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Trends in Microbiology

Ecological and evolutionary solutions to the plasmid paradox

--Manuscript Draft--

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Abstract:	<p>Plasmids are common features of bacterial genomes, but theoretically they should not exist: Due to the costs associated with plasmid maintenance, non-beneficial plasmids should be purged by negative selection, whereas even under positive selection plasmid-encoded beneficial genes should be captured to the bacterial chromosome, followed by loss of the redundant plasmid. In the decade since we described this apparent “plasmid paradox” a range of ecological and evolutionary solutions have been shown to operate in bacterial populations and communities, explaining the widespread distribution and stable persistence of plasmids. We conclude, therefore, that the theoretical plasmid paradox has now been solved. The current challenge for the field, however, is to better understand how these solutions operate in natural bacterial communities to explain and predict the dynamics and distributions of plasmids and the horizontal gene transfer that they mediate in bacterial (pan)genomes.</p>
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1 **Ecological and evolutionary solutions to the plasmid paradox**

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8 **Keywords:** plasmid; mobile genetic element; horizontal gene transfer; pangenome;
9 accessory genome

10

11 **Abstract**

12 Plasmids are common features of bacterial genomes, but theoretically they should not
13 exist: Due to the costs associated with plasmid maintenance, non-beneficial plasmids
14 should be purged by negative selection, whereas even under positive selection plasmid-
15 encoded beneficial genes should be captured to the bacterial chromosome, followed by
16 loss of the redundant plasmid. In the decade since we described this apparent “plasmid
17 paradox” a range of ecological and evolutionary solutions have been shown to operate
18 in bacterial populations and communities, explaining the widespread distribution and
19 stable persistence of plasmids. We conclude, therefore, that the theoretical plasmid
20 paradox has now been solved. The current challenge for the field, however, is to better
21 understand how these solutions operate in natural bacterial communities to explain and
22 predict the dynamics and distributions of plasmids and the horizontal gene transfer that
23 they mediate in bacterial (pan)genomes.

24 **Introduction**

25 Plasmids are (usually circular) extrachromosomal genetic elements that encode their
26 own replication, maintenance and, in the case of conjugative plasmids, the conjugative
27 pilus that enables horizontal transfer of the plasmid to other cells [1]. In addition to
28 encoding these plasmid-related functions, plasmids often also carry a cargo of
29 accessory genes that confer a wide range of ecological traits upon the bacterial host,
30 including resistance to stressors and metabolic capacities [2]. As such, plasmids play a
31 key role in bacterial evolution by transferring these ecologically important genes
32 between lineages [3].

33

34 Although plasmids are common features of bacterial genomes, theoretically they should
35 not exist, a situation described as the “plasmid paradox” [4]: Plasmids place a metabolic
36 burden upon host cells for their maintenance and expression and often cause other
37 types of cellular disruptions leading to fitness costs (summarised in [5,6]).

38 Consequently, plasmids should be purged from bacterial populations by negative
39 selection [7]. Yet even when plasmid encoded accessory functions are sufficiently
40 beneficial to outweigh these costs (and thus the encoding plasmid is positively
41 selected), theory predicts that the accessory genes should be captured to the bacterial
42 chromosome, thereby allowing loss of the redundant plasmid [7]. Addiction systems,
43 such as toxin-antitoxin systems, are also no guarantee of long-term survival due the
44 possibility of these systems also being captured by the chromosome, enabling loss of
45 the plasmid without suffering ill effects of the toxin [8].

46

47 Over the past decade, new theory and experiments have shown that a range of
48 solutions to the plasmid paradox operate in bacterial populations and communities
49 (Figure 1). In this review, we summarise the ecological and evolutionary solutions
50 explaining the widespread existence and stable maintenance of plasmids in bacterial
51 genomes.

52

53 **Ecological solutions**

54 *Solution 1: Infectious transmission*

55 Rather than persisting long term in any given genome, plasmids may potentially survive
56 at the population level by horizontal transmission, passing from cell to cell as an
57 infectious element [9]. Early mathematical models of plasmid-host dynamics usually
58 assumed, based upon the available data [10], that rates of plasmid conjugation were too
59 low to sustain plasmid maintenance by infectious transmission [7]. However, more
60 recent studies have shown that some plasmids have conjugation rates that are
61 sufficiently high to enable their persistence in bacterial populations even in the face of
62 high fitness costs and, consequently, strong negative selection. For example, Lopatkin
63 *et al* [11] showed conjugation mediated persistence of nine antibiotic resistance
64 plasmids from six major incompatibility groups in experimental *Escherichia coli*
65 populations in the absence of antibiotics.

66

67 Even more remarkably, the environmental mercury resistance plasmid, pQBR57, was
68 capable of driving itself to fixation in experimental *Pseudomonas fluorescens*
69 populations through infectious transmission in both liquid broth [12] and soil habitats

70 [13] without mercury selection. Notably, similar dynamics were not observed in a
71 congeneric host, *Pseudomonas putida*, where pQBR57 was less costly but went extinct
72 due to a lower conjugation rate in this host [13]. In environments that experience
73 occasional pulses of positive selection, even low rates of conjugation can enhance
74 plasmid survival by enabling low level persistence during the periods between pulses,
75 which then select for clonal expansion of plasmid carriers [14].

