



The
University
Of
Sheffield.

This is a repository copy of *Antibacterial activity of Mn(i) and Re(i) tricarbonyl complexes conjugated to a bile acid carrier molecule*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/175295/>

Version: Supplemental Material

Article:

Betts, J.W., Roth, P., Patrick, C.A. et al. (4 more authors) (2020) Antibacterial activity of Mn(i) and Re(i) tricarbonyl complexes conjugated to a bile acid carrier molecule. *Metalomics*, 12 (10). pp. 1563-1575. ISSN 1756-5901

<https://doi.org/10.1039/d0mt00142b>

This is a pre-copyedited, author-produced version of an article accepted for publication in *Metalomics* following peer review. The version of record [Jono W Betts, Patrick Roth, Calum A Patrick, Hannah M Southam, Roberto M La Ragione, Robert K Poole, Ulrich Schatzschneider, Antibacterial activity of Mn(I) and Re(I) tricarbonyl complexes conjugated to a bile acid carrier molecule, *Metalomics*, Volume 12, Issue 10, October 2020, Pages 1563–1575] is available online at: <https://doi.org/10.1039/d0mt00142b>

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

Antibacterial activity of Mn(I) and Re(I) tricarbonyl complexes conjugated to a bile acid carrier molecule

Jono W. Betts,^a Patrick Roth,^b Calum A. Patrick,^c Hannah Southam,^c Roberto La Ragione,^a
Robert K. Poole,^c and Ulrich Schatzschneider^{b*}

^a Department of Pathology and Infectious Diseases, School of Veterinary Medicine,
University of Surrey, Guildford, United Kingdom

^b Institut für Anorganische Chemie, Julius-Maximilians-Universität Würzburg, Am Hubland,
D-97074 Würzburg, Germany

^c Department of Molecular Biology and Biotechnology, The University of Sheffield, United
Kingdom

* Corresponding author: eMail: ulrich.schatzschneider@uni-wuerzburg.de;
Tel: +49 931 31 83636; Fax: +49 931 31 84605

Supporting Information

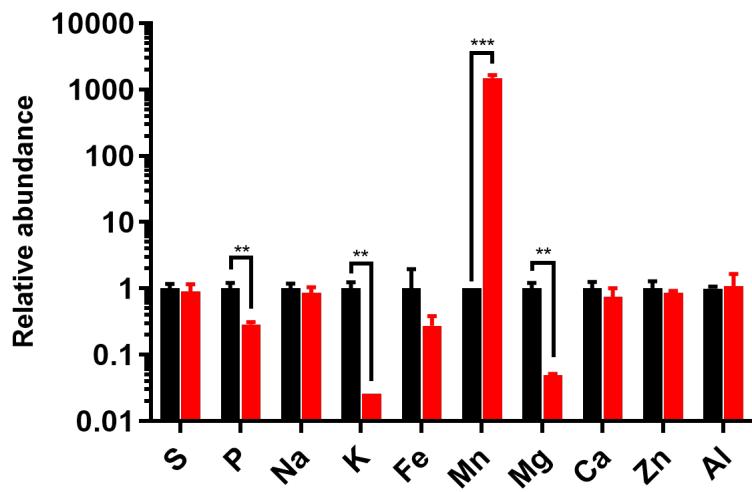


Figure S1. ICP-MS determination of the intracellular metal content of *E. coli* treated with $[\text{Mn}(\text{CO})_3(\text{bqpa}-\kappa^3\text{N})]\text{Br}$ (red bars) relative to the metal content of control samples treated with 1% DMSO (black bars). Data shown are mean of three biological replicates and errors bars represent the standard deviation. *, **, and *** denote *p*-values of 0.01–0.05, 0.001–0.01, and < 0.001 respectively, by Student's *t*-test.

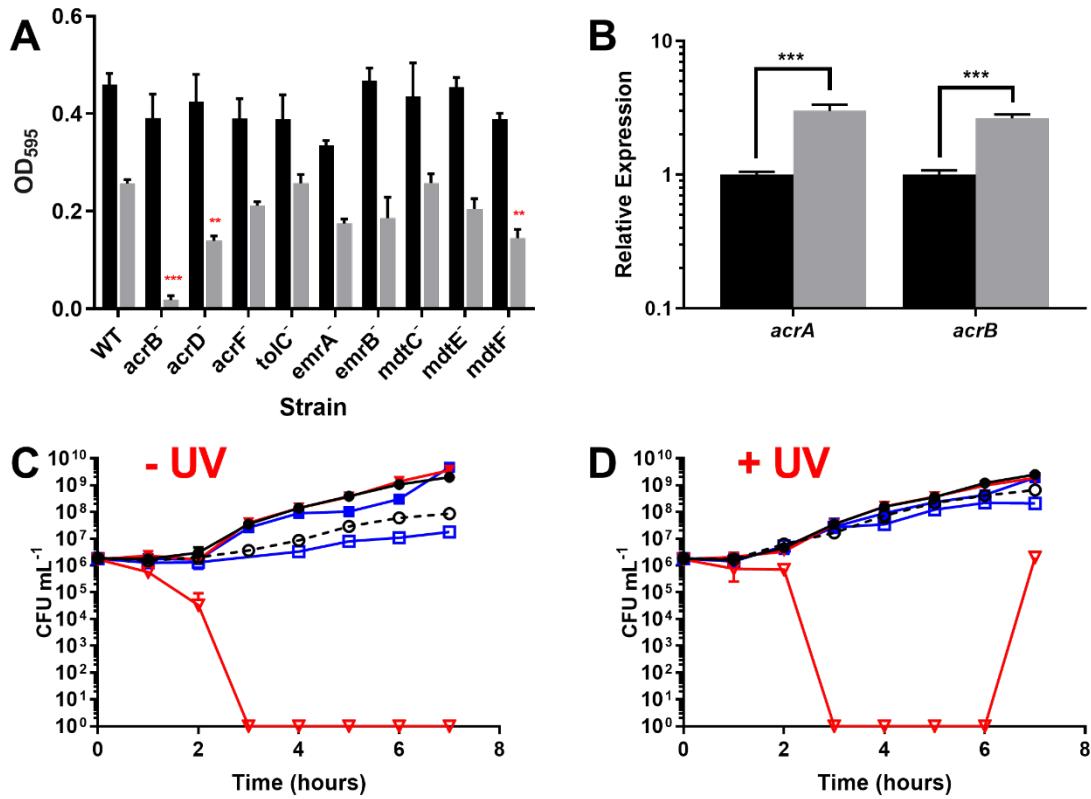


Figure S2. A) 5 h growth of *E. coli* strains in the absence (black bars) and presence (grey bars) of $[\text{Mn}(\text{CO})_3(\text{bqpa}-\kappa^3\text{N})]\text{Br}$. Values are means of three biological repeats with error bars indicating standard deviation. B) RT-PCR showing the relative expression of the *E. coli* genes *acrA* and *acrB* in cells grown in the absence (black bars) and presence (grey bars) of $[\text{Mn}(\text{CO})_3(\text{bqpa}-\kappa^3\text{N})]\text{Br}$. Expression levels are reported relative to the levels determined for untreated cells and normalized to the *rrsA* housekeeping gene. Data are plotted as means of results from three biological replicates (each consisting of three technical replicates) with standard deviations shown as error bars. C) Growth of *E. coli* without and D) with UV activation of $[\text{Mn}(\text{CO})_3(\text{bqpa}-\kappa^3\text{N})]\text{Br}$. Black lines represent wildtype (BW25113), red lines represent *acrB*⁻ and blue line represent *acrD*⁻; solid points indicate the 1% DMSO control and unfilled points indicate cells treated with 105 μM $[\text{Mn}(\text{CO})_3(\text{bqpa}-\kappa^3\text{N})]\text{Br}$. *, **, and *** denote *p*-values of 0.01–0.05, 0.001–0.01, and < 0.001 respectively, by Student's *t*-test.

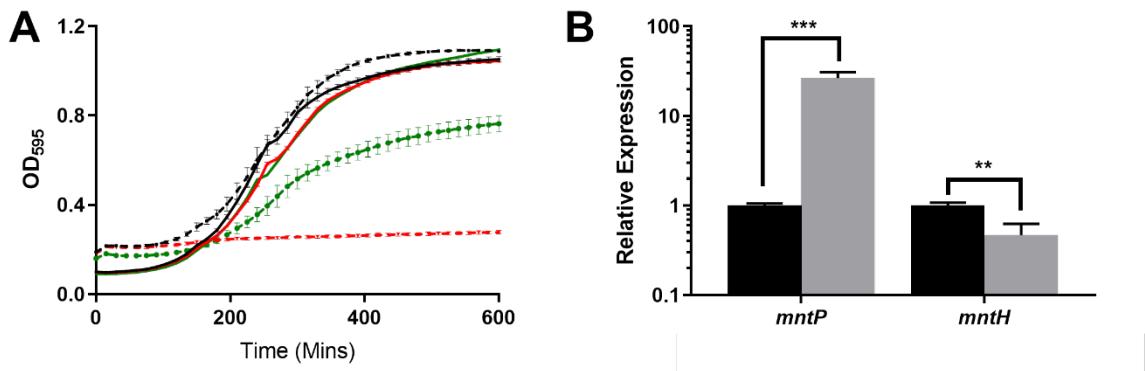
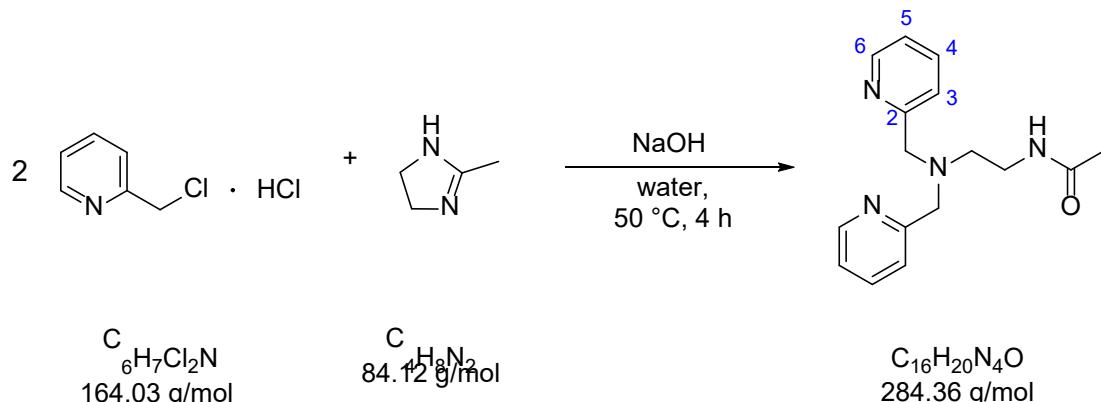


