RESEARCH ARTICLE

Open Access



DNA methylation changes in cord blood and the developmental origins of health and disease – a systematic review and replication study

Loubna Akhabir^{1,2†}, Randa Stringer^{1,2†}, Dipika Desai¹, Piush J Mandhane³, Meghan B Azad⁴, Theo J Moraes⁵, Padmaja Subbarao^{2,5}, Stuart E Turvey⁶, Guillaume Paré^{1,7,8,9,10}, Sonia S. Anand^{1,2,10*} and for the NutriGen Alliance

Abstract

Background: Environmental exposures *in utero* which modify DNA methylation may have a long-lasting impact on health and disease in offspring. We aimed to identify and replicate previously published genomic loci where DNA methylation changes are attributable to *in utero* exposures in the NutriGen birth cohort studies Alliance.

Methods: We reviewed the literature to identify differentially methylated sites of newborn DNA which are associated with the following five traits of interest maternal diabetes, pre-pregnancy body mass index (BMI), diet during pregnancy, smoking, and gestational age. We then attempted to replicate these published associations in the Canadian Healthy Infant Longitudinal Development (CHILD) and the South Asian birth cohort (START) cord blood epigenome-wide data.

Results: We screened 68 full-text articles and identified a total of 17 cord blood epigenome-wide association studies (EWAS) of the traits of interest. Out of the 290 CpG sites reported, 19 were identified in more than one study; all of them associated with maternal smoking. In CHILD and START EWAS, thousands of sites associated with gestational age were identified and maintained significance after correction for multiple testing. In CHILD, there was differential methylation observed for 8 of the published maternal smoking sites. No other traits tested (i.e., folate levels, gestational diabetes, birthweight) replicated in the CHILD or START cohorts.

Conclusions: Maternal smoking during pregnancy and gestational age are strongly associated with differential methylation in offspring cord blood, as assessed in the EWAS literature and our birth cohorts. There are a limited number of reported methylation sites associated in more than two independent studies related to pregnancy. Additional large studies of diverse populations with fine phenotyping are needed to produce robust epigenome-wide data in order to further elucidate the effect of intrauterine exposures on the infants' methylome.

Keywords: Cord blood; methylation; EWAS; prenatal, In utero, CHILD, START, Nutrigen

*Correspondence: anands@mcmaster.ca [†]Loubna Akhabir and Randa Stringer contributed equally to this work. ¹⁰ Department of Health Research Methods, Evidence, and Impact, McMaster University, Hamilton, Ontario, Canada Full list of author information is available at the end of the article



Background

The Developmental Origins of Health and Disease (DOHaD) is an area of research that investigates the influence of pregnancy and early life exposures on the growth, development, and health of the offspring. Fetal

© The Author(s) 2022. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, wisit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

programming is a process whereby the *in utero* environment may prime a fetus for an expected environment after delivery, which may have a significant impact on lifelong health. It is thought that epigenetic mechanisms, in particular DNA methylation, mediate fetal programming [1, 2] and may modify trajectories of health and disease throughout the offspring's lifetime [1, 3, 4]. Technological advances have enabled expansive evaluations of epigenetic modifications [1] so that researchers are now able to interrogate the epigenome for signs of fetal programming.

Maternal nutrition and chemical exposures are associated with epigenetic changes which may lead to the development of a variety of non-communicable diseases. It is thought that these epigenetic changes are responsible for what is described as the developmental origins of health and disease whereby early life environmental stresses influence life-long disease risk [5]. For instance, the maternal famine exposure in the Dutch Hunger Winter of 1944 to 1945 and Chinese famine of 1959 to 1961, is linked with an increased incidence of type-2 diabetes in the offspring [6]. This phenomenon of "developmental programming" has also been shown in less severe exposures such as in response to a dysglycemic *in utero* environment and has been linked to altered DNA methvlation patterns in placental and infant tissues [7, 8]. For instance, differential methylation and gene expression have been observed at the IGF2/H19 locus, an imprinted region involved in metabolic programming, in the blood samples of offspring exposed to intrauterine hyperglycemia in human and animal studies [9, 10]. Maternal obesity, excess pregnancy weight gain, and gestational diabetes mellitus (GDM) are risk factors for negative health outcomes in the offspring. There is a mounting body of evidence that several other exposures including Page 2 of 9

maternal smoking and nutrition also lead to adverse child health outcomes, potentially through modification of their DNA methylation profile [11-13].

We sought to identify and replicate previously published genomic loci where DNA methylation changes are attributable to *in utero* exposures, including maternal GDM, maternal pre-pregnancy body mass index (BMI), diet during pregnancy, maternal smoking, and to perinatal traits such as gestational age and birthweight.

First, we searched the literature to identify robustly associated methylation sites for these exposures/traits of interest, and second, we attempted to replicate these robust published associations in two independent prospective birth cohort studies of South Asians and white Caucasians in which epigenome-wide data are available.

Methods

Literature review

We conducted a generalized search in the PubMed database to identify CpG probes associated with our 5 exposures of interest and perinatal traits: prenatal nutrition (especially dietary patterns), maternal smoking, maternal or infant dysglycemia (especially GDM), gestational age, and maternal pre-pregnancy BMI. The following search was used: "cord blood" AND "DNA methylation" AND "pregnancy" AND ("diet" OR "nutrition" OR "dietary pattern" OR "diabetes" OR "dysglycemia" OR "glucose" OR "insulin" OR "smoking" OR "gestational age" OR "weight" OR "BMI"). The literature search was conducted in January 2020. Figure 1 delineates the steps and articles inclusion.

