



Deposited via The University of Leeds.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/180780/>

Version: Accepted Version

---

**Article:**

Le Feunteun, S, Mackie, AR and Dupont, D (2020) In silico trials of food digestion and absorption: how far are we? *Current Opinion in Food Science*, 31. pp. 121-125. ISSN: 2214-7993

<https://doi.org/10.1016/j.cofs.2020.04.006>

---

© 2020, Elsevier. This manuscript version is made available under the CC-BY-NC-ND 4.0 license <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

**Reuse**

This article is distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs (CC BY-NC-ND) licence. This licence only allows you to download this work and share it with others as long as you credit the authors, but you can't change the article in any way or use it commercially. More information and the full terms of the licence here: <https://creativecommons.org/licenses/>

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.

1 ***In silico* trials of food digestion and absorption: How far**  
2 **are we?**

3 **Steven Le Feunteun<sup>1,\*</sup>, Alan Robert Mackie<sup>2</sup> and Didier Dupont<sup>1</sup>**

4

5 <sup>1</sup> INRAE, Agrocampus Ouest, UMR STLO, F-35042 Rennes, France

6 <sup>2</sup> School of Food Science and Nutrition, University of Leeds, Leeds, LS2 9JT, UK

7 \* [steven.le-feunteun@inra.fr](mailto:steven.le-feunteun@inra.fr)

8

9 **Abstract**

10 In recent years, experimental research on the mechanisms of food digestion in the  
11 gastrointestinal tract has strengthened our knowledge on the effect of food on human health.  
12 A number of mathematical models have been proposed to rationalize our understanding on  
13 the related mechanisms. One common suggestion is that *in silico* models could be  
14 interconnected and used in the future to predict the effect of food systems (liquid or solid,  
15 inner microstructure, state of nutrients...) on various metabolic responses. This paper aims  
16 to provide a brief overview of the latest developments in this young but promising field of  
17 research.

18

## 19 **Introduction**

20 The increase in diet-related diseases has fuelled the need to improve our knowledge on the  
21 fate of foods, or meals, in the gastrointestinal (GI) tract. Over the past 15 years, research in  
22 this field has provided new insights into why and how the structure of the food we eat can  
23 affect the kinetics and extent of nutrient absorption [1,2]. As with pharmaceuticals, the dose  
24 and timing of nutrient arrival in the blood stream during digestion has important metabolic  
25 consequences. Examples include both deleterious and beneficial repercussions, as for  
26 instance the increased risk of type-2 diabetes for diets with high glycaemic index [3], or the  
27 stimulation of protein muscle synthesis above a threshold of leucine in the peripheral blood  
28 [4]. To advance further in our understanding of the relationship between foods, or diets, with  
29 the overall functioning of the human gastrointestinal (GI) tract and post-absorptive processes,  
30 systems modelling of digestion appears as a promising means. Such an approach can be used  
31 to tackle the diversity of the mechanisms that take place during digestion, and could be very  
32 valuable for predictive purposes. This has already happened in the pharmaceutical area, in  
33 which the concept of “*in silico* clinical trials” has emerged as a new tool in the drug regulatory  
34 process [5,6]. This paper therefore questions the possibility of advancing towards  
35 establishing models of food digestion and nutrient absorption that could help predict the  
36 metabolic responses to foods and meals. It starts by describing some attempts that have been  
37 proposed in the past, before reviewing the latest developments and the remaining challenges  
38 related to this field of research.

## 39 **Physiologically based kinetic (PBK) modelling of food digestion: Where do we stand?**

40 Predictive models of food’s GI transit, hydrolysis and absorption can be built with classical  
41 engineering approaches, in which the digestive system is split into a number of anatomical  
42 compartments (e.g. one or several gastric and intestinal sub-compartments, peripheral blood,  
43 etc.) and into a series of unit operations to model the physicochemical processes that take  
44 place [7]. In the pharmaceutical area, this strategy has led to a number of physiologically  
45 based kinetic (PBK) models to predict the absorption of orally administered pharmaceuticals  
46 [5], or for safety assessment of chemicals: cosmetics, food additives, pesticides, etc. [8].  
47 Thanks to their capability to predict the overall internal exposure to a chemical and on its  
48 ability to elicit a biological response, these models are becoming more and more essential  
49 before *in vivo* experiments can be undertaken.

