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Harnessing the membrane potential to combat cancer progression

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Abstract

Rapid fluctuations in the plasma membrane potential (V_m) provide the basis underlying the action potential waveform in electrically excitable cells; however, a growing body of literature shows that the V_m is also functionally instructive in non-excitable cells, including cancer cells. Various ion channels play a key role in setting and fine tuning the V_m in cancer and stromal cells within the tumor microenvironment, raising the possibility that the V_m could be targeted therapeutically using ion channel-modulating compounds. Emerging evidence points to the V_m as a viable therapeutic target, given its functional significance in regulating cell cycle progression, migration, invasion, immune infiltration and pH regulation. Several compounds are now undergoing clinical trials and there is increasing interest in therapeutic manipulation of the V_m via application of pulsed electric fields. The purpose of this article is to update the reader on the significant recent and ongoing progress to elucidate the functional significance of V_m regulation in tumors, to highlight key remaining questions and the prospect of future therapeutic targeting. In particular, we focus on key developments in understanding the functional consequences of V_m alteration on tumor development via the activation of small GTPase (K-Ras and Rac1) signalling, as well as the impact of V_m changes within the heterogeneous tumor microenvironment on immune cell function and cancer progression.

Introduction

The plasma membrane potential (V_m), the voltage difference between the cytosol and the extracellular environment, is determined by the unequal distribution and differential permeability of key ions including Na⁺, K⁺, Ca²⁺ and Cl⁻. Rapid fluctuations in V_m provide the basis underlying the action potential waveform in electrically excitable cells; however, a growing body of literature shows that the V_m is also functionally instructive in non-excitable cells, including cancer cells. In addition to this, increasing evidence points to the V_m of various organelles, e.g. mitochondria¹, endosomes², playing key roles in regulating cellular behaviour, as well as likely interplay between plasma V_m and organellar V_m underlying unique signalling axes³. In our 2013 review article⁴, we summarised current understanding in the field relating to V_m and its role in cancer progression. Since 2013, research in this area has progressed rapidly. The purpose of the present article is to update the reader on these key developments, with a focus on the role of the plasma V_m in cancer progression, and how it may present a unique therapeutic target.

V_m depolarization in cancer cells

Studies over a number of years have shown a remarkable correlation between V_m and malignancy, such that cancer cells have a more depolarized V_m compared to healthy normal cells (see⁴ for references to individual studies dating back to 1959) (Figure 1). This correlation may, at least in part, be due to the fact that cancer cells are often highly proliferative, compared to terminally differentiated somatic cells, and thus V_m depolarization may be associated with a proliferative state. Indeed, V_m depolarization has been shown to initiate mitosis whereas hyperpolarization induced mitotic arrest⁵, and artificial depolarization of *Xenopus laevis* embryos has been shown to promote malignant transformation, providing a sustaining proliferative signal⁶. Moreover, an exquisite rhythmic relationship exists between V_m and the cell cycle⁷. In addition to general depolarization seen in cancer cells

versus healthy normal counterparts, the V_m has been shown to fluctuate during cell cycle progression. V_m hyperpolarization has been shown to occur at the G₁/S checkpoint, remaining fairly hyperpolarized during the S phase, before becoming depolarised during the G₂/M transition.

A multiplicity of different ion channels has been shown to contribute to the fine tuning of V_m in cancer cells, thus promoting proliferation⁴. Of particular note is the involvement of various classes of K⁺ channels, several of which serve not only as cancer biomarkers, but also as regulators of cell cycle progression and cellular proliferation, in part due to regulation of proproliferative signalling via altered intracellular Ca^{2+ 8}. These ion channel-dependent oscillations in V_m observed in cancer cells have led to the 'Celex hypothesis', which argues that K⁺ channels are expressed on cancer cells relatively early on in tumor development, but as the disease progresses to a more invasive/metastatic phenotype, K⁺ channel expression is downregulated and replaced by an upregulation of voltage-gated Na⁺ channels (VGSCs)⁹. There is growing evidence in support of the notion that metastatic cancer cells are themselves electrically excitable and capable of firing action potentials^{10–12}. In addition, the Celex hypothesis fits with the general depolarization of V_m observed in metastatic cancer cells versus non-metastatic cells at steady state¹³. Furthermore, the persistent inward Na⁺ current carried by VGSCs expressed on metastatic cancer cells would be expected to contribute to this depolarization, and this, together with downregulated K⁺ channel activity, promotes metastatic invasion¹⁴.

