Article:
Dey, Nidhi S, Senaratne, Sujai, Somaratne, Vijani et al. (13 more authors) (2021) Early reduction in PD-L1 expression predicts faster treatment response in human cutaneous leishmaniasis. Journal of Clinical Investigation. e142765. ISSN: 1558-8238

https://doi.org/10.1172/JCI142765

JCI The Journal of Clinical Investigation

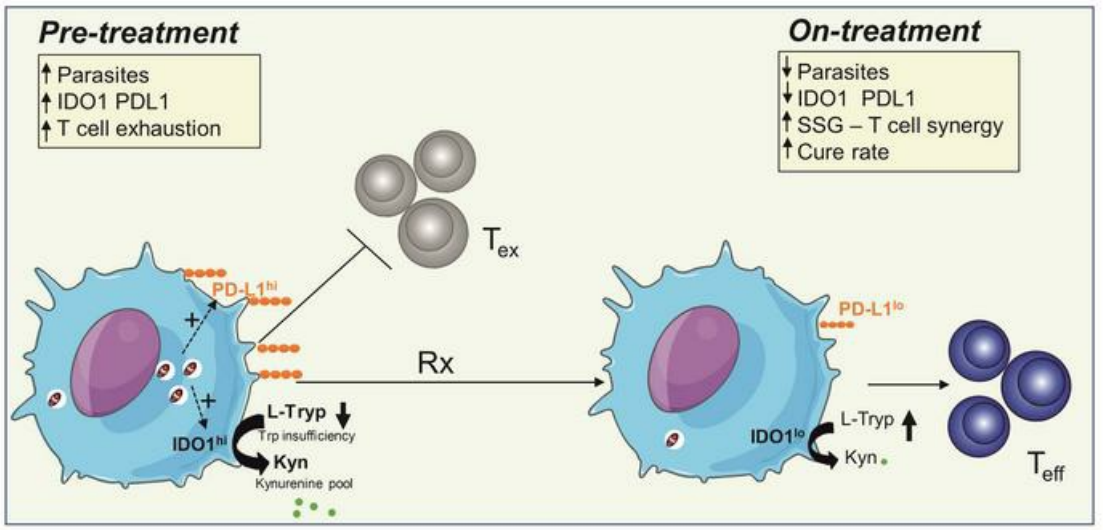
Early reduction in PD-L1 expression predicts faster treatment response in human cutaneous leishmaniasis

Nidhi S. Dey, … , Paul M. Kaye, Shalindra Ranasinghe

J Clin Invest. 2021. https://doi.org/10.1172/JCI142765.

Concise Communication In-Press Preview Immunology Infectious disease

Graphical abstract



Early reduction in PD-L1 expression predicts faster treatment response in human cutaneous leishmaniasis.

Nidhi S. Dey[1‡], Sujai Senaratne[2‡], Vijani Somaratne[3], Nayani Madarasinghe[4], Bimalka Seneviratne[5], Sarah Forrester[1], Marcela Montes De Oca[1], Luiza Campos Reis[6], Srija Moulik[7], Pegine Walrad[8], Mitali Chatterjee[7], Hiro Goto[6,9], Renu Wickremasinghe[2], Dimitris Lagos[1], Paul M. Kaye[1*] and Shalindra Ranasinghe[2*].

[1]York Biomedical Research Institute, Hull York Medical School, University of York, UK.
[2]Department of Parasitology, University of Sri Jayewardenepura, Sri Lanka.
[3]Dermatology Unit, District General Hospital Embilipitiya, Sri Lanka
[4]Dermatology Unit, Teaching Hospital Anuradhapura, Sri Lanka.
[5]Deparment of Pathology, University of Sri Jayewardenepura, Sri Lanka
[6]Instituto de Medicina Tropical de São Paulo, Faculdade de Medicina, Universidade de São Paulo, Brazil.
[7]Department of Pharmacology, Institute of Postgraduate Medical Education and Research, Kolkata, India.
[8]York Biomedical Research Institute, Dept. of Biology, University of York, UK.
[9]Departamento de Medicina Preventiva, Faculdade de Medicina, Universidade de São Paulo, São Paulo, Brazil

[‡]NSD and SS contributed equally to this work.

*Correspondence:
1. Paul M. Kaye

Hull York Medical School, University of York, Heslington, York - YO10 5DD, UK

Tel: +44 1904 328840, paul.kaye@york.ac.uk

2. Dr. Shalindra Ranasinghe

Department of Parasitology, University of Sri Jayewardenepura, Gangodawila, Nugegoda - 10250, Sri Lanka

Tel: +94 11 2801028; ishalindra@sjp.ac.lk

Conflict of interest: The authors have declared that no conflict of interest exists.

Abstract

Cutaneous leishmaniasis (CL) is caused by *Leishmania donovani* in Sri Lanka. Pentavalent antimonials (e.g. sodium stibogluconate; SSG) remain first line drugs for CL with no new effective treatments emerging. We studied whole blood and lesion transcriptomes from Sri Lankan CL patients at presentation and during SSG treatment. From lesions but not whole blood, we identified differential expression of immune-related genes, including immune checkpoint molecules, after onset of treatment. Using spatial profiling and RNA-FISH, we confirmed reduced expression of PD-L1 and IDO1 proteins on treatment in lesions of a second validation cohort and further demonstrated significantly higher expression of these checkpoint molecules on parasite-infected compared to non-infected lesional $CD68^+$ monocytes / macrophages. Crucially, early reduction in PD-L1 but not IDO1 expression was predictive of rate of clinical cure (HR = 4.88) and occurred in parallel with reduction in parasite load. Our data support a model whereby the initial anti-leishmanial activity of antimonial drugs alleviates checkpoint inhibition on T cells, facilitating immune-drug synergism and clinical cure. Our findings demonstrate that PD-L1 expression can be used as a predictor of rapidity of clinical response to SSG treatment in Sri Lanka and support further evaluation of PD-L1 as a host directed therapeutic in leishmaniasis.

