

This is a repository copy of *Neutrophils as Orchestrators in Tumor Development and Metastasis Formation*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/175020/>

Version: Published Version

Article:

Kalafati, Lydia, Mitroulis, Ioannis, Verginis, Panayotis et al. (2 more authors) (2020) Neutrophils as Orchestrators in Tumor Development and Metastasis Formation. *Frontiers in Oncology*. p. 581457. ISSN: 2234-943X

<https://doi.org/10.3389/fonc.2020.581457>

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:

<https://creativecommons.org/licenses/>

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Neutrophils as Orchestrators in Tumor Development and Metastasis Formation

Lydia Kalafati^{1,2}, Ioannis Mitroulis^{2,3}, Panayotis Verginis^{4,5}, Triantafyllos Chavakis¹ and Ioannis Kourtzelis^{6*}

¹ Institute for Clinical Chemistry and Laboratory Medicine, University Hospital and Faculty of Medicine, Technische Universität Dresden, Dresden, Germany, ² National Center for Tumor Diseases, Partner Site Dresden and German Cancer Research Center, Heidelberg, Germany, ³ Department of Hematology and Laboratory of Molecular Hematology, Department of Medicine, Democritus University of Thrace, Alexandroupolis, Greece, ⁴ University of Crete, School of Medicine, Heraklion, Greece, ⁵ Institute of Molecular Biology and Biotechnology, Foundation for Research and Technology-Hellas, Heraklion, Crete, Greece, ⁶ York Biomedical Research Institute, Hull York Medical School, University of York, York, United Kingdom

OPEN ACCESS

Edited by:

Panagiota S. Filippou,
Teesside University, United Kingdom

Reviewed by:

Cecilia Garlanda,
Humanitas University, Italy
Jadwiga Jablonska,
Essen University Hospital, Germany

*Correspondence:

Ioannis Kourtzelis
ioannis.kourtzelis@york.ac.uk

Specialty section:

This article was submitted to
Molecular and Cellular Oncology,
a section of the journal
Frontiers in Oncology

Received: 08 July 2020

Accepted: 10 November 2020

Published: 10 December 2020

Citation:

Kalafati L, Mitroulis I, Verginis P,
Chavakis T and Kourtzelis I (2020)
Neutrophils as Orchestrators
in Tumor Development and
Metastasis Formation.
Front. Oncol. 10:581457.
doi: 10.3389/fonc.2020.581457

Several lines of clinical and experimental evidence suggest that immune cell plasticity is a central player in tumorigenesis, tumor progression, and metastasis formation. Neutrophils are able to promote or inhibit tumor growth. Through their interaction with tumor cells or their crosstalk with other immune cell subsets in the tumor microenvironment, they modulate tumor cell survival. Here, we summarize current knowledge with regards to the mechanisms that underlie neutrophil-mediated effects on tumor establishment and metastasis development. We also discuss the tumor-mediated effects on granulopoiesis and neutrophil precursors in the bone marrow and the involvement of neutrophils in anti-tumor therapeutic modalities.

Keywords: neutrophil, tumor, metastasis, immune modulation, cancer-dependent granulopoiesis, trained immunity, innate immune memory, trained granulopoiesis

INTRODUCTION

Neutrophils comprise the majority of leukocytes in humans and are considered the first immune cell population to respond against infectious and inflammatory insults (1–5). This innate immune cell type fine-tunes the armament of host defense through modulating phagocytosis and intracellular killing of pathogens, release of proteases and antimicrobial peptides from their granules, as well as formation of neutrophil extracellular traps (NETs) (6–9). In addition, neutrophils mediate interactions between innate and adaptive immunity by shaping antigen presentation (10, 11) and the production of chemokines and cytokines (12, 13). The generation of neutrophils from their myeloid precursors, designated as granulopoiesis, takes place in the bone marrow, where neutrophils accumulate until they are released in the circulation in a timely and tightly controlled process (14–16). Billions of neutrophils are produced daily under steady state conditions (17). However, certain types of stress such as exposure to inflammatory or infectious agents or cancer result in emergency granulopoiesis that induces a rapid increase in neutrophil production (1, 18). Neutrophils may have gained less attention than other immune cells in the study of anti-tumor immunity due to their relatively short lifespan. However, neutrophil survival is much longer than initially thought; they can remain alive for at least 5 days in the

circulation (19). In addition, neutrophils are generated in high numbers daily and recent findings point to substantial neutrophil heterogeneity (20). Recent evidence thus suggests their involvement in shaping of pro-tumor and anti-tumor responses (21). For instance, neutrophils promote the formation of the pre-metastatic niche and neutrophils from mice with early-stage tumors display increased migratory activity compared to neutrophils from tumor-free animals (22). On the other hand, neutrophils with certain phenotypic characteristics have been associated with enhanced tumor suppression. Specifically, a subset of tumor-associated neutrophils from patients diagnosed with early-stage human lung cancer bears antigen-presentation activity thereby facilitating anti-tumor immunity (23). To further support the dual and context-dependent role of neutrophils in tumors, low-density neutrophils have been shown to be more immunosuppressive and to promote cancer progression as compared to high-density neutrophils (24). In addition, neutrophil plasticity and localization at the tumor site depends not only on intrinsic cues, but also on the type and the stage of the tumor (25). Here, we discuss the neutrophil-dependent mechanisms that may affect suppression or progression of primary tumors and establishment of metastasis.

