

This is a repository copy of Exploring the Conformational Landscape and Stability of Aurora A Using Ion-Mobility Mass Spectrometry and Molecular Modeling.

White Rose Research Online URL for this paper: <a href="https://eprints.whiterose.ac.uk/183559/">https://eprints.whiterose.ac.uk/183559/</a>

Version: Supplemental Material

#### Article:

Tomlinson, LJ, Batchelor, M orcid.org/0000-0001-6338-5698, Sarsby, J et al. (5 more authors) (2022) Exploring the Conformational Landscape and Stability of Aurora A Using Ion-Mobility Mass Spectrometry and Molecular Modeling. Journal of the American Society for Mass Spectrometry, 33 (3). pp. 420-435. ISSN 1044-0305

https://doi.org/10.1021/jasms.1c00271

© 2022 American Society for Mass Spectrometry. Published by American Chemical Society. All rights reserved. This is an author produced version of an article published in Journal of The American Society for Mass Spectrometry. Uploaded in accordance with the publisher's self-archiving policy.

### Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

#### **Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



## **Supporting Information**

# Exploring the conformational landscape and stability of Aurora A using ion-mobility mass spectrometry and molecular modelling

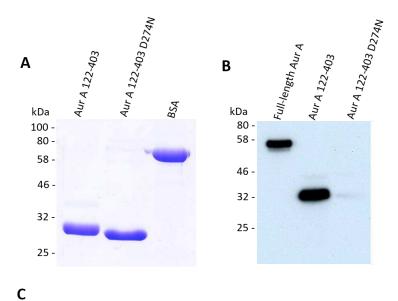
Lauren J. Tomlinson<sup>1, 3</sup>, Matthew Batchelor<sup>2</sup>, Joscelyn Sarsby<sup>1</sup>, Dominic P. Byrne<sup>3</sup>, Philip Brownridge<sup>1</sup>, Richard Bayliss<sup>2</sup>, Patrick A. Eyers<sup>3</sup> and Claire E. Eyers<sup>1, 3\*</sup>.

<sup>&</sup>lt;sup>1</sup> Centre for Proteome Research, Department of Biochemistry & Systems Biology, Institute of Systems, Molecular & Integrative Biology, University of Liverpool, Crown Street, Liverpool, L69 7ZB, U.K.

<sup>&</sup>lt;sup>2</sup> Astbury Centre for Structural Molecular Biology, School of Molecular and Cellular Biology, Faculty of Biological Sciences, University of Leeds, Leeds, LS2 9JT, U.K.

<sup>&</sup>lt;sup>3</sup> Department of Biochemistry & Systems Biology, Institute of Systems, Molecular & Integrative Biology, University of Liverpool, Crown Street, Liverpool, L69 7ZB, U.K.

<sup>\*</sup>Correspondence: Claire E. Eyers (ceyers@liverpool.ac.uk)



Peptide sequence	Modification	Charge	m/z (Da)	DeltaM (ppm)	lon score	ptmRS
EVEIQsHLR	S186 (Phospho)	2	595.78	1.16	22	100
ELQKLsKFDEQR	S226 (Phospho)	3	534.26	1.89	32	100
DIKPENLLLGsAGELK	S266 (Phospho)	2	888.96	0.99	61	100
IADFGWsVHAPSSR	S278 (Phospho)	2	805.36	1.05	73	100
RttLAGTLDYLPPEmIEGR	T287 (Phospho); T288 (Phospho); M300 (Oxidation)	2	1155.01	-3.84	43	99.75; 100

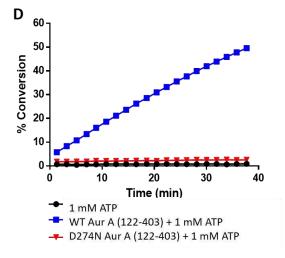
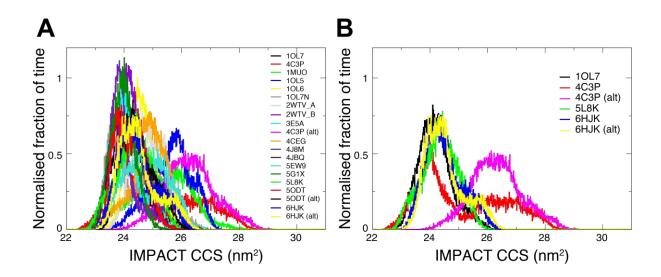