Introduction

Every year, approximately 600,000 – 1 million new cases of cutaneous leishmaniasis (CL) occur, with a broad global distribution, often leading to stigma and reduced life chances and placing a burden on health services (1-3). Treatment options for CL have changed little in over 70 years, since pentavalent antimonial drugs were first introduced, and there are limited new treatments on the horizon (3). Sri Lanka is endemic for CL, with the first autochthonous case being reported in 1992 (4). Sri Lankan CL is caused by *Leishmania donovani* zymodeme MON-37 parasite (5-7), usually associated with visceral leishmaniasis in other endemic countries. Current treatment for CL in Sri Lanka involves weekly intra-lesional or daily intra-muscular administration of sodium stibogluconate (SSG), with or without cryotherapy, based on the site and size of the lesion and response to treatment. Cure often takes many months, and some patients may fail to respond completely or withdraw from treatment (8).

Most of our understanding of the host immune response in CL stems from experimental models, and human disease is much less understood (9). Immune checkpoint molecules have been implicated in disease progression in pre-clinical models (10-17), but their role in human CL has not been explored. It is widely proposed that immune-drug synergy is required for effective treatment and that host directed therapy (HDT) may have a future role in patient management (18-20), but few validated targets have emerged. Here, we searched for early correlates of treatment response that might be used to stratify patient response. Our results indicate an intimate relationship between intracellular parasitism and immune checkpoint molecule expression, with PD-L1 emerging as a promising target for HDT in Sri Lanka.

Results and Discussion

We first conducted a targeted transcriptomic analysis of the lesion site in a test cohort of 6 patients with typical homogeneous nodulo-ulcerative CL lesions (3 females, 3 males; mean age ± standard deviation, 34 ± 11 years; (Supplemental Figures 1-3 and Supplemental Table 1). Principal component analyses of lesion transcriptomic data showed separation of pre- and on-treatment samples in most patients (Figure 1A) and 120 differentially expressed genes were identified (DEGs; FDR adjusted p-value<0.01; Figure 1B). In contrast, no DEGs were identified by RNA-seq in whole blood (Supplemental Figure 4) suggesting that unlike CL caused by *L. braziliensis* (21), CL due to *L. donovani* in Sri Lanka is not accompanied by an overt systemic immune response.

Following treatment, the majority of DEGs in dermal lesions were downregulated (87%; 105/120) suggesting a reduction in inflammation following treatment (105 downregulated, 15 upregulated; Figure 1B and Supplemental Table 3). Genes for cellular functions and regulation, chemokines, membrane receptors, T cell function and regulation were amongst the top 20 DEGs (Figure 1C). Further, STRING analysis (22) identified Lymphocyte migration (GO: 0002687, FDR= 1.06E-14; including interferon inducible chemokines like *CXCL9, CXCL10, CXCL11*, *CCL19, CCL8)*) and regulators of immune response (GO: 0002684, FDR=1.94E-11; including *IDO1, LAG3* and *CD274/PDL1*) as highly enriched pathways (Figure 1D). Transcripts of inflammatory mediators including *CXCL10*, *GZMB*, *CCL2* and *CCR7* (receptor for CCL19), previously shown to be associated with other forms of murine (23-25) or human CL (26-28) were also downregulated with initiation of treatment (Supplemental Table 3).

We next conducted multiplexed antibody digital spatial profiling (29) for 59 immune targets, selecting regions of interest (ROIs) based on expression of $CD3^+$ and/or $CD68^+$ (Supplemental Figure 5 and Figure 2, A-F). The t-SNE dimensional reduction on a total of 33 regions of interest (ROIs) analysed from three patients (P4, P6 and P7) (Figure 2G) indicated a considerable degree of inter-patient heterogeneity in pre-treatment lesional protein profiles, but with clear discrimination for each patient between pre- and on-treatment ROIs. Upon treatment, IDO1 and PD-L1 as well as PD-1 were selectively reduced in expression (Figure 2, H and I). STRING analysis of all discoveries based on FDR (5%) also indicated significant enrichment in GO: 002684, as well as a pathway associated with regulation of T cell activation (GO: 0050863; Supplemental Figure 6, A-B).

As IDO1 and PD-L1 have been targeted in cancer immunotherapy and hold promise for drug re-purposing, we next sought to further validate these findings using quantitative IHC in an independent cohort of CL patients (5 females, 18 males; mean age ± standard deviation, 44 ± 11 years; time to diagnosis 7.76 ± 8.2 months; Supplemental Figures 7 and 8 and Supplemental Table 4) sampled at baseline and after 4 weeks of treatment. Using an accepted cut-off of >5% of cells being positive (30), all patients (n=23) expressed IDO1 (Histochemical (H)-score (31) median = 81.2; range 16 - 165) and 20/23 patients had a reduction in the abundance of $IDO1^+$ cells on treatment (H-score median = 32; range 1 – 171; p=0.0023; Figure 2J). All patients were PD-L1 positive at presentation (n=23; H-score median = 82.8; range 12-164) and 20/23 patients exhibited a reduction in the number of PD-L1 expressing cells on treatment (Figure 2J; H-score median = 36.7; range 12.3-36.7; p=0.0008). Collectively, these data indicate that IDO1 and PD-L1 are highly expressed in the lesions of Sri Lankan CL patients and reduction in expression of these two checkpoint molecules represents an early response to SSG.