NEUTROPHILS CONTRIBUTE TO TUMOR PROGRESSION

Emerging evidence suggests that neutrophils modulate cancer-associated inflammation. Importantly, inflammation is a hallmark of cancer (26) and represents an essential contributor to the development of many tumors (27). Neutrophils are present in several types of human tumors and neutrophil accumulation in certain tumors is correlated with poor prognosis (27–29). Inflammatory mediators can affect plasticity of tumor-associated neutrophils and polarize them towards either pro-tumor or anti-tumor phenotype (25, 30, 31) (**Figure 1**). Fridlender et al. have shown that blockade of transforming growth factor β (TGF β) signaling leads to increased neutrophil influx in the tumor. More importantly, these infiltrated neutrophils acquire an anti-tumor phenotype suggesting that TGF β polarizes neutrophils toward a pro-tumor phenotype (30). In addition, the pro-tumoral role of neutrophils has been associated with promotion of angiogenesis (32, 33). Tumor expansion requires the development of new blood vessels that ensure sufficient supply of oxygen and nutrients. Tumor-infiltrating neutrophils are a source of matrix metalloprotease 9 (MMP-9) promoting remodeling of extracellular matrix (ECM) and neovascularization (34). Along

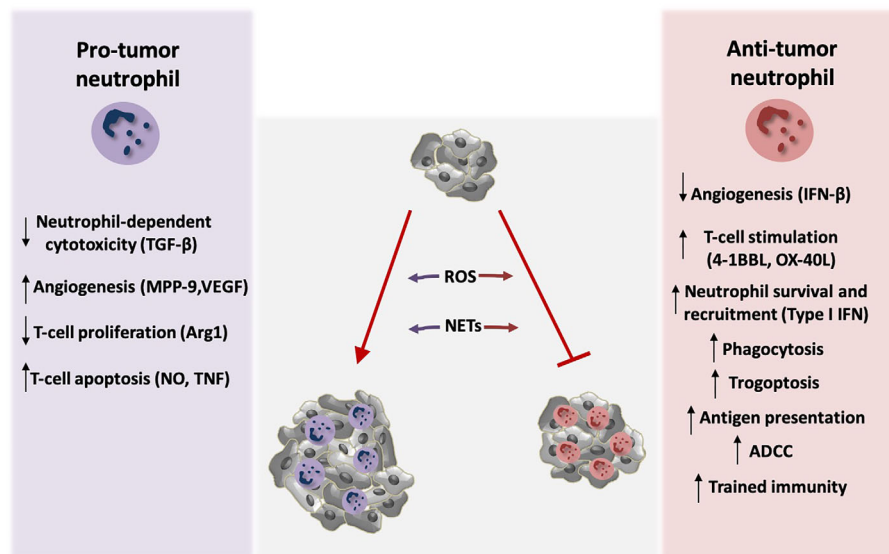


FIGURE 1 | Neutrophil activity modulates tumor growth. Neutrophils exert both tumor-promoting and tumor suppressive functions. TGF- β signaling induces polarization of neutrophils towards pro-tumor phenotype, by blocking direct neutrophil-dependent tumor cell killing. Activation of tumor angiogenesis is stimulated by the production of the neutrophil-derived pro-angiogenic factors MMP-9 and VEGF, whereas endogenous IFN- β downregulates these factors resulting in inhibition of angiogenesis. Neutrophils also modulate anti-tumor T-cell responses. Arginase 1 (Arg1) secretion by neutrophils inhibits T-cell proliferation. Moreover, nitric oxide (NO), and tumor necrosis factor (TNF) derived by neutrophils induce T-cell apoptosis. On the contrary, neutrophils activate T-cell proliferation and anti-tumor function through the production of co-stimulatory molecules such as 4-1BBL and OX-40L. Type I IFN signaling induces neutrophil-mediated tumor suppression by increasing their survival and recruitment in the tumor. Neutrophils can also kill tumor cells directly via antibody dependent cell-mediated cytotoxicity (ADCC) and trogoptosis. In addition, they phagocytose tumor cells and mediate antigen presentation resulting in enhanced anti-tumor immunity. Induction of trained immunity has also been described to promote neutrophil-dependent tumor suppression. ROS production and formation of NETs by neutrophils play a dual role in tumor expansion in a context-dependent manner. Specifically, neutrophils produce ROS that leads to genetic instability and carcinogenesis. On the other hand, ROS can mediate tumor cell killing. NETs contain MMP-9, cathepsin G, and neutrophil elastase that promote tumor growth, but in parallel NET formation primes T-cells and leads to enhanced anti-tumor responses.

