

This is a repository copy of *How the insect central complex could coordinate multimodal navigation*.

White Rose Research Online URL for this paper: https://eprints.whiterose.ac.uk/181601/

Version: Accepted Version

Article:

Sun, X., Yue, S. and Mangan, M. orcid.org/0000-0002-0293-8874 (2021) How the insect central complex could coordinate multimodal navigation. eLife, 10. e73077.

https://doi.org/10.7554/elife.73077

© 2021 The Authors. This article is distributed under the terms of the Creative Commons Attribution License permitting unrestricted use and redistribution provided that the original author and source are credited (http://creativecommons.org/licenses/by/4.0/).

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here: https://creativecommons.org/licenses/

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/

How the insect central complex could coordinate multimodal navigation

Xuelong Sun^{1,2}, Shigang Yue^{2,1†}, Michael Mangan^{3†}

- ⁵ ¹Machine Life and Intelligence Research Centre, School of Mathematics and Information
- Science, Guangzhou University, China; ²Computational Intelligence Lab & L-CAS, School
- of Computer Science, University of Lincoln, United Kingdom; ³Sheffield Robotics,
- [®] Department of Computer Science, University of Sheffield, Sheffield, United Kingdom
- Abstract The central complex of the insect midbrain is thought to coordinate insect guidance
 strategies. Computational models can account for specific behaviours but their applicability
 across sensory and task domains remains untested. Here we assess the capacity of our previous
 model (*Sun et al., 2020*) of visual navigation to generalise to olfactory navigation and its
 coordination with other guidance in flies and ants. We show that fundamental to this capacity is
 the use of a biologically-plausible neural copy-and-shift mechanism that ensures sensory
 information is presented in a format compatible with the insect steering circuit regardless of its
- Information is presented in a format compatible with the insect steering circuit regardless of it source. Moreover, the same mechanism is shown to allow the transfer cues from
- source. Moreover, the same mechanism is shown to allow the transfer cues from
 unstable/egocentric to stable/geocentric frames of reference providing a first account of the
- mechanism by which foraging insects robustly recover from environmental disturbances. We
- propose that these circuits can be flexibly repurposed by different insect navigators to address
- their unique ecological needs.
- Introduction

22

23

7

Recently, it has been proposed that the repertoire of robust navigation behaviours displayed by 24 insects (Webb and Wystrach, 2016; Wehner, 2019) can be traced to the well conserved brain region 25 known as the central complex (CX) (Honkanen et al., 2019: Hulse et al., 2021). The evidence to 26 support this hypothesis includes: the discovery of the insect head-direction system in the CX that 27 tracks the animal's current heading relative to external (Heinze, 2014; Seelig and Jayaraman, 2015; 28 Kim et al., 2019: Hardcastle et al., 2021) or self-motion (Green et al., 2017: Turner-Evans et al., 29 2017) cues; the innervation of the fan-shaped body (FB) region of the CX with sensory information 30 relevant to different orientation strategies (Hu et al., 2018: Franconville et al., 2018: Hulse et al., 31 2021; Shiozaki et al., 2020); the well-preserved columnar structure that is well suited to computing 32 desired headings for vector navigation tasks (Stone et al., 2017; Honkanen et al., 2019; Le Moël 33 et al.. 2019: Lvu et al., 2020); and the identification of a neural steering circuit in the FB capable of 34 computing motor commands that reduce the offset between the current heading and a desired 35 heading (Stone et al., 2017; Honkanen et al., 2019; Rayshubskiy, 2020). Computational models 36 of this architecture have produced realistic path integration (Stone et al., 2017; Gkanias et al., 37 2019) and trap-lining behaviours (Le Moël et al., 2019), and simple conceptual extensions have been outlined that could account for long-distance migratory behaviour (Honkanen et al., 2019).

40 Yet, for the CX to be considered a general navigation centre, it must additionally be capable of

*For correspondence:

syue@lincoln.ac.uk; m.mangan@sheffield.ac.uk; xsun@lincoln.ac.uk

[†]Joint last authorship

- (i) generating of gradient ascent/descent behaviours that rely on spatially-varying but rotationally-
- invariant sensory cues (e.g. odour gradients) (ii) co-ordinating competing guidance systems into a
 - single meaningful motor command (iii) generalise across sensory modalities and task spaces.

We recently demonstrated how the steering circuit could be adapted to ascent gradients of visual familiarity when augmented by a neural *'copy-and-shift'* mechanism that converts temporal changes in spatially sampled sensory information into an orientation signal (*Sun et al., 2020*).

Specifically the mechanism firstly conject the animal's current heading from the head direction cells

in the protocerebral bridge (PB) to desired heading networks in the FB. At the same time the signal

- undergoes a lateral *shift* in proportion to any undesired change in sensory valence as measured
- by the MB output neurons (Aso et al., 2014: Li et al., 2020: Hulse et al., 2021). Thus, the animal will
- 51 continue on its current heading until an undesirable change in sensory valence is experienced at
- ⁵² which point the shift mechanism will create an offset between the current and desired headings
- ⁵³ causing the steering circuit to initiate a change of direction. The architecture of the CX already pos-
- sesses neural substrates ideally suited for both the 'copy' and shift' functions: head direction cells
- are known to transmit their output into the ring structures of the central body (*Stone et al., 2017*;
 Honkanen et al., 2019) as needed for *copy* stage: and neural mechanisms that laterally shift the
- Honkanen et al., 2019) as needed for copy stage; and neural mechanisms that laterally shift the head direction cells in response to sensory feedback (e.g. the self-motion cues (Turner-Evans et al.,
- ⁵⁷ head direction cells in response to sensory feedback (e.g. the self-motion cues (*Turner-Evans et al.*,
 2017: Green et al., 2017), the visual cues (*Kim et al.*, 2019: Fisher et al., 2019)) are well established
- 2017; Green et al., 2017), the visual cues (*Kim et al., 2019*; Fisher et al., 2019)) are well established
 as required for the *shift* stage. Crucially, the complete '*copy-and-shift*' mechanism explains how the
- ⁶⁰ CX steering circuit (see *Figure 1*) could exploit sensory gradients that provide no instantaneous
- orientation information for navigation.

43

We also demonstrated neural mechanisms that coordinate between different guidance strate-62 gies (Sun et al., 2020). Specifically we added a contextual-switching mechanism (see Figure 1) that 63 triggers specific guidance strategies depending on the context, e.g. switching from path integration 64 unfamiliar surroundings to visual route-following in familiar terrain. As a final stage, we revealed 65 how ring attractor circuits (Touretzky, 2005; Sun et al., 2018) (see Figure 1) that we hypothesise 66 exist in the fan-shaped body provide an ideal substrate for optimally integrating cues that exist 67 within a shared context (e.g. path integration and visual homing in unfamiliar contexts). The 'copy-68 and-shift' mechanism again plays a crucial role in this capacity as it "transfers" orientation outputs 69 into a shared frame of reference. For example, when ascending gradients temporal changes in vi-70 sual familiarity are translated into heading commands relative to the head-direction system which 71

then share a frame of reference with the path integration system.

This biologically-constrained model of the insect midbrain was shown capable of generating 73 realistic visual navigation behaviours of desert ants through the coordinated action of visual route 74 following (RF), visual homing (VH) and path integration (PI) modules partially addressing two of the 75 requirements listed above (Sun et al., 2020). In this study, we extend our analysis of the model, and 76 in particular the 'copy-and-shift' mechanism, to assess if it can address the latter issue of generalisa-77 tion across and between sensory and task domains. The following sections first assess whether the 78 model can be easily reapplied to the olfactory tasks of chemotaxis and odour-gated anemotaxis 70 (plume-following) in laboratory-like settings. We then probe whether the same integration mech-80 anisms can generalise to odour-gated switching in both flies and desert ants. Finally, we provide 81 the first account of how the central complex could transfer orientation cues from an egocentric to 82 a geocentric frame of reference which we propose can enhance the robustness of navigation. 83

84 Results

- ⁸⁶ Here we assess the ease with which our visual navigation model generalises to olfactory navigation
- 87 tasks.

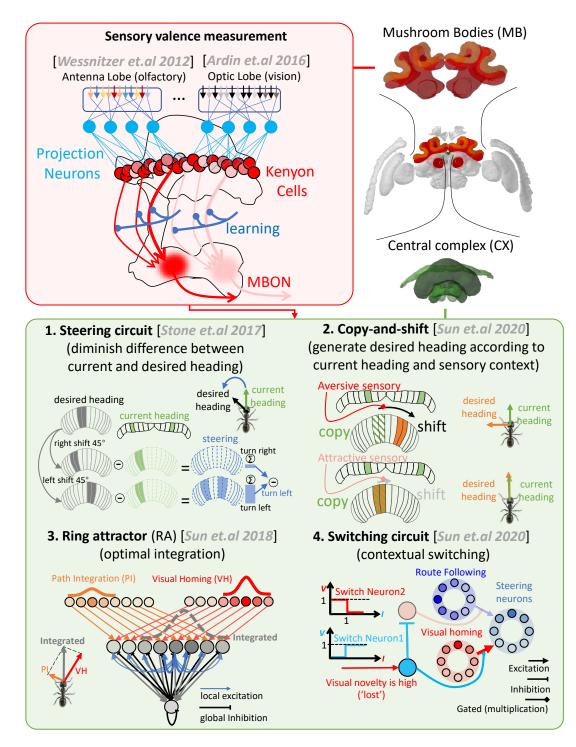


Figure 1. Schematic overview of the MB-CX model first presented in (*Sun et al., 2020*) and re-applied here to multimodal guidance.. The upper right panel depicts the two key brain areas modelled (Mushroom bodies in red, Central in green). The upper left panel (red background) outlines the role of the MBs in measuring valence of odour (*Wessnitzer et al., 2012*) and visual (*Ardin et al., 2016*) cues. The lower panel (green background) introduces the 4 CX sub-circuits integrated in the previous model. (1) The steering circuit proposed to exist in the CPU1 neurons of the CX that computes the turning angle that minimises the difference between the current heading (from the PB) and desired heading (from the CPU4 cells) (*Stone et al., 2017*). (2) The copy-and-shift mechanism creates a desired heading from non-directional cues by simply copying the current heading and shifting it in proportion to the change in sensory valance.