Inclusion criteria were: (i) studies in cord blood, (ii) studies reporting epigenome-wide associations and (iii) number of cases greater than 50. Targeted studies, those that reported statistically non-significant results,



or EWAS in peripheral blood or placental tissue were excluded.

All abstracts were then screened for our inclusion and exclusion criteria, with some studies moving on for full text evaluation and possible inclusion in the review. Our search returned a total of 101 results which were screened at the abstract level. Of these, 68 full-text articles were accessed for closer inspection, and a final of 19 studies were identified for inclusion in our review.

Study population, EWAS, and Replication Study

The NutriGen study, is an alliance of four birth cohorts, and has been described previously [14]. Briefly, the NutriGen study aims to investigate nutrition and environmental determinants of maternal and childhood health, with a focus on genetic, epigenetic, and microbiome contributions. Cord blood samples were collected at birth for genetic and cardiometabolic analysis. The present report deals with data from two of the NutriGen cohorts in whom EWAS was conducted: the South Asian birth cohort (START) [15] and the Canadian Healthy Infant Longitudinal Development (CHILD) [16] (Additional File 1). Newborn anthropometric information was collected at the time of birth and cord blood collected where possible.

All pregnant women completed a food frequency questionnaire (FFQ) to assess prenatal diet and were assessed for anthropometric and other health measures (Additional File 1). Principal component (PC) analysis of FFQ data was used to derive individual scores representing dietary patterns. Individual nutrient information regarding fatty acid intake was used, including polyunsaturated fatty acids (PUFAs), saturated fats, and the ratio of PUFAs to saturated fats (P:S).

Two diet quality scores, the modified Alternative Healthy Eating Index (mAHEI) and the DOHaD score developed by the NutriGen group, were calculated and used in analysis. Dietary variables were adjusted for total energy consumption when applicable.

These variables have been harmonized across cohorts for comparisons. A harmonized GDM variable was available in both START and CHILD. Maternal smoking and pre-pregnancy BMI were only available in CHILD; whereas in START there were no smokers during pregnancy, and BMI was measured in the second trimester.

Epigenome Wide Association Study (EWAS)

DNA was extracted from 512 START and 511 CHILD cord blood samples and subjected to bisulfite conversion. The samples were hybridized to the Illumina Human-Methylation450K beadchip array. Raw data generated by the Illumina iScan software were processed using the minfi R package [17]. Quality control and pre-processing

were performed in each cohort separately. Samples were excluded if they showed > 0.01 failed probes or had a discrepancy between genetic sex and reported sex. Probes were excluded based on >0.05 missingness, and if they contained a SNP or showed cross-reactivity [18]. Clean data were obtained for 393,400 probes in 491 START samples and for 393,449 probes in 511 CHILD samples. ReFaCTor reference-free algorithm was used to obtain principal components of cell composition [19, 20]. For CHILD and START cohorts separately, cord blood DNA methylation was assessed for association using multivariate linear regression with the following exposures of interest / prenatal traits: (1) diet, including dietary patterns, macronutrient intake, diet quality scores, (2) gestational age, (3) GDM, and (4) smoking during pregnancy (CHILD only), and pre-pregnancy BMI (CHILD only). The following variables were included as covariates in the association models: maternal age, infant sex, gestational age, study centre, processing time (CHILD only), smoking (CHILD only), and cellular composition. Statistical analyses were conducted in R version 3.2.0. The Bonferroni correction was used to adjust for multiple testing, with a significance threshold of $p < 1.27 \times 10^{-7}$. EWAS analysis in CHILD was restricted to the European subset (N=295), due to low sample size from other ethnic groups. Gene set enrichment analysis for gestational age significant results was performed using the R package 'enrichr' [21].

Replication study

Where data were available, we used the START and CHILD DNA methylation to conduct a targeted replication analysis based on the findings of the literature review. Analyses were repeated similarly as in the EWAS using linear regression models, for only those sites identified in the literature as significantly associated with a given outcome.

Multivariate linear regression models were used to test for association between DNA methylation β values at each site and the exposures/traits variables. The association models were adjusted, as appropriate, for factors suspected to influence DNA methylation: maternal age, infant sex, gestational age, study centre, processing time (CHILD only), smoking (CHILD only), and cell composition. The Bonferroni correction was used to adjust for multiple testing, using the number of probes tested for each replication analysis. We adjusted for appropriate covariates for each replication analysis. Specifically, for the CHILD maternal smoking replication analysis, we adjusted for maternal age, sex, gestational age, study centre, cellular composition. For the folate level replication analysis in START, we adjusted for maternal age, sex, gestational age, total homocysteine levels, vitamin B12 levels, RBC folate levels, study centre and cellular composition.

For the birthweight replication in CHILD and START, we adjusted for maternal age, sex, smoking, gestational age, study centre, and cellular composition.

Estimates for diet, birthweight, GDM and gestational age from CHILD and START models were meta-analyzed using the 'meta' R package inverse variance method, to obtain fixed and random effect estimates as well as a heterogeneity statistic.

Results

Literature review

Using the keywords for traits of interest, we identified 68 studies for which we read the full text publications, 41 of them were EWAS. After excluding studies with less than 50 cases, 31 remained. Nineteen of these studies reported significant methylation results and were included in our analysis.

