

This is a repository copy of *Cell Wall Composition Impacts Structural Characteristics of the Stems and Thereby Biomass Yield*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/188746/>

Version: Accepted Version

Article:

López-Malvar, Ana, Santiago, Rogelio, Souto, Xose Carlos et al. (3 more authors) (2022) Cell Wall Composition Impacts Structural Characteristics of the Stems and Thereby Biomass Yield. *Journal of Agricultural and Food Chemistry*. 3136–3141. ISSN: 1520-5118

<https://doi.org/10.1021/acs.jafc.1c06986>

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

**Cell Wall Composition Impacts Structural Characteristics of the Stems and thereby
Biomass Yield**

López-Malvar, Ana^{1*}; Santiago, Rogelio^{1,2}; Souto, Xose Carlos ³; Barros-Rios, Jaime⁴;
Gómez, Leonardo D.⁵; Malvar, Rosa Ana²

¹Facultad de Biología, Departamento de Biología Vegetal y Ciencias del Suelo,
Universidade de Vigo, As Lagoas Marcosende, 36310 Vigo, Spain. Agrobiología
Ambiental, Calidad de Suelos y Plantas (UVIGO), Unidad Asociada a la MBG (CSIC),
Vigo, Spain

²Misión Biológica de Galicia (CSIC), Pazo de Salcedo, Carballeira 8, 36143 Pontevedra,
Spain

³E.E. Forestales, Dpto. Ingeniería Recursos Naturales Y Medio Ambiente, Universidade
de Vigo, 36005, Pontevedra, Spain

⁴Division of Plant Sciences and Technology, College of Agriculture Food and Natural
Resources and Interdisciplinary Plant Group, Christopher S. Bond Life Sciences Center,
University of Missouri, Columbia, MO 65211, USA

⁵CNAP, Department of Biology, University of York, Heslington YO10 5DD, York,

UK*corresponding autor: alopezmalvar@uvigo.es

Abstract

Maize stalks support leaves and reproductive structures, functionally support water and nutrient transport; besides their anatomical and biochemical characteristics have been described as a plant defense against stress, also impacting economically important applications. In this study, we evaluated agronomical and stem description traits in a subset of maize inbred lines that showed variability for cell wall composition in the internodes. Overall, a great proportion of lignin subunit G and a low concentration of *p*-coumaric acid and lignin subunit S is beneficial for greater rind puncture resistance and taller plants, with greater biomass yield. Also, the greater the proportions of subunit H, the longest the internode. By last, the lower the total hemicellulose content the greater the rind puncture resistance. Our results confirmed the effect of the cell wall on agronomic and stalk traits which would be useful in applied breeding programs focused on biomass yield improvement.

Keywords: *Zea mays*, cell wall, stem characteristics, biomass, maize, plant architecture

Introduction

The structure and function of the plant cell wall are controlled by how each of its components interacts within the cell wall. This strong assembly, apart from providing structural support and rigidity to the cell, determining its size and shape, also provides resistance to abiotic and biotic stresses and communication among cells ^{1,2}. Furthermore, the framework constituted by the cell wall is closely related to the growth and fitness of the plant and is expected to determine the functional characteristics of the stem, which are closely related to yield. ^{3,4}.

From a breeder point of view, the first goal of the crop improvement is, on the one hand, to obtain increased grain yield, considered as the potential of the grain production and increased biomass yield expressed as tons of biomass produced per hectare ^{5,6}. In maize, increases in maize grain have been accompanied by increases in biomass, which indicates that breeding for biomass yield would not compromise grain yield ⁷. Furthermore, increases in biomass or stover yield have been also a target trait for biofuel production and forage digestibility ^{8,9}.

Because cell walls constitute more than 50% of the dry biomass weight; improvement of biomass relies largely on the cell wall components and anatomical arrangement of the stems, conditioning also plant height ⁹⁻¹². However, increases in plant height must have to deal with stem lodging losses. Stem lodging, caused by the bending or breaking of the stalk, is greatly impacted by stalk strength and stem morphological traits, therefore it could be said that maize stem strength impacts both grain yield and silage quality ^{10,13,14}.

Research on cell wall composition and its influence in basic and applied aspects of maize stem strength would be important steps in maize breeding and improvement ^{10,15}.

58 Overall, in the current study, we evaluated agronomical and stem description traits in a
59 subset of maize inbred lines that showed variability for cell wall composition, being the
60 main goal to identify cell wall components that can be used in applied breeding
61 programs.

Materials and Methods

Plant Material and experimental design

A set of 20 inbred lines was tested through two consecutive years (2016, 2017) in Pontevedra (Spain, 42° 24' 22.3" N, 8° 38' 28.16" W, 20 m above sea level). The set of inbred lines evaluated can be subdivided into three subsets: (i) inbred lines included in previous evaluations for resistance to *Sesamia nonagrioides* or *Ostrinia nubilalis*, (ii) inbreds used in hybrids combinations for bioenergy and for silage, (iii) inbreds that perform well in hybrid combinations. A complete and detailed description of the inbreds evaluated could be found in ¹⁶.

In both trials, the set was evaluated following a random block design with three repetitions. In 2017, the set was reduced to nineteen because there was not enough stock for the inbred line PB130. The experimental plots consisted of three rows, with 15 double-kernel hills each, with a total surface of 0.14 m² per plot, with a final density of ~ 70,000 plants ha⁻¹ after thinning. The trials were maintained with local agronomical practices.

