



This is a repository copy of *Genetic variance in fitness indicates rapid contemporary adaptive evolution in wild animals*.

White Rose Research Online URL for this paper:
<https://eprints.whiterose.ac.uk/188184/>

Version: Accepted Version

Article:

Bonnet, T., Morrissey, M.B., de Villemereuil, P. et al. (37 more authors) (2022) Genetic variance in fitness indicates rapid contemporary adaptive evolution in wild animals. *Science*, 376 (6596). pp. 1012-1016. ISSN 0036-8075

<https://doi.org/10.1126/science.abk0853>

This is the author's version of the work. It is posted here by permission of the AAAS for personal use, not for redistribution. The definitive version was published in *Science* on 26th May 2022, Vol 376, Issue 6596 pp.1012-1016, <https://doi.org/10.1126/science.abk0853>.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Title: Genetic variance in fitness indicates rapid contemporary adaptive evolution in wild animals

Authors: Timothée Bonnet^{1*}, Michael B. Morrissey², Pierre de Villemereuil^{3,4}, Susan C. Alberts⁵, Peter Arcese⁶, Liam D. Bailey⁷, Stan Boutin⁸, Patricia Brekke⁹, Lauren J. N. Brent¹⁰,
5 Glauco Camenisch¹¹, Anne Charmantier¹², Tim H. Clutton-Brock^{13,14}, Andrew Cockburn¹,
David W. Coltman⁸, Alexandre Courtiol⁷, Eve Davidian⁷, Simon R. Evans^{15,16}, John G. Ewen⁹,
Marco Festa-Bianchet¹⁷, Christophe de Franceschi¹², Lars Gustafsson¹⁶, Oliver P. Höner⁷,
Thomas M. Houslay¹³, Lukas F. Keller¹¹, Marta Manser^{11,14}, Andrew G. McAdam¹⁸, Emily
McLean¹⁹, Pirmin Nietlisbach²⁰, Helen L. Osmond¹, Josephine M. Pemberton²¹, Erik Postma²²,
10 Jane M. Reid^{23,24}, Alexis Rutschmann⁴, Anna W. Santure⁴, Ben C. Sheldon¹⁵, Jon Slate²⁵, Céline
Teplitsky¹², Marcel E. Visser²⁶, Bettina Wachter⁷, Loeske E.B. Kruuk^{1,21}

Affiliations:

¹Research School of Biology, Australian National University; Canberra ACT 2600, Australia.

²School of Biology, University of St Andrews; St Andrews, Fife, United Kingdom.

³Institut de Systématique, Évolution, Biodiversité (ISYEB), École Pratique des Hautes Études, PSL, MNHN, CNRS, SU, UA; Paris, France.

⁴School of Biological Sciences, University of Auckland; Auckland, New Zealand.

⁵Departments of Biology and Evolutionary Anthropology, Box 90338, Duke University; Durham NC 27708, USA.

⁶Forest and Conservation Sciences, University of British Columbia; Vancouver, British Columbia, Canada.

⁷Departments of Evolutionary Ecology and Evolutionary Genetics, Leibniz Institute for Zoo and Wildlife Research; Alfred-Kowalke-Str. 17, 10315 Berlin, Germany.

⁸Department of Biological Sciences, University of Alberta; 11455 Saskatchewan Drive, Edmonton, AB, T6G 2E9, Canada.

⁹Institute of Zoology, Zoological Society of London; Nuffield Building, Regents Park, London NW1 4RY, United Kingdom.

¹⁰Centre for Research in Animal Behaviour, University of Exeter; United Kingdom.

¹¹Department of Evolutionary Biology and Environmental Studies, University of Zurich; Zurich, Switzerland.

¹²Centre d'Écologie Fonctionnelle et Évolutive, Université de Montpellier, CNRS, EPHE, IRD; Montpellier, France.

¹³Department of Zoology, University of Cambridge; Downing Street, Cambridge CB2 3EJ, United Kingdom.

¹⁴Mammal Research Institute, University of Pretoria; South Africa.

¹⁵Edward Grey Institute, Department of Zoology, University of Oxford; United Kingdom.

¹⁶Department of Ecology and Genetics/Animal Ecology, Uppsala University; Uppsala, Sweden.

¹⁷Département de Biologie, Université de Sherbrooke; Sherbrooke, Québec, Canada.

¹⁸Department of Ecology and Evolutionary Biology; University of Colorado, Boulder, CO 80309, USA.

