



Deposited via The University of Sheffield.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/176076/>

Version: Accepted Version

---

**Article:**

Howard, F. and Muthana, M. (2020) Designer nanocarriers for navigating the systemic delivery of oncolytic viruses. *Nanomedicine*, 15 (1). pp. 93-110. ISSN: 1743-5889

<https://doi.org/10.2217/nnm-2019-0323>

---

© 2021 Future Science Group. This is an author-produced version of a paper subsequently published in *Nanomedicine*. Uploaded in accordance with the publisher's self-archiving policy.

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.

# Nanomedicine

## Designer nanocarriers for navigating the systemic delivery of oncolytic viruses

Journal:	<i>Nanomedicine</i>
Manuscript ID	NNM-2019-0323.R1
Manuscript Type:	Review
Keywords:	Cancer/oncology, Immunology/Infectious diseases, Metal nanoparticles, Nanoparticles, Viruses

SCHOLARONE™  
Manuscripts

---

## Article Body Template

**PLEASE DO NOT INCLUDE ANY IDENTIFYING INFORMATION IN THE MAIN BODY OF THE MANUSCRIPT**

- **Abstract:**

Nanotechnology is paving the way for new carrier systems designed to overcome the greatest challenges of oncolytic virotherapy (OV); systemic administration and subsequent implications of immune responses and specific cell binding and entry. Systemic administration of oncolytic agents is vital for disseminated neoplasms, however transition of nanoparticles (NP) to virotherapy has yielded modest results. Their success relies on how they navigate the merry-go-round of often-contradictory phases of nanoparticle delivery: circulatory longevity, tissue permeation and cellular interaction, with many studies postulating design features optimal for each phase. This review discusses the optimal design of nanoparticles for OV transport within these phases, to determine whether improved virotherapeutic efficacy lies in the pharmacokinetic/pharmacodynamics characteristics of the NP-OV complexes rather than manipulation of the virus and targeting ligands.

- **Lay abstract:** optional – N/A

- **Graphical abstract:** optional – N/A

- **Video abstract:** optional – N/A

- **Keywords:** Oncolytic virotherapy, nanomedicine, nanoparticles, nanotargeting, immunotherapy, magnetic guidance

- **Main body of text:**

### 1. Introduction

Cancer killing oncolytic viruses (OV) are a promising treatment modality for cancer. However, variable clinical response rates [1] have shifted the impetus towards various carrier strategies for their systemic delivery including liposomes, polymers, cell carriers and metallic nanoparticles to name a few. Carrier systems are not new in the field of oncology with many small molecule inhibitors (SMIs) embracing their advantages including biocompatibility, increased endocytosis, solubility and extended circulation time compared to free anticancer drugs, whilst simultaneously reducing systemic toxicity [2]. Successful examples include liposomal doxorubicin and nanoparticle albumin bound paclitaxel, which have progressed to the clinic. Viruses face similar pharmacokinetic and pharmacodynamic (PK/PD) challenges to SMI's which have been mitigated to a certain extent by the transfer of carrier systems to the field of virotherapy. To successfully reach target tumour cells by a passive system the ideal therapeutic agent must traverse three major phases for nanoparticle drug delivery for oncology; systemic circulation and reticuloendothelial system (RES) interaction, extravasation and tumour penetration and interaction with

---

## Article Body Template

target cells. The physical characteristics (shape and size) and chemical characteristics (composition and charge) of nanoparticles are significant determinants of their functionalities within these three phases yet they have contradictory requirements leading to a paradoxical merry-go-round (Figure 1) that often requires sacrifice of design features optimal for at least one of the other phases. From this perspective it is even more striking how similar in both composition and mode of action the current raft of available nanoparticles are (Figure 2). We will explore whether this represents a conscious decision by investigators to focus on one phase they believe to be the most important or whether other limiting factors such as fabrication technologies have driven these decisions. This review will discuss the PK/PD of nanoparticles and whether this knowledge can be applied for the generation of virotherapeutic complexes endeavouring to reconcile the opposing determinants of their success.

### 2. Pharmacokinetics of nanoparticles for oncology

The influence that the characteristics of nanoparticles have on their transport and interaction with cells is well known and has been extensively reviewed by Ernsting et al [3] within the context of oncology. The key points are represented in Figure 3 and briefly discussed below.

#### Morphology

Size and shape influences nanoparticle transport behaviour including how they interact with the endothelial cells of blood vessels [4] and capillaries wall for extravasation before being removed from the blood either by the RES or filtered by the lungs, liver and spleen. Particles larger than 5 $\mu$ m are trapped within the capillary beds of the liver and between 1-5 $\mu$ m they are phagocytosed by Kupffer cells [5]. Circulating nanoparticles exceeding 100nm in diameter are rapidly phagocytosed by hepatic and splenic macrophages, as opsonisation by serum proteins increases with size [3, 6]. However, to enhance extravasation into tissues, nanoparticles approaching 100nm are more likely to marginate towards the periphery of the blood vessel [7]. Radiolabelled metal organic frameworks (MOFs) of 60nm demonstrated longer blood circulation and over 50% higher tumour accumulation than 130 nm MOFs [8]. Formation of nanoparticle aggregates is also a concern for nanodrugs due to the risk of embolism and changes in biodistribution, with aggregates of polystyrene aggregates demonstrating a higher propensity for the reticuloendothelial systems [3, 9]. However, aggregation of aerosolised gold nanoparticles demonstrated

---

## Article Body Template

1 significantly faster cellular uptake than single AuNPs at the air-blood barrier interface using a multicellular  
2 lung system [10] once again forcing investigators onto the merry-go-round of decision-making as to which  
3 delivery phase is most important to them (Figure 1).

4  
5  
6  
7  
8  
9  
10  
11  
12 Once within the tissue, size also affects permeation and cell internalisation. Small nanoparticles show high  
13 permeation rate but also are rapidly cleared from the tumour by RES leading to poor accumulation (Figure  
14 3) [11]. Never is size more important than when considering transport across the blood-brain barrier for  
15 treatment of brain neoplasms. A recent study in a mouse model of orthotopic glioblastoma multiforme  
16 (GBM) demonstrated higher distribution of gold nanoparticles (AuNPs) within tumorous tissue compared  
17 to normal tissue when perfused via the carotid artery. Pertinently, nanoparticle size affected the  
18 permeation of nanoparticles with 10nm AuNPs widely distributed throughout the brain tumour, whereas  
19 50 and 100 nm AuNPs were located near the blood vessels [12]. By using carotid infusion, the investigators  
20 were able to bypass renal filtration, however, clinically this would be considered a highly invasive  
21 procedure with usual standard of care recommending intravenous administration therefore exposing  
22 these particles to rapid elimination from the circulation.

23  
24  
25  
26  
27  
28  
29  
30  
31 NP size may affect the uptake efficiency and kinetics, the internalisation mechanism (eg. clathrin or  
32 caveolin mediated endocytosis and phagocytosis) [13] and also the subcellular distribution. A size-  
33 dependent uptake in different cell lines has been observed as well as size dependent cytotoxicity which is  
34 extensively reviewed by Shang et al [14]. They concluded that the optimal size for active cellular uptake  
35 required a NP core size of 30-50nm. However this is within the range that NP's are rapidly cleared by the  
36 RES following systemic administration. Recently it has also been postulated that tumour volume can  
37 selectively change tumour uptake of nanoparticles of varying size and that this depends on the frequency  
38 of interaction of particles with the perivascular extracellular matrix for smaller nanoparticles, whereas  
39 transport of larger nanomaterials is dominated by Brownian motion [15], adding a further layer of  
40 complexity to nanocarrier design.

41  
42  
43  
44  
45  
46  
47  
48  
49 There is also evidence that the size of NPs can influence the therapeutic effect. Naked AuNPs can inhibit  
50 the function of pro-angiogenic heparin-binding growth factors (HB-GFs) and subsequent intracellular  
51 signalling events. Using AuNPs of 10, 20 and 30nm, investigators demonstrated that the specific inhibitory  
52 effects of AuNPs towards HB-GFs are size dependent, bigger nanoparticles being more efficacious at  
53 inhibiting proliferation of HUVEC and NIH3T3 cells *in vitro* (100% inhibition at 5nM/L) [16].

---

## Article Body Template

The shape effect is also known to influence fluid dynamics including lateral drift of particles towards the blood vessel wall, how particles interact with tumour capillaries during transport and their role in intracellular uptake [17]. However, it is only in recent years that this characteristic has begun to draw interest, most likely previously hindered by the lack of fabrication technology. In terms of circulation time, investigations have demonstrated, using Tobacco Mosaic Virus (TMV) in mice, that spherical NPs are cleared more rapidly from tissues in comparison to nano-rods of the same chemical composition [18]. Moreover, TMV has been used to generate a computational simulation to model nanoparticle diffusion within a small segment of solid tumour between capillaries [19]. They concluded that smaller aspect ratios (AR) (rod-shaped) NPs have higher diffusion and accumulate more easily in the tumour tissue, however higher aspect ratio NPs possess enhanced margination (the ability of particles to migrate towards blood vessel walls in blood flow), increased transport across tissue membranes and reduced clearance by phagocytosis. Likewise, Lee et al (2009) observed that whilst elongated particles (small AR) had a greater propensity to marginate in linear laminar flow, this could only be achieved through the application of external forces (gravitational, magnetic) within blood microcirculation [20]. Internalisation of particles was dependent on both the shape and absolute size and/or volume of the particle with larger rod-shaped particles internalised by HeLa cells 4 times faster than symmetrical particles and smaller rod-shaped particles [21], possibly due to the larger surface areas in contact with the cell membrane. Spherical metallic nanoparticles displayed greater cytotoxicity compared to rod-shaped [22] and star-shaped [23] nanoparticles in human skin fibroblasts although results from these studies could also be attributed to size and surface chemistry.