Figure S1. Expression and purification of wild-type (WT) phosphorylated Aur A (122-403) and catalytically inactive D274N Aur A (amino acids 122-403). (A) SDS-PAGE gel of purified WT and D274N Aur A (122-403) proteins. His-Aur A proteins were purified using a Nickel HisTrap HP column, prior to tag cleavage and gel filtration. Aur A (122-403) has a predicted MW of 32 kDa. BSA was used as a control (B) Immunoblot of full-length Aur A (1-403), WT and D274N Aur A (122-403). Phosphorylation at the activation site in the T-loop (T288) was evaluated using an anti-phospho T288 antibody (CST); 200 ng of protein was loaded per well. (C) LC-MS/MS phosphosite mapping of WT Aur A detailing the peptide sequence identified, the site(s) of modification and the confidence associated with both (MASCOT lon score and ptmRS scores respectively). Data is representative of three independent experiments. (D) Enzyme activity assay of WT and D274N Aur A (122-403) in the presence of 1 mM ATP (10 ng protein, 2  $\mu$ M substrate peptide).



**Figure S2. Collision cross section distributions from all-atom simulations of Aur A.** Overlaid normalised distributions for (A) all simulations, and (B) example structures 1OL7 (DFG-in), 5L8K (DFG-up), 6FHK (DFG-out) and 4C3P (DFG-in, A-loop out). The bulk of trajectories contribute to a first major peak at ~24 nm² and extend between 22.5 and 27 nm². A handful of models, particularly those of 4C3P, extend the range to higher CCS values contributing to a second peak and the tail of the CCS distribution.

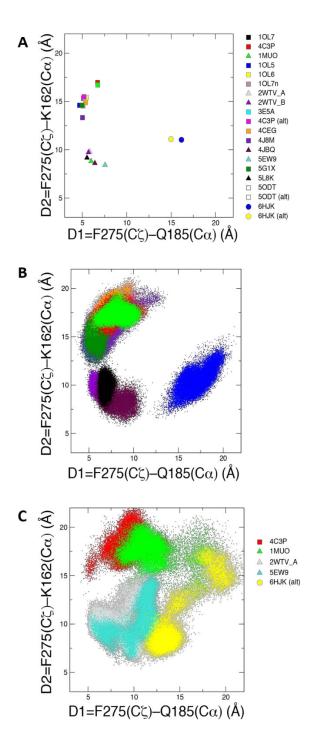


Figure S3. Designation of DFG motif conformations using the D1/D2 criteria introduced by Modi and Dunbrack (PNAS, 2019, 10.1073/pnas.1814279116). (A) Positions in the D1/D2 plot for each of the starting structures. Three clear groups are observed: the DFG-in group of 12 structures (squares) positioned in the upper left quadrant; the DFG-up/inter group of 6 structures (triangles) in the lower left quadrant; and the two DFG-out 6HJK structures (circles) in the lower right quadrant. (B, C) The range of the D1/D2 plot explored for each simulation. In (B), the majority of simulations remain reasonably close to their initial position and maintain their DFG conformation. In (C), five trajectories are more mobile on the D1/D2 plot and exhibit DFG motif conformations further away from their initial structure, including some conformations in the upper right quadrant of the D1/D2 plot that are not observed in kinase crystal structures.