50 The same principles can be applied to relate some nutritional considerations (e.g. extent and  
51 kinetics of nutrient absorption) with food or meal digestion. PBK models of the GI transit  
52 and absorption of meals, from stomach to colon with consideration of all kinds of  
53 macronutrients, were proposed for pigs quite a long time ago [9,10]. The latest version of this  
54 model [10] seemed to accurately predict the digestibility of the main food components, lipid  
55 excepted, as well as the absorption profile of the studied nutrients (glucose, amino acids, and  
56 volatile fatty acids) over a large variety of diets. The same strategy could thus be adapted for  
57 human physiology. One lack, however, is that they do not directly account for the physical  
58 properties of ingested foods or meals. Hence, these are not directly suitable to predict food  
59 structure effects. In another study, a PBK model dedicated to dairy protein digestion and  
60 absorption in mini-pigs showed that the great differences in the kinetics of amino acid  
61 absorption experimentally observed for differently structured dairy matrices of identical  
62 composition could be fairly reproduced by considering contrasting gastric behaviours and  
63 emptying kinetics [11]. By distinguishing the fraction of gastric content that is ready to be  
64 emptied into the small intestine from the one that is not ready yet (e.g. large food particles),  
65 this model provided a first attempt to integrate food structure considerations into PBK models  
66 of food digestion.

67 These examples are interesting in that they can be used to predict the dose and timing of  
68 nutrient arrival in the blood stream in response to a given food or a meal, and could be easily  
69 connected with nutritional models that aim to predict the metabolic fate and consequences of  
70 absorbed amino-acids [12], lipid products [13], or sugars [14] in the fed state. However, to  
71 build a PBK model of food transit and absorption that becomes truly relevant for predictive  
72 purposes, there remains a clear need to more directly relate the properties of the foods or  
73 meals with: (i) the kinetics of enzymatic hydrolysis, and (ii) the GI transit, in particular the  
74 gastric emptying kinetics. As further reviewed, a number of models are now becoming  
75 available to tackle these issues.

### 76 **How to account for key properties of macronutrients in the modelling of enzymatic** 77 **hydrolysis and GI transit?**

78 Current developments in the modelling of enzymatic hydrolysis mostly originate from the  
79 ongoing efforts to take into account the key properties of the main nutrients (i.e. proteins,  
80 lipids and starch). In this field, we may shed light on the results recently obtained from  
81 approaches that consider multiple species within each type of macronutrient, rather than  
82 trying to model an average behaviour. For instance, a model accounting for the fatty acid

83 composition of an oil-in-water emulsion, and for rate constants that are specific for each fatty  
84 acid residue, not only allowed a better fitting of the *in vitro* disappearance of triacylglycerols  
85 but also proved very efficient in predicting the individual bioaccessibility profiles of each  
86 oil's fatty acid [15]. Another multi-response model has been successfully applied to the  
87 hydrolysis profiles of the main lipid categories (triglycerides, monoglycerides, and free fatty  
88 acids) for a total of 28 different data sets of emulsion intestinal *in vitro* digestions [16].  
89 Comparable approaches have also been proposed to model the hydrolysis kinetics of proteins  
90 [17,18], and starch [19–21], where consideration is made that different rates of hydrolysis  
91 can be assumed for different subclasses of the considered substrate, *i.e.* more or less resistant  
92 and/or accessible fractions. These examples are of particular interest because they provide a  
93 direct means of incorporating both compositional and structural information in enzymatic  
94 hydrolysis models, with non-empirical relations between the model parameters and the food  
95 properties. Since they all rely on the same principles and are relatively simple to build or  
96 modify, these approaches could thus be assembled and interconnected to enable predictive  
97 modelling of the release kinetics of all kinds of nutrient from food material(s). The large body  
98 of experimental data already existing in the literature could be used to identify the most  
99 appropriate model structures and tune their parameterization.

