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**Amino acid availability is not essential for lifespan extension by dietary restriction in
the fly**

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Abstract

Dietary restriction (DR) is one of the most potent ways to extend health- and lifespan. Key progress in understanding the mechanisms of DR, and ageing more generally, was made when dietary protein, and more specifically essential amino acids (EAA), were identified as the dietary component to restrict to obtain DR's health and lifespan benefits. This role of dietary amino acids has influenced work on ageing mechanisms, especially in nutrient sensing, e.g. Tor and insulin(-like) signalling networks. Experimental biology in *Drosophila melanogaster* has been instrumental in generating and confirming the hypothesis that EAA availability is important in ageing. Here, we expand on previous work testing the involvement of EAA in DR through large scale (N=6,238) supplementation experiments across four diets and two genotypes in female flies. Surprisingly, we find that EAA are not essential to DR's lifespan benefits. Importantly, we do identify the fecundity benefits of EAA supplementation suggesting the supplemented EAA were bioavailable. Furthermore, we find that the effects of amino acids on lifespan vary by diet and genetic line studied and that at our most restricted diet fecundity is constrained by other nutrients than EAA. We suggest that DR for optimal health is a concert of nutritional effects, orchestrated by genetic, dietary and other environmental interactions. Our results question the universal importance of amino acid availability in the biology of ageing and DR.

Keywords: ageing, reaction norm, drosophila, diet, nutrients

Introduction

A key step in understanding the mechanisms underlying dietary restriction (DR) is to identify which components of the diet cause increased longevity and decreased fecundity, and whether different nutrients are responsible for these related but potentially separate responses(1–3). When the precise nutrients that cause the health benefits of DR are understood, it is probable more precise molecular mechanisms of DR can be distilled. In addition, the health benefits of DR can potentially be separated from its side-effects such as those on reproduction, or it could be that these effects cannot be separated as they are so physiologically entwined(3–5).

Despite considerable research we still do not know the exact nutrient, or combination of nutrients, responsible for lifespan extension seen under DR(1,3,6,7). Even for one of the most tractable and studied model organisms in this regard, the fruit fly (*Drosophila melanogaster*), the precise effects of separate dietary components are far from fully elucidated(8–12). A growing line of research has, however, identified protein restriction as the causative route for lifespan extension in a wide range of species including flies(10,13–15). These experiments including those using the Geometric Framework, in which the ratio and amounts of macronutrients in semi- or fully defined diets are varied, demonstrated that the macronutrient protein is the main dietary axis determining lifespan. In flies, in which this paradigm has been most rigorously applied, experimental artefacts curtailing lifespan due to capillary feeding have been suggested. Yet, food delivery via vials, void of these artefacts, yielded similar conclusions(16,17).

The most conclusive evidence for protein as the main dietary component determining lifespan comes from supplementation studies of protein to a DR diet (in the form of casein) which nullified DR's longevity effect(13,18). One of the most influential studies in this field used a

range of supplementation studies to pinpoint restriction of essential amino acids (EAA) as responsible for the DR(5) ^{replicated in} (19). Supplementing the DR diet with EAAs led to an increase in fecundity and decrease in lifespan phenocopying the fully fed diet. Of these EAA, Methionine alone was shown to determine the fecundity response to DR, with lifespan being determined by a combination of Methionine and other EAA. This insight fits with work on rodents, in which restriction of Methionine can extend lifespan(20,21). In flies, restriction of Methionine extends lifespan, but only under conditions of low amino acid status(22). In mice, EAA supplementation nullified DR's lifespan extension(23) and restriction of particular amino-acids, namely branched-chained, can confer health and lifespan benefits(1,7,24).

The role of EAA in DR further fits with the identification of TOR (Target of Rapamycin) and IGF (Insulin-like Growth Factor) as important cellular signalling pathways in the molecular biology of ageing(25), as these pathways sense nutrient availability(1,26). There is thus a central line of literature within the biology of ageing field that implicates amino acid availability as a determinant of ageing(27,28). This concept has influenced how we view the mechanisms of ageing more generally. Although, in the fly, surprisingly, DR has additive benefits to lifespan on top of those gained by TOR suppression(29) and reduced insulin-like signalling(30). Moreover, it has been suggested using demographic analysis that TOR, somatotrophic signalling (e.g. IGF) and DR might use different mechanisms to extend lifespan(31,32). Recent insights implicating other nutrients in the DR reproduction and longevity response(8), the emerging criticism of the importance of dietary protein over calories in mice(33) and the demonstrated independent effects of sugar(11,34–37) whilst largely absent in the geometric framework(2,10,14), have now cast doubt on the universal importance of EAA in DR.