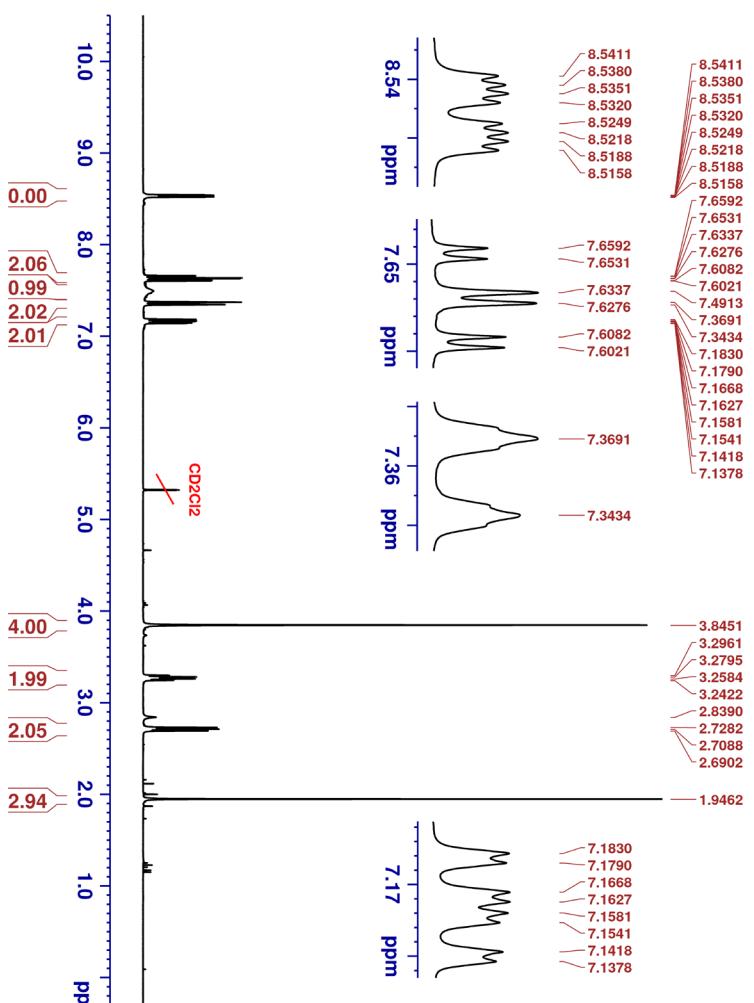
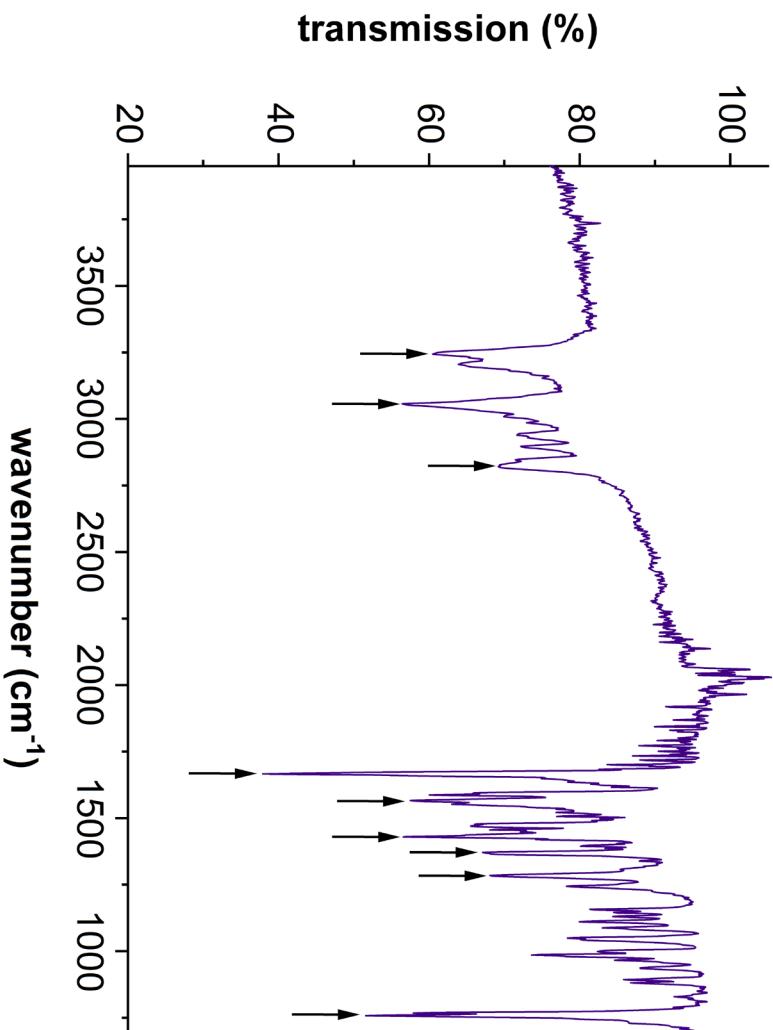
Figure S3. A) Growth curves of *E. coli* strains with (solid line) and without (dashed line) addition of $[\text{Mn}(\text{bqpa-}\kappa^3\text{N})(\text{CO})_3]\text{Br}$ (105 μM); WT (BW25113) is represented by the black lines, *mntP*⁺ is represented by the red lines and *mntR*⁺ is represented by the green lines and B) RT-PCR of *mntP* and *mntH* indicates activation of the Mn regulator MntR, as shown by the increased in expression of *mntP* and the decrease in expression of *mntH* in response to treatment with $[\text{Mn}(\text{bqpa-}\kappa^3\text{N})(\text{CO})_3]\text{Br}$ (grey) compared to the 1% DMSO control (black).

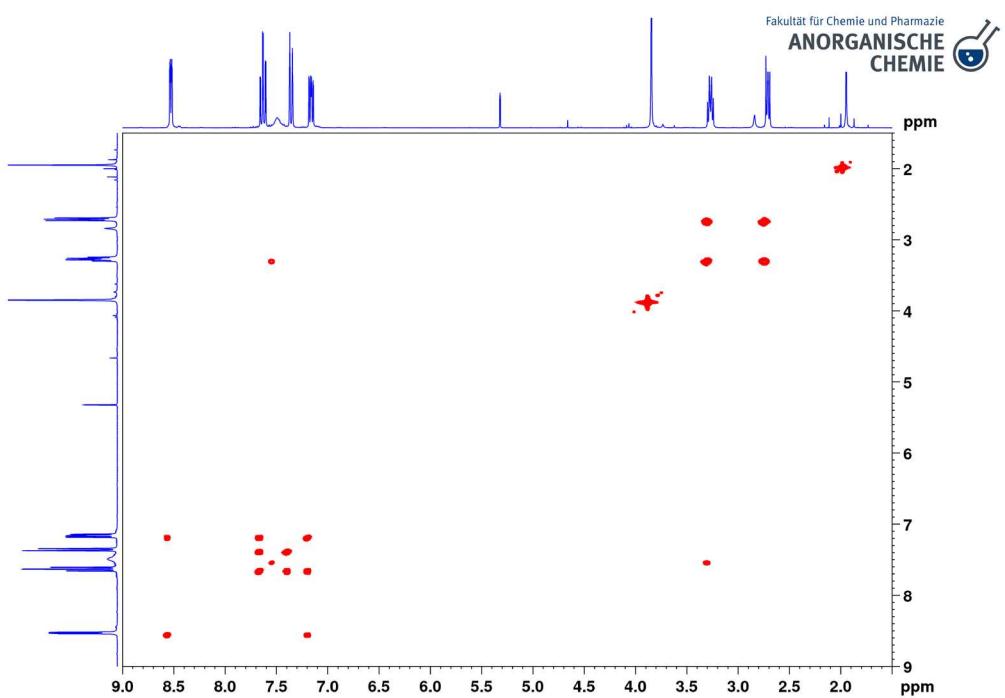
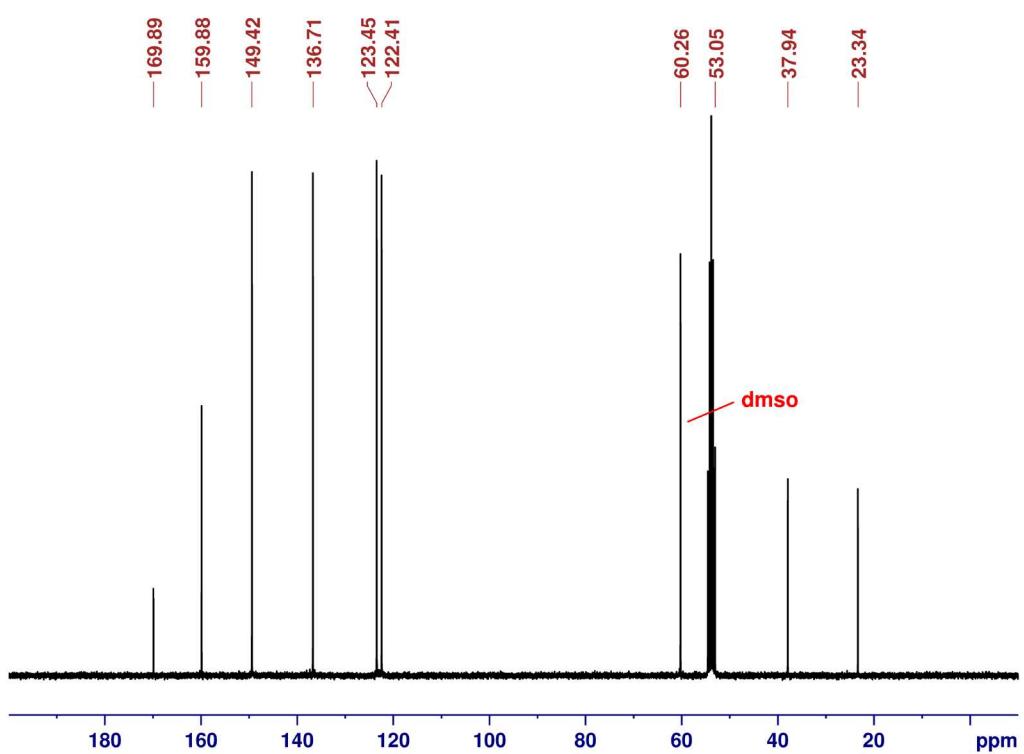
Ligand synthesis