Agronomic Traits

Biomass yield. Seventy days after silking, considering it as days from planting until half of the plants in the plot showed visible silks; plots were harvested. Two to ten plants without ears from each plot were collected, weighed and chopped from which a stover sample was collected (sample fresh weight) for estimating the percentage of stover dry matter. For that, the fresh stover was pre-dried (35 °C) in a forced air-drying chamber, dried on a stove (60 °C), and again weighed after a week (sample dry weight).

Determination of Biomass yield in Mg ha⁻¹ was done as it follows:

$$84 \quad \text{Biomass yield } \left(\frac{\text{Mg}}{\text{ha}}\right) = \frac{\text{weight of fresh sample (g)} * \text{sample dry weight (g)}}{\text{Surface (m}^2\text{)} * \text{sample fresh weight(g)} \times 100}$$

85 The surface was calculated as the number of plants per plot multiplied by the space
86 between rows (0.80 m) and the space between plants (0.18 m). Following this equation
87 biomass yield corresponds to the maximum yield.

88 **Stem Lodging.** Calculated at harvest as the sum of broken plants (split underneath the
89 main ear) divided by the total number of plants in the plot, was calculated. Stem lodging
90 is expressed in percentage.

91 **Stem descriptions traits**

92 Rind puncture resistance, the total number of internodes, and internode diameter were
93 recorded 55 days after flowering, the rest of the stem description traits 70 days after
94 flowering. A more detailed description of the methodology can be found in López-
95 Malvar et al. 16

96 Briefly, plant height was calculated as the mean of plant height (in cm) measured from
97 the base of the plant until the flag leaf, of five plants per plot; internode length was
98 calculated as the total number of internodes divided by the height of the plant; in five
99 plants, rind puncture resistance was the measured of the maximum force required to
100 puncture the rind (in kg/section) on one side of the stalk using an Accuforce Cadet Force
101 Gauge (Ametek, Mansfield and Green Division, Largo, FL); from the same five
102 internodes, using an electronic caliper, the diameter was recorded, in millimeters.

103 **Biochemical Traits**

The complete characterization of the cell wall was performed in the second internode below the main ear from five plants per plot, collected 55 days after silking. The complete description of the methodology can be found in López-Malvar et al. ¹⁶

Briefly, cellulose was quantified in crude cell walls by the Updegraff method ¹⁸; hemicellulose composition was determined using high-performance anion exchange chromatography (HPAEC) (Carbopac PA-10; Dionex, Camberley, Surrey, UK) as described previously by Jones et al. (2003), it included the quantification of glucose, galactose, fucose, arabinose, rhamnose, xylose, mannose, arabinose:xylose ratio glucuronic and galacturonic acid (the sum of all of them would be considered further on as total hemicellulose content); total lignin content was determined by Klason Lignin protocol ²¹; subunit composition was determined by thioacidolysis followed by Gas chromatography–mass spectrometry (GC–MS)²²; cell wall-bound hydroxycinnamates quantification was performed by High-Performance Liquid Chromatography (HPLC) following the protocol described in Santiago and col. ²³

Statistical Analysis

Contrast analysis

The SAS mixed model procedure (PROC MIXED) of the SAS program (version 9.4) ²⁵ was used for the individual and combined analyses of variance for each trait. Using the combined data for the analysis across years, the best linear unbiased estimators (BLUES) for each inbred line was calculated. We considered as fixed effects inbred lines and as random effects years, replication within years, and lines × year. We used Fisher's protected least significant difference (LSD) for means comparison.

126 After that analysis, inbred lines were qualitatively classified, according to their BLUEs,
127 in high, intermediate, and low groups for agronomic and stem traits (Table 1); high and
128 low groups differing $p < 0.05$. With the qualitative dataset, mean comparisons for groups
129 with contrasting values were performed to look for differences in cell wall composition.

130 Table 1. Inbred lines under study were qualitatively classified according to the BLUES
131 for biomass yield and stem description traits evaluated in 2016 and 2017.

Inbred	Plant Height (cm)	Internode Length (cm)	Internode Diameter (mm)	Rind Puncture Resistance (kg/section)	Stover Yield (Mg/ha)
A509	Low	Intermediate	Intermediate	Low	Low
A632	High	Low	Low	Intermediate	High
A654	Low	Low	Intermediate	Intermediate	Intermediate
C103	High	High	Intermediate	High	High
CO348	Low	Low	Intermediate	High	Intermediate
CO384	High	Low	Intermediate	Intermediate	High
CO442	High	Low	Intermediate	Low	High
CO444	Intermediate	Low	Low	Intermediate	Intermediate
EC212	Intermediate	Low	Intermediate	High	Intermediate
EP105	High	Intermediate	Intermediate	Intermediate	Intermediate
EP125	High	High	Intermediate	Low	Low
EP17	High	Low	High	High	High
EP42	Intermediate	Low	Low	Intermediate	Intermediate
EP47	High	High	High	Intermediate	High
EP53	Low	High	Intermediate	High	Low
EP86	High	Intermediate	Low	Low	Low
F473	Low	Low	Low	Intermediate	Low
PB130	Low	Low	High	Intermediate	Intermediate

W182B	Low	Low	High	Low	Low
W64A	Intermediate	Low	High	Intermediate	Intermediate

132

133 **Multiple linear regression analysis**

134 For understanding the relationship between agronomic, stem description traits, and cell
135 wall components we performed a multiple linear regression model using the BLUEs. For
136 this analysis, we used, in SAS ²⁵, the stepwise method following the PROC REG
137 procedure. Variables with a significance value less than 0.15 were not selected to take
138 part in the regression model. We considered as dependent variables agronomic and stem
139 description traits; as independent variables, we considered cell wall components.