Though in vitro studies have indicated that intracellular parasitism by *Leishmania* could impact on the expression of immune checkpoint molecules (32-34), this has not been established in situ during human disease. To address this question, we combined IHC with RNA-FISH (35) to identify *Amastin* transcripts (as a surrogate for viable amastigotes) with a bespoke StrataQuest image analysis pipeline (Supplemental Figure 9, A-F). In 7 patients studied that were *Amastin*$^+$ at presentation (Supplemental Methods, Supplemental Table 5), PD-L1 expression co-localised with CD68$^+$ macrophages (Figure 3A, Supplemental Figure 10C) and parasitized cells were both PD-L1$^+$ and PD-L1$^-$ (Figure 3A). We binned the Amastin$^+$ PDL1$^+$ and Amastin $^-$PDL1$^+$ cells based on PD-L1 mean fluorescent intensity (Figure 3, B-D) and found that cells containing abundant *Amastin* transcripts expressed more PD-L1 than cells with less or no *Amastin* transcripts (Figure 3, B-E, Supplemental Figure 9, G-L and Supplemental Figure 10). To independently corroborate this observation, we showed that a Sri Lankan strain of *L. donovani* was also capable of inducing up-regulation of PD-L1 expression on human monocyte-derived macrophages in vitro (Supplemental Figure 11, A-F) , as previously described for *L. major* (34). Similarly, IDO1 extensively co-localised with CD68$^+$ cells (Supplemental Figure 11A) and both IDO1$^+$CD68$^+$ and IDO1$^-$CD68$^+$ cells were infected (Supplemental Figure 11B). Using a similar gating strategy (Supplemental Figure 11C-H; n=3 patients), we found that cells with abundant *Amastin* transcripts expressed more IDO1 than those with fewer or no *Amastin* transcripts (Supplemental Figure 11, I-K). These data show that, although a notable population of uninfected CD68$^+$ cells contribute to PD-L1 and IDO-1 expression within CL lesions, intracellular parasitism leads to heightened expression of these checkpoint molecules in lesional monocytes and macrophages.

Finally, we tested whether reduction in IDO1 or PD-L1 expression early during therapy could be used as a prognostic marker for treatment response. Patients with the greatest reduction in

PD-L1 expression (i.e. greater than the geomean of the pre-treatment: on-treatment expression ratio; n=12 patients) (Figure 4, A-B) cured earlier than those that had lower or no reduction in PD-L1 expression (p=0.015). Patients with lower PD-L1 expression after 4 weeks of treatment (i.e. lower than the geomean of on-treatment expression; n=12 patients) also cured faster (p=0.0045; Figure 4B). We assessed the association of PD-L1 with disease cure rate using univariate Cox Proportional Hazard regression (Supplemental Figure S13A; Hazard Ratio (HR) = 3.96, p=0.008). Upon adjustment for age and gender of the participants, HR increased to 4.88 (p= 0.007; Figure 4D), indicating that patients that maximally reduced PD-L1 expression upon treatment were about 5 times more likely to cure earlier. Conversely, patients remaining parasite PCR^+ at 4 weeks post treatment had a significantly longer cure time (Figure 4E) and higher PD-L1 expression (Figure 4F). Surprisingly, reduction in IDO1 expression, calculated as either pre-treatment: on-treatment expression ratio or IDO1 expression at 4 weeks (n=12 vs 11), did not correlate with cure rate (Supplemental Figure 13, B and C). Thus, the relationship between declining PD-L1 expression and rate of cure (Figure 4, E-F) appears selective.

We conclude that expression of IDO1 and PD-L1 immune checkpoint molecules is a common feature of Sri Lankan CL and that intracellular parasitism is associated with heightened expression of these immunoregulatory proteins in lesional macrophages. Tissue expression of both IDO1 and PD-L1 reduces significantly within 2-4 weeks of treatment onset and well in advance of clinical cure, and a reduction in PD-L1 is associated with a more rapid therapeutic response. The elevated expression of negative immune regulators on macrophages at the lesion site, as shown here, has clear parallels with tumour-associated macrophages (36) and extends our understanding of how *Leishmania* parasites influence the function of their host cell during human disease (37). Though longitudinal sampling of the

same macrophage population was not possible, it seems likely that reduction of PD-L1 expression is facilitated by the leishmanicidal action of SSG, suggesting a model for drug-immune synergy whereby early rounds of SSG treatment reduce intracellular parasite burden leading to reduced checkpoint inhibition and re-engagement of T cell effector function. Our data, together with strong pre-clinical evidence of an inhibitory role of PD-L1 in various forms of leishmaniasis (10, 12, 38) supports the candidacy of PD-L1 blockade as an adjunct HDT in Sri Lankan CL. In addition, our data suggest the possibility that changes in PD-L1 expression early after treatment could be considered as a biomarker to trigger drug tapering or drug cessation.

Methods

Information is provided in Supplemental methods.

Study approval

The study was conducted in accords with the principles of the Declaration of Helsinki and was approved by the Ethical Review Committee of the Faculty of Medical Sciences, University of Jayewardenepura (Ref: 780/13 & 52/17) and the Department of Biology, University of York. Written informed consent, including for lesion photographs, was received from participants prior to inclusion in this study.

Author Contributions

NSD, SS, VS, NM, BS, MMDO, LR, SM and SR conducted experiments. NSD and SF performed data analysis. NSD and PMK wrote the manuscript. PW, MC, HG, RW, DL, PMK and SR were involved in conceptualisation and securing funding and PMK and SR supervised the study. The order of the co–first authors was determined by their relative contribution to this study.

207 **Acknowledgements**

208 The authors thank Dr. Pushpa Ilanngasinghe (histopathologist, Teaching Hospital
209 Anuradhapura, Sri Lanka), Dr. Dawei Chen (phlebotomist) and Karen Hogg (flow cytometry
210 expert) at University of York, technical support at Nanostring Technologies, TissueGnostics
211 and Centre for Genomic Research, University of Liverpool. This work was supported by
212 funding from the UK Medical Research Council / UK Aid Global Challenges Research Fund
213 (MR/P024661/1 to PMK, SR, HG and MC) and a Wellcome Trust Senior Investigator Award
214 (WT104726 to PMK). The funders had no role in the design or conduct of the study or the
215 decision to publish.