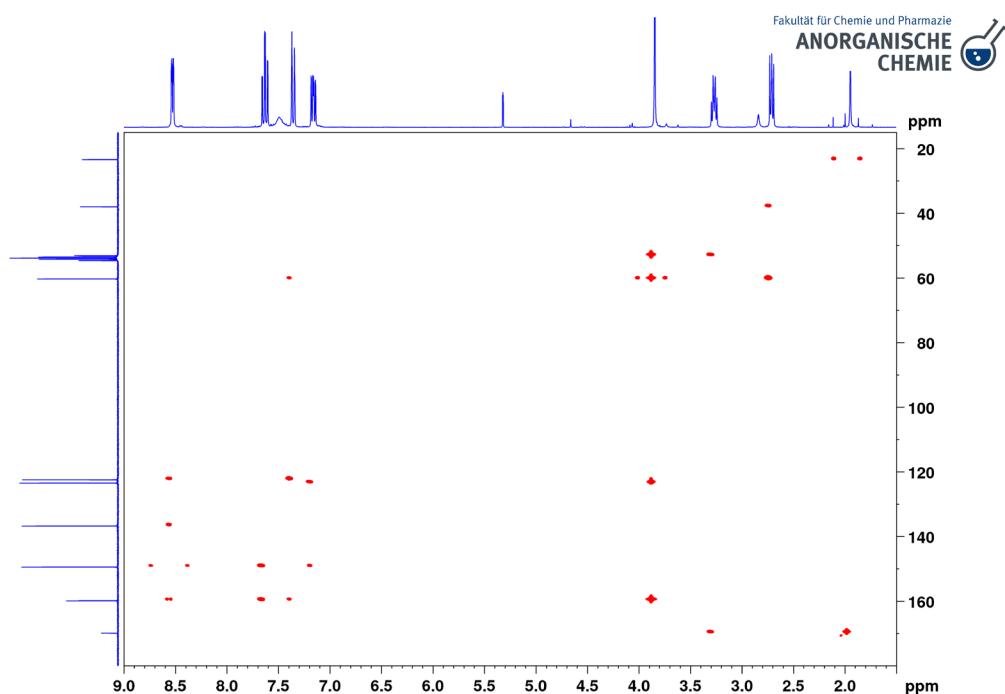
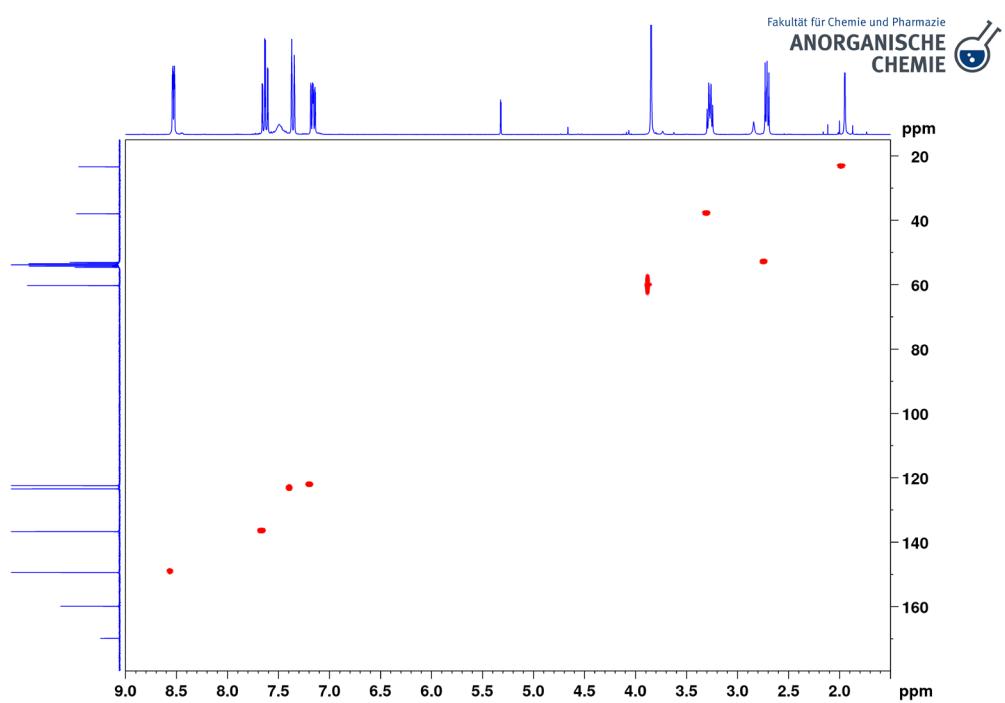
Synthesis of *N*-(2-(bis(pyridin-2-ylmethyl)amino)ethyl)acetamide (**bpen**^{COCH₃})¹ **3**