Here we report on an EAA supplementation experiment, using four different diets and two different genotypes. The diets used varied in both yeast and sugar content. We studied females only, the focal sex in the fly DR literature and exhibiting an additional relevant phenotype, namely egg laying, but recognise sex-specific effects of diet that warrant future study(10,38,39). Overall, we find that adding EAA had little effect on lifespan, but that small effects on lifespan of EAA depended on which diet it is supplemented to (note, previous work exclusively supplemented DR diets) and on the genetic line studied. In contrast, we did find the expected increase in fecundity when EAAs were supplemented, and these effects were similar across the two genetic lines studied. Our results question the universal importance of EAA in DR, by revisiting and expanding prior influential work on the fly in this area.

Results & Discussion

We tested the effect on lifespan of EAA addition across four diets, varying in yeast (protein) and sugar, in two genotypes. These diets represent ad libitum and restricted diets used in our laboratory(40,41) and in a principal study in the fly(5) that showed EAA restriction is essential for DR. The effects on lifespan of EAA addition were dependent on the genetic line tested and on the specific diet it was supplemented to (significant three-way interaction between diet, line and EAA addition, $\chi^2=11.8$, $df=4$, $P=0.019$, $N=6,238$). Within the *yw* genotype the direction and magnitude of the effect of EAA varied depending on the diet which it was supplemented to (interaction: $\chi^2=10.9$, $df=3$, $P=0.012$, $N=3,103$). The overall effect on lifespan of EAA addition was however only a fraction of that induced by diet, a log hazard of 0.19, compared to hazard differentials of -1.35 and 1.08 when yeast is manipulated in the diet inducing a DR pro-longevity response (Table S1). Within genotype *I95* the effect of EAA addition did not vary significantly with the diet it was supplemented to ($\chi^2=3.45$, $df=3$, $P=0.33$, $N=3,135$). Again, the effect of EAA on log hazard was modest (0.31 ± 0.13 ,

P=0.015) in comparison to the effects induced by varying yeast concentration, which induced a DR pro-longevity response (-0.66; 2.76, Table S1). It is evident therefore that the effects of EAA addition did not recapitulate the effects of DR (Figure 1), suggesting EAA are at best only partially responsible for the DR longevity responses we observed.

Even though we used the exact concentration and composition of EAA as was previously shown to explain DR in the fly(5) ^{replicated in (19)} a question could remain whether the supplementation had any substantial physiological impact in our experiments. The data collected on age-specific fecundity demonstrate, however, that egg laying is increased substantially with EAA supplementation. This effect, in contrast to the effect on lifespan, is similar in magnitude to that of varying yeast concentration in the diet (Figure 2). Overall egg laying was increased in both genotypes to a similar degree by amino-acid supplementation (*yw*: 1.11 ± 0.19 , $P < 0.0001$; *I95*: 0.89 ± 0.13 , $P = 0.003$, Figure 2), irrespective of the diet it was supplemented to (interaction diet by EAA addition, $P > 0.16$). For both genotypes analyses of the granularity of the patterns with age resulted in models including interactions of age with diet and with EAA supplementation, suggesting fecundity becomes differentially constrained with age depending on the nutritional environment (Table S2, Figure 2). Overall, these results suggest EAA supplementation fully rescues the loss of egg production when yeast concentration in the food is lowered, in contrast to the effects of yeast on lifespan.

Egg laying on the lowest yeast concentration (2%) was so low that counting via image analysis proved unreliable. Still, manual counting of eggs also revealed an increase in egg laying at this diet of lowest nutritional value when EAA were supplemented (Figure 3). This increase in egg laying was however modest and did not come near any of the egg production seen at the higher yeast diets (*yw*: 0.60 ± 0.10 ; *I95*: 0.41 ± 0.05 at 2%, Figure 3, versus at 8%, Figure 2, *yw*: 2.88 ± 0.19 ; *I95*: 2.37 ± 0.25). That egg production at these lowest yeast

conditions was not rescued by EAA supplementation suggests other nutrients (present in yeast) limit egg production at this dietary condition. Arguably an even higher EAA dosage could increase egg laying at these diets. However, egg laying is reduced by 5-fold when yeast concentration is lowered, whereas the maximum estimated available EAA at a higher yeast concentration is only 3.1 times higher than the pure EAA supplemented (Table S3).

Similarly, survival could reduce when a higher dose of EAA was supplemented, yet, the dietary modulation of log mortality hazard is 5.7 to 8.9 times that of EAA supplementation. Restriction of other nutrients than EAA therefore must explain a substantial part of the pro-longevity response of DR we observed.

Carbohydrates (sugar) are currently considered to play a marginal role in determining longevity under DR, at least when considered within the geometric framework(10,14). Often geometric framework studies are conducted on a single genotype only(40,41). We detect here a genotype specific effect of sugar on lifespan (Figure 1) that is interestingly accompanied by not a reduction but a non-significant increase in egg laying (Figure 2). Effects of sugar in flies have recently been attributed to water balance(34), but are less likely responsible for our results here, as we supplemented water to all experimental groups(41). As such, our results might mirror the effects of sugar on metabolic health as observed in mice(42).