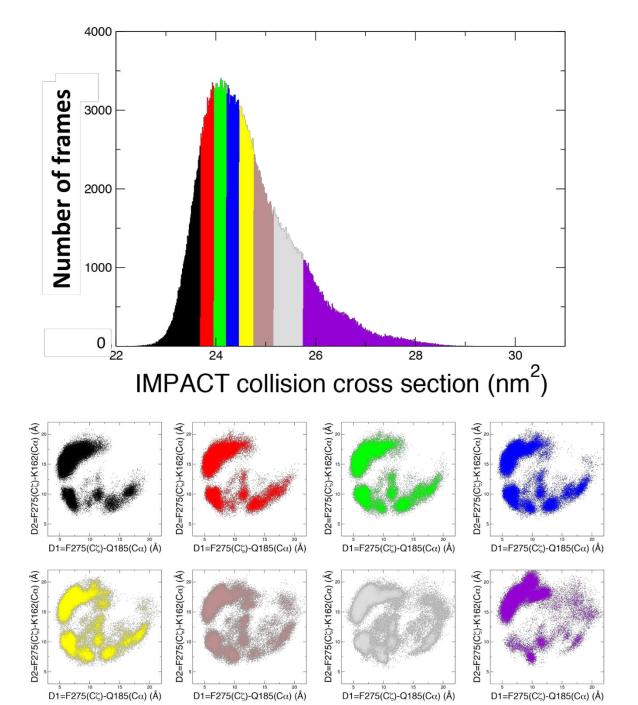


Figure S4. Collision cross section distributions for the combined all-atom simulation trajectories of Aur A. The distribution has been divided up into eight CCS regions with ~80,000 structures in each region. The contribution of different DFG motif conformations in each region is described using a D1/D2 plot. A high density of DFG-in models is observed in all eight regions on the CCS profile, as shown by the density of structures in the upper left quadrant of the D1/D2 diagram. DFG-in is the most common DFG motif conformer of the initial structures (12 of 20) and includes the A-loop extended 4C3P model. DFG-up/inter and DFG-out conformers are also seen across all eight regions but are less frequently observed at the highest CCS regions. The upper-right quadrant of the D1/D2 plot is more frequently occupied in the highest CCS regions, suggesting that uncategorised DFG motif conformers (not observed in crystal structures) with a more open structure contribute to the high CCS range of the profile.

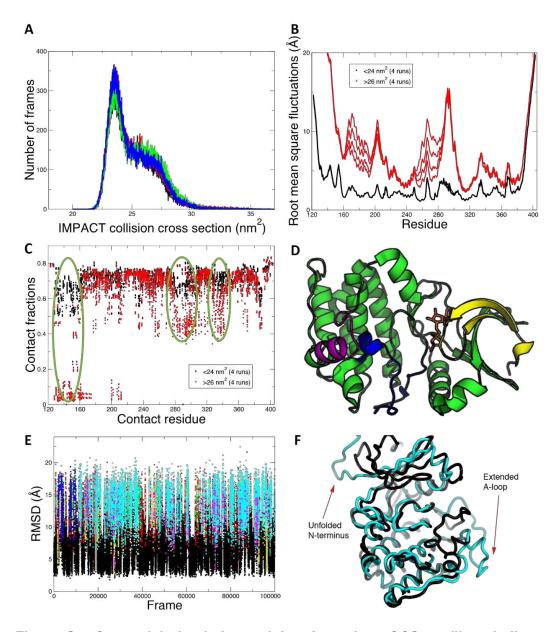
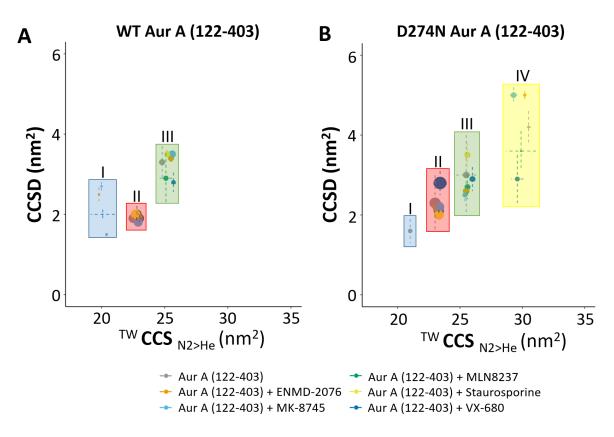


Figure S5. Gō model simulations of Aur A produce CCS profiles similar to those of experimental IM-MS conformers II and III, and suggest that structures with an extended A-loop and unfolded N-terminus may contribute to the higher CCS (conformer III) region. (A) Overlaid CCS distributions for 4 independent Gō model simulations display the two-component profile akin to those of the combined all-atom simulations and the experimental CCS distributions for conformer II and III. Simulation trajectory frames were separated out into first peak (<24 nm², mainly 'conformer II') and second peak (>26 nm², mainly 'conformer III') groups and then analysed to give: the root mean square fluctuations (RMSF) (B), and the native contact fractions (C). RMSF values are generally higher, marking more variable regions/structures, for the conformer III type (red) ensemble of structures compared to the conformer II-type (black) structures, but especially so in the N-lobe and around the A-loop region. Native contacts are also most likely broken for conformer III-type structures at the N-terminus, and the A-loop and contacting regions, highlighted in (D). An RMSD-based cluster analysis for one of the trajectories, gave representative structures of well-populated clusters with an unfolded N-terminus and extended A-loop region (cyan, E, F).