### Surface characteristics

To overcome the general lack of specificity of nanoparticles and poor biodistribution a myriad of coatings exist purporting enhanced functionalisation capabilities, biocompatibility and aggregation reduction. The most successful strategy in this endeavor has been the wide-spread use of polyethylene glycol (PEG), a water-soluble synthetic polymer that creates a stable hydration layer resistant to protein adsorptions to underlying surfaces. However an increase in PEGylated proteins and PEG-modified NPs including liposomes and micelles has seen a correlation of anti-PEG antibodies with loss of therapeutic efficacy both in the clinic [24, 25] and in animal models [26, 27]. What once was used as a shield against opsonisation and immunosurveillance is now a recognised determinant for their clearance [28] resulting in loss of

---

## Article Body Template

therapeutic efficacy and increase in adverse effects. Biomimetic coatings may represent the future, where nanoparticles are recognised as “self” using membrane materials such as red blood cell membrane-coated nanoparticles which may enhance glioma therapy efficacy [29] and has demonstrated 29% overall retention after 24 hours within the circulation of mice compared to bare NPs that showed negligible signal at the first 2 minute timepoint [30]. Surface coatings can also change the charge of the NPs and exert a major effect on their properties including drug loading and release rates, systemic circulation and absorption [31]. As shown in Figure 3, charged NPs are more prone to serum protein opsonisation and interactions with negatively charged plasma membranes compared to neutral NPs yet cationic NPs display enhanced cellular uptake, once again forcing the investigator to choose between enhanced circulatory time or target cell interactions (Figure 1).

### 3. Virotherapy

Dysfunctional cellular processes that are the markers of neoplasms provide the ideal environment for viral infection including sustained proliferation, resistance to cell death, the evasion of growth suppressors and immune destruction, genome instability and DNA damage stress [32]. The attraction to virotherapy in comparison to conventional treatments is the unique ability of OV to only kill cancer cells. For example, inherently selective RNA viruses such as the measles virus vaccine strain MV\_SPUD, reovirus and Newcastle disease virus take advantage of tumour cell resistance to interferon [33], whilst vaccinia virus, adenovirus, Herpes simplex virus (HSV) and polioviruses exploit the deletion of viral genes [34-37], that are necessary for replication in normal cells but expendable in cancer cells. Further advantages, summarized in Table 1, include the OV ability to target multiple oncogenic pathways and use multiple means for cytotoxicity thereby avoiding cross-resistance encountered in standard anticancer therapies with minimal side effects due to tumour specific replication. Whilst they are not without their risks (including virus replication, insertional mutagenesis, loss of specificity, immunogenicity and adverse effects) [38, 39] a predicted compound annual growth rate of 24.9% between 2018-2025 [40] and multitude of companies and academic sectors either emerging or investing in this field [1] is testament to the exciting success of T-Vec as a treatment for melanoma [41-43] and more recently the clinical trial involving safety assessments of Parvovirus as a treatment for glioblastoma [44]. A number of studies have provided evidence for the natural oncolytic activity of viruses and have been extensively reviewed for

---

## Article Body Template

intratumoral administration elsewhere [45-48], but crucially, it is well understood that to be clinically relevant systemic delivery is optimum. Not only is it a relatively simple procedure but could also potentially facilitate treatment of disseminated disease and hard to reach tumours including those beyond the blood brain barrier. Whilst systemic administration of viruses has achieved efficacious results preclinically, including vesicular stomatitis virus [49-52], Newcastle disease virus [53, 54], reovirus [55], lentivirus [56] and herpes simplex virus [57, 58], they have translated poorly into humans [59, 60].

Despite the many advantages coupled with a spectrum of virus types, tropism and targeting pathways there are a number of obstacles to overcome to ensure clinical translation against both solid and disseminated tumours. These include the role of innate and adaptive immunity; the effect of viral tropism towards the liver resulting in liver toxicity and clearance and the physical barriers including tumour extracellular matrix and limited extravasation of OV's due to high interstitial fluid pressure within the tumour. The heterogeneity in the clinical response to OV therapies also suggests that a one-pronged approach to treatment may facilitate relapse and metastasis in heterogenous tumours, through the incomplete eradication of neoplastic cells [61]. Combination therapies have demonstrated synergistic antitumour effects [62] for example the facilitation of virion assembly and upregulation of CAR expression by paclitaxel (PTX) for Ad internalisation elicited by PTX-conjugated micelle-coated oncolytic adenovirus [63] whilst Tong et al., demonstrated that both intratumoral heterogeneity as well as extensive interpatient heterogeneity impacts the potential of Maraba virus as an oncolytic agent for ovarian cancer [64]. Oncolytic virotherapy therefore exists in a paradoxical situation whereby many of their attractive features also serve in their destruction e.g. immune responses designed to remove unrecognisable microorganisms yet also augment immune cell death (ICD) and cancer cell destruction.

Whilst genetic engineering of viruses has sought to overcome the problems associated with cell targeting and entry, this technique requires extensive modification of the capsid to incorporate new moieties, which is a laborious process and risks generating non-infectious or dysfunctional virus. In comparison, chemical engineering is simple and straightforward, since nanomaterials can be complexed with viruses through chemical conjugation or electrostatic interactions. It is this versatility that has attracted many investigators to utilise the advantages that NPs such as liposomes, polymers and cell carriers convey, yet despite the wealth of data described earlier regarding PK/PD of these NPs, there is a distinct pattern of strategies that have emerged.

---

## Article Body Template

### 4. Strategies employed by nanoparticles to enhance virotherapy delivery

As mentioned, there are three predominant stages for systemic NP delivery in oncology which has led to the three most important modifiable strategies for increasing concentrations of therapeutics at tumour targets; 1) shielding within the circulation; 2) tumour targeting; and 3) cell entry (Figure 4).

#### Shielding

Evading the humoral immune response is one of the biggest challenges facing systemic administration of OV's such that failure to do so will result in virus neutralisation before they have reached the target. The liver and spleen rapidly clear systemically administered viruses from the circulation by opsonisation with antibodies, complement and coagulation factors resulting in sequestration by the mononuclear phagocytic system (MPS). Yet viral replication, the release of pro-inflammatory cytokines and the accumulation of inflammatory cells to the tumour microenvironment (TME) are critical to tumour cell death [65, 66]. Consequently, an interplay between OV activity and an inflammatory response is believed to aid in the therapeutic efficacy of OVs in an *in vivo* environment. Some viruses have managed to overcome these challenges by disguising themselves within host cells for delivery to their target cell population including the human immune deficient virus (HIV) which utilises dendritic cells and macrophages that naturally migrate to the lymph nodes for delivery to CD4+ T cells [67]. Cell to cell spread of viruses also enables evasion of antibody neutralisation [68, 69]. Investigators are again taking inspiration from these evolved survival mechanisms to optimise cell carrier systems in a number of tumour models as a way of disguising OV from host defences prior to deposition at its target cell population (Figure 3). Avoiding virus sequestration by the MPS is achievable by shielding virus particles with chemically modified coat proteins. Probably one of the most studied uses of nanoparticles for virotherapy is the decoration of viruses with various polymers/micelles [63, 70-74], dendrimers [75, 76], liposomes [77-80], and cells [81, 82]. Alternatively, pre-administration of compounds designed to deplete serum factors (Cobra venom factor [83], cyclophosphamide [84, 85]), saturate scavenger receptors (polyinosinic acid) [86, 87] and diminish splenic macrophages using clodronate-loaded liposomes [88, 89] have been used to enhance therapeutic outcome by downregulating specific compartments of the immune system. Ultimately these strategies require extensive optimisation and understanding of the concomitant effects.

---

## Article Body Template

Shielding of the viruses via masking their surfaces with NPs requires some method of complexation of the two components. The zeta potential/surface charge is therefore probably one of the most important factors for carrier systems that do not involve encapsulation of viruses. To the best of our knowledge, complexation of viruses with NPs has been predominantly via spontaneous electrostatic interactions for the formation of safe complexes at a charge neutralised ratio [90-92]. However, in these examples, very little is known regarding their final shape, uniformity and stability with binding success often only assessed by changes in the net surface charge and particle size. There are very few examples where the coexistence of the various components have been verified by investigating chemical characteristics on surface composition and the bond types participating in the complexation [93-95]. One also needs to consider how strong these interactions are and whether they are displaced following injection into a turbulent fluidics system and displacement by the presence of other proteins such as serum albumin.

It is interesting that electrostatic interaction is the only method that has been utilised by investigators with examples for exploiting these interactions further by the addition of cationic coatings (eg TAT peptide), especially when covalent binding may provide a more stable bond and offers flexibility in terms of available residues and reversible linkers that can be activated under certain conditions and thereby act as triggers for more targeted delivery. One such reason may be evidence that bioconjugation in nanoparticle-based drug delivery systems alters their properties with glutaraldehyde protein cross-linking influencing the induced antibody responses at several levels *in vivo* [96] which would be an important consideration due to the inter-dependence of the immune response in the efficacy of virotherapy.

### Tumour targeting/recognition

If OV's manage to avoid detection in the systemic circulation their next challenge is to accumulate at their tumour target (Figure 4). This passive transport and non-specific accumulation of drugs in solid tumours has long been attributed to the enhanced permeability and retention (EPR) effect whereby a combination of their so called "leaky" vasculature for tissue entry together with a lack of well-defined lymphatic systems results in increased retention times [97]. Strategies for a more specific method of targeting tumour cells (often called active targeting), is to either recognise a determinant expressed directly on/by the tumour cell, by recognising the microenvironment created by the tumour (homing macrophages to

---

## Article Body Template

hypoxic areas [82], trafficking mesenchymal stem cells to areas of tumour-associated cytokine production [98, 99] or endothelial cells to tumour-associated angiogenesis [100]), or by exploiting intrinsic trafficking to specific tissues or organs (e.g. nonspecific T cells which naturally home to lymphoid organs) (Figure 3). One mechanistic explanation for intrinsic tumour tropism of viruses is linked to the inability of many cancer cells to respond properly to pro-inflammatory or antiviral cytokines [101]. For example, defects in the interferon (IFN) responses of cancer cells allows VSV to replicate, even in the presence of IFN where in normal primary mouse cells, the replication of VSV is strongly inhibited by IFN. One common method of genetically modifying an oncolytic virus candidate to increase tumour tropism is to delete or modify the viral genes responsible for countering cytokine-mediated immune responses such as members of the interleukin (IL) family, erythropoietin and interferons to name a few [102].

Functionalising the surfaces of NPs rather than modifying the viruses is also a common approach for re-targeting these drug delivery systems including the addition of ligands such as luteinizing hormone releasing hormone receptors [103] in breast cancer and anti-CD47 in pancreatic cancer cell targeting [104]. Redirection of virion-liposome complexes containing Moloney leukaemia virus (MMLV) to vascular endothelium by incorporation of antibodies for the endocytic receptors CD71 and CD62E/P [105] has been achieved and ovarian cancer growth has been ceased by systemic administration of a liposome-encapsulated adenovirus-encoding endostatin which decreased angiogenesis and increased tumour apoptosis [79]. Similarly, oncolytic adenovirus complexed with EGFR targeted dendrimer can be efficiently internalised by EGFR positive tumours [75], further validating the benefits of this strategy. Ultimately even these ligand-bearing NP-OV complexes rely on passive transport and whilst the EPR effect is widely held to increase nanotherapeutic delivery to tumours over normal organs owing to their defective tumour vasculature, the heterogeneity of the EPR effect in cancer [106] often offers less than a 2-fold increase in nanodrug delivery compared with critical normal organs, resulting in drug concentrations that are not sufficient for curing most cancers [107].