These studies reported 259 significantly associated DNA methylation probes for the following traits: (1) birthweight (12 probes) and gestational age (22 probes), (2) childhood adiposity (6 probes), (3) GDM (96 probes) and maternal diabetes (16 probes), (4) maternal obesity (3 probes), (5) maternal eating disorder (2 probes), (6) maternal polyunsaturated fatty acids (PUFA) (6 probes) and trans fatty acids (FA) (5 probes) at preconception, (7) maternal plasma folate levels (48 probes) and (8) maternal smoking during pregnancy (60 probes).

The 259 identified sites were significantly associated with their trait after correction for multiple testing. Bonferroni correction was used for correction in 9/18 publications, while 9/18 used FDR. One article reported 18 methylation sites associated with GDM without any multiple testing correction. We added the non-corrected GDM sites to our list of interest for a total number of 277 sites to include in our analysis.

Among the 277 sites, only 19 replicated in at least one other study. All of these were maternal smokingassociated probes. A list of the probes is provided in the Additional Table 1. Additional Table 2 details all studies identified, and the probes extracted. We also noted that the birthweight publication [22] reported robust findings from a large meta-analysis of EWAS; and we included 12 sites where the association persisted in childhood peripheral blood in our sites to test for replication.

EWAS

CHILD and START study participants' characteristics are shown in Table 1.

We observed no epigenome-wide significant differential DNA methylation for diet, maternal smoking or maternal GDM, after correction for multiple testing in separate cohort analyses or in a meta-analysis of the cohorts. Gestational age was highly associated with thousands of sites in both cohorts independently. The meta-analysis yielded 5,732 significant associations after Bonferroni correction for multiple testing. Gene set enrichment analyses of the significant results did not yield any statistically significant enriched Gene Ontology (GO) pathway (Additional Table 3). Table 2 indicates the top 10 sites identified in the meta-analysis. All 5,732 significant results are listed in Additional Table 4. A volcano plot of the results is provided as Additional Fig. 1.

Targeted replication

Using the probes identified using the literature review, we conducted replication analyses in START and CHILD for plasma folate levels (48 sites) and maternal smoking (25 sites), respectively. Both single study analyses and metaanalyses were conducted for birthweight (12 sites) and gestational age (279 sites).

Gestational age in CHILD and START meta-analysis was significantly associated with 37% of the sites identified in the literature, all with consistent direction of effects (Additional Table 5).

Eight of the maternal smoking probes showed statistically significant association with the exposure in CHILD, after correction with multiple testing (Table 3). START mothers were all non-smokers.

While no site survived Bonferroni correction for multiple testing in the single study analyses of birthweight, 3 sites showed significant negative association

Table 1 Characteristics of	CHILD and STAR	FEWAS participants
----------------------------	----------------	--------------------

Characteristics	START (South Asians)	CHILD (European subset)
Sample size	491	295
Maternal age (years, mean \pm SD)	30.9 ± 3.9	32.7±4.4
Infant sex (% Female)	51.9	46.1
Gestational age (weeks, mean \pm SD)	39.2±1.3	39.5 ± 1.3
Birth weight (grams, mean \pm SD)	$3,265 \pm 454.5$	3,498.6±485.9
Smoking (N /%)	0/0	23 / 7.8

in meta-analysis after Bonferroni correction. No significant heterogeneity was detected between CHILD and START in this analysis, therefore fixed-effect estimates are reported (Table 4)

There were no significant associations between START methylation and maternal plasma folate levels at the 48 sites from the Joubert et al. study [23]. Similarly, none of the identified GDM probes exhibited significant

association with GDM in our data. All replication results including null associations are provided in Additional Table 6

Discussion

In this report, we show that the cord blood EWAS literature based on individual CpG probe methylation differential in relation with our prenatal exposures of interest,

Table 2 Top 10 sites most significantly associated with gestational age in	the meta-analysis of START and CHILD
--	--------------------------------------

Site Chr		Position	Gene	Estimate (CI)	Nominal P value	Adjusted P value	
cg11932158	3	155,422,129	PLCH1	-0.019 (-0.022, -0.016)	4.8×10^{-47}	1.9×10^{-41}	
cg18623216	3	155,421,970	PLCH1	-0.021 (-0.024, -0.018)	3.7×10^{-46}	1.5×10^{-40}	
cg16103712	8	99,023,869	MATN2	-0.023 (-0.026, -0.020)	1.2×10^{-45}	4.8×10^{-40}	
cg17133774	1	6,198,667	CHD5	-0.018 (-0.020, -0.015)	4.1×10^{-45}	1.6×10^{-39}	
cg12713583	19	940,724	ARID3A	-0.022 (-0.025, -0.019)	6.3×10^{-45}	2.5×10^{-39}	
cg04347477	12	125,002,007	NCOR2	-0.019 (-0.022, -0.016)	5.3×10^{-41}	2.1×10^{-35}	
cg08817867	17	19,656,554	-	-0.018 (-0.020, -0.015)	5.4×10^{-39}	2.1×10^{-33}	
cg02001279	19	940,967	ARID3A	-0.013 (-0.015, -0.011)	7.7×10^{-39}	3.0×10^{-33}	
cg08412913	16	85,429,522	-	-0.011 (-0.013, -0.010)	1.3×10^{-35}	4.9×10^{-30}	
cg06870470	19	11,315,767	DOCK6	-0.019 (-0.022, -0.016)	8.2×10^{-35}	3.2×10^{-29}	