Functional consequences of V_m alteration in tumor development

Despite the myriad different studies pointing to a link between V_m depolarization and cell proliferation, the mechanism(s) involved have, until relatively recently, remained elusive. In 2015, an elegant mechanism providing a direct link between V_m and mitogenic signaling was described for the first time¹⁵. In this model, V_m depolarization induced by the activities of

specific ion channels resulted in the redistribution of negatively charged phospholipids (phosphatidylserine and phosphatidylinositol 4,5-bisphosphate) within the inner leaflet of the phospholipid bilayer of the plasma membrane. This redistribution of phospholipids resulted in nanoclustering of the phosphatidylserine-anchored small guanosine triphosphate hydrolase (GTPase) K-Ras, leading to its activation and induction of the rapidly accelerated fibrosarcoma (RAF)–mitogen-activated protein kinase (MAPK) cascade, promoting proliferation. The authors thus delineated a mechanism by which K-Ras can function as a 'field effect transistor', linking the V_m to mitogenic intracellular signalling pathways¹⁵.

A growing body of evidence implicates V_m in the regulation of other cellular processes that are integral to tumor development and the metastatic cascade, in addition to cancerous transformation and increased proliferation. For example, V_m depolarization may also promote apoptosis resistance: Kv1.5-dependent Vm hyperpolarization in cancer cell lines has been shown to inhibit voltage-gated Ca2+ channels, reducing intracellular Ca2+ levels and therefore inhibiting activation of nuclear factor of activated T-cells (NFAT), increasing apoptosis³. In addition, V_m fluctuations can also affect cancer cell migration; in part the mechanisms are likely dependent on V_m-mediated alteration of Ca²⁺ signalling, in turn leading to cytoskeletal reorganization^{16,17}. V_m-dependent regulation of cancer cell migration may also be Ca^{2+} -independent: we recently showed that $Na_v 1.5$ -dependent V_m depolarization promotes redistribution of the phosphatidylserine-anchored small GTPase Rac1 at the leading edge of migrating metastatic breast cancer cells, resulting in Rac1 activation, cytoskeletal reorganisation and acquisition of a motile phenotype¹⁸. Thus, V_mdependent activation of small GTPase signalling can not only increase mitogenic signalling¹⁵, but can also promote migration in response to changes in the ionic tumor microenvironment (TME)¹⁸.

V_m depolarization may also promote cancer cell invasion. Similar to depolarizationdependent migration, evidence suggests that persistent inward Na⁺ current through VGSCs

enhances cellular invasive capacity¹³. Increased cytosolic Na⁺ resulting from this persistent inward current has itself been shown to have a multiplicity of consequences on cancer cell behaviour¹⁹, including altered pH buffering capacity, leading to activation of pH-dependent cysteine cathepsins and increased invasion²⁰. An additional hitherto unexplored consequence of V_m depolarization is on the regulation of cancer stem cell differentiation. Various studies have shown that the V_m can regulate differentiation of human mesenchymal stem cells, with depolarization likely maintaining cells in an undifferentiated state²¹. It is not yet clear whether such a mechanism exists in the context of cancer stem cells; however, V_m depolarization may serve as a survival mechanism to preserve energy, as has been shown in immune cells, where high extracellular K⁺ maintains T cell stemness²².