this line, reduction of tumor-associated angiogenesis was observed after neutrophil depletion (35). Neutrophils also produce the major pro-angiogenic factor vascular endothelial growth factor (VEGF) regulating tumor-associated angiogenesis (36).

The immunosuppressive function of neutrophils may also contribute to their tumor-promoting function. Neutrophils mediate the suppression of CD8⁺ T cell proliferation and activation by inducing their apoptosis in a manner dependent on nitric oxide and TNF production (37). Furthermore, upregulation of arginase 1 in neutrophils inhibits T-cell proliferation (36, 38) thereby promoting immunosuppression and tumor evasion. Consistently, neutrophil depletion in a mouse model of lung cancer resulted in increased CD8⁺ T cell activation and in decreased tumor burden (30). Furthermore, neutrophils exert their protumorigenic activity by releasing oxygen and nitrogen free radicals that promote genetic instability and carcinogenesis (39–41).

Neutrophils are able to generate neutrophil extracellular traps (NETs). These structures contain extracellular fibers composed of chromatin, histones, and other proteins (42, 43). Except from their established role in host-pathogen interactions (42), NETs modulate cancer-associated procoagulant activity (44) and promote tumor growth (45, 46) by including tumor-promoting components such as MMP-9, cathepsin G (47) and neutrophil elastase (45, 46). In addition, presence of NETs in patients diagnosed with cancer has been associated with poor prognosis (48) and blockade of IL17-mediated NET generation resulted in increased responsiveness to immune checkpoint blockade in pancreatic ductal adenocarcinoma (49).

TUMOR-SUPPRESSIVE ACTIVITY OF NEUTROPHILS

Besides their pro-tumorigenic role, neutrophils can function as tumor suppressors boosting anti-tumor activity (**Figure 1**). Neutrophils have the capacity to generate reactive oxygen species (ROS) by the NADPH oxidase complex and mediate anti-tumor responses (50–52). In an autochthonous mouse tumor model, tumor oxygenation levels differentially affected neutrophil function, and inhibition of tumor hypoxia was associated with enhanced neutrophil dependent-tumor cell killing as a result of ROS production (53).

Antibody dependent cell-mediated cytotoxicity (ADCC) represents another way by which neutrophils may kill tumor cells. In particular, neutrophils express several Fc receptors (FcRs), such as FcγRI (CD64), FcγRIIa (CD32), FcγRIIIa (CD16a), and FcγRIIIb (CD16b) that recognize tumor cell-specific antibodies and mediate ADCC (52, 54, 55). In addition, neutrophil phagocytosis of opsonized tumor cells enhances anti-tumor activity (56) as shown with human tumor cells (57). Neutrophil trogoptosis has been described to exert tumor suppressive activity (58). Specifically, neutrophils target and destroy tumor cells that are opsonized with therapeutic monoclonal antibodies in a process that involves tumor cell lysis mediated by trogocytosis (58–60).

Type I interferons contribute to the anti-tumor effects of neutrophils. Endogenous interferon-β (IFN-β) has been shown to inhibit angiogenesis by downregulating the proangiogenic factors VEGF and MMP-9 in tumor-infiltrating neutrophils (61). Consistently, type I IFN signaling mediates neutrophil-dependent anti-tumor activity by modulating neutrophil survival and recruitment into the tumor (62, 63). Furthermore, neutrophils contribute to the activation of the IFN-γ pathway that enhances anti-tumor activity mediated by the activity of CD4⁺CD8⁺ unconventional αβ T-cells. In agreement with these findings, neutrophil infiltration in certain types of tumors was linked to better clinical outcome (64). Interestingly, NET formation has been also associated with inhibition in tumor growth. Specifically, NETs prime T-cells and play potential role in cancer immunoediting and enhancement of antitumor responses (48, 52).

Up-regulation of antigen presentation can mediate neutrophil-dependent anti-tumor activity. Beauvillain et al. have shown that neutrophils process and present antigens to T-cells (65), thereby enhancing T-cell mediated antitumor responses (66, 67). Along the same line, a subset of neutrophils from patients diagnosed with early-stage human lung cancer has exhibited up-regulated antigen-presenting activity. This neutrophil subpopulation originates from specific bone marrow progenitors upon exposure to IFNγ and GM-CSF signaling (23). Neutrophils can additionally promote T-cell responses *via* the production of the co-stimulatory molecules 4-1BBL and OX-40L, which enhance proliferation and activation of CD4⁺ and CD8⁺ T-cells and increase their cytotoxic capacity in a model of lung cancer (68).