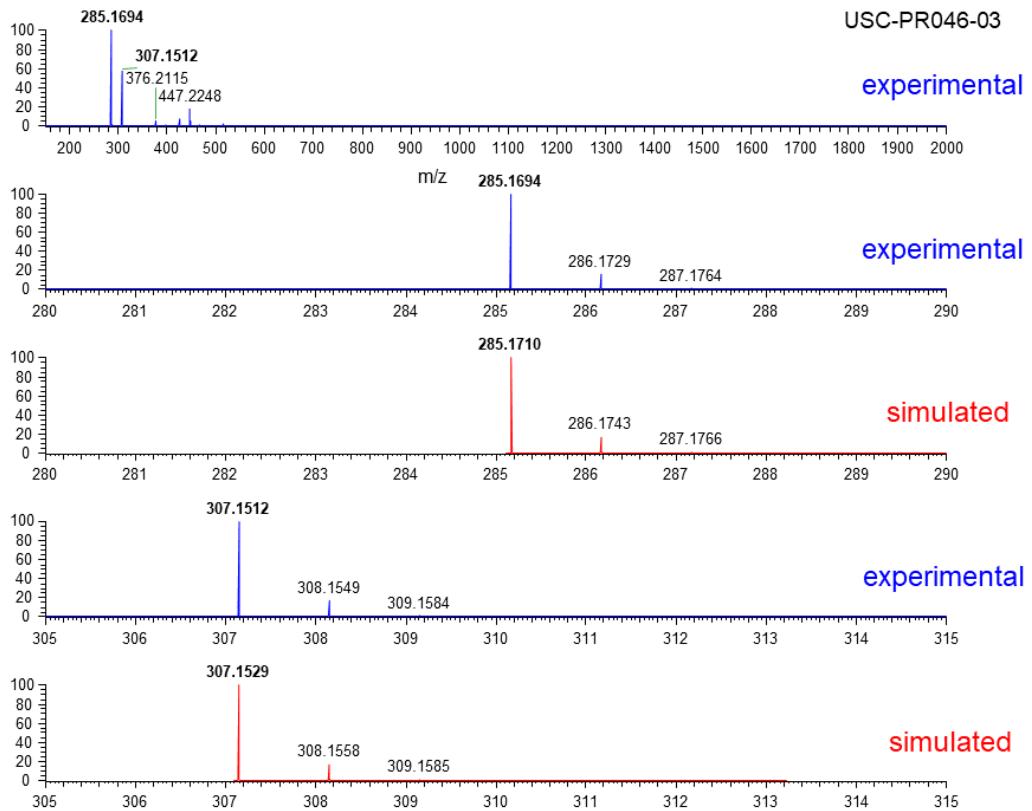


2-Methyl-4,5-dihydro-1*H*-imidazole (5.14 g, 61.1 mmol) was dissolved in water (5 mL) and heated to 75 °C for 2 h. The resulting yellow solution was cooled to room temperature and added to a solution of 2-(chloromethyl)pyridine hydrochloride (20.05 g, 122.2 mmol) in water (50 mL). The mixture was heated to 50 °C and then, with vigorous stirring, 10 M aqueous sodium hydroxide (24.5 mL, 245 mmol) was slowly added over 2 h, resulting in a colour change from light brown to violet-red. After complete addition, heating was continued for another 2 h and the reaction mixture then cooled to room temperature. The solution was extracted with dichlormethane (4 x 50 mL), the combined organic phases dried over sodium sulfate, the solvent removed under vacuum. The resulting dark red oil was dissolved in ethyl acetate (15 mL) and applied to a glas frit filled with neutral aluminium oxide. The yellow-orange product was eluted with ethyl acetate (approx. 300 mL) and the solvent removed under vacuum to obtain the product as a beige solid. Yield: 50% (8.70 g, 30.6 mmol). **IR** (ATR): $\tilde{\nu} = 3244$ (m), 3056 (m), 2821 (w), 1666 (s), 1566 (m), 1430 (m), 1370 (w), 1284 (w), 758 (s) cm^{-1} ; **¹H NMR** (300.18 MHz, CD₂Cl₂): $\delta = 8.53$ (ddd, 2H, $^3J_{\text{H}6,\text{H}5} = 4.9$ Hz, $^4J_{\text{H}6,\text{H}4} = 1.8$ Hz, $^5J_{\text{H}6,\text{H}3} = 0.9$ Hz, py-H6), 7.63 (dt, 2H, $^3J_{\text{H}4,\text{H}5/\text{H}3} = 7.7$ Hz, $^4J_{\text{H}4,\text{H}6} = 1.8$ Hz, py-H4), 7.49 (s, 1H, NH), 7.36 (d, 2H, $^3J_{\text{H}3,\text{H}4} = 7.7$ Hz, py-H3), 7.16 (ddd, 2H, $^3J_{\text{H}5,\text{H}4} = 7.5$ Hz, $^3J_{\text{H}5,\text{H}6} = 4.9$ Hz, $^4J_{\text{H}5,\text{H}3} = 1.2$ Hz, py-H5), 3.85 (s, 4H, py-CH₂), 3.27 (q, 2H, $^3J = 5.4$ Hz, CH₂NHAc), 2.71 (t, 2H, $^3J = 5.7$ Hz, (Py-CH₂)₂NCH₂), 1.95 (s, 3H, CH₃) ppm; **¹³C NMR** (75.48 MHz, CD₂Cl₂): $\delta = 169.89$ (C=O) 159.88 (py-C2), 149.42 (py-C6), 136.71 (py-C4), 123.45 (py-C3), 122.41 (py-C5), 60.26 (py-CH₂), 53.05 ((py-CH₂)₂NCH₂), 37.94 (CH₂NHAc), 23.34 (CH₃) ppm; **MS** (ESI⁺, CH₃OH): *m/z* = 285.1694 [M+H]⁺, 307.1513 [M+Na]⁺; **Elemental analysis (%)** calcd. for C₁₆H₂₀N₄O: C 67.58, H 7.09, N 19.70; found (%): C 67.32, H 7.07, N 19.70.

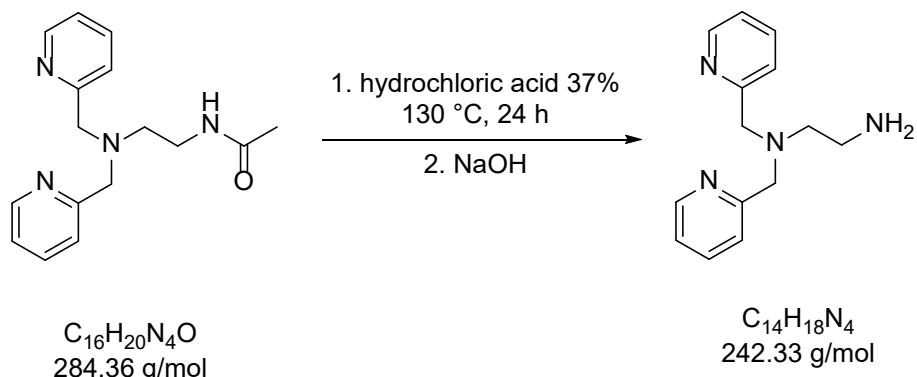




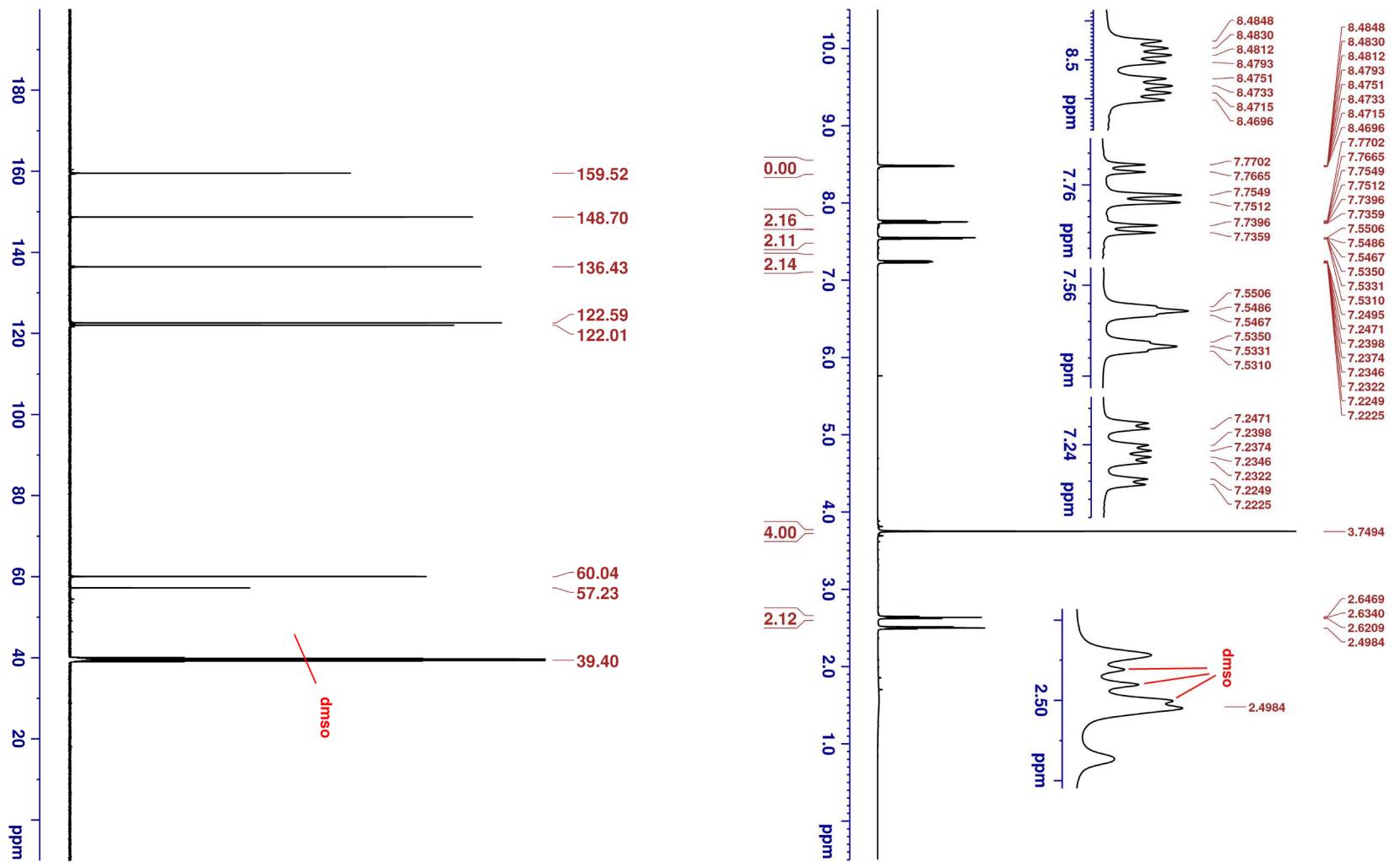




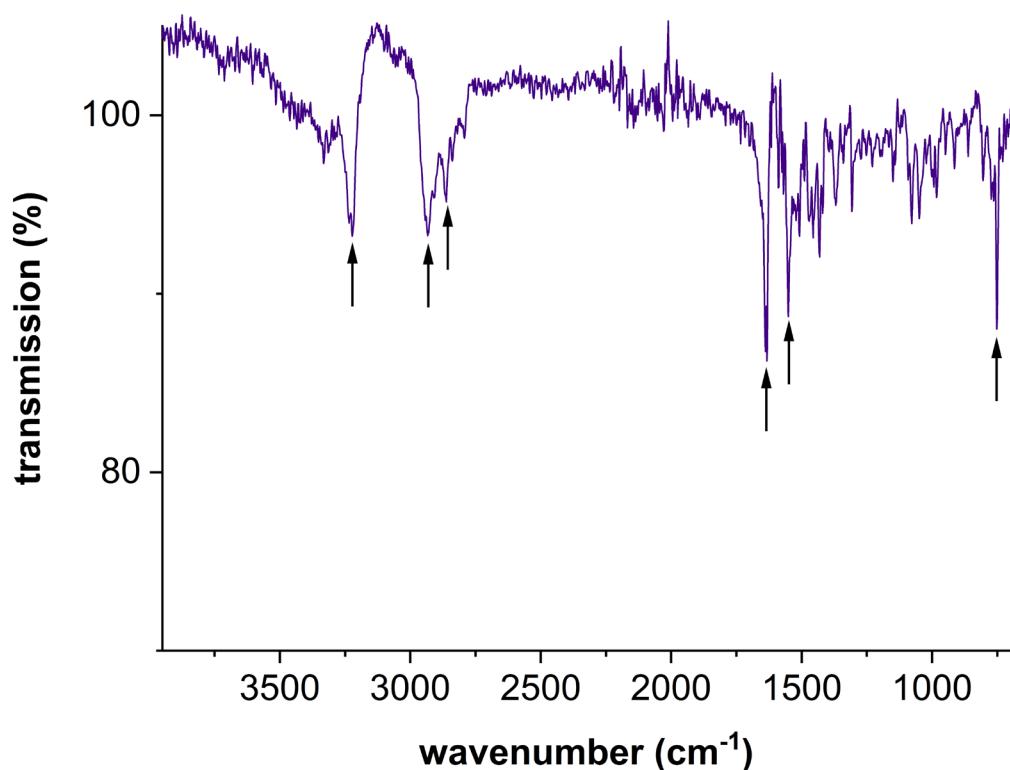
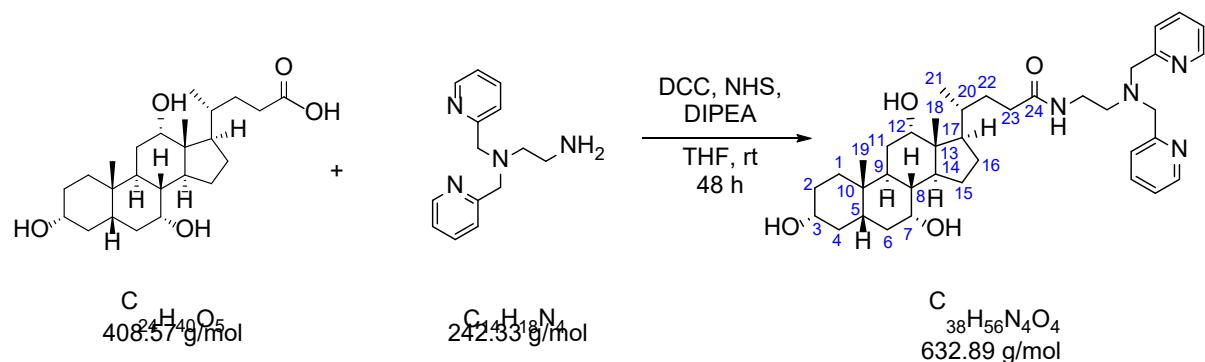
Synthesis of *N,N*-bis(pyridin-2-ylmethyl)ethan-1,2-diamine (**bpen**)¹ 4