100 Gastric emptying is another key determinant of the overall nutrient absorption kinetics. It is  
101 well known that this process is, in first approximation, governed by a feedback mechanism  
102 controlling the flux of calories delivered to the proximal small intestine. Indeed, it was shown  
103 more than 40 years ago that gastric emptying of meals can be fairly predicted from the meal's  
104 initial volume and caloric density [22]. More recently, Moxon and co-workers proposed a  
105 gastric emptying model for nutrient liquid meals that include a nutrient feedback mechanism,  
106 and which further takes into account the variations of the chyme viscosity upon gastric  
107 secretions and emptying [23]. Overall, this model closely simulated *in vivo* gastric emptying  
108 patterns of liquid meals varying by both their nutrient content and viscosity. Although the  
109 predictive capability of this model needs to be validated further, it certainly constitutes a  
110 much more elegant basis than mass action laws or empirical equations to predict the gastric  
111 emptying of liquid meals in PBK models of food digestion. Overall, almost all pieces seem  
112 now available to develop PBK models that could predict the main nutritional responses to  
113 liquid foods.

114 **Three-dimensional computational models of food breakdown and mixing**

115 Food is not simply a soup of nutrients. It has structure that is complex and multiscale,  
116 requiring phenomena occurring at a larger scale to be taken into account for solid foods. The  
117 comminution of food and mixing with GI fluids are primarily important at the oral and gastric  
118 stages, where food pieces are larger and less diluted by the digestive secretions. Food  
119 fragment size and structure may not only impact enzymatic hydrolysis but also gastric  
120 emptying, which largely controls the overall kinetics of nutrient uptake [24]. It is thus  
121 important to develop models that could predict food fragmentation and mixing in the upper  
122 part of the GI tract. These mechanisms are generally simulated using computational solid  
123 mechanics and fluid dynamics.

124 During the oral phase, solid foods are broken down and lubricated with saliva. Food oral  
125 processing therefore governs the size distribution of food particles that reach the stomach,  
126 which are typically up to several millimetres in size. To enhance our understanding of the  
127 relationship between food structure and oral breakdown during mastication, latest results  
128 from both finite element [25] and meshfree [26,27] three-dimensional (3D) methods are  
129 rather encouraging. When combined with experimental data from model food materials, they  
130 enabled fair predictions of crack initiation and propagation [25], and of food fragment sizes  
131 produced during chewing [26]. This latter model, proposed by Harrison and co-workers, used  
132 smoothed particle hydrodynamics (SPH) to predict the mechanical behaviour and breakdown  
133 of two agar gels during mastication. It is particularly interesting in that several chewing  
134 cycles are considered and the predicted fracture damage and particle size distributions are  
135 directly related to the measured properties of their food materials. These computational  
136 models still remain in their early stages but seem extendable to a variety of food structures  
137 with different rheological properties. In the future, it is therefore possible that these  
138 approaches could be used to predict fragment sizes of oral boli from known or measured food  
139 properties and anatomical considerations.

140 At the gastric stage, pioneer 3D computational models of the entire stomach date from the  
141 early 2010s, by Ferrua and Singh [28,29] and Imai et al. [30]. These models considered the  
142 case of a closed pylorus to investigate gastric mixing of liquid meals of different viscosity.  
143 More recently, Harrison and co-workers have also developed a SPH based model to simulate  
144 gastric mixing and emptying of aqueous solutions when the pylorus remains fully open [31],  
145 and the team of Imai has just investigated how the coordination between pyloric closure and  
146 antral contraction affects the emptying of liquid contents [32]. By considering gastric  
147 emptying, these new models represent an important step forward in improving our

148 understanding of gastric digestion. However, despite their great merits, current gastric  
149 computational models do not yet account for secretions and enzymatic reaction(s) and have  
150 only focused on liquids, with no or few discrete solid particles. Several important  
151 developments thus remain to be performed before a comprehensive multiphysics model of  
152 the stomach can be used to predict the gastric breakdown, mixing and emptying of solid foods  
153 [33].