Results

Inbred lines differed significantly for biomass yield and stem description traits. There were no significant differences for stem lodging, so it was not included nor in the contrast analysis, nor the multiple linear regression (Supplementary Table 1).

Contrast analysis

Significant differences between high and low contrast groups for every trait are shown in Table 2. Values for non-significant traits in the contrast analysis are included in Supplementary Table 2. Inbred lines presenting the greatest biomass yield showed the lower concentration of PCA, low proportion of lignin subunits S and H, low S:G ratio, and on the opposite greater proportion of subunit G.

In the same way, but attending to the plant height, the taller plants presented lower proportions of lignin subunit S and S:G ratio, and higher proportions of lignin subunit G, however, the H subunit showed the opposite trend for biomass yield.

Regarding to internode description traits, the greater proportion of subunit H, the longest the internode, in accordance with plant height results. Contrast groups for internode diameter did not differ for any cell wall trait. Finally, inbred lines showing the greater resistance to puncture were the ones showing the greatest cellulose content and the greatest proportions of subunit G; and the lowest concentrations of cell wall-bound hydroxycinnamates (namely PCA and diferulates), lowest total hemicellulose content (galactose, glucuronic and galacturonic acid, arabinose, xylose, and mannose) and lowest proportions of lignin subunit H and S:G ratio (Table 2).

Table 2. Contrast analysis of inbred lines attending to contrasting values of biomass yield and agronomic stem description traits. Only cell wall components that significantly differ among groups are included.

	Classification Group			
Cell Wall Component	High	Intermediate	Low	LSD
Biomass yield (Mg/ha)				
PCA (mg/g)	11.54	12.77	13.94	0.907
S subunit (%)	55.28	57.83	57.83	0.982
S:G ratio	1.317	1.44	1.479	0.052
G subunit (%)	42.17	39.27	39.27	0.851
H subunit (%)	2.548	1.95	2.910	0.052
Plant height (cm)				
S subunit (%)	55.65	57.26	58.88	0.98
S:G ratio	1.34	1.42	1.52	0.052
G subunit (%)	41.64	40.75	38.85	0.89
H subunit (%)	2.72	2.28	2.28	0.38
Internode length (cm)				
H subunit (%)	2.95	2.26	2.26	0.44
Rind puncture resistance (kg/section)				
PCA (mg/g)	11.38	12.70	13.90	0.99
DFA 8-5-l (mg/g)	0.046	0.060	0.062	0.008
DFA 8-5-b (mg/g)	0.088	0.105	0.105	0.156

DFA 5-5 (mg/g)	0.067	0.087	0.086	0.012
DFAT (mg/g)	0.274	0.338	0.326	0.045
Cellulose (mg/g)	441.63	441.63	382.11	34.324
Galactose (mg/g)	4.855	5.600	8.285	2.926
Galacturonic acid (mg/g)	6.734	9.230	10.166	2.422
Glucuronic acid (mg/g)	2.472	2.748	3.936	1.935
Arabinose (mg/g)	8.002	9.218	12.248	3.016
Mannose (mg/g)	2.429	2.586	3.699	0.80
Xylose (mg/g)	20.87	24.21	24.75	3.051
H Subunit (%)	2.248	2.356	2.703	0.398
G subunit (%)	41.15	40.73	39.77	1.041
S:G ratio	1.39	1.40	1.45	0.06
<p>LSD: Least Square Distance ($P \leq 0.05$)</p> <p>PCA: <i>p</i>-coumaric acid; DFA 8-5-l: Diferulic acid 8-5-Linear; DFA 8- o-4: Diferulic acid 8-O-4; DFA 8-5: Diferulic acid 8-5; DFA85b: Diferulic acid 8-5-Benzofuran; DFAT: Total diferulic acids</p> <p>* some missing data for individual traits and inbreds could interfere in the final ratio calculations of the groups</p>				

165

166 Multiple Linear Regression

167 We found that a greater proportion of lignin subunit G and greater Total Hemicellulose
168 Content (mainly galactose) increase biomass yield; on the opposite, greater Galacturonic
169 acid and arabinose:xylose ratio decrease biomass yield (Table 3).

We found that 34% of the variance for plant height was affected lignin subunit S, with a negative effect. In the case of internode length and internode diameter, no variable met the 0.15 significance level to be included in the model (Table 3). Rind puncture resistance was mainly affected by Galacturonic acid concentration and Arabinose:Xylose Ratio, negatively, and positively by Glucose reporting 53 % of the variation for rind puncture resistance (Table 3).

Table 3. Multiple linear regression model (using stepwise selection) of biomass yield and stem description traits on cell wall composition of a set of inbred lines evaluated in 2016 and 2017.