¹⁹Oxford College of Emory University; Oxford, GA 30054, USA.

²⁰School of Biological Sciences, Illinois State University; Normal, IL 61790-4120, USA.

²¹Institute of Evolutionary Biology, University of Edinburgh; Edinburgh, United Kingdom.

²²Centre for Ecology and Conservation, University of Exeter; Penryn, TR4 7AS, United Kingdom.

²³Centre for Biodiversity Dynamics, NTNU; Trondheim, Norway.

²⁴School of Biological Sciences, University of Aberdeen; Aberdeen, United Kingdom.

5 ²⁵Ecology and Evolutionary Biology, School of Biosciences, University of Sheffield; Sheffield, S10 2TN, United Kingdom.

²⁶Department of Animal Ecology, Netherlands Institute of Ecology (NIOO-KNAW); P.O. Box 50, 6700 AB Wageningen, The Netherlands.

*Corresponding author. Email: timotheebonnetc@gmail.com

10

Abstract: The rate of adaptive evolution, the contribution of selection to genetic changes that increase mean fitness, is determined by the additive genetic variance in individual relative fitness. To date, there are few robust estimates of this parameter from natural populations, and it is therefore unclear whether adaptive evolution can play a meaningful role in short-term population dynamics. We applied new quantitative genetic methods to long-term datasets from 15 19 wild bird and mammal populations, and found that, while estimates vary between populations, additive genetic variance in relative fitness is often substantial, and on average double previous estimates. We show that these rates of contemporary adaptive evolution can impact population dynamics, and hence that natural selection has the potential to partly mitigate effects of current 20 environmental change.

One-Sentence Summary: Genetic variance in fitness in 19 wild vertebrate populations suggests adaptive evolution is currently common and rapid.

Main Text:

How fast are wild populations currently evolving in response to natural selection? The rate of adaptive evolution in nature is both of fundamental theoretical importance, and also of increasing practical relevance given the clear impact of human activities on the environment experienced by wild organisms (1). Numerous examples of phenotypic and genetic changes for traits under selection (2-5) suggests that adaptive evolution commonly occurs in wild populations over contemporary timescales, although many studies have found that trait changes do not correspond to adaptive expectations and even suggest evolutionary stasis (6, 7). However, analysis of the evolution of specific traits is unlikely to represent the overall rate of adaptation of a population, as natural selection acts on many traits concurrently. Instead, a comprehensive assessment of the rate of adaptive evolution in a population needs to integrate adaptive genetic changes across all traits that determine individual fitness, the contribution of an individual to the gene pool of the next generation. According to Fisher's Fundamental Theorem of Natural Selection, the per-generation proportional change in mean absolute fitness caused by natural selection is given by the additive genetic variance in relative fitness, $V_{A(w)}$ (8-10). In non-technical terms, $V_{A(w)}$ is the extent of heritable (transmitted from parents to offspring) genetic differences in the ability to reproduce. The realized change in mean fitness between generations may not exactly be reflected by $V_{A(w)}$, because of concurrent effects of genetic mutations, gene flow, environmental change or gene-environment interactions (8, 9, 11). Nonetheless, a non-zero value of $V_{A(w)}$ indicates that, all else being equal, natural selection contributes to an increase in mean fitness (8, 9). It also indicates that at least some of the traits that determine individual fitness are currently evolving in response to selection. Thus, $V_{A(w)}$ is arguably the most important evolutionary parameter in any population (9, 12).

Robust estimation of $V_A(w)$ requires accurate measures both of individual fitness and of pairwise genetic relatedness for large numbers of individuals. Such data are difficult to collect for wild populations of animals or plants (13). Moreover, their analysis is made challenging by the distribution of individual fitness, which generally does not conform well to common statistical methods (14). Consequently, our knowledge of $V_A(w)$ in natural populations is currently limited: two reviews report estimates of $V_A(w)$ from 16 populations of 13 plant and (non-human) animal species with fitness measured over complete lifetimes (12, 14; we discuss these results alongside our own below). However, notwithstanding possible issues specific to each analysis (such as the omission of important non-genetic sources of similarity between relatives), most of these estimates were obtained from Gaussian models (for exceptions see 10) which generally do not fit the distribution of fitness well. In natural populations, the distribution of fitness of all individuals is typically both highly right-skewed, with most individuals having low values but a few having very high values, and zero-inflated, with an excess of zeroes over and above that otherwise expected (zero-inflation may for example be generated by high levels of juvenile mortality). Estimates of $V_A(w)$ from Gaussian models, and their associated uncertainty, may thus be unreliable (14, 15).