216

References

1. Pires M, Wright B, Kaye PM, da Conceicao V, and Churchill RC. The impact of leishmaniasis on mental health and psychosocial well-being: A systematic review. *PLoS One.* 2019;14(10):e0223313.
2. Alvar J, Velez ID, Bern C, Herrero M, Desjeux P, Cano J, et al. Leishmaniasis worldwide and global estimates of its incidence. *PLoS One.* 2012;7(5):e35671.
3. Drugs for Neglected Diseases initiative. About leishmaniasis. https://dndi.org/diseases/cutaneous-leishmaniasis/.
4. Athukorale DN, Seneviratne JK, Ihalamulla RL, and Premaratne UN. Locally acquired cutaneous leishmaniasis in Sri Lanka. *J Trop Med Hyg.* 1992;95(6):432-3.
5. Karunaweera ND, Pratlong F, Siriwardane HV, Ihalamulla RL, and Dedet JP. Sri Lankan cutaneous leishmaniasis is caused by Leishmania donovani zymodeme MON-37. *Trans R Soc Trop Med Hyg.* 2003;97(4):380-1.
6. Ranasinghe S, Zhang WW, Wickremasinghe R, Abeygunasekera P, Chandrasekharan V, Athauda S, et al. Leishmania donovani zymodeme MON-37 isolated from an autochthonous visceral leishmaniasis patient in Sri Lanka. *Pathog Glob Health.* 2012;106(7):421-4.
7. Zhang WW, Ramasamy G, McCall LI, Haydock A, Ranasinghe S, Abeygunasekara P, et al. Genetic analysis of Leishmania donovani tropism using a naturally attenuated cutaneous strain. *PLoS Pathog.* 2014;10(7):e1004244.
8. Refai FW, Madarasingha NP, Fernandopulle R, and Karunaweera N. Nonresponsiveness to standard treatment in cutaneous leishmaniasis: A case series from Sri Lanka. *Trop Parasitol.* 2016;6(2):155-8.
9. Scott P, and Novais FO. Cutaneous leishmaniasis: immune responses in protection and pathogenesis. *Nat Rev Immunol.* 2016;16(9):581-92.
10. da Fonseca-Martins AM, Ramos TD, Pratti JES, Firmino-Cruz L, Gomes DCO, Soong L, et al. Immunotherapy using anti-PD-1 and anti-PD-L1 in Leishmania amazonensis-infected BALB/c mice reduce parasite load. *Sci Rep.* 2019;9(1):20275.
11. Brown JA, Titus RG, Nabavi N, and Glimcher LH. Blockade of CD86 ameliorates Leishmania major infection by down-regulating the Th2 response. *J Infect Dis.* 1996;174(6):1303-8.
12. Liang SC, Greenwald RJ, Latchman YE, Rosas L, Satoskar A, Freeman GJ, et al. PD-L1 and PD-L2 have distinct roles in regulating host immunity to cutaneous leishmaniasis. *Eur J Immunol.* 2006;36(1):58-64.
13. Greenwald RJ, McAdam AJ, Van der Woude D, Satoskar AR, and Sharpe AH. Cutting edge: inducible costimulator protein regulates both Th1 and Th2 responses to cutaneous leishmaniasis. *J Immunol.* 2002;168(3):991-5.
14. Mou Z, Muleme HM, Liu D, Jia P, Okwor IB, Kuriakose SM, et al. Parasite-derived arginase influences secondary anti-Leishmania immunity by regulating programmed cell death-1-mediated CD4+ T cell exhaustion. *J Immunol.* 2013;190(7):3380-9.
15. Makala LH, Baban B, Lemos H, El-Awady AR, Chandler PR, Hou DY, et al. Leishmania major attenuates host immunity by stimulating local indoleamine 2,3-dioxygenase expression. *J Infect Dis.* 2011;203(5):715-25.
16. Okwor I, Xu G, Tang H, Liang Y, Fu YX, and Uzonna JE. Deficiency of CD40 Reveals an Important Role for LIGHT in Anti-Leishmania Immunity. *J Immunol.* 2015;195(1):194-202.
17. Akiba H, Miyahira Y, Atsuta M, Takeda K, Nohara C, Futagawa T, et al. Critical contribution of OX40 ligand to T helper cell type 2 differentiation in experimental leishmaniasis. *J Exp Med.* 2000;191(2):375-80.
18. Dalton JE, and Kaye PM. Immunomodulators: use in combined therapy against leishmaniasis. *Expert Rev Anti Infect Ther.* 2010;8(7):739-42.
19. Sbaraglini ML, Vanrell MC, Bellera CL, Benaim G, Carrillo C, Talevi A, et al. Neglected Tropical Protozoan Diseases: Drug Repositioning as a Rational Option. *Curr Top Med Chem.* 2016;16(19):2201-22.
20. Rao SPS, Barrett MP, Dranoff G, Faraday CJ, Gimpelewicz CR, Hailu A, et al. Drug Discovery for Kinetoplastid Diseases: Future Directions. *ACS Infect Dis.* 2019;5(2):152-7.