154 Peristaltic and segmentation contractions in the small intestine have also been simulated with  
155 computational fluid dynamics to predict the flow and mixing at different length scales [34–  
156 37]. These studies have notably shown that the transport of nutrients from the intestinal lumen  
157 to the wall can be significantly reduced when the apparent viscosity of digesta is high [34],  
158 and that the motion of villi at the gut wall can significantly enhance the mixing in the  
159 proximity of epithelium cells [35–37]. Overall, these works have provided important insights  
160 to better understand the rate limiting steps of nutrient absorption, and on the possible effect  
161 of the mechanical properties of food.

162 Computational modelling of food breakdown and mixing in the GI tract has already provided  
163 important findings. If more work remains to be performed, in particular at the oral and gastric  
164 stages, this field of research is still young and ongoing developments in numerical methods  
165 rapidly improve the accuracy and speed of complex simulation scenarios. It is therefore  
166 expectable that these approaches will become more and more accurate and reliable in the  
167 forthcoming years.

### 168 **Alternative approaches to predict the breakdown and transit of solid foods**

169 In relation to the objective of predicting the main metabolic responses to a food or a meal  
170 from *in silico* modelling, it may not always be necessary to simulate the 3D-spatiotemporal  
171 evolution of the GI content. Predictions of food transit in one spatial dimension (*i.e.* along  
172 the GI tract) should be adequate for most nutritional considerations. From a systems  
173 modelling perspective, as in PBK models, this strategy also facilitates model computations  
174 and interconnections.

175 A number of research groups continue to use and adapt models developed in the fields of  
176 engineering and biophysics to predict the behaviour of food in gastric conditions [7]. Recent  
177 examples include modelling work on mass transfer and absorption in the intestine [38], or on  
178 the physical-chemistry of gastric digestion of solid foods to predict: their swelling [39], their

179 softening [40], their breakdown into particles [41], their acid and moisture uptake and  
180 buffering capacity [42–44]. Recently, a more integrative approach has also been proposed by  
181 Sicard and co-workers to model of the gastric digestion of meat proteins [45]. They built a  
182 reaction-diffusion model that accounts for a number of mechanisms: pepsin and proton  
183 diffusion in bolus particles, pepsin activity as a function of pH, the buffering capacity of  
184 meat, and indirectly for the movement of particles with secretion via a mass transfer  
185 coefficient at the meat particle–gastric fluid interface. As discussed by the authors, the current  
186 version of their model still has room for improvement, in particular by computing the  
187 progressive reduction of the particle size in relation to gastric emptying kinetics. Because the  
188 structure of this model has a very general character, this work appears suitable to be  
189 transposed to other types of solid foods, meanwhile it could also be integrated into PBK  
190 models of food digestion and absorption. In the future, further developments might enable  
191 prediction of the gastric digestion and emptying of solid foods from physical considerations  
192 in place of the classically employed empirical equations. This represents the main modelling  
193 challenge to overcome before predictive models of solid food digestion and absorption could  
194 become a reality.

### 195 **The essential role of experimental data**

196 Another bottleneck in establishing a comprehensive model of food digestion is the need to  
197 rely on relevant experimental data and knowledge. Because *in vitro* experiments use well-  
198 controlled conditions, they provide a very good framework to test some modelling  
199 assumptions, in particular with regards to the effects of food composition and structure on  
200 enzymatic and disintegration kinetics. Recent reviews have also discuss the ability and limits  
201 of both static [46] and dynamic [47] *in vitro* experiments to reproduce *in vivo* observations.  
202 Nonetheless, *in vitro* experiments do not reflect the reality of *in vivo* digestion, which is  
203 regulated by both neural and hormonal feedback mechanisms.