NEUTROPHILS MODULATE METASTATIC DISSEMINATION OF CANCER CELLS

Detachment and escape of tumor cells from the primary tumor represents the initial step of metastasis that is followed by intravasation into the blood and lymphatic system, extravasation, and colonization of tumor cells to distant organs or draining lymph nodes (69). Metastasis is associated with increased mortality (70, 71). Neutrophils affect not only the growth of primary tumors but also orchestrate the metastatic potential of cancer cells (72, 73). Specifically, large body of evidence supports that neutrophils contribute to the initiation phase of metastatic dissemination (69, 74) (**Figure 2** and **Supplementary Table 1**). In addition, primary tumor growth has been associated with accumulation of neutrophils in distant organs before the arrival of the disseminated tumor cells to the site designated as premetastatic niche (75–77). Along the same line, primary tumor cells release factors that render distinct sites more prone to become metastatic sites. The accumulation of neutrophils at these sites is dependent on the growth factor granulocyte-colony stimulating factor (G-CSF) in several tumor models (75, 76, 78). G-CSF promotes the pro-metastatic phenotype of neutrophils by inducing BV8 expression in neutrophils (75, 79) that in turn enhances angiogenesis and

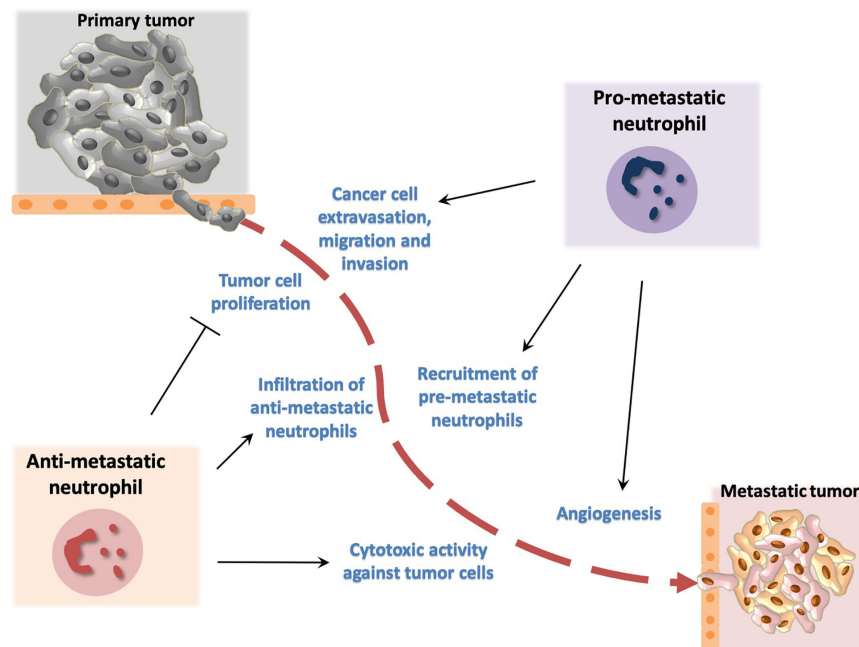


FIGURE 2 | Dual role of neutrophils in metastasis development. Neutrophils promote metastasis by facilitating cancer cell extravasation from primary tumor, migration to the metastatic site and invasion in secondary tumors. In addition, neutrophils promote angiogenesis that has been associated with development of metastasis. Neutrophil activity has been also implicated in inhibition of metastasis. In particular, neutrophils have been described to block cancer cell proliferation and to exert cytotoxic activity thereby affecting tumor cell survival.

cancer cell migration (80, 81). Additionally, neutrophils cooperate with $\gamma\delta$ T-cells, in an interleukin-17/G-CSF dependent manner to facilitate breast cancer metastasis. Depletion of neutrophils in an experimental model of metastatic breast cancer in mice led to a decrease of metastatic burden in both lymph nodes and lungs (75). The interaction of neutrophils with endothelial cells also enhances metastasis by facilitating tumor cell extravasation into the circulation (82–86).