76

77 Higher rates of conjugative horizontal transmission are required to enable plasmid
78 persistence as the fitness costs of plasmid carriage increase [15]. In addition, other
79 sources of selection on bacterial populations can increase the relative importance of
80 conjugation for plasmid survival. For example, predation by phages can limit plasmid
81 persistence by reducing bacterial density and thus the opportunity for conjugation [16].
82 Similarly, conjugation was required for the persistence of an antibiotic resistance
83 plasmid in bacterial populations that were being predated by a protist [17].

84

85 Infectious transmission through conjugation can, of course, only explain the persistence
86 of the fraction of plasmids that are self-transferable and encode their own conjugative
87 apparatus. Plasmids that are not self-transferable must either rely on other plasmids to
88 mobilize them [18], or on other mechanisms of cell-to-cell transmission, such as
89 packaging and transfer of plasmid DNA by phages [19] or natural transformation [20].
90 The relative contribution of these alternative transfer mechanisms to the persistence of
91 non-self-transferable plasmids versus the other stability mechanisms we discuss below
92 is unknown.

93

94 *Solution 2: Plasmid properties vary across host backgrounds*

95 Models of plasmid persistence typically assume that plasmid properties are inherent to
96 the plasmid. However, key properties that determine plasmid persistence are
97 increasingly being shown to vary between bacterial species and even between closely
98 related conspecific bacterial strains. Plasmid segregation [21] and conjugation rates
99 [22–25] vary extensively across genomic backgrounds. The causes of this variation in
100 plasmid properties are largely unknown, but recent studies suggest that this variability
101 among coexisting bacterial strains could affect plasmid stability in bacterial
102 communities.

103

104 Using hospital isolates of *E. coli* and *Klebsiella pneumoniae* and a co-circulating
105 antibiotic resistance plasmid, [23] have shown that the fitness effects of plasmid
106 acquisition were on average costly but ranged from negative to positive effects on
107 bacterial fitness among strains in the absence of antibiotics. A similar pattern has been
108 independently observed in a diverse collection of *E. coli* environmental and clinical
109 isolates for an unrelated antibiotic resistance plasmid [22]. Combining their plasmid
110 fitness effects data with a simple mathematical model, Alonso-Del Valle *et al* [23]
111 showed that introducing variable plasmid fitness effects among strains increased the
112 stability of low cost plasmids. In effect, the variability of plasmid fitness effects meant
113 that in some hosts, the plasmid causes no fitness cost (and indeed may increase
114 fitness), correspondingly reducing the dependence upon infectious transmission for
115 plasmid survival. Interestingly, this stabilising effect of variable plasmid fitness costs

116 strengthens with increasing bacterial community diversity. Variable fitness effects
117 cannot, however, explain the persistence of high cost plasmids. Where mean fitness
118 costs are high, the distribution of plasmid fitness effects may result in fewer geotypes
119 able to serve as permissible hosts, and thus high cost plasmids are still predicted to
120 critically rely on infectious transmission for their maintenance [23]. Nevertheless,
121 variation in fitness effects may allow certain host genotypes to act as refugia for
122 plasmids, enabling their maintenance in the community.

123

124 *Solution 3: Interactions with other plasmids*

125 Plasmid persistence can also be affected by the presence of other plasmids in the same
126 genome. Coinfecting plasmids can affect transfer efficiency through mobilisation and co-
127 integration [18,26–28] of both non-conjugative [29] and conjugative [30] plasmids.

128

129 Using plasmid coinfection experiments, San Millan *et al* [29] showed that for 5 out of 6
130 plasmids their fitness cost was lower in coinfection than expected from the cost of each
131 plasmid when measured alone. This is an example of synergistic epistasis, and could
132 lead to improved plasmid persistence through coinfection by weakening negative
133 selection. This may help to explain why bacterial genomes contain multiple plasmids
134 more often than is expected by chance [29].

135

136 A similar pattern of synergistic epistasis of plasmid fitness costs has been observed for
137 two large conjugative plasmids [30]. However, this study also suggested that finding
138 synergistic epistasis was somewhat dependent upon how fitness was measured, such

139 that it was more likely to be observed when competing plasmid bearing cells against
140 plasmid free cells, than when coinfecting cells were competed against single-infected
141 cells. Nevertheless, a recent theoretical study modelling how plasmid coinfection
142 affected stability found that epistatic interactions in particular are likely to determine the
143 persistence of plasmids in bacterial populations, while variation in other plasmid
144 properties (such as conjugation and segregation rates) were less important [31]. Thus,
145 more experimental tests across a wider range of plasmid-host combinations are
146 urgently needed to test the generality of the role for plasmid-plasmid interactions in
147 determining plasmid stability.

