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Associational resistance through intercropping reduces yield losses to soil-borne pests and diseases

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Summary

- Associational resistance to herbivore and pathogen attack is a well documented ecological phenomenon and, if applied to agriculture, may reduce impact of pests and diseases on crop yields without recourse to pesticides.
- The value of associational resistance through intercropping, planting multiple crops alongside each other, as a sustainable control method remains unclear, due to variable outcomes reported in the published literature. We performed a meta-analysis to provide a quantitative assessment of benefits of intercropping for target plant resistance to plant-parasitic nematodes and soil-borne diseases.
- We found that intercropping reduced damage to focal crops from nematodes by 40% and disease incidence by 55%. Intercropping efficacy varied with biological variables, such as field fertilisation status and intercrop family, and methodology, including whether study samples were potted or in fields.
- Nematode control using intercropping was sufficient to offset reductions in focal crop yield from intercrop presence, making intercropping a viable agricultural tool. We identify key drivers for underpinning the success of intercropping and indicate areas for future research to improve efficacy. This study also highlights the potential benefits of harnessing ecological knowledge on plant–enemy interactions for improving agricultural and landscape sustainability.

Introduction

Plants are at the centre of a complex network of interactions, including those with competing individuals and species, as well as natural enemies, both above and below ground (Johnson *et al.*, 2016). The nature of these interactions between plants and their herbivores and pathogens is altered by close-proximity neighbouring plants, particularly for heterospecifics (Tahvanainen & Root, 1972). Associational resistance is a recognised, widespread ecological interaction whereby specific plant associations with unpalatable neighbours decrease the likelihood of detection by, and vulnerability to, herbivore attack; in other cases, known as associational susceptibility, interactions with neighbouring plants lead to an increase in such attacks (Barbosa *et al.*, 2009).

Ecological intensification, the practice of utilising natural ecological processes to replace intensive anthropogenic inputs in agricultural systems, has been proposed as a means to increase the environmental sustainability of agriculture (Bommarco *et al.*, 2013). Future changes in climate, population and patterns of consumption are predicted to require an increase over current crop production of 100% by 2050 (Tilman *et al.*, 2011), which clearly cannot be delivered sustainably by devoting more land to agriculture or by increasing already high levels of external inputs. Crop losses due to weeds, insects, nematodes and pathogens

exacerbate this problem, standing at 26–40%, even with the best current control methods (Oerke & Dehne, 2004; Oerke, 2006). Furthermore, these losses are predicted to increase under climate change (Deutsch *et al.*, 2018). The application of associational resistance approaches could be a sustainable method to address some of the current and are likely to be future crop losses to pests.

Much research on the influence of close neighbours to plant resistance has focused on aboveground herbivores in natural systems, but plants come under attack from many other types of enemies, especially in the monocultures and high input environments of intensive agriculture. Plant natural enemies in soil, such as plant-parasitic nematodes and soil-borne diseases, reduce the yields of many crops worldwide (Shigaki *et al.*, 1998; Oerke & Dehne, 2004; Marimuthu *et al.*, 2013) and are particularly difficult to control through pesticide application (Matthiessen & Kirkegaard, 2006). Some pathogenic organisms, such as potato cyst nematodes (*Globodera* spp.), can persist in soil in a resistant state for decades (Evans & Stone, 1977), quiescent until soil pesticide concentrations are reduced to ineffective levels. Adsorption and degradation of pesticides in agricultural soils can prevent sufficient exposure for control (Munnecke, 1972), particularly in the context of restrictions on their use due to the impacts on environmental and human health (Popp *et al.*, 2013). There is an urgent need for alternatives to current practice, alternatives based

on approaches that apply a knowledge of plant ecology and biology to improve food crop production sustainably (Hartley, 2018; Pretty *et al.*, 2018). Many of these approaches, such as intercropping, in which more than one plant species is cultivated in the same plot at the same time, do not require major or technically difficult advances so could be implemented quickly, and with more research into underpinning ecological mechanisms they could be more widely adopted (Royal Society, 2009).

Some planting regimes can diversify agricultural systems, including temporal diversification through expanded rotations to include cover crops or spatial diversification via cultivar mixing and intercropping (Reiss & Drinkwater, 2018). Of these potential options, intercropping has been identified as a potential sustainable strategy for maximising crop yields while combating pest damage and disease, including those caused by soil-borne organisms (Malézieux *et al.*, 2009; Boudreau, 2013). Intercropping, applying the framework of associational resistance, may influence pest and disease damage through (1) modifying soil nutrient and water availability, (2) interfering with pest capacity to detect/recognise focal crop root structures, (3) inducing defences in focal crops through chemical cues released by the intercrop, and (4) provision of habitat for known natural enemies of focal crop pests (Barbosa *et al.*, 2009). Associational susceptibility is driven by the converse of these functional behaviours.

Studies of the value of intercropping for sustainable pest control have found mixed and sometimes conflicting results, reflecting the complex and variable nature of associational resistance and susceptibility. For example, intercropping cucumber with hot pepper, castor or crown daisy reduced root knot nematode (*Meloidogyne incognita*) attack, whereas damage increased with hairy vetch, common zinnia, and Baikal skullcap intercrops (Dong *et al.*, 2012). Intercropping can also lead to variable outcomes for focal crop diseases. For example, anise and garlic intercrops reduced damping-off and root rot in lentil regardless of whether the causal pathogen was *Rhizoctonia solani* or *Fusarium solani*, whereas intercropping with onion only protected lentil from *R. solani* (Abdel-Monaim & Abo-Elyousr, 2012).

This variability in published results is perhaps unsurprising given that intercrops can affect nematode damage or disease incidence through a range of mechanisms that, combined with the complex nature of plant–soil interactions in agroecosystems, is likely to cause significant variability in outcomes. These mechanisms can be broadly divided into three categories (following Trenbath, 1993): (1) direct impacts on the survival, activity, or reproduction of the pest/pathogen; (2) indirect impacts on the pest/pathogen via changes in the focal plant species; and (3) indirect impacts on the pest-pathogen via changes in the soil community (including natural enemies) or in the soil environment. Given the conflicting but burgeoning literature, a robust quantitative statistical review of the effects of intercropping on plant-parasitic nematode damage and on soil-borne disease incidence is needed and timely but has not been attempted to date. This sort of contingency is a barrier to uptake of sustainable approaches in agriculture systems (Doheny-Adams *et al.*, 2018; Hartley, 2018), so a better understanding of the factors driving variability in outcomes is vital.

Our study was designed to test whether intercropping is an effective agricultural tool to reduce nematode and pathogen impacts on focal crop yields. We addressed this knowledge gap using meta-analysis, an approach that has been used successfully to synthesise results from studies in field environments (e.g. Denno *et al.*, 2008; Letourneau *et al.*, 2011; Johnson *et al.*, 2012). Specifically, we used meta-analysis, combined with meta-regression, to investigate mechanisms underpinning the relationship between intercropping and damage to focal crops from parasitic nematodes or soil-borne diseases. The role of any given mechanism will be context-dependent, and we explored the influence of these contextual factors through meta-regression, a technique that quantifies the heterogeneity between experimental outcomes by fitting a range of moderator variables (Berkey *et al.*, 1995).

Considering the mechanisms of action for associational resistance, we hypothesised that resource availability, plant density/diversity as well as both intercrop and pathogen species identity/guild are primary drivers of interactions between intercrop plants, focal crop plants and herbivores/diseases in agricultural systems. For instance, we might expect greater intercrop planting density to create a more challenging ‘maze’, making it more difficult for pests to detect and move to focal crop plant roots. Similarly, intercrop plants known to produce bioactive compounds may be more likely to be effective in conferring associational resistance. At this time, exact ecological mechanisms of control remain contested in ecosystems displaying associational resistance (Agrawal *et al.*, 2006), so these parameters were targeted to provide insight into underlying mechanisms and to determine the effectiveness of intercropping. We therefore tested the influence of a range of both biological and agronomic factors on damage or disease incidence in intercropped vs nonintercropped systems, including intercrop plant family, fertiliser inputs, pathogen inoculum source (i.e. laboratory cultured or field soil population), nematode species, pathogen type, timing of inoculation and intercrop addition, experiment duration and plant density. Furthermore, we tested whether the beneficial effects of intercropping on focal crop yield through reductions in the losses due to nematode damage or disease incidence were sufficient to offset yield losses due to the presence of the intercrop, so determining whether intercropping can be an environmentally and economically sustainable alternative to chemical control of soil-borne pests and diseases.

Description

Literature search and study selection

We performed literature searches in the Web of Science (Core Collection; www.webofknowledge.com), the British Library (theses; www.explore.bl.uk), the Indian Citation Index (ICI; www.indiancitationindex.com), and the National Library of Australia Trove (theses and conference proceedings; www.trove.nla.gov.au) databases using the following topic keywords: intercrop* OR “inter-crop*” OR cocrop* OR “co-crop*” OR (mixed crop*) OR interplant* OR “inter-plant*” OR bicrop* OR “bi-crop” OR

polycultur* OR “poly-cultur*” OR dicultur* OR “di-cultur*” OR (cover crop*) or companion* AND disease* OR patho* OR nemat*. Full search query strings are provided in supplementary information, under Supporting Information Notes S1). The final search of the Web of Science database was performed on 30 June 2020. The final searches in the British Library and Trove were performed by 11 June 2020, and the ICI was last searched on 4 August 2013. We did not place time limits on any of the searches, therefore studies from the full timespan in each database were searched: 1900–2020 for Web of Science; 1800–2020 for the British Library; 2004–2013 for ICI; unspecified to 2020 for Trove. We also used backwards and forwards citation following.

To be included, experiments were required to compare (1) damage caused by plant-parasitic nematodes or disease incidence on a primary (focal) crop species grown alone as a monocrop (control) with (2) damage or incidence on the same focal crop when grown intercropped with one other plant species (treatment). Experiments had to be replicated ($n \geq 2$) with randomly assigned treatments.