Here, we address the gap in our knowledge of the values of $V_A(w)$ in the wild and its implications in terms of adaptation, trait evolution and population dynamics. We apply new Bayesian quantitative genetic methods to data from long-term studies of 19 free-living vertebrate populations with high-quality lifetime reproduction and multi-generational relatedness data. Covering more populations and species than all previous studies combined, these 19 populations of 15 different species (6 birds and 9 mammals) have contrasting ecologies, life histories and social systems (10, SI Table S1-2) and are located in diverse terrestrial biomes and continents (Fig. 1). Our analysis is restricted to birds and mammals because of their predominance among

long-term studies with suitable data (13). The populations have been monitored for between 11 and 63 years, providing fitness records for 561 fully monitored cohorts totaling 249,430 individuals of both sexes (10). For all data-sets used here, an individual's fitness was measured as 'lifetime breeding success', the total number of offspring produced over its lifetime, irrespective of offspring survival. While there are numerous definitions of fitness, each motivated by different theoretical frameworks (16), measuring fitness as lifetime breeding success corresponds most closely to a life-cycle-calibrated 'zygote-to-zygote' definition of individual fitness, consistent with quantitative genetic theory (17). Individuals were identified soon after birth or hatching, and fitness was estimated for all known individuals in each population, including the often large proportion that died as juveniles (10). We modeled absolute lifetime breeding success using a quantitative genetic form of mixed effects model known as an 'animal model' (18), assuming that lifetime breeding success followed zero-inflated over-dispersed Poisson distributions and including relevant covariates (such as inbreeding, genetic group, sex and cohort. See 10, Table S3-4, Text S1 for model details, Fig. S1-2 for evaluation of model goodness of fit, Text S2, Fig. S3 for prior distribution). The zero-inflated Poisson models were fitted to absolute fitness data and the resulting parameter estimates, obtained on link-function scales, were then back-transformed to derive estimates of $V_A(w)$ and other components of variances for relative fitness on the scale of the data (15). We first ran one model for each study population, and subsequently combined results into a meta-analysis (10).

We found evidence for additive genetic variance in relative fitness in multiple populations. Our models provided estimates of $V_A(w)$ with posterior modes ranging from 0.003 to 0.497 (Fig 2A). The 95% credible intervals (95%CI) for $V_A(w)$ excluded values below 0.001 in ten of the 19 populations, and excluded values below 0.01 in eight (thresholds explained in caption of Fig. 2A, Text S2, S3). Therefore, there was clear evidence that selection contributed to genetic changes

that would increase mean fitness in roughly half of the study populations (9, 19). Across populations, the median of the posterior modes for $V_A(w)$ was 0.100 and the meta-analytic mean of $V_A(w)$ was 0.185, 95%CI [0.088; 0.303]. There was also considerable variation among populations, with a meta-analytic among-population standard deviation in $V_A(w)$ of 0.11, 95%CI [0.01; 0.26]. The median and mean values of $V_A(w)$ were about four and two times larger than those of previous estimates (previous median 0.0225; previous mean 0.092; 12, 14). Our values can be considered large given theoretical considerations (SI Text S3, Fig. S4). Our estimates were robust to the modeling of possible confounders: inbreeding, sex, linear environmental changes in mean fitness, gene-flow due to immigration, variance among cohorts and among mothers (10) and also mother-by-cohort interactions, social group effects (SI Text S4, Table S5, Fig. S5) and the social inheritance of social dominance within families (SI Text S5, Fig. S6-7). For completeness, we also present estimates relating to an alternative formulation of Fisher's Fundamental Theorem expressing change in terms of absolute fitness (Text S6, Fig. S8).

Previous work on adaptive evolution has often focused on the heritability of fitness, $h^2(w) = V_A(w)/V_P(w)$, where $V_P(w)$ is the phenotypic variance in relative fitness, or 'opportunity for selection' (20). However, $h^2(w)$ may be a poor measure of the overall rate of adaptive evolution (20). In natural conditions, stochastic or unaccounted environmental variation is expected to dominate variation in individual fitness, even in the presence of large deterministic sources of variation in fitness (21), so that $h^2(w)$ may be small even when $V_A(w)$ is large (21, 22). In line with this expectation, we found that $h^2(w)$ was generally small, with a meta-analytic average of 2.99%, 95%CI [0.80; 6.60%] and a value of less than 1% in 11 populations (Fig. 2B), similar to previous estimates of $h^2(w)$ (14). Nevertheless, estimates of $h^2(w)$ were of similar magnitude to the proportion of variance explained by maternal effect and cohort variances (Fig. 2B, SI Text S7, Table S6-10 for parameter estimates on different scales). Furthermore, $h^2(w)$ was highly

variable between populations and was sometimes substantial, with posterior modes ranging from 0.019% to 17.1%.