Stepwise Selection			
Biomass yield (Mg/ha)		R² Partial	R²
Subunit G (%)		0.31	0.31
Arabinose:Xylose Ratio		0.14	0.46
Total Hemicellulose (mg/g)		0.09	0.55
Galacturonic Acid (mg/g)		0.07	0.62
Galactose (mg/g)		0.07	0.69
Model	Biomass yield: $-16.11997 + 0.48330 \cdot G - 5.68752 \cdot \text{ARA:XYL} + 0.55854 \cdot \text{Galactose} - 0.38920 \cdot \text{Galacturonic Acid} + 0.03730 \cdot \text{Total Hemicellulose}$		
Plant height (cm)		R² Partial	R²

Subunit S (%)		0.34	0.34
Model	Plant height= 469.3563-5. 72250*S		
Rind puncture resistance (kg/section)		R ² Partial	R ²
Galacturonic acid (mg/g)		0.36	0.36
Arabinose:Xylose Ratio (mg/g)		0.11	0.47
Glucose (mg/g)		0.07	0.53
Model	Rind puncture resistance: 3.83618 + 0.01978*Glucose - 0.15986*Galacturonic Acid -0.58579*Arabinose:Xylose Ratio		
R ² : Total % of the variance explained by the model; R ² partial: % of the variance explained by each trait.			

Discussion

Our results confirm that biomass, stem strength, and other stem features such as plant height or internode length rest on the organization and composition of the stem cell walls. Secondary cell wall formation, characterized by lignin deposition, seems to play a central role in maize stem characteristics.

Contrast analysis

In the contrast analysis, we noted that it is not the total lignin content, but the lignin subunit composition the trait that most influences differences groups of lines classified and in high and low groups for plant biomass and stem architecture; the lignin with higher proportions of subunit G in detriment of subunit S, is valuable for both increased biomass yield, plant height and rind puncture resistance (Table 2). The composition and proportion of the subunits highly influence the molecular structure of lignin. It affects the degree of crosslinking with the polysaccharides and also the branching of the polymer, affecting, as it has been demonstrated, economically important processes such as biofuel production and digestibility^{16,26}. In addition, some other related phenylpropanoids also contributed to biomass yield and anatomical traits of the stems. We have shown that a great concentration of PCA in the cell wall is unfavorable for increasing biomass yield and rind puncture resistance. Most PCA is bound to S units in lignin, esterified to the γ -position of phenylpropanoid sidechains²⁷. The PCA acylation influences the bonding mode of S lignin units and on the spatial organization of lignin, and by consequence also on the way that lignin and polysaccharides interact²⁸. In this sense, S-type lignin presents a more linear structure²⁹ with almost no branching and with a lesser degree of polymerization; lignin G is more condensed than lignin S³⁰. In

our case, S-type lignin, is detrimental for increases in biomass yield, plant height, and rind puncture resistance.

The network formed by the fibres within the cell wall (cellulose-lignin-hemicellulose) is believed to define the functional properties of the stems ³¹. We found that increases in cellulose would favor greater rind puncture resistance, therefore stalk strength, while greater concentrations of total hemicellulose content would be disadvantageous for rind puncture resistance. Increases in rind puncture resistance and formation of the cortex tissue have been closely related with cellulose and lignin deposition, serving as structural support to the cell wall ³². Moreover, cellulose compositional features, such as crystallinity, have been related to stalk lodging and stalk strength, which could be associated with rind puncture resistance, as previously mentioned ¹⁵. It has been also proved the positive association between the quantity of cellulose amorphous regions and the arabinose-substitution of xylans; also the negative effect that increasing levels of arabinose have in cellulose crystallinity has been demonstrated ^{33,34}. In the contrast analysis, the group of inbred lines presenting the higher rind puncture resistance present a reduced arabinose:xylose ratio and thus a reduction in the arabinose content. In a cell wall presenting a low concentration of arabinose, the hemicellulose and cellulose chains tend to interact through of hydrogen bonds, which would contribute more crystalline cellulose, which is more uniform, ordered, and hard; which could indicate a greater resistance to puncture. Contrary to cellulose, hemicelluloses are not chemically uniform. Xylan containing β -(1,4)-linked xylose residues, is one of the most complex heteroxylans in the fibre of maize ³⁵. Based on Appeldoorn et al. ³⁶ and Van Eylen et al. ³⁷ a reduced incidence of uronic acid, acetic acid, and arabinose side groups in glucuronoarabinoxylans would drive changes in the properties of the cell wall. Contrast

analysis showed that the presence of more glucuronic acid and galacturonic acid may contribute to a less strengthen stalk, less resistant to puncture.

Finally, the mechanical resistance granted by DFAs would make us think in a cell wall with greater strength and higher tissue toughness, would also present a greater resistance to the penetrometer, however, regarding our contrast analysis, the group of inbred lines showing the greatest rind puncture resistance showed the lowest concentrations of diferulates. But our results are in accordance with the ones obtained by Manga-Robles et al.¹⁵ in a previous study. They observed a significantly higher level in diferulic individual dimers in inbred lines showing low rind penetrometer strength. Attending to the plasticity of the cell wall we may argue that some of the other components of the cell wall have a more significant part in the strengthening and support like, for this panel of inbred lines, great cellulose content or lignin presenting low S:G ratio, which would increase rind puncture resistance.

Multiple linear Regression

Mainly, the results obtained in the multiple linear regression analysis support the ones obtained in the contrast analysis. Again, the influence of lignin subunit composition and how PCA acetylation of lignin subunit affected the final lignin structure, showed significant effects on biomass yield and plant height. Lignin with a greater proportion of subunit G may be beneficial for greater biomass yield, and lignin presenting lower proportions of subunit S would produce taller plants.

We have already mentioned how the fibre proportion of the cell wall takes part in determining stem anatomical characteristics, according to our results the structural support granted by Total Hemicellulose content produces greater biomass yields.