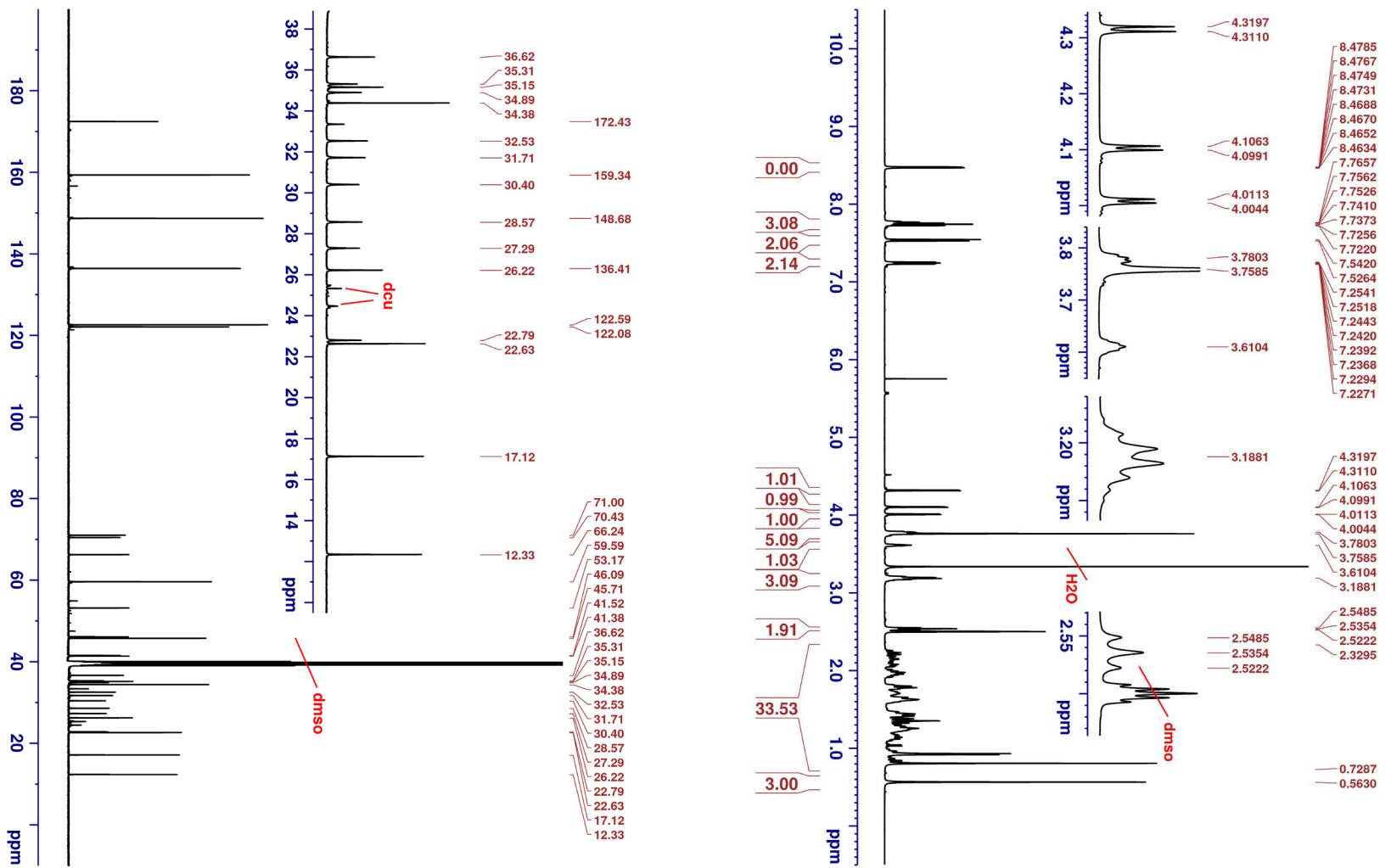


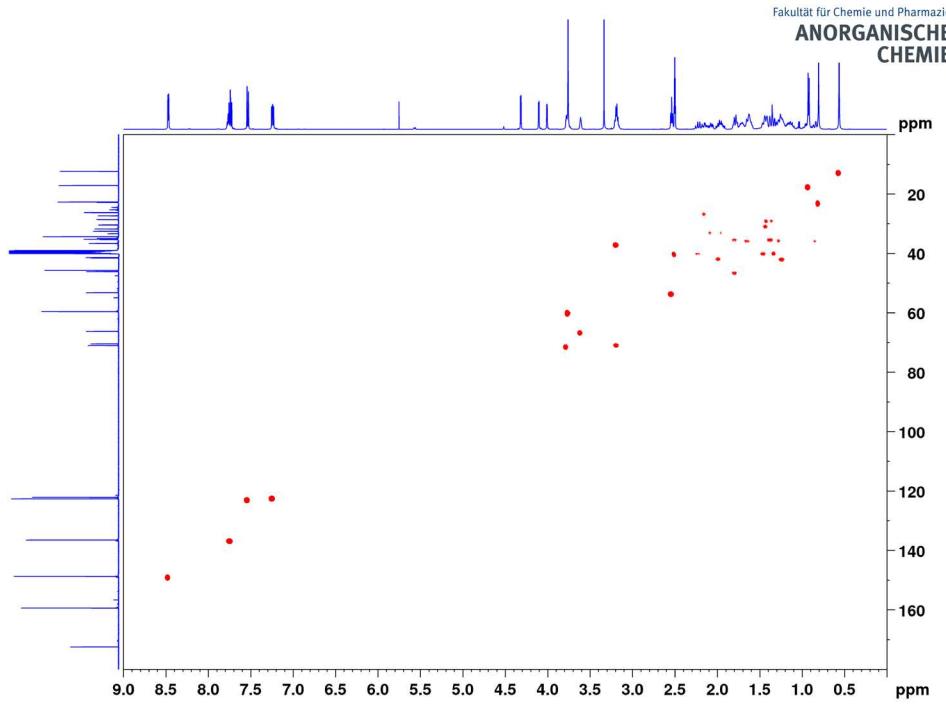
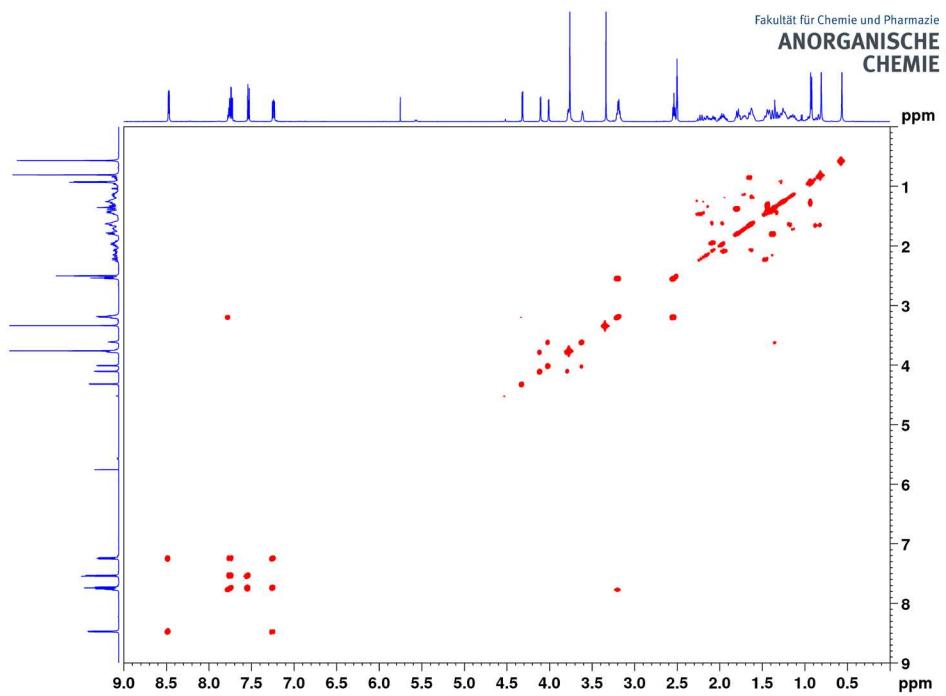
N-(2-(Bis(pyridin-2-ylmethyl)amino)ethyl)acetamide (4.73 g, 16.6 mmol) was dissolved in concentrated hydrochloric acid (50 mL) and then heated to 130 °C for 24 h. The resulting pale yellow solution was carefully diluted with water (150 mL) and then solid sodium hydroxide was added to adjust to pH 10. The aqueous solution was extracted with dichlormethane (3 x 50 mL) and the combined organic phases were dried over sodium sulfate. After removal of the solvent under vacuum, the product was obtained as a pale yellow oil. Since the compound is prone to decomposition, it was directly used in the next step with only limited characterization. Yield: 96% (3.89 g, 16.0 mmol). **¹H NMR** (500.13 MHz, DMSO-*d*₆): δ = 8.48 (ddd, 2H, ³J_{H6,H5} = 4.9 Hz, ⁴J_{H6,H4} = 1.8 Hz, ⁵J_{H6,H3} = 0.9 Hz, py-H6), 7.75 (dt, 2H, ³J_{H4,H5/H3} = 7.7 Hz, ⁴J_{H4,H6} = 1.9 Hz, py-H4), 7.54 (td, 2H, ³J_{H3,H4} = 7.7 Hz, ⁴J_{H3,H5} = 1.0 Hz, ⁵J_{H3,H6} = 1.0 Hz, py-H3), 7.24 (ddd, 2H, ³J_{H5,H4} = 7.5 Hz, ³J_{H5,H6} = 4.9 Hz, ⁴J_{H5,H3} = 1.2 Hz, py-H5), 3.75 (s, 4H, py-CH₂), 2.63 (t, 2H, ³J = 6.5 Hz, CH₂NH₂), 2.50 (t, 2H, ³J = 6.5 Hz, (py-CH₂)₂NCH₂, overlapping with solvent peak) ppm; **¹³C NMR** (125.76 MHz, DMSO-*d*₆): δ = 159.52 (py-C2), 148.70 (py-C6), 136.43 (py-C4), 122.59 (py-C3), 122.01 (py-C5), 60.04 (py-CH₂), 57.23 ((py-CH₂)₂NCH₂), 39.40 (CH₂NH₂, overlapping with solvent peak) ppm.