Both genotypes tested also responded differently to EAA supplementation in terms of lifespan, but similarly in terms of egg production (Figure 1-3). Genetic differences in the response to DR(43) are of high interest, for translation⁴² and to understand the mechanisms of DR(44). Until now however such differences have not suggested a differential response to specific nutrients. Our results suggest there is the potential for genetic variance in how nutrients shape lifespan. We recognise only two lines were tested here. Our study was not set up to estimate genetic variance, but rather to replicate and expand prior work in this area.

Note, however, that the differences between lines we detect are in line with recent work in mice(38). Organisms should be viewed as a hierarchical set of physiological reaction norms(45) to a range of nutrients. DR becomes apparent when one of these reactions is limiting, and this need not be at the same level or the same nutrients for each genotype(41).

Conclusions

Previous work on the fly has been instrumental to shape the idea that dietary protein intake underlies the pro-health and pro-longevity benefits of DR across organisms, including our own species. Importantly, this idea has also shaped how EAA availability might determine longevity through altered nutrient signalling. Our results now suggest that amino acid availability does not explain DR universally. These results are in line with recent experiments in mice that question the dominance of dietary protein in determining longevity, albeit with observable benefits to health(7). Importantly, these insights now warrant a re-appreciation of how and in which circumstances specific nutrients, including amino acids, determine longevity and other key life history traits, such as reproduction. The metabolic networks that fuel bodily functions form a plastic network that shape phenotypic reactions to nutritional availability. The effects of nutrition on health are therefore probable to depend on genetic(38,40,41,44,46), dietary(3,47) and other environmental(48,49) interactions. Similar or differential pro-longevity physiology underlying DR could be triggered by differential nutritional interactions, and such could explain inconsistencies in our current understanding of DR and the mechanisms of ageing.