148

149 *Solution 4: Source-sink spillover transmission*

150 In bacterial communities containing multiple species that vary in their proficiency as
151 plasmid hosts, plasmids can persist in species that would otherwise be incapable of
152 sustaining them due to spillover transmission from other more proficient host species
153 (i.e., those species that do stably maintain the plasmid within their own population). In
154 soil microcosms, whereas the mercury resistance plasmid pQBR57 went extinct in *P.*
155 *putida* populations without mercury selection, the plasmid was consistently detected at
156 appreciable levels in *P. putida* when grown in a community alongside a more proficient
157 host, *P. fluorescens*, which acted as a source of plasmid spillover transmission [13].

158

159 Experiments in more complex communities by Cairns *et al* [32] suggest that spillover
160 transmission of an antibiotic resistance plasmid occurred preferentially to higher
161 abundance taxa and close phylogenetic relatives of the donor species. Interestingly,

162 transmission to lower abundance taxa was enhanced in spatially structured
163 environments, presumably by increasing cell-to-cell contacts and thus opportunities for
164 conjugation. Moreover, by increasing plasmid abundance, low antibiotic concentrations
165 increased the diversity of bacterial taxa acquiring the plasmid.

166

167 The impact of spillover transmission on plasmid spread in communities varies with the
168 plasmid donor species, such that plasmids reach higher community-level abundances
169 when introduced by a proficient plasmid host [33]. Conversely, plasmid transmission
170 within a given species' population can be impaired when living in a community
171 alongside less proficient plasmid hosts due to the dilution effect [26], which could
172 potentially destabilise the plasmid if efficient infectious transmission is required for its
173 stability in the source host population.

174

175 Spillover from proficient plasmid hosts into less proficient host species could increase
176 the probability of plasmids adapting to better persist in less proficient host species,
177 similar to host shifts in infectious disease [34,35]. For example by reducing their fitness
178 cost in this new host or enhancing their conjugation rate from this host [36], thus
179 broadening the host range of the plasmid.

180

181 **Evolutionary solutions**

182 *Solution 5: Compensatory evolution*

183 The fitness costs of acquiring a plasmid can be overcome by compensatory evolution to
184 ameliorate the cost and thus reduce or negate negative selection on the plasmid.

185 Compensatory mutations have been observed to evolve repeatedly in laboratory
186 populations of diverse bacteria-plasmid associations [8,30,37–43]. Compensatory
187 mutations can occur on the plasmid [40,41] or the chromosome [30,37–39,43] or on
188 both [8,43]. Theoretically, plasmid encoded compensatory mutations should be more
189 successful if they also reduce plasmid costs in transconjugant cells as the benefits are
190 carried in linkage with plasmid transfer [44]. However, chromosomal compensatory
191 mutations appear to be more commonly observed across the studies performed to date
192 [8,30,37–39]. This may simply be a consequence of the chromosome being larger and
193 containing more genes than the plasmid, and thus offering a bigger mutational target,
194 but it is possible that other selective forces or limitations may contribute to this pattern.
195 For example, in diverse communities, variation in plasmid fitness effects across host
196 backgrounds [23] may weaken selection for plasmid-encoded compensatory mutation.
197
198 Importantly, compensatory evolution occurs both with or without positive selection for
199 plasmid encoded functions [30,37], confirming that even when a costly plasmid's net
200 fitness effect is beneficial (because the benefit outweighs the cost), the fitness cost of
201 plasmid maintenance remains and must be ameliorated to prevent plasmid loss driven
202 by chromosomal capture. Indeed, positive selection has been shown to enhance
203 compensatory evolution by increasing the population size of plasmid-carrying cells [38],
204 which increases the likelihood of acquiring a compensatory mutation. Studies that have
205 tracked the temporal dynamics of compensatory evolution suggest that there is, in
206 effect, a race between the processes of plasmid loss, chromosomal capture, and
207 compensatory evolution [37,38,45]. Nonetheless, compensatory evolution can be

208 extremely rapid, in some cases occurring during the outgrowth of transconjugant
209 colonies within 48-72 hours [39].

210

211 A potential explanation for the speed of compensatory evolution is that amelioration
212 often only requires a single mutation, although some examples of large-scale genetic
213 changes, including large plasmid deletions, have been observed [40,46,47]. Comparing
214 multiple independent populations shows that compensatory mutations tend to be
215 focused within one or a few genetic targets [8,30,37–41]. This pattern of requiring only
216 one or a few targeted mutations to ameliorate fitness costs suggests that these
217 plasmids cause their fitness costs through specific genetic conflicts occurring between
218 one or few plasmid genes and one or few chromosomal genes. The targets of
219 compensatory mutations are often associated with the SOS response, and in particular
220 helicase genes have acquired compensatory mutation across several host-plasmid
221 pairs [38,41–43]. In addition, genes that interact deleteriously with incoming plasmids
222 are often themselves horizontally acquired accessory genes already present in the
223 chromosome [42,48,49].

224

225 Certain chromosomal compensatory mutations have been shown to ameliorate the
226 fitness costs of multiple plasmids in a cell, potentially enabling these bacterial lineages
227 to become hotspots of plasmid-plasmid recombination [42,50–52]. However, even after
228 compensatory evolution to negate their fitness costs, the coexistence of plasmids in a
229 cell may not be stable if they encode the same ecological function. This is because

230 positive selection has been shown to discriminate between the relative fitness benefits
231 of coexisting plasmids, only retaining the most beneficial [50].