Chr chromosome, *Cl* confidence interval; Adjusted p value: Bonferroni-corrected p value; bolded (or italic) CpG sites are also top 10 significant associations in the separate START (or CHILD) analysis; bolded and italic sites were in the top 10 significant associations in both cohort specific analyses. Analyses were adjusted for maternal age, sex, study centre and cellular composition. Estimates are expressed as the change in methylation beta value per one unit change in gestational age in weeks

Table 3 Replicated maternal smoking-associated sites in CHILD

Site	Chr	Position	Gene	Estimate (se)	Nominal <i>P</i> value	Adjusted P value	Literature Estimate (se)	Author (PMID)
cg05549655	15	75,019,143	CYP1A1	0.027 (0.006)	3.8×10^{-6}	9.4 × 10 ⁻⁵	0.019 (0.001)	Joubert et al. 27,040,690
cg11924019	15	75,019,283	CYP1A1	0.043 (0.010)	2.2×10^{-5}	5.6×10^{-4}	0.025 (0.002)	Joubert et al. 27,040,690
cg22549041	15	75,019,251	CYP1A1	0.047 (0.011)	3.5×10^{-5}	8.9×10^{-4}	0.066 (0.004)	Joubert et al. 27,040,690
cg23067299	5	323,907	AHRR	0.036 (0.008)	4.7×10^{-5}	0.001	0.030 (0.002)	Joubert et al. 27,040,690
cg22132788	7	45,002,486	MYO1G	0.051 (0.013)	7.9×10^{-5}	0.002	0.023 (0.001)	Joubert et al. 27,040,690
cg18092474	15	75,019,302	CYP1A1	0.041 (0.010)	1.2×10^{-4}	0.003	0.051 (0.003)	Joubert et al. 27,040,690
cg12803068	7	45,002,919	MYO1G	0.060 (0.016)	2.8×10^{-4}	0.007	0.070 (0.003)	Joubert et al. 27,040,690
cg12101586	15	75,019,203	CYP1A1	0.034 (0.009)	4.1×10^{-4}	0.010	0.045 (0.003)	Joubert et al. 27,040,690

Chr chromosome; Estimate: regression coefficient from association analysis; se: standard error; Adjusted P: Bonferroni-corrected p value. Analyses were adjusted for maternal age, sex, gestational age, study centre and cellular composition. The estimates are expressed as the average difference in methylation beta values between smokers and non-smokers mothers

Site	Chr	Position	Gene	Estimate (CI)	Nominal P value	Adjusted P value	Literature estimate (CI)	Author (PMID)
 cg26995690	13	36,346,376	DCLK1	-13.8 (-22.4, -5.	.2) 0.0017	0.020	-48.7 (-62.9, -34.5)	Kupers et al. 31,015,461
cg00637745	2	121,497,334	-	-35.9 (-58.6, -13.	.3) 0.0019	0.023	-20.9 (-26.8, -15.1)	Kupers et al. 31,015,461
cg07133097	2	121,497,538	-	-21.9 (-35.8, -7.	.9) 0.0021	0.025	-29.2 (-37.5, -20.8)	Kupers et al. 31,015,461

Chr: chromosome; Cl: confidence interval; Adjusted P value: Bonferroni-corrected p value; Published estimate: coefficients from Kupers at al birthweight meta-analysis of all ancestries (PMID: 31,015,461). Analyses were adjusted for maternal age, sex, maternal smoking, gestational age, study centre and cellular composition. The estimates are expressed as the change in methylation beta values per one unit change in birthweight (grams)

is limited in the number of publications, as well as in the replication of associated probes. Out of the 101 publications retrieved, nineteen articles reported cord blood EWAS with significant differentially methylated sites. Most the published significant probes were only found in one study. Only 19 out of 290 probes replicated in at least another study, and all of the replicated probes were associated with maternal smoking.

CHILD and START EWAS yielded no association which survived statistical correction, except for probes associated with gestational age. However, meta-analysis of the 2 CHILD and START identified 3 significant birthweight sites.

While the use of methylation arrays allows for hypothesis-free investigations, it requires greater number of samples in order to have enough statistical power to detect effects after correcting for the multiple hypotheses testing. Many published studies have shown interesting associations between metabolic maternal measures and cord blood methylome but were underpowered [24]. Our literature review, which aimed to identify robust differentially methylated sites associated with exposures of interest and perinatal traits, showed that among hundreds of associated probes, only 19 replicated in more than one study, all of which are associated with maternal smoking. In the subset of exposure/traits we explored in our study, we showed that 4 exposures/traits show strong associations with DNA methylation: maternal smoking, maternal plasma folate levels, gestational age and birthweight, as defined by statistical significance in EWAS with greater than 50 cases and using Bonferroni correction for multiple testing. While the folate and birthweight sites originated from meta-analyses of multiple studies ^{13,14}, the 86 GDM-associated probes we identified were from a small EWAS study of 88 cases [25], where FDR was used for correction (instead of Bonferroni). This study reported some FDR-corrected findings of small effect sizes.

Guided by the literature identified methylation sites, we were able to replicate associations at sites related to maternal smoking, gestational age, and birthweight, but not for GDM or plasma folate levels. Cigarette smoking was one of the strongest influential factors for the methylome as compared to other exposures. Its modulation of DNA methylation and gene expression seems to persist over time and is associated with a variety of negative health outcomes [26].