Figures

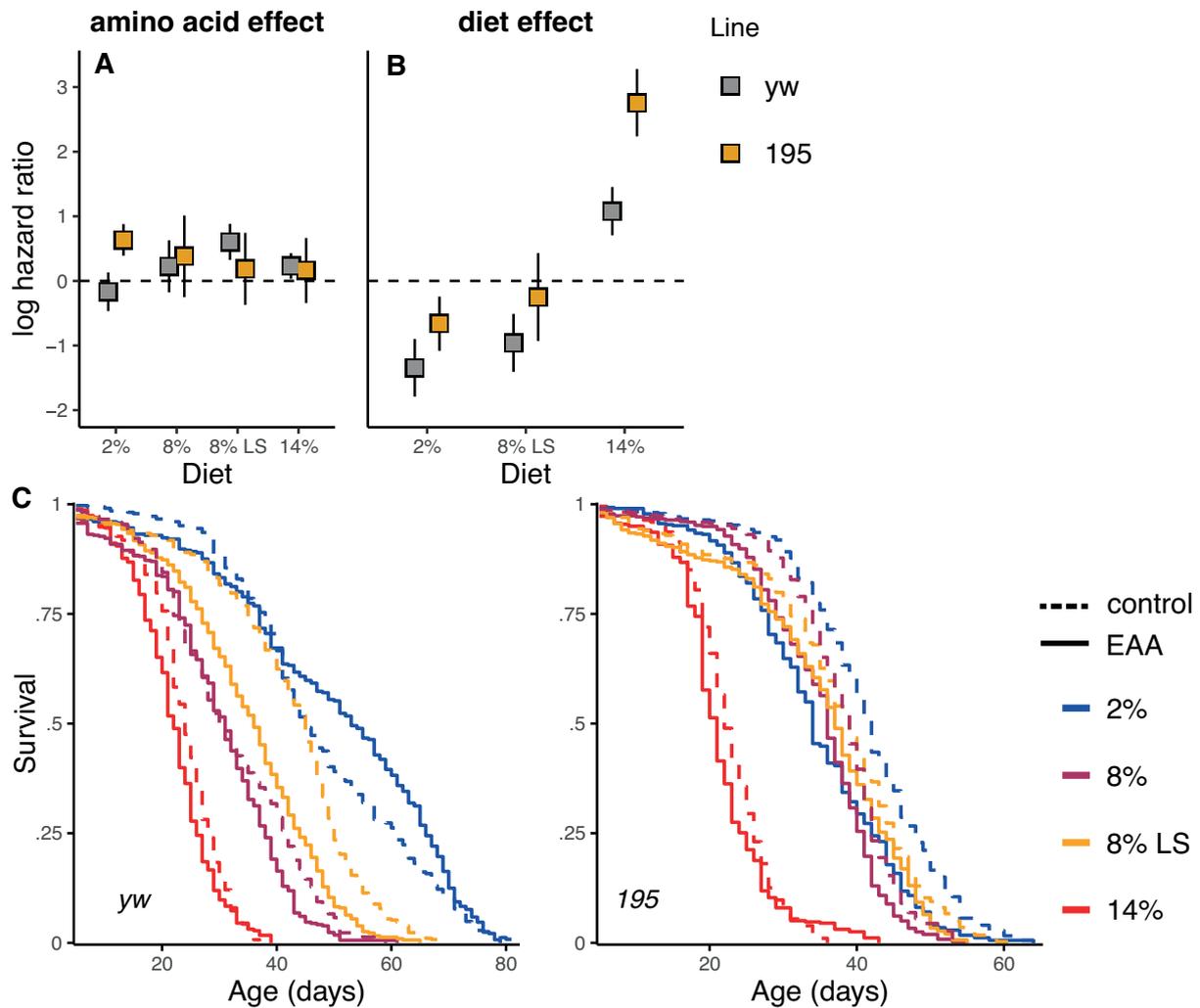


Figure 1. A, B: Modulation of lifespan by EAA supplementation is small in comparison to diet, but varies by genotype. **A:** Log hazard ratios and their 95% CIs for both genotypes associated with amino-acid supplementation per diet (left, reference is control), and overall diet effects (right, reference 8%). Hazard ratios provide a quantitative estimate of risk across circumstances that are directly comparable. Note that higher hazard ratios indicate higher risk and that the logarithm is plotted. **C:** Survival curves underlying the hazard estimates. Colour indicates the diets used with solid lines indicating amino acid supplementation, dash lines indicate control. Clearly EAA supplementation did not revert the lifespan gains achieved by modulating dietary yeast concentration.

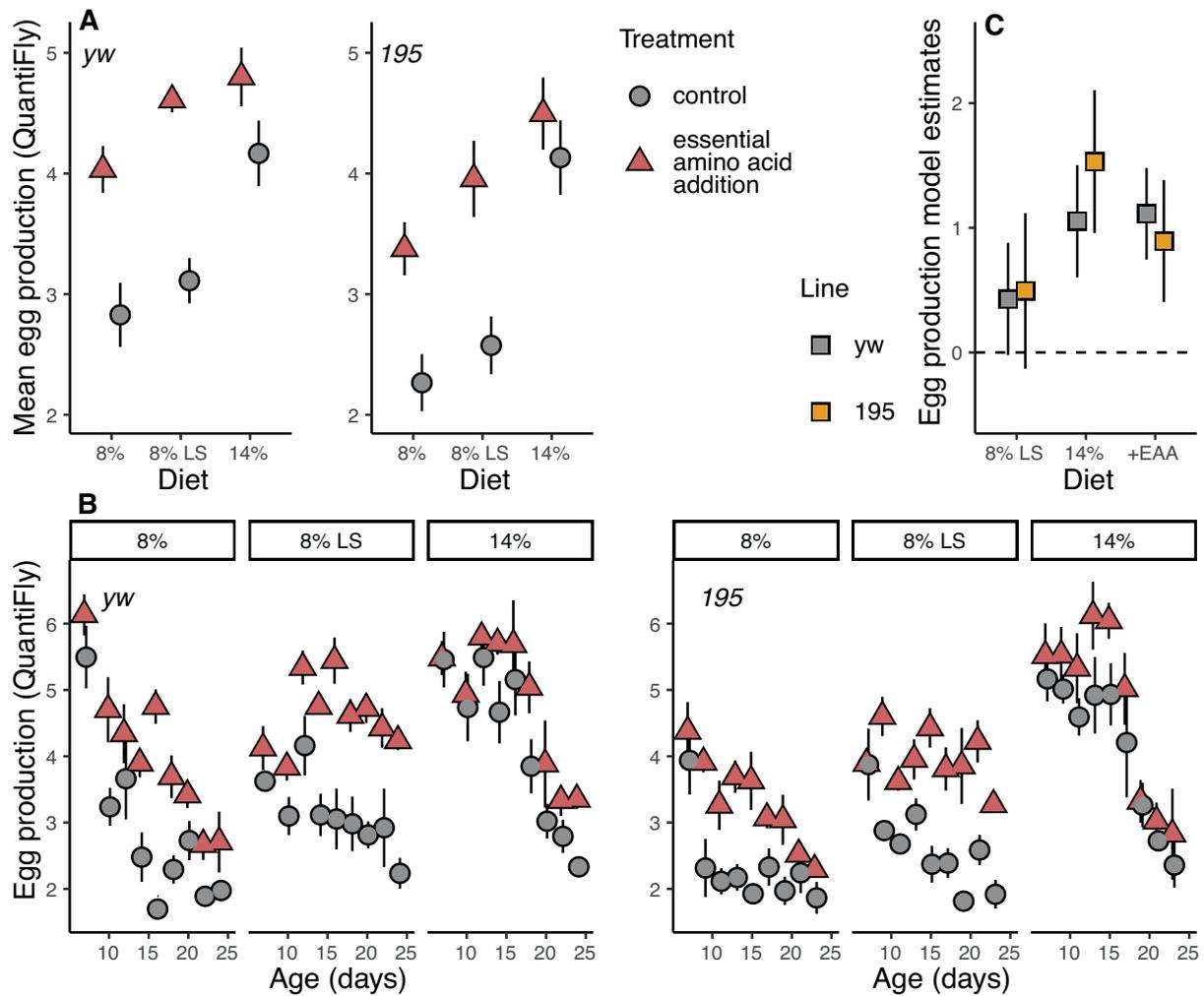


Figure 2. A: Mean egg production per fly (across 2 days) as measured using Quantifly. EAA supplementation increased egg laying on all diets and mimicked the fecundity gain from increased dietary yeast. **B:** Age-specific fecundity plots per line per diet. Supplementation effects on fecundity vary by line, diet and age. **C:** Model estimates analogous to log hazard ratio plots, clearly showing a similar effect of EAA supplementation and increased yeast concentration (14%) on egg laying.