We contacted the author if a relevant paper did not report the data needed. When the necessary data was reported only in a figure, GRAPHGRABBER v.1.5.5 (Dedross & Boardley, 2009) software was used to extract the values.

We found 52 studies containing data that met the relevance criteria for nematode damage, and calculated effect sizes for 326 experiments (Table S1). For soil-borne disease, we found 28 studies that met the relevance criteria, yielding 117 effect sizes (Table S1).

Effect size and variance

Metric choice A common metric was needed to compare results from studies that used different variables, constructs or descriptors to measure nematode damage or disease incidence. We used response ratio (R), defined as the mean value for the damage/disease found within the treatment (intercrop) group divided by the mean value for the control (focal monocrop) group, as the effect size. L , defined as the natural logarithm of R , was used in the meta-analysis calculations along with a nonparametric variance estimate that allowed more experiments to be included, giving greater statistical power (Mayerhofer *et al.*, 2012). L was used because, unlike R , it responds to changes in the numerator or denominator equally and in a linear fashion. The sampling distribution of L is also more normal than that of R in small samples (Hedges *et al.*, 1999). The equation for L is:

$$L = \log_e R = \log_e(\bar{x}_T/\bar{x}_C) \quad \text{Eqn 1}$$

where \bar{x}_T is the mean value for the treatment (intercrop) group and \bar{x}_C is the mean value for the control (focal monocrop) group (Hedges *et al.*, 1999).

The equation for the nonparametric variance of L is:

$$v_L = (n_T + n_C)/(n_T n_C) \quad \text{Eqn 2}$$

where n_T is the sample size for the treatment group, and n_C is the sample size for the control group (Mayerhofer *et al.*, 2012).

Crop yield effect size Using the yield of the focal crop in the monocrop control and intercrop treatment respectively, we calculated a yield effect size for each study that reported relevant yield data. Further analyses using meta-regression (please refer to ‘Meta-regressions’ in the Description section) explored whether intercropping-related changes in damage are associated with yield. A summary analysis of the focal crop yield effect size was not performed, as the experiments collected were only a nonrepresentative subset of the available experiments on yield and intercropping in the presence of nematodes.

Response ratios with zeroes The response ratio (R) could not be calculated for experiments in which the monocropping treatment had a damage or disease value of zero. To avoid introducing bias against experiments in which the focal monocrop control had no observed damage or disease but the intercrop treatment did, we used the highest value of L in the final dataset in place of the infinitely large effect size that would have been calculated otherwise in these cases. Similarly, for experiments in which the intercropping treatment had a disease or damage value of zero (meaning L could not be calculated), we used the lowest value of L to avoid bias against experiments in which intercrop treatment reduced damage or disease below detectable levels.

Experiment moderators

We collected details on experiment moderators for each experiment, including focal crop and intercrop species/density, water status, inoculum type and timing, as well as nematode species/lifestyle and/or pathogen genus (Table S1). We also described each experiment as either contained (in pots) or uncontained (in the ground, field-based) and characterised the measurement construct utilised, whether these were direct measures or generalised indices (e.g. direct measures might include root damage/infection and/or plant mortality, whereas nematode/disease indices generalised individual plant results over a field area using a defined scale).

To ensure correct grouping, we used the current accepted binomial name for all species, using ‘The Plant List’, database (www.theplantlist.org) to check plant species, and multiple sources to check nematode and pathogen species (Table S2). Table S2 also contains the references for the host status and susceptibility (of the intercrop in each experiment to the relevant nematode/pathogen) variables.

Models

Summary random-effects meta-analyses To allow generalisation of the results from this meta-analysis we performed random-effects meta-analyses on the nematode and disease datasets when calculating the overall mean effect sizes (Hedges & Vevea, 1998). In addition, as intercropping is not expected to be identical in all agroecosystems, the random-effects model is more appropriate because it does not assume that the true effect is identical in every experiment.

The estimated mean true effect sizes and estimated heterogeneity values were calculated using the *rma.mv* function in R in the METAFOR package (Viechtbauer, 2010). Each observed effect size was weighted by the inverse of its variance, with Experiment ID within Study ID included as nested random effects to include nonindependence of effect sizes from the same study and when multiple effect sizes were calculated using a shared experimental control or treatment.

Meta-regressions We used a mixed-effects model for the meta-regressions, retaining Experiment ID within Study ID as random effects but also with moderators as fixed effects, specified as a formula in the *mods* argument in the *rma.mv* function (Viechtbauer, 2010). For the heterogeneity estimator, we used maximum likelihood so that the model fit statistics could be compared during step-wise model reduction. Base models, or meta-regressions, were initially performed including moderators for which all available experiments provided data. These included nematode experiments from uncontained experiments (NU1; inclusive of 'Measurement construct + Co-crop family + Conditions + Water status + Fertilisation status'), nematode experiments in contained/potted experiments (NC1) and their disease counterparts (DU1, DC1) (Tables S3, S4). Meta-regressions that initially included a moderator for which not every experiment in the full nematode or disease dataset had a reported value were run on subsets of the data so that those moderators could be investigated (e.g. NU3, for which nematode lifestyle was included along with all NU1 moderators) (Table S4). We also carried out summary meta-analyses on each subset, to ascertain if the overall mean effect size was affected by the number of experiments included.

After running the initial model for a given subset, we removed the moderator with the highest nonsignificant *P*-value and ran the reduced model. Results of the two models were compared using the *anova* function. We reduced the model step-wise until all the moderators were significant or until reducing the model further explained a significantly lower amount of the heterogeneity in the data, according to the likelihood ratio test (Viechtbauer, 2010). If the moderator unique to that data subset dropped out, we abandoned the meta-regression as the other moderators would already have been tested on a larger data subset (e.g. NU1, NC1, DU1 or DC1). All analyses were performed in R using the METAFOR package.

Assessing bias

A 'file drawer analysis' was performed for the data(sub)set of each meta-regression that yielded interesting results (and were therefore reported and discussed), to give the number of experiments ('fail-safe number') with an average effect size of zero that would have to be added to the dataset to render the estimated mean true effect nonsignificant (Rosenthal, 1979). Funnel plots were also produced and examined for evidence of bias (Figs S1–S3). The file drawer analyses and funnel plots were performed in R using the METAFOR package.

Results

The data set used across all meta-analyses included 138 uncontained (in field) plus 188 contained (potted) nematode experiments and 89 uncontained plus 28 contained soil-borne disease experiments. These published findings included 45 focal crops (limited due to the priority focus on soil pests and pathogens), 21 intercropping families, seven nematode genera inclusive of 16 nematode species, and 10 pathogen genera, including bacterial, fungal and oomycete varieties (Table S1). Beyond crop and pathogen type, other data from field and glasshouse trials were included as moderators (e.g. fertilisation and water status of the experiment, inoculum type (artificial or natural), and timing of intercrop planting relative to the focal crop) (Table S1). The combination of these moderators allowed us to determine which associational resistance mechanisms were most consistently effective within the agroecosystem context: (1) fertiliser and water status of the experiments allowed us to test the impact arising from modification of soil nutrient and water availability, (2) planting density and duration allowed us to examine the potential of physical barriers to interfere with pest capacity to detect/recognise the focal crop root structures, (3) intercrop family investigated the induction of defences in focal crops through chemical cues released by the intercrop, and (4) analysis of pest/pathogen genera tested the provision/reduction of habitat for focal crop pests/diseases and known natural enemies.

Intercropping impacts on pest/disease damage reduction and implications for focal crop yield

Both disease incidence and pest damage were significantly reduced through intercropping. Soil-borne disease damage to the focal crop within in-field studies was reduced by 55% due to intercropping (with a 95% confidence interval (CI) of 67 to 38%) and 44% (95% CI of 57 to 28%) in contained experiments (Table S3). Nematode damage to the focal crop was reduced by 40% (95% CI of 55 to 20%) in uncontained, field-based studies and 42% (95% CI of 54 to 28%) in contained, pot-bound experiments (Table S3).

Nineteen studies that reported nematode damage data also contained yield information, for 76 experiments in total (and 11 studies, 66 experiments for disease; Table S1). The data published in these studies can be used to estimate how much damage reduction in intercropped fields is required to deliver equivalent focal crop yields to monocrops. Based on damage effect to focal crop yield regressions we estimate that anything more effective than a +15% enhancement in nematode damage (Fig. 1) results in improved focal crop yields from intercropped fields, compared with those obtained from monocrop planting. Whereas disease reduction is apparent in published studies, disease effect size was not significantly correlated to yield size and therefore estimates of reduced focal crop yield, necessary to obtain improved focal crop yield, were not possible.

Identifying field and method variables that affect intercrop impact

The use of meta-regressions allowed us to identify those experimental and field variables that were most influential. Within

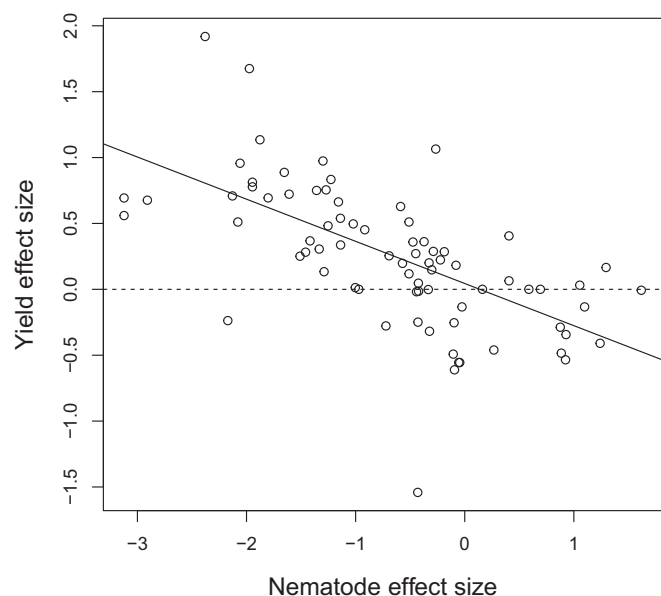


Fig. 1 Plot of yield against nematode damage (both log-response ratios where ± 1.0 equals a deviation of 100%). The solid line is the regression line that corresponds to $(\log_e(\text{Yield response ratio}) = -0.320 \times \log_e(\text{Damage response ratio}) + 0.044$. Slope confidence intervals (95%) are -0.476 to -0.164 ; while 95% CI for the intercept are -0.150 to 0.238 . The dashed line (100% yield) marks where focal crop yield was the same in both monocrop and intercrop treatments. The solid regression line passes through 100% yield at a damage response ratio of 1.15. Assuming that the regression line is accurate, this suggests improving or maintaining focal crop yields in intercrops is possible even with an increase of up to 15% in focal crop damage under intercropping.

field-based experiments the variables that were most influential in intercrop control of nematodes included intercrop family and field fertilisation status. Within the intercrop families most commonly reported, Asteraceae, Pedaliaceae and Solanaceae were found to be the most significantly effective in their capacity to moderate nematode damage (Fig. 2). Unfertilised fields showed significantly better outcomes (less nematode damage) compared with fertilised fields (Fig. 2).