Figure 1 (continued). (3) Ring attractor networks can automatically and optimally integrate orientation cues from disparate sources into a single readout. Our model uses RAs to integrate both compass and desired heading signals. (4) Context-dependent switches multiplex systems at a high level (e.g when 'lost' engages visual homing (VH) but not route following (RF)). Images of the brain regions are adapted from the insect brain database (*Heinze et al., 2021*)-https://www.insectbraindb.org.

88 Chemotaxis of odour gradients

Adult and larvae fruit-flies readily climb rewarding odour gradients by modulating their heading 89 direction in direct response to the temporal change in odour concentration (Gomez-Marin et al., 2010: Nagel and Wilson, 2011: Kim et al., 2011: Schulze et al., 2015: Jung et al., 2015) mirroring our model's approach to visual homing. Moreover, the neural pathways of olfactory processing are well established and only differ from our model in their sensory origins (antennal lobe (AL) to the lateral horn (I H) (Gupta and Stopfer, 2012; Roussel et al., 2014) and mushroom bodies (MBs) (Aso et al., 2014; Hulse et al., 2021)) before connecting to the CX through direct or indirect (hypo-95 thetically via superior medial protocerebrum (SMP) (Plath et al., 2017: Hulse et al., 2021: Li et al., 2020)) neural pathways. Thus by simply changing the input from optic to antennal lobes and the 97 processing region from the MB to the LH and MB (see *Figure 2*A (left panel)) our model is able to 98 adapt its heading to align with the positive odour gradient over successive steps (see Figure 2B (left 90 panel)). Note that here we simply take the valence output of the MB as the odour concentration, 100 buy any other equivalent measurement (such as the degree of attraction) could work along with 101 the 'copy-and-shift' mechanism. *Figure 2*C (left panel) demonstrates the realistic chemotaxis be-102 haviour generated by the model in a classic 'volcano' environment (lung et al., 2015: Schulze et al., 103 2015). Figure Supplement 1 provides similarly realistic paths in other odour landscapes. It should 104 be noted that there are neural pathways not included in the model that directly link odour input 105 to motor outputs that may play a role in chemotactic guidance (Green et al., 2019: Rayshubskiv, 106 2020: Scaplen et al., 2021). Indeed while larvae possess a MB and LH assemblies they do not a fully 107 developed CX as modelled here (Ibrahim et al., 2018; Gowda et al., 2021). Analysis of behavioural 108 deficiencies in animals with CX-knockouts would offer crucial insights into the role of the CX for 109 chemotactic behaviours. 110

Anemotaxis in odour plumes

In moving air-flows adult fruit flies pinpoint olfactory sources by anemotaxis whereby individuals 112 align with the upwind direction allowing them to approach the hidden odour source (Kennedy and 113 Marsh. 1974: Rutkowski et al., 2009: van Breugel and Dickinson, 2014). Insects sense wind direction 114 through deflections of their antennae (Yorozu et al., 2009: Patella and Wilson, 2018: Okubo et al., 115 2020) with the wedge projection neurons (WPNs) converting their inputs (via antennal mechanosen-116 sory & motor centre (AMMC) pathway in *Figure 2*B (right panel)) into a direction relative to the 117 animals current heading (Suver et al., 2019) (see Figure Supplement 2). The WPN output is then 118 transmitted to the FB of the CX via the lateral accessory lobe (LAL) -> noduli (NO) pathway (Hulse 119 et al., 2021: Matheson et al., 2021) (Figure 2B (right panel)). The 'copy-and-shift' mechanism again 120 provides the ideal bridge between input signal and steering circuit. By simply driving the direction 121 and magnitude of the 'shift' by the WPN response when a rewarding odour is detected (Figure 2A 122 (right panel)) the model turns the agent upwind (see *Figure 2*B (right panel)). *Figure 2*C (right panel) 123 shows an example path of a simulated fly navigating a classic laboratory environment with an 124 odour plume into which rewarding odour is toggled ON and OFF (for a simulation of a group agents 125 see Figure Supplement 3), which demonstrates realistic odour-driven anemotaxis behaviour. 126 Taken together the above data demonstrates the capacity of the model to generalise from visual 127

to olfactory navigation without significant alteration.

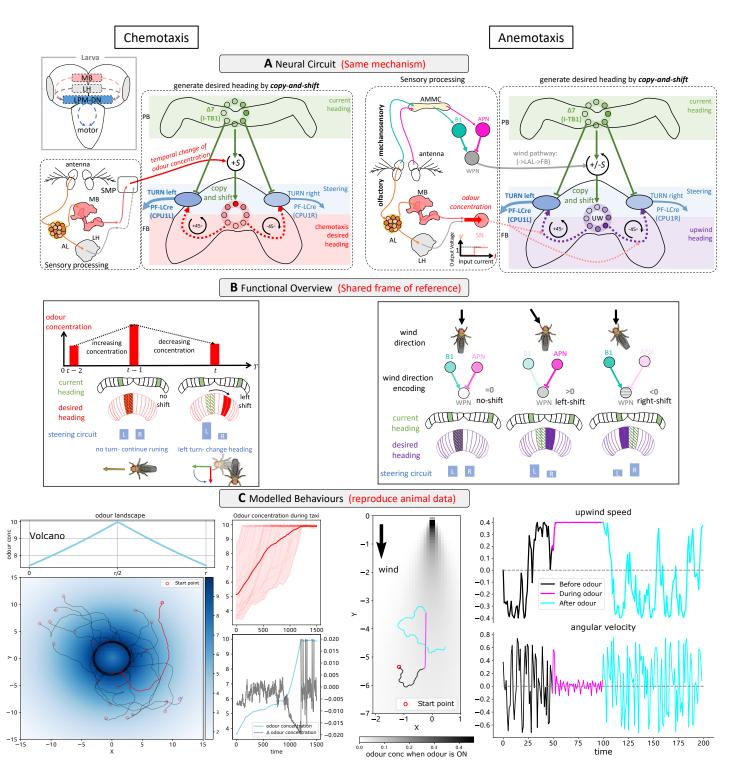


Figure 2. Modelling olfactory navigation in flies using a 'copy-and-shift' mechanism: chemotaxis (left side) and anemotaxis (right side). (A): Schematic diagrams of the neural circuits generating current-desired heading pairings for chemotaxis and anemotaxis. The *copy-and-shift* mechanism is only different in how the shift is realised: for chemotaxis, the temporal change of the odour concentration produces turns of different magnitude in a predefined direction, which for anemotaxis the wedge projection neuron (WPN) provide both turning magnitude and direction to steer the animal upwind. The corresponding hypothesised functional map of larvae brain is inserted in the left panel showing that the olfactory descending neurons LPM-DN may play similar role as the CX. *Figure 2 continued on next page*

Figure 2 (continued). (B): Schematic diagram explaining the model functions. For chemotaxis, decreasing odour concentration will shift the desired heading from current heading causing the steering circuit to initiate a turn. For anemotaxis, the WPN neurons subtract the activation of the antennal mechanosensory and motor centre (AMMC) projection neuron (APN) from that of B1 that directly shifts the desired heading to align with the upwind direction. Note that the two mechanisms share a frame of reference. (C): Example behaviours generated by the model. Realistic chemotaxis behaviour is shown (left) in a 'Volcano' odour landscape. On the right, realistic anemotaxis (magenta path segment) are shown when odour is 'ON' vs undirected motion (black and cyan path segments) when odour is 'OFF'. Upwind speed and angular velocity of the example agent are shown on the right panel. Note the obvious higher upwind translational velocity and low angular velocity during the presence of the odour indicates surges upwind.

Figure 2-Figure supplement 1. The simulation results of chemotaxis model with odour landscape of 'Linear'.

Figure 2-Figure supplement 2. Simulation of wind direction encoding.

Figure 2-Figure supplement 3. Simulation results of a group of agents (N = 20) driven by the odour-driven anemotaxis model.

¹²⁹ Coordination of guidance behaviours by linking frames of reference

- 130 With the model shown to generalise from visual to olfactory navigation tasks, we now assess it
- ability to co-ordinate guidance strategies across sensory domains.
- 132 Contextual switching between olfactory guidance behaviours
- 133 In reality insects utilise both the chemotaxis and anemotaxis strategies outlined above. Across
- species and environments (laminar odour gradient or turbulent odour plume), a distinct behavioural
- trigger is reported at the onset (ON-response) or loss (OFF-response) of sensory valence (moths
- 136 (Kennedy and Marsh, 1974; Rutkowski et al., 2009), flying fruit flies (van Breugel and Dickinson,
- 2014), walking flies (Steck et al., 2012; Bell and Wilson, 2016; Álvarez-Salvado et al., 2018)). Specifi-
- cally, in the presence of the attractive odour animals apply anemotaxis and surge upwind but when
- the attractive odour is lost they engage in a chemotactic-like search to recover the plume. This
- problem is analogous with the contextual switching using in our previous model to select between ON- and OFF-route navigation strategies (*Wystrach et al., 2012*), *Figure 3*A (left panel) depicts how
- ¹⁴¹ ON- and OFF-route navigation strategies (*Wystrach et al., 2012*). *Figure 3*A (left panel) depicts how ¹⁴² the CX switching circuit can be easily reconfigured to be triggered by the instantaneous change
- the CX switching circuit can be easily reconfigured to be triggered by the instantaneous change of odour concentration fitting with the reported ON- and OFF-responses (*Álvarez-Salvado et al.*,
- 2018). Note that we here assume that the ON- and OFF-response are driven by the output neu-
- rons of the odour processing brain regions (i.e., MBON or LHON) that could compute the temporal
- changes of odour concentration (Dolan et al., 2018; Hulse et al., 2021; Matheson et al., 2021). Fig-
- ure 3B (left panel) illustrates simulated ON- and OFF- responses that are supplied to the model and
- their behavioural consequence. *Figure 3*C (left panel) demonstrates realistic olfactory navigation
- behaviour similar to the behavioural data in *Álvarez-Salvado et al.* (2018). See also the simulation
- results of a 20-agents group demonstrating similar performance in *Figure Supplement 1*.