What do our estimates of $V_A(w)$ imply about the evolution of traits in our study populations?

$V_A(w)$ is the partial increase in fitness expected to result from the combined responses to

5 selection across heritable traits (23). Therefore, a non-zero $V_A(w)$, as in at least half of our study populations, implies that for one or several traits, the responses to selection tend to cause

adaptive change, although the total change may be affected by mutations or environmental

change (19). The value of $V_A(w)$ sets an upper bound for the possible per-generation response to selection of any trait (19). Given the meta-analytic estimate of $V_A(w)=0.185$, and a trait with a

10 heritability of 0.3 (an average value for trait heritability in wild populations, 24), the maximal

rate of response to selection is 0.24 standard deviations per generation (10, 19). Across our 19

populations, the upper bound of response to selection for a trait with a heritability of 0.30 varies

from 0.05, 95%CI [0.01;0.13], to 0.39, 95%CI [0.29;0.50] standard deviations. These upper

bounds are substantial: for comparison, in natural populations the rates of phenotypic change,

15 irrespective of whether the change is known to be adaptive, are rarely above 0.03 standard

deviations (around 10% of estimates), and only very rarely above 0.13 standard deviations

(around 5% of estimates; 2). Evolutionary studies of wild populations, including several

conducted in our study populations, have often failed to detect phenotypic change in response to

current selection (5, 6, 25). Our results may therefore appear at odds with these observations.

20 However, attempts to estimate genetic evolution of traits, as opposed to just phenotypic trends,

remain rare and under-powered (25). Genetic evolution of traits may be masked at the

phenotypic level, either because phenotypic plasticity hides genetic change (6) or because direct

evolution is counterbalanced by the evolution of ‘indirect genetic effects’, that is, the effect of

other individuals’ genotypes (26). Moreover, approaches to estimating genetic change for a trait,

such as estimation of trends in individual genetic merit ('breeding values') (27) or by estimation of polygenic scores (28), may have limited statistical power. Finally, if $V_A(w)$ is ultimately driven by the cumulative effects of many traits evolving in response to selection, the evolutionary change in each trait will be small and even more difficult to identify statistically.

5 Any or all of these scenarios could prevent observed rates of phenotypic change in single traits reaching the upper bound of what might be possible given the observed levels of $V_A(w)$.

Irrespective of the rates of adaptive evolution in the potentially many traits that contribute to $V_A(w)$, our estimates of their combined effect, summarized in $V_A(w)$, indicate that adaptive evolution may have substantially affected recent population dynamics (see Text S6, S8, Fig. S8).

10 For instance, in a thought experiment assuming that no forces oppose adaptive evolution and that $V_A(w)$ remains constant, 11 out of our 19 populations would recover from an arbitrary one-third reduction in fitness in fewer than 10 generations (SI Text S8). Moreover, the median $V_A(w)$ of 0.10 means that in half the populations, natural selection tends to increase mean absolute fitness of at least 10% every generation. Such a change would lead to exponential population growth if not counterbalanced. Yet none of our study populations showed any exponential increase in
15 population size such as predicted by the thought experiment (SI Text S9). This indicates that any adaptive evolution was countered by simultaneous deleterious effects of other processes such as mutation, gene flow, or environmental changes (19). The presence of these counterbalancing forces, as well as potential changes in future selective pressures and the potential instability of
20 $V_A(w)$ in future environments, make it impossible to project whether the contemporary adaptive evolution that our results indicate is sufficiently fast and lasting to ensure population persistence. Other studies that focused on specific traits, rather than on the net effect of selection on fitness, suggest that short-term phenotypic changes in response to climate change are overall insufficient to ensure the persistence of populations (29, 30). Crucially, however, our finding that most

populations harbor biologically meaningful levels of additive genetic variance in fitness indicates that the machinery of adaptive evolution often operates at a substantial pace on generation-to-generation time-scales. Without ongoing adaptive genetic changes, these populations would presumably have had, often substantially, lower growth rates over recent generations.