Within contained studies, nematode species was an important variable in determining intercropping impact, with *P. neglectus* and *C. xenoplax* reported as the most impacted (reduced focal crop damage) and *M. incognita* and *M. javanica* as the least affected species of those commonly studied (Fig. 3). The method by which damage, or impact, was assessed is also a significant variable in contained studies. Those studies that assessed roots, either via directly measuring root damage or assessing damage against a root damage index (RDI) showed substantially greater impacts from intercropping, while those that explored nonroot specific outcomes (e.g. nematode soil population or plant mortality) reported less impact. (Fig. 3).

Disease control variables that directly and significantly influenced intercrop outcomes within in-field, or uncontained, studies included intercrop family and focal crop density in the intercrop treatment relative to that in the monocrop control. Pathogen type was not a significant variable within the meta-analysis, suggesting that pathogen type was much less important to intercropping yield outcomes compared with intercrop and focal crop density. Of the most studied intercrop families,

Co-crop family/Fertiliser

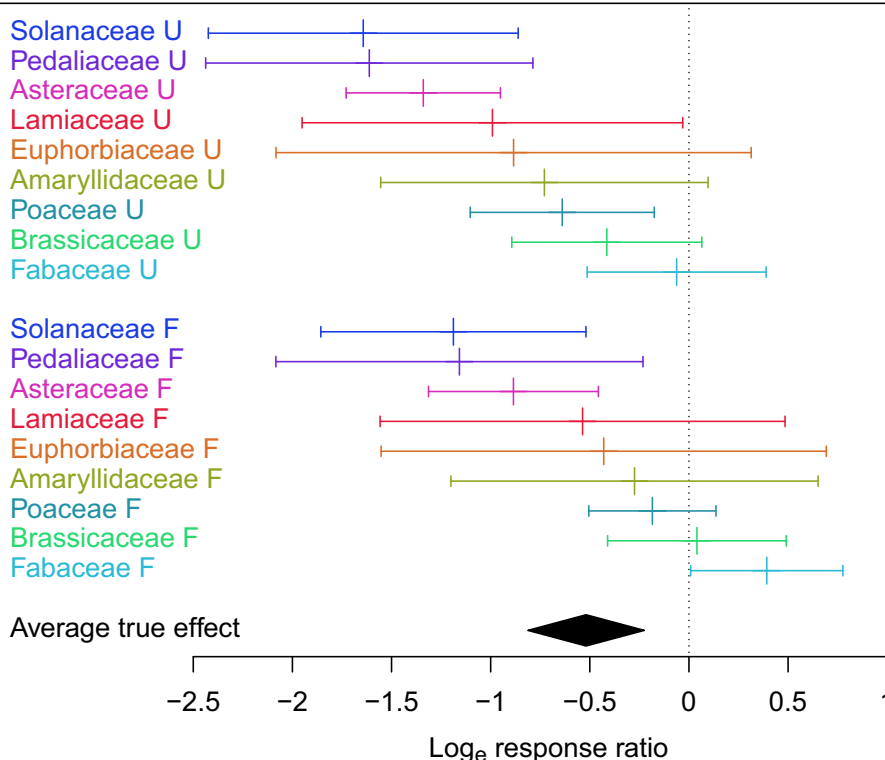


Fig. 2 Predicted effects of intercropping on nematode damage with combinations of the moderators retained in the final model for the NU1 dataset, showing selected families (F, fertilised; U, unfertilised). Values below 1 show reduced nematode damage in intercrop studies. Error bars represent 95% confidence intervals.

Amaryllidaceae was more effective than either Poaceae or Fabaceae (Fig. 4). Methodologically, only the measurement construct was found to be significant in influencing perceived focal crop outcomes, with disease incidence indicative of greater impact relative to studies that utilised a disease index (Fig. 4). Contrary to that observed with nematode intercrop effects, neither disease type nor field fertilisation significantly influenced intercrop reduction of disease damage to focal crop.

The overall summary response ratio estimates for the full uncontained (NU1) and contained (NC1) nematode datasets were quite close, 0.60 and 0.55, respectively (Table S3), suggesting that experiments in pots may slightly overestimate the effectiveness of intercropping against nematodes. For the full uncontained (DU1) and contained (DC1) disease datasets, however, the overall summary true effect response ratio estimates were 0.45 and 0.56, respectively (Table S3), indicating that experiments in pots substantively underestimated the effectiveness of intercropping against soil-borne diseases.

Impacts of publication bias on model outcomes

Publication bias towards positive outcomes does not appear to have had a significant effect on our results: the fail-safe numbers for the NU1, NC2, and DU1 datasets were 2250, 9773, and 3513, respectively, and no bias in the datasets was detected in the funnel plots (Figs S1–S3).

Discussion

We found strong quantitative evidence in support of intercropping as an effective method of control for both plant-parasitic nematodes and soil-borne diseases.

Nematode impacts were reduced by 40% in-field experiments, and 45% in pot studies (Table S3). The meta-regression analysing the effects of intercropping on focal crop yield showed that anything below a 15% increase in nematode impacts was associated with an equal or greater yield in the intercropped fields (Fig. 1). This counterintuitive result suggests that, in intercropping systems, moderate increases in nematode damage are disconnected from yield outcomes. This indicates that other mechanisms arising from associational resistance may benefit yield, not solely pest reduction. However, the calculated regression line has substantial confidence intervals for both slope and intercept (Fig. 1) creating possible outcomes that range from (1) effectively no impact on yield outcomes in intercrop systems, (2) slightly reduced focal crop outcomes where intercrops are used, or (3) an even greater disconnect between nematode damage and focal crop yield in intercrop systems. Disease outcomes were reduced significantly through intercropping, by 55% across uncontained field studies but only 44% when experiments in pots were analysed (Table S3).

Drivers of variability in outcomes

Variability in the effectiveness of intercropping as a control strategy is driven by substantially different factors for nematodes

compared with disease. Both nematode and pathogenic microbial survival are determined by soil texture, moisture, temperature and pH (Menzies, 1963; Wallace, 1966). On species-specific bases both are also affected by plant released volatiles (van Agtmaal *et al.*, 2018; de Boer *et al.*, 2019; Sikder & Vestergård, 2020). It seems likely that the differences in field outcomes between the two originate from the capacity of nematodes to orient and move towards host species based on volatile cues (Turnbull *et al.*, 2001; Turlings *et al.*, 2012; Yang & van Elsas, 2018). Meta-regression of published outcomes reveals that effective nematode control is determined primarily by nematode species in contained studies, or by intercrop family and soil nutrient status (whether the experiment incorporated fertilisation) in uncontained experiments. By contrast, the only variables that significantly influenced disease outcomes (and only within uncontained studies) included intercrop family and density of focal crop planting in the intercrop relative to that in the focal monocrop.

Intercrop family Intercrop family explained a significant fraction of the efficacy of intercropping against nematodes in uncontained field studies. For example, interplanting with Asteraceae, Pedaliaceae or Solanaceae species was associated with better nematode control, whereas Poaceae, Brassicaceae and Fabaceae intercrops were associated with less effective control. Intercrop root exudates can lead to associational resistance to nematodes through several mechanisms: (1) acting as biocidal compounds, so killing pests, or reducing their fitness; (2) acting as an attractant, so trapping pests away from the focal crop; (3) acting as a repellent, reducing pest population near the focal crop; or (4) modifying hatching rates, creating phenological mismatch between crops and early-stage feeding (Sikder & Vestergård, 2020). Biocidal effects can include the housing of organisms such as nematode-detering endoroot bacteria, or, as for the Brassicaceae, the production of biofumigant compounds such as glucosinolates, which are metabolic precursors to bioactive isothiocyanates (Brennan *et al.*, 2020). It is interesting to note the limited efficacy of the Brassicaceae in this meta-analysis, despite their production of glucosinolates and their frequent use as biofumigants. Nematodes have been shown to be attracted to plant root volatiles (Rasmann *et al.*, 2005; Turlings *et al.*, 2012), and trap crops, including Asteraceae, have been shown to be effective in this context through a combination of lack of susceptibility and reduction of nematode hatch (Tsay *et al.*, 2004).

Examples in which exudates can operate by combinations of these mechanisms are common, making these intercrops particularly effective. For example, lauric acid, exuded from *Chrysanthemum coronarium* roots, has been shown to be both attractive and lethal to *M. incognita* second-stage juveniles, reducing galling on intercropped tomato plants (Dong *et al.*, 2014). Some root exudates may accumulate, or persist at residual concentrations, after intercropping. Microbial parasites or allelochemicals may still be effective in reducing nematode numbers or effect, even at lower concentrations and could provide protection after intercrop removal (Mohan *et al.*, 2020; Sikder & Vestergård, 2020).