¹⁵¹ Optimally integrating navigation behaviours across sensory domains

In barren salt-pans, homing desert ants follow their path integrator to their pest area before re-152 lying on nest-odour plumes for their final approach (Buehlmann et al., 2012). Ants bypass the 153 nests of conspectics that diffuse similar odours (CO_2) until reaching the nest locale (**Buehlmann**) 154 et al., 2012) indicating use of a sophisticated integration strategy beyond simple switching outlined 155 above. Rather, ants instead appear to weight their PI output relative to the home-vector length in 156 a similar fashion to their integration of path integration and visual cues (Wystrach et al., 2015: 157 Legge et al., 2014) as was realised in our previous model using ring attractor networks (Touretzky, 158 2005: Sun et al., 2018, 2020). Figure 3A (right panel) depicts the augmentation of our odour-gated 150 anemotaxis model with a ring attractor circuit to optimally integrate PI and olfactory navigation 160 outputs. These adaptations are in accordance with the olfactory navigation mechanisms (chemo-161 taxis and anemotaxis) proposed to be used by ants by Wolf and Wehner (2000, 2005). Note that the 162 desired headings recommended by odour homing (OH, or chemotaxis) and upwind direction (UW, 163 or odour-gated anemotaxis) are gated by the OFF and ON response and weighted by the odour 164 concentration signal prior to being injected into the ring attractor to be combined with Pl. Figure 3B 165

- (right panel) illustrates how the various desired heading signals are optimally integrated by the ring 166
- attractor network before being sent as input to the steering circuit. *Figure 3C* shows homing paths 167
- generated by the model following simulated displacements left or right of the regular feeder which 168
- closely match those of real ants (*Buehlmann et al., 2012*). Note that there is an additional odour 169
- plume diffused by a simulated conspecific nest positioned near the release points which causes 170
- some distraction before the simulated ants continue to the real nest site. In the absence of the 17
- distractor nest paths are much more direct (see **Figure Supplement 3**) 172
- Taken together these data demonstrate that the CX possess the neural mechanisms to flexibly 173
- coordinate the various guidance behaviours observed in insects across sensory domains support-174
- ing its role as the navigation centre (Honkanen et al., 2019: Hulse et al., 2021). 175

A mechanism for transferring between orientation frames of reference 176

The optimal integration model detailed above is reliant on the *copy-and-shift* mechanism firstly 177

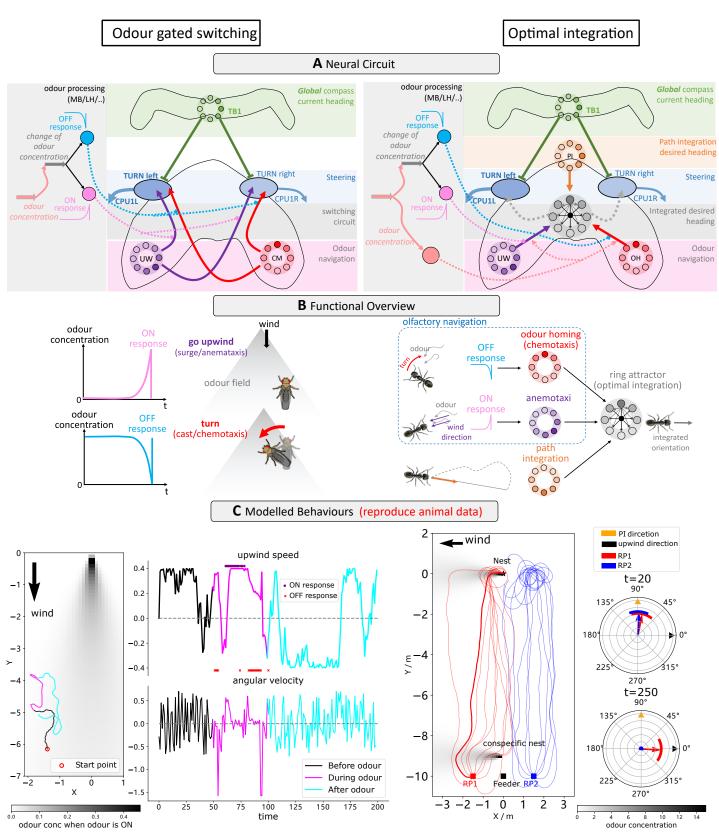
- ensuring that all orientation cues are presented in a shared frame of reference. Recall that the 178
- desired headings for path integration, chemotaxis, and anemotaxis are all defined in relation to 170
- the animal's global head direction. In the following analysis we assess whether this frame-changing 180
- capacity can also provide benefits for navigational robustness. 181
- From egocentric wind direction to geocentric celestial compass 182
- Desert ants travel to and from familiar feeder locations via visually guided routes (Kohler and 183 Wehner, 2005: Mangan and Webb, 2012) but wind gusts can blow them off course. Wystrach 184 and Schwarz (2013) reported that in the instant prior to displacement ants assume a stereotypical 185 'clutching' pose during which they transfer their egocentric measure of wind direction (indicating 186 the direction in which they are about to be blown) into a geocentric frame of reference given by 187 their celestial compass. Displaced ants then utilise this celestial compass memory to guide their 188 path directly towards their familiar route (*Figure 4*A (left panel)). Such a strategy is easily accounted 180 for by the 'copy-and-shift' mechanism as seen in **Figure 4**B (left panel). That is, during the clutch 190 pose the celestial compass heading is *copied*, and *shifted* by the activation of the WPN encoding 191 the upwind direction relative to the animal's heading to create a desired heading that points back 192 along the direction of travel. This desired heading is maintained in a working memory during dis-
- 193 placement before activation to guide the agent back to the familiar route region (see simulated 194
- navigating paths in *Figure 4*C (left panel)). 195

From visual context to geocentric celestial compass 196

Similarly, homing desert ants captured just before entering their nest and released in unfamiliar 197 visual surroundings initially dash back along the celestial compass heading in which they were 198 travelling (Wvstrach et al., 2013) (Figure 4A (right panel)). Note that this differs from the behaviour of ants lacking path integration cues and displaced from other locations along the route. Those ants have no preferred direction of travel following displacement according to the observation 201 (Wystrach et al., 2013). This indicates that sight of the nest surroundings could be considered a 202 'special circumstance' in a similar way to the 'clutching' pose mentioned above. *Figure 4*B (right 203 panel) depicts how this behaviour could also arise from the 'copy-and-shift' mechanism. That is, 204 when there is a significant drop of visual novelty (as might only be experienced after a displacement 205 from the nest), the compass direction is again *copied* and *shifted* by a predetermined amount, this 206 case 180 degrees. This creates a new desired heading that can be stored in working memory that 207 will cause the initial search to be focused in the direction from which the animals just travelled 208 (Figure 4C (right panel)). 209

In summary, the data above demonstrates the flexibility of the 'copy-and-shift' mechanism to 210 transfer directional cues from an unstable frame of reference such as the wind direction to a sta-211 ble frame of reference such as the global celestial compass which can be used at a later time. We 212 proposed that this transfer is triggered by special sensory experience and motivational state of 213

7 of 21



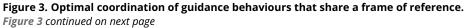


Figure 3 (continued). (A): Schematic diagrams of the integration circuits. Left: temporal change in odour concentration based ON and OFF-responses drives the switching circuit to select between chemotaxis or anemotaxis strategies. Right: ring attractor network integrate multiple cues weighted by sensory valence. (B): Functional explanations of the model. Left: On-responses trigger upwind turns while OFF-responses trigger chemotaxis leading the animal back into the odour plume. Right: ring attractors serves as the optimal integration circuit to mediating between anemotaxis, chemotaxis and path integration systems. (C): Example behaviours generated by the model in an anemotaxis, and ant homing task. Left part of the left panel shows the trajectory of the one simulated fly, the upwind speed and angular velocity of the agent are shown in the right part. The time at which ON- and OFF- responses are triggered are shown by purple dots and red stars respectively. The left panel of the right side data shows paths of simulated ants when guided by PI and odour cues. Groups headings are also shown at *t* = 20 (early in the route when PI dominates) and *t* = 250 (later in the route when olfactory navigation begin to dominate as PI vector length is low).

Figure 3-Figure supplement 1. The simulation results of a 20-agents group driven by the ON- and OFF-response based switching model. **Figure 3-Figure supplement 2.** Sensory perception and neural activities of the highlighted ant driven by the proposed model.

Figure 3-Figure supplement 3. Simulation results where there is no conspecific nest near the releasing points with comparison to (C) right panel. **Figure 3-video 1.** The animation showing the simulation process including homing trajectories, dynamic neural activation, odour measurement etc.

- the animal, that could be driven by some of the numerous tangential inputs from multiple up-
- stream brain regions to the FB (Franconville et al., 2018; Hulse et al., 2021) forming a contextually
- dependent guidance network. This again extends the repertoire of guidance behaviour that the
- mechanism can account for and further supports to the role of the central complex as a navigationcentre.