204 Hence, to develop *in silico* models of food digestion that become physiologically relevant,  
205 there is a clear need to rely on *in vivo* data, and more particularly on human data whenever  
206 possible. However, most of the previously described models rely on a small sample of the  
207 physiology literature. Since there is no *in vivo* database that modellers could use to build and  
208 evaluate their models, one of the main challenges they face is to find, extract and assess the  
209 relevant quantities from articles across various scientific fields. This is complicated by the  
210 fact that modellers do not always have a background in animal or human digestion  
211 physiology, meanwhile experts in physiology and metabolism are not necessarily aware of

212 the needs and constraints of modelling. To advance, the communities of experimentalists and  
213 modellers will have to collaborate more closely to identify and gather relevant experimental  
214 data sets and knowledge for model development and evaluation. In this context, modelling  
215 should also serve as a knowledge-based system, in which our understanding of the  
216 mechanisms and of their relationships is organized and can be incrementally improved and  
217 complete.

## 218 **Conclusion**

219 Modelling the digestive processes is challenging and research in this area is currently very  
220 active. The scope of existing models spread from molecular mechanisms up to systems view  
221 approaches with increasing efforts to relate food properties to key mechanisms of food  
222 digestion. Previously proposed PBK models of food digestion and absorption could largely  
223 benefit from recent advances in the modelling of food structure effects on digestion, from the  
224 enzymatic hydrolysis of macronutrients up to the impact of food macrostructure. All the  
225 pieces seem now available to start building *in silico* models that could predict the main  
226 metabolic responses to liquid foods and meals, although more work remains to be done for  
227 the case of solid foods.

228

229 **References**

230 Papers of particular interest, published within the period of review, have been highlighted as:

231

232 \* of special interest

233

234 **Moxon et al. (23):** This article presents a mathematical model for the gastric emptying of  
 235 liquid foods that relies on a nutrient feedback mechanism in the proximal small intestine.  
 236 Although this idea is not new (e.g. Ref. 22), this study is probably the first in recent years to  
 237 explicitly build a gastric emptying model upon this assumption. The model presented  
 238 provided good fits to experimental data for both high- and low-viscosity liquid meals with  
 239 high- and low-nutrient content.

240

241 **Brandstaeter et al. (33):** This review sums-up our current understanding of the mechanics  
 242 of the human stomach and delineates the challenges in mathematical and computational  
 243 modelling that remain to be addressed in this emerging field. It also illustrates potential  
 244 applications of a computational multiphysics model of the human stomach in areas ranging  
 245 from medicine to food industries.

246

247 \*\* of outstanding interest

248

249 **Lamond et al. (7):** This book chapter reviews the mechanics of human digestion from an  
 250 engineering viewpoint. It provides an engineering analysis of the gut, which includes  
 251 dimensional analysis and identification of the key parameters and phenomena that determine  
 252 the rate and extent of food digestion. The use of mathematical models and computational  
 253 fluid dynamics are also discussed for the study of digestive processes, in particular gastric  
 254 and intestinal flow and mixing.

255

256 **Sicard et al. (45):** This study presents a reaction–diffusion mathematical model that has been  
 257 developed to predict the gastric digestion of meat proteins. It is one of the most advanced  
 258 attempt to model the gastric hydrolysis of a solid food by the number of mechanisms it takes  
 259 into account: pepsin and proton diffusion in bolus particles, the buffering capacity of the  
 260 meat, the pepsin activity as a function of pH... This paper also reports simulation work  
 261 predicting the effects of pepsin concentration, pH variations and meat particle size and list  
 262 several possible improvements, especially in the way it accounts for the time-course change  
 263 in particle size.

264

265

266

267 1. Dupont D, Le Feunteun S, Marze S, Souchon I: **Structuring food to control its disintegration in**  
 268 **the gastrointestinal tract and optimize nutrient bioavailability.** *Innovative Food Science &*  
 269 *Emerging Technologies* 2018, **46**:83–90.

270 2. Gouseti O, Bornhorst G, Bakalis S, Mackie A (Eds): *Interdisciplinary Approaches to Food*  
 271 *Digestion.* Springer International Publishing; 2019.