### Nano entry into cell targets

The final hurdle for OV's once they have reached their tumour target is the mechanisms for cell entry (Figure 4). Whilst the natural tropism towards tumour cells by certain viruses is selected by their

---

## Article Body Template

phenotype (immortality, DNA instability etc), cell entry relies on specific signalling pathways i.e. CAR expression is required for internalisation of adenoviruses [108] whilst glycoprotein D is an essential component of HSV-1 entry apparatus [109]. These signalling pathways can be enhanced for either increased recognition or enhanced viral replication. For example a genetically modified version of the measles virus (MV) vaccine strain which already demonstrated tropism towards many different types of cancer cells through the highly expressed CD46 receptors induces greater oncolytic activity against renal cancer cells [71]. Conversely, liposomal mediated entry of viruses are independent of viral receptors as they attach and reform with the cell membrane. This entry method has proven to enhance infectivity of liposomal HSV-1 even in the presence of neutralising antibodies following systemic administration for liver metastases [78]. Magnetofection is a membrane-receptor independent mechanism for hard-to-transfect biological models that uses a magnetic field to transfect cells [110], this technique has also demonstrated enhanced adenovirus uptake and increased time to full oncolytic effect *in vitro* and *in vivo* with consequential significant inhibition of tumour growth in a murine xenograft of human pancreatic carcinoma following intratumoural administration [90].

Whilst cell carriers may provide OV's with the ideal disguise for avoiding immunosurveillance, their successful delivery is complicated by a number of considerations including the kinetics of viral replication and release and kinetics of trafficking of the cell carrier from the site of injection to the tumour location. Cell carriers that support viral replication provide the added advantage of amplification overcoming issues including limits of clinical grade virus production and increase the therapeutic index and the likeliness of overcoming neutralising antibodies [82, 111]. Timing of replication and release is important as accumulation of viral proteins at the surface of the infected cell (e.g. VSV G protein) or creation of IHC complexes can alert the immune system resulting in premature clearance. Mitigation of this risk could involve either co-ordination of the lag time following initial viral infection of a cell and release of progeny virions with transport to target tissue or initiation of viral replication and release only at the targeted tumour. Shielding viruses within immune cells such as T cells avoids eliciting an immune response but they do not support all viruses and can be refractory to viral infection *in vivo* [112]. Mesenchymal stem cells can also support therapeutic adenoviral replication [113] yet more recently neural stem cells have demonstrated enhanced viral loading with significant increases in survival rates administered intracranially in an orthotopic glioma model [114] as well as significant reduction in omental tumour burden in an orthotopic model of ovarian cancer [115]. Whilst promising, both these examples utilised

---

## Article Body Template

direct administration of the treatment to the tumour site, not to mention the extraction of neural cells requiring invasive stereotactical surgery.

The toxic environments associated with tumours also influence cell entry and therapeutic effects for example hypoxia, which is associated with resistance to radiotherapy and chemotherapy despite enhanced delivery of these drugs via carriers. Liposomal formulations of SMI's doxorubicin [116], daunorubicin [117] and paclitaxel [118] have demonstrated success in the clinic although augmentation of efficacy above drug alone is debatable [119, 120]. This could be attributed to the characteristics of the carrier including their size (too large and they are captured by the reticuloendothelial system or too small and they are excreted in urine). In addition, reduced cellular uptake and cellular adaptations can also compromise the effectiveness of the chemotherapy [121] due to exposure to toxic conditions. Investigators have sought to adapt these situations to their advantage by using them as molecular switches for the release or activation of therapies e.g. Hypoxia activated pro-drugs (HAPs) such as TH-302 and PR-104, reviewed by Baran and Konopleva [122]. Fortunately, hypoxia seems to exert little or no effect on the replication of some oncolytic viruses [123]. Replication of adenoviruses from both groups B and C is inhibited, yet replication of the herpes simplex virus G207 strain was enhanced in brain and breast cancer cell lines by a 74% increase in cytotoxicity of hypoxic MCF 7 cells [124] and a 3.6-fold increase in G207 viral titres by hypoxic U87 glioblastoma cells [125]. This idea of activating switches has also been applied to MNP's for controlled drug release. Thermodegradation of polymer caps by application of an oscillating magnetic field to mesoporous silica nanoparticles has unblocked nanochannels containing drugs [126, 127], thereby releasing therapeutic cargo on demand in the desired location. Applying this strategy to OV could provide protection against immunosurveillance and allow targeted release thereby potentially increasing biocompatibility and efficacy.

Despite these strategies for enhanced virus protection and target recognition, perhaps the biggest limitation for any systemically administered therapy is its reliance on passive delivery with evidence demonstrating that nanoparticles (NPs) displaying decreased blood circulation time usually display concomitant reduced tumour uptake and efficacy [3]. It is therefore feasible that the similarities displayed by the current raft of available NPs (spherical, 80-120nm diameter, membranous) is a reflection of this (Figure 2). Thus would an active guidance/steering system allow for more radical designs of NP-OV

---

## Article Body Template

complexes by incorporating a more holistic approach encompassing the optimum features from all three strategies (Figure 4).

### Designer OV-nanocomplexes

Utilisation of viruses as a treatment modality in cancer have so far focused on viral tropism to specific tumour types for intratumoural administration or the addition of specific receptors for target recognition following systemic administration. Despite the multitude of drug delivery systems available they are all based on this principle of additional recognition moieties to overcome the limitations of their passive transport and discriminate between healthy and neoplastic cells. The literature therefore suggests that there is a requirement that the virus be “matched” to its target in order to achieve efficacy. However, these solutions lack a propelling force to a specific area as well as to penetrate tumours beyond diffusion limits. Systemic administration of naked virus as well as functionalised nanoparticles are inefficient as they still rely on the EPR bias (whereby the tumour vasculature is considered more “leaky” than that of normal organs) to navigate the vasculature. Perhaps it is therefore unsurprising that despite the wealth of knowledge regarding the pharmacokinetics of NPs within each phase of NP drug delivery there is a trend towards spherical particles of 80-130 nm diameter with lipid-based outer coatings (Figure 2). Would application of an external driving force allow radical divergence away from these parameters and translate to improved efficacy?

Application of external magnets have sought to overcome this problem of passive targeting. Magnetic chemotherapy has improved drug delivery for monotherapies such as doxorubicin [128], photothermal ablation with cetuximab [129] and melanin [130], dual targeting combining ligands with therapeutics [131], as well as utilising their function to overcome the toxic effects of hypoxia in HCT116 colorectal xenografts [132]. A similar methodology has been employed by investigators for regional delivery of cells including macrophages preloaded with magnetic iron particles [111], mesenchymal stem cells [133] and stem cells for the treatment of lesions [134]. Cisplatin loaded magnetic liposomes were able to overcome the drawback of low drug encapsulation efficiency by embedding magnetite nanoparticles in the liposomal membrane and the pharmacokinetics study in rats was able to sustain bioavailability in the body circulation compared to free drug [135]. Drug-containing liposomes covalently attached to magnetotactic bacteria were magnetically guided to HCT116 colorectal xenografts, with 55% of the bacteria localised to hypoxic regions [132]. Not to mention biocompatible magnetic nanoparticles (MNPs) to improve cancer

---

### Article Body Template

diagnostics have been demonstrable in magnetic resonance imaging [136], radionuclide therapy [137] and hyperthermia [138-140].

The ability of MNP's to be guided externally to the target tissue could therefore circumvent the reliance on specific viral tropism or specific ligands for efficacious virotherapy, evidence for which comes from a study using ectoenzyme ALPP decorated MNPs for selectively binding cancer cells without involving specific receptor interactions or antibodies [141]. A few examples of magnetically enhancing viruses are demonstrating this potential whereby adenovirus complexed with iron oxide increased transduction efficiency in CAR-negative MCF tumours, culminating in increased cancer cell killing and intracellular replication of Ad [90, 91]. Whilst demonstrating enhanced cellular interaction with adenoviruses, these studies were performed in nude mice with intratumoural administration of virotherapy at an acute timepoints.

One of the few studies to utilise magnetic guidance of systemically administered OV's boosted HSV oncolytic activity in xenograft model of prostate cancer [142]. By disguising the OV, Sephrivir, in macrophages loaded with iron oxide NPs, investigators were able to direct them to primary and metastatic tumours using pulsed field magnetic gradients resulting in increased tumour macrophage infiltration and reduction in tumour burden. One consideration when applying magnetic gradients is their effect on cells. A large gradient (up to 1 GT/m) can significantly change the membrane potential of the cell and thus have a significant impact on not only the properties and biological functionality of cells but also cell fate [143].

In terms of nanoparticle design, the use of magnetic field gradients could allow a departure from the traditional spherical, 100nm particles which are optimal for passive circulatory interactions and margination. For example, the increased circulation times demonstrated by rod-shaped particles [18] are controversial as nanocarriers due to their poor margination but application of external forces including magnets are expected to increase margination of rod-shaped magnetic particles [20] thereby enhancing their potential to reach tumour targets via increased circulation time, increased extravasation and magnetic steering. By overcoming some of the issues concerning circulatory transport, it may allow investigators to focus nanocarrier design on the characteristics for augmentation of the latter stages, namely tissue permeation and cell entry.

---

## Article Body Template

One of the limitations to investigate the interdependence of shape and size on the pharmacokinetics of magnetic nanoparticles (MNPs) has been their fabrication and production. The chemical synthesis of MNPs offers little control over uniform shape and size distribution and require coatings for biocompatibility and functionalisation. Conversely, biologically derived MNPs called magnetosomes, extracted from magnetotactic bacteria, display highly uniform size and shape. Medical applications in oncology have so far exploited the magnetic properties of magnetosomes for magnetic resonance imaging (MRI) contrast agents [144] and magnetic hyperthermia [138]. More recently, magnetosomes have established themselves as contenders for delivery of chemotherapies. Magnetosomes loaded with DOX demonstrated comparable tumour inhibition versus DOX alone against hepatocellular carcinoma (HCC) but significantly enhanced mortality rates by reducing cardiac toxicity to DOX alone [145]. By cross-linking chemotherapies to the surface of magnetosomes Deng et al and Liu et al have generated slow releasing formulations of cytosine arabinoside (Ara-C) for acute leukemia treatment [146, 147] and co-administration of DOX with siRNA using magnetosomes protected siRNA from degradation in serum resulting in synergistic cytotoxicity *in vitro* [148]. The ability to manipulate magnetosome transport via non-invasive, external application of magnetic fields has been confirmed by Tang et al whereby human papillomavirus type E7 and Ig-Fc fragment (pSLC-E7-Fc) combined with magnetosomes generated antigen-specific cytotoxic T lymphocyte activity with resultant tumour inhibition in a murine metastatic lung model (average pulmonary metastatic tumour weight of 343.6 mg vs 58.9mg) [149].