The plasma folate study from which we extracted probes, was performed in European subjects, whereas the START subjects are of South Asian descent. This may explain the lack of replication, as DNA methylation of certain areas of the genome does vary across different ancestral populations, due to a combination of different allele frequencies and complex gene-environment interactions [27–29]. Ethnicity has also been documented to be an independent determinant of folate status in a study which controlled for folate intake [30]. Canadian South-Asian women in particular have been found to have high levels of B12 deficiency compared to women of other ethnic backgrounds; and B12 and folate are metabolically inter-dependent [31].

A recent systematic review of preterm birth cohorts included only ethnically homogeneous populations (i.e. Europeans), preventing comparisons between ethnically diverse epigenomes [32]. In a replication study of the association of chronic diseases with published differentially methylated sites in 120 Batswana South African men, 86% of the associations replicated in terms of consistent size and direction of effect (but not significant p values). There were 14% of African population-specific associations which were not previously observed in European published sites [33]. Ethnic-specific methylation changes were also observed in smoking sites [34] and in cancer pathway regions at birth [35]. The epigenetic clock (epigenetic biomarkers of aging) has been proposed as a way to elucidate disease burden differential in different population since it is strongly associated with genetic race/ethnicity [36]. However, this association differs when using different epigenetic clocks, comprised of different CpG sites [37]; this warrants continued research efforts in elucidating the epigenetic/genetic race association. Using 3 different epigenetic clocks, it was shown that African-Americans generally have more wide-spread methylation changes than white Caucasians and that sex also interacts with genetic race in determining epigenetic age [38]. Together, these data certainly warrant more studies of differential methylation profiles in non-white subjects and the relationship of methylation profiles with environmental exposures in different populations.

The DOHaD framework stipulates the importance of pre-conception and early life in future health outcomes. Maternal obesity and diabetes, for instance, are known risk factors for poor fetal outcomes such as fetal/infant death, low birthweight, impaired lipid profiles and higher BMI in childhood [39–41]. Animal and human studies (including extreme food deprivation studies) have shown that DNA methylation is a major mechanism by which intrauterine and environmental exposures influence fetal development and future child health [6, 42, 43].

The life course of non-communicable disease highlights an opportune period in early life where interventions could have a high impact on life-long health and disease [5]. This early 'plasticity' period is the target study period of birth cohorts. A challenge is the need for well-phenotyped birth cohort with ample sample size. We previously summarized our findings on the effect of maternal and infant nutrition on cardiometabolic risk, and developed a harmonization tool to enable standardized nutrition research in different birth cohorts [44].

Strengths and Limitations: Strengths of our study include: (a) our interrogation of probes was guided by the literature to reduce number of statistical tests, (b) testing for replication in two cohorts including one of South Asian ethnicity, and (c) specificity of the START GDM phenotype which was ascertained directly by glucose oral tests, which increases the accuracy of the exposure. A main limitation of our study is the statistical power. The number of subjects of START and CHILD included in the EWAS was relatively small, which likely contributes to the few sites identified in our analysis. Another limitation of our study is the use of beta values instead of M-values for analysis of differential methylation using regression models. Despite our use of the SWAN normalization to correct noise in beta values, our results may be influenced by heteroscedasticity of beta-values of probes in the high and low methylation range. In addition, enrichment analysis of the gestational age results was performed using the Enrichr method which may be biased due to uneven representation of probes in the DNA methylation array, hence the lack of significant enrichment results may be due to this bias. Lastly, we used a reference-free unsupervised method for adjusting differential methylation analysis for cell composition, ReFACTor which was shown to improve performance on the Houseman cord blood reference dataset [20]. A good reference-based alternative would be to combine 4 MACS and FACS-sorted datasets and use the Identifying Optimal Libraries (IDOL) filter as described here [45]; which could have improved performance of the cell composition adjustment.

Conclusions

Our study confirms the strong effect of maternal smoking during pregnancy on the infant's methylome. Gestational age is also highly associated with differential methylation in 2 independent cohorts of different ethnicity, as well in their meta-analysis. Some previously identified birthweight sites were replicated in the meta-analysis of our 2 cohorts. There is a clear need for larger studies in diverse populations in order to establish the real effects of *in utero* exposures on infant and child health. Published EWAS associations should be reviewed carefully, for sample size and statistical correction; as well as compared to other studies to increase confidence in existing data.

Abbreviations

BMI: body mass index; CHILD: Canadian healthy infant longitudinal development; Dohad: Developmental Origins of Health and Disease; EWAS: epigenome-wide association study; FDR: false discovery rate; FFQ: food frequency questionnaire; GDM: gestational diabetes mellitus; START: South Asian birth cohort; T2DM: type-2 diabetes mellitus.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12864-022-08451-6.

Additional file 1. Supplementary information. This file contains a description of the NutriGen alliance studies,data and samples collection details and sample and probe quality control procedures.

Additional file 2. List of associated probes from the literature search. Information about the total methylation sites in the published literature.

Additional file 3. List of studies identified in literature search. Details of all studies identified in literature search.

Additional file 4. Gene set enrichment analysis for gestational age associated genes. Results of gene set enrichment analysis for genes associated with gestational age in meta-analysis of CHILD and START.

Additional file 5. Results of EWAS meta-analysis of gestational age. A list of significant gestational age meta-analysis association results.

Additional file 6. Meta-analysis association of Gestational age with methylation in CHILD and START. Results of meta-analysis association analysis of Gestational age in CHILD and START with methylation in sites identified in the literature.

Additional file 7. Replication association results. Significant and null results from replication efforts for all traits.