272 3. Bhupathiraju SN, Tobias DK, Malik VS, Pan A, Hruby A, Manson JE, Willett WC, Hu FB: **Glycemic**  
 273 **index, glycemic load, and risk of type 2 diabetes: results from 3 large US cohorts and an**  
 274 **updated meta-analysis.** *Am J Clin Nutr* 2014, **100**:218–232.

- 275 4. Rieu I, Balage M, Sornet C, Giraudet C, Pujos E, Grizard J, Mosoni L, Dardevet D: **Leucine**  
276 **supplementation improves muscle protein synthesis in elderly men independently of**  
277 **hyperaminoacidaemia**. *The Journal of Physiology* 2006, **575**:305–315.
- 278 5. Zhuang X, Lu C: **PBPK modeling and simulation in drug research and development**. *Acta*  
279 *Pharmaceutica Sinica B* 2016, **6**:430–440.
- 280 6. Pappalardo F, Russo G, Tshinanu FM, Viceconti M: **In silico clinical trials: concepts and early**  
281 **adoptions**. *Brief Bioinformatics* 2018, doi:10.1093/bib/bby043.
- 282 7. Lamond AR, Janssen AEM, Mackie A, Bornhorst GM, Bakalis S, Gouseti O: **An engineering**  
283 **perspective on human digestion**. In *Interdisciplinary Approaches to Food Digestion*. Edited by  
284 Gouseti O, Bornhorst GM, Bakalis S, Mackie A. Springer International Publishing; 2019:255–273.
- 285 8. Madden JC, Pawar G, Cronin MTD, Webb S, Tan Y-M, Paini A: **In silico resources to assist in the**  
286 **development and evaluation of physiologically-based kinetic models**. *Computational*  
287 *Toxicology* 2019, **11**:33–49.
- 288 9. Bastianelli D, Sauvant D, Rérat A: **Mathematical modeling of digestion and nutrient absorption**  
289 **in pigs**. *Journal of animal science* 1996, **74**:1873–1887.
- 290 10. Strathe AB, Danfær A, Chwalibog A: **A dynamic model of digestion and absorption in pigs**.  
291 *Animal Feed Science and Technology* 2008, **143**:328–371.
- 292 11. Le Feunteun S, Barbe F, Remond D, Menard O, Le Gouar Y, Dupont D, Laroche B: **Impact of the**  
293 **dairy matrix structure on milk protein digestion kinetics: Mechanistic modelling based on**  
294 **mini-pig in vivo data**. *Food and Bioprocess Technology* 2014, **7**:1099–1113.
- 295 12. Fouillet H, Juillet B, Gaudichon C, Mariotti F, Tome D, Bos C: **Absorption kinetics are a key factor**  
296 **regulating postprandial protein metabolism in response to qualitative and quantitative**  
297 **variations in protein intake**. *Am J Physiol-Regul Integr Comp Physiol* 2009, **297**:R1691–R1705.
- 298 13. Jelic K, Hallgreen CE, Colding-Jørgensen M: **A model of NEFA dynamics with focus on the**  
299 **postprandial state**. *Ann Biomed Eng* 2009, **37**:1897.
- 300 14. Rozendaal YJ, Maas AH, van Pul C, Cottaar EJ, Haak HR, Hilbers PA, van Riel NA: **Model-based**  
301 **analysis of postprandial glycemic response dynamics for different types of food**. *Clinical*  
302 *Nutrition Experimental* 2018, **19**:32–45.
- 303 15. Giang TM, Gaucel S, Brestaz P, Anton M, Meynier A, Trelea IC, Le Feunteun S: **Dynamic modeling**  
304 **of in vitro lipid digestion: Individual fatty acid release and bioaccessibility kinetics**. *Food*  
305 *Chemistry* 2016, **194**:1180–1188.
- 306 16. Verkempinck SHE, Salvia-Trujillo L, Infantes Garcia MR, Hendrickx ME, Grauwet T: **From single**  
307 **to multiresponse modelling of food digestion kinetics: The case of lipid digestion**. *Journal of*  
308 *Food Engineering* 2019, **260**:40–49.