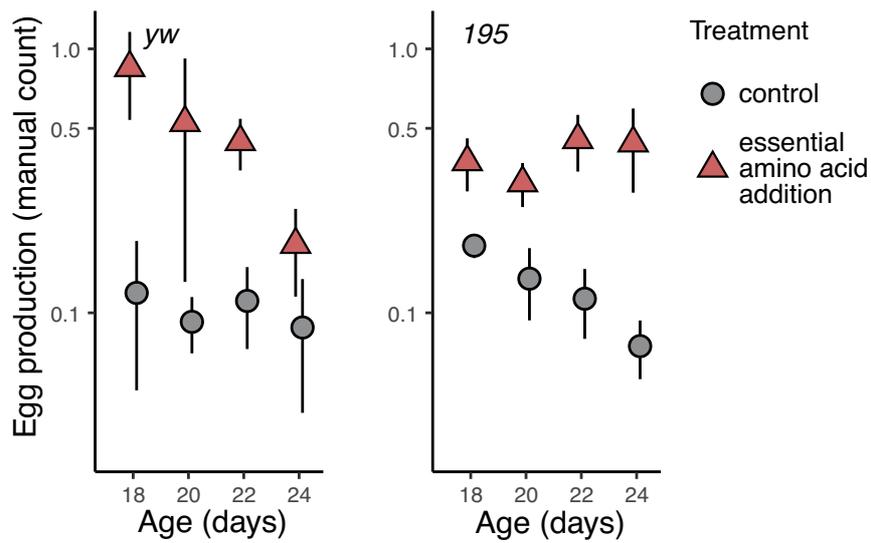


Figure 3. Mean egg production per fly on the lowest yeast diets (2%) from manual counts (across 2 days). EAA supplementation increased egg production but egg laying remains far lower than fecundity seen at the higher yeast diets (Figure 2), suggesting egg production is limited by other nutrients than EAA at this diet.

Methods

Experiments used two genetic lines of *Drosophila melanogaster* - *195* from the DGRP⁴⁹, and the yellow white (*yw*) lab strain. All flies were cultured on rich yeast media(40) (8% autolysed yeast, 13% table sugar, 6% cornmeal, 1% agar, 0.225% nipagin and 0.4% propanoic acid). Cooked fly media was stored for up to two weeks at 4-6°C, and warmed to 25°C before use. Experimental diets consisted of 3% cornmeal, 1% agar and 0.225% nipagin, with dietary yeast and sugar varied to create the following experimental diets: low protein (2% yeast + 13% sugar), high protein (8% yeast + 13% sugar), high protein, low sugar (8% yeast + 5% sugar) and very high protein, low sugar (14% yeast + 5% sugar). The latter two diets are similar to the DR and fully-fed diets used in work that showed EAA are fundamental to DR⁵. Each of these diets was cooked with and without added essential amino acids, at the concentrations as in previous work(5,19) (Table S3). Experiments were conducted in a climate-controlled environment with a 12:12 hour light-dark cycle, temperature at 25°C and 50-60% relative humidity.

Longevity

Flies were grown in bottles on rich media and incubated at 25°C. In each of these bottles, 10 females (with 2 males) were allowed to lay eggs for 3 days. Bottles were given water daily if media appeared dry during larval development. When offspring began to eclose, individuals were transferred to mating bottles, where they were left to mate for ~48 hours. This was repeated every day until all flies had eclosed to generate age-matched cohorts. Flies were sorted under carbon dioxide anaesthesia (Flystuff Flowbuddy; <5L/min), and females were transferred to demography cages, specially designed to allow removal of deceased flies, and changing of food vial with minimal disturbance to living flies(40,50). As in extreme conditions, and in some genotypes water availability can confound dietary effects(41) all flies

were supplemented with a vial of water agar ^{as in} (41). Censusing was conducted every two days, with the food and water vial changed each time. Living flies which were stuck to the food vial or died as a result of sticking to the food or becoming trapped in any part of the cage, and flies which escaped, were right-censored.

Fecundity

After 6 days of being on the experimental diets, vials from demography cages were photographed using a webcam under custom LED-lighting and images were analysed using QuantiFly(51); machine learning software for automated image analysis for egg counting. Vials from cages on 2% yeast were counted manually using a dissection microscope, when it became apparent egg counts were too low to be reliably analysed with the QuantiFly setup.