Magnetosomes have proven themselves equally successful as existing nanocarriers which may not be surprising when we compare their structures – cubooctohedral, 30-50nm diameter surrounded by phospholipid membranes extracted from MSR-1, MC-1 and AMB-1 strains. However, a multitude of other strains have been identified that synthesise rod, bullet and cuboidal magnetosomes that could provide further opportunities to investigate the effect of shape on targeted delivery of therapies. In fact, could magnetosomes provide the ideal opportunity to optimise a carrier system that is a) not reliant on passive delivery and the EPR effect but magnetic steering for targeting and margination; b) therefore shifts the focus to particle design for permeation and cell entry; c) removes the confounding influences of composition (chemical characteristics, charge) allowing direct comparison of shape alone.

---

## Article Body Template

### Conclusions

Systemically delivered oncolytic viruses exist in a paradoxical scenario whereby immunosurveillance can both enhance and destroy the therapeutic potential and whereby tumour angiogenesis both facilitates viral entry but is dependent on tumour type and status. Whilst nanoparticles have demonstrated improved efficacy their design primarily focuses on optimal conditions for circulatory survival due to a reliance on the EPR effect for passive delivery. The use of active guidance systems such as MRI together with magnetic nanoparticles may allow investigators to step off the merry-go-round and select NP-OV complex designs for cell entry alone. This could mediate a complete departure from traditional nanoparticle sizes and shapes and open the door for a pan-nanocarrier for oncology.

### Future prospects

The opposing design requirements within each of the three phases required for successful NP delivery indicates that there will eventually be a trade-off whereby investigators will be forced into decisions biasing one phase over another. To date this has involved intratumoural administration of OV's only; high titres for systemic administration to account for immunosurveillance; and the addition of specific targeting moieties. Unfortunately this has resulted in ineffective OV therapy for disseminated tumours; adverse effects; and precision virotherapy for single tumour types respectively. More ambitious OV-NP complexes are required (such as the design postulated in Figure 4) that encompass multiple strategies to overcome this challenge of systemic administration. Equally, the methodology required to both generate and assess NP-OV complex stability (including 3D morphology, bond strength and hydrodynamics) must be compatible within both biological and material sciences.

- **Future Perspective:** N/A
- **Executive Summary:** (bulleted summary points that illustrate the main conclusions made throughout the article. Less than 400 words).

---

## Article Body Template

### Introduction

- Oncolytic viruses (OV) are an attractive prospect due to their two-pronged attack mechanism: direct cell lysis and amplification of an anti-tumour immune response. Paradoxically this also results in their removal by the reticuloendothelial system when delivered systemically.
- There are three phases that OV's must navigate for successful nanoparticle delivery; circulatory longevity, tissue permeation and cellular interaction. Nanoparticles including liposomes, polymers and metallic particles are mitigating the circulatory interactions using shielding techniques and chemical modifications of their surfaces have incorporated moieties for the detection of specific tumour ligands for increased cell recognition. Despite these advances, the variable response rates evident in both primary and disseminated neoplasms suggests that efficacy relies on a multi-faceted approach.

### Pharmacokinetics of nanoparticles for oncology

- It is well known that nanoparticle shape, size and composition determines their fate during each of these phases and particular characteristics have been postulated for their success. Unfortunately these characteristics are not always conducive for each phase.
- The reliance on passive delivery to the tumour target via the EPR effect has biased the current raft of nanocarriers to be optimal for navigating circulatory complications such as avoiding immune/serum protein interactions and extravasation (spherical, ~100nm diameter, neutral charge). Therefore their limited success could be attributed to suboptimal characteristics required for the latter stages of nanoparticle delivery.

### Strategies employed by nanoparticles to enhance virotherapy delivery

- The ability to actively target tumours by external guidance systems (eg. magnetic gradients) may shift the focus to characteristics required for tissue permeation and retention by circumventing the need to rely on the EPR effect. Similarly the use of environmental stimuli (eg. hypoxia) as triggers for the unveiling of specific characteristics via polymeric switches allows nanoparticles to straddle the needs of the various phases.
- The effects of shape, size, charge etc pose another problem when trying to draw comparisons due to fabrication limitations and inherent compositional differences in the various nanocarriers available.
- Ultimately, successful systemic delivery of OV's relies on how we reconcile the different needs of each phase on the merry-go-round that is nanoparticle delivery.

### OR

**Summary Points (Research articles & Company profiles only):** 8–10 bullet point sentences highlighting the key points of the article.

---

## Article Body Template

- **Figure legends**

**Figure 1. The nanocarrier design merry-go-round.** Optimal nanocarrier characteristics for each phase of systemic delivery of cancer therapies. AR = aspect ratio.

**Figure 2.** Currently used nanocarriers demonstrating significant similarities; spherical morphology, synthetic or biological phospholipid coating and overlapping diameters.

**Figure 3. Factors controlling pharmacokinetics and biodistribution of nanoparticles (NP).** **Size:** Large NPs (1) will not be able to enter the tumour through leaky vasculature and demonstrate increased blood protein deposition as a result of increased surface area resulting in rapid clearance. Small NPs show high permeation rate (2) but also are rapidly cleared from the tumour by RES (3) leading to poor accumulation. Shape of NPs (4) affects tumour cell internalisation and determines interaction with RES, PK and tumour retention as a result of surface curvature and altered hydrodynamic behaviour. Aggregates can cause different organ distribution. **Surface characteristics:** Polymer coated NPs may shield NPs from neutralising Abs and RES but repeated administration can induce anti-PEG antibodies (5). Negatively charged NPs exhibit strong RES uptake whilst cationic NPs induce serum protein aggregation exhibit increased uptake by cells (6). Neutral NPs can travel up to three times more distance than charged analogues (7), distribute more evenly and exhibit least RES interaction for the longest circulation. PEGylation can shield charge effects (8) preventing opsonisation and can be shed after tumour extravasation to expose cationic particles that can interact with target cells (9). **Regime:** Multiple treatments could either increase generation of neutralising antibodies (10) to NPs resulting in reduced efficacy or suppress RES activity during the first dose but reduce clearance and increase toxicity of subsequent doses (11). Distribution of NPs is dependent also on exposure route (12).

**Figure 4.** Current methods for virotherapeutic targeting of tumours and how to manipulate these further to create the ideal NP-OV complex.

- **Table Legends**

**Table 1:** Multiple oncolytic pathways of oncolytic viruses

**Table 1: Multiple oncolytic pathways of oncolytic viruses**

Advantage	Description	Virus	Ref
-----------	-------------	-------	-----

---

**Article Body Template**

Direct Oncolysis	Consequence of viral replication and virion release.	HSV, VV, VSV	[150]
	Cytotoxic proteins synthesised by the virus e.g. induction of apoptosis by adenovirus E4-ORF3 protein expression.	Ad	[151]
Reprogramming the tumour microenvironment	Release of DAMPs and TAAs into bloodstream thereby switching an immunogenically “cold” tumour (low abundance of tumour antigens, tumour infiltrating lymphocytes and an immunosuppressive tumour environment) to a ‘hot’ immunogenic tumour.	HSV	[152, 153]
	Activation of innate and adaptive anti-cancer immune responses, thereby inducing immunogenic cell death (ICD) in cancer cells.	Ad, MV	[154-158]
	Mediate the killing of uninfected cancer cells by destruction of tumour blood vessels.	VSV, VV	[159-161]
Enhance anti-tumour effects	Amplification of therapy overtime due to virion release and immune interaction in comparison to classical drug pharmacokinetics which decrease with time.	Ad	[162]
Inhibit relapse and metastasis	Incorporation of transgenes (eg. GM-CSF, IFN $\alpha/\beta$ , IL-12, CCL5).	HSV, Ad, VSV, VV	[163]

DAMPs = damage-associated molecular patterns, TAAs = tumour associated antigens, VSV = vesicular stomatitis virus, VV = vaccinia virus, Ad = adenovirus, MV = measles virus

**References:**

1. Duffy MR, Fisher KD, Seymour LW. Making Oncolytic Virotherapy a Clinical Reality: The European Contribution. *Hum Gene Ther* 28(11), 1033-1046 (2017).
2. Iwamoto T. Clinical application of drug delivery systems in cancer chemotherapy: review of the efficacy and side effects of approved drugs. *Biol Pharm Bull* 36(5), 715-718 (2013).
3. Ernsting MJ, Murakami M, Roy A, Li SD. Factors controlling the pharmacokinetics, biodistribution and intratumoral penetration of nanoparticles. *J Control Release* 172(3), 782-794 (2013).
4. Setyawati MI, Tay CY, Docter D, Stauber RH, Leong DT. Understanding and exploiting nanoparticles' intimacy with the blood vessel and blood. *Chem Soc Rev* 44(22), 8174-8199 (2015).
5. Caldorera-Moore M, Guimard N, Shi L, Roy K. Designer nanoparticles: incorporating size, shape and triggered release into nanoscale drug carriers. *Expert Opin Drug Deliv* 7(4), 479-495 (2010).
6. Fang C, Shi B, Pei YY, Hong MH, Wu J, Chen HZ. In vivo tumor targeting of tumor necrosis factor- $\alpha$ -loaded stealth nanoparticles: effect of MePEG molecular weight and particle size. *Eur J Pharm Sci* 27(1), 27-36 (2006).
7. Decuzzi P, Lee S, Bhushan B, Ferrari M. A theoretical model for the margination of particles within blood vessels. *Ann Biomed Eng* 33(2), 179-190 (2005).