- 309 17. Barros RM, Xavier Malcata F: **A kinetic model for hydrolysis of whey proteins by cardosin A**  
310 **extracted from Cynara Cardunculus**. *Food Chemistry* 2004, **88**:351–359.
- 311 18. Kondjoyan A, Daudin J-D, Santé-Lhoutellier V: **Modelling of pepsin digestibility of myofibrillar**  
312 **proteins and of variations due to heating**. *Food Chemistry* 2015, **172**:265–271.
- 313 19. Li H, Dhital S, Gidley MJ, Gilbert RG: **A more general approach to fitting digestion kinetics of**  
314 **starch in food**. *Carbohydrate Polymers* 2019, **225**:115244.
- 315 20. Edwards CH, Warren FJ, Milligan PJ, Butterworth PJ, Ellis PR: **A novel method for classifying**  
316 **starch digestion by modelling the amylolysis of plant foods using first-order enzyme kinetic**  
317 **principles**. *Food Funct* 2014, **5**:2751–2758.
- 318 21. Meraz M, Alvarez-Ramirez J, Vernon-Carter EJ, Reyes I, Hernandez-Jaimes C, Martinez-Martinez  
319 F: **A two competing substrates Michaelis–Menten kinetics scheme for the analysis of in vitro**  
320 **starch digestograms**. *Starch - Stärke* [date unknown], **n/a**:1900170.
- 321 22. Hunt JN, Stubbs DF: **The volume and energy content of meals as determinants of gastric**  
322 **emptying**. *The Journal of Physiology* 1975, **245**:209–225.
- 323 23. Moxon TE, Nimmegeers P, Telen D, Fryer PJ, Van Impe J, Bakalis S: **Effect of chyme viscosity and**  
324 **nutrient feedback mechanism on gastric emptying**. *Chemical Engineering Science* 2017,  
325 **171**:318–330.
- 326 24. Weber E, Ehrlein H-JR: **Relationships between gastric emptying and intestinal absorption of**  
327 **nutrients and energy in mini pigs**. *Digestive Diseases and Sciences* 1998, **43**:13.
- 328 25. Skamniotis CG, Elliott M, Charalambides MN: **On modeling the large strain fracture behaviour**  
329 **of soft viscous foods**. *Physics of Fluids* 2017, **29**:121610.
- 330 26. Harrison SM, Eyres G, Cleary PW, Sinnott MD, Delahunty C, Lundin L: **Computational modeling**  
331 **of food oral breakdown using smoothed particle hydrodynamics**. *Journal of Texture Studies*  
332 2014, **45**:97–109.
- 333 27. Harrison SM, Cleary PW, Eyres G, D. Sinnott M, Lundin L: **Challenges in computational**  
334 **modelling of food breakdown and flavour release**. *Food Funct* 2014, **5**:2792–2805.
- 335 28. Ferrua MJ, Singh RP: **Modeling the fluid dynamics in a human stomach to gain insight of food**  
336 **digestion**. *J Food Sci* 2010, **75**:R151–R162.
- 337 29. Ferrua MJ, Singh RP: **Understanding the fluid dynamics of gastric digestion using**  
338 **computational modeling**. *Procedia Food Science* 2011, **1**:1465–1472.
- 339 30. Imai Y, Kobayashi I, Ishida S, Ishikawa T, Buist M, Yamaguchi T: **Antral recirculation in the**  
340 **stomach during gastric mixing**. *American Journal of Physiology-Gastrointestinal and Liver*  
341 *Physiology* 2013, **304**:G536–G542.