Besides, we found that reduced arabinose:xylose ratio, and lower concentrations of galacturonic acid decrease both Rind Puncture Resistance (accordance with contrast analysis) and Biomass Yield. As previously explained for the contrast analysis results, the influence of matrix polysaccharides (total hemicellulose content) has been confirmed to affect rind puncture resistance; and in the same way, could affect biomass yield. It has been demonstrated the negative relationship between arabinose content and cellulose crystallinity. The intra and intermolecular hydrogen bridges within the cellulose have as a result a crystalline configuration that gives cellulose mechanical solidity, which may be beneficial for biomass increases ³⁸.

In this representative material, S-type lignin accompanied by increases in *p*-coumaric acid would be in detriment of biomass yield, plant height, and rind puncture resistance, whereas, cell walls richer in cellulose and with a lower proportion of total hemicellulose, would be beneficial for stalk strength (Figure 1). These results prove that cell wall composition clearly influences structural characteristics of the maize stems and thereby can be useful to improve maize biomass yield.

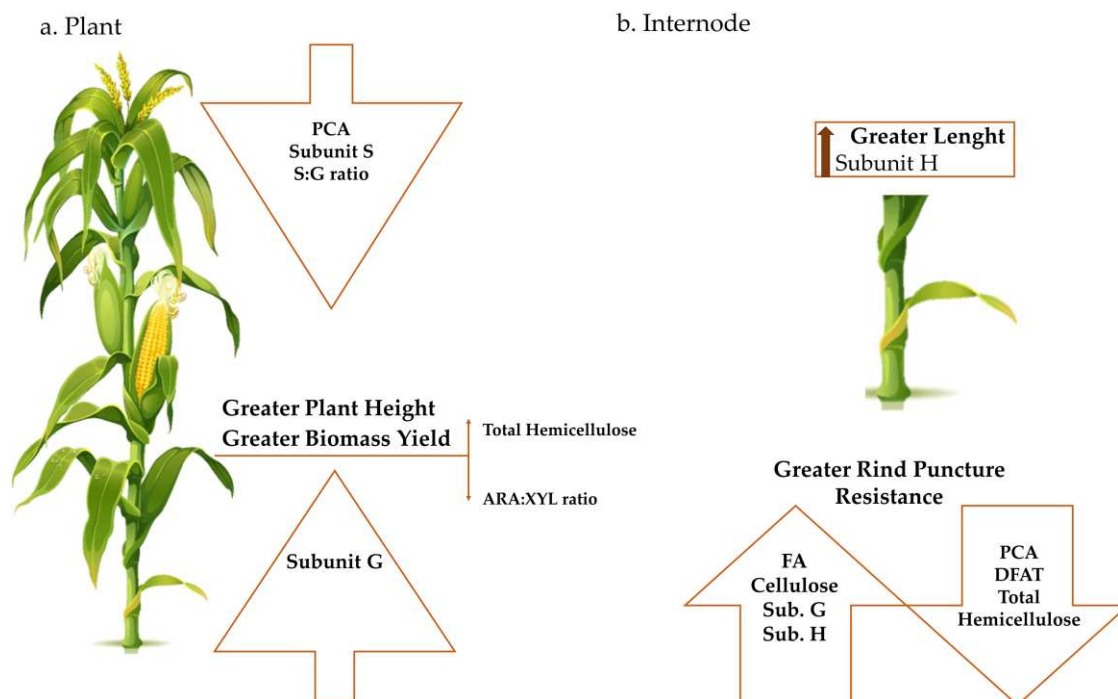


Figure 1. Graphical summary of the results obtained. a: Results concerning the whole plant; b: results concerning the second internode below the main ear.

List of Abbreviations

TFA: Trifluoroacetic acid; HPAEC: high-performance anion exchange chromatography; GC-MS: Gas chromatography-mass-spectrometry; BLUES: Best Linear Unbiased Estimators; FA: Ferulic acid; PCA: p-coumaric acid; DFA 8-5-l: Diferulic acid 8-5-Linear; DFA 8- o-4: Diferulic acid 8-O-4; DFA 8-5: Diferulic acid 8-5; DFA85b: Diferulic acid 8-5-Benzofuran; DFAT: Total diferulic acids; G: Subunit; GLUA: Glucuronic Acid

Supplementary material

Supplementary Table 1: Means of 20 inbred lines evaluated for agronomic and stem description traits.

Supplementary Table 2. Contrast analysis of inbred lines attending to contrasting values of biomass yield and agronomic stem description traits. Means for cell wall components with **non-significant differences** among groups are included.

Acknowledgements

We would like to thank Dr. Rachael Simister for the technical assistant of matrix polysaccharides analyses. We are grateful to Prof. Simon J McQueen Mason for their support at the CNAP, University of York, UK and the Dixon laboratory at University of North Texas for support with the lignin compositional analyses.

Funding

This research has been developed in the frame of the 'Agri-Food Research and Transfer Centre of the Water Campus (CITACA) at the University of Vigo (Spain), which is economically supported by the Galician Government and in the Misión Biológica de Galicia-CSIC. It was funded by the "Plan Estatal de Ciencia y Tecnología de España" (projects RTI2018-096776-B-C21, and RTI2018-096776-B-C22 co-financed with European Union funds under the FEDER program). The funding body played no role in study design, data analysis, and manuscript preparation.

Authors' contributions

RAM and RS conceived the study. RS, RAM, and AL, participated in the experimental design, carried out the field trials, and participated in sample collection; AL carried out biochemical determinations and statistical analysis; AL wrote the draft. JBR and LG

301 assisted on biochemical analysis, results and discussion. XCS contributed to the results
302 discussion. All authors read and approved the final manuscript.