232

233 Compensatory evolution is a particularly important mechanism enabling the long-term
234 maintenance and survival of non-conjugative plasmids, which unless spread horizontally
235 by other mechanisms (e.g. packaging by temperate phages) rely entirely upon their
236 efficiency of vertical transmission at cell division [38].

237

238 *Solution 6: Piggybacking on niche adaptation*

239 Compensatory mutations are defined as those that specifically reduce the plasmid
240 fitness cost, and thus will not be selected in bacterial populations evolved without the
241 plasmid where they provide no benefit. Recent experiments by Kloos *et al* [53] reveal a
242 different class of mutations that are generally beneficial, and as such evolve in both
243 plasmid-carrying and plasmid-free cells, but nonetheless have the effect of
244 pleiotropically reducing the fitness cost of the plasmid. Using clinical isolates of *E. coli*
245 with two different multidrug resistance plasmid, Kloos *et al* [53] showed that mutations in
246 bacterial global regulators of metabolism reduced the plasmid fitness costs by causing a
247 net downregulation of plasmid gene expression. Crucially, these mutations increased
248 bacterial growth in the environment in which they evolved regardless of plasmid
249 carriage, confirming that in addition to reducing plasmid fitness costs these mutations
250 were generally adaptive in this niche.

251

252 **Conclusion**

253 The plasmid paradox has inspired a large body of research during the past decade. This
254 work demonstrates that there are multiple ecological and evolutionary mechanisms that
255 can explain the persistence of plasmids in bacterial genomes. These mechanisms may
256 also often work in concert, such that a plasmid may undergo compensatory evolution to
257 negate its fitness cost whilst also still transmitting horizontally by conjugation, with
258 potentially synergistic benefits for the plasmid which is no longer subject to negative
259 selection. While the theoretical plasmid paradox is now effectively solved, the challenge
260 for the field is to translate these insights to better understand plasmid dynamics in
261 natural microbial communities.

262

263 **Future directions**

264 With the plasmid paradox now solved, where next for plasmid ecology and evolution?
265 Below we outline some of the key open research questions for the field (see also
266 Outstanding Questions):

- 267 • **What is the relative importance of these plasmid stability mechanisms in**
268 **nature?** Most experimental work on plasmid stability to date has used relatively
269 simplified laboratory systems, which do not reflect the complexity of plasmid-host
270 interactions in natural communities and environments (although see [32,54] for
271 experimental set-ups that approach more natural levels of diversity and habitat).
272 Studies tracking the ecological and evolutionary dynamics of plasmid-host
273 interactions in natural communities are required to determine the relative
274 contributions of the various mechanisms of plasmid stability observed in the lab
275 to dynamics occurring in nature. A recent example of such a study tracked a

276 carbapenem resistance plasmid in hospitalised patients, demonstrating pervasive
277 plasmid horizontal transmission between bacterial lineages and species,
278 suggesting a potentially important role for spillover transmission in this natural
279 setting [55].

280 ● **What are the effects of these plasmid stability mechanisms on the**
281 **dynamics of horizontal gene transfer?** Understanding how different
282 mechanisms of plasmid stability impact rates of horizontal gene transfer and
283 gene mobilisation will be important for predicting the spread of accessory genes
284 in natural communities. For example, in environments where plasmids persist by
285 infectious transmission, higher rates of interspecies gene mobilisation have been
286 observed compared to communities in environments where plasmids persisted
287 mostly by vertical transmission [56]. Conversely, in some studies, compensatory
288 evolution has been associated with reduced conjugative ability [37,57], and thus
289 potentially reduced rates of HGT. More generally, whether compensatory
290 mutations are plasmid- or chromosomally encoded could alter rates of horizontal
291 gene transfer in complex microbial communities, and in turn different
292 compensatory mechanisms could be favoured depending on the host and
293 plasmid community diversity [44].

294 ● **What drives the distribution of plasmids in bacterial (pan)genomes?**
295 Understanding the molecular causes of the fitness costs of plasmids across a
296 wider diversity of plasmid-host interactions could enable the compatibility of
297 plasmid-host pairs to be predicted from sequence data. This could in turn
298 improve prediction of strains that pose particular risks of stably acquiring

299 plasmids and e.g. becoming multidrug resistant or acting as hubs of plasmid
300 recombination and dissemination. For example, comparative genomic studies
301 have revealed associations between plasmid carriage and chromosomal
302 regulatory sequences that suggest past compensatory evolution, and may
303 indicate other strains with the same allelic variants that are “pre-adapted” to be
304 proficient plasmid hosts [58]. How these patterns of chromosome-plasmid
305 compatibility interact with selection for the genes encoded on plasmids will shape
306 the flow of plasmids and the genes they mobilise in bacterial pangenomes [59].