Additional file 8. Volcano plot of gestational age EWAS results. Volcano plot showing gestational age EWAS meta-analysis results. Red horizontal line shows the Bonferroni correction threshold. The Y axis represents -log10 of the P values. The Y axis represents the regression coefficient of the association analysis of methylation change. *No correction for cell composition; no significant findings; gestational clock panels; sex-stratified analysis; no individual CpG analysis.

Acknowledgements

We are grateful to all the families who took part in this study, and the whole START and CHILD teams, which includes interviewers, nurses, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, and receptionists.

We acknowledge the valuable contribution of the Genetic and Molecular Epidemiology Laboratory (GMEL), affiliated with Hamilton Health Sciences and McMaster University. All epigenetic assessments including sample handling, and all technical handling was performed by technical staff of GMEL. The members of the Nutrigen Alliance are: Sonia S. Anand; Stephanie (A) Atkinson; Meghan (B) Azad; Allan B. Becker; Jeffrey Brook; Judah A Denburg; Dipika Desai; Russell J. de Souza; Milan Gupta; Michael Kobor; Diana L. Lefebvre; Wendy Lou; Piushkumar J. Mandhane; Sarah McDonald; Andrew Mente; David Meyre; Theo J. Moraes; Katherine Morrison; Guillaume Paré; Malcolm R. Sears; Padmaja Subbarao; Koon K. Teo; Stuart E. Turvey; Julie Wilson; Salim Yusuf; Stephanie Atkinson; Gita Wahi; Michael A. Zulyniak.

Authors' contributions

LA. and R.S. contributed to conceptualization, performed the literature review, all analysis of epigenetic data, all other investigations, and writing of first draft and subsequent editing. D.D. contributed to data curation, securing funding and project administration. PJ.M., M.B.A., T.J.M., P.S. and S.E.T. contributed to data curation, funding acquisition and critical review of manuscript. G.P. and S.A.A. were responsible for study conceptualization, supervision, data curation, funding acquisition, and resources and financial support to L.A. and R.S. All authors participated in manuscript.

Funding

This work was supported by funding by a Canadian Institute for Health Research (CIHR) Grant in Food and Health Population Health Research grant (RFA# 201301FH6; 2013–2018). START data were collected as part of a bilateral ICMR /CIHR funded programme (INC-109205) and HSF Canada Grant in Aid (NA7283). CIHR provided funds for CHILD (AEC85761) data collection, with additional funding from AllerGen NCE Inc. The funding bodies had no role in the design of the study and collection, analysis, and interpretation of data and in writing the manuscript. S.S.A. holds a Canada Research Chair Tier 1 in Ethnic Diversity and Cardiovascular Disease and a Heart and Stroke Foundation / Michael G. De Groote Chair in Population Health Research. S.E.T. holds a Canada Research Chair Tier 1 in Pediatric Precision Health. M.B.A. holds a Canada Research Chair in the Developmental Origins of Chronic Disease. The funders did not have any role in the study design, data collection, analysis, interpretation or in preparation of the manuscript.

Availability of data and materials

All relevant data are within the paper and its Additional Information files. Raw data cannot be shared publicly because of lack of participant consent but are available upon reasonable request from the corresponding author.

Declarations

Ethics approval and consent to participate

Written informed consent was obtained from all participants, including from parents or legal guardians for minors. Study ethics approval were obtained from the Hamilton Integrated Research Ethics Board independently for the CHILD (REB 07–2929) and START studies (REB 10–640) in Canada.

Consent for publication

Not applicable.

Competing interests

The authors have no competing interests.

Author details

¹Population Health Research Institute, David Braley Cardiac, Vascular and Stroke Research Institute, 237 Barton Street East, Hamilton, ON, Canada. ²Department of Medicine, McMaster University, 1280 Main Street West, L8S 4K1 Hamilton, ON, Canada. ³Department of Pediatrics, University of Alberta, Edmonton, AB, Canada. ⁴Department of Pediatrics and Child Health, Children's Hospital Research Institute of Manitoba, University of Manitoba, Winnipeg, MB, Canada. ⁵Department of Pediatrics, Hospital for Sick Children, ON, Toronto, Canada. ⁶Department of Pediatrics, BC Children's Hospital, University of British Columbia, Vancouver, BC, Canada. ⁷Thrombosis and Atherosclerosis Research Institute, David Braley Cardiac, Vascular and Stroke Research Institute, 237 Barton Street East, Hamilton, ON, Canada. ⁸Department of Pathology and Molecular Medicine, Michael G. DeGroote School of Medicine, McMaster University, 1280 Main Street West, Hamilton, ON, Canada. ⁹Department of Clinical Epidemiology & Biostatistics, McMaster University, 1280 Main Street West, Hamilton, ON, Canada. ¹⁰Department of Health Research Methods, Evidence, and Impact, McMaster University, Hamilton, Ontario, Canada.