Oncostatin M functions as a pro-angiogenic factor that promotes metastasis. In particular, co-culture experiments have demonstrated that exposure of neutrophils to granulocyte-macrophage colony stimulating factor (GM-CSF) results in production of oncostatin M that contributes to metastasis in a model of breast cancer (87). The glycosaminoglycan hyaluronan, a major component of the ECM that is produced by various types of tumor cells, activates neutrophils through TLR4 signaling and promotes malignant cell migration (88). Furthermore, macrophage migration inhibitory factor (MIF) promotes neutrophil chemotaxis that in turn leads to increased migratory capacity of tumor cells in an *in vitro* model of head and neck cancer (89). Another *in vitro* study using a model of renal cell carcinoma revealed higher recruitment of neutrophils towards tumor cells that was associated with enhanced cancer cell migration and invasion in a manner dependent on a VEGF/hypoxia inducible factor 2a signaling (90). A metastasis-promoting role for neutrophils has been observed in a model of bladder cancer, in which infiltrating neutrophils contribute to cancer cell invasion *via* mediating an upregulation of androgen receptor signals (91).

Besides their impact on primary tumor growth, NETs may contribute to metastasis and blockade of NET formation resulted in decreased tumor metastasis in mice (92, 93).

In agreement with the plasticity and context-specific phenotype of neutrophils, some neutrophil depletion studies have resulted in increased incidence of metastasis (94). Specifically, the chemokine CCL2 has been shown to promote activation of neutrophils towards an anti-metastatic phenotype in a mouse model of lung metastasis. These neutrophils acquired tumor cytotoxic activity that was mediated by production of ROS and H_2O_2 (94). Along the same line, the proto-oncogene MET has been linked to neutrophil-dependent inhibition of primary tumor growth and metastasis formation. Deletion of *Met* in neutrophils resulted in decreased neutrophil infiltration and nitric oxide-dependent tumor cell killing and reduced metastasis (95). In addition, thrombospondin 1 (Tsp1) that is derived by $Gr1^+$ bone marrow myeloid cells may restrain metastasis. Consistently, deficiency in proteases that mediate Tsp1 degradation was associated with decreased metastasis dissemination (96).

THE TUMOR-RELATED IMPACT ON NEUTROPHIL PRODUCTION IN THE BONE MARROW

Aberrant myelopoiesis is a hallmark of cancer (97). Tumor-associated inflammation reprograms hematopoiesis in the bone

marrow *via* acting on hematopoietic stem and progenitor cells (HSPCs) (98). These cells are responsible for the maintenance of hematopoiesis and give rise to all hematopoietic cells through several steps of differentiation (14). Inflammatory stimuli, including those associated with cancer, activate HSPCs, thus promoting their proliferation and myeloid cell priming (16, 99). Studies in patients with cancer (100) and tumor-bearing mice (101) have demonstrated that the tumor environment drives a myeloid bias in HSPCs resulting in enhanced production of cells of the myeloid lineage, at the expense of cells of lymphoid lineage, which has been correlated with disease prognosis in different types of malignancy (102, 103). Circulating hematopoietic progenitor subsets were increased in patients with cancer compared to age-matched healthy subjects (100). Interestingly, the same study has reported enhanced frequency of granulocyte-macrophage myeloid progenitors (GMPs) in the circulation of patients with cancer, further suggesting the myeloid priming of hematopoiesis (100). Increased frequency of myeloid-biased HSPCs residing in the spleen has also been reported in tumor-bearing mice (101). These cells were responsive to the myelopoiesis-driving growth factor GM-CSF, which resulted in production of myeloid cells with pro-tumorigenic properties (101).

Further studies have implicated the myeloid lineage growth factors GM-CSF and G-CSF in the generation of increased numbers of neutrophils in cancer. In a mouse model of invasive breast carcinoma, tumor cell-derived G-CSF can activate bone marrow hematopoietic progenitors, driving myeloid differentiation, and production of neutrophils with T-cell suppressive properties (78). G-CSF can induce mobilization of granulocytes from the bone marrow, which in turn results in their accumulation to distal tissues, supporting metastasis (76). A study in a mouse model of pancreatic ductal adenocarcinoma has shown that tumor-derived GM-CSF regulates the generation of immunosuppressive Gr1⁺CD11b⁺ myeloid cells (104). Mutations in the oncogenic gene KRAS were shown to drive the increased production of GM-CSF by pancreatic ductal endothelial cells, thus further fueling myelopoiesis (105). Except from the myeloid lineage growth factors, TNF supports tumor-associated aberrant myelopoiesis. TNF released by activated CD4⁺ T cells in tumor-bearing mice drives emergency myelopoiesis and generation of both monocytic and granulocytic myeloid cells with immunosuppressive properties (106).

Recent studies have identified unipotent neutrophil precursors that expand in the bone marrow and circulation of tumor-bearing mice (107, 108). These neutrophil precursors have immunosuppressive and tumor-promoting characteristics, as shown in a mouse melanoma model (107). Such circulating neutrophil precursors were also identified in patients with melanoma (107). Using a xenograft osteosarcoma model, it was demonstrated that these neutrophil precursors promote tumor growth (107). Taken together, cancer is associated with aberrant myelopoiesis, which usually results in the generation of neutrophil precursors and neutrophils with tumor-promoting potential.