Impact of V_m changes in the tumor microenvironment

Tumor progression is regulated by the crosstalk among components of the TME, which consists of tumor cells, immune cells, extracellular matrix (ECM) and stromal cells. Although the ionic composition of the TME is less well studied and therefore the functional roles of V_m in the various constituent TME cell types remains unclear, high local extracellular [K⁺] within mouse and human tumors has been shown to suppresses cluster of differentiation (CD) 8⁺ T cell effector function in necrotic areas of the TME²³, highlighting the relevance of ionic reprogramming to tumor-infiltrating immune cells. This is supported by an *in vitro* study showing that high extracellular [K⁺] inhibits T cell proliferation, reduces pro-inflammatory cytokine production, including interferon (IFN)-γ and interleukin (IL)-2, and increases expression of the programmed death-1 (PD-1) receptor on T cells²⁴, leading to immunosuppression in PD-1 ligand (PD-L1)-expressing tumors.

Tumor ionic activity is closely related to T cell activation, differentiation, proliferation and apoptosis via Ca^{2+} signaling²⁵. For example, K_{Ca} and K_V channels maintain a hyperpolarized V_m and increase the driving force for Ca^{2+} entry via the Ca^{2+} release-activated Ca^{2+} (CRAC)

channel Orai1, which in turn tunes T cell immunity²⁶. Compared to quiescent T cells, stimulated T cells have distinct expression profiles of K_{Ca} and K_v channels, and blocking those channels impairs CD4⁺ T cell proliferation and IL-2 secretion^{27,28}, and blockade of K_{Ca} 3.1 channels inhibits the migration of CD3⁺ T cells²⁹. High availability of Na⁺ in the TME¹⁹ may increase the driving force for Na⁺ entry into T cells via the Ca²⁺-activated cation channel, transient receptor potential cation channel subfamily M (TRPM)4, whose activation leads to a depolarized phenotype and decreases cytosolic [Ca²⁺], which may reduce IL-2 production and therefore dampen tumoricidal activity³⁰. Furthermore, high extracellular [K⁺], [Ca²⁺] (both due to apoptosis/necrosis) and high [Na⁺] in tumor hypoxic/necrotic regions, together with the abundance of H₂O₂, which activates TRPM2, can lead to Ca²⁺ influx-induced T cell apoptosis³¹. Together, these results raise the intriguing possibility that hyperpolarizing the V_m of cytotoxic effector cells may increase their efficiency in combating tumor cells.

For cancer cells to avoid immune eradication, there is usually a dysregulation in the interaction and balance between the effector immune cell and the regulatory immune cell population. Among the latter, tumor-associated macrophages (TAMs) are widely present within the TME, exhibiting a tumor-promoting (sometimes known as M2) phenotype. Blocking K_{Ca}3.1 channels on TAMs with the clotrimazole analogue TRAM-34 when co-cultured with colorectal cancer cells directed the TAMs towards an anti-inflammatory M2 phenotype³². Additionally, TRAM-34-treated mice exhibited a significant reduction in plasma pro-inflammatory cytokine levels, including IL-2, IL-6 and tumor necrosis factor- α (TNF- α), and increased the plasma levels of the anti-inflammatory cytokine IL-10³³. Interestingly, TRAM-34 also increased IL-10 expression and secretion in immunosuppressive CD4⁺CD25⁺ regulatory T (T_{reg}) cells in an inflammatory bowel disease mouse model³⁴. Future studies should investigate whether hyperpolarized V_m in immune cells leads to a reduction in immunosuppression in the TME.

Whether V_m itself can functionally regulate immune cell recruitment to the TME is not well understood. However, some evidence suggests that TMEs with an aberrant ionic composition may affect the chemotaxis of lymphocytes and other immune cell types. For example, migration of human lung mast cells towards various chemoattractants including C-X-C motif chemokine ligand 10 (CXCL10) is attenuated upon K_{Ca} channel blockade³⁵. In addition, activating K_{Ca}3.1 channels on both CD11c^{high}CD11b^{low} and CD11c^{low}CD11b^{high} lung dendritic cells hyperpolarized their V_m, and blocking these channels with TRAM-34 impaired cysteine-cysteine chemokine ligand (CCL)19/CCL21-induced dendritic cell transmigration *in vitro*³⁶.