219 Discussion

To summarise, we have shown how the CX-based steering circuit augmented with a copy-and-shift 220 functionality can generate realistic odour-based chemotaxis and anemotaxis behaviours adding 221 to the path integration, visual homing, visual route following, and long-range migrations explained 222 previously (Stone et al., 2017: Honkanen et al., 2019: Sun et al., 2020). We have also outlined CX-223 based mechanisms that can coordinate guidance cues across sensory domains using biologically-224 realistic context-dependent switches and ring attractor networks. Finally, we demonstrated how 225 the *copy-and-shift* mechanism can facilitate the transfer of orientation cues between unstable to 226 stable frames of references. By triggering such a transfer under specific environmental conditions 227 insects can increase the robustness of their guidance repertoire. The model presented can thus 228 be considered as a general navigation model extending across multiple behavioural tasks (align-229 ment with rotationally-varying compass, visual route or wind cues; and gradient ascent of spatially 230 varving but rotationally-invariant cues such as odour and visual memories) experienced in multiple 231 contexts. Taken together the results add further validation to the claim that the central complex 232 acts as the seat of navigation coordination in insects. 233

The central complex is as ancient as insects themselves (Homberg, 2008: Strausfeld, 2009) and 234 is highly conserved across different species solving different navigational tasks (Honkanen et al., 235 2019: Hulse et al., 2021). This fixed circuitry thus appears optimised to receive input from a variety 236 of sensory sources and return a similar variety of navigational behaviours applicable across con-237 texts. Indeed Dovle and Csete (2011) posits that such 'bowtie' (or hourglass) architectures are also 238 observed in the decision making circuits of the mammalian brain (Redgrave et al., 1999: Humphries 239 and Prescott. 2010) and function by providing "constraints that deconstrain" (see Figure 5A). That 240 is, the fixed circuitry of the CX constrains the format of the sensory input but decontrains the ap-241 plication domains of the output behaviours. Through interpreting various navigation behaviours 242 through the lens of the 'copy-and-shift' mechanism, our model can be considered an example of 243 such bowtie structure within the CX (*Figure 5*B). 244

This study has explored the behavioural consequences of the mechanisms using abstracted neural implementations, raising the question as whether they can be realised in insect brains. Regarding the *copy-and-shift* mechanism, lateralised neural connections and synapse-plasticity that shift the head-direction output relative to sensory input (i.e. nudge the activation 'bump' within

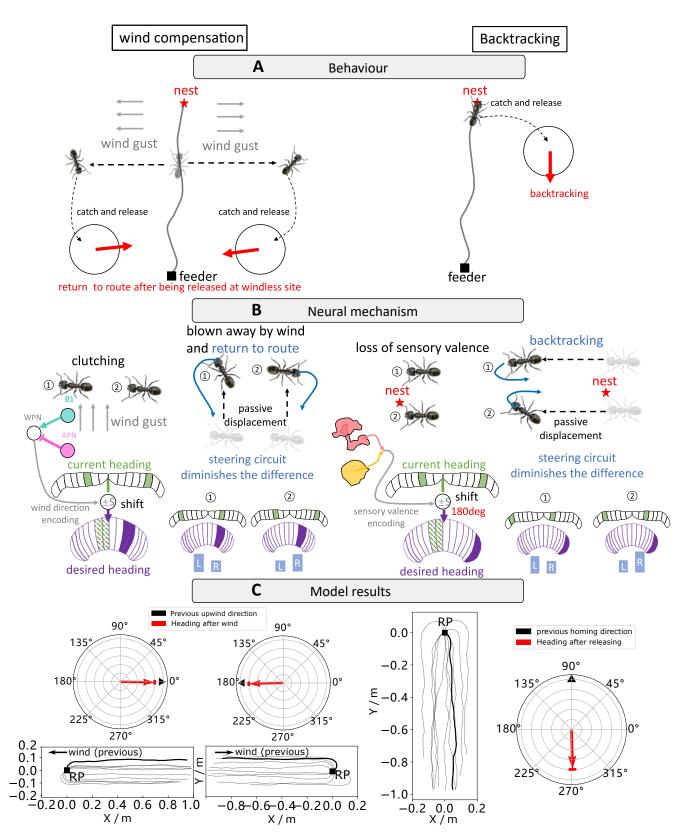


Figure 4. Navigating using egocentric and geocentic frames of reference.

Figure 4 continued on the next page

Figure 4 (continued). (A): Wind compensation and backtracking behaviour of navigating ants. Left panel illustrates the wind compensation behaviour where ants reorientate to the direction from which they were blown off course but with respect to their celestial compass (*Wystrach and Schwarz, 2013*). Right subfigure shows backtracking behaviours whereby homing desert ants captured just before entering their nest and released in unfamiliar visual surroundings initially dash back along the celestial compass heading in which they were travelling (*Wystrach et al., 2013*). (B): The proposed neural mechanism showing how the behaviours in (A) could be recreated. Wind-compensation is implemented by using the *copy-and-shift* to *copy* their heading compass stored in the CX when clutching and *shift* by an amount degree determined by the activation of WPN neurons to form the working memory (desired heading) for later navigation. Backtracking is modelled in identical way except that the *shift* is constant 180°. (C): The simulation results of our model. In each panel, the navigating trajectories and initial headings of the simulated ants are shown. Simulated ants guided by the model are all heading to the expected orientation as observed in real behavioural experiments (*Wystrach and Schwarz, 2013*; *Wystrach et al., 2013*).

a population of neurons) have already been mapped (Seelig and Javaraman, 2015; Green et al., 240 2017; Kim et al., 2019; Fisher et al., 2019) and modelled (Cope et al., 2017) demonstrating the fea-250 sibility of such computation. More recently, Goulard et al. (2021) presented a CX-based navigation 251 model that includes a biologically realistic neural pathway that is functionally similar to the *cop*-252 and-shift mechanism proposed here. The same study also outlined how a short-term memory of 253 a desired heading could be maintained in the FB of the CX via synapse-weight modulation after 254 the original guidance cue is removed, that could support the wind-compensation and backtrack-255 ing behaviours described above. Our model hypothesises the existence of a ring attractor network 256 to optimally integrate desired heading cues which we suggest could be realised in the complex 257 intra-connections within the FB and the Noduli (NO) (Hulse et al., 2021: Savre et al., 2021). We also hypothesise that different populations of PFN neurons in the CX simultaneously store the distinct 259 desired headings computed by the independent navigation systems (e.g., PI-based home vector is stored in CPU4 neurons (a subset of PFNs) (Stone et al., 2017; Hulse et al., 2021; Savre et al., 2021)). 261 Further, the hypothetical context-switching introduced could be achieved by the recently mapped 262 FB-NOc neurons found in the bees (Savre et al., 2021). 263 It is also worth noting that the simulated odour perception utilised here is very simplistic. For 264 example, we assume that the odour stimulus (with or without a laminar air-flow) forms a stable 265 gradient, which while reflecting the laboratory settings in behavioural studies (Gomez-Marin et al., 266 2010: Gomez-Marin and Louis, 2012: Álvarez-Salvado et al., 2018), simplifies the spatiotemporally 267 complex plumes in naturalistic settings where odour encounters are intermittent, occurring ran-268 domly as brief bursts (Murlis et al., 2000; Webster and Weissburg, 2001). We do note however, 260 that more stable odour gradients have been mapped to the desert surfaces upon which desert 270 ants forage (Buehlmann et al., 2015). Regardless, insect olfactory receptor neurons (ORNs) and 271 projection neurons (PNs) posses adaption (Kaissling et al., 1987; Nagel and Wilson, 2011), and di-272 visive gain control (Luo et al., 2010: Olsen et al., 2010: Gorur-Shandilva et al., 2017) mechanisms 273 that normalise and smooth noisy olfactory inputs. It is interesting to note that the visual gradients 274 can often present data in a similar noisy fashion (personal observation) and thus raises the ques-275 tion as to whether similar processing steps are applied across modalities. Indeed, this hypothesis is 276 supported by identification of shared early sensory processing principles across sensory modalities 277 (Wilson, 2013), especially the vision and olfactory in insects (My et al., 2012) and mammals (Cleland, 278 **2010**). Another interesting point is to the temporal presentation of information (e.g. continual or 279 discrete) and how this might affect aspects such as optimal integration of cues. We suggest that 280 optimal integration would not be unduly affected as sampling over longer time scales would sim-281 ply reduce the strength of the more sparsely samples cues to the ring attractor. Moreover, there 282 may be benefits in sampling less as it could smooth out local noise in sensory gradients. Investi-283 gation of these questions through modelling studies that add more realistic sensory processing in 284 more realistic sensory settings (odour: (Demir et al., 2020), vision: (Millward et al., 2021)) is vital to 285 answering these questions. 286 Despite growing agreement on the functional role of the CX in insect navigation (Honkanen 287 et al., 2019: Hulse et al., 2021), a number of issues remain. Firstly, as well as innervating the CX. 288

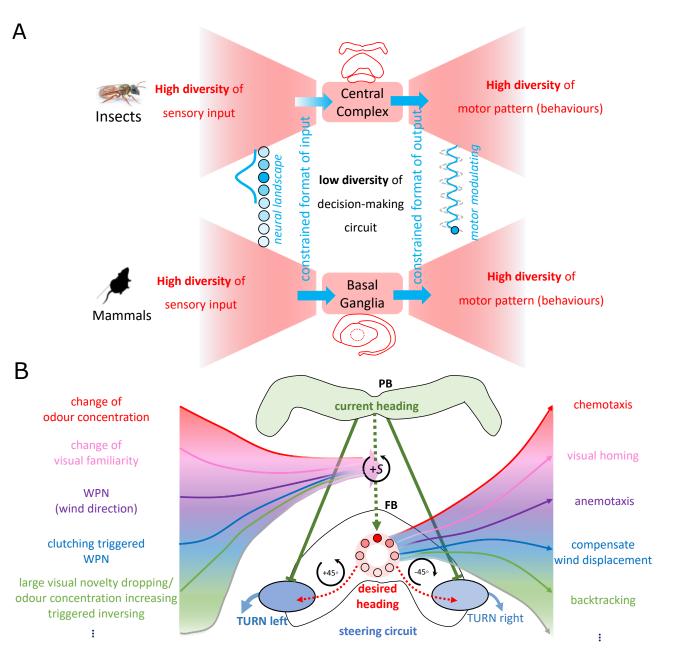


Figure 5. The 'bowtie/hourglass' architecture (*Doyle and Csete, 2011***) of biological control system.** (A) The control systems of insect navigation (top) and mammalian decision-making (bottom) are epitomised by the 'bowtie' architecture, proposing that fixed brain circuitry constrains the format of the sensory input (fanning in to the knot) but decontrains the application domains of the output behaviours (fanning out of the bowtie). Photo of sweet bee *Megalopta genalis* is from Ajay Narendra. (B) The proposed mapping of the bowtie architecture to the CX for insect navigation. Specially, the copy-and-shift mechanism (regarded as the knot of the bowtie thus constrains the representation) reused to generate different desired headings across sensory and task domains (deconstrains the motor pattern thus allows for high diversity of behaviours).