- 307 ● **What kinds of genes become mobilised by plasmids and why?** Plasmids are
308 notorious for encoding and transferring antibiotic resistance genes among
309 lineages, thus contributing to the antimicrobial resistance crisis [60]. But plasmids
310 also carry other kinds of functions, such as metabolic, virulence, and symbiosis
311 genes [61,62]. One property that potentially unifies the kinds of traits that are
312 more frequently plasmid versus chromosomally encoded is that their fitness
313 benefits are strongly environmentally contingent, such that being encoded on
314 plasmids might enable their recurrent gain, loss, and regain depending upon
315 local selection pressures [63]. Moreover, genes with social effects, such as
316 cooperative or spiteful traits, have been suggested to be enriched on plasmids
317 [64]. Recent theory suggests that whether genes are plasmid or chromosomally
318 encoded tends to be under positive frequency dependent selection, leading to
319 priority effects such that once mobilised a moderately beneficial plasmid encoded
320 trait will tend to prevent the invasion of a chromosomal version, especially when
321 the trait is moderately beneficial across multiple bacterial species [65].

322 • **What are the effects of plasmids beyond horizontal gene transfer?** Plasmids
323 play other roles in bacterial ecology and evolution besides the transfer of
324 accessory genes [66] but these remain relatively poorly understood. Key
325 examples include the manipulation of bacterial phenotypes through gene
326 regulatory cross-talks [67], which can alter bacterial lifestyle (e.g. from planktonic
327 to biofilm growth [68,69]) or physiology (e.g. from aerobic to anaerobic
328 metabolism [22]). In addition, because plasmids are often present in multiple
329 copies within the cell, this can alter the evolutionary dynamics and evolvability of
330 plasmid-encoded versus chromosomally encoded genes [70].

331

332

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339

340 **References**

- 341 1 Thomas, C.M. and Nielsen, K.M. (2005) Mechanisms of, and barriers to, horizontal
342 gene transfer between bacteria. *Nat. Rev. Microbiol.* 3, 711–721
- 343 2 Norman, A. *et al.* (2009) Conjugative plasmids: vessels of the communal gene pool.
344 *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 364, 2275–2289

- 345 3 Hall, J.P.J. *et al.* (2017) Sampling the mobile gene pool: innovation via horizontal
346 gene transfer in bacteria. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 372,
- 347 4 Harrison, E. and Brockhurst, M.A. (2012) Plasmid-mediated horizontal gene transfer
348 is a coevolutionary process. *Trends Microbiol.* 20, 262–267
- 349 5 Baltrus, D.A. (2013) Exploring the costs of horizontal gene transfer. *Trends Ecol.*
350 *Evol.* 28, 489–495
- 351 6 San Millan, A. and MacLean, R.C. (2017) Fitness Costs of Plasmids: a Limit to
352 Plasmid Transmission. *Microbiol Spectr* 5,
- 353 7 Bergstrom, C.T. *et al.* (2000) Natural selection, infectious transfer and the existence
354 conditions for bacterial plasmids. *Genetics* 155, 1505–1519
- 355 8 Stalder, T. *et al.* (2017) Emerging patterns of plasmid-host coevolution that stabilize
356 antibiotic resistance. *Sci. Rep.* 7, 4853
- 357 9 Lili, L.N. *et al.* (2007) The persistence of parasitic plasmids. *Genetics* 177, 399–405
- 358 10 Gordon, D.M. (1992) Rate of plasmid transfer among *Escherichia coli* strains
359 isolated from natural populations. *J. Gen. Microbiol.* 138, 17–21
- 360 11 Lopatkin, A.J. *et al.* (2017) Persistence and reversal of plasmid-mediated antibiotic
361 resistance. *Nat. Commun.* 8, 1689
- 362 12 Stevenson, C. *et al.* (2017) Gene mobility promotes the spread of resistance in
363 bacterial populations. *ISME J.* 11, 1930–1932
- 364 13 Hall, J.P.J. *et al.* (2016) Source–sink plasmid transfer dynamics maintain gene
365 mobility in soil bacterial communities. *Proceedings of the at*
366 <<https://www.pnas.org/content/113/29/8260.short>>
- 367 14 Stevenson, C. *et al.* (2018) Plasmid stability is enhanced by higher-frequency

368 pulses of positive selection. *Proc. Biol. Sci.* 285,
369 15 Stewart, F.M. and Levin, B.R. (1977) The Population Biology of Bacterial Plasmids:
370 A PRIORI Conditions for the Existence of Conjugationally Transmitted Factors.
371 *Genetics* 87, 209–228

372 16 Harrison, E. *et al.* (2015) Bacteriophages limit the existence conditions for
373 conjugative plasmids. *MBio* 6, e00586

374 17 Cairns, J. *et al.* (2016) Conjugation is necessary for a bacterial plasmid to survive
375 under protozoan predation. *Biol. Lett.* 12, 20150953

376 18 Dionisio, F. *et al.* (2019) Interactions between plasmids and other mobile genetic
377 elements affect their transmission and persistence. *Plasmid* 102, 29–36

378 19 Rodríguez-Rubio, L. *et al.* (2020) Extensive antimicrobial resistance mobilization via
379 multicopy plasmid encapsidation mediated by temperate phages. *J. Antimicrob.*
380 *Chemother.* 75, 3173–3180

