et al. 2009, Zhang et al. 2012, Kadim
 1081 et al. 2014);

1082 Partial negative effect= (Holm and Fletcher 1997, Sandercock 2001, Debut et al.
 1083 2003, Maria et al. 2006, Lu et al. 2007, Saha et al. 2013, Shi et al. 2017, Imik et al.
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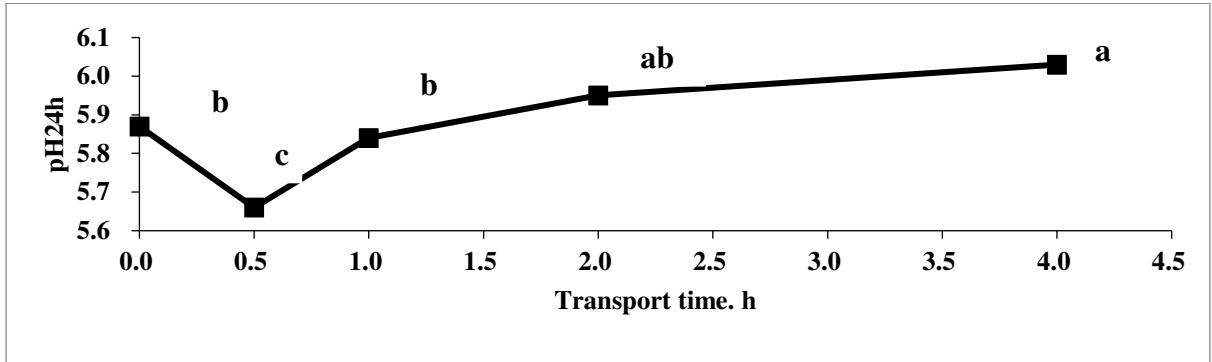
1085 No effect= (Froning et al. 1978, Owens et al. 2000, Zeferino et al. 2013);

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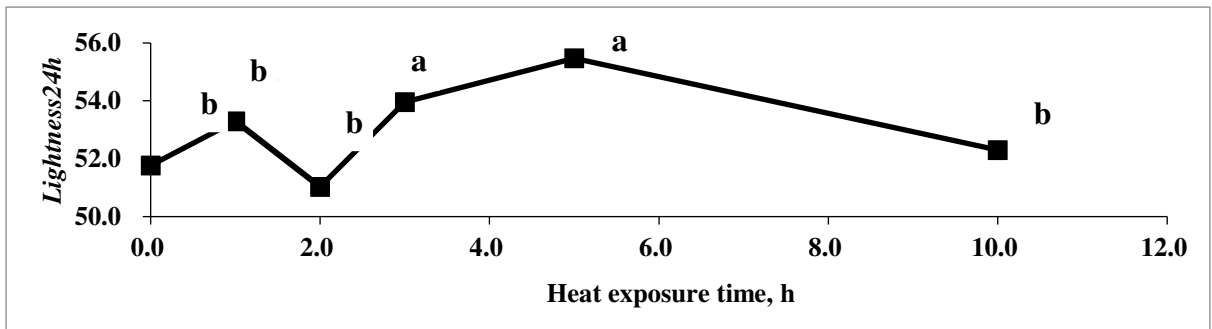
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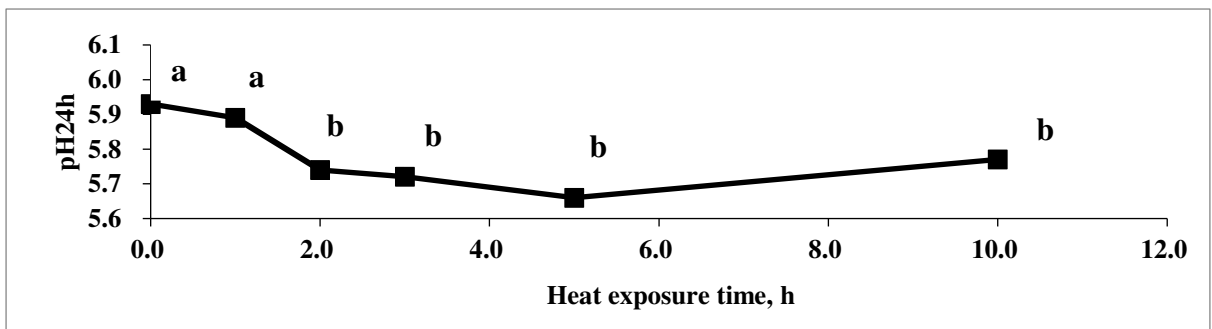
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1090 **Figure 2.** Effect of transport time on *pectoralis major* muscle pH and lightness (L*)
 1091 at 24 hrs post-slaughter (pH24h, Lightness24h respectively) for AA broiler chickens
 1092 (market age) exposed to different transportation conditions (40-42°C); n=21, 21, 21,
 1093 17, and 14 chickens for control, 0.5h, 1h, 2h, and 4h transport groups, respectively
 1094 (Adapted from Xing et al., 2016). a-c means the significant difference ($P < 0.05$)
 1095 within different transport time