- ²⁸⁹ both visual and olfactory cues are also transferred directly to motor centres (*Rayshubskiy, 2020*;
- 290 Scaplen et al., 2021; Green et al., 2019) providing redundant information streams. One possibility
- is that the direct pathways are used for fast reflex-like movements, whereas the CX pathway is
- ²⁹² responsible for higher-level guidance that requires learning and integration of multiple elemental
- ²⁹³ guidance systems (*Currier et al., 2020; Matheson et al., 2021*). This view is consistent with *Stein*-
- *beck et al.* (2020) who demonstrate that the lateral-accessory-lobes (LAL), downstream of the CX,
- ²⁹⁵ possess neural structures well suited to integrating outputs of the fast and the slow pathways (For

Drosophila larvae, there should be equivalent neural circuity functioning similarly as the CX involved 296 pathway (probably with the olfactory descending neurons PDM-DN (Ibrahim et al., 2018: Gowda 297 et al., 2021)) and direct pathway (probably with Odd neurons (Slater et al., 2015; Gowda et al., 298 **2021**))). Future work is needed to merge these concepts into a single computational framework. Secondly, there is the question as to whether insects maintain a single or multiple head direction 300 signals in the PB. In our previous model (Sun et al., 2020), we introduced a global celestial compass 301 used by VH and PI behaviours and a local visual compass for RF. In this study, we relied solely on 302 the global celestial compass, but wind direction sensing from the WPN neurons are known to feed 303 into the head direction cells (Okubo et al., 2020: Hulse et al., 2021) which could facilitate a local 304 compass similar to our previous terrestrial compass. The utility and biological realism of the multi-305 compass hypothesis deserves further investigation. Thirdly, insects possess a MB in each brain 306 hemisphere posing the question as to their combined role. Le Möel and Wystrach (2020): Wys-307 trach et al. (2020) offer the hypothesis that MBs form an opponent memory system that can drive 308 visual route following by balancing the difference in their outputs. This approach can be easily ex-309 tended to incorporate both attractive and repulsive MB output neurons extending the application 310 space and robustness of navigation. Integration of dual MB inputs represents an obvious next ex-311 tension of the model presented here. Finally, the model presented here is unique in the format of 312 the sensory data input to the MBs, and the behavioural strategies that the MBs generate. Specifi-313 cally, we propose that the MBs process rotationally-invariant but spatially-varying cues (e.g. odour 314 and visual familiarity gradients) and are thus responsible for generating gradient ascent/descent 315 behaviours such as visual homing and chemotaxis via operant connections to the CX. In contrast, 316 all rotationally-varving cues (e.g. wind-direction, visual route memories, and celestial compass) in-317 nervate the CX directly via alternate pathways (e.g. LAL). This separation of sensory information is 318 fundamental to the flexibility of the model presented to create the array of behaviours presented 319 and offers a testable hypothesis for future work. Such insights will be invaluable for refinement of 320 our understanding of the robust navigation behaviours facilitated by the insect minibrain. 321

322 Methods and Materials

All simulations and network models are implemented by Python 3.5 and external libraries-numpy,

matplotlib, *scipy*, *opencv* etc. The source code of the simulation and plotting figures are available via Github.

Odour field

327 As the basic sensory input, the spatial concentration distribution of the odour field is simulated

simply and based on the scaled exponential functions, with required changes according to the

329 wind dynamics.

330 Odour field without wind

³³¹ For the simulations in the laminar odour environment (i.e. no wind) as that in *Figure 2*(left panel),

the landscape of the odour concentration CON_a are modelled for 'volcano' shape:

$$CON_o = \begin{cases} ke^{\tau(r/2-d)} & if \ d > r/2\\ ke^{\tau(d-r/2)} & otherwise \end{cases}$$
(1)

333 and for 'linear' shape:

$$CON_o = \begin{cases} ke^{\tau(r/2-d)} & if \quad d > r/2\\ k - 0.2e^{\tau(d-r/2)} & otherwise \end{cases}$$
(2)

where *d* is the distance from the position (x, y) to the odour source (x_s, y_s) . Thus, $d = \sqrt{(x - x_s)^2 + (y - y_s)^2}$.

 $_{335}$ k is the scale factor, r is the radius of the odour source and τ is decay factor.

- 336 Odour field with wind
- ³³⁷ To simplify the simulation of the odour plume dynamics, all the simulations in this study are con-
- ³³⁸ ducted under the condition of constant wind speed *u* and wind direction $\theta_{w'}$ and we assume that
- the odour plume will ideally flow to the downwind area, i.e., the odour concentration in the upwind
- area will always be zero. The source of the odour constantly emits at the rate q, Then the odour
- concentration at position (x, y) can be calculated by:

$$CON_{o} = \begin{cases} \frac{q}{u\sigma_{xy}\sqrt{2\pi}}e^{-\frac{d^{2}}{2\pi\sigma_{xy}}} & if\cos\theta > 0\\ 0 & otherwise \end{cases}$$
(3)

where $d = \sqrt{(x - x_s)^2 + (y - y_s)^2} \sin \theta$ is the projected distance from the odour source. And σ_{xy} is calculated by $\sigma_{xy} = K_s d$ where $K_s \in [0.5, 0.3, 0.2, 0.15, 0.1]$ is the tuning factor determined by the stability of the odour. And θ is the angel between the vector pointing from the position to the source and the wind direction, so can be computed by:

$$\theta = \arccos \frac{(x - x_s)(u \cos \theta_w) + (y - y_s)(u \sin \theta_w)}{\sqrt{(x - x_s)^2 + (y - y_s)^2}u}$$
(4)

346 Neural model

- ³⁴⁷ We use the simple firing rate to model the neurons in the proposed networks, where the output
- $_{348}$ firing rate C is a sigmoid function of the input I if there is no special note. In the following descrip-
- tions and formulas, a subscript is used to represent the layers or name of the neuron while the
- ³⁵⁰ superscript is used to represent the value at a specific time or with a specific index.

351 Current heading

- In our previous model, there are two compass references derived from different sensory informa-
- tion (Sun et al., 2020), but in this paper, only the global compass, (i.e. the activation of I-TB1/Δ7
- ³⁵⁴ neuron) is used here because navigation behaviours reproduced in this study are all assumed us-
- ing the global compass as the external direction reference. For the details of the modelling of
- global current heading $(I_{I-TB1}^{t,j})$ see our previous paper (Sun et al., 2020).
- 357 Steering circuit
- ³⁵⁸ The steering neurons (the same as previous paper (Sun et al., 2020) but presented here for conve-
- nience), i.e., CPU1 neurons (C_{CPU1}^{i} , i = 0, 1, 2...15) receive excitatory inputs from the desired heading
- C_{DH}^{i} , i = 0, 1, 2...15) and inhibitory inputs from the current heading (C_{CH} , i = 0, 1, 2...15) to generate
- ³⁶¹ the turning signal:

$$C_{ST}^{i} = C_{DH}^{i} - C_{CH}^{i} \qquad i = 0, 1, ...15$$
(5)

The turning angle is determined by the difference of the activation summations between left (i = 0, 1, 2...7) and right (i = 8, 9, 10...15) set of CPU1 neurons:

$$\theta_M = k_{motor} (\sum_{i=0}^{7} C_{CPU1} - \sum_{i=8}^{15} C_{CPU1})$$
(6)

364 Upwind direction encoding

³⁶⁵ The upwind direction is decoded as the activation of UW neurons copied and shifted from heading

- neurons (I-TB1), the value of this shifting is determined by the angular difference between the
- ³⁶⁷ current heading (θ_h) and wind direction (θ_w) encoded by the firing rate of WPN neuron. And the

value of WPN neuron is defined as the difference of the antennal deflection encoded by B1 and

369 APN neurons as:

$$C_{WPN} = C_{APN} - C_{B1} = \sin\left(\theta_w - \theta_h + \pi\right) - \sin\left(-(\theta_w - \theta_h + \pi)\right) \tag{7}$$

	$< Thr_{off}$	$> Thr_{off} < Thr_{on}$	$> Thr_{on}$
$< Thr_o$	Random	Random	ON
$> Thr_o$	OFF	ON	ON

Table 1. 'Truth table' of the ON and OFF response of the modelled fly odour navigation. The column lists the state of sensed odour concentration while the row indicates the state of the changing of odour concentration.

Then population activation of upwind direction neurons (UW) can be calculated by:

$$C_{UW} == C_{I-TB1}^{j}, j = \begin{cases} i + offset & if i + offset \le 7\\ i + offset - 7 & otherwise \end{cases}$$
(8)

³⁷¹ Fly- ON and OFF response based switching circuit

2

³⁷² Different navigation strategy will dominate the motor system according to the sensory inputs, i.e.,

in this study, the change of perceived odour concentration. This coordination is modelled as a

- contextual switching that is very similar with the mechanism with SN1 and SN2 neuron involved in
- our previous model (*Sun et al., 2020*) to define the final output of odour navigation (C_{ON}):

$$C_{ON}^{i} = \begin{cases} C_{chemo}^{i} & if \quad OFF \ response\\ C_{anemo}^{i} & if \quad ON \ response \end{cases}$$
(9)

And how the sensory information determine the response is shown in *Table 1*, where Random means no reliable sensory input is available, the agent will move forward to a random direction.

378 OFF response- chemotaxis

The chemotaxis model is adapted from the previous visual homing model (Sun et al., 2020) by

changing the change of visual familiarity signal from the MBON neuron (ΔC_{MBON}) to the change of the odour concentration to determine the shifting value, thus the desired heading of chemotaxis is:

$$C^{i}_{chemo} = C^{j}_{I-TB1}, j = \begin{cases} i + offset & if i + offset \le 7\\ i + offset - 7 & otherwise \end{cases} \quad i = 0, 1, ...7$$
(10)

Note that, in our previous visual navigation model (*Sun et al., 2020*), *i*, *j* both are integer for the ease of computing, thus, the shifting resolution is 45°, but here to more accurately model the desired heading and to achieve better performance, the shifting resolution was set to be 4.5° by interpolating neuron activation of I-TB1 from 8 to 80 then down-sampling to 8 to generate shifted desired heading.

The relationship between the ΔC_o and the of f set is shown as following:

$$offset = \begin{cases} 0 & if \quad \Delta C_o < 0\\ \min(\lfloor k_{chemo} \Delta C_o \rfloor, 3) & otherwise \end{cases}$$
(11)

Then the desired heading of OH will be fed into the steering circuit to compare with the current heading to generate the motor command.