5 **References**

1. F. Pelletier, D. W. Coltman. *BMC Biol.* **16**, 7 (2018).
2. A. P. Hendry, *Eco-Evolutionary Dynamics* (Princeton University Press, 2017), chap. 3, Adaptation.
3. J. N. Thompson, *Relentless Evolution*. (University of Chicago Press, 2013).
- 10 4. P. Karell, K. Ahola, T. Karstinen, J. Valkama, J. E. Brommer. *Nat. Comm.* **2**, 208 (2011).
5. T. Bonnet, et al. *PLOS Biol.* **17**, e3000493 (2019).
6. J. Merilä, B. C. Sheldon, L. E. B. Kruuk. *Genetica* **112**, 199–222 (2001).
7. B. Pujol, et al. *Trends Ecol. Evol.* **33**, 337–346 (2018).
8. G. R. Price. *Ann. Hum. Genet.* **36**, 129–140 (1972).
- 15 9. A. Grafen. *J. Theor. Biol.* **456**, 175–189 (2018).
10. Material and methods are provided as supplementary information.
11. D. N. Fisher, A. G. McAdam. *Evol. Lett.* **3**, 4–14 (2019).
12. A. Burt. *Evolution* **49**, 1–8 (1995).
13. T. H. Clutton-Brock, B. C. Sheldon. *Trends Ecol. Evol.* **25**, 562–573 (2010).
- 20 14. A. P. Hendry, D. J. Schoen, M. E. Wolak, J. M. Reid. *Annu. Rev. Ecol. Evol.* **49**, 457–476 (2018).

15. T. Bonnet, M. B. Morrissey, L. E. B. Kruuk. *J. Hered.* **110**, 383–395 (2019).
16. B.-E. Sæther, S. Engen. *Trends Ecol. Evol.* **30**, 273–281 (2015).
17. J. D. Hadfield. The evolution of parental care, N. J. Royle, P. T. Smiseth, M. Kölliker, eds. (Oxford Univ. Press, 2012), pp. 267–284.
- 5 18. L. E. B. Kruuk. *Phil. Trans. R. Soc. B* **359**, 873–90 (2004).
19. B. Walsh, M. Lynch, *Evolution and Selection of Quantitative Traits* (Oxford University Press, 2018), chap. 6, Theorems of natural selection.
20. T. F. Hansen, C. Pélabon, D. Houle. *Evol. Biol.* **38**, 258–277 (2011).
21. R. E. Snyder, S. P. Ellner. *Am. Nat.* **191**, E90–E107 (2018).
- 10 22. T. Price, D. Schluter. *Evolution* **45**, 853–861 (1991).
23. R. Fisher, *The Genetical Theory of Natural Selection* (Clarendon Press, Oxford, 1930), first edn.
24. E. Postma, *Quantitative Genetics in the Wild*, A. Charmantier, D. Garant, L. E. B. Kruuk, eds. (Oxford University Press, 2014), pp. 16–33, first edn.
- 15 25. J. Merilä, A. P. Hendry. *Evol. Appl.* **7**, 1–14 (2014).
26. P. Bijma. *Heredity* **112**, 61–9 (2014).
27. J. D. Hadfield, A. J. Wilson, D. Garant, B. C. Sheldon, L. E. B. Kruuk. *Am. Nat.* **175**, 116–125 (2010).
28. J. P. Beauchamp. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 7774–7779 (2016).
- 20 29. L. Browne, J. W. Wright, S. Fitz-Gibbon, P. F. Gugger, V. L. Sork. *Proc. Natl. Acad. Sci. U.S.A.* **116**, 25179–25185 (2019).

30. V. Radchuk, et al. Nat. Comm. **10**, 3109 (2019).

Acknowledgments: We acknowledge the people, organizations and traditional owners on whose land the study populations were monitored. We also thank numerous fieldworkers and funding bodies; see SI Text S10 for full acknowledgments related to each study. This work was supported by computational resources provided by the Australian Government through the National Computational Infrastructure (NCI) under the ANU Merit Allocation Scheme. We thank Ashley E. Latimer for graphic design, Luis-Miguel Chevin and Jarrod Hadfield for suggestions on early versions of this work, and Bruce Walsh and three anonymous reviewers for comments on the manuscript.

Funding: The long-term studies presented here were funded as follows (see details in SI Text S10).