21. Farias Amorim C, F ON, Nguyen BT, Nascimento MT, Lago J, Lago AS, et al. Localized skin inflammation during cutaneous leishmaniasis drives a chronic, systemic IFN-gamma signature. *PLoS Negl Trop Dis.* 2021;15(4):e0009321.
22. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. *Nucleic Acids Res.* 2019;47(D1):D607-D13.
23. Goncalves R, Zhang X, Cohen H, Debrabant A, and Mosser DM. Platelet activation attracts a subpopulation of effector monocytes to sites of Leishmania major infection. *J Exp Med.* 2011;208(6):1253-65.
24. Leon B, Lopez-Bravo M, and Ardavin C. Monocyte-derived dendritic cells formed at the infection site control the induction of protective T helper 1 responses against Leishmania. *Immunity.* 2007;26(4):519-31.
25. Ives A, Ronet C, Prevel F, Ruzzante G, Fuertes-Marraco S, Schutz F, et al. Leishmania RNA virus controls the severity of mucocutaneous leishmaniasis. *Science.* 2011;331(6018):775-8.
26. Novais FO, Carvalho LP, Passos S, Roos DS, Carvalho EM, Scott P, et al. Genomic profiling of human Leishmania braziliensis lesions identifies transcriptional modules associated with cutaneous immunopathology. *J Invest Dermatol.* 2015;135(1):94-101.
27. Santos Cda S, Boaventura V, Ribeiro Cardoso C, Tavares N, Lordelo MJ, Noronha A, et al. CD8(+) granzyme B(+)-mediated tissue injury vs. CD4(+)IFNgamma(+)-mediated parasite killing in human cutaneous leishmaniasis. *J Invest Dermatol.* 2013;133(6):1533-40.
28. Boussoffara T, Boubaker MS, Ben Ahmed M, Mokni M, Feriani S, Ben Salah A, et al. Activated cytotoxic T cells within zoonotic cutaneous leishmaniasis lesions. *Immun Inflamm Dis.* 2019;7(3):95-104.
29. Beechem JM. High-Plex Spatially Resolved RNA and Protein Detection Using Digital Spatial Profiling: A Technology Designed for Immuno-oncology Biomarker Discovery and Translational Research. *Methods Mol Biol.* 2020;2055:563-83.
30. Powles T, Eder JP, Fine GD, Braiteh FS, Loriot Y, Cruz C, et al. MPDL3280A (anti-PD-L1) treatment leads to clinical activity in metastatic bladder cancer. *Nature.* 2014;515(7528):558-62.
31. Igarashi T, Teramoto K, Ishida M, Hanaoka J, and Daigo Y. Scoring of PD-L1 expression intensity on pulmonary adenocarcinomas and the correlations with clinicopathological factors. *ESMO Open.* 2016;1(4):e000083.
32. Roy S, Saha S, Gupta P, Ukil A, and Das PK. Crosstalk of PD-1 signaling with the SIRT1/FOXO-1 axis during the progression of visceral leishmaniasis. *J Cell Sci.* 2019;132(9).
33. Donovan MJ, Tripathi V, Favila MA, Geraci NS, Lange MC, Ballhorn W, et al. Indoleamine 2,3-dioxygenase (IDO) induced by Leishmania infection of human dendritic cells. *Parasite Immunol.* 2012;34(10):464-72.
34. Filippis C, Arens K, Noubissi Nzeteu GA, Reichmann G, Waibler Z, Crauwels P, et al. Nivolumab Enhances In Vitro Effector Functions of PD-1(+) T-Lymphocytes and Leishmania-Infected Human Myeloid Cells in a Host Cell-Dependent Manner. *Front Immunol.* 2017;8:1880.
35. Wang F, Flanagan J, Su N, Wang LC, Bui S, Nielson A, et al. RNAscope: a novel in situ RNA analysis platform for formalin-fixed, paraffin-embedded tissues. *J Mol Diagn.* 2012;14(1):22-9.
36. Cassetta L, and Pollard JW. Targeting macrophages: therapeutic approaches in cancer. *Nat Rev Drug Discov.* 2018;17(12):887-904.
37. Kaye P, and Scott P. Leishmaniasis: complexity at the host-pathogen interface. *Nat Rev Microbiol.* 2011;9(8):604-15.
38. Joshi T, Rodriguez S, Perovic V, Cockburn IA, and Stager S. B7-H1 blockade increases survival of dysfunctional CD8(+) T cells and confers protection against Leishmania donovani infections. *PLoS Pathog.* 2009;5(5):e1000431.