Many intercrops produce nematocidal root exudates but, even when these compounds are not actively exuded from roots,

Nematode sp./Construct

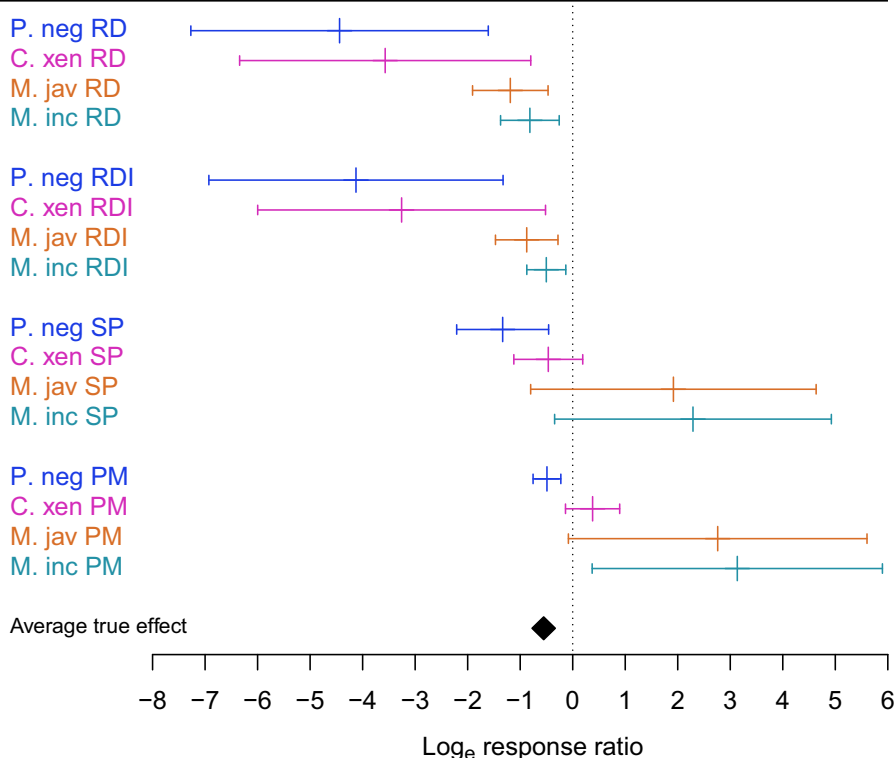


Fig. 3 Predicted effects of intercropping on nematode damage with combinations of the moderators retained in the final model for the NC2 dataset, showing moderator categories most represented in the data. Measurement constructs and nematode species: RD, root damage; RDI, root damage index; SP, soil population; PM, plant mortality; P. neg, *Pratylenchus neglectus*; C. xen, *Crictonemoides xenoplax*; M. jav, *Meloidogyne javanica*; M. inc, *M. incognita*. Error bars represent 95% confidence intervals.

nematodes may still be exposed to these allelochemicals when feeding upon, or penetrating, intercrop roots or encountering allelochemicals in the soil after root degradation. Examples include root tissues and exudates from Asteraceae family species that have been shown to contain compounds with nematocidal properties (Sánchez De Viala *et al.*, 1998; Mohny *et al.*, 2009; Weidenhamer *et al.*, 2009), at levels sufficient to reduce nematode damage and reproduction (Salem & Osman, 1988) and are lethal and inhibit egg hatching (Siddiqui & Alam, 1987; Tsay *et al.*, 2004).

Many plants produce root exudates that are not toxic but act as sensory stimulants, attracting or repulsing nematodes. For example, Solanaceae treatments were more effective than nearly all other intercrop families in nematode control (Fig. 2). Solanaceae may be particularly effective against nematodes because they emit methyl salicylate (MSA) (Murungi *et al.*, 2018). MSA has been shown to be an effective attractant for *M. incognita* (Wuyts *et al.*, 2006) and also acts a phytohormone, released during herbivory (Lin *et al.*, 2017). Furthermore, exposure to MSA can confer resistance to agricultural pests (Bar-Nun & Mayer, 2008) and its metabolism regulates plant defence signalling and systemic acquired resistance in exposed plants (Chen *et al.*, 2019). Solanaceae therefore appears to confer multiple beneficial outcomes that build upon MSA attraction to nematodes, signalling other plants to prime defences and nematocidal action. Another example of a multi-impact signalling effect can be found in intercropping with brassicaceous species, currently used as trap crops in rotations with sugar beet. *Brassica* spp. are beneficial intercrops, as their roots are attractive to, and can be

invaded by, sugar beet cyst nematodes, but the sex ratio of the subsequent generation is heavily skewed towards males, leading to population decline (Caubel & Chaubet, 1985; Lelivelt & Hoogendoorn, 1993; Ratnadass *et al.*, 2012).

Impacts of changing focal crop planting density Whereas intercrop family was an important factor in determining disease reduction in uncontained studies, effects from interplanting on soil-borne disease were greater when the density of focal crop plants in the intercrop treatment were kept closer to densities in focal monocrop control. This may be evidence of the ‘dilution effect’, whereby disease reduction is achieved by decreasing the frequency of hosts rather than absolute density (Civitello *et al.*, 2015). Interplanting maize at a spacing of 5 cm decreased red crown rot in soybean more than a spacing of 20 cm (Gao *et al.*, 2014). They suggested that greater density of maize roots reduced root-to-root transmission of the pathogen, in this case due to higher planting density rather than enhanced root growth.

Dispersal of pathogens across the crop system is a key determinant of disease incidence and spread (Tack *et al.*, 2014). Therefore, interception of propagules travelling between roots could be an effective intercropping mechanism, leading to a lower incidence of disease and a process shown to be effective against aerial pathogens (Bouws & Finckh, 2008; Fernández-Aparicio *et al.*, 2011). Size and location of intercrop root systems are important in reducing disease incidence. For example, bacterial wilt was only reduced when cowpea was planted between tomato within rows, not when it was planted between the rows of tomato, or

Relative density/Construct/Co-crop family

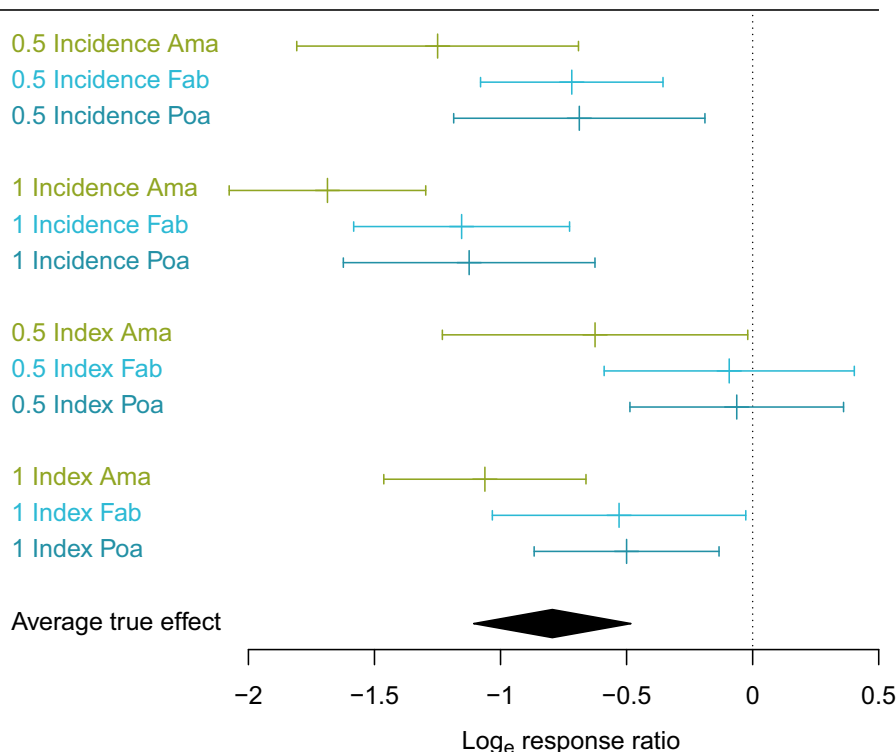


Fig. 4 Predicted effects of intercropping on disease damage with combinations of the moderators retained in the final model for the DU1 dataset, showing relative density set to 0.5 or 1 and the co-crop families most represented in the data (Ama, Amaryllidaceae; Fab, Fabaceae; Poa, Poaceae). Error bars represent 95% confidence intervals.

when Welsh onion (which had smaller root systems) was interplanted with tomato (Michel *et al.*, 1997).

Greater plant density also causes shading (reducing soil temperatures) and greater evapotranspiration (reducing soil moisture), producing less favourable conditions for infection and disease development (Robinson *et al.*, 1987; Olasantan, 1988; Yadav & Lalramliana, 2012). Retention of focal crop density could make intercropping more attractive to growers, as yield per unit area could be maintained while also reducing disease incidence.

Susceptibility of nematode species Intercropping treatment impact in contained experiments was significantly influenced by the species of nematode involved. Contained studies are likely to restrain nematode mobility, enforcing the interaction with non-volatile root exudates that may not travel far from the root or diffuse volatiles concentrated within contained soils (Erb *et al.*, 2013). Studies on nematode susceptibility to root metabolites show substantial variation in their responses, only partially attributable to nematode lifestyle. Such species-dependent resistance to repelling and/or toxic compounds explains the key role of nematode species identity in outcomes (Sikder & Vestergård, 2020).

Perhaps surprisingly, neither host status nor susceptibility of intercrop species to nematode attack was retained as significant in any of the meta-regressions in which they were included, suggesting that disrupting nematode movement by adding a nonhost as intercrop to reduce nematode damage is ineffective. However, information on host status/susceptibility was unavailable for nearly half the experiments included in the full nematode dataset,

limiting the scope of this result. The unimportance of intercrop species' host status or susceptibility level is positive from an agronomic point of view, as finding suitable nonhost species for intercropping might be challenging, especially for pests with wide host ranges such as the root knot nematode, *Meloidogyne incognita*. Host status/susceptibility may be important in intercrop impacts on soil-borne disease transmission, but we were unable to assess this.