381 20 Hasegawa, H. *et al.* (2018) Horizontal Plasmid Transfer by Transformation in
382 *Escherichia coli*: Environmental Factors and Possible Mechanisms. *Front. Microbiol.*
383 9, 2365

384 21 De Gelder, L. *et al.* (2007) Stability of a promiscuous plasmid in different hosts: no
385 guarantee for a long-term relationship. *Microbiology* 153, 452–463

386 22 Dunn, S. *et al.* (2021) Limited and Strain-Specific Transcriptional and Growth
387 Responses to Acquisition of a Multidrug Resistance Plasmid in Genetically Diverse
388 *Escherichia coli* Lineages. *mSystems* 6,

389 23 Alonso-Del Valle, A. *et al.* (2021) Variability of plasmid fitness effects contributes to
390 plasmid persistence in bacterial communities. *Nat. Commun.* 12, 2653

391 24 Kottara, A. *et al.* (2018) Variable plasmid fitness effects and mobile genetic element
392 dynamics across *Pseudomonas* species. *FEMS Microbiol. Ecol.* 94,
393 25 Sheppard, R.J. *et al.* (2020) The role of hosts, plasmids and environment in
394 determining plasmid transfer rates: A meta-analysis. *Plasmid* 108, 102489
395 26 Kottara, A. *et al.* 18-May-(2021) , The dilution effect limits plasmid horizontal
396 transmission in multispecies bacterial communities. , *bioRxiv*, 2021.05.18.444624
397 27 Gama, J.A. *et al.* (2017) Multiple plasmid interference - Pledging allegiance to my
398 enemy's enemy. *Plasmid* 93, 17–23
399 28 Gama, J.A. *et al.* (2017) Co-resident plasmids travel together. *Plasmid* 93, 24–29
400 29 San Millan, A. *et al.* (2014) Positive epistasis between co-infecting plasmids
401 promotes plasmid survival in bacterial populations. *ISME J.* 8, 601–612
402 30 Carrilero, L. *et al.* 30-Sep-(2020) , Positive selection inhibits plasmid coexistence in
403 bacterial genomes. , *bioRxiv*, 2020.09.29.318741
404 31 Gama, J.A. *et al.* (2020) Plasmid Interactions Can Improve Plasmid Persistence in
405 Bacterial Populations. *Front. Microbiol.* 11, 2033
406 32 Cairns, J. *et al.* (2018) Ecology determines how low antibiotic concentration impacts
407 community composition and horizontal transfer of resistance genes. *Commun Biol*
408 1, 35
409 33 Kottara, A. *et al.* (2021) The proficiency of the original host species determines
410 community-level plasmid dynamics. *FEMS Microbiol. Ecol.* 97,
411 34 Benmayor, R. *et al.* (2009) Host mixing and disease emergence. *Curr. Biol.* 19,
412 764–767
413 35 Chabas, H. *et al.* (2018) Evolutionary emergence of infectious diseases in

414 heterogeneous host populations. *PLoS Biol.* 16, e2006738

415 36 Kottara, A. *et al.* (2016) Multi-host environments select for host-generalist
416 conjugative plasmids. *BMC Evol. Biol.* 16, 70

417 37 Harrison, E. *et al.* (2015) Parallel compensatory evolution stabilizes plasmids
418 across the parasitism-mutualism continuum. *Curr. Biol.* 25, 2034–2039

419 38 San Millan, A. *et al.* (2014) Positive selection and compensatory adaptation interact
420 to stabilize non-transmissible plasmids. *Nat. Commun.* 5, 5208

421 39 Hall, J.P.J. *et al.* (2020) Extremely fast amelioration of plasmid fitness costs by
422 multiple functionally diverse pathways. *Microbiology* 166, 56–62

423 40 Porse, A. *et al.* (2016) Survival and Evolution of a Large Multidrug Resistance
424 Plasmid in New Clinical Bacterial Hosts. *Mol. Biol. Evol.* 33, 2860–2873

425 41 Sota, M. *et al.* (2010) Shifts in the host range of a promiscuous plasmid through
426 parallel evolution of its replication initiation protein. *ISME J.* 4, 1568–1580

427 42 Loftie-Eaton, W. *et al.* (2017) Compensatory mutations improve general
428 permissiveness to antibiotic resistance plasmids. *Nat Ecol Evol* 1, 1354–1363

429 43 Loftie-Eaton, W. *et al.* (2016) Evolutionary Paths That Expand Plasmid Host-Range:
430 Implications for Spread of Antibiotic Resistance. *Mol. Biol. Evol.* 33, 885–897

431 44 Zwanzig, M. *et al.* (January/February 2019) Mobile Compensatory Mutations
432 Promote Plasmid Survival. *mSystems* DOI: 10.1128/mSystems.00186-18

433 45 Harrison, E. *et al.* Rapid compensatory evolution promotes the survival of
434 conjugative plasmids. , *Mobile genetic elements*, 6. May-(2016) , e1179074

435 46 Lee, M.-C. and Marx, C.J. (2012) Repeated, selection-driven genome reduction of
436 accessory genes in experimental populations. *PLoS Genet.* 8, e1002651