---

**Article Body Template**

8. Duan D, Liu H, Xu M *et al.* Size-Controlled Synthesis of Drug-Loaded Zeolitic Imidazolate Framework in Aqueous Solution and Size Effect on Their Cancer Theranostics in Vivo. *ACS Appl Mater Interfaces* doi:10.1021/acsami.8b17660 (2018).
9. Mohr K, Sommer M, Baier G *et al.* Aggregation behaviour of polystyrene-nanoparticles in human blood serum and its impact on the in vivo distribution in mice. *Journal of Nanomedicine & Nanotechnology* 5(2), (2014).
10. Durantie E, Vanhecke D, Rodriguez-Lorenzo L *et al.* Biodistribution of single and aggregated gold nanoparticles exposed to the human lung epithelial tissue barrier at the air-liquid interface. *Part Fibre Toxicol* 14(1), 49 (2017).
11. Tang L, Yang X, Yin Q *et al.* Investigating the optimal size of anticancer nanomedicine. *Proc Natl Acad Sci U S A* 111(43), 15344-15349 (2014).
12. Kang JH, Cho J, Ko YT. Investigation on the effect of nanoparticle size on the blood-brain tumour barrier permeability by in situ perfusion via internal carotid artery in mice. *J Drug Target* 27(1), 103-110 (2019).
13. Kettiger H, Schipanski A, Wick P, Huwyler J. Engineered nanomaterial uptake and tissue distribution: from cell to organism. *Int J Nanomedicine* 8 3255-3269 (2013).
14. Shang L, Nienhaus K, Nienhaus GU. Engineered nanoparticles interacting with cells: size matters. *J Nanobiotechnology* 12 5 (2014).
15. Sykes EA, Dai Q, Sarsons CD *et al.* Tailoring nanoparticle designs to target cancer based on tumor pathophysiology. *Proc Natl Acad Sci U S A* 113(9), E1142-1151 (2016).
16. Arvizo RR, Rana S, Miranda OR, Bhattacharya R, Rotello VM, Mukherjee P. Mechanism of anti-angiogenic property of gold nanoparticles: role of nanoparticle size and surface charge. *Nanomedicine* 7(5), 580-587 (2011).
17. Truong NP, Whittaker MR, Mak CW, Davis TP. The importance of nanoparticle shape in cancer drug delivery. *Expert Opin Drug Deliv* 12(1), 129-142 (2015).
18. Bruckman MA, Randolph LN, Vanmeter A *et al.* Biodistribution, pharmacokinetics, and blood compatibility of native and PEGylated tobacco mosaic virus nano-rods and -spheres in mice. *Virology* 449 163-173 (2014).
19. Chariou PL, Lee KL, Pokorski JK, Saidel GM, Steinmetz NF. Diffusion and Uptake of Tobacco Mosaic Virus as Therapeutic Carrier in Tumor Tissue: Effect of Nanoparticle Aspect Ratio. *J Phys Chem B* 120(26), 6120-6129 (2016).
20. Lee SY, Ferrari M, Decuzzi P. Shaping nano-/micro-particles for enhanced vascular interaction in laminar flows. *Nanotechnology* 20(49), 495101 (2009).
21. Gratton SE, Ropp PA, Pohlhaus PD *et al.* The effect of particle design on cellular internalization pathways. *Proc Natl Acad Sci U S A* 105(33), 11613-11618 (2008).
22. Favi PM, Valencia MM, Elliott PR *et al.* Shape and surface chemistry effects on the cytotoxicity and cellular uptake of metallic nanorods and nanospheres. *J Biomed Mater Res A* 103(12), 3940-3955 (2015).
23. Favi PM, Gao M, Johana Sepulveda Arango L *et al.* Shape and surface effects on the cytotoxicity of nanoparticles: Gold nanospheres versus gold nanostars. *J Biomed Mater Res A* 103(11), 3449-3462 (2015).
24. Armstrong JK, Hempel G, Koling S *et al.* Antibody against poly(ethylene glycol) adversely affects PEG-asparaginase therapy in acute lymphoblastic leukemia patients. *Cancer* 110(1), 103-111 (2007).

---

**Article Body Template**

25. Garay RP, El-Gewely R, Armstrong JK, Garratty G, Richette P. Antibodies against polyethylene glycol in healthy subjects and in patients treated with PEG-conjugated agents. *Expert Opin Drug Deliv* 9(11), 1319-1323 (2012).
26. Hsieh YC, Wang HE, Lin WW *et al.* Pre-existing anti-polyethylene glycol antibody reduces the therapeutic efficacy and pharmacokinetics of PEGylated liposomes. *Theranostics* 8(11), 3164-3175 (2018).
27. Fix SM, Nyankima AG, Mcsweeney MD, Tsuruta JK, Lai SK, Dayton PA. Accelerated Clearance of Ultrasound Contrast Agents Containing Polyethylene Glycol is Associated with the Generation of Anti-Polyethylene Glycol Antibodies. *Ultrasound Med Biol* 44(6), 1266-1280 (2018).
28. Zhang P, Sun F, Liu S, Jiang S. Anti-PEG antibodies in the clinic: Current issues and beyond PEGylation. *J Control Release* 244(Pt B), 184-193 (2016).
29. Fu S, Liang M, Wang Y *et al.* Dual-Modified Novel Biomimetic Nanocarriers Improve Targeting and Therapeutic Efficacy in Glioma. *ACS Appl Mater Interfaces* doi:10.1021/acsami.8b18664 (2018).
30. Hu CM, Zhang L, Aryal S, Cheung C, Fang RH, Zhang L. Erythrocyte membrane-camouflaged polymeric nanoparticles as a biomimetic delivery platform. *Proc Natl Acad Sci U S A* 108(27), 10980-10985 (2011).
31. Honary S, Zahir F. Effect of zeta potential on the properties of nano-drug delivery systems - A review (Part 2). *Tropical Journal of Pharmaceutical Research* 12(2), 265-273 (2013).
32. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 144(5), 646-674 (2011).
33. Russell SJ. RNA viruses as virotherapy agents. *Cancer Gene Ther* 9(12), 961-966 (2002).
34. Haddad D. Genetically Engineered Vaccinia Viruses As Agents for Cancer Treatment, Imaging, and Transgene Delivery. *Front Oncol* 7 96 (2017).
35. Sokolowski NA, Rizos H, Diefenbach RJ. Oncolytic virotherapy using herpes simplex virus: how far have we come? *Oncolytic Virother* 4 207-219 (2015).
36. Alemany R. Oncolytic Adenoviruses in Cancer Treatment. *Biomedicines* 2(1), 36-49 (2014).
37. Brown MC, Dobrikova EY, Dobrikov MI *et al.* Oncolytic polio virotherapy of cancer. *Cancer* 120(21), 3277-3286 (2014).
38. Assessment of adenoviral vector safety and toxicity: report of the National Institutes of Health Recombinant DNA Advisory Committee. *Hum Gene Ther* 13(1), 3-13 (2002).
39. Matsuda T, Karube H, Aruga A. A Comparative Safety Profile Assessment of Oncolytic Virus Therapy Based on Clinical Trials. *Ther Innov Regul Sci* 52(4), 430-437 (2018).
40. Global Oncolytic Virus Therapy Market Size, Status and Forecast 2018-2025. (2019).
41. Johnson DB, Puzanov I, Kelley MC. Talimogene laherparepvec (T-VEC) for the treatment of advanced melanoma. *Immunotherapy* 7(6), 611-619 (2015).
42. Killock D. Skin cancer: T-VEC oncolytic viral therapy shows promise in melanoma. *Nat Rev Clin Oncol* 12(8), 438 (2015).
43. Kohlhapp FJ, Zloza A, Kaufman HL. Talimogene laherparepvec (T-VEC) as cancer immunotherapy. *Drugs Today (Barc)* 51(9), 549-558 (2015).
44. Geletneky K, Hajda J, Angelova AL *et al.* Oncolytic H-1 Parvovirus Shows Safety and Signs of Immunogenic Activity in a First Phase I/IIa Glioblastoma Trial. *Mol Ther* 25(12), 2620-2634 (2017).
45. Donnelly OG, Errington-Mais F, Prestwich R *et al.* Recent clinical experience with oncolytic viruses. *Curr Pharm Biotechnol* 13(9), 1834-1841 (2012).

---

**Article Body Template**

46. Lawler SE, Speranza MC, Cho CF, Chiocca EA. Oncolytic Viruses in Cancer Treatment: A Review. *JAMA Oncol* 3(6), 841-849 (2017).
47. Motalleb G. Virotherapy in cancer. *Iran J Cancer Prev* 6(2), 101-107 (2013).
48. Maroun J, Munoz-Alia M, Ammayappan A, Schulze A, Peng KW, Russell S. Designing and building oncolytic viruses. *Future Virol* 12(4), 193-213 (2017).
49. Ebert O, Harbaran S, Shinozaki K, Woo SL. Systemic therapy of experimental breast cancer metastases by mutant vesicular stomatitis virus in immune-competent mice. *Cancer Gene Ther* 12(4), 350-358 (2005).
50. Naik S, Nace R, Barber GN, Russell SJ. Potent systemic therapy of multiple myeloma utilizing oncolytic vesicular stomatitis virus coding for interferon-beta. *Cancer Gene Ther* 19(7), 443-450 (2012).
51. Ozduman K, Wollmann G, Piepmeier JM, Van Den Pol AN. Systemic vesicular stomatitis virus selectively destroys multifocal glioma and metastatic carcinoma in brain. *J Neurosci* 28(8), 1882-1893 (2008).
52. Rommelfanger DM, Wongthida P, Diaz RM *et al.* Systemic combination virotherapy for melanoma with tumor antigen-expressing vesicular stomatitis virus and adoptive T-cell transfer. *Cancer Res* 72(18), 4753-4764 (2012).
53. Phuangsab A, Lorence RM, Reichard KW, Peeples ME, Walter RJ. Newcastle disease virus therapy of human tumor xenografts: antitumor effects of local or systemic administration. *Cancer Lett* 172(1), 27-36 (2001).
54. Schirmacher V, Griesbach A, Ahlert T. Antitumor effects of Newcastle Disease Virus in vivo: local versus systemic effects. *Int J Oncol* 18(5), 945-952 (2001).
55. Kottke T, Thompson J, Diaz RM *et al.* Improved systemic delivery of oncolytic reovirus to established tumors using preconditioning with cyclophosphamide-mediated Treg modulation and interleukin-2. *Clin Cancer Res* 15(2), 561-569 (2009).
56. Ma JL, Han SX, Zhao J *et al.* Systemic delivery of lentivirus-mediated secretable TAT-apoptin eradicates hepatocellular carcinoma xenografts in nude mice. *Int J Oncol* 41(3), 1013-1020 (2012).
57. Hodish I, Tal R, Shaish A *et al.* Systemic administration of radiation-potentiated anti-angiogenic gene therapy against primary and metastatic cancer based on transcriptionally controlled HSV-TK. *Cancer Biol Ther* 8(5), 424-432 (2009).
58. Leoni V, Gatta V, Palladini A *et al.* Systemic delivery of HER2-retargeted oncolytic-HSV by mesenchymal stromal cells protects from lung and brain metastases. *Oncotarget* 6(33), 34774-34787 (2015).
59. Auffinger B, Ahmed AU, Lesniak MS. Oncolytic virotherapy for malignant glioma: translating laboratory insights into clinical practice. *Front Oncol* 3 32 (2013).
60. Kaur B, Chiocca EA, Cripe TP. Oncolytic HSV-1 virotherapy: clinical experience and opportunities for progress. *Curr Pharm Biotechnol* 13(9), 1842-1851 (2012).
61. Zeng W, Hu P, Wu J *et al.* The oncolytic herpes simplex virus vector G47 effectively targets breast cancer stem cells. *Oncol Rep* 29(3), 1108-1114 (2013).
62. Simpson GR, Relph K, Harrington K, Melcher A, Pandha H. Cancer immunotherapy via combining oncolytic virotherapy with chemotherapy: recent advances. *Oncolytic Virother* 5 1-13 (2016).
63. Kasala D, Lee SH, Hong JW *et al.* Synergistic antitumor effect mediated by a paclitaxel-conjugated polymeric micelle-coated oncolytic adenovirus. *Biomaterials* 145 207-222 (2017).