Field fertilisation Soil and plant nutrient status are key drivers of ecological interactions between plants and their natural enemies (Holopainen *et al.*, 1995). Associational susceptibility is often perceived to be driven by competition for nutrients between plants and reduced resource in focal crop plants. Here we found, perhaps surprisingly, that increasing resource availability through fertilisation reduced nematode control efficacy of intercropping in field experiments. There are several potential explanations for this surprising outcome. Plants can respond to nutrient (particularly N/P) stress by increasing root length/branching and releasing root exudates to access/forage for new soil nutrient pools (Khan *et al.*, 2016). This increased root volume occupies a greater soil volume, leading to increased interception of pests by intercrop roots and/or enhanced exposure to allelopathic root exudates or root-altered soil conditions. As with other types of plant defence (De Long *et al.*, 2016), nutrient levels affect composition and quantity of root exudates. Nutrient-rich plants have been shown to release fewer defensive volatiles (Fernandez-Martinez *et al.*, 2018). When maize was grown under

phosphorus-limiting conditions, their roots produced more cinnamic acid (with strong activity against the soybean red crown rot pathogen *Cylindrocladium parasiticum*) and more salicylic acid (inducing resistance against pathogens) (Gutjahr & Paszkowski, 2009; Gao *et al.*, 2014). Plant nutrient status also creates indirect effects on pests: enhanced competition under reduced nutrients with a second plant species can cause morphological or physiological changes in a focal crop, such that it becomes a less suitable host (Trenbath, 1993).

Conclusion

Intercropping can be an effective tool for reducing nematode damage and disease in agricultural fields, with averaged reductions in impacts on the performance of the focal crop of 40% and 55% respectively. Our study also showed that treatments in fields with nematode pests will improve when using reduced field fertilisation approaches, whereas diseased field outcomes will be improved with greater planting densities. Generally, across all intercropping systems the intercrop has a significant impact on outcomes, and further characterisation of specific modes of associational resistance derived for each intercrop/pest pairing is needed to provide confidence in focal crop yield outcomes.

Future research should avoid experiments with potted plants. Our analysis demonstrates that outcomes from contained experiments are not indicative of *in situ* outcomes or mechanisms of action. Similarly, although our results point to effective pest and disease control, it remains unclear whether even greater nematode control can be achieved through reduced, intermediate nutrient content in agricultural fields. Further in-field research into intermediate fertilisation status is needed to clarify this question. Similarly, quantifying maximally effective planting densities will require substantially more research to understand whether these can be generalised across a range of agroecosystems, or whether they are crop/pathogen specific.

Overall, our study demonstrates the potential of associational resistance, via intercropping and other mechanisms, to reduce the impact of soil pests and disease on crops. It provides support to farming practices based on ecological approaches to deliver more sustainable productive agricultural systems.



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Author contributions

VGAC performed the literature search and the meta-analyses. VGAC wrote the first draft of the manuscript, with equal editorial input from KRR and SEH after the first draft.

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Data availability

Data sharing is not applicable to this article as no new data were created or analysed in this study. All materials obtained through systematic review and which supports the meta-analysis findings of this study are described in the [Supplementary Material](#) of this article.

References

- Abdel-Monaim MF, Abo-Elyousr KAM. 2012. Effect of preceding and intercropping crops on suppression of lentil damping-off and root rot disease in New Valley – Egypt. *Crop Protection* 32: 41–46.
- Agrawal AA, Lau JA, Hamback PA. 2006. Community heterogeneity and the evolution of interactions between plants and insect herbivores. *The Quarterly Review of Biology* 81: 349–376.
- van Agtmaal H, Straathof AL, Termorshuizen A, Lievens B, Hoffland E, de Boer W. 2018. Volatile-mediated suppression of plant pathogens is related to soil properties and microbial community composition. *Soil Biology and Biochemistry* 117: 164–174.
- Barbosa P, Hines J, Kaplan I, Martinson H, Szczepaniec A, Szendrei Z. 2009. Associational resistance and associational susceptibility: having right or wrong neighbors. *Annual Review of Ecology, Evolution, and Systematics* 40: 1–20.
- Bar-Nun M, Mayer AM. 2008. Methyl jasmonate and methyl salicylate, but not cis-jasmone, evoke defenses against infection of *Arabidopsis thaliana* by *Orobanche aegyptiaca*. *Weed Biology and Management* 8: 91–96.
- Berkey CS, Hoaglin DC, Mosteller F, Colditz GA. 1995. A random-effects regression model for meta-analysis. *Statistics in Medicine* 14: 395–411.
- de Boer W, Li X, Meisner A, Garbeva P. 2019. Pathogen suppression by microbial volatile organic compounds in soils. *FEMS Microbiology Ecology* 95: fiz105.
- Bommarco R, Kleijn D, Potts SG. 2013. Ecological intensification: harnessing ecosystem services for food security. *Trends in Ecology & Evolution* 28: 230–238.
- Boudreau MA. 2013. Diseases in intercropping systems. *Annual Review of Phytopathology* 51: 499–519.
- Bouws H, Finckh MR. 2008. Effects of strip intercropping of potatoes with non-hosts on late blight severity and tuber yield in organic production. *Plant Pathology* 57: 916–927.
- Brennan RJB, Glaze-Corcoran S, Wick R, Hashemi M. 2020. Biofumigation: an alternative strategy for the control of plant parasitic nematodes. *Journal of Integrative Agriculture* 19: 1680–1690.
- Caubel G, Chaubet B. 1985. Hatching and multiplication of beet cyst nematode *Heterodera schachtii* Schmidt on oilseed rape and forage radish. *Agronomie* 5: 463–466.
- Chen L, Wang WS, Wang T, Meng XF, Chen TT, Huang XX, Li YJ, Hou BK. 2019. Methyl salicylate glucosylation regulates plant defense signaling and systemic acquired resistance. *Plant Physiology* 180: 2167–2181.
- Civittello DJ, Cohen J, Fatima H, Halstead NT, Liriano J, McMahon TA, Ortega CN, Sauer EL, Sehgal T, Young S *et al.* 2015. Biodiversity inhibits parasites: broad evidence for the dilution effect. *Proceedings of the National Academy of Sciences, USA* 112: 8667–8671.
- De Long JR, Sundqvist MK, Gundale MJ, Giesler R, Wardle DA. 2016. Effects of elevation and nitrogen and phosphorus fertilization on plant defence compounds in subarctic tundra heath vegetation. *Functional Ecology* 30: 314–325.
- Dedross P, Boardley A. 2009. *GRAPHGRABBER, v. 1.5.5*. Henley-on-Thames, UK: Quintessa.
- Denno RF, Gruner DS, Kaplan I. 2008. Potential for entomopathogenic nematodes in biological control: a meta-analytical synthesis and insights from trophic cascade theory. *Journal of Nematology* 40: 61–72.