437 47 Modi', R.I. *et al.* (1991) Plasmid macro-evolution: selection of deletions during
438 adaptation in a nutrient- limited environment. *Genetica* 84, 195–202

439 48 Hall, J.P.J. *et al.* 12-Apr-(2021) , Plasmid fitness costs are caused by specific
440 genetic conflicts. , *bioRxiv*, 2021.04.10.439128

441 49 San Millan, A. *et al.* (2015) Interactions between horizontally acquired genes create
442 a fitness cost in *Pseudomonas aeruginosa*. *Nat. Commun.* 6, 6845

443 50 Carrilero, L. *et al.* (2021) Positive Selection Inhibits Plasmid Coexistence in
444 Bacterial Genomes. *MBio* 12,

445 51 Jordt, H. *et al.* (2020) Coevolution of host-plasmid pairs facilitates the emergence of
446 novel multidrug resistance. *Nat Ecol Evol* DOI: 10.1038/s41559-020-1170-1

447 52 Santos-Lopez, A. *et al.* 11-Sep-(2017) , Compensatory evolution facilitates the
448 acquisition of multiple plasmids in bacteria. , *bioRxiv*, 187070

449 53 Kloos, J. *et al.* (2021) Piggybacking on niche-adaptation improves the maintenance
450 of multidrug resistance plasmids. *Mol. Biol. Evol.* DOI: 10.1093/molbev/msab091

451 54 Hall, J.P.J. *et al.* (2020) The Impact of Mercury Selection and Conjugative Genetic
452 Elements on Community Structure and Resistance Gene Transfer. *Front. Microbiol.*
453 11, 1846

454 55 León-Sampedro, R. *et al.* (2021) Pervasive transmission of a carbapenem
455 resistance plasmid in the gut microbiota of hospitalized patients. *Nat Microbiol* 6,
456 606–616

457 56 Hall, J.P.J. *et al.* (2017) Positive selection inhibits gene mobilization and transfer in
458 soil bacterial communities. *Nature ecology & at*
459 <<https://www.nature.com/articles/s41559-017-0250-3>>

460 57 Heuer, H. *et al.* (2006) Frequent conjugative transfer accelerates adaptation of a
461 broad-host-range plasmid to an unfavorable *Pseudomonas putida* host: Adaptation
462 of a plasmid to an unfavorable host. *FEMS Microbiol. Ecol.* 59, 738–748

463 58 McNally, A. *et al.* (2016) Combined Analysis of Variation in Core, Accessory and
464 Regulatory Genome Regions Provides a Super-Resolution View into the Evolution
465 of Bacterial Populations. *PLoS Genet.* 12, e1006280

466 59 Whelan, F.J. *et al.* (2021) Evidence for selection in the abundant accessory gene
467 content of a prokaryote pangenome. *Mol. Biol. Evol.* DOI: 10.1093/molbev/msab139

468 60 San Millan, A. (2018) Evolution of Plasmid-Mediated Antibiotic Resistance in the
469 Clinical Context. *Trends Microbiol.* 26, 978–985

470 61 Stasiak, G. *et al.* (2014) Functional relationships between plasmids and their
471 significance for metabolism and symbiotic performance of *Rhizobium*
472 *leguminosarum* bv. *trifolii*. *J. Appl. Genet.* 55, 515–527

473 62 Ahmer, B.M. *et al.* (1999) The virulence plasmid of *Salmonella typhimurium* is self-
474 transmissible. *J. Bacteriol.* 181, 1364–1368

475 63 Niehus, R. *et al.* (2015) Migration and horizontal gene transfer divide microbial
476 genomes into multiple niches. *Nat. Commun.* 6, 8924

477 64 Rankin, D.J. *et al.* (2011) What traits are carried on mobile genetic elements, and
478 why? *Heredity* 106, 1–10

479 65 Lehtinen, S. *et al.* (2021) Evolutionary mechanisms that determine which bacterial
480 genes are carried on plasmids. *Evol Lett* 5, 290–301

481 66 Rodríguez-Beltrán, J. *et al.* (2021) Beyond horizontal gene transfer: the role of
482 plasmids in bacterial evolution. *Nat. Rev. Microbiol.* 19, 347–359

- 483 67 Vial, L. and Hommais, F. (2020) Plasmid-chromosome cross-talks. *Environ.*
484 *Microbiol.* 22, 540–556
- 485 68 Ghigo, J.M. (2001) Natural conjugative plasmids induce bacterial biofilm
486 development. *Nature* 412, 442–445
- 487 69 Parashar, V. *et al.* (2013) A plasmid-encoded phosphatase regulates *Bacillus*
488 *subtilis* biofilm architecture, sporulation, and genetic competence. *J. Bacteriol.* 195,
489 2437–2448
- 490 70 San Millan, A. *et al.* (2016) Multicopy plasmids potentiate the evolution of antibiotic
491 resistance in bacteria. *Nat Ecol Evol* 1, 10