NEUTROPHIL-ASSOCIATED ANTI-TUMOR THERAPEUTIC APPROACHES

Given their involvement in the shaping of pro-tumor or anti-tumor activity, neutrophils may serve as a therapeutic target in the context of tumor progression. For instance, blockade of the recruitment of pro-tumorigenic neutrophils into tumor may represent a promising strategy against tumor expansion (109). Along the same line, administration of a neutralizing antibody against the neutrophil chemokine interleukin 8 (IL-8) that can be secreted by tumor cells resulted in decreased primary tumor growth and metastasis in models of melanoma and lung cancer (110). Inhibition of the CXC chemokine receptor 2 (CXCR2), a major receptor for IL-8, also led to decreased neutrophil presence in tumors and was associated with tumor suppression (111). Additionally, blockade of CXCR2 demonstrated anti-metastatic effect and led to increased efficacy of either immunotherapy in a model of pancreatic ductal adenocarcinoma (112) or chemotherapy in breast carcinoma (113). Neutrophil depletion led to increased sensitivity to radiation therapy in a mouse model of sarcoma (114). The ratio of CD8⁺ T-cells to neutrophils within the tumor of patients with non-small cell lung cancer has been suggested as a marker indicative of immune checkpoint inhibitor efficacy (115). In addition, accumulation of Gr1⁺CD11b⁺ cells that is mediated by G-CSF-induced mobilization (116, 117) or not (118) was associated with decreased tumor responsiveness after therapeutic inhibition of angiogenesis.

On the other hand, there are reports suggesting a beneficial impact of neutrophils by promoting tumor elimination. Neutrophils were shown to mediate T-cell anti-tumor activity in early stages of human lung cancer (68). Additionally, neutrophils derived from healthy donors have demonstrated tumor cell killing potential (119). Combination of radiation therapy with G-CSF administration has also resulted in neutrophil-dependent anti-tumor immunity as shown in syngeneic mouse tumor models (120).

Manipulation of the phenotype of tumor-associated neutrophils can be exploited as a potential anti-tumor therapeutic approach. Inhibition of TGF β signaling promotes reprogramming of tumor-associated neutrophils, shifting their actions from pro-tumor to anti-tumor (30). Deficiency of TGF β signaling in myeloid cells has also resulted in inhibition of metastasis that was associated with enhanced anti-tumor immunity (121). Additionally, priming of tumor-associated neutrophils with IFN γ and TNF contributed to alterations in the polarization of neutrophils rendering them from tumor promoters to tumor suppressors (122).

Moreover, recent evidence suggests that trained immunity may confer anti-tumor properties in neutrophils. Trained immunity represents memory of the innate immune system (123). In particular, exposure of innate immune cells to certain stimuli, such as the microbial component β -glucan or the Bacillus Calmette–Guérin vaccine leads to enhanced responsiveness to subsequent homologous or heterologous triggers (123, 124). Epigenetic reprogramming of myeloid cells

and their progenitors in the bone marrow represent major components of innate immune memory (125–127). Trained innate immunity may boost neutrophil-dependent tumor suppression. Specifically, β -glucan-induced trained immunity led to epigenetic reprogramming of granulopoiesis towards generation of neutrophils with an anti-tumor phenotype. Trained granulopoiesis was mediated by type I IFN signaling, while the tumor suppressive activity of “trained” neutrophils was mediated by enhanced ROS production. The therapeutic potential of ‘trained’ neutrophils was confirmed by the decreased tumor growth in mice that received neutrophils from β -glucan-treated donor mice (128).

CONCLUDING REMARKS

Myeloid cells play an important role in the modulation of tumor growth. Neutrophils not only respond against pathogens and inflammatory stimuli, but also orchestrate tumor-associated immune responses. Different polarization signals can affect neutrophil plasticity and in turn lead to either promotion or suppression of primary tumors or metastasis. Neutrophils can affect cancer progression by interacting directly with tumor cells or indirectly with other immune cell types. Importantly, tumor-associated inflammation has a substantial impact in neutrophil production in the bone marrow that is a key determinant in tumor growth.