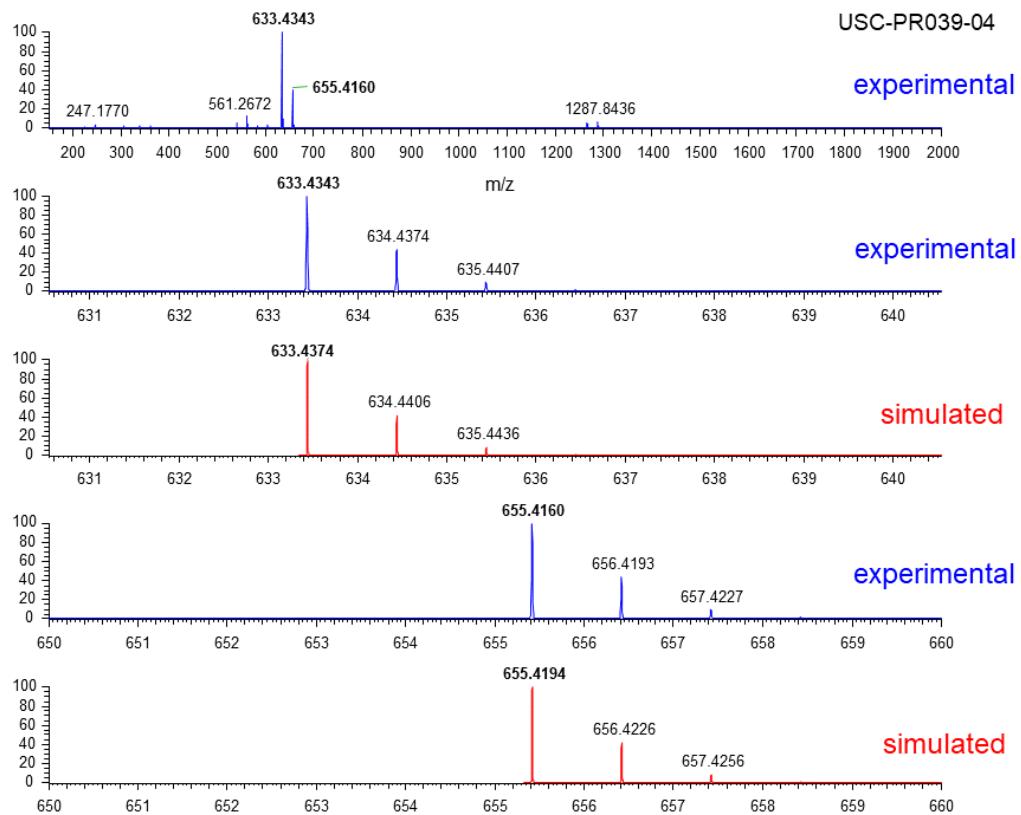
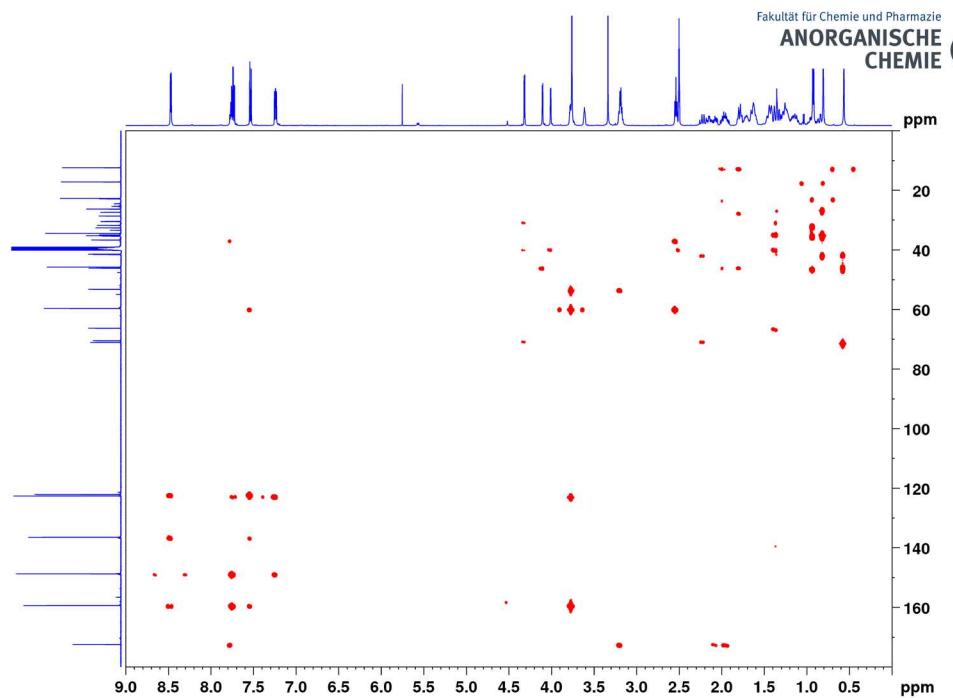


**Spectroscopic data for *N*-(2-(bis(pyridin-2-ylmethyl)amino)ethyl)cholamide
(bpen^{cholamid}) 6**

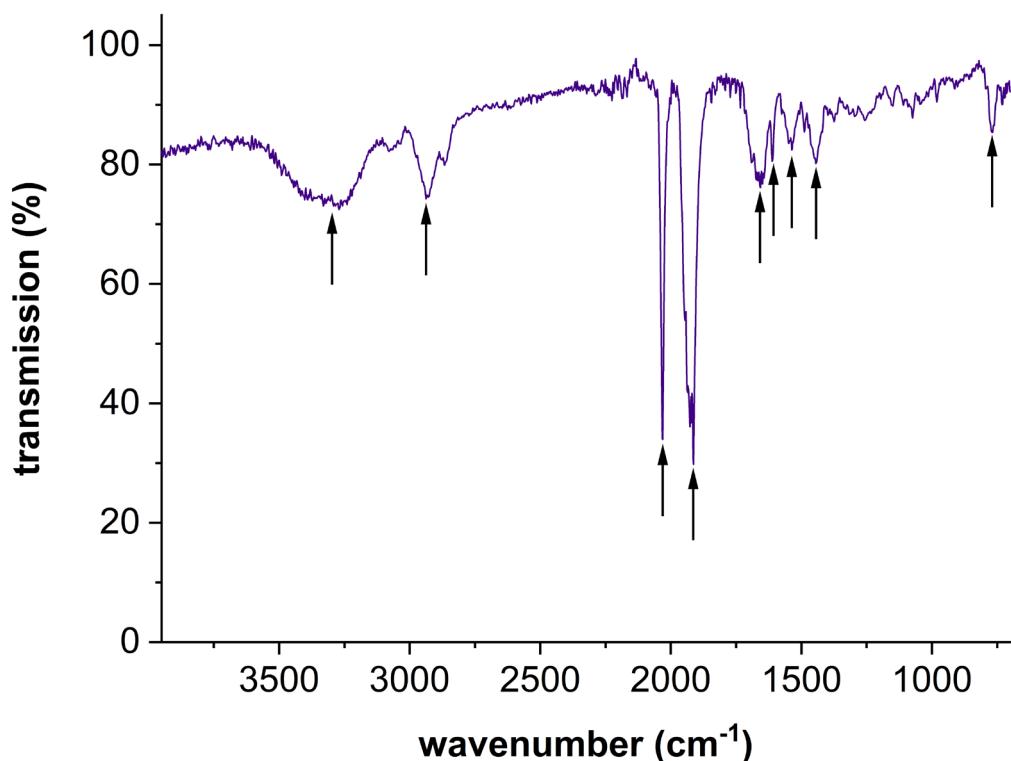
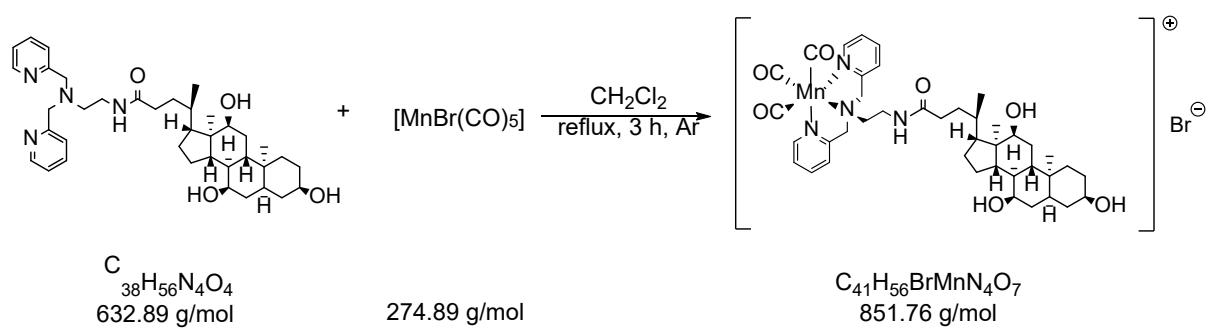