Statistical analysis

Lifespan data were analysed using time-to-event mixed-effects Cox proportional hazard models, with cage as a random term, implemented in ‘coxme’ in R(52). Full models and comparisons with and without interaction terms between the dietary treatments (supplementation and diet coded as two separate independent variables) and line were used to test the main hypotheses. Estimates of individual models ran within each line and within each diet are presented as these will be less sensitive to deviations from proportionality of hazards and provide the best estimates of individual effects. Egg laying was analysed using linear mixed effects models in ‘lmer’(53) in R, with cage as random term. Models on age-specific fecundity were simplified using backward selection, using the step function from ‘lmerTest’(54). Egg counts were divided by the total flies in the cage at the time of fecundity measurement to correct for any differences in mortality, although uncorrected results yielded qualitatively similar results. In text, \pm indicates standard error.

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Reference List

1. Babygirija R, Lamming DW. The regulation of healthspan and lifespan by dietary amino acids. *Transl Med Aging*. 2021;5:17-30. doi:10.1016/j.tma.2021.05.001.
2. Solon-Biet SM, Wahl D, Raubenheimer D, Cogger VC, Le Couteur DG, Simpson SJ. The geometric framework: An approach for studying the impact of nutrition on healthy aging. *Drug Discov Today Dis Model*. 2018;27:61-68. doi:10.1016/j.ddmod.2019.03.002.
3. Wali JA, Milner AJ, Luk AWS, et al. Impact of dietary carbohydrate type and protein-carbohydrate interaction on metabolic health. *Nat Metab*. June 2021:1-19. doi:10.1038/s42255-021-00393-9.
4. Hoedjes KM, Rodrigues MA, Flatt T. Amino acid modulation of lifespan and reproduction in *Drosophila*. *Curr Opin Insect Sci*. 2017;23:118-122. doi:10.1016/j.cois.2017.07.005.
5. Grandison RC, Piper MDW, Partridge L. Amino-acid imbalance explains extension of lifespan by dietary restriction in *Drosophila*. *Nature*. 2009;462(7276):1061-1064. doi:10.1038/nature08619.
6. Yap YW, Rusu PM, Chan AY, et al. Restriction of essential amino acids dictates the systemic metabolic response to dietary protein dilution. *Nat Commun*. 2020;11(1):1-13. doi:10.1038/s41467-020-16568-z.
7. Richardson NE, Konon EN, Schuster HS, et al. Lifelong restriction of dietary branched-chain amino acids has sex-specific benefits for frailty and life span in mice. *Nat Aging*. 2021;1(1):73-86. doi:10.1038/s43587-020-00006-2.

8. Zanco B, Mirth CK, Sgrò CM, Piper MDW. A dietary sterol trade-off determines lifespan responses to dietary restriction in *Drosophila melanogaster* females. *Elife*. 2021;10:1-20. doi:10.7554/eLife.62335.
9. Tatar M, Post S, Yu K. Nutrient control of *Drosophila* longevity. *Trends Endocrinol Metab*. 2014;25(10):509-517. doi:10.1016/j.tem.2014.02.006.
10. Jensen K, McClure C, Priest NK, Hunt J. Sex-specific effects of protein and carbohydrate intake on reproduction but not lifespan in *Drosophila melanogaster*. *Aging Cell*. 2015;14(4):605-615. doi:10.1111/accel.12333.
11. Troen AM, French EE, Roberts JF, et al. Lifespan modification by glucose and methionine in *Drosophila melanogaster* fed a chemically defined diet. *Age (Omaha)*. 2007;29(1):29-39. doi:10.1007/s11357-006-9018-4.
12. Krittika S, Yadav P. An overview of two decades of diet restriction studies using *Drosophila*. *Biogerontology*. 2019;20(6):723-740. doi:10.1007/s10522-019-09827-0.
13. Min KJ, Tatar M. *Drosophila* diet restriction in practice: Do flies consume fewer nutrients? *Mech Ageing Dev*. 2006;127(1):93-96. doi:10.1016/j.mad.2005.09.004.
14. Lee KP, Simpson SJ, Clissold FJ, et al. Lifespan and reproduction in *Drosophila*: New insights from nutritional geometry. *Proc Natl Acad Sci*. 2008;105(7):2498-2503. doi:10.1073/pnas.0710787105.
15. Mair W, Piper MDW, Partridge L. Calories do not explain extension of life span by dietary restriction in *Drosophila*. *PLoS Biol*. 2005;3(7):1305-1311.

doi:10.1371/journal.pbio.0030223.