References

- (1) Carpita, N. C.; Defernez, M.; Findlay, K.; Wells, B.; Shoue, D. A.; Catchpole, G.; Wilson, R. H.; McCann, M. C. Cell Wall Architecture of the Elongating Maize Coleoptile. *Plant Physiol.* **2001**, 127 (2), 551–565.
<https://doi.org/10.1104/pp.010146>.
- (2) Houston, K.; Tucker, M. R.; Chowdhury, J.; Shirley, N.; Little, A. The Plant Cell Wall: A Complex and Dynamic Structure as Revealed by the Responses of Genes under Stress Conditions. *Front. Plant Sci.* **2016**, 7 (AUG2016), 1–18.
<https://doi.org/10.3389/fpls.2016.00984>.
- (3) Brulé, V.; Rafsanjani, A.; Pasini, D.; Western, T. L. Hierarchies of Plant Stiffness. *Plant Sci.* **2016**, 250, 79–96. <https://doi.org/10.1016/j.plantsci.2016.06.002>.
- (4) Scheller, H. V.; Ulvskov, P. Hemicelluloses. *Annu. Rev. Plant Biol.* **2010**, 61, 263–289. <https://doi.org/10.1146/annurev-arplant-042809-112315>.
- (5) Vermerris, W.; Saballos, A.; Ejeta, G.; Mosier, N. S.; Ladisch, M. R.; Carpita, N. C. Molecular Breeding to Enhance Ethanol Production from Corn and Sorghum Stover. *Crop Sci.* **2007**, 47 (SUPPL. DEC.).
<https://doi.org/10.2135/cropsci2007.04.0013IPBS>.
- (6) Sinclair, T. R.; Purcell, L. C.; Sneller, C. H. Crop Transformation and the Challenge to Increase Yield Potential. *Trends Plant Sci.* **2004**, 9 (2), 70–75.
<https://doi.org/10.1016/j.tplants.2003.12.008>.
- (7) Mazaheri, M.; Heckwolf, M.; Vaillancourt, B.; Gage, J. L.; Burdo, B.; Heckwolf, S.; Barry, K.; Lipzen, A.; Ribeiro, C. B.; Kono, T. J. Y.; Kaeppler, H. F.; Spalding, E. P.; Hirsch, C. N.; Robin Buell, C.; de Leon, N.; Kaeppler, S. M. Genome-Wide Association Analysis of Stalk Biomass and Anatomical Traits in Maize. *BMC*

- 327 *Plant Biol.* **2019**, 19 (1), 1–17. <https://doi.org/10.1186/s12870-019-1653-x>.
- 328 (8) López-Malvar, A.; Butron, A.; Malvar, R. A.; McQueen-Mason, S. J.; Faas, L.;
 329 Gómez, L. D.; Revilla, P.; Figueroa-Garrido, D. J.; Santiago, R. Association
 330 Mapping for Maize Stover Yield and Saccharification Efficiency Using a
 331 Multiparent Advanced Generation Intercross (MAGIC) Population. *Sci. Rep.*
 332 **2021**, 11 (1), 1–9. <https://doi.org/10.1038/s41598-021-83107-1>.
- 333 (9) Heckwolf, S.; Heckwolf, M.; Kaeppler, S. M.; de Leon, N.; Spalding, E. P. Image
 334 Analysis of Anatomical Traits in Stalk Transections of Maize and Other Grasses.
 335 *Plant Methods* **2015**, 11 (1). <https://doi.org/10.1186/s13007-015-0070-x>.
- 336 (10) Wang, Y.; Li, J. Genes Controlling Plant Architecture. *Curr. Opin. Biotechnol.*
 337 **2006**, 17 (2), 123–129. <https://doi.org/10.1016/j.copbio.2006.02.004>.
- 338 (11) Hansey, C. N.; Lorenz, A. J.; de Leon, N. Cell Wall Composition and Ruminant
 339 Digestibility of Various Maize Tissues across Development. *Bioenergy Res.* **2010**,
 340 3 (1), 28–37. <https://doi.org/10.1007/s12155-009-9068-4>.
- 341 (12) Brancourt-Hulmel, M.; Arnoult, S.; Cézard, L.; El Hage, F.; Gineau, E.; Girones,
 342 J.; Griveau, Y.; Jacquemont, M. P.; Jaffuel, S.; Mignot, E.; Mouille, G.; Lapierre,
 343 C.; Legée, F.; Méchin, V.; Navard, P.; Vo, L. T. T.; Reymond, M. A Comparative
 344 Study of Maize and Miscanthus Regarding Cell-Wall Composition and Stem
 345 Anatomy for Conversion into Bioethanol and Polymer Composites. *Bioenergy*
 346 *Res.* **2021**. <https://doi.org/10.1007/s12155-020-10239-z>.
- 347 (13) Peng, C.; Wang, X.; Feng, T.; He, R.; Zhang, M.; Li, Z.; Zhou, Y.; Duan, L. System
 348 Analysis of MIRNAs in Maize Internode Elongation. *Biomolecules* **2019**, 9 (9).
 349 <https://doi.org/10.3390/biom9090417>.
- 350 (14) Ordas, B.; Malvar, R. A.; Santiago, R.; Butron, A. QTL Mapping for

351 Mediterranean Corn Borer Resistance in European Flint Germplasm Using
 352 Recombinant Inbred Lines. *BMC Genomics* **2010**, 11 (1).
 353 <https://doi.org/10.1186/1471-2164-11-174>.