Received: 20 June 2021 Accepted: 7 March 2022 Published online: 19 March 2022

References

- Safi-Stibler S, Gabory A. Epigenetics and the Developmental Origins of Health and Disease: Parental environment signalling to the epigenome, critical time windows and sculpting the adult phenotype. SI Chromatin Dyn Regen. 2020;1:172–80.
- Briana DD, Malamitsi-Puchner A. Perinatal Biomarkers Implying 'Developmental Origins of Health and Disease' Consequences in Intrauterine Growth Restriction. Acta Paediatr. 2020;1(7):1317–22.
- Simeoni U, Armengaud J-B, Siddeek B, Tolsa J-F. Perinatal Origins of Adult Disease. Neonatology. 2018;113(4):393–9.
- Rodger EJ, Chatterjee A. The Epigenomic Basis of Common Diseases. Clin Epigenetics. 2017;9(1):5.
- Hanson MA, Gluckman PD. Early Developmental Conditioning of Later Health and Disease: Physiology or Pathophysiology? Physiol Rev. 2014;94(4):1027–76.
- Zimmet P, Shi Z, El-Osta A, Ji L. Epidemic T2DM, Early Development and Epigenetics: Implications of the Chinese Famine. Nat Rev Endocrinol. 2018;14(12):738–46.

- Chu AHY, Godfrey KM. Gestational Diabetes Mellitus and Developmental Programming. Ann Nutr Metab. 2021/01/19 ed. 2020;76(Suppl 3):4–15.
- Godfrey KM, Costello PM, Lillycrop KA. The Developmental Environment, Epigenetic Biomarkers and Long-Term Health. J Dev Orig Health Dis. 2015;6(5):399–406.
- Ding G-L, Wang F-F, Shu J, Tian S, Jiang Y, Zhang D, et al. Transgenerational Glucose Intolerance With Igf2/H19 Epigenetic Alterations in Mouse Islet Induced by Intrauterine Hyperglycemia. Diabetes. 2012;61(5):1133–42.
- Su R, Wang C, Feng H, Lin L, Liu X, Wei Y, et al. Alteration in Expression and Methylation of IGF2/H19 in Placenta and Umbilical Cord Blood Are Associated with Macrosomia Exposed to Intrauterine Hyperglycemia. PLoS ONE. 2016;11(2):e0148399.
- 11. Franzago M, Santurbano D, Vitacolonna E, Stuppia L. Genes and Diet in the Prevention of Chronic Diseases in Future Generations. Int J Mol Sci. 2020;21(7):2633. https://doi.org/10.3390/ijms21072633.
- Jaddoe VWV, de Jonge LL, van Dam RM, Willett WC, Harris H, Stampfer MJ, et al. Fetal Exposure to Parental Smoking and the Risk of Type 2 Diabetes in Adult Women. Diabetes Care. 2014;37(11):2966–73.
- Goyal D, Limesand SW, Goyal R. Epigenetic Responses and the Developmental Origins of Health and Disease. J Endocrinol. 2019;242(1):T105–19.
- de Souza RJ, Zulyniak MA, Desai D, Shaikh MR, Campbell NC, Lefebvre DL, et al. Harmonization of Food-Frequency Questionnaires and Dietary Pattern Analysis in 4 Ethnically Diverse Birth Cohorts. J Nutr. 2016;146(11):2343–50.
- Anand SS, Vasudevan A, Gupta M, Morrison K, Kurpad A, Teo KK, et al. Rationale and Design of South Asian Birth Cohort (START): a Canada-India Collaborative Study. BMC Public Health. 2013;28(1):79.
- Moraes TJ, Lefebvre DL, Chooniedass R, Becker AB, Brook JR, Denburg J, et al. The Canadian Healthy Infant Longitudinal Development Birth Cohort Study: Biological Samples and Biobanking. Paediatr Perinat Epidemiol. 2015;29(1):84–92.
- Aryee MJ, Jaffe AE, Corrada-Bravo H, Ladd-Acosta C, Feinberg AP, Hansen KD, et al. Minfi: a Flexible and Comprehensive Bioconductor Package for the Analysis of Infinium DNA Methylation Microarrays. Bioinforma Oxf Engl. 2014;15(10):1363–9.
- Chen Y, Lemire M, Choufani S, Butcher DT, Grafodatskaya D, Zanke BW, et al. Discovery of cross-reactive probes and polymorphic CpGs in the Illumina Infinium HumanMethylation450 microarray. Epigenetics. 2013;8(2):203–9.
- Rahmani E, Zaitlen N, Baran Y, Eng C, Hu D, Galanter J, et al. Sparse PCA Corrects for Cell Type Heterogeneity in Epigenome-Wide Association Studies. Nat Methods. 2016;13(5):443–5.
- Rahmani E, Zaitlen N, Baran Y, Eng C, Hu D, Galanter J, et al. Correcting for Cell-Type Heterogeneity in DNA Methylation: a Comprehensive Evaluation. Nat Methods. 2017;28(3):218–9.
- Chen EY, Tan CM, Kou Y, Duan Q, Wang Z, Meirelles GV, et al. Enrichr: interactive and collaborative HTML5 gene list enrichment analysis tool. BMC Bioinformatics. 2013;14:128–8.
- Küpers LK, Monnereau C, Sharp GC, Yousefi P, Salas LA, Ghantous A, et al. Meta-Analysis of Epigenome-Wide Association Studies in Neonates Reveals Widespread Differential DNA Methylation Associated with Birthweight. Nat Commun. 2019;10(1):1893.
- Joubert BR, den Dekker HT, Felix JF, Bohlin J, Ligthart S, Beckett E, et al. Maternal Plasma Folate Impacts Differential DNA Methylation in an Epigenome-Wide Meta-Analysis of Newborns. Nat Commun. 2016;7:10577.
- Yeung EH, Guan W, Mumford SL, Silver RM, Zhang C, Tsai MY, et al. Measured Maternal Prepregnancy Anthropometry and Newborn DNA Methylation Epigenomics. 2019;11(2):187–98.
- Haertle L, El Hajj N, Dittrich M, Müller T, Nanda I, Lehnen H, et al. Epigenetic signatures of gestational diabetes mellitus on cord blood methylation. Clin Epigenetics. 2017;9:28.
- Joehanes R, Just AC, Marioni RE, Pilling LC, Reynolds LM, Mandaviya PR, et al. Epigenetic Signatures of Cigarette Smoking. Circ Cardiovasc Genet. 2016;9(5):436–47.
- 27. Fraser HB, Lam LL, Neumann SM, Kobor MS. Population-specificity of human DNA methylation. Genome Biol. 2012;13(2):R8.
- 28. Mozhui K, Smith AK, Tylavsky FA. Ancestry dependent DNA methylation and influence of maternal nutrition. PLoS ONE. 2015;10(3):e0118466.
- Chu S-K, Yang H-C. Interethnic DNA methylation difference and its implications in pharmacoepigenetics. Epigenomics. 2017;9(11):1437–54.