³⁹¹ ON-response- dour-gated Anemotaxis

- As shown in Table 1, when the ON response is determined, the agent will follow the upwind direc-
- tion, thus the desired heading input to steering circuit should be the upwind direction encoded by
- 394 UM neuron ((8)):

$$C_{anemo}^{i} = C_{UW}^{i} \tag{12}$$

- 395 Ants- integration with PI
- ³⁹⁶ The modelling of ants' odour navigation integrated with PI can be regarded as the extension of
- ³⁹⁷ the fly's odour navigation and an application of the unified model. Specifically, the final output of
- ³⁹⁸ olfactory navigation is determined by the ON and OFF response (see *Table 1*), and then is integrated
- with PI via RA like that in the optimal integration of PI and VH:

$$\tau \frac{dC_{IN}}{dt} = -C_{IN} + g\left(\sum_{j=1}^{n} W_{E2E}^{ji} C_{IN}^{j} + X_{1}^{i} + X_{2}^{i} + W_{I2E} C_{UI}\right) \qquad i = 0, 1, \dots 7.$$
(13)

Where W_{E2E}^{ji} is the recurrent connections from j^{th} neuron to i^{th} neuron, g(x) is the activation function that provides the non-linear property of the neuron:

$$g(c) = max(0, \rho + c) \tag{14}$$

Where ρ denotes the offset of the function. Thus the X1 should be:

$$X_1^i = C_{PI}^i \qquad i = 0, 1, ...7$$
(15)

and *X*2 in (13) should be:

$$X_{2}^{i} = \begin{cases} k_{o}CON_{o}C_{OH}^{i} & if \quad OFF \ response\\ k_{o}CON_{o}C_{anemo}^{i} & if \quad ON \ response \end{cases}$$
(16)

Then the output of optimal integration (OI) of the RA acts as the only desired heading input to the steering circuit:

$$C_{DH}^{0-7} = C_{OI} W_{DH2CPU1L}$$

$$C_{DH}^{8-15} = C_{OI} W_{DH2CPU1R}$$
(17)

As only the global compass is needed in this study's modelling. Thus the input of current heading will always be the excitation of the I-TB1 neuron:

$$\begin{cases} C_{CH}^{0-7} = C_{I-TB1} \\ C_{CH}^{8-15} = C_{I-TB1} \end{cases}$$
(18)

The output of the steering circuit (i.e., the summed activation of the left and right CPU1 neurons) is used to generate the turning command in the way that is same as (6).

410 Simulations

In all simulations, at each time step, the simulated agent (walking fly or ant) will sense the odour

sensory based on its current location and then update neural activation to generate the desired

moving direction and finally move one step to that direction. *Equation 6* gives the turning angle of

the agent, thus the instantaneous "velocity" (*v*) at every step can be computed by:

$$\boldsymbol{v}^{t} = S_{L}[\cos\theta_{M}^{t}, \sin\theta_{M}^{t}]$$
⁽¹⁹⁾

Where S_L is the step length with the unit of centimetres. Note that we haven't defined the time accuracy for every step of the simulations, thus the unit of the velocity in this implementation is cm/step rather than cm/s. Then the position of agent P^{t+1} in the Cartesian coordinates for the is updated by:

$$^{+1} = \boldsymbol{P}^t + \boldsymbol{v}^t \tag{20}$$

The position of odour sources in all simulations are all set to (0,0), i.e., $x_s = 0$, $y_s = 0$. Other main parameters are listed in **Table 2**. Note that in each simulation, the speed of agent is set constant.

P^t

- 421 Fly- Chemotaxis
- 422 To test the performances of the chemotaxis behaviour, 5 simulated agents with randomly gen-
- erated heading direction starts from 5 randomly generated locations in the zone of (-12 < x <
- 12, -12 < y < 12), and then driven by the model for 1500 steps. Then we run this simulation for 4
- times in two different odour landscapes ('volcano' and 'linear') to get the results shown in *Figure 2*
- (right panel) and *Figure Supplement 1*.

		Fly-Chemotaxis			Elv Integrated	Apt Integrated
		volcano	Linear	– Fly-Anemotaxis	Fly-Integrated	Ant-Integrated
odour	k	10	10			
	τ	0.1	0.1		/	
	r	6	6			
	q	/	/	10.0	10.0	20
wind	и	no wind		10.0	10.0	10.0
	$w_{ heta}$			$-\pi/2$	$-\pi/2$	π
	Thr_o				0.001	1.2
model	Thr_{on}	/		/	0.02	0.5
	Thr_{off}				-0.0002	-0.0002
&	k _o				/	0.5
	k _{chemo}	100.0	100.0	/	100.0	100.0
simulation	k _{motor}	1.0	1.0	1.5	1.5	1.0
	S_L	0.02	0.02	0.4	0.4	0.05
	Heading	random	random	random	random	$0-2\pi$

Table 2. The detailed parameters settings for the simulations in this study.

Flv- Anemotaxis 427

To reproduce the behavioural data in Álvarez-Salvado et al. (2018), the odour was only set on 428

during the second a quarter of total time (e.g. if the agent is set to run 200 steps, then the odour-429

on time will in 50-100 steps). Four agents with randomly generated heading starts from randomly 430

generated locations in the zone of (-1.5 < x < 1.5, -13 < y < -5), and then guided by the model to 431

run 200 steps. The simulation was conducted for 5 times. 432

Fly-Integrated ON and OFF Response 433

The whole simulation settings are the same as that in the last section except for some model pa-434

rameters listed in *Table 2*, as this simulation is conducted to verify the integrated model. 435

Ants- Odour Navigation Integrated with PI 436

To reproduce the behavioural data in **Buehlmann et al.** (2012), we first generate PI memory en-437

coding the home vector with 10m length and $\pi/2$ direction. Then at each release point ((-1.5, -10) 438

and (1.5, -10)), we released 10 simulated full-vector (10m-long and pointing to $\pi/2$) ants with dif-439

ferent initial headings sampled uniformly from $0 - 2\pi$, see also **Table 2**. Note that the simulation 440

settings with/without additional odour plume diffused by conspecific nest are identical so list as 441 one column in Table 2.

442

Ants- wind compensation and backtracking 443

The guick implementations of using 'copy-and-shift' mechanism to model the wind compensation 444

and backtracking behaviour follow the same step: first, generate the desired headings by shifting 445

the current heading by the WPN activation for the wind compensation and by 180° for backtrack-446

ing respectively; second, release the simulated ant at the same releasing point but with random 447

headings (uniform distribution in $0 - 2\pi$). Motion-related parameters are set identically as that of 448

Ants- Odour Navigation Integrated with PI. 449

Acknowledgements 450

This research has received funding from the European Union's Horizon 2020 research and innova-451

tion programme under the Marie Sklodowska-Curie grant agreement No 778062, ULTRACEPT and 452

No 691154, STEP2DYNA. 453

- **454 References**
- 455 Álvarez-Salvado E, Licata AM, Connor EG, McHugh MK, King BM, Stavropoulos N, Victor JD, Crimaldi JP, Nagel
- KI. Elementary sensory-motor transformations underlying olfactory navigation in walking fruit-flies. Elife.
- 457 2018; 7:e37815.
- Ardin P, Peng F, Mangan M, Lagogiannis K, Webb B. Using an insect mushroom body circuit to encode route
 memory in complex natural environments. PLoS computational biology. 2016; 12(2):e1004683.
- 400 Aso Y, Sitaraman D, Ichinose T, Kaun KR, Vogt K, Belliart-Guérin G, Plaçais PY, Robie AA, Yamagata N, Schnait-
- mann C, et al. Mushroom body output neurons encode valence and guide memory-based action selection
- 462 in Drosophila. Elife. 2014; 3:e04580.
- Bell JS, Wilson RI. Behavior reveals selective summation and max pooling among olfactory processing channels.
 Neuron. 2016; 91(2):425–438.
- van Breugel F, Dickinson MH. Plume-tracking behavior of flying Drosophila emerges from a set of distinct
 sensory-motor reflexes. Current Biology. 2014; 24(3):274–286.
- Buehlmann C, Graham P, Hansson BS, Knaden M. Desert ants use olfactory scenes for navigation. Animal
 Behaviour. 2015; 106:99–105.
- **Buehlmann C**, Hansson BS, Knaden M. Path integration controls nest-plume following in desert ants. Current Biology. 2012; 22(7):645–649.
- 471 Cleland TA. Early transformations in odor representation. Trends in neurosciences. 2010; 33(3):130–139.
- 472 Cope AJ, Sabo C, Vasilaki E, Barron AB, Marshall JA. A computational model of the integration of landmarks and
 473 motion in the insect central complex. PloS one. 2017; 12(2):e0172325.
- **Currier TA**, Matheson AM, Nagel KI. Encoding and control of orientation to airflow by a set of Drosophila fan-shaped body neurons. Elife. 2020; 9:e61510.
- **Demir M**, Kadakia N, Anderson HD, Clark DA, Emonet T. Walking Drosophila navigate complex plumes using stochastic decisions biased by the timing of odor encounters. Elife. 2020; 9:e57524.
- Dolan MJ, Belliart-Guérin G, Bates AS, Frechter S, Lampin-Saint-Amaux A, Aso Y, Roberts RJ, Schlegel P, Wong
- A, Hammad A, et al. Communication from learned to innate olfactory processing centers is required for
 memory retrieval in Drosophila. Neuron. 2018: 100(3):651–668.
- Doyle JC, Csete M. Architecture, constraints, and behavior. Proceedings of the National Academy of Sciences.
 2011: 108(Supplement 3):15624–15630.
- Fisher YE, Lu J, D'Alessandro I, Wilson RI. Sensorimotor experience remaps visual input to a heading-direction
 network. Nature. 2019; 576(7785):121–125.
- Franconville R, Beron C, Jayaraman V. Building a functional connectome of the Drosophila central complex.
 Elife. 2018; 7:e37017.
- **Gkanias E**, Risse B, Mangan M, Webb B. From skylight input to behavioural output: a computational model of the insect polarised light compass. PLoS computational biology. 2019; 15(7):e1007123.
- **Gomez-Marin A**, Duistermars B, Frye MA, Louis M. Mechanisms of odor-tracking: multiple sensors for enhanced perception and behavior. Frontiers in cellular neuroscience. 2010; 4:6.
- **Gomez-Marin A**, Louis M. Active sensation during orientation behavior in the Drosophila larva: more sense than luck. Current opinion in neurobiology. 2012; 22(2):208–215.
- **Gorur-Shandilya S**, Demir M, Long J, Clark DA, Emonet T. Olfactory receptor neurons use gain control and complementary kinetics to encode intermittent odorant stimuli. Elife. 2017; 6:e27670.
- 497 //doi.org/10.1371/journal.pcbi.1009383, doi: 10.1371/journal.pcbi.1009383.
- **Gowda SBM**, Salim S, Mohammad F. Anatomy and Neural Pathways Modulating Distinct Locomotor Behaviors
- in Drosophila Larva. Biology. 2021; 10(2). https://www.mdpi.com/2079-7737/10/2/90, doi: 10.3390/biol-
- 500 ogy10020090.