16. Lee KP. Dietary protein: Carbohydrate balance is a critical modulator of lifespan and reproduction in *Drosophila melanogaster*: A test using a chemically defined diet. *J Insect Physiol.* 2015;75:12-19. doi:10.1016/j.jinsphys.2015.02.007.
17. Kim K, Jang T, Min KJ, Lee KP. Effects of dietary protein:carbohydrate balance on life-history traits in six laboratory strains of *Drosophila melanogaster*. *Entomol Exp Appl.* 2020;168(6-7):482-491. doi:10.1111/eea.12855.
18. Bruce KD, Hoxha S, Carvalho GB, et al. High carbohydrate–low protein consumption maximizes *Drosophila* lifespan. *Exp Gerontol.* 2013;48(10):1129-1135. doi:10.1016/j.exger.2013.02.003.
19. Emran S, Yang M, He X, Zandveld J, Piper MDW. Target of rapamycin signalling mediates the lifespan-extending effects of dietary restriction by essential amino acid alteration. *Aging (Albany NY).* 2014;6(5):390-398. doi:10.18632/aging.100665.
20. Miller RA, Buehner G, Chang Y, Harper JM, Sigler R, Smith-Wheelock M. Methionine-deficient diet extends mouse lifespan, slows immune and lens aging, alters glucose, T4, IGF-I and insulin levels, and increases hepatocyte MIF levels and stress resistance. *Aging Cell.* 2005;4(3):119-125. doi:10.1111/j.1474-9726.2005.00152.x.
21. Zimmerman J. Nutritional control of aging. *Exp Gerontol.* 2003;38(1-2):47-52. doi:10.1016/S0531-5565(02)00149-3.
22. Lee BC, Kaya A, Ma S, et al. Methionine restriction extends lifespan of *Drosophila melanogaster* under conditions of low amino-acid status. *Nat Commun.*

2014;5:3592. doi:10.1038/ncomms4592.

23. Yoshida S, Yamahara K, Kume S, et al. Role of dietary amino acid balance in diet restriction-mediated lifespan extension, renoprotection, and muscle weakness in aged mice. *Aging Cell*. 2018;17(4):12796. doi:10.1111/acer.12796.

24. Fontana L, Cummings NE, Arriola Apelo SI, et al. Decreased Consumption of Branched-Chain Amino Acids Improves Metabolic Health. *Cell Rep*. 2016;16(2):520-530. doi:10.1016/j.celrep.2016.05.092.

25. Johnson SC. Nutrient sensing, signaling and ageing: The role of IGF-1 and mTOR in ageing and age-related disease. In: *Subcellular Biochemistry*. Vol 90. Springer New York; 2018:49-97. doi:10.1007/978-981-13-2835-0_3.

26. Shimobayashi M, Hall MN. Multiple amino acid sensing inputs to mTORC1. *Cell Res*. 2016;26(1):7-20. doi:10.1038/cr.2015.146.

27. Canfield CA, Bradshaw PC. Amino acids in the regulation of aging and aging-related diseases. *Transl Med Aging*. 2019;3:70-89. doi:10.1016/j.tma.2019.09.001.

28. Mirzaei H, Suarez JA, Longo VD. Protein and amino acid restriction, aging and disease: From yeast to humans. *Trends Endocrinol Metab*. 2014;25(11):558-566. doi:10.1016/j.tem.2014.07.002.

29. Bjedov I, Toivonen JM, Kerr F, et al. Mechanisms of Life Span Extension by Rapamycin in the Fruit Fly *Drosophila melanogaster*. *Cell Metab*. 2010;11(1):35-46. doi:10.1016/j.cmet.2009.11.010.

30. Min KJ, Yamamoto R, Buch S, Pankratz M, Tatar M. *Drosophila* lifespan control by dietary restriction independent of insulin-like signaling. *Aging Cell*.

2008;7(2):199-206. doi:10.1111/j.1474-9726.2008.00373.x.

31. Garratt M, Nakagawa S, Simons MJP. Life-span Extension With Reduced Somatotrophic Signaling: Moderation of Aging Effect by Signal Type, Sex, and Experimental Cohort. *Journals Gerontol Ser A*. 2017;72(12):1620-1626. doi:10.1093/gerona/glx010.

32. Garratt M, Nakagawa S, Simons MJPMJP. Comparative idiosyncrasies in life extension by reduced mTOR signalling and its distinctiveness from dietary restriction. *Aging Cell*. 2016;15(4):737-743. doi:10.1111/accel.12489.

33. Speakman JR, Mitchell SE, Mazidi M. Calories or protein? The effect of dietary restriction on lifespan in rodents is explained by calories alone. *Exp Gerontol*. 2016;86:28-38. doi:10.1016/j.exger.2016.03.011.

34. van Dam E, van Leeuwen LAG, dos Santos E, et al. Sugar-Induced Obesity and Insulin Resistance Are Uncoupled from Shortened Survival in Drosophila. *Cell Metab*. 2020;31(4):710-725.e7. doi:10.1016/j.cmet.2020.02.016.

35. Chandegra B, Tang JLY, Chi H, Alic N. Sexually dimorphic effects of dietary sugar on lifespan, feeding and starvation resistance in Drosophila. *Aging (Albany NY)*. 2017;9(12):2521-2528. doi:10.18632/aging.101335.