- Deutsch CA, Tewksbury JJ, Tigchelaar M, Battisti DS, Merrill SC, Huey RB, Naylor RL. 2018. Increase in crop losses to insect pests in a warming climate. *Science* 361: 916–919.
- Doheny-Adams T, Ellis S, Wade R, Lilley CJ, Barker A, Atkinson HJ, Urwin PE, Redeker KR, Hartley SE. 2018. Constant isothiocyanate-release potentials across biofumigant seeding rates. *Journal of Agricultural and Food Chemistry* 66: 5108–5116.
- Dong L, Huang C, Huang L, Li X, Zuo Y. 2012. Screening plants resistant against *Meloidogyne incognita* and integrated management of plant resources for nematode control. *Crop Protection* 33: 34–39.
- Dong L, Li X, Huang L, Gao Y, Zhong L, Zheng Y, Zuo Y. 2014. Lauric acid in crown daisy root exudate potentially regulates root-knot nematode chemotaxis and disrupts Mi-flp-18 expression to block infection. *Journal of Experimental Botany* 65: 131–141.
- Erb M, Huber M, Robert CAM, Ferrieri AP, Machado RAR, Arce CCM. 2013. The role of plant primary and secondary metabolites in root-herbivore behaviour, nutrition and physiology. In: Johnson SN, Hiltbold I, Turlings TCJ, eds. *Advances in insect physiology: behaviour and physiology of root herbivores*. Amsterdam, the Netherlands: Elsevier, 53–95.
- Evans K, Stone AR. 1977. A review of the distribution and biology of the potato cyst-nematodes *Globodera rostochiensis* and *G. pallida*. *PANS* 23: 178–189.
- Fernández-Aparicio M, Emeran AA-M, Rubiales D. 2011. Inter-cropping faba bean with berseem, fenugreek or oat can contribute to broomrape management. *Grain Legumes* 56: 31.
- Fernandez-Martinez M, Lluisa J, Filella I, Niinemets U, Arneth A, Wright IJ, Loreto F, Penuelas J. 2018. Nutrient-rich plants emit a less intense blend of volatile isoprenoids. *New Phytologist* 220: 773–784.
- Gao X, Wu M, Xu R, Wang X, Pan R, Kim H-JJ, Liao H. 2014. Root interactions in a maize/soybean intercropping system control soybean soil-borne disease, red crown rot. *PLoS ONE* 9: e95031.
- Gutjahr C, Paszkowski U. 2009. Weights in the balance: jasmonic acid and salicylic acid signaling in root-biotroph interactions. *Molecular Plant–Microbe Interactions* 22: 763–772.
- Hartley SE. 2018. Agroecological approaches to sustainable intensification. In: Campanhola C, Pandey S, eds. *Sustainable food and agriculture: an integrated approach*. Rome, Italy: Academic Press, 179–184.
- Hedges LV, Gurevitch J, Curtis PS. 1999. The meta-analysis of response ratios in experimental ecology. *Ecology* 80: 1150–1156.
- Hedges LV, Vevea JL. 1998. Fixed- and random-effects models in meta-analysis. *Psychological Methods* 3: 486–504.
- Holopainen JK, Rikala R, Kainulainen P, Oksanen J. 1995. Resource partitioning to growth, storage and defence in nitrogen-fertilized Scots pine and susceptibility of the seedlings to the tarnished plant bug *Lygus rugulipennis*. *New Phytologist* 131: 521–532.
- Johnson SN, Clark KE, Hartley SE, Jones TH, McKenzie SW, Koricheva J. 2012. Aboveground-belowground herbivore interactions: a meta-analysis. *Ecology* 93: 2208–2215.
- Johnson SN, Erb M, Hartley SE. 2016. Roots under attack: contrasting plant responses to below- and aboveground insect herbivory. *New Phytologist* 210: 413–418.
- Khan MA, Gemenet DC, Villordon A. 2016. Root system architecture and abiotic stress tolerance: current knowledge in root and tuber crops. *Frontiers in Plant Science* 7: 1584.
- Lelivelt CLC, Hoogendoorn J. 1993. The development of juveniles of *Heterodera schachtii* in roots of resistant and susceptible genotypes of *Sinapis alba*, *Brassica napus*, *Raphanus sativus* and hybrids. *Netherlands Journal of Plant Pathology* 99: 13–22.
- Letourneau DK, Armbricht I, Rivera BS, Lerma JM, Carmona EJ, Daza MC, Escobar S, Galindo V, Gutiérrez C, López SD *et al.* 2011. Does plant diversity benefit agroecosystems? A synthetic review. *Ecological Applications* 21: 9–21.
- Lin Y, Qasim M, Hussain M, Akutse KS, Avery PB, Dash CK, Wang L. 2017. The herbivore-induced plant volatiles methyl salicylate and menthol positively affect growth and pathogenicity of entomopathogenic fungi. *Scientific Reports* 7: 40494.
- Malézieux E, Crozat Y, Dupraz C, Laurans M, Makowski D, Ozier-Lafontaine H, Rapidel B, de Tournonet S, Valantin-Morison M. 2009. Mixing plant species in cropping systems: concepts, tools and models: a review. *Agronomy for Sustainable Development* 29: 43–62.
- Marimuthu S, Ramamoorthy V, Samiyappan R, Subbian P. 2013. Intercropping system with combined application of *Azospirillum* and *Pseudomonas fluorescens* reduces root rot incidence caused by *Rhizoctonia bataticola* and increases seed cotton yield. *Journal of Phytopathology* 161: 405–411.
- Matthiessen JN, Kirkegaard JA. 2006. Biofumigation and enhanced biodegradation: opportunity and challenge in soilborne pest and disease management. *Critical Reviews in Plant Sciences* 25: 235–265.
- Mayerhofer MS, Kernaghan G, Harper KA. 2012. The effects of fungal root endophytes on plant growth: a meta-analysis. *Mycorrhiza* 23: 119–128.
- Menzies JD. 1963. Microbial plant pathogens in soil. *The Botanical Review* 29: 79–122.
- Michel VV, Wang J-F, Midmore DJ, Hartman GL. 1997. Effects of intercropping and soil amendment with urea and calcium oxide on the incidence of bacterial wilt of tomato and survival of soil-borne *Pseudomonas solanacearum* in Taiwan. *Plant Pathology* 46: 600–610.
- Mohan S, Kumar KK, Sutar V, Saha S, Rowe J, Davies KG. 2020. Plant root-exudates recruit hyperparasitic bacteria of phytonematodes by altered cuticle aging: implications for biological control strategies. *Frontiers in Plant Science* 11: 763.
- Mohney BK, Matz T, Lamoreaux J, Wilcox DS, Gimsing AL, Mayer P, Weidenhamer JD. 2009. In situ silicone tube microextraction: a new method for undisturbed sampling of root-exuded thiophenes from marigold (*Tagetes erecta* L.) in soil. *Journal of Chemical Ecology* 35: 1279–1287.
- Munnecke DE. 1972. Factors affecting the efficacy of fungicides in soil. *Annual Review of Phytopathology* 10: 375–398.
- Murungi LK, Kirwa H, Coyne D, Teal PEA, Beck JJ, Torto B. 2018. Identification of key root volatiles signaling preference of tomato over spinach by the root knot nematode *Meloidogyne incognita*. *Journal of Agricultural and Food Chemistry* 66: 7328–7336.
- Oerke E-C. 2006. Crop losses to pests. *The Journal of Agricultural Science* 144: 31–43.
- Oerke E-C, Dehne H-W. 2004. Safeguarding production—losses in major crops and the role of crop protection. *Crop Protection* 23: 275–285.
- Olasantan FO. 1988. The effects on soil temperature and moisture content and crop growth and yield of intercropping maize with melon (*Colocynthis vulgaris*). *Experimental Agriculture* 24: 67–74.
- Popp J, Pető K, Nagy J. 2013. Pesticide productivity and food security. A review. *Agronomy for Sustainable Development* 33: 243–255.
- Pretty J, Benton TG, Bharucha ZP, Dicks LV, Flora CB, Godfray HCJ, Goulson D, Hartley S, Lampkin N, Morris C *et al.* 2018. Global assessment of agricultural system redesign for sustainable intensification. *Nature Sustainability* 1: 441–446.
- Rasmann S, Köllner TG, Degenhardt J, Hiltbold I, Töpfer S, Kuhlmann U, Gershenson J, Turlings TCJ. 2005. Recruitment of entomopathogenic nematodes by insect-damaged maize roots. *Nature* 434: 732–737.
- Ratnadass A, Fernandes P, Avelino J, Habib R. 2012. Plant species diversity for sustainable management of crop pests and diseases in agroecosystems: a review. *Agronomy for Sustainable Development* 32: 273–303.
- Reiss ER, Drinkwater LE. 2018. Cultivar mixtures: a meta-analysis of the effect of intraspecific diversity on crop yield. *Ecological Applications* 28: 62–77.
- Robinson MP, Atkinson HJ, Perry RN. 1987. The influence of soil moisture and storage time on the motility, infectivity and lipid utilization of second stage juveniles of the potato cyst nematodes *Globodera rostochiensis* and *G. pallida*. *Revue de Nématologie* 10(3): 343–348.
- Rosenthal R. 1979. The file drawer problem and tolerance for null results. *Psychological Bulletin* 86: 638–641.
- Royal Society. 2009. *Reaping the benefits: science and the sustainable intensification of global agriculture*. London, UK: Royal Society.
- Salem FM, Osman GY. 1988. Effectiveness of tagetes natural exudates on *Meloidogyne javanica* (Chitwood) nematode. *Anzeiger für Schädlingskd. Pflanzenschutz Umweltschutz* 61: 17–19.
- Sánchez De Viala S, Brodie BB, Rodríguez E, Gibson DM. 1998. The potential of thiurubrine C as a nematocidal agent against plant-parasitic nematodes. *Journal of Nematology* 30: 192–200.