- Green J, Adachi A, Shah KK, Hirokawa JD, Magani PS, Maimon G. A neural circuit architecture for angular inte gration in Drosophila. Nature. 2017; 546(7656):101–106.
- Green J, Vijayan V, Pires PM, Adachi A, Maimon G. A neural heading estimate is compared with an internal goal
 to guide oriented navigation. Nature neuroscience. 2019; 22(9):1460–1468.
- Gupta N, Stopfer M. Functional analysis of a higher olfactory center, the lateral horn. Journal of Neuroscience.
 2012; 32(24):8138–8148.
- Hardcastle BJ, Omoto JJ, Kandimalla P, Nguyen BCM, Keleş MF, Boyd NK, Hartenstein V, Frye MA. A visual
 pathway for skylight polarization processing in Drosophila. Elife. 2021; 10:e63225.
- 609 Heinze S. Polarized-light processing in insect brains: recent insights from the desert locust, the monarch
- butterfly, the cricket, and the fruit fly. In: *Polarized light and polarization vision in animal sciences* Springer;
 2014.p. 61–111.
- Heinze S, Jundi B, Berg BG, Homberg U, Menzel R, Pfeiffer K, Hensgen R, Zittrell F, Tedore K. A unified platform to manage , share , and archive morphological and functional data in insect neuroscience. eLife. 2021;
 10:e65376. doi: 10.7554/eLife.65376.
- **Homberg U.** Evolution of the central complex in the arthropod brain with respect to the visual system. Arthropod structure & development. 2008; 37(5):347–362.
- Honkanen A, Adden A, da Silva Freitas J, Heinze S. The insect central complex and the neural basis of naviga tional strategies. Journal of Experimental Biology. 2019; 222(Suppl 1).
- **Hu W**, Peng Y, Sun J, Zhang F, Zhang X, Wang L, Li Q, Zhong Y. Fan-shaped body neurons in the Drosophila brain regulate both innate and conditioned nociceptive avoidance. Cell reports. 2018; 24(6):1573–1584.
- Hulse BK, Haberkern H, Franconville R, Turner-Evans DB, Takemura S, Wolff T, Noorman M, Dreher M, Dan
 C, Parekh R, Hermundstad AM, Rubin GM, Jayaraman V. A connectome of the Drosophila central complex
 reveals network motifs suitable for flexible navigation and context-dependent action selection. eLife. 2021;
- 524 10:e66039.
- Humphries MD, Prescott TJ. The ventral basal ganglia, a selection mechanism at the crossroads of space,
 strategy, and reward. Progress in neurobiology. 2010; 90(4):385–417.
- Ibrahim T, Avinash K, David T, Fessner ND, James WT, Marta Z, Albert C, Matthieu L. Sensorimotor pathway
 controlling stopping behavior during chemotaxis in the Drosophila melanogaster larva. eLife. 2018; 7:e38740.
 doi: 10.7554/eLife.38740.
- Jung SH, Hueston C, Bhandawat V. Odor-identity dependent motor programs underlie behavioral responses
 to odors. Elife. 2015; 4:e11092.
- Kaissling KE, Strausfeld CZ, Rumbo E. Adaptation processes in insect olfactory receptors: mechanisms and
 behavioral significance. Annals of the New York Academy of Sciences. 1987; 510(1):104–112.
- Kennedy JS, Marsh D. Pheromone-regulated anemotaxis in flying moths. Science. 1974; 184(4140):999–1001.
- **Kim AJ**, Lazar AA, Slutskiy YB. System identification of Drosophila olfactory sensory neurons. Journal of computational neuroscience. 2011; 30(1):143–161.
- Kim SS, Hermundstad AM, Romani S, Abbott L, Jayaraman V. Generation of stable heading representations in
 diverse visual scenes. Nature. 2019; 576(7785):126–131.
- Kohler M, Wehner R. Idiosyncratic route-based memories in desert ants, Melophorus bagoti: how do they
 interact with path-integration vectors? Neurobiology of learning and memory. 2005; 83(1):1–12.
- Le Moël F, Stone T, Lihoreau M, Wystrach A, Webb B. The central complex as a potential substrate for vector based navigation. Frontiers in psychology. 2019; 10:690.
- Le Möel F, Wystrach A. Opponent processes in visual memories: A model of attraction and repulsion in navigating insects' mushroom bodies. PLoS computational biology. 2020; 16(2):e1007631.
- Legge EL, Wystrach A, Spetch ML, Cheng K. Combining sky and earth: desert ants (Melophorus bagoti) show
 weighted integration of celestial and terrestrial cues. Journal of Experimental Biology. 2014; 217(23):4159–4166.

- Li F, Lindsey JW, Marin EC, Otto N, Dreher M, Dempsey G, Stark I, Bates AS, Pleijzier MW, Schlegel P, et al. The connectome of the adult Drosophila mushroom body provides insights into function. Elife. 2020; 9:e62576.
- Luo SX, Axel R, Abbott L. Generating sparse and selective third-order responses in the olfactory system of the
 fly. Proceedings of the National Academy of Sciences. 2010; 107(23):10713–10718.
- Lyu C, Abbott L, Maimon G. A neuronal circuit for vector computation builds an allocentric traveling-direction
 signal in the Drosophila fan-shaped body. bioRxiv. 2020; .
- Mangan M, Webb B. Spontaneous formation of multiple routes in individual desert ants (Cataglyphis velox).
 Behavioral Ecology. 2012; 23(5):944–954.
- Matheson AM, Lanz AJ, Licata AM, Currier TA, Syed MH, Nagel KI. Organization of central circuits for windguided olfactory navigation. bioRxiv. 2021; .
- Millward BF, Maddock S, Mangan M. CompoundRay: An open-source tool for high-speed and high-fidelity
 rendering of compound eyes. bioRxiv. 2021; .
- Mu L, Ito K, Bacon JP, Strausfeld NJ. Optic glomeruli and their inputs in Drosophila share an organizational
 ground pattern with the antennal lobes. Journal of Neuroscience. 2012; 32(18):6061–6071.
- Murlis J, Willis MA, Cardé RT. Spatial and temporal structures of pheromone plumes in fields and forests.
 Physiological Entomology. 2000; 25(3):211–222. https://onlinelibrary.wiley.com/doi/abs/10.1046/j.1365-3032.
 2000.00176.x, doi: https://doi.org/10.1046/j.1365-3032.2000.00176.x.
- Nagel KI, Wilson RI. Biophysical mechanisms underlying olfactory receptor neuron dynamics. Nature neuro science. 2011; 14(2):208–216.
- Okubo TS, Patella P, D'Alessandro I, Wilson RI. A neural network for wind-guided compass navigation. Neuron.
 2020; .
- **Olsen SR**, Bhandawat V, Wilson RI. Divisive normalization in olfactory population codes. Neuron. 2010; 66(2):287–299.
- Patella P, Wilson RI. Functional maps of mechanosensory features in the Drosophila brain. Current Biology.
 2018; 28(8):1189–1203.
- **Plath JA**, Entler BV, Kirkerud NH, Schlegel U, Galizia CG, Barron AB. Different roles for honey bee mushroom
- bodies and central complex in visual learning of colored lights in an aversive conditioning assay. Frontiers
 in behavioral neuroscience. 2017; 11:98.
- 876 Rayshubskiy A. Neural control of steering in walking Drosophila. PhD thesis, Harvard University; 2020.
- Redgrave P, Prescott TJ, Gurney K. The basal ganglia: a vertebrate solution to the selection problem? Neuro science. 1999; 89(4):1009–1023.
- **Roussel E**, Carcaud J, Combe M, Giurfa M, Sandoz JC. Olfactory coding in the honeybee lateral horn. Current
 biology. 2014; 24(5):561–567.
- Rutkowski AJ, Quinn RD, Willis MA. Three-dimensional characterization of the wind-borne pheromone tracking
 behavior of male hawkmoths, Manduca sexta. Journal of Comparative Physiology A. 2009; 195(1):39–54.
- Sayre ME, Templin R, Chavez J, Kempenaers J, Heinze S. A projectome of the bumblebee central complex. eLife.
 2021; 10.
- Scaplen KM, Talay M, Fisher JD, Cohn R, Sorkaç A, Aso Y, Barnea G, Kaun KR. Transsynaptic mapping of
 Drosophila mushroom body output neurons. Elife. 2021; 10:e63379.
- Schulze A, Gomez-Marin A, Rajendran VG, Lott G, Musy M, Ahammad P, Deogade A, Sharpe J, Riedl J, Jarriault
 D, et al. Dynamical feature extraction at the sensory periphery guides chemotaxis. Elife. 2015; 4:e06694.
- Seelig JD, Jayaraman V. Neural dynamics for landmark orientation and angular path integration. Nature. 2015;
 521(7551):186–191.
- Shiozaki HM, Ohta K, Kazama H. A multi-regional network encoding heading and steering maneuvers in
 Drosophila. Neuron. 2020; 106(1):126–141.