---

**Article Body Template**

64. Tong JG, Valdes YR, Sivapragasam M *et al.* Spatial and temporal epithelial ovarian cancer cell heterogeneity impacts Maraba virus oncolytic potential. *BMC Cancer* 17(1), 594 (2017).
65. Filley AC, Dey M. Immune System, Friend or Foe of Oncolytic Virotherapy? *Front Oncol* 7 106 (2017).
66. Gujar S, Pol JG, Kim Y, Lee PW, Kroemer G. Antitumor Benefits of Antiviral Immunity: An Underappreciated Aspect of Oncolytic Virotherapies. *Trends Immunol* 39(3), 209-221 (2018).
67. Dayton AI. Hitting HIV where it hides. *Retrovirology* 5 15 (2008).
68. Di Lorenzo C, Angus AG, Patel AH. Hepatitis C virus evasion mechanisms from neutralizing antibodies. *Viruses* 3(11), 2280-2300 (2011).
69. Schiffner T, Sattentau QJ, Duncan CJ. Cell-to-cell spread of HIV-1 and evasion of neutralizing antibodies. *Vaccine* 31(49), 5789-5797 (2013).
70. Myers R, Coviello C, Erbs P *et al.* Polymeric Cups for Cavitation-mediated Delivery of Oncolytic Vaccinia Virus. *Mol Ther* 24(9), 1627-1633 (2016).
71. Nosaki K, Hamada K, Takashima Y *et al.* A novel, polymer-coated oncolytic measles virus overcomes immune suppression and induces robust antitumor activity. *Mol Ther Oncolytics* 3 16022 (2016).
72. Oh E, Oh JE, Hong J *et al.* Optimized biodegradable polymeric reservoir-mediated local and sustained co-delivery of dendritic cells and oncolytic adenovirus co-expressing IL-12 and GM-CSF for cancer immunotherapy. *J Control Release* 259 115-127 (2017).
73. Price DL, Li P, Chen CH *et al.* Silk-elastin-like protein polymer matrix for intraoperative delivery of an oncolytic vaccinia virus. *Head Neck* 38(2), 237-246 (2016).
74. Na Y, Choi JW, Kasala D *et al.* Potent antitumor effect of neurotensin receptor-targeted oncolytic adenovirus co-expressing decorin and Wnt antagonist in an orthotopic pancreatic tumor model. *J Control Release* 220(Pt B), 766-782 (2015).
75. Yoon AR, Kasala D, Li Y *et al.* Antitumor effect and safety profile of systemically delivered oncolytic adenovirus complexed with EGFR-targeted PAMAM-based dendrimer in orthotopic lung tumor model. *J Control Release* 231 2-16 (2016).
76. Grunwald GK, Vetter A, Klutz K *et al.* Systemic image-guided liver cancer radiovirotherapy using dendrimer-coated adenovirus encoding the sodium iodide symporter as theranostic gene. *J Nucl Med* 54(8), 1450-1457 (2013).
77. Sakurai F, Inoue S, Kaminade T *et al.* Cationic liposome-mediated delivery of reovirus enhances the tumor cell-killing efficiencies of reovirus in reovirus-resistant tumor cells. *Int J Pharm* 524(1-2), 238-247 (2017).
78. Shikano T, Kasuya H, Sahin TT *et al.* High therapeutic potential for systemic delivery of a liposome-conjugated herpes simplex virus. *Curr Cancer Drug Targets* 11(1), 111-122 (2011).
79. Yang L, Wang L, Su XQ *et al.* Suppression of ovarian cancer growth via systemic administration with liposome-encapsulated adenovirus-encoding endostatin. *Cancer Gene Ther* 17(1), 49-57 (2010).
80. Yoshida T, Mizuno M, Taniguchi K, Nakayashiki N, Wakabayashi T, Yoshida J. Rat glioma cell death induced by cationic liposome-mediated transfer of the herpes simplex virus thymidine kinase gene followed by ganciclovir treatment. *J Surg Oncol* 76(1), 19-25 (2001).
81. Balvers RK, Belcaid Z, Van Den Hengel SK *et al.* Locally-delivered T-cell-derived cellular vehicles efficiently track and deliver adenovirus delta24-RGD to infiltrating glioma. *Viruses* 6(8), 3080-3096 (2014).

---

**Article Body Template**

82. Muthana M, Giannoudis A, Scott SD *et al.* Use of macrophages to target therapeutic adenovirus to human prostate tumors. *Cancer Res* 71(5), 1805-1815 (2011).
83. Evgin L, Acuna SA, Tanese De Souza C *et al.* Complement inhibition prevents oncolytic vaccinia virus neutralization in immune humans and cynomolgus macaques. *Mol Ther* 23(6), 1066-1076 (2015).
84. Fulci G, Breyman L, Gianni D *et al.* Cyclophosphamide enhances glioma virotherapy by inhibiting innate immune responses. *Proc Natl Acad Sci U S A* 103(34), 12873-12878 (2006).
85. Lun XQ, Jang JH, Tang N *et al.* Efficacy of systemically administered oncolytic vaccinia virotherapy for malignant gliomas is enhanced by combination therapy with rapamycin or cyclophosphamide. *Clin Cancer Res* 15(8), 2777-2788 (2009).
86. Haisma HJ, Kamps JA, Kamps GK, Plantinga JA, Rots MG, Bellu AR. Polyinosinic acid enhances delivery of adenovirus vectors in vivo by preventing sequestration in liver macrophages. *J Gen Virol* 89(Pt 5), 1097-1105 (2008).
87. Liu YP, Tong C, Dispenzieri A, Federspiel MJ, Russell SJ, Peng KW. Polyinosinic acid decreases sequestration and improves systemic therapy of measles virus. *Cancer Gene Ther* 19(3), 202-211 (2012).
88. Fulci G, Dmitrieva N, Gianni D *et al.* Depletion of peripheral macrophages and brain microglia increases brain tumor titers of oncolytic viruses. *Cancer Res* 67(19), 9398-9406 (2007).
89. Denton NL, Chen CY, Hutzen B *et al.* Myelolytic Treatments Enhance Oncolytic Herpes Virotherapy in Models of Ewing Sarcoma by Modulating the Immune Microenvironment. *Mol Ther Oncolytics* 11 62-74 (2018).
90. Tresilwised N, Pithayanukul P, Mykhaylyk O *et al.* Boosting oncolytic adenovirus potency with magnetic nanoparticles and magnetic force. *Mol Pharm* 7(4), 1069-1089 (2010).
91. Choi JW, Park JW, Na Y *et al.* Using a magnetic field to redirect an oncolytic adenovirus complexed with iron oxide augments gene therapy efficacy. *Biomaterials* 65 163-174 (2015).
92. Almstatter I, Mykhaylyk O, Settles M *et al.* Characterization of magnetic viral complexes for targeted delivery in oncology. *Theranostics* 5(7), 667-685 (2015).
93. Wojcik M, Lewandowski W, Krol M *et al.* Enhancing anti-tumor efficacy of Doxorubicin by non-covalent conjugation to gold nanoparticles - in vitro studies on feline fibrosarcoma cell lines. *PLoS One* 10(4), e0124955 (2015).
94. Lim EK, Sajomsang W, Choi Y *et al.* Chitosan-based intelligent theragnosis nanocomposites enable pH-sensitive drug release with MR-guided imaging for cancer therapy. *Nanoscale Res Lett* 8(1), 467 (2013).
95. Korin E, Froumin N, Cohen S. Surface analysis of nanocomplexes by X-ray photoelectron spectroscopy (XPS). *ACS Biomaterials Science & Engineering* 3 882-889 (2017).
96. Soldemo M, Adori M, Stark JM *et al.* Glutaraldehyde Cross-linking of HIV-1 Env Trimers Skews the Antibody Subclass Response in Mice. *Front Immunol* 8 1654 (2017).
97. Fang J, Nakamura H, Maeda H. The EPR effect: Unique features of tumor blood vessels for drug delivery, factors involved, and limitations and augmentation of the effect. *Adv Drug Deliv Rev* 63(3), 136-151 (2011).
98. Hoyos V, Del Bufalo F, Yagyu S *et al.* Mesenchymal Stromal Cells for Linked Delivery of Oncolytic and Apoptotic Adenoviruses to Non-small-cell Lung Cancers. *Mol Ther* 23(9), 1497-1506 (2015).
99. Parker Kerrigan BC, Shimizu Y, Andreeff M, Lang FF. Mesenchymal stromal cells for the delivery of oncolytic viruses in gliomas. *Cytotherapy* 19(4), 445-457 (2017).