Figure legends

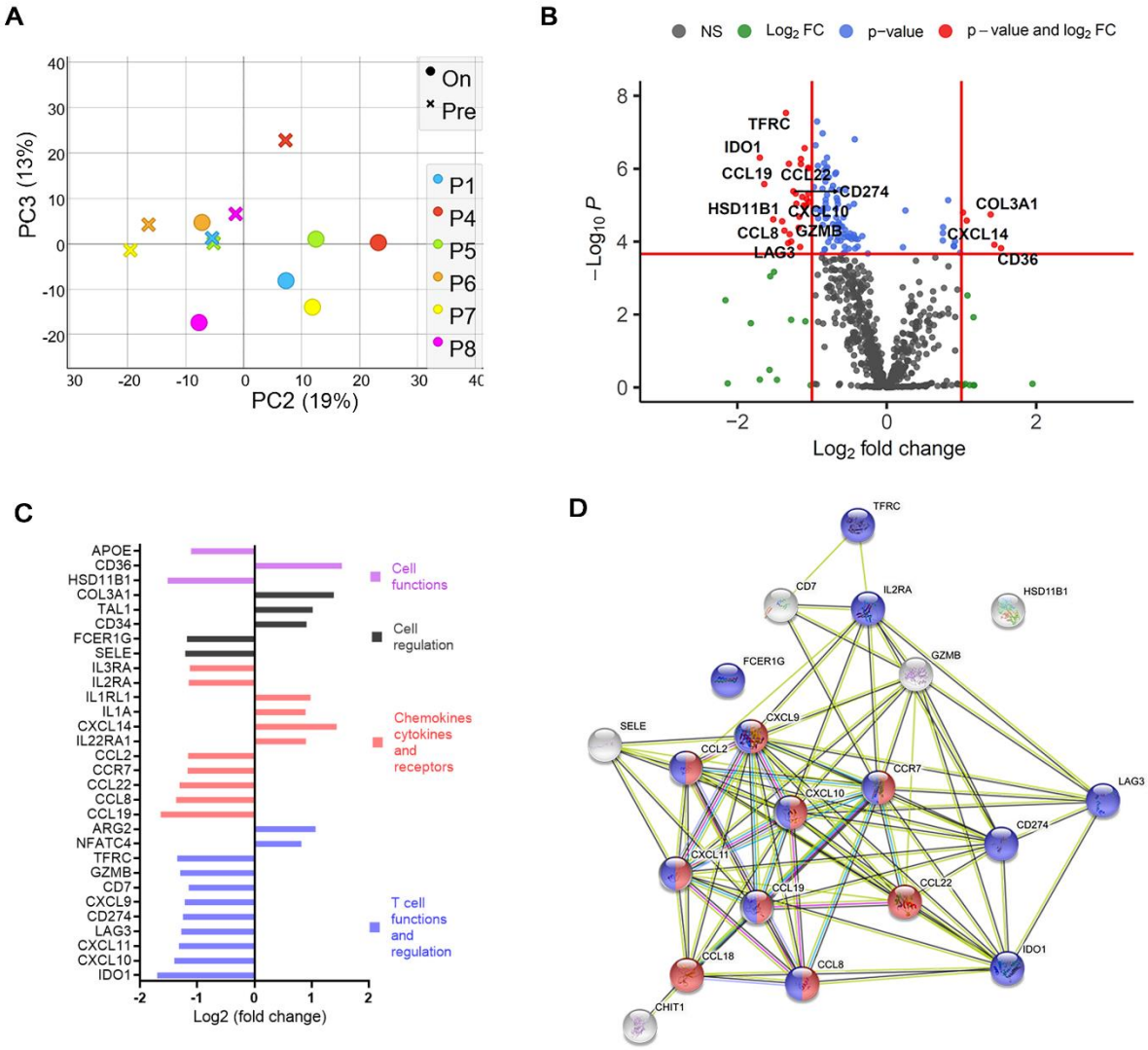


Figure 1. Differential expression and network analysis of genes regulated by drug treatment in lesions of Sri Lankan CL patients.

Immune-targeted tissue transcriptomics was conducted on tissue sections from test cohort patients comparing transcriptomes at presentation and on treatment. (**A**) Principal component analysis was performed to show differences between pre- and on-treatment transcriptome of each patient based on 770 gene nCounter PanCancer Immunology Panel (n=6) **(B)** Differentially expressed genes comparing pre-treatment biopsies with biopsies taken after two weeks on treatment (SSG). Cut off (red line) drawn at equivalent of adjusted p-value =0.01 and Log (Fold change) of 1**(C)** Top 30 genes that changed in expression on SSG treatment. **(D)** STRING protein-protein interaction network (22)(https://string-db.org) analysis of genes listed in **Supplemental Table 3** down-regulated on SSG treatment. Pathways represent GO: 0072676, Lymphocyte migration (red spheres) and GO: 0002684, positive regulation of immune system process (blue spheres). Top 20 genes are shown (Log2fold change ≥1.15) for clarity.

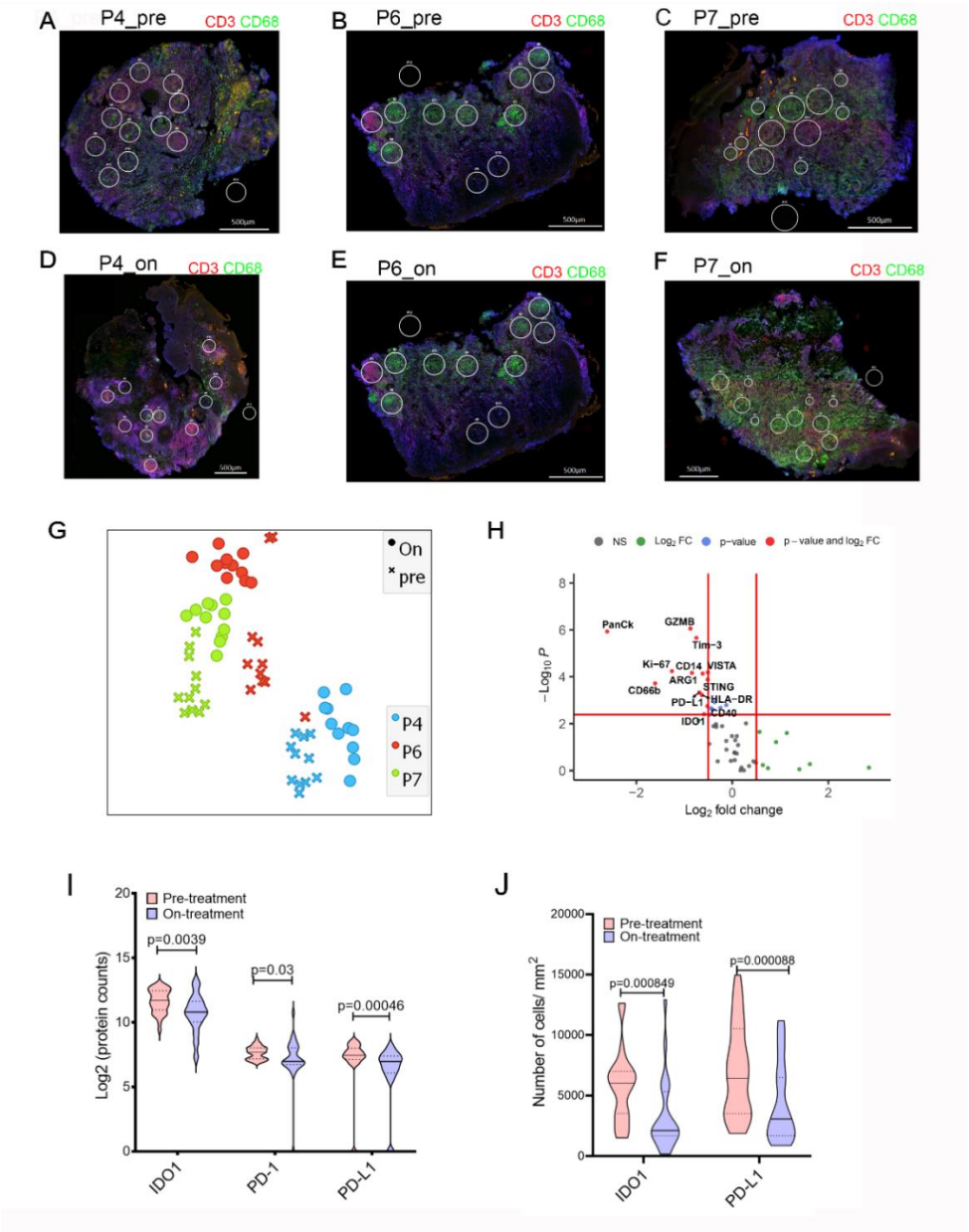


Figure 2. Digital Spatial Profiling (DSP) of CL lesions.