- Shigaki T, Gray FA, Delaney RH, Koch DW. 1998. Evaluation of host resistance and intercropping for management of the northern root-knot nematode in sainfoin, *Onobrychis viciifolia*. *Journal of Sustainable Agriculture* 12: 23–39.
- Sikder MM, Vestergård M. 2020. Impacts of root metabolites on soil nematodes. *Frontiers in Plant Science* 10: 1792.
- Tack AJM, Hakala J, Petaja T, Kulmala M, Laine A-L. 2014. Genotype and spatial structure shape pathogen dispersal and disease dynamics at small spatial scales. *Ecology* 95(2014): 703–714.
- Tahvanainen JO, Root RB. 1972. The influence of vegetational diversity on the population ecology of a specialized herbivore, *Phyllotreta cruciferae* (Coleoptera: Chrysomelidae). *Oecologia* 10: 321–346.
- The Plant List. 2010. Version 1. [WWW document] URL <http://www.theplantlist.org/> [accessed 31 December 2012].
- Tilman D, Balzer C, Hill J, Befort BL. 2011. Global food demand and the sustainable intensification of agriculture. *Proceedings of the National Academy of Sciences, USA* 108: 20260–20264.
- Trenbath BR. 1993. Intercropping for the management of pests and diseases. *Field Crops Research* 34: 381–405.
- Tsay TT, Wu ST, Lin YY. 2004. Evaluation of Asteraceae plants for control of *Meloidogyne incognita*. *Journal of Nematology* 36: 36–41.
- Turlings TCJ, Hiltbold I, Rasmann S. 2012. The importance of root-produced volatiles as foraging cues for entomopathogenic nematodes. *Plant and Soil* 358: 51–60.
- Turnbull GA, Morgan JAW, Whipps JM, Saunders JR. 2001. The role of bacterial motility in the survival and spread of *Pseudomonas fluorescens* in soil and in the attachment and colonisation of wheat roots. *FEMS Microbiology Ecology* 36: 21–31.
- Viechtbauer W. 2010. Conducting meta-analyses in R with the METAFOR package. *Journal of Statistical Software* 36: 1–48.
- Wallace HR. 1966. Factors influencing the infectivity of plant parasitic nematodes. *Proceedings of the Royal Society B: Biological Sciences* 164: 592–614.
- Weidenhamer JD, Boes PD, Wilcox DS. 2009. Solid-phase root zone extraction (SPRE): a new methodology for measurement of allelochemical dynamics in soil. *Plant and Soil* 322: 177–186.
- Wuyts N, Swennen R, De Waele D. 2006. Effects of plant phenylpropanoid pathway products and selected terpenoids and alkaloids on the behaviour of the plant-parasitic nematodes *Radopholus similis*, *Pratylenchus penetrans* and *Meloidogyne incognita*. *Nematology* 8: 89–101.
- Yadav AK, Lalramliana. 2012. Soil moisture effects on the activity of three entomopathogenic nematodes (Steinernematidae and Heterorhabditidae) isolated from Meghalaya, India. *Journal of Parasitic Diseases* 36: 94–98.
- Yang P, van Elsas JD. 2018. Mechanisms and ecological implications of the movement of bacteria in soil. *Applied Soil Ecology* 129: 112–120.
- rusticana*) roots against *Meloidogyne incognita*. *Journal of Agricultural and Food Chemistry* 61: 4723–4727.
- Alam MM, Saxena SK, Khan AM. 1977. Influence of interculture of marigold and murgosa with some vegetable crops on plant growth and nematode population. *Acta Botanica Indica* 5: 33–39.
- Ali SS, Kumar R, Naimuddin, Sing B. 2008. Management of plant parasitic nematodes infesting chickpea through intercropping with non-leguminous crops. *Trends in Bioscience* 1: 8–12.
- Ameen HH, Hasabo SA. 1995. Effect of intercropping *Asparagus scandens* with sour orange seedling in comparison with nematicidal and root exudate treatments on *Tylenchulus semipenetrans* larvae. *Anzeiger für Schädlingskd. Pflanzenschutz Umweltschutz* 68: 129–130.
- Arim OJ, Waceke JW, Waudu SW, Kimenju JW. 2006. Effects of *Canavalia ensiformis* and *Mucuna pruriens* intercrops on *Pratylenchus zeae* damage and yield of maize in subsistence agriculture. *Plant and Soil* 284: 243–251.
- Autrique A, Potts MJ. 1987. The influence of mixed cropping on the control of potato bacterial wilt (*Pseudomonas solanacearum*). *The Annals of Applied Biology* 111: 125–133.
- Barto EK, Cipollini D. 2009. Half-lives and field soil concentrations of *Alliaria petiolata* secondary metabolites. *Chemosphere* 76: 71–75.
- Bateman GL, Hornby D. 1999. Comparison of natural and artificial epidemics of take-all in sequences of winter wheat crops. *The Annals of Applied Biology* 135: 555–571.
- Bhan M, McSorley R, Chase CA. 2010. Effect of cropping system complexity on plant-parasitic nematodes associated with organically grown vegetables in Florida. *Nematropica* 40: 53–70.
- Bongers AMT. 2014. *Tylenchorhynchus brassicae* Siddiqi, 1961. *Fauna Europaea*. [WWW document] URL <https://fauna-eu.org> [accessed 14 November 2014].
- Butler J, Garratt MPD, Leather SR. 2012. Fertilisers and insect herbivores: a meta-analysis. *The Annals of Applied Biology* 161: 223–233.
- Chiamolera F, Dias-Arieira C, Souto E. 2012. Susceptibility of winter crops to *Pratylenchus brachyurus* and effect on the nematode population in the maize crop. *Nematropica* 42: 267–275.
- Curto G, Dallavalle E, Lazzeri L. 2005. Life cycle duration of *Meloidogyne incognita* and host status of Brassicaceae and Capparaceae selected for glucosinolate content. *Nematology* 7: 203–212.
- Desaeger J, Rao MR. 2001. The potential of mixed covers of *Sesbania*, *Tephrosia* and *Crotalaria* to minimise nematode problems on subsequent crops. *Field Crops Research* 70: 111–125.
- Dong L, Huang C, Huang L, Li X, Zuo Y. 2012. Screening plants resistant against *Meloidogyne incognita* and integrated management of plant resources for nematode control. *Crop Protection* 33: 34–39.
- Egunjobi OA. 1984. Effects of intercropping maize with grain legumes and fertilizer treatment on populations of *Pratylenchus brachyurus* Godfrey (Nematoda) and on the yield of maize (*Zea mays* L.). *Protection Ecology* 6: 153–167.
- El-Haddad SA, Omar MNA, El-Kattan MH. 2003. Comparative studies on some components of integrated management on soil borne plant pathogens affecting cucumber, grown under protected agriculture. In: Abou-Hadid A, ed. *Proceedings of the International Symposium on the horizons of using organic matter substrates in horticulture, presented in Cairo, Egypt April 2022*. *Acta Horticulturae*, 219–226.
- El-Hamawi MH, Youssef MMA, Zawam HS. 2004. Management of *Meloidogyne incognita*, the root-knot nematode, on soybean as affected by marigold and sea ambrosia (damsisa) plants. *Journal of Pest Science* 77: 95–98.
- Ferris H. 1999a. *Nemabase search engine for the host status of plants to nematodes* (revised 13 July 2012). [WWW document] URL <http://plpnemweb.ucdavis.edu/nemaplex/Nemabase2010/NemabasePlantQuery.aspx> [accessed 2 January 2013].
- Ferris H. 1999b. *Meloidogyne hapla* (revised 13 April 2011). [WWW document] URL <http://plpnemweb.ucdavis.edu/nemaplex/taxadata/g076s2.htm> [accessed 2 January 2013].
- Ferris H. 1999c. *Meloidogyne incognita* (revised 15 November 2012). [WWW document] URL <http://plpnemweb.ucdavis.edu/nemaplex/taxadata/g076s3.htm> [accessed 1 January 2013].

Appendix A

Citations used in the meta-analysis

- Aballay E, Flores P, Insunza V. 2001. Nematicidal effect of eight plant species on *Xiphinema americanum* sensu lato in *Vitis vinifera*, var. Cabernet Sauvignon in Chile. *Nematropica* 31: 95–102.
- Abd-Elgawad MM, Saad FF. 1989. Nematode population dynamics on common bean as affected by intercropping with maize. *Beiträge zur Tropischen Landwirtschaft und Veterinärmedizin* 27: 443–448.
- Abdel-Monaim MF, Abo-Elyousr KAM. 2012. Effect of preceding and intercropping crops on suppression of lentil damping-off and root rot disease in New Valley – Egypt. *Crop Protection* 32: 41–46.
- Adegbite AA, Adesiyun SO, Agbaje GO, Omoloye AA. 2005. Host suitability of crops under yam intercrop to root-knot nematode (*Meloidogyne incognita* Race 2) in south-western Nigeria. *Journal of Agriculture and Rural Development in the Tropics and Subtropics* 106: 113–118.
- Aissani N, Tedeschi P, Maietti A, Brandolini V, Garau VL, Caboni P. 2013. Nematicidal activity of allylisothiocyanate from horseradish (*Armoracia*