**Figure S6. Conformational space adopted by Aur A.** (A, B) Proportional scatter plots (CCS (nm²) versus CCSD (nm²)) for the different conformational states (as determined by Gaussian fitting in Fig. 5) for WT (A) and D274N (B) Aur A (122-403). Size of dot representative of area.

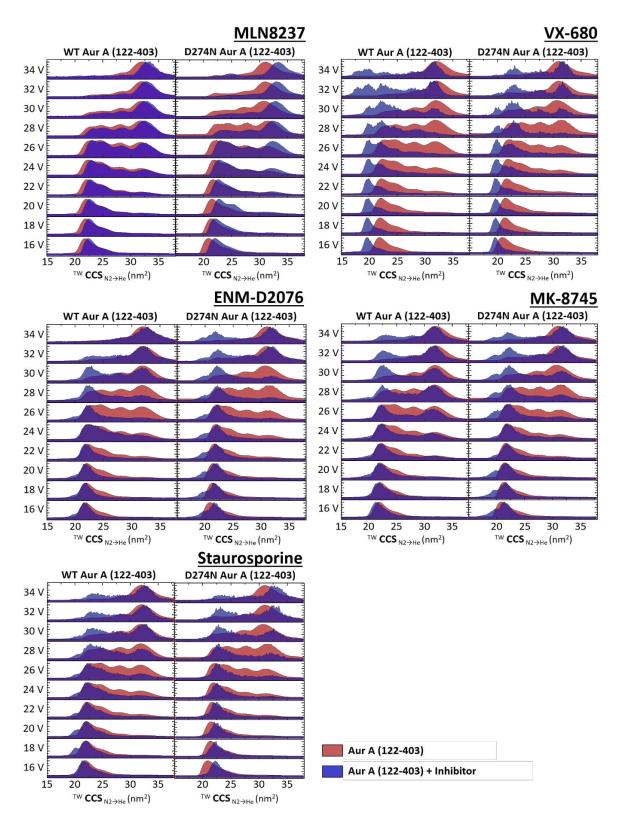


Figure S7. Collision-induced unfolding profiles of Aur A in the absence and presence of inhibitors. The isolated 11+ charge state of WT (left) and D274N (right) Aur A (122-403) in the presence (blue) or absence (red) of 10-fold molar excess of inhibitor were subject to CIU using a stepped collision energy (CE) between 16 and 34 V (two-volt intervals). Data analysis was carried out in MassLynx 4.1, generating mountain plots using Origin (Version 2016). Presented are data from an average of three replicates.