Future perspectives: therapeutic V_m targeting

The key role of various ion channels in setting and fine tuning the V_m in cancer and stromal cells within the TME (Figure 2) raises the possibility that the V_m could be targeted therapeutically using ion channel-modulating compounds. Indeed, a large body of preclinical research highlights a range of ion channel-modulating drugs as potential anti-cancer therapeutics, although the efficacy of many of these compounds is likely due to functional consequences in addition to V_m alteration³⁷. Nonetheless, the emerging evidence highlighted in this Perspectives article points to the V_m as a viable therapeutic target, given its functional significance in regulating a range of intrinsic tumor features, including cell cycle progression, migration, invasion, immune infiltration and pH regulation (Figure 3). Excitingly, several of these ion transport-modulating interventions are now undergoing clinical trials^{19,37}. Furthermore, there is increasing interest in therapeutic manipulation of cancer cell behaviour (and disruption of the V_m) via application of pulsed electric fields^{38,39}. Clearly, for such interventions to be effective at targeting the V_m, the wider context of tissue-, organ-wide and long-range endogenous electric fields needs to be considered⁴⁰, as well as the possibility that cancer cells in certain tumor types may function as an electrical syncytium via gapjunctional communication⁴¹. A barrier to progression in this area remains the absence of

reliable methods to accurately quantify small V_m changes *in vitro* and *in vivo*, although encouraging progress is now being made in this regard^{42,43}. In addition, further work is required to address whether the V_m-dependent effects reported thus far are generalisable across different tumor types. Finally, for such (drug-based or electrical) therapies to be effective, adequate selectivity for the lesion *vs.* healthy normal tissue is of paramount importance. Such selectivity may be achieved for electric field interventions by appropriate electrode replacement, however, challenges remain with respect to systemic drug application, when target ion channels may play a key role in regulating V_m in normal tissues, e.g. during mammary gland development/remodelling *vs.* cancer^{18,44}. Further work is required to establish the effect(s) of local and systemic V_m-modulating therapies on both stromal cells in the TME (e.g. infiltrating immune cells) and surrounding normal cells. In conclusion, significant progress continues at pace to elucidate the functional significance of V_m regulation in tumors, and although key questions remain, there is the exciting prospect of its realization as a future therapeutic target.

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Authorship confirmation statement

WB had the original idea for this study. MY and WB wrote the manuscript. Both authors contributed to the interpretation and revising the manuscript. Both authors approved the final submitted version of the manuscript.

Authors' disclosure

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References

- Pedersen SF, Flinck M, Pardo LA. The Interplay between Dysregulated Ion Transport and Mitochondrial Architecture as a Dangerous Liaison in Cancer. Int J Mol Sci;22.
 Epub ahead of print May 14, 2021. DOI: 10.3390/ijms22105209.
- 2. Jung J, Venkatachalam K. TRPML1 and RAS-driven cancers exploring a link with great therapeutic potential. Channels 2019;13:374–381.
- Bonnet S, Archer SL, Allalunis-Turner J, et al. A mitochondria-K+ channel axis is suppressed in cancer and its normalization promotes apoptosis and inhibits cancer growth. Cancer Cell 2007;11:37–51.
- Yang M, Brackenbury WJ. Membrane potential and cancer progression. Front Physiol 2013;4:185.