- Perry CA, Renna SA, Khitun E, Ortiz M, Moriarty DJ, Caudill MA. Ethnicity and Race Influence the Folate Status Response to Controlled Folate Intakes in Young Women. J Nutr. 2004;134(7):1786–92.
- Jeruszka-Bielak M, Isman C, Schroder TH, Li W, Green TJ, Lamers Y. South Asian Ethnicity Is Related to the Highest Risk of Vitamin B12 Deficiency in Pregnant Canadian Women. Nutrients. 2017;9(4):317. https://doi.org/10. 3390/nu9040317.
- Collier A-RY, Ledyard R, Montoya-Williams D, Qiu M, Dereix AE, Farrokhi MR, Hacker MR, Burris HH. Racial and ethnic representation in epigenomic studies of preterm birth: a systematic review. Epigenomics. 2021;13(21):1735-1746.
- Cronjé HT, Elliott HR, Nienaber-Rousseau C, Pieters M. Replication and Expansion of Epigenome-Wide Association Literature in a Black South African Population. Clin Epigenetics. 2020;12(1):6.
- Elliott HR, Tillin T, McArdle WL, Ho K, Duggirala A, Frayling TM, et al. Differences in Smoking Associated DNA Methylation Patterns in South Asians and Europeans. Clin Epigenetics. 2014;6(1):4–4.
- Adkins RM, Krushkal J, Tylavsky FA, Thomas F. Racial Differences in Gene-Specific DNA Methylation Levels are Present at Birth. Birt Defects Res A Clin Mol Teratol. 2011;91(8):728–36.
- Horvath S, Gurven M, Levine ME, Trumble BC, Kaplan H, Allayee H, et al. An epigenetic clock analysis of race/ethnicity, sex, and coronary heart disease. Genome Biol. 2016;17(1):171.
- Crimmins EM, Thyagarajan B, Levine ME, Weir DR, Faul J. Associations of Age, Sex, Race/Ethnicity, and Education With 13 Epigenetic Clocks in a Nationally Representative U.S. Sample: The Health and Retirement Study. J Gerontol Ser A. 2021 [cited 2021 Mar 5];(glab016). Available from: https://doi.org/10.1093/gerona/glab016.
- Tajuddin SM, Hernandez DG, Chen BH, Noren Hooten N, Mode NA, Nalls MA, et al. Novel Age-Associated DNA Methylation Changes and Epigenetic Age Acceleration in Middle-Aged African Americans and Whites. Clin Epigenetics. 2019;11(1):119.
- Aune D, Saugstad OD, Henriksen T, Tonstad S. Maternal body mass index and the risk of fetal death, stillbirth, and infant death: a systematic review and meta-analysis. JAMA. 2014;16(15):1536–46.
- Gaillard R, Steegers EAP, Duijts L, Felix JF, Hofman A, Franco OH, et al. Childhood cardiometabolic outcomes of maternal obesity during pregnancy: the Generation R Study. Hypertens Dallas Tex 1979. 2014;63(4):683–91.
- Oostvogels AJJM, Stronks K, Roseboom TJ, van der Post JAM, van Eijsden M, Vrijkotte TGM. Maternal prepregnancy BMI, offspring's early postnatal growth, and metabolic profile at age 5–6 years: the ABCD Study. J Clin Endocrinol Metab. 2014;99(10):3845–54.
- Hardikar AA, Satoor SN, Karandikar MS, Joglekar MV, Puranik AS, Wong W, et al. Multigenerational Undernutrition Increases Susceptibility to Obesity and Diabetes that Is Not Reversed after Dietary Recuperation. Cell Metab. 2015;22(2):312–9.
- Legoff L, D'Cruz SC, Tevosian S, Primig M, Smagulova F. Transgenerational Inheritance of Environmentally Induced Epigenetic Alterations during Mammalian Development. Cells. 2019;8(12):1559. https://doi.org/10. 3390/cells8121559.
- 44. de Souza RJ, Zulyniak MA, Stearns JC, Wahi G, Teo K, Gupta M, et al. The influence of maternal and infant nutrition on cardiometabolic traits: novel findings and future research directions from four Canadian birth cohort studies. Proc Nutr Soc. 2019;78(3):351–61.
- Gervin K, Salas LA, Bakulski KM, van Zelm MC, Koestler DC, Wiencke JK, et al. Systematic Evaluation and Validation of Reference and Library Selection Methods for Deconvolution of Cord Blood DNA Methylation Data. Clin Epigenetics. 2019;11(1):125.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