DSP was performed on tissue sections from test cohort individuals comparing ROIs from pre and on-treatment biopsies**. (A-F)** ROIs on $CD3^+$ and/or $CD68^+$ rich areas from pre and on-treatment biopsies from patients P4, P6 and P7 (CD68, green; CD3, red; Syto13, blue). 20x magnification; scale bar, 500µm **(G)** t-SNE plot based on 20 PCA loadings coloured on patient ID. (**H**) Differential protein expression analysis comparing pre-treatment to on-treatment ROIs. Red lines indicate adjusted p value cut off of 1% (Mann-Whitney test with FDR correction based on Benjamini, Krieger, and Yekutieli two stage set-up method) and and Log2FC = 0.5 (n=33 ROIs) (**I**) IDO1, PD-1 and PD-L1 expression in pre- and on-treatment ROIs. Mann Whitney rank test (n=33 ROIs). (**J**) Immunohistochemistry (IHC) was performed on sections from patients pre and on-treatment from the validation cohort and quantitated using StrataQuest (see Methods) (n=23). Wilcoxon matched-pairs signed rank test. Dotted lines show upper and lower quantile in **I-J,** median by solid line.

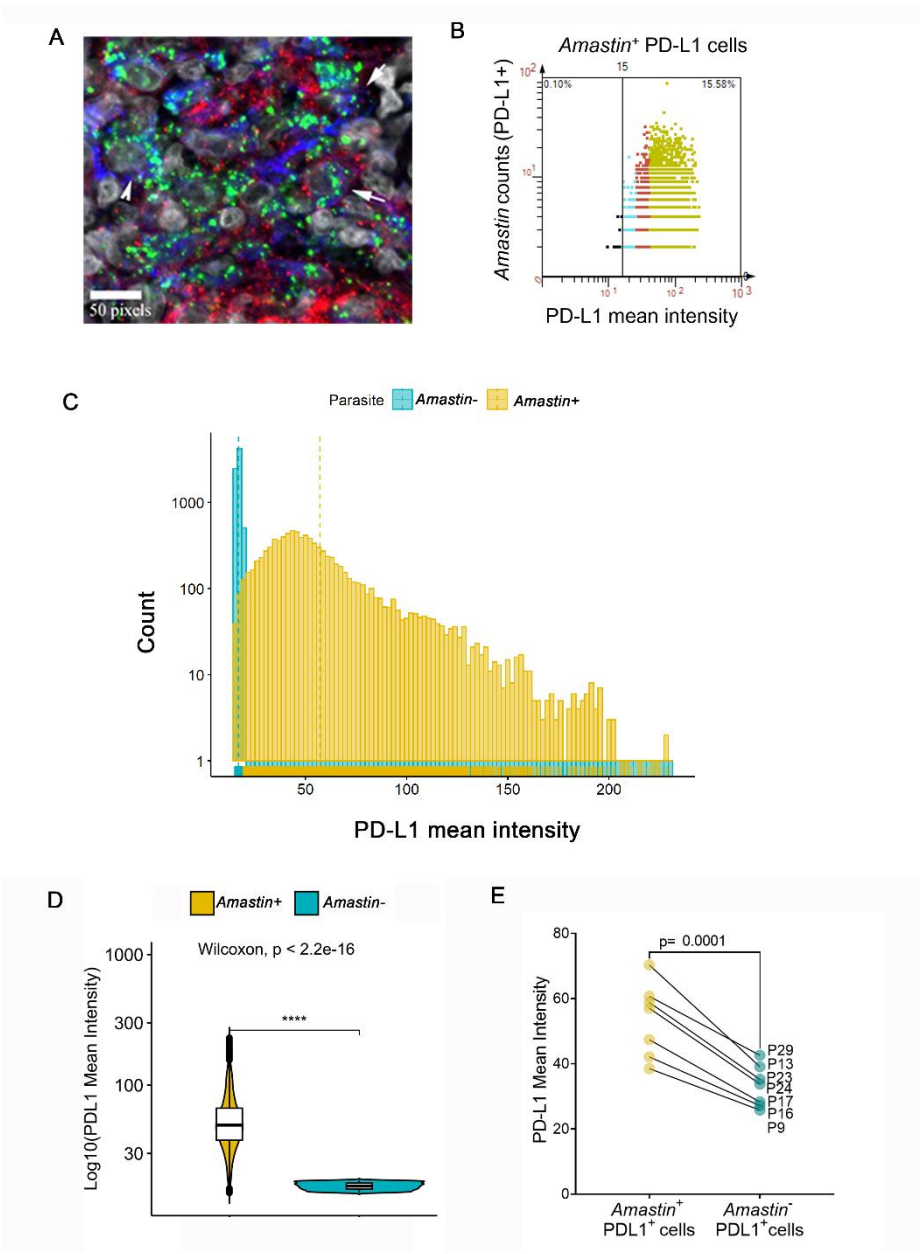


Figure 3 Imunofluorescence analyses of PD-L1 in infected and uninfected cells

Dual IHC-FISH using an *Amastin* probe was performed on pre-treatment sections of patients enrolled in the validation cohort. **(A)** A 400x confocal image showing infection of PD-L1^{+}CD68^{+} (arrows) and PD-L1^{-}CD68^{+} (arrowhead) cells. Scale bar, 50 pixels **(B)** Relationship between PD-L1 expression and parasite burden (*Amastin* dot count). Scattergram from a representative patient (P24 at presentation) showing *Amastin*$^{+}$ low (cyan), medium (red) and high (green) PD-L1 expressing cells with respect to parasite abundance. **(C)** Fluorescence intensity distributions of infected and uninfected PD-L1 cells **(D)** Mean fluorescent intensity of PD-L1 expression on *Amastin*$^{-}$ cells compared to *Amastin*$^{+}$ cells from a representative patient P24. The upper and lower whisker represents highest and lowest value that is within 1.5 * interquartile range. N=9159 parasite positive cells and N=41520 for parasite negative cells. Significance score was generated using Wilcoxon signed rank test. **(E)** PD-L1 expression on *Amastin*$^{+}$PD-L1^{+} cells vs. *Amastin*$^{-}$PD-L1^{+} cells (n=7 patients). Significance score was generated using Students two-tailed paired t-test after testing for normality using Shapiro Wilk and Kolkogorov-Smirnov tests.