- Ferris H. 1999d. *Meloidogyne javanica* (revised 13 April 2011). [WWW document] URL <http://plpnemweb.ucdavis.edu/nemaplex/taxadata/g076s4.htm> [accessed 2 January 2013].
- Ferris H. 1999e. *Pratylenchus zaei* (revised 25 October 2012). [WWW document] URL <http://plpnemweb.ucdavis.edu/nemaplex/taxadata/g105s6.htm> [accessed 2 January 2013].
- Ferris H. 1999f. *Rotylenchulus reniformis* (revised 30 November 2012). [WWW document] URL <http://plpnemweb.ucdavis.edu/nemaplex/taxadata/g116s2.htm> [accessed 2 January 2013].
- Fokunang CN, Dixon AGO, Ikotun T, Asiedu R, Tembe EA, Akem CN. 2001. In vitro, greenhouse and field assessments of cassava lines for resistance to anthracnose disease caused by *Colletotrichum gloeosporioides* f.sp. *manihotis*. *Mycopathologia* 154: 191–198.
- Fraser S, Woodward S, Brown AV. 2016. Inter- and intraspecific variation in susceptibility to dothistroma needle blight in Britain. How susceptible are *Pinus sylvestris* and *Pinus contorta*? *Forest Pathology* 46: 534–546.
- Gangwar RK, Chaudhary RG, Kumar K. 2009. Effect of cultural practices and edaphic environment on chickpea wilt caused by *Fusarium oxysporum* f. sp. *ciceri*. *Journal of Food Legumes* 22: 273–275.
- Gao X, Wu M, Xu R, Wang X, Pan R, Kim H-JJ, Liao H. 2014. Root interactions in a maize/soybean intercropping system control soybean soil-borne disease, red crown rot. *PLoS ONE* 9: e95031.
- Germani G, Plenchette C. 2004. Potential of *Crotalaria* species as green manure crops for the management of pathogenic nematodes and beneficial mycorrhizal fungi. *Plant and Soil* 266: 333–342.
- Goel SR, Gupta DC. 2004. Effect of intercropping of nematotoxic plant on tomato in different types of soil against root-knot nematode, *Meloidogyne javanica*. *Haryana Journal of Horticultural Sciences* 33: 142–145.
- Goel SR, Gupta DC, Mehla CP. 2005. Studies on the effect of intercropping of onion on tomato and population dynamics of root knot nematode (*Meloidogyne javanica*) under field conditions. *Haryana Journal of Horticultural Sciences* 34: 164–165.
- Griffin GD. 1994. Effect of single and interplantings on pathogenicity of *Pratylenchus penetrans* and *P. neglectus* to alfalfa and crested wheatgrass. *Journal of Nematology* 26: 460–466.
- Griffin GD, Jensen KB. 1997. Differential effects of *Pratylenchus neglectus* populations on single and interplantings of alfalfa grasses. *Journal of Nematology* 29: 82–89.
- Hackney R, Dickerson O. 1975. Marigold, castor bean, and chrysanthemum as controls of *Meloidogyne incognita* and *Pratylenchus alleni*. *Journal of Nematology* 7: 84–90.
- Hage-Ahmed K, Krammer J, Steinkellner S. 2013. The intercropping partner affects arbuscular mycorrhizal fungi and *Fusarium oxysporum* f. sp. *lycopersici* interactions in tomato. *Mycorrhiza* 23: 543–550.
- Kaitera J, Nuorteva H. 2006. Susceptibility of *Ribes* spp. to pine stem rusts in Finland. *Forest Pathology* 36: 225–246.
- Kaşkavalcı G, Tüzıl Y, Dura O, Öztekin GB. 2009. Effects of alternative control methods against *Meloidogyne incognita* in organic tomato production. *Ekoloji* 18: 23–31.
- Kloos JP, Tulog B, Tumapon AS. 1987. Effects of intercropping potato on bacterial wilt. *Philippine Agriculturist* 70: 83–90.
- Lai R, You M, Jiang L, Lai B, Chen S, Zeng W, Jiang D. 2011. Evaluation of garlic intercropping for enhancing the biological control of *Ralstonia solanacearum* in flue-cured tobacco fields. *Biocontrol Science and Technology* 21: 755–764.
- Lemańczyk G. 2009. Fungal diseases on roots and stem bases of spring rye cultivated in pure stand or in mixtures with other crops. *Phytopathologia* 53: 31–41.
- Lemańczyk G. 2010. The problem of root and stem base health of oat (*Avena sativa* L.) cultivated in mixture with spring rye (*Secale cereale* L.). *Journal of Plant Protection Research* 50: 398–401.
- Marimuthu S, Ramamoorthy V, Samiyappan R, Subbian P. 2013. Intercropping system with combined application of *Azospirillum* and *Pseudomonas fluorescens* reduces root rot incidence caused by *Rhizoctonia bataticola* and increases seed cotton yield. *Journal of Phytopathology* 161: 405–411.
- Mashela PW, Pofu KM. 2017. Nematode resistance to tropical *Meloidogyne* species in *Moringa oleifera*. *Research on Crops* 18: 513–517.
- McIntyre BD, Gold CS, Kashaia IN, Ssali H, Night G, Bwamiki DP. 2001. Effects of legume intercrops on soil-borne pests, biomass, nutrients and soil water in banana. *Biology and Fertility of Soils* 34: 342–348.
- McSorley R. 1999. Host suitability of potential cover crops for root-knot nematodes. *Journal of Nematology* 31: 619–623.
- Meyer JR, Zehr EI, Meagher RL Jr, Salvo SK. 1992. Survival and growth of peach trees and pest populations in orchard plots managed with experimental ground covers. *Agriculture, Ecosystems and Environment* 41: 353–363.
- Mori T, Fujiyoshi T, Matsusaki H. 2011. Comparative influence of root wounds on proliferation of *Ralstonia solanacearum* and bacterial wilt disease in plant species with varying resistances. *Journal of Food, Agriculture and Environment* 9: 641–645.
- Oduor-Owino P. 1993. Effects of aldicarb, *Datura stramonium*, *Datura metel* and *Tagetes minuta* on the pathogenicity of root-knot nematodes in Kenya. *Crop Protection* 12: 315–317.
- Oduor-Owino P, Sikora RA, Waudo SW, Schuster RP. 1996. Effects of aldicarb and mixed cropping with *Datura stramonium*, *Ricinus communis* and *Tagetes minuta* on the biological control and integrated management of *Meloidogyne javanica*. *Nematologica* 42: 127–130.
- Olabiya T, Oyedunmade E. 2007. Marigold (*Tagetes erecta* L.) as interplant with cowpea for the control of nematode pests. *African Crop Science Conference Proceedings* 8: 1075–1078.
- Olasantun FO. 2001. Optimum plant populations for okra (*Abelmoschus esculentus*) in a mixture with cassava (*Manihot esculenta*) and its relevance to rainy season-based cropping systems in south-western Nigeria. *The Journal of Agricultural Science* 136: 207–214.
- Peterson JK, Harrison HF. 2002. Suppression effect of *Capsicum chinense* Jacq. on Southern rootknot nematode [*Meloidogyne incognita* (Kofoid and White) Chitwood] in peppers and tomato. *Allelopathy Journal* 9: 59–62.
- Potter JW, McKeown AW. 2002. Inhibition of *Pratylenchus penetrans* by intercropping of *Rudbeckia hirta* and *Lycopersicon esculentum* in pot cultivation. *Phytoprotection* 83: 115–120.
- Powers LE, McSorley R, Dunn RA, Montes A. 1994. The agroecology of a cucurbit-based intercropping system in the Yeguar Valley of Honduras. *Agriculture, Ecosystems and Environment* 48: 139–147.
- Ren L, Su S, Yang X, Xu Y, Huang Q, Shen Q. 2008. Intercropping with aerobic rice suppressed *Fusarium* wilt in watermelon. *Soil Biology and Biochemistry* 40: 834–844.
- Riaz T, Khan SN, Javaid A. 2009. Effect of co-cultivation and crop rotation on corm rot disease of *Gladiolus*. *Scientia Horticulturae* 121: 218–222.
- Roskov Y, Abucay L, Orrell T, Nicolson D, Kunze T, Culham A, Bailly N, Kirk P, Bourgoin T, DeWalt RE, Decock W, De Wever A, eds. 2014. *Species 2000 & ITIS Catalogue of Life*. [WWW document] URL <https://www.catalogueoflife.org/col> [accessed 29 October 2014].
- Salem FM, Osman GY. 1988. Effectiveness of tagetes natural exudates on *Meloidogyne javanica* (Chitwood) nematode. *Anzeiger für Schädlingskd. Pflanzenschutz Umweltschutz* 61: 17–19.
- Shanks CH, Chamberlain JD. 1993. Strawberry fruit yield and vegetative growth and pest populations in plantings with and without cover crops. *HortScience* 28: 1172–1173.
- Sharma GC, Bajaj BK. 1998. Effect of inter-cropping bell-pepper with ginger on plant parasitic nematode populations and crop yields. *The Annals of Applied Biology* 133: 199–205.
- Sharma GC, Kashyap AS. 2004. Effect of intercropping on the nematode population and yield of peach (*Prunus persica* Batsch.). *Indian Journal of Horticulture* 61: 273–275.
- Sharma GC, Kashyap AS. 2009. Effect of different intercrops on the nematode populations and yield of apricot var. New Castle. *Indian Journal of Horticulture* 66: 420–421.
- Shigaki T, Gray FA, Delaney RH, Koch DW. 1998. Evaluation of host resistance and intercropping for management of the northern root-knot nematode in sainfoin, *Onobrychis viciifolia*. *Journal of Sustainable Agriculture* 12: 23–39.
- Siddiqui M, Alam M. 1987. Control of plant-parasitic nematodes by intercropping with *Tagetes minuta*. *Nematologia Mediterranea* 15: 205–211.

- Speijer PR, Kajumba C, Ssango F. 1999. East African highland banana production as influenced by nematodes and crop management in Uganda. *International Journal of Pest Management* 45: 41–49.
- Sundararaju P. 2005. Effect of marigold, *Tagetes erecta* intercropped with banana against root-lesion nematode, *Pratylenchus coffeae*. *Indian Journal of Nematology* 35: 123–126.
- Tanda AS, Atwal AS. 1988. Effect of sesame intercropping against the root-knot nematode (*Meloidogyne incognita*) in okra. *Nematologica* 34: 484–492.
- Theunissen J, Schelling G. 2000. Undersowing carrots with clover: Suppression of carrot rust fly (*Psila rosae*) and cavity spot (*Pythium* spp.) infestation. *Biological Agriculture and Horticulture* 18: 67–76.
- Timper P, Davis RF, Tillman PG. 2006. Reproduction of *Meloidogyne incognita* on winter cover crops used in cotton production. *Journal of Nematology* 38: 83–89.
- Tsay TT, Wu ST, Lin YY. 2004. Evaluation of Asteraceae plants for control of *Meloidogyne incognita*. *Journal of Nematology* 36: 36–41.
- Ushiki J, Tahara S, Hayakawa Y, Tadano T. 1998. Medicinal plants for suppressing soil-borne plant diseases II. Suppressive effect of *Geranium pratense* L. on common scab of potato and identification of the active compound. *Soil Science & Plant Nutrition* 44: 157–165.
- Vrain T, DeYoung R, Hall J, Freyman S. 1996. Cover crops resistant to root-lesion nematodes in raspberry. *HortScience* 31: 1195–1198.
- Walker JT. 1971. Populations of *Pratylenchus penetrans* relative to decomposing nitrogenous soil amendments. *Journal of Nematology* 3: 43–49.
- Whittington DP, Zehr EI. 1992. Populations of *Criconebella-xenoplax* on peach interplanted with certain herbaceous plants. *Journal of Nematology* 24: 688–692.
- Wu H, Wang C-J, Bian X-W, Zeng S-Y, Lin K-C, Wu B, Zhang G-A, Zhang X. 2011. Nematicidal efficacy of isothiocyanates against root-knot nematode *Meloidogyne javanica* in cucumber. *Crop Protection* 30: 33–37.
- Yang Y, Fengzhi W, Shouwei L. 2011. Allelopathic effects of root exudates of Chinese onion accessions on cucumber yield and *Fusarium oxysporum* f. sp. *cucumerinum*. *Allelopathy Journal* 27: 75–85.
- Yassin MY, Ismail AE. 1994. Effect of *Zinnia elegans* as a mix-crop along with tomato against *Meloidogyne incognita* and *Rotylenchulus reniformis*. *Anzeiger für Schädlingskunde, Pflanzenschutz, Umweltschutz* 67: 41–43.
- Zehr EI, Aitken JB, Scott JM, Meyer JR. 1990. Additional hosts for the ring nematode, *Criconebella xenoplax*. *Journal of Nematology* 22: 86–89.
- Zehr EI, Lewis SA, Bonner MJ. 1986. Some herbaceous hosts of the ring nematode (*Criconebella xenoplax*). *Plant Disease* 70: 1066–1069.

Supporting Information

Additional Supporting Information may be found online in the Supporting Information section at the end of the article.

Fig. S1 Funnel plot analysis for the ‘Nematode Uncontained’ model 1 (Table S4).

Fig. S2 Funnel plot analysis for the ‘Nematode Contained’ model 2 (Table S4).

Fig. S3 Funnel plot analysis for the ‘Disease Uncontained’ model 1 (Table S4).

Notes S1 Search query strings.

Table S1 Moderator variables, with categories/ranges and number of experiments that reported them.

Table S2 Sources of information on currently accepted binomials, plant host status and susceptibility, and standard agronomic practices.

Table S3 Summary meta-analyses results.

Table S4 Name, size, initial and (where reached) final model specifications, and moderator omnibus test results for each meta-regression performed using the nematode and disease data.

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