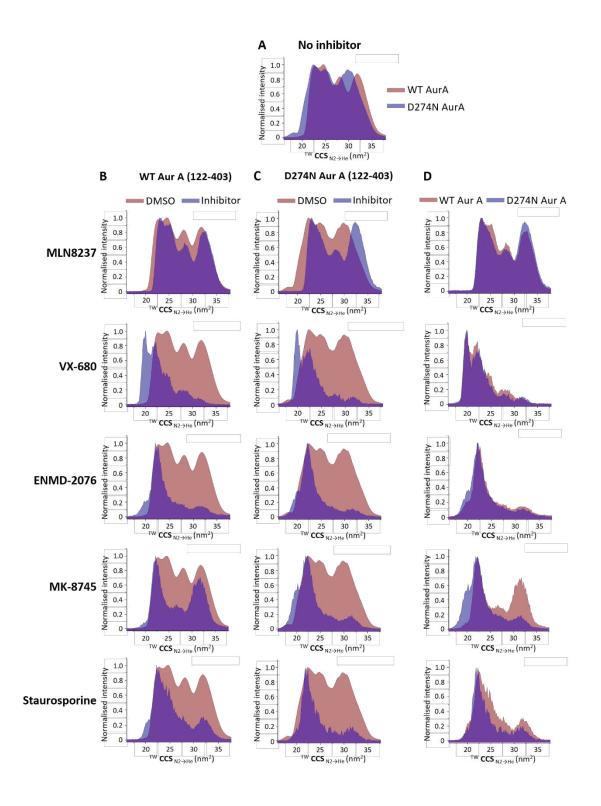


Figure S8. Collision-induced unfolding profiles of Aur A in the absence and presence of inhibitors. The isolated 11+ charge state at 26 V collision energy of (A) Aur A (122-403) with no inhibitor. (B) WT Aur A (122-403) and (C) D274N Aur A (122-403) in the presence (blue) or absence (red) of 10-molar excess of inhibitor. (D) Aur A (122-403) in the presence of inhibitor for WT (red) and D274N (blue). Data analysis was carried out in MassLynx 4.1 and line plots were generated using Origin (Version 2016). Presented are data from an average of three replicates.

PDB	<sup>1</sup> CCS (nm <sup>2</sup> )	Crystal description	
1OL5	22.6-23.0	phos, TPX2, DFG-in	
1OL6	22.3-22.8	dephos, D274N, DFG-in	
10L7	22.2–22.7	phos, DFG-in	
10L7N	22.1–22.6	based on 1OL7, dephos, D274N, DFG-in	
3E5A	22.5–22.9	phos, VX-680, TPX2, DFG-in	
4C3P	25.0–25.5	dephos, TPX2, dimer, DFG-in, A-loop open	
4C3P (alt.)	24.3–24.8	dephos, TPX2, dimer, DFG-in, A-loop open	
4J8M	23.1–23.7	dephos, T287D, CD532, DFG-in	
4CEG	21.1–21.6	phos, C290A, C393A, DFG-in	
5G1X	21.8–22.2	phos, N-MYC, DFG-in	
5ODT	23.2–23.7	dephos, D274N, TACC3, DFG-in	
5ODT (alt.)	23.9–24.3	dephos, D274N, TACC3, DFG-in	
1MUO	23.2-23.6	dephos, adenosine, DFG-up	
2WTV	23.4–23.7	phos, MLN8054, chain A, DFG-up	
2WTV	22.6-23.0	phos, MLN8054, chain B, DFG-up	
4JBQ	21.9–22.5	dephos, VX-680, DFG-up	
5EW9	22.8–23.4	phos, MK-5108, DFG-up	
5L8K	22.3–22.7	phos, vNAR, DFG-up	
6HJK	22.9–23.4	dephos, L210C, ASDO2, DFG-out	
6HJK (alt.)	23.0–23.4	dephos, L210C, ASDO2, DFG-out	
4F5S	39.3–40.1	BSA, calibrant	

Table S1. Collision cross section estimates made using IMPACT on a single initial structural model of Aur A (122–403) for each of the listed PDB codes. DFG-in models are grouped in blue, DFG-up (or inter) models are grouped in yellow, and two alternatively built starting structures for the only available DFG-out Aur A structure (6HJK) are green.

<sup>&</sup>lt;sup>1</sup> As IMPACT is a stochastic method, CCS estimates were performed 200 times on each structure to give a range of values.

Inhibitor	Mass (Da)	Structure	Binding configuration
MLN8237 (Alisertib)	518.92		DFG-out partial (TR-FRET)
(Allsertib)		HO, NH	Closely-related MLN8054
			DFG-up (X-ray; PDB 2WTV)
		CIFN	DFG-in (X-ray; PDB 2WTW)
VX-680	464.59	N-NH	DFG-out partial (TR-FRET)
(Tozasertib)		HZ N S	DFG-in with TPX2 (X-ray; PDB 3E5A)
			DFG-up without TPX2 (X-ray; PDB 4JBQ)
ENMD-2076	375.47	T N N N N N N N N N N N N N N N N N N N	DFG-in (TR-FRET)
MK-8745	431.91	O N N N N N N N N N N N N N N N N N N N	DFG-out (TR-FRET)
Staurosporine	466.53	HN O	ND

Table S2. Aur A inhibitors employed in this study. ND: Not Determined