Montpellier and Corsica blue tits: Observatoire de Recherche Montpelliérain de l'Environnement (OSU-OREME), Agence Nationale de la Recherche (ANR), European Research Council (ERC);

Wytham great tits: Biotechnology and Biological Sciences Research Council, European Research Council (ERC), and the UK Natural Environment Research Council (NERC).

Mandarte song sparrows: Natural Sciences and Engineering Research Council of Canada, Swiss National Science Foundation, European Research Council (ERC), Norwegian Research Council;

Gotland collared flycatchers: Swedish Research Council (VR) and Swedish Research Council for Environment, Agricultural Sciences and Spatial Planning (FORMAS);

Hihi: the New Zealand Department of Conservation, the Hihi Recovery Group Research England, Royal Society of New Zealand;

Canberra superb fairy-wrens: the Australian Research Council (ARC);

Amboseli baboons: the US National Science Foundation, the US National Institute on Aging, the Princeton Center for the Demography of Aging, the Chicago Zoological Society, the Max Planck Institute for Demographic Research, the L.S.B. Leakey Foundation, and the National Geographic Society;

Cayo Santiago macaques: the National Center for Research Resources and the Office of Research Infrastructure Programs of the National Institutes of Health;

Graubünden Snow voles: the Swiss National Science Foundation;

Kluane Red squirrels: Natural Sciences and Engineering Research Council (NSERC) and the National Science Foundation (NSF);

Ram Mountain bighorn sheep: Natural Sciences and Engineering Research Council (NSERC);

The Isle of Rum red deer and St Kilda Soay sheep: the Natural Environment Research Council (NERC);

Kalahari Meerkats: European Research Council (ERC), Human Frontier Science Program, the University of Zurich, the Swiss National Science Foundation, the Mammal Research Institute at the University of Pretoria, South Africa;

Ngorongoro spotted hyenas: the Leibniz Institute for Zoo and Wildlife Research, the Deutsche Forschungsgemeinschaft, the Deutscher Akademischer Austauschdienst, the Max Planck Society, the Werner Dessoir Stiftung.

Author contributions:

- 5 Conceptualization: TB, LEBK
- Data curation: all authors
- Methodology: TB, MBM, PdV, LEBK
- Formal analysis: TB
- Writing – original draft: TB, LEBK
- 10 Writing – review & editing: all authors

Competing interests: Authors declare that they have no competing interests.

Data and materials availability: All code and data are available in the supplementary materials.

Supplementary Materials

Materials and Methods

- 15 Supplementary Text S1 to S10
- Figs. S1 to S10
- Tables S1 to S10
- References (30–240)
- Data S1
- 20 Code S1

Fig. 1. Locations of the 19 long-term population studies. From top to bottom and then left to right: bsR = bighorn sheep on Ram Mountain, ssS = Soay sheep on St Kilda, rdR = red deer on the Isle of Rum, gtW = great tits in Wytham Woods, gtH = great tits in Hoge Veluwe, cfG = collared flycatchers on Gotland, svG = snow voles in Graubünden, rsK=red squirrels in Kluane, 5 btR = blue tits at la Rouvière, spM= song sparrows on Mandarte Island, btP = blue tits at Pirio, btM = blue tits at Muro, rmC = rhesus macaques at Cayo Santiago, ybA = yellow baboons at Amboseli, hhT = hihi on Tiritiri Matangi Island, shN = spotted hyenas in the Ngorongoro Crater, mkK = meerkats in the Kalahari, sfC = superb fairy-wrens in Canberra, hhK = hihi at Karori.

Fig. 2. Additive genetic variance and other components of variance in relative fitness.

10 Panels show posterior distributions of each parameter: **(A)** additive genetic variance in relative fitness, $V_A(w)$; **(B)** proportion of phenotypic variance in fitness due to different variance components: additive genetic variance, i.e., heritability (red), maternal effect variance (light blue), cohort variance (dark green). Species are ordered by phylogenetic proximity. Each distribution has an area of 1 but is scaled arbitrarily on the y-axis to aid comparison. Asterisks: * 15 indicates that the 95%CI of a variance component does not overlap 0.001 (approximately the mode of the prior distribution for $V_A(w)$, Text S2); ** that the 95%CI does not overlap 0.01 (the approximate threshold between small and moderate rates of adaptive evolution, Text S3); asterisks are about absolute variance values, not proportions of variance. See Fig. 1 caption for full population names.