492

493

494

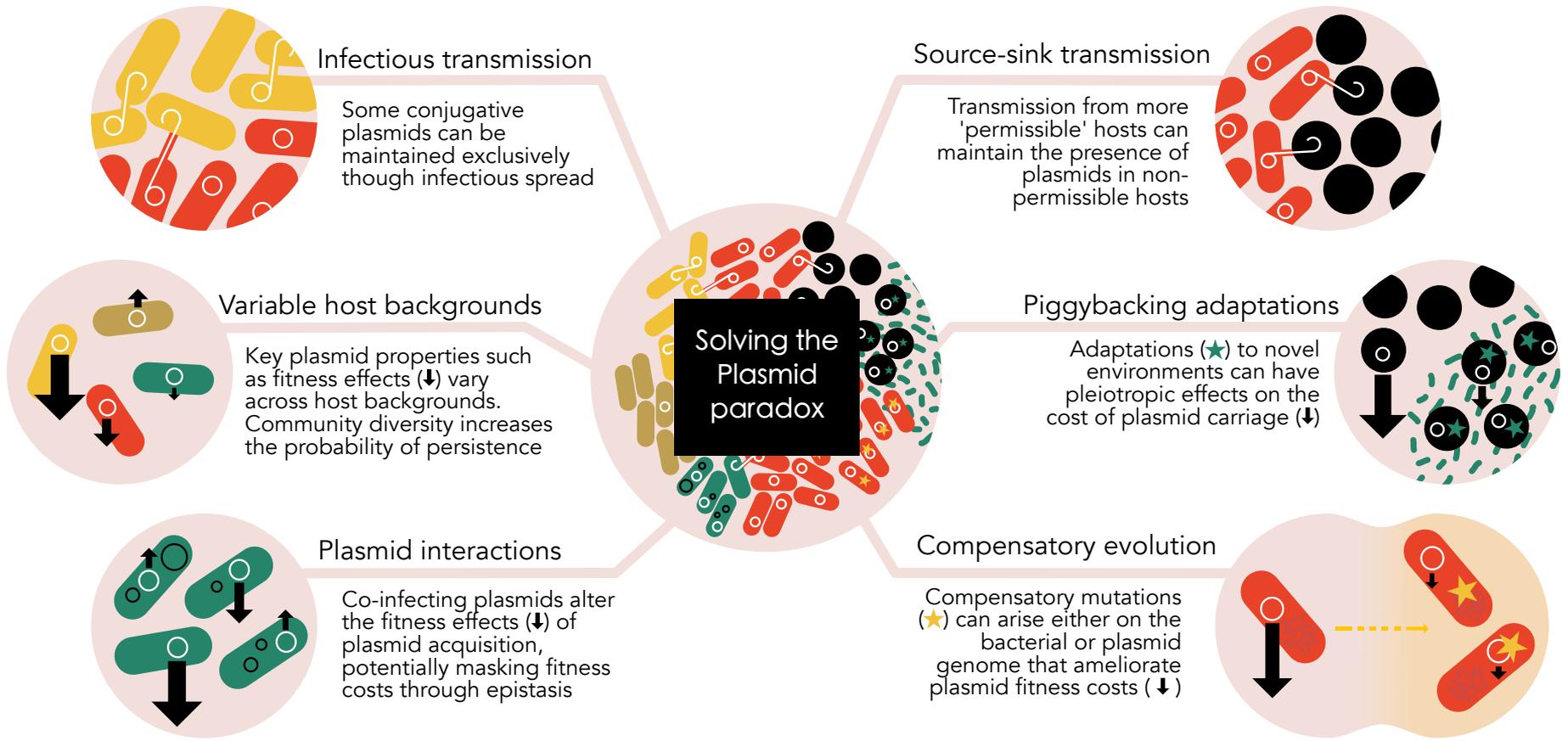
495 **Figure Legend**

496

497 **Figure 1 | Ecological and evolutionary mechanisms of plasmid stability**

498 Over the past decade theoretical and experimental studies have discovered a range of
499 ecological and evolutionary mechanisms that enable plasmids to persist in bacterial
500 populations and communities. Cells of the same colour belong to the same species,
501 whereas those of a different colour belong to a different species or lineage. Plasmids
502 are shown as white rings, and where these connect cells, this denotes conjugative
503 transfer. Solid arrows denote fitness costs, and are scaled by the magnitude of the
504 fitness cost. Dotted lines denote evolutionary changes.

Figure



- Plasmids are common in bacterial genomes, but theoretically they should not exist, giving rise to the plasmid paradox
- Recent studies show that multiple mechanisms can explain the long-term persistence of plasmids in bacterial genomes, solving the plasmid paradox
- Ecological solutions to the plasmid paradox include infectious transmission and variation in fitness costs between lineages
- Evolutionary solutions to the plasmid paradox include compensatory evolution and piggybacking niche adaptation

- What is the relative importance of these plasmid stability mechanisms in nature?
- What are the effects of these plasmid stability mechanisms on the dynamics of horizontal gene transfer?
- What drives the distribution of plasmids in bacterial (pan)genomes?
- What kinds of genes become mobilised by plasmids and why?
- What are the effects of plasmids beyond horizontal gene transfer?