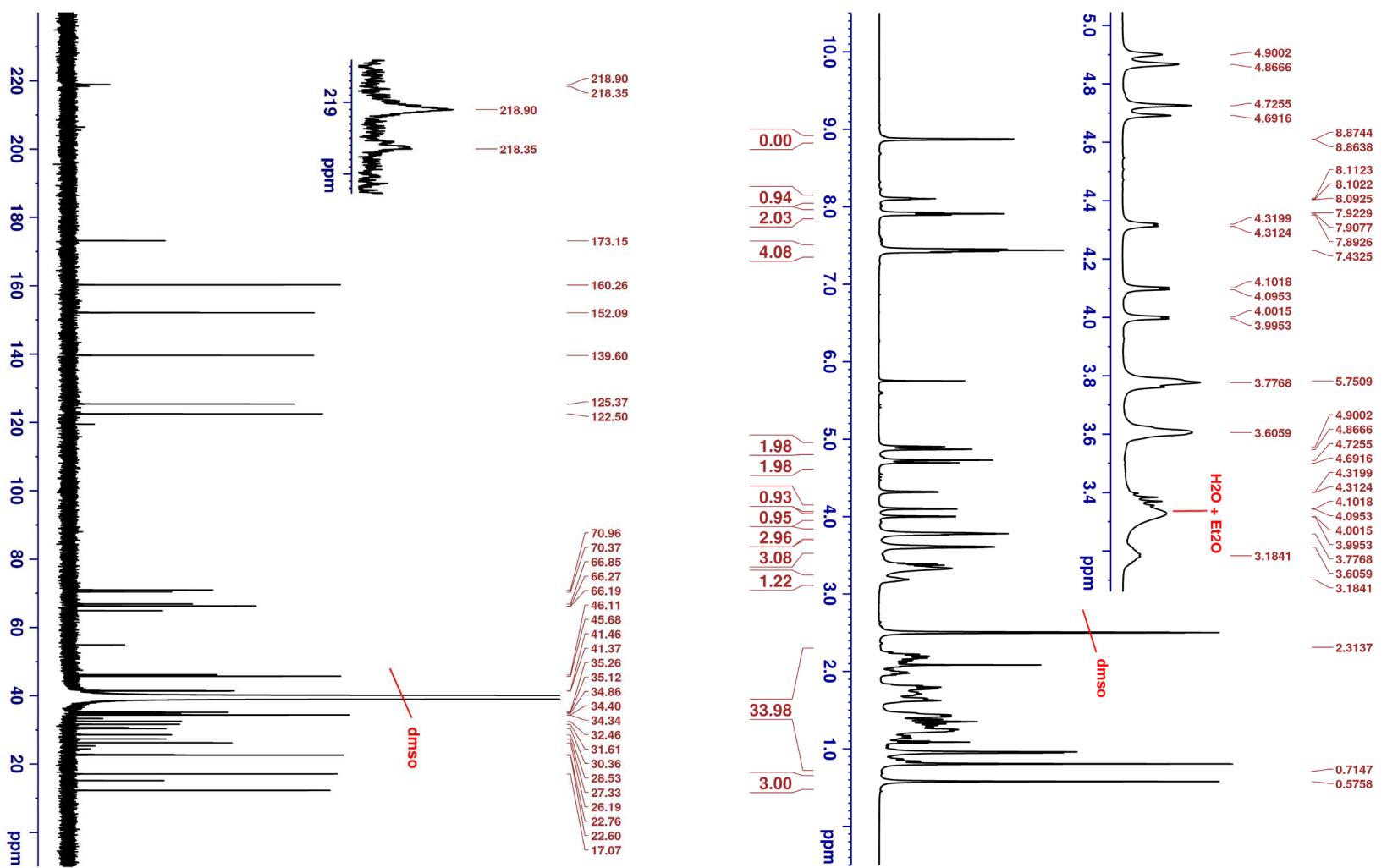




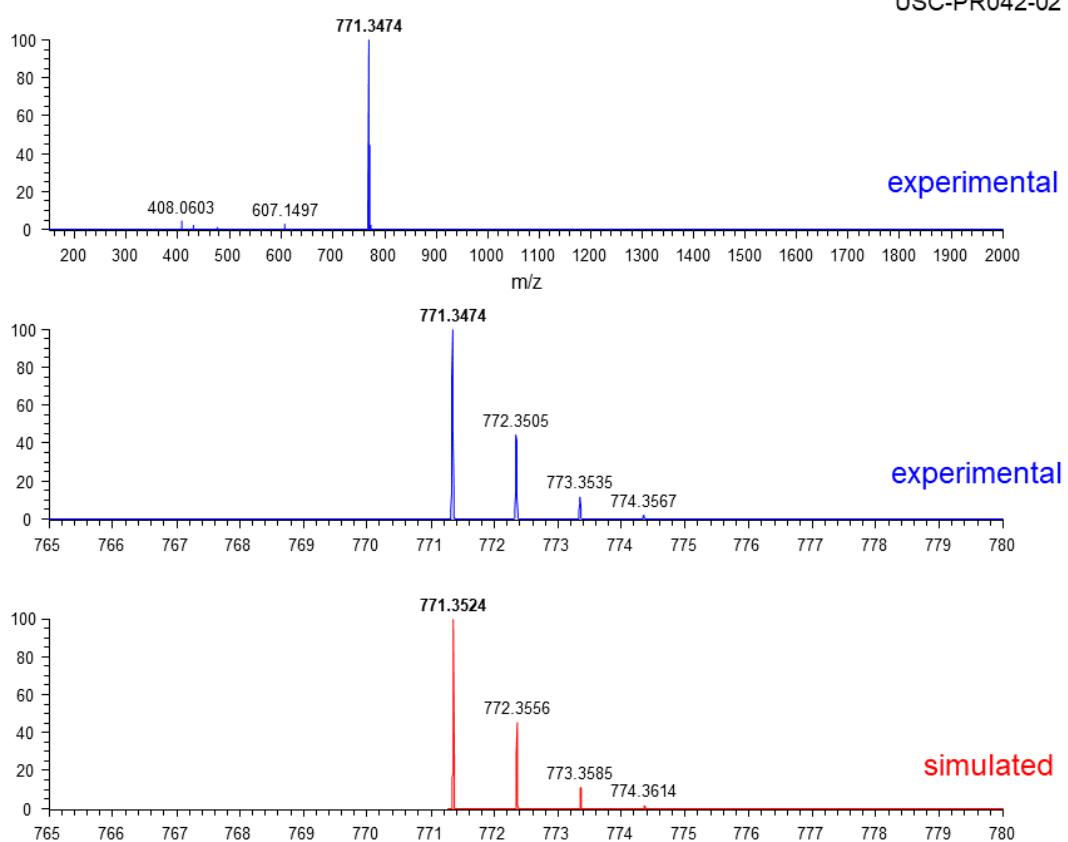


Spectroscopic data for $[\text{Mn}(\text{bpen}^{\text{cholamid}}-\kappa^3\text{N})(\text{CO})_3]\text{Br}$ 9