36. Keipert S, Voigt A, Klaus S. Dietary effects on body composition, glucose metabolism, and longevity are modulated by skeletal muscle mitochondrial uncoupling in mice. *Aging Cell*. 2011;10(1):122-136. doi:10.1111/J.1474-9726.2010.00648.X.

37. Schulz TJ, Zarse K, Voigt A, Urban N, Birringer M, Ristow M. Glucose Restriction Extends Caenorhabditis elegans Life Span by Inducing Mitochondrial

- Respiration and Increasing Oxidative Stress. *Cell Metab.* 2007;6(4):280-293.
doi:10.1016/J.CMET.2007.08.011.
38. Green CL, Pak HH, Richardson NE, et al. Sex and genetic background define the metabolic, physiologic, and molecular response to protein restriction. *Cell Metab.* 2022;34(2):209-226.e5. doi:10.1016/J.CMET.2021.12.018.
39. Wu Q, Yu G, Cheng X, et al. Sexual dimorphism in the nutritional requirement for adult lifespan in *Drosophila melanogaster*. *Aging Cell.* 2020;19(3):e13120. doi:10.1111/ACEL.13120.
40. McCracken AW, Adams G, Hartshorne L, Tatar M, Simons MJ. The hidden costs of dietary restriction: Implications for its evolutionary and mechanistic origins. *Sci Adv.* 2020;6(8):eaay3047. doi:10.1126/sciadv.aay3047.
41. McCracken AW, Buckle E, Simons MJ. The relationship between longevity and diet is genotype dependent and sensitive to desiccation in *Drosophila melanogaster*. *J Exp Biol.* 2020;223(23):jeb.230185. doi:10.1242/jeb.230185.
42. Andres-Hernando A, Orlicky DJ, Kuwabara M, et al. Deletion of Fructokinase in the Liver or in the Intestine Reveals Differential Effects on Sugar-Induced Metabolic Dysfunction. *Cell Metab.* 2020;32(1):117-127.e3.
doi:10.1016/J.CMET.2020.05.012.
43. Unnikrishnan A, Matyi S, Garrett K, et al. A Reevaluation of the Effect of Dietary Restriction on Different Recombinant Inbred (RI) Lines of Male and Female Mice. *bioRxiv.* 2021. doi:10.1101/2021.06.25.449984.
44. Jin K, Wilson KA, Beck JN, et al. Genetic and metabolomic architecture of

variation in diet restriction-mediated lifespan extension in *Drosophila*. *PLoS Genet.* 2020;16(7):e1008835. doi:10.1371/journal.pgen.1008835.

45. Flatt T. Plasticity of lifespan: A reaction norm perspective. *Proc Nutr Soc.* 2014;73(4):532-542. doi:10.1017/S0029665114001141.

46. Dick KB, Ross CR, Yampolsky LY. Genetic variation of dietary restriction and the effects of nutrient-free water and amino acid supplements on lifespan and fecundity of *Drosophila*. *Genet Res (Camb)*. 2011;93(4):265-273. doi:10.1017/S001667231100019X.

47. Ma C, Mirth CK, Hall MD, Piper MDW. Amino acid quality modifies the quantitative availability of protein for reproduction in *Drosophila melanogaster*. *J Insect Physiol.* March 2020:104050. doi:10.1016/j.jinsphys.2020.104050.

48. Adler MI, Cassidy EJ, Fricke C, Bonduriansky R. The lifespan-reproduction trade-off under dietary restriction is sex-specific and context-dependent. *Exp Gerontol.* 2013;48(6):539-548. doi:10.1016/j.exger.2013.03.007.

49. Zajitschek F, Zajitschek SRK, Friberg U, Maklakov AA. Interactive effects of sex, social environment, dietary restriction, and methionine on survival and reproduction in fruit flies. *Age (Omaha)*. 2013;35(4):1193-1204. doi:10.1007/s11357-012-9445-3.

50. Good TP, Tatar M. Age-specific mortality and reproduction respond to adult dietary restriction in *Drosophila melanogaster*. *J Insect Physiol.* 2001;47(12):1467-1473. doi:10.1016/S0022-1910(01)00138-X.

51. Waithe D, Rennert P, Brostow G, Piper MDW. QuantiFly: Robust trainable

software for automated *Drosophila* egg counting. *PLoS One*. 2015;10(5):1-16.

doi:10.1371/journal.pone.0127659.

52. Therneau TM, Grambsch PM, Shane Pankratz V, Shane PANKRATZ V.

Penalized Survival Models and Frailty. *J Comput Graph Stat*. 2003;12(1):156-175.

doi:10.1198/1061860031365.

53. Bates D, Mächler M, Bolker BM, Walker SC. Fitting linear mixed-effects

models using lme4. *J Stat Softw*. 2015;67(1):1-48. doi:10.18637/jss.v067.i01.

54. Kuznetsova A, Brockhoff PB, Christensen RHB. lmerTest Package: Tests in

Linear Mixed Effects Models . *J Stat Softw*. 2017;82(13):1-26.

doi:10.18637/jss.v082.i13.