---

**Article Body Template**

- 1  
2  
3  
4  
5  
6 100. Uldry E, Faes S, Demartines N, Dormond O. Fine-Tuning Tumor Endothelial Cells to Selectively  
7 Kill Cancer. *Int J Mol Sci* 18(7), (2017).
- 8 101. Mcfadden G, Mohamed MR, Rahman MM, Barte E. Cytokine determinants of viral tropism. *Nat*  
9 *Rev Immunol* 9(9), 645-655 (2009).
- 10 102. Jhavar SR, Thandoni A, Bommareddy PK *et al.* Oncolytic Viruses-Natural and Genetically  
11 Engineered Cancer Immunotherapies. *Front Oncol* 7 202 (2017).
- 12 103. Zhou J, Leuschner C, Kumar C, Hormes JF, Soboyejo WO. Sub-cellular accumulation of magnetic  
13 nanoparticles in breast tumors and metastases. *Biomaterials* 27(9), 2001-2008 (2006).
- 14 104. Trabulo S, Aires A, Aicher A, Heeschen C, Cortajarena AL. Multifunctionalized iron oxide  
15 nanoparticles for selective targeting of pancreatic cancer cells. *Biochim Biophys Acta* 1861(6),  
16 1597-1605 (2017).
- 17 105. Tan PH, Xue SA, Wei B, Holler A, Voss RH, George AJ. Changing viral tropism using  
18 immunoliposomes alters the stability of gene expression: implications for viral vector design.  
19 *Mol Med* 13(3-4), 216-226 (2007).
- 20 106. Maeda H. Vascular permeability in cancer and infection as related to macromolecular drug  
21 delivery, with emphasis on the EPR effect for tumor-selective drug targeting. *Proc Jpn Acad Ser B*  
22 *Phys Biol Sci* 88(3), 53-71 (2012).
- 23 107. Nakamura Y, Mochida A, Choyke PL, Kobayashi H. Nanodrug Delivery: Is the Enhanced  
24 Permeability and Retention Effect Sufficient for Curing Cancer? *Bioconjug Chem* 27(10), 2225-  
25 2238 (2016).
- 26 108. Mcdonald D, Stockwin L, Matzow T, Blair Zajdel ME, Blair GE. Coxsackie and adenovirus receptor  
27 (CAR)-dependent and major histocompatibility complex (MHC) class I-independent uptake of  
28 recombinant adenoviruses into human tumour cells. *Gene Ther* 6(9), 1512-1519 (1999).
- 29 109. Campadelli-Fiume G, Petrovic B, Leoni V *et al.* Retargeting Strategies for Oncolytic Herpes  
30 Simplex Viruses. *Viruses* 8(3), 63 (2016).
- 31 110. Pereyra AS, Mykhaylyk O, Lockhart EF *et al.* Magnetofection Enhances Adenoviral Vector-based  
32 Gene Delivery in Skeletal Muscle Cells. *J Nanomed Nanotechnol* 7(2), (2016).
- 33 111. Muthana M, Scott SD, Farrow N *et al.* A novel magnetic approach to enhance the efficacy of cell-  
34 based gene therapies. *Gene Ther* 15(12), 902-910 (2008).
- 35 112. Kim J, Hall RR, Lesniak MS, Ahmed AU. Stem Cell-Based Cell Carrier for Targeted Oncolytic  
36 Virotherapy: Translational Opportunity and Open Questions. *Viruses* 7(12), 6200-6217 (2015).
- 37 113. Komarova S, Kawakami Y, Stoff-Khalili MA, Curiel DT, Pereboeva L. Mesenchymal progenitor  
38 cells as cellular vehicles for delivery of oncolytic adenoviruses. *Mol Cancer Ther* 5(3), 755-766  
39 (2006).
- 40 114. Ahmed AU, Tyler MA, Thaci B *et al.* A comparative study of neural and mesenchymal stem cell-  
41 based carriers for oncolytic adenovirus in a model of malignant glioma. *Mol Pharm* 8(5), 1559-  
42 1572 (2011).
- 43 115. Mooney R, Majid AA, Batalla-Covello J *et al.* Enhanced Delivery of Oncolytic Adenovirus by  
44 Neural Stem Cells for Treatment of Metastatic Ovarian Cancer. *Mol Ther Oncolytics* 12 79-92  
45 (2019).
- 46 116. Batist G, Ramakrishnan G, Rao CS *et al.* Reduced cardiotoxicity and preserved antitumor efficacy  
47 of liposome-encapsulated doxorubicin and cyclophosphamide compared with conventional  
48 doxorubicin and cyclophosphamide in a randomized, multicenter trial of metastatic breast  
49 cancer. *J Clin Oncol* 19(5), 1444-1454 (2001).
- 50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

---

**Article Body Template**

- 1  
2  
3  
4  
5  
6 117. Gill PS, Wernz J, Scadden DT *et al.* Randomized phase III trial of liposomal daunorubicin versus  
7 doxorubicin, bleomycin, and vincristine in AIDS-related Kaposi's sarcoma. *J Clin Oncol* 14(8),  
8 2353-2364 (1996).
- 9 118. Koudelka S, Turanek J. Liposomal paclitaxel formulations. *J Control Release* 163(3), 322-334  
10 (2012).
- 11 119. Frenkel V, Etherington A, Greene M *et al.* Delivery of liposomal doxorubicin (Doxil) in a breast  
12 cancer tumor model: investigation of potential enhancement by pulsed-high intensity focused  
13 ultrasound exposure. *Acad Radiol* 13(4), 469-479 (2006).
- 14 120. Williams G, Cortazar P, Pazdur R. Developing drugs to decrease the toxicity of chemotherapy. *J*  
15 *Clin Oncol* 19(14), 3439-3441 (2001).
- 16 121. Cosse JP, Michiels C. Tumour hypoxia affects the responsiveness of cancer cells to  
17 chemotherapy and promotes cancer progression. *Anticancer Agents Med Chem* 8(7), 790-797  
18 (2008).
- 19 122. Baran N, Konopleva M. Molecular Pathways: Hypoxia-Activated Prodrugs in Cancer Therapy. *Clin*  
20 *Cancer Res* 23(10), 2382-2390 (2017).
- 21 123. Hay JG. The potential impact of hypoxia on the success of oncolytic virotherapy. *Curr Opin Mol*  
22 *Ther* 7(4), 353-358 (2005).
- 23 124. Pin RH, Reinblatt M, Fong Y. Employing tumor hypoxia to enhance oncolytic viral therapy in  
24 breast cancer. *Surgery* 136(2), 199-204 (2004).
- 25 125. Aghi MK, Liu TC, Rabkin S, Martuza RL. Hypoxia enhances the replication of oncolytic herpes  
26 simplex virus. *Mol Ther* 17(1), 51-56 (2009).
- 27 126. Saint-Cricq P, Deshayes S, Zink JI, Kasko AM. Magnetic field activated drug delivery using  
28 thermodegradable azo-functionalised PEG-coated core-shell mesoporous silica nanoparticles.  
29 *Nanoscale* 7(31), 13168-13172 (2015).
- 30 127. Chen P-J, Hu S-H, Hsiao C-S, Chen Y-Y, Liu D-M, Chen S-Y. Multifunctional magnetically  
31 removable nanogated lids of Fe<sub>3</sub>O<sub>4</sub>-capped mesoporous silica nanoparticles for intracellular  
32 controlled release and MR imaging. *Journal of Materials Chemistry* 21(8), 2535-2543 (2011).
- 33 128. Lu YJ, Lin PY, Huang PH *et al.* Magnetic Graphene Oxide for Dual Targeted Delivery of  
34 Doxorubicin and Photothermal Therapy. *Nanomaterials (Basel)* 8(4), (2018).
- 35 129. Lu Q, Dai X, Zhang P *et al.* Fe<sub>3</sub>O<sub>4</sub>@Au composite magnetic nanoparticles modified with  
36 cetuximab for targeted magneto-photothermal therapy of glioma cells. *Int J Nanomedicine* 13  
37 2491-2505 (2018).
- 38 130. Zhang L, Sheng D, Wang D *et al.* Bioinspired Multifunctional Melanin-Based Nanoliposome for  
39 Photoacoustic/Magnetic Resonance Imaging-Guided Efficient Photothermal Ablation of Cancer.  
40 *Theranostics* 8(6), 1591-1606 (2018).
- 41 131. Lu YJ, Wei KC, Ma CC, Yang SY, Chen JP. Dual targeted delivery of doxorubicin to cancer cells  
42 using folate-conjugated magnetic multi-walled carbon nanotubes. *Colloids Surf B Biointerfaces*  
43 89 1-9 (2012).
- 44 132. Felfoul O, Mohammadi M, Taherkhani S *et al.* Magneto-aerotactic bacteria deliver drug-  
45 containing nanoliposomes to tumour hypoxic regions. *Nat Nanotechnol* 11(11), 941-947 (2016).
- 46 133. Yun WS, Choi JS, Ju HM *et al.* Enhanced Homing Technique of Mesenchymal Stem Cells Using  
47 Iron Oxide Nanoparticles by Magnetic Attraction in Olfactory-Injured Mouse Models. *Int J Mol*  
48 *Sci* 19(5), (2018).
- 49 134. Chen J, Huang N, Ma B *et al.* Guidance of stem cells to a target destination in vivo by magnetic  
50 nanoparticles in a magnetic field. *ACS Appl Mater Interfaces* 5(13), 5976-5985 (2013).
- 51  
52  
53  
54  
55  
56  
57  
58  
59  
60