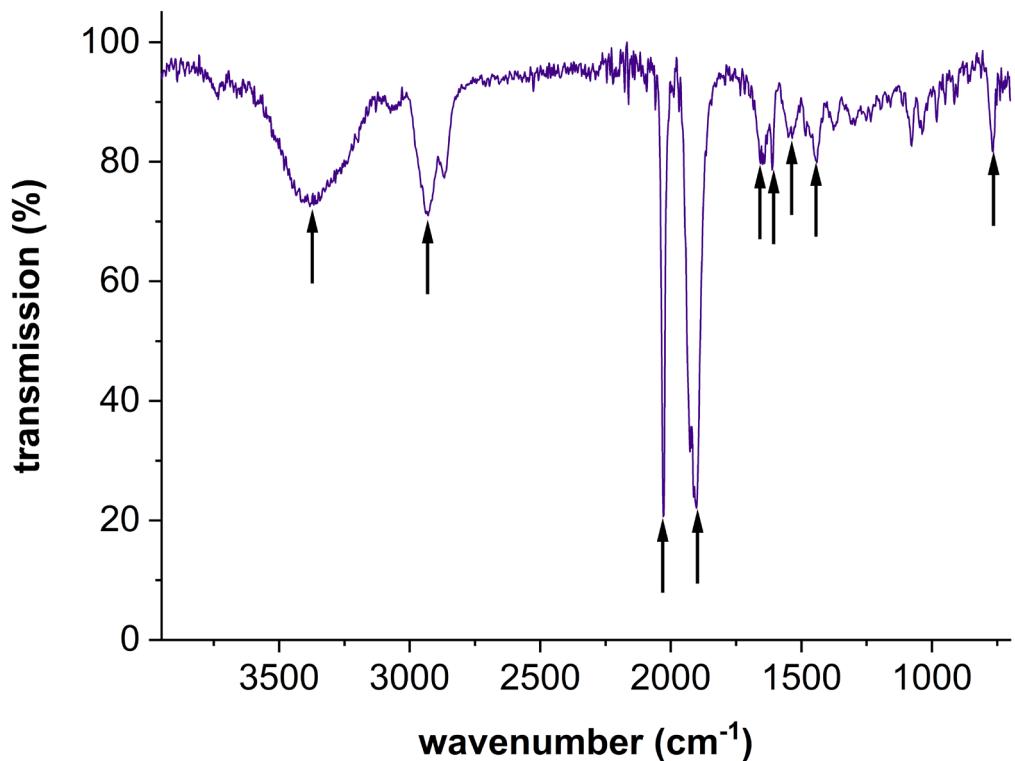
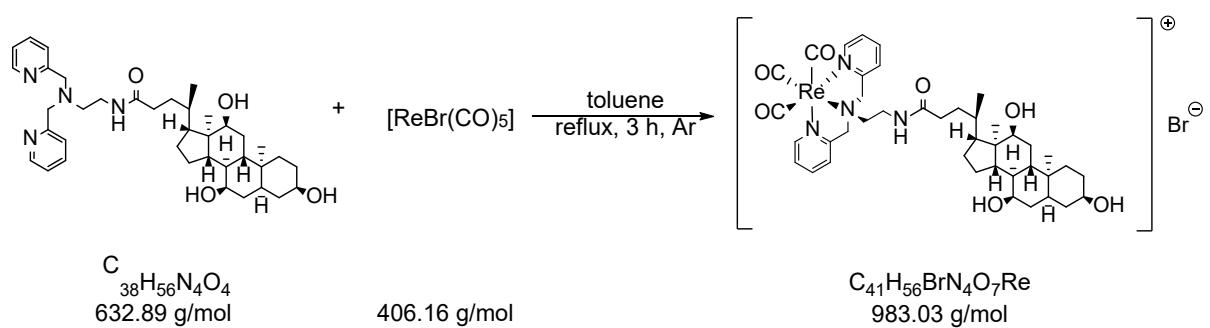


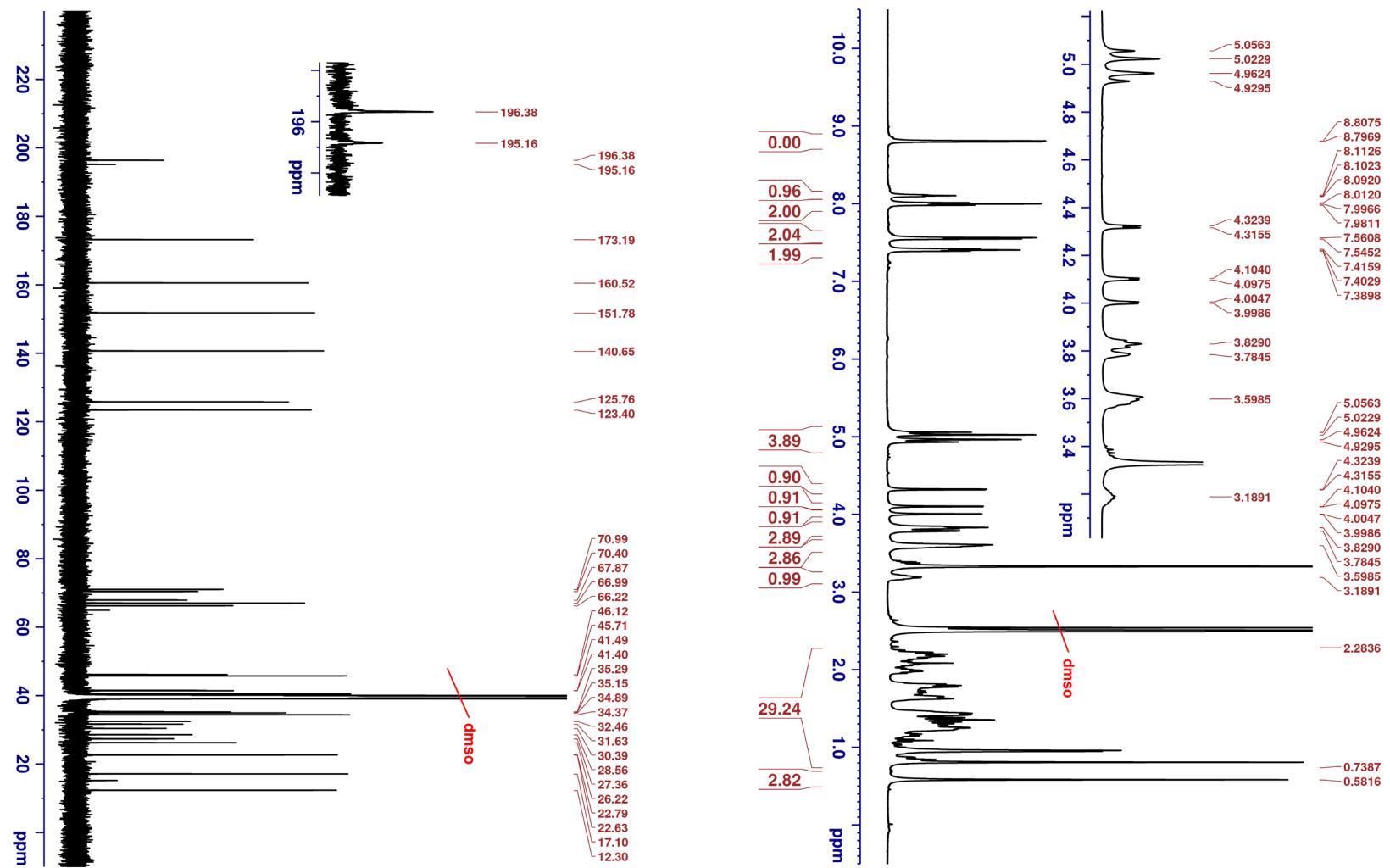


USC-PR042-02

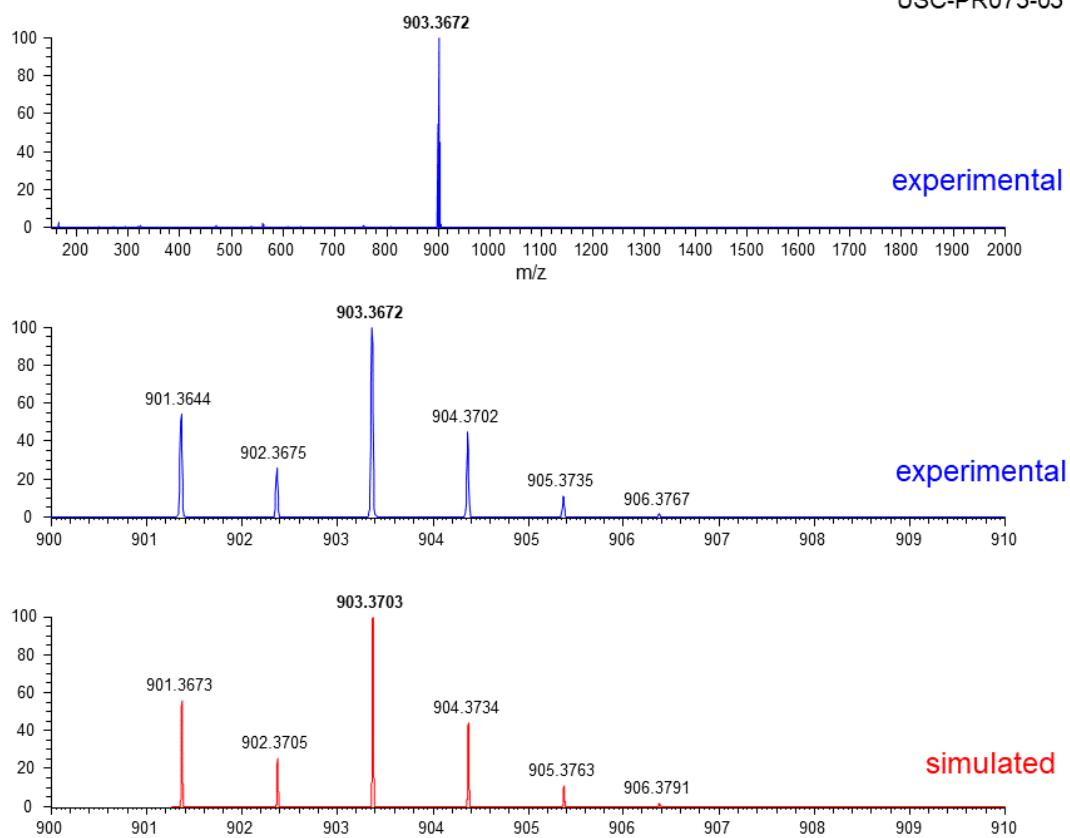


Spectroscopic data for $[\text{Re}(\text{bpn}^{\text{cholamid}}-\kappa^3\text{N})(\text{CO})_3]\text{Br}$ 10





USC-PR073-03



References

- Y.-H. Chiu and J. W. Canary, *Inorg. Chem.*, 2003, **42**, 5107.