REFERENCES

- Chavakis T, Mitroulis I, Hajishengallis G. Hematopoietic progenitor cells as integrative hubs for adaptation to and fine-tuning of inflammation. *Nat Immunol* (2019) 20:802–11. doi: 10.1038/s41590-019-0402-5
- Kourtzelis I, Mitroulis I, von Renesse J, Hajishengallis G, Chavakis T. From leukocyte recruitment to resolution of inflammation: the cardinal role of integrins. *J Leukoc Biol* (2017) 102:677–83. doi: 10.1189/jlb.3MR0117-024R
- Mitroulis I, Alexaki VI, Kourtzelis I, Ziogas A, Hajishengallis G, Chavakis T. Leukocyte integrins: role in leukocyte recruitment and as therapeutic targets in inflammatory disease. *Pharmacol Ther* (2015) 147:123–35. doi: 10.1016/j.pharmthera.2014.11.008
- Kubes P. The enigmatic neutrophil: what we do not know. *Cell Tissue Res* (2018) 371:399–406. doi: 10.1007/s00441-018-2790-5
- Phillipson M, Kubes P. The Healing Power of Neutrophils. *Trends Immunol* (2019) 40:635–47. doi: 10.1016/j.it.2019.05.001
- Kruger P, Saffarzadeh M, Weber AN, Rieber N, Radsak M, von Bernuth H, et al. Neutrophils: Between host defence, immune modulation, and tissue injury. *PLoS Pathog* (2015) 11:e1004651. doi: 10.1371/journal.ppat.1004651
- Nauseef WM, Borregaard N. Neutrophils at work. *Nat Immunol* (2014) 15:602–11. doi: 10.1038/ni.2921
- Kolaczowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nat Rev Immunol* (2013) 13:159–75. doi: 10.1038/nri3399
- Cowland JB, Borregaard N. Granulopoiesis and granules of human neutrophils. *Immunol Rev* (2016) 273:11–28. doi: 10.1111/imr.12440
- Vono M, Lin A, Norrby-Teglund A, Koup RA, Liang F, Lore K. Neutrophils acquire the capacity for antigen presentation to memory CD4(+) T cells in vitro and ex vivo. *Blood* (2017) 129:1991–2001. doi: 10.1182/blood-2016-10-744441
- Meindert SM, Baker G, van Wijk S, Beuger BM, Geissler J, Jansen MH, et al. Neutrophils acquire antigen-presenting cell features after phagocytosis of IgG-opsonized erythrocytes. *Blood Adv* (2019) 3:1761–73. doi: 10.1182/bloodadvances.2018028753
- Tecchio C, Cassatella MA. Neutrophil-derived chemokines on the road to immunity. *Semin Immunol* (2016) 28:119–28. doi: 10.1016/j.smim.2016.04.003
- Tecchio C, Scapini P, Pizzolo G, Cassatella MA. On the cytokines produced by human neutrophils in tumors. *Semin Cancer Biol* (2013) 23:159–70. doi: 10.1016/j.semcancer.2013.02.004
- Mitroulis I, Kalafati L, Hajishengallis G, Chavakis T. Myelopoiesis in the Context of Innate Immunity. *J Innate Immun* (2018) 10:365–72. doi: 10.1159/000489406
- Mitroulis I, Chen LS, Singh RP, Kourtzelis I, Economopoulou M, Kajikawa T, et al. Secreted protein Del-1 regulates myelopoiesis in the hematopoietic stem cell niche. *J Clin Invest* (2017) 127:3624–39. doi: 10.1172/JCI92571
- Mitroulis I, Kalafati L, Bornhauser M, Hajishengallis G, Chavakis T. Regulation of the Bone Marrow Niche by Inflammation. *Front Immunol* (2020) 11:1540. doi: 10.3389/fimmu.2020.01540
- Yvan-Charvet L, Ng LG. Granulopoiesis and Neutrophil Homeostasis: A Metabolic, Daily Balancing Act. *Trends Immunol* (2019) 40:598–612. doi: 10.1016/j.it.2019.05.004
- Manz MG, Boettcher S. Emergency granulopoiesis. *Nat Rev Immunol* (2014) 14:302–14. doi: 10.1038/nri3660
- Pillay J, den Braber I, Vriskoop N, Kwast LM, de Boer RJ, Borghans JA, et al. In vivo labeling with $^2\text{H}_2\text{O}$ reveals a human neutrophil lifespan of 5.4 days. *Blood* (2010) 116:625–7. doi: 10.1182/blood-2010-01-259028
- Rubio-Ponce A, Hidalgo A, Ballesteros I. How to bridle a neutrophil. *Curr Opin Immunol* (2020) 68:41–7. doi: 10.1016/j.coi.2020.08.003
- Kim J, Bae JS. Tumor-Associated Macrophages and Neutrophils in Tumor Microenvironment. *Mediators Inflamm* (2016) 2016:6058147. doi: 10.1155/2016/6058147
- Patel S, Fu S, Mastio J, Dominguez GA, Purohit A, Kossenkova A, et al. Unique pattern of neutrophil migration and function during tumor

AUTHOR CONTRIBUTIONS

LK, IM, PV, TC, and IK have made substantial, direct, and intellectual contribution to the manuscript. All authors contributed to the article and approved the submitted version.