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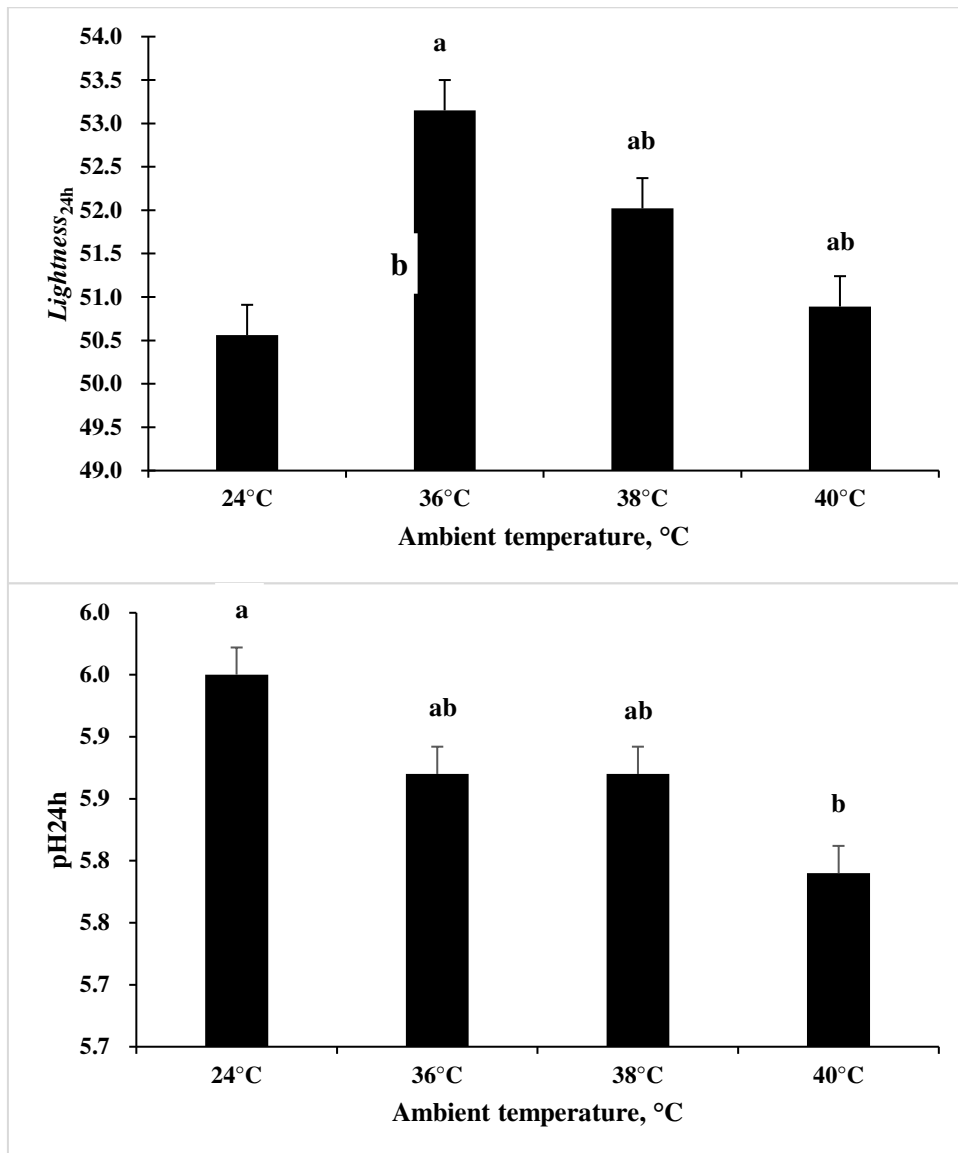


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Figure 3. Effect of time of exposure to acute HS (40°C) before slaughter on *pectoralis major* muscle pH and lightness at 24 hrs post-slaughter (pH24h, Lightness24h respectively) for AA broiler (28 days age) (n=10). Adapted from Tang et al.(2013). ab, indicates a significant difference ($P < 0.05$) between heat exposure times.