354 (15) Manga-Robles, A.; Santiago, R.; Malvar, R. A.; Moreno-González, V.; Fornalé, S.;
 355 López, I.; Centeno, M. L.; Acebes, J. L.; Álvarez, J. M.; Caparros-Ruiz, D.; Encina,
 356 A.; García-Angulo, P. Elucidating Compositional Factors of Maize Cell Walls
 357 Contributing to Stalk Strength and Lodging Resistance. *Plant Sci.* **2021**, 307
 358 (December 2020). <https://doi.org/10.1016/j.plantsci.2021.110882>.

359 (16) López-Malvar, A.; Malvar, R. A.; Gomez, L. D.; Barros-rios, J.; Pereira-crespo, S.
 360 Elucidating The Multifunctional Role of The Cell Wall Components in The
 361 Maize Exploitation. *BMC Plant Biol.* **2021**, 21, 251.

362 (17) Mérida, H.; Encina, A.; Álvarez, J.; Acebes, J. L.; Caparrós-Ruiz, D. Unraveling
 363 the Biochemical and Molecular Networks Involved in Maize Cell Habituation to
 364 the Cellulose Biosynthesis Inhibitor Dichlobenil. *Mol. Plant* **2010**, 3 (5), 842–853.
 365 <https://doi.org/10.1093/mp/ssq027>.

366 (18) Updegraff, D. M. Semimicro Determination of Cellulose Inbiological Materials.
 367 *Anal. Biochem.* **1969**, 32 (3), 420–424. [https://doi.org/10.1016/S0003-2697\(69\)80009-](https://doi.org/10.1016/S0003-2697(69)80009-6)
 368 6.

369 (19) Dische, Z. Color Reactions of Carbohydrates. In *Methods in carbohydrate*
 370 *chemistry, Vol.1*; Whistler, R.L. and Wolfrom, M. L. E., Ed.; Academic Press: New
 371 York, 1962; pp 475–514.

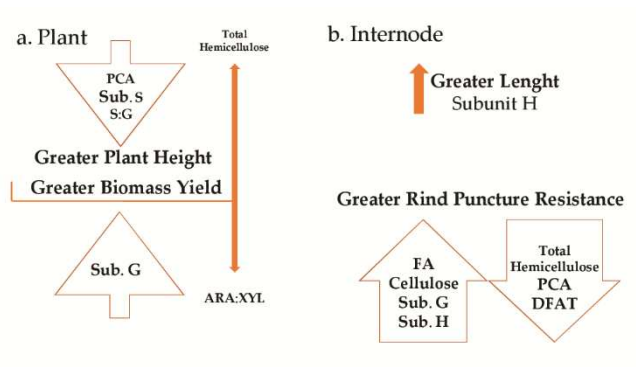
372 (20) Jones, L.; Milne, J. L.; Ashford, D.; McQueen-Mason, S. J. Cell Wall Arabinan Is
 373 Essential for Guard Cell Function. *Proc. Natl. Acad. Sci. U. S. A.* **2003**, 100 (20),
 374 11783–11788. <https://doi.org/10.1073/pnas.1832434100>.

- 375 (21) Dence, C. W. The Determination of Lignin. **1992**, 33–61.
 376 https://doi.org/10.1007/978-3-642-74065-7_3.
- 377 (22) Barros, J.; Escamilla-Trevino, L.; Song, L.; Rao, X.; Serrani-Yarce, J. C.; Palacios,
 378 M. D.; Engle, N.; Choudhury, F. K.; Tschaplinski, T. J.; Venables, B. J.; Mittler, R.;
 379 Dixon, R. A. 4-Coumarate 3-Hydroxylase in the Lignin Biosynthesis Pathway Is
 380 a Cytosolic Ascorbate Peroxidase. *Nat. Commun.* **2019**, *10* (1), 1–11.
 381 <https://doi.org/10.1038/s41467-019-10082-7>.
- 382 (23) Santiago, R.; López-Malvar, A.; Souto, C.; Barros-Ríos, J. Methods for
 383 Determining Cell Wall-Bound Phenolics in Maize Stem Tissues. *J. Agric. Food*
 384 *Chem.* **2018**, *66* (5), 1279–1284. <https://doi.org/10.1021/acs.jafc.7b05752>.
- 385 (24) Waldron, K. W.; Parr, A. J.; Ng, A.; Ralph, J. Cell Wall Esterified Phenolic
 386 Dimers: Identification and Quantification by Reverse Phase High Performance
 387 Liquid Chromatography and Diode Array Detection. *Phytochem. Anal.* **1996**, *7*
 388 (6), 305–312. [https://doi.org/10.1002/\(SICI\)1099-1565\(199611\)7:6<305::AID-](https://doi.org/10.1002/(SICI)1099-1565(199611)7:6<305::AID-PCA320>3.0.CO;2-A)
 389 [PCA320>3.0.CO;2-A](https://doi.org/10.1002/(SICI)1099-1565(199611)7:6<305::AID-PCA320>3.0.CO;2-A).
- 390 (25) SAS Institute. Base SAS 9.4 Procedures Guide. *SAS iNstitute* **2015**.
- 391 (26) Méchin, V.; Argillier, O.; Menanteau, V.; Barrière, Y.; Mila, I.; Rollet, B.; Lapierre,
 392 C. Relationship of Cell Wall Composition to in Vitro Cell Wall Digestibility of
 393 Maize Inbred Line Stems. *J. Sci. Food Agric.* **2000**, *80* (5), 574–580.
 394 [https://doi.org/10.1002/\(SICI\)1097-0010\(200004\)80:5<574::AID-JSFA575>3.0.CO;2-](https://doi.org/10.1002/(SICI)1097-0010(200004)80:5<574::AID-JSFA575>3.0.CO;2-R)
 395 [R](https://doi.org/10.1002/(SICI)1097-0010(200004)80:5<574::AID-JSFA575>3.0.CO;2-R).
- 396 (27) Ralph, J.; Hatfield, R. D.; Quideau, S.; Helm, R. F.; Grabber, J. H.; Jung, H. J. G.
 397 Pathway of P-Coumaric Acid Incorporation into Maize Lignin As Revealed by
 398 NMR. *J. Am. Chem. Soc.* **1994**, *116* (21), 9448–9456.