- 342 31. Harrison SM, Cleary PW, Sinnott MD: **Investigating mixing and emptying for aqueous liquid**  
343 **content from the stomach using a coupled biomechanical-SPH model.** *Food & Function* 2018,  
344 **9:3202–3219.**
- 345 32. Ishida S, Miyagawa T, O’Grady G, Cheng LK, Imai Y: **Quantification of gastric emptying caused**  
346 **by impaired coordination of pyloric closure with antral contraction: a simulation study.**  
347 *Journal of The Royal Society Interface* 2019, **16:20190266.**
- 348 33. Brandstaeter S, Fuchs SL, Aydin RC, Cyron CJ: **Mechanics of the stomach: A review of an**  
349 **emerging field of biomechanics.** *GAMM-Mitteilungen* 2019, **42:e201900001.**
- 350 34. Love RJ, Lentle RG, Asvarujanon P, Hemar Y, Stafford KJ: **An expanded finite element model of**  
351 **the intestinal mixing of digesta.** *Food Digestion* 2013, **4:26–35.**
- 352 35. de Loubens C, Lentle RG, Hulls C, Janssen PWM, Love RJ, Chambers JP: **Characterisation of**  
353 **mixing in the proximal duodenum of the rat during longitudinal contractions and comparison**  
354 **with a fluid mechanical model based on spatiotemporal motility data.** *PLoS ONE* 2014,  
355 **9:e95000.**
- 356 36. Wang Y, Brasseur JG, Banco G, Webb AG, Ailiani AC, Neuberger T: **A multiscale lattice**  
357 **Boltzmann model of macro- to micro-scale transport, with applications to gut function.**  
358 *Philosophical Transactions of the Royal Society A* 2010, **368:2863–2880.**
- 359 37. Lentle RG, Janssen PWM, DeLoubens C, Lim YF, Hulls C, Chambers P: **Mucosal microfolds**  
360 **augment mixing at the wall of the distal ileum of the brushtail possum.** *Neurogastroenterology*  
361 *& Motility* 2013, **25:881-e700.**
- 362 38. Moxon TE, Gouseti O, Bakalis S: **In silico modelling of mass transfer & absorption in the human**  
363 **gut.** *Journal of Food Engineering* 2016, **176:110–120.**
- 364 39. van der Sman RGM, Houlder S, Cornet S, Janssen A: **Physical chemistry of gastric digestion of**  
365 **proteins gels.** *Current Research in Food Science* 2019, doi:10.1016/j.crf.2019.11.003.
- 366 40. Drechsler KC, Bornhorst GM: **Modeling the softening of carbohydrate-based foods during**  
367 **simulated gastric digestion.** *Journal of Food Engineering* 2018, **222:38–48.**
- 368 41. Drechsler KC, Ferrua MJ: **Modelling the breakdown mechanics of solid foods during gastric**  
369 **digestion.** *Food Research International* 2016, **88:181–190.**
- 370 42. Luo Q, Zhan W, Boom RM, Janssen AEM: **Interactions between acid and proteins under in vitro**  
371 **gastric condition – a theoretical and experimental quantification.** *Food Funct* 2018, **9:5283–**  
372 **5289.**
- 373 43. Mennah-Govela YA: **Acid and moisture uptake into red beets during in vitro gastric digestion**  
374 **as influenced by gastric pH.** *Food Biophysics* [date unknown],

- 375 44. Mennah-Govela YA, Singh RP, Bornhorst GM: **Buffering capacity of protein-based model food**  
376 **systems in the context of gastric digestion.** *Food Funct* 2019, **10**:6074–6087.
- 377 45. Sicard J, Mirade P-S, Portanguen S, Clerjon S, Kondjoyan A: **Simulation of the gastric digestion**  
378 **of proteins of meat bolus using a reaction–diffusion model.** *Food & Function* 2018, **9**:6455–  
379 6469.
- 380 46. Bohn T, Carriere F, Day L, Amelie D, Egger L, Freitas D, Golding M, Le Feunteun S, Macierzanka  
381 A, Menard O, et al.: **Correlation between in vitro and in vivo data on food digestion. What can**  
382 **we predict with static in vitro digestion models?** *Critical Reviews in Food Science and Nutrition*  
383 2018, **58**:2239–2261.
- 384 47. Dupont D, Alric M, Blanquet-Diot S, Bornhorst G, Cueva C, Deglaire A, Denis S, Ferrua M,  
385 Havenaar R, Lelieveld J, et al.: **Can dynamic in vitro digestion systems mimic the physiological**  
386 **reality?** *Crit Rev Food Sci Nutr* 2019, **59**:1546–1562.
- 387