- Slater G, Levy P, Chan KLA, Larsen C. A Central Neural Pathway Controlling Odor Tracking in Drosophila.
 Journal of Neuroscience. 2015; 35(5):1831–1848. https://www.jneurosci.org/content/35/5/1831, doi:
- 595 10.1523/JNEUROSCI.2331-14.2015.
- **Steck K**, Veit D, Grandy R, i Badia SB, Mathews Z, Verschure P, Hansson BS, Knaden M. A high-throughput behavioral paradigm for Drosophila olfaction-The Flywalk. Scientific reports. 2012; 2:361.
- Steinbeck F, Adden A, Graham P. Connecting brain to behaviour: a role for general purpose steering circuits
 in insect orientation? Journal of Experimental Biology. 2020; 223(5):jeb212332.
- 600 Stone T, Webb B, Adden A, Weddig NB, Honkanen A, Templin R, Wcislo W, Scimeca L, Warrant E, Heinze S. An
- anatomically constrained model for path integration in the bee brain. Current Biology. 2017; 27(20):3069– 3085.
- Strausfeld NJ. Brain organization and the origin of insects: an assessment. Proceedings of the Royal Society
 B: Biological Sciences. 2009; 276(1664):1929–1937.
- **Sun X**, Mangan M, Yue S. An analysis of a ring attractor model for cue integration. In: *Conference on Biomimetic and Biohybrid Systems* Springer; 2018. p. 459–470.
- Sun X, Yue S, Mangan M. A decentralised neural model explaining optimal integration of navigational strategies
 in insects. Elife. 2020; 9:e54026.
- Suver MP, Matheson AM, Sarkar S, Damiata M, Schoppik D, Nagel KI. Encoding of wind direction by central
 neurons in Drosophila. Neuron. 2019; 102(4):828–842.
- **Touretzky DS**. Attractor network models of head direction cells. Head direction cells and the neural mechanisms of spatial orientation. 2005; p. 411–432.
- Turner-Evans D, Wegener S, Rouault H, Franconville R, Wolff T, Seelig JD, Druckmann S, Jayaraman V. Angular
 velocity integration in a fly heading circuit. Elife. 2017; 6:e23496.
- Webb B, Wystrach A. Neural mechanisms of insect navigation. Current Opinion in Insect Science. 2016; 15:27–
 39.
- 617 Webster DR, Weissburg MJ. Chemosensory guidance cues in a turbulent chemical odor plume. Limnology and
- Oceanography. 2001; 46(5):1034–1047. https://aslopubs.onlinelibrary.wiley.com/doi/abs/10.4319/lo.2001.46.5.
 1034, doi: https://doi.org/10.4319/lo.2001.46.5.1034.
- Wehner R. The Cataglyphis Mahrèsienne: 50 years of Cataglyphis research at Mahrès. Journal of Comparative
 Physiology A. 2019; 205(5):641–659.
- Wessnitzer J, Young JM, Armstrong JD, Webb B. A model of non-elemental olfactory learning in Drosophila.
 Journal of computational neuroscience. 2012; 32(2):197–212.
- Wilson RI. Early olfactory processing in Drosophila: mechanisms and principles. Annual review of neuroscience.
 2013; 36:217–241.
- Wolf H, Wehner R. Pinpointing food sources: olfactory and anemotactic orientation in desert ants, Cataglyphis
 fortis. Journal of Experimental Biology. 2000; 203(5):857–868.
- Wolf H, Wehner R. Desert ants compensate for navigation uncertainty. Journal of Experimental Biology. 2005;
 208(22):4223–4230.
- Wystrach A, Schwarz S. Ants use a predictive mechanism to compensate for passive displacements by wind.
 Current Biology. 2013; 23(24):R1083–R1085.
- Wystrach A, Beugnon G, Cheng K. Ants might use different view-matching strategies on and off the route.
 Journal of Experimental Biology. 2012; 215(1):44–55.
- Wystrach A, Buehlmann C, Schwarz S, Cheng K, Graham P. Rapid aversive and memory trace learning during
 route navigation in desert ants. Current Biology. 2020; .
- Wystrach A, Mangan M, Webb B. Optimal cue integration in ants. Proceedings of the Royal Society B: Biological
 Sciences. 2015; 282(1816):20151484.
- Wystrach A, Schwarz S, Baniel A, Cheng K. Backtracking behaviour in lost ants: an additional strategy in their
 navigational toolkit. Proceedings of the Royal Society B: Biological Sciences. 2013; 280(1769):20131677.
- Yorozu S, Wong A, Fischer BJ, Dankert H, Kernan MJ, Kamikouchi A, Ito K, Anderson DJ. Distinct sensory representations of wind and near-field sound in the Drosophila brain. Nature. 2009; 458(7235):201–205.

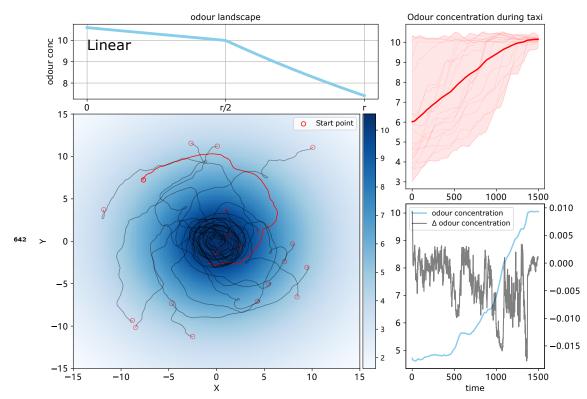


Figure 2-Figure supplement 1. The simulation results of chemotaxis model with odour landscape of 'Linear'. The odour field model and navigating trajectories are shown on the left whilst the perceived odour concentration and the temporal change of the highlighted agent are shown on the right.

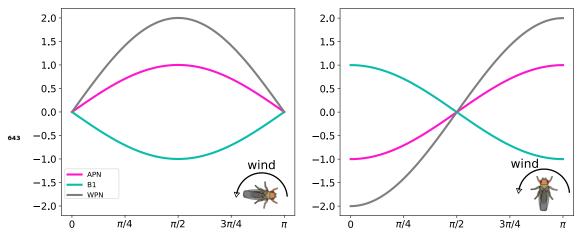


Figure 2-Figure supplement 2. Neural responses of the wind direction encoding neurons with different animal headings (0 and $\pi/2$) and the wind direction stimuli is swept from 0 to π .

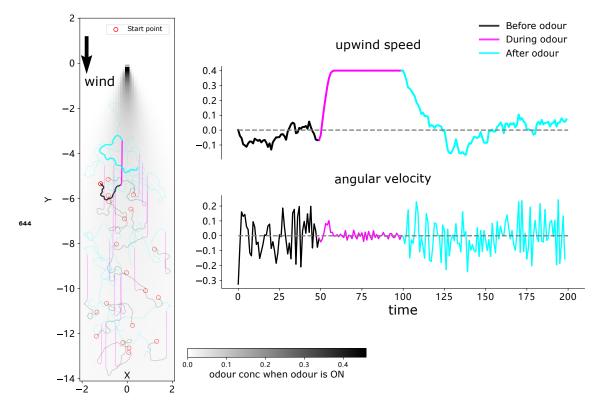


Figure 2–Figure supplement 3. Trajectories of each agents (highlighted one corresponding to that shown in **Figure 2**), mean upwind speed and angular velocity of 20 simulated agents are shown.

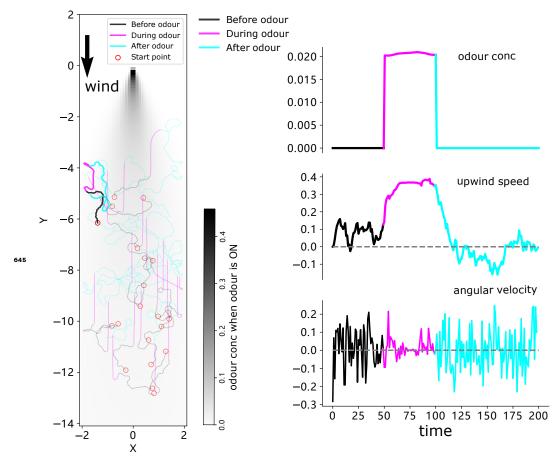


Figure 3–Figure supplement 1. The simulation results of 20 agents. Trajectories are shown on the left with highlighted one corresponding to that of **Figure 3**, mean perceived odour concentration, upwind speed and angular velocity are plotted on the right.

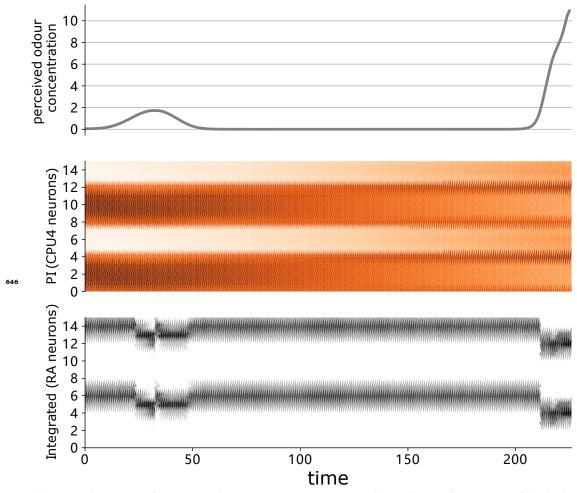


Figure 3–Figure supplement 2. The instantaneous sensory value and neural activation of the highlighted agent in **Figure 3**C (right panel) during homing. From top to bottom, the value of perceived odour concentration, the activation of PI memory neurons (CPU4) and the ring attractor excitation neurons. Note that the output of the ring attractor neurons combines injected cues as expected.

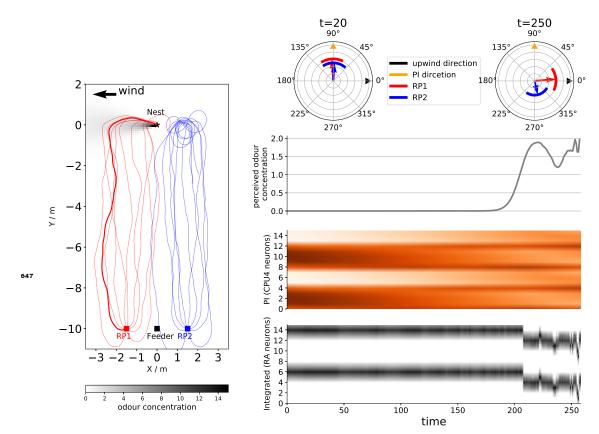


Figure 3–Figure supplement 3. Left part draws the simulated ants' homing paths and the group mean headings at t = 20 (when PI dominated) and t = 250 (when olfactory navigation should dominate the steering) are shown on the right. The instantaneous sensory value and neural activation of highlighted agent in the left panel during homing on shown on the right hand panel: from top to bottom, the value of perceived odour concentration, the activation of PI memory neurons (CPU4) and the ring attractor excitation neurons.