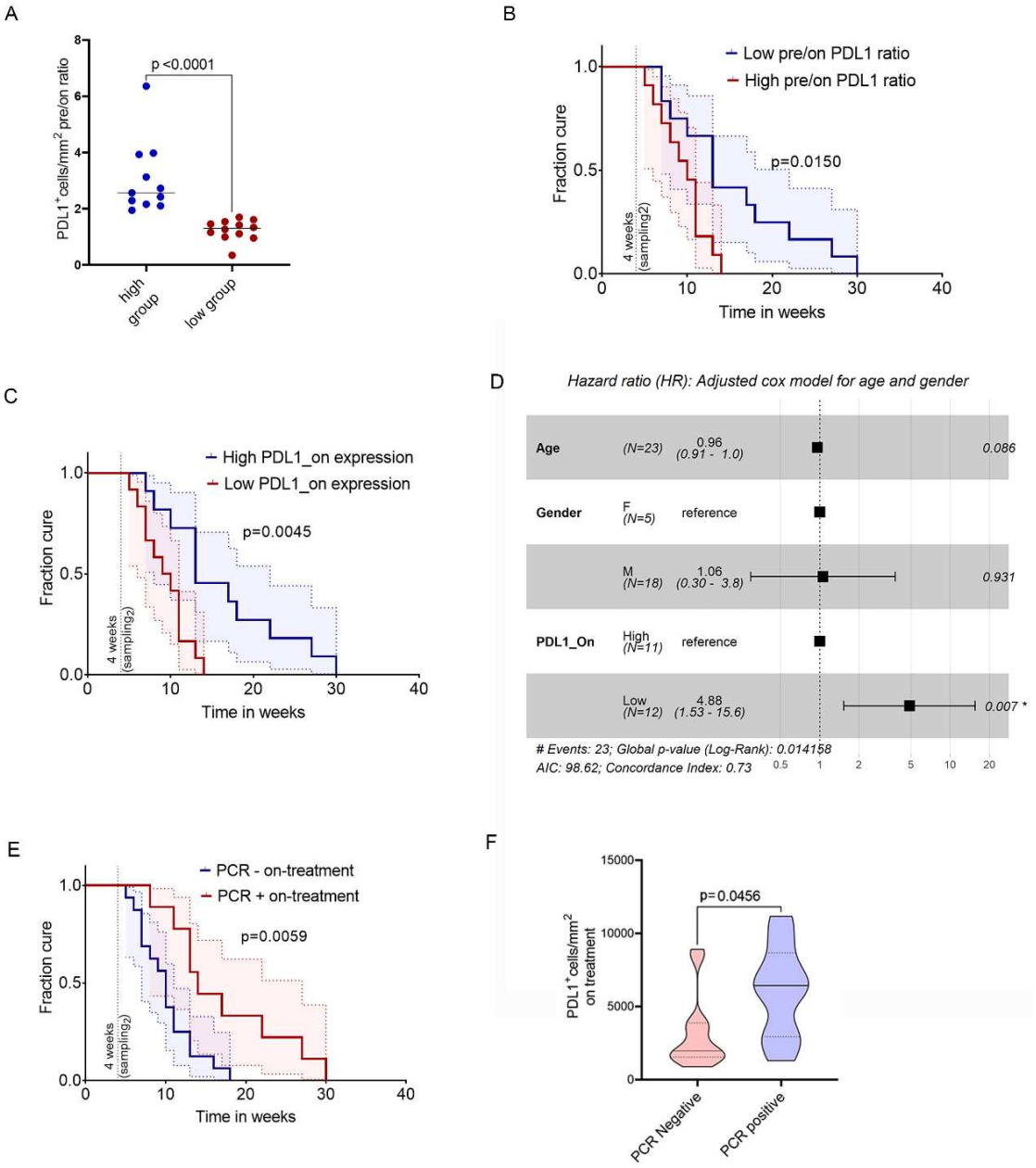


Figure 4 Clinical correlates of PDL1 reduction on treatment in CL patients

(A) Patients (validation cohort; n=23) were stratified based on high (>geomean value; n=11) and low (< geomean value; n=12) pre-: on-treatment expression ratio. **(B)** Kaplan-Meier curve based on pre-:on-treatment ratio of PD-L1 expression (high vs low). **(C)** Patients stratified based on on-treatment expression of PD-L1 (> geomean value; n=11 vs < geomean value; n=12). **(D)** Multivariate Cox Proportional Hazards model plotted as a forest plot. p-values for each covariate represent Wald statistic value and overall statistical significance is also indicated. **(E)** Patients stratified by *LITS1* PCR status (n= 9 PCR^+ vs n=14 PCR^- or $^{+/-}$ (equivocal)) on treatment. **(F)** PD-L1 expression in *LITS* PCR^+ vs. PCR^- individuals on treatment. Dotted lines show upper and lower quantile, solid line shows median. P-value generated using two-tailed Mann-Whitney test. Vertical line drawn in **B, C, E** on the X axis shows time when on-treatment biopsies were collected. Curves in **B, C, E** were compared using Log-rank (Mantel-Cox) test. Blue and red shaded area show 95% CI of the two groups.