---

**Article Body Template**

- 1  
2  
3  
4  
5  
6 135. Toro-Cordova A, Flores-Cruz M, Santoyo-Salazar J *et al.* Liposomes Loaded with Cisplatin and  
7 Magnetic Nanoparticles: Physicochemical Characterization, Pharmacokinetics, and In-Vitro  
8 Efficacy. *Molecules* 23(9), (2018).
- 9 136. Li L, Jiang W, Luo K *et al.* Superparamagnetic iron oxide nanoparticles as MRI contrast agents for  
10 non-invasive stem cell labeling and tracking. *Theranostics* 3(8), 595-615 (2013).
- 11 137. Hauser AK, Mitov MI, Daley EF, McGarry RC, Anderson KW, Hilt JZ. Targeted iron oxide  
12 nanoparticles for the enhancement of radiation therapy. *Biomaterials* 105 127-135 (2016).
- 13 138. Alphandery E, Faure S, Seksek O, Guyot F, Chebbi I. Chains of magnetosomes extracted from  
14 AMB-1 magnetotactic bacteria for application in alternative magnetic field cancer therapy. *ACS*  
15 *Nano* 5(8), 6279-6296 (2011).
- 16 139. Alphandery E, Idbaih A, Adam C *et al.* Chains of magnetosomes with controlled endotoxin  
17 release and partial tumor occupation induce full destruction of intracranial U87-Luc glioma in  
18 mice under the application of an alternating magnetic field. *J Control Release* 262 259-272  
19 (2017).
- 20 140. Le Fevre R, Durand-Dubief M, Chebbi I *et al.* Enhanced antitumor efficacy of biocompatible  
21 magnetosomes for the magnetic hyperthermia treatment of glioblastoma. *Theranostics* 7(18),  
22 4618-4631 (2017).
- 23 141. Du X, Zhou J, Xu B. Ectoenzyme switches the surface of magnetic nanoparticles for selective  
24 binding of cancer cells. *J Colloid Interface Sci* 447 273-277 (2015).
- 25 142. Muthana M, Kennerley AJ, Hughes R *et al.* Directing cell therapy to anatomic target sites in vivo  
26 with magnetic resonance targeting. *Nat Commun* 6 8009 (2015).
- 27 143. Zablotskii V, Polyakova T, Lunov O, Dejneka A. How a High-Gradient Magnetic Field Could Affect  
28 Cell Life. *Sci Rep* 6 37407 (2016).
- 29 144. Erdal E, Demirbilek M, Yeh Y *et al.* A Comparative Study of Receptor-Targeted Magnetosome  
30 and HSA-Coated Iron Oxide Nanoparticles as MRI Contrast-Enhancing Agent in Animal Cancer  
31 Model. *Appl Biochem Biotechnol* 185(1), 91-113 (2018).
- 32 145. Sun JB, Duan JH, Dai SL *et al.* In vitro and in vivo antitumor effects of doxorubicin loaded with  
33 bacterial magnetosomes (DBMs) on H22 cells: the magnetic bio-nanoparticles as drug carriers.  
34 *Cancer Lett* 258(1), 109-117 (2007).
- 35 146. Deng Q, Liu Y, Wang S *et al.* Construction of a Novel Magnetic Targeting Anti-Tumor Drug  
36 Delivery System: Cytosine Arabinoside-Loaded Bacterial Magnetosome. *Materials (Basel)* 6(9),  
37 3755-3763 (2013).
- 38 147. Liu YG, Dai QL, Wang SB, Deng QJ, Wu WG, Chen AZ. Preparation and in vitro antitumor effects  
39 of cytosine arabinoside-loaded genipin-poly-L-glutamic acid-modified bacterial magnetosomes.  
40 *Int J Nanomedicine* 10 1387-1397 (2015).
- 41 148. Long RM, Dai QL, Zhou X *et al.* Bacterial magnetosomes-based nanocarriers for co-delivery of  
42 cancer therapeutics in vitro. *Int J Nanomedicine* 13 8269-8279 (2018).
- 43 149. Tang YS, Wang D, Zhou C *et al.* Bacterial magnetic particles as a novel and efficient gene vaccine  
44 delivery system. *Gene Ther* 19(12), 1187-1195 (2012).
- 45 150. Davola ME, Mossman KL. Oncolytic viruses: how "lytic" must they be for therapeutic efficacy?  
46 *Oncoimmunology* 8(6), e1581528 (2019).
- 47 151. Brestovitsky A, Sharf R, Mittelman K, Kleinberger T. The adenovirus E4orf4 protein targets PP2A  
48 to the ACF chromatin-remodeling factor and induces cell death through regulation of SNF2h-  
49 containing complexes. *Nucleic Acids Res* 39(15), 6414-6427 (2011).
- 50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

---

### Article Body Template

152. Ito N, Demarco RA, Mailliard RB *et al.* Cytolytic cells induce HMGB1 release from melanoma cell lines. *J Leukoc Biol* 81(1), 75-83 (2007).
153. Haanen J. Converting Cold into Hot Tumors by Combining Immunotherapies. *Cell* 170(6), 1055-1056 (2017).
154. Kohlhapp FJ, Kaufman HL. Molecular Pathways: Mechanism of Action for Talimogene Laherparepvec, a New Oncolytic Virus Immunotherapy. *Clin Cancer Res* 22(5), 1048-1054 (2016).
155. Chaurasiya S, Chen NG, Fong Y. Oncolytic viruses and immunity. *Curr Opin Immunol* 51 83-90 (2018).
156. Diaconu I, Cerullo V, Hirvonen ML *et al.* Immune response is an important aspect of the antitumor effect produced by a CD40L-encoding oncolytic adenovirus. *Cancer Res* 72(9), 2327-2338 (2012).
157. Donnelly OG, Errington-Mais F, Steele L *et al.* Measles virus causes immunogenic cell death in human melanoma. *Gene Ther* 20(1), 7-15 (2013).
158. Workenhe ST, Mossman KL. Oncolytic virotherapy and immunogenic cancer cell death: sharpening the sword for improved cancer treatment strategies. *Mol Ther* 22(2), 251-256 (2014).
159. Breitbach CJ, De Silva NS, Falls TJ *et al.* Targeting tumor vasculature with an oncolytic virus. *Mol Ther* 19(5), 886-894 (2011).
160. Hou W, Chen H, Rojas J, Sampath P, Thorne SH. Oncolytic vaccinia virus demonstrates antiangiogenic effects mediated by targeting of VEGF. *Int J Cancer* 135(5), 1238-1246 (2014).
161. Angarita FA, Acuna SA, Ottolino-Perry K, Zerhouni S, Mccart JA. Mounting a strategic offense: fighting tumor vasculature with oncolytic viruses. *Trends Mol Med* 19(6), 378-392 (2013).
162. Thorne SH, Tam BY, Kirn DH, Contag CH, Kuo CJ. Selective intratumoral amplification of an antiangiogenic vector by an oncolytic virus produces enhanced antivascular and anti-tumor efficacy. *Mol Ther* 13(5), 938-946 (2006).
163. Chiocca EA, Rabkin SD. Oncolytic viruses and their application to cancer immunotherapy. *Cancer Immunol Res* 2(4), 295-300 (2014).

- **Reference annotations:** authors should highlight 6–8 references that are of particular significance to the subject under discussion as “\* of interest” or “\*\* of considerable interest”, and provide a brief (1–2 line) synopsis.



Figure 1

175x152mm (150 x 150 DPI)

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

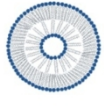



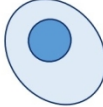
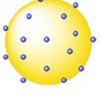
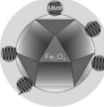

Lipid nanocarriers		Polymeric nanocarriers		Organic nanocarriers	Metal nanocarriers		
Liposome	Micelles	Polymer	Dendrimer	Cells	Gold nanoparticle	Magnetosome	Iron oxide nanoparticle
							
25-2500nm	2-20nm	10-1000nm	1-100nm	25000nm	5-400nm	30-120nm	1-100nm

Figure 2

266x75mm (150 x 150 DPI)

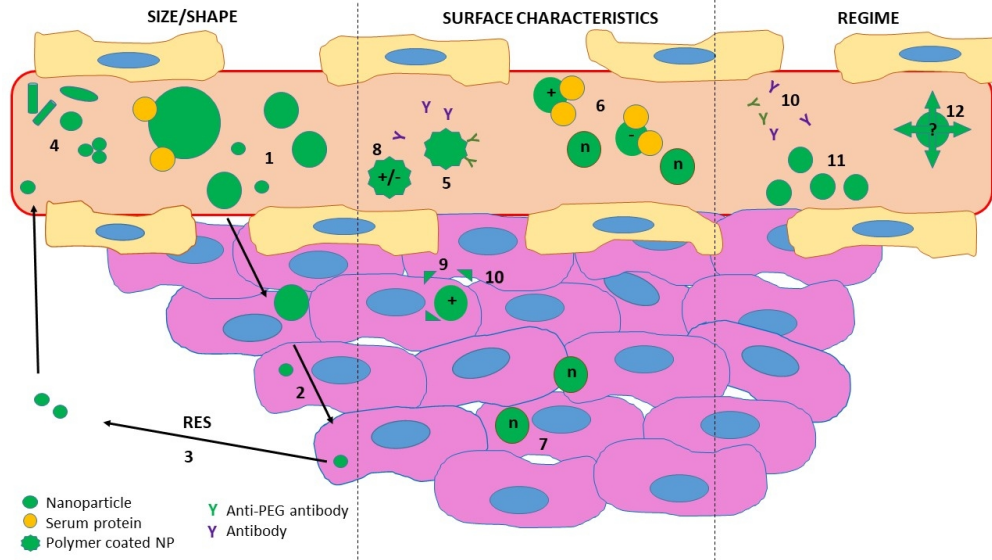
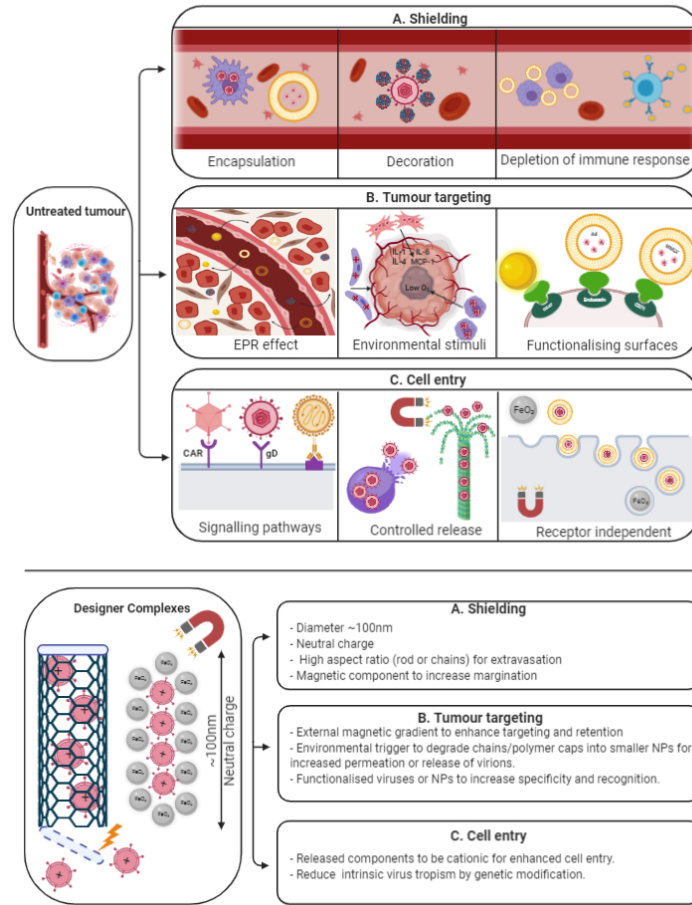


Figure 3

338x190mm (96 x 96 DPI)



CREATED WITH  
**bio**  
**RENDER**  
 WWW.BIORENDER.COM

Figure 4