- 5. Cone CD Jr, Tongier M Jr. Control of somatic cell mitosis by simulated changes in the transmembrane potential level. Oncology 1971;25:168–182.
- Lobikin M, Chernet B, Lobo D, et al. Resting potential, oncogene-induced tumorigenesis, and metastasis: the bioelectric basis of cancer in vivo. Phys Biol 2012;9:065002.
- Wonderlin WF, Strobl JS. Potassium channels, proliferation and G1 progression. J Membr Biol 1996;154:91–107.
- Pardo LA, Stuhmer W. The roles of K(+) channels in cancer. Nat Rev Cancer 2014;14:39–48.
- Djamgoz MBA. Biophysics of cancer: Cellular excitability ("CELEX") hypothesis of metastasis. J Clin Exp Oncol;s1 . Epub ahead of print 2014. DOI: 10.4172/2324-9110.s1-005.
- Cabello M, Ge H, Aracil C, et al. Extracellular Electrophysiology in the Prostate Cancer Cell Model PC-3. Sensors ;19 . Epub ahead of print January 3, 2019. DOI: 10.3390/s19010139.
- Ribeiro M, Elghajiji A, Fraser SP, et al. Human Breast Cancer Cells Demonstrate Electrical Excitability. Front Neurosci 2020;14:404.
- 12. McCallum GA, Shiralkar J, Suciu D, et al. Chronic neural activity recorded within breast tumors. Sci Rep 2020;10:14824.
- Djamgoz MBA, Fraser SP, Brackenbury WJ. In Vivo Evidence for Voltage-Gated Sodium Channel Expression in Carcinomas and Potentiation of Metastasis. Cancers ;11
 Epub ahead of print 2019. DOI: 10.3390/cancers11111675.
- Fraser SP, Tesi A, Bonito B, et al. Potassium Channel Blockage and Invasiveness of Strongly Metastatic Prostate and Breast Cancer Cells. Bioelectricity 2021;3:215–220.

- 15. Zhou Y, Wong CO, Cho KJ, et al. Membrane potential modulates plasma membrane phospholipid dynamics and K-Ras signaling. Science 2015;349:873–876.
- Rizaner N, Onkal R, Fraser SP, et al. Intracellular calcium oscillations in strongly metastatic human breast and prostate cancer cells: control by voltage-gated sodium channel activity. Eur Biophys J 2016;45:735–748.
- Schwab A, Fabian A, Hanley PJ, et al. Role of ion channels and transporters in cell migration. Physiol Rev 2012;92:1865–1913.
- Yang M, James AD, Suman R, et al. Voltage-dependent activation of Rac1 by Nav 1.5 channels promotes cell migration. J Cell Physiol 2020;235:3950–3972.
- 19. Leslie TK, James AD, Zaccagna F, et al. Sodium homeostasis in the tumour microenvironment. Biochim Biophys Acta Rev Cancer 2019;1872:188304.
- Brisson L, Driffort V, Benoist L, et al. NaV1.5 Na(+) channels allosterically regulate the NHE-1 exchanger and promote the activity of breast cancer cell invadopodia. J Cell Sci 2013;126:4835–4842.
- Sundelacruz S, Moody AT, Levin M, et al. Membrane Potential Depolarization Alters Calcium Flux and Phosphate Signaling During Osteogenic Differentiation of Human Mesenchymal Stem Cells. *Bioelectricity* 2019;1:56–66.
- Vodnala SK, Eil R, Kishton RJ, et al. T cell stemness and dysfunction in tumors are triggered by a common mechanism. Science 2019;363:eaau0135.
- Eil R, Vodnala SK, Clever D, et al. Ionic immune suppression within the tumour microenvironment limits T cell effector function. Nature 2016;537:539–543.
- 24. Ong ST, Ng AS, Ng XR, et al. Extracellular K+ Dampens T Cell Functions: Implications for Immune Suppression in the Tumor Microenvironment. Bioelectricity 2019;1:169–179.