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Figure 4. Effect of different temperature of acute heat stress on *pectoralis major* meat lightness and pHu at 24 hrs post-slaughter (pH24h, Lightness24h respectively) for AA broiler (42 days) (n=12) (Zhang et al., 2019). ab, indicates a significant difference ($P < 0.05$) between heat exposure times.

Table 1. The effect of acute and chronic heat stress on ruminant growth performance and meat quality

Authors	Breed	Treatment	Muscle	Observation
Kadim et al. (2008)	Goat: Omani, Somali Sheep: Somali, Merino	Hot season (35°C, 47%RH) Cool season (21°C, 59%RH)	Psoas major and minor	Hot season decreased meat lightness (24 h postmortem), WHC, increased MFI and pHu.
Kadim et al. (2014)	Dhofari goats	Summer transportation (6 hrs. 42°C)	Longissimus thoracis et lumborum	Pre-slaughter transportation during high ambient temperatures increased meat pHu decreased meat lightness, WHC and WBSF at 48 h postmortem.
Ponnampalam et al. (2016)	Poll Dorset x [Border Leicester x Merino] lamb	HS:28-40°C TN:18-21°C 1 week	longissimus lumborum	HS had no significant influence on pHu of meat and slaughter or carcass weight. TBARS also has no significant different.
Archana et al. (2018)	Osmanabadi and Salem Black goat	Out-shade and in-shade feeding 28 to 40 °C and 29 to 58%RH, 1 month	Longissimus thoracis et lumborum	HS significant increased meat pHu (24 h postmortem), WBSF (12 h postmortem) and decreased lightness at 12 h postmortem.

Kadim et al. (2004)	Omani beef	Hot season 34.3±1.67 °C, 48.87.57% RH	Longissimus thoracis	Ambient temperatures of approximately 35°C elevated meat pHu, darkened meat colour (lightness reduced) and decreased cooking loss than those under 21°C.
		Cold season 21.2±1.40 °C, 57.91.61% RH		

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HS=heat stress, TN=thermoneutral, RH=relative humidity, WHC=water holding capacity, MFI=myofibril fragmentation index, WBSF= warner–bratzler shear force, TBARS=thiobarbituric acid reactive substances.

Table 2. The effect of acute and chronic heat stress on broiler and pig meat quality

Author	Breed	Stress treatment	Muscle	Observation
Xing et al. (2015)	Arbor Acres broiler	Summer transportation; 0-4 h.	Pectoralis major	0.5h summer transpiration had the most negative effect on broiler meat quality (lightness, cooking loss and drip loss) but not include redness and yellowness. Meat quality parameters got closer to control group as transportation time and death rate increased.
Sandercock et al. (2001)	Broiler (35d, 63d age)	HS:32°C, 75% RH TN:21°C, 75%RH 2 h.	Pectoralis major	HS only increased meat drip loss (age of 35 days) and hemorrhage score, but had no effect on colour score and sensory test results.
Tang et al. (2013)	Arbor Acres broiler	HS:37°C; 0 to 10 h TN:22°C.	Pectoralis major	Acute HS had significant effect on chicken meat quality. Meat had the poorest quality under 2 h acute HS condition. As exposure time of HS extended, the negative effect of HS gradually decreased.

Zhang et al. (2019)	Arbor Acres broiler	HS:36°C, 38°C, 40°C; TN:25°C 1 h., 2 h.	Pectoralis major	36°C HS significant increased chicken meat lightness and cooking loss. In 40°C temperature, chicken meat quality parameters had no significant difference with control group (25°C)
Lu et al. (2007)	Arbor Acres broiler, AA Beijing You chicken, BJY	HS:34°C TN:21°C 3 weeks.	Pectoralis major	The data showed that chronic heat exposure had negative effects on growth performance, breast yield, and meat quality of AA broilers, but it had no significant influence on growth and meat quality in the local, slow-growing chickens.
Aksit et al. (2006)	Broilers	HS: 34°C TN:22°C -28°C 4 weeks.	Pectoralis major	22-28°C and 34°C HS both had negative effect on chicken meat pHu and lightness and growth performance including food consumption and conversion.
Shi et al. (2017)	American Landrace barrows	HS:35°C, 78% RH TN:22 °C, 81%RH 30 days.	Longissimus dorsi	HS significantly decreased feed intake (P<0.05), but meat quality had no difference with control group (22°C), except intramuscular fat contend.