- 399 (28) Barrière, Y.; Méchin, V.; Riboulet, C.; Guillaumie, S.; Thomas, J.; Bosio, M.; Fabre,
400 F.; Goffner, D.; Pichon, M.; Lapierre, C.; Martinant, J. P. Genetic and Genomic
401 Approaches for Improving Biofuel Production from Maize. *Euphytica* **2009**, *170*
402 (1), 183–202. <https://doi.org/10.1007/s10681-009-9923-6>.
- 403 (29) Kishimoto, T.; Uraki, Y.; Ubukata, M. Easy Synthesis of β -O-4 Type Lignin
404 Related Polymers. *Org. Biomol. Chem.* **2005**, *3* (3), 1067–1073.
- 405 (30) Anderson, N. A.; Tobimatsu, Y.; Ciesielski, P. N.; Ximenes, E.; Ralph, J.;
406 Donohoe, B. S.; Ladisch, M.; Chapple, C. Manipulation of Guaiacyl and
407 Syringyl Monomer Biosynthesis in an Arabidopsis Cinnamyl Alcohol
408 Dehydrogenase Mutant Results in Atypical Lignin Biosynthesis and Modified
409 Cell Wall Structure. *Plant Cell* **2015**, *27* (8), 2195–2209.
410 <https://doi.org/10.1105/tpc.15.00373>.
- 411 (31) Wilson, J. R.; Hatfield, R. D. Structural and Chemical Changes of Cell Wall
412 Types during Stem Development: Consequences for Fibre Degradation by
413 Rumen Microflora. *Aust. J. Agric. Res.* **1997**, *48* (2), 165–180.
414 <https://doi.org/10.1071/A96051>.
- 415 (32) Xue, J.; Zhao, Y.; Gou, L.; Shi, Z.; Yao, M.; Zhang, W. How High Plant Density of
416 Maize Affects Basal Internode Development and Strength Formation. *Crop Sci.*
417 **2016**, *56* (6), 3295–3306. <https://doi.org/10.2135/cropsci2016.04.0243>.
- 418 (33) Li, F.; Ren, S.; Zhang, W.; Xu, Z.; Xie, G.; Chen, Y.; Tu, Y.; Li, Q.; Zhou, S.; Li, Y.;
419 Tu, F.; Liu, L.; Wang, Y.; Jiang, J.; Qin, J.; Li, S.; Li, Q.; Jing, H. C.; Zhou, F.;
420 Gutterson, N.; Peng, L. Arabinose Substitution Degree in Xylan Positively
421 Affects Lignocellulose Enzymatic Digestibility after Various NaOH/H₂SO₄
422 Pretreatments in Miscanthus. *Bioresour. Technol.* **2013**, *130*, 629–637.

<https://doi.org/10.1016/j.biortech.2012.12.107>.

- (34) Marriott, P. E.; Gómez, L. D.; Mcqueen-Mason, S. J. Unlocking the Potential of Lignocellulosic Biomass through Plant Science. *New Phytol.* **2016**, 209 (4), 1366–1381. <https://doi.org/10.1111/nph.13684>.
- (35) Saha, B. C.; Bothast, R. J. Pretreatment and Enzymatic Saccharification of Corn Fiber. *Appl. Biochem. Biotechnol. - Part A Enzym. Eng. Biotechnol.* **1999**, 76 (2), 65–77. <https://doi.org/10.1385/ABAB:76:2:65>.
- (36) Appeldoorn, M. M.; Kabel, M. A.; Van Eylen, D.; Gruppen, H.; Schols, H. A. Characterization of Oligomeric Xylan Structures from Corn Fiber Resistant to Pretreatment and Simultaneous Saccharification and Fermentation. *J. Agric. Food Chem.* **2010**, 58 (21), 11294–11301. <https://doi.org/10.1021/jf102849x>.
- (37) Van Eylen, D.; van Dongen, F.; Kabel, M.; de Bont, J. Corn Fiber, Cobs and Stover: Enzyme-Aided Saccharification and Co-Fermentation after Dilute Acid Pretreatment. *Bioresour. Technol.* **2011**, 102 (10), 5995–6004. <https://doi.org/10.1016/j.biortech.2011.02.049>.
- (38) Kumar, M.; Turner, S. Plant Cellulose Synthesis: CESA Proteins Crossing Kingdoms. *Phytochemistry* **2015**, 112 (1), 91–99. <https://doi.org/10.1016/j.phytochem.2014.07.009>.



449

450