- 25. Trebak M, Kinet J-P. Calcium signalling in T cells. Nat Rev Immunol 2019;19:154–169.
- Cahalan MD, Chandy KG. The functional network of ion channels in T lymphocytes. Immunol Rev 2009;231:59–87.
- Beeton C, Wulff H, Barbaria J, et al. Selective blockade of T lymphocyte K(+) channels ameliorates experimental autoimmune encephalomyelitis, a model for multiple sclerosis.
 Proc Natl Acad Sci U S A 2001;98:13942–13947.
- Liu Q-H, Fleischmann BK, Hondowicz B, et al. Modulation of Kv channel expression and function by TCR and costimulatory signals during peripheral CD4(+) lymphocyte differentiation. J Exp Med 2002;196:897–909.
- 29. Kuras Z, Yun Y-H, Chimote AA, et al. KCa3.1 and TRPM7 channels at the uropod regulate migration of activated human T cells. PLoS One 2012;7:e43859.
- Launay P, Cheng H, Srivatsan S, et al. TRPM4 regulates calcium oscillations after T cell activation. Science 2004;306:1374–1377.
- 31. Hara Y, Wakamori M, Ishii M, et al. LTRPC2 Ca2+-permeable channel activated by changes in redox status confers susceptibility to cell death. Mol Cell 2002;9:163–173.
- 32. Xu H, Lai W, Zhang Y, et al. Tumor-associated macrophage-derived IL-6 and IL-8 enhance invasive activity of LoVo cells induced by PRL-3 in a KCNN4 channeldependent manner. BMC Cancer 2014;14:330.
- Xu R, Li C, Wu Y, et al. Role of KCa3.1 Channels in Macrophage Polarization and Its Relevance in Atherosclerotic Plaque Instability. Arterioscler Thromb Vasc Biol 2017;37:226–236.
- 34. Ohya S, Matsui M, Kajikuri J, et al. Increased Interleukin-10 Expression by the Inhibition of Ca2+-Activated K+ Channel KCa3.1 in CD4+CD25+ Regulatory T Cells in the Recovery Phase in an Inflammatory Bowel Disease Mouse Model. J Pharmacol Exp

Ther 2021;377:75–85.

- Cruse G, Duffy SM, Brightling CE, et al. Functional KCa3.1 K+ channels are required for human lung mast cell migration. Thorax 2006;61:880–885.
- Shao Z, Makinde TO, Agrawal DK. Calcium-Activated Potassium Channel KCa3.1 in Lung Dendritic Cell Migration. *American Journal of Respiratory Cell and Molecular Biology* 2011;45:962–968.
- Capatina AL, Lagos D, Brackenbury WJ. Targeting Ion Channels for Cancer Treatment: Current Progress and Future Challenges. Rev Physiol Biochem Pharmacol;2020/09/01.
 Epub ahead of print September 1, 2020. DOI: 10.1007/112_2020_46.
- Nuccitelli R. Application of Pulsed Electric Fields to Cancer Therapy. Bioelectricity 2019;1:30–34.
- Voloshin T, Schneiderman RS, Volodin A, et al. Tumor Treating Fields (TTFields)
 Hinder Cancer Cell Motility through Regulation of Microtubule and Acting Dynamics.
 Cancers ;12 . Epub ahead of print October 17, 2020. DOI: 10.3390/cancers12103016.
- Payne SL, Levin M, Oudin MJ. Bioelectric Control of Metastasis in Solid Tumors. Bioelectricity 2019;1:114–130.
- 41. Venkatesh HS, Morishita W, Geraghty AC, et al. Electrical and synaptic integration of glioma into neural circuits. Nature 2019;573:539–545.
- Rühl P, Langner JM, Reidel J, et al. Monitoring of compound resting membrane potentials of cell cultures with ratiometric genetically encoded voltage indicators. Commun Biol 2021;4:1164.
- 43. Quicke P, Sun Y, Arias-Garcia M, et al. Membrane voltage fluctuations in human breast cancer cells. *bioRxiv* 2021;2021.12.20.473148.

44. Silver BB, Zhang SX, Rabie EM, et al. Substratum stiffness tunes membrane voltage in mammary epithelial cells. J Cell Sci . Epub ahead of print June 11, 2021. DOI: 10.1242/jcs.256313.



Figure 1. The membrane potential (V_m) scale across different cell types. Scale adapted from ⁴.



Figure 2. Altered membrane potential (V_m) in the tumor microenvironment (TME). Depolarized V_m in both cancer and immune cells in the TME can promote cancer progression by different mechanisms. PD-1: programmed cell death protein 1; PD-L1: programmed death-ligand 1. Figure created with BioRender.com.



Figure 3. Summary of the key features of tumor development proposed to be regulated by the V_m .