FUNDING

IK is supported by a start-up funding from the Hull York Medical School of University of York and the Royal Society research grant RGS\R2\202032.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fonc.2020.581457/full#supplementary-material>

SUPPLEMENTARY TABLE 1 | The impact of neutrophil activity on metastasis formation.

- progression. *Nat Immunol* (2018) 19:1236–47. doi: 10.1038/s41590-018-0229-5
23. Singhal S, Bhojnagarwala PS, O'Brien S, Moon EK, Garfall AL, Rao AS, et al. Origin and Role of a Subset of Tumor-Associated Neutrophils with Antigen-Presenting Cell Features in Early-Stage Human Lung Cancer. *Cancer Cell* (2016) 30:120–35. doi: 10.1016/j.ccell.2016.06.001
 24. Sagiv JY, Michaeli J, Assi S, Mishalian I, Kisos H, Levy L, et al. Phenotypic diversity and plasticity in circulating neutrophil subpopulations in cancer. *Cell Rep* (2015) 10:562–73. doi: 10.1016/j.celrep.2014.12.039
 25. Mishalian I, Bayuh R, Levy L, Zolotarov L, Michaeli J, Fridlender ZG. Tumor-associated neutrophils (TAN) develop pro-tumorigenic properties during tumor progression. *Cancer Immunol Immunother* (2013) 62:1745–56. doi: 10.1007/s00262-013-1476-9
 26. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013
 27. Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature* (2008) 454:436–44. doi: 10.1038/nature07205
 28. Bodey B, Bodey BJr., Siegel SE, Luck JV, Kaiser HE. Immunophenotypic characterization of human primary and metastatic melanoma infiltrating leukocytes. *Anticancer Res* (1996) 16:3439–46.
 29. Caruso RA, Bellocchio R, Pagano M, Bertoli G, Rigoli L, Inferriera C. Prognostic value of intratumoral neutrophils in advanced gastric carcinoma in a high-risk area in northern Italy. *Mod Pathol* (2002) 15:831–7. doi: 10.1097/01.MP.0000020391.98998.6B
 30. Fridlender ZG, Sun J, Kim S, Kapoor V, Cheng G, Ling L, et al. Polarization of tumor-associated neutrophil phenotype by TGF-beta: "N1" versus "N2" TAN. *Cancer Cell* (2009) 16:183–94. doi: 10.1016/j.ccr.2009.06.017
 31. Fridlender ZG, Albelda SM. Tumor-associated neutrophils: friend or foe? *Carcinogenesis* (2012) 33:949–55. doi: 10.1093/carcin/bgs123
 32. Tazzyman S, Lewis CE, Murdoch C. Neutrophils: key mediators of tumour angiogenesis. *Int J Exp Pathol* (2009) 90:222–31. doi: 10.1111/j.1365-2613.2009.00641.x
 33. Seignez C, Phillipson M. The multitasking neutrophils and their involvement in angiogenesis. *Curr Opin Hematol* (2017) 24:3–8. doi: 10.1097/MOH.0000000000000300
 34. Nozawa H, Chiu C, Hanahan D. Infiltrating neutrophils mediate the initial angiogenic switch in a mouse model of multistage carcinogenesis. *Proc Natl Acad Sci U S A* (2006) 103:12493–8. doi: 10.1073/pnas.0601807103
 35. Benelli R, Morini M, Carrozzino F, Ferrari N, Minghelli S, Santi L, et al. Neutrophils as a key cellular target for angiostatin: implications for regulation of angiogenesis and inflammation. *FASEB J* (2002) 16:267–9. doi: 10.1096/fj.01-0651fje
 36. Kusumanto YH, Dam WA, Hoppers GA, Meijer C, Mulder NH. Platelets and granulocytes, in particular the neutrophils, form important compartments for circulating vascular endothelial growth factor. *Angiogenesis* (2003) 6:283–7. doi: 10.1023/B:AGEN.0000029415.62384.ba
 37. Michaeli J, Shaul ME, Mishalian I, Hovav AH, Levy L, Zolotriov L, et al. Tumor-associated neutrophils induce apoptosis of non-activated CD8 T-cells in a TNFalpha and NO-dependent mechanism, promoting a tumor-supportive environment. *Oncoimmunology* (2017) 6:e1356965. doi: 10.1080/2162402X.2017.1356965
 38. Rotondo R, Barisione G, Mastracci L, Grossi F, Orenzo AM, Costa R, et al. IL-8 induces exocytosis of arginase 1 by neutrophil polymorphonuclears in non-small cell lung cancer. *Int J Cancer* (2009) 125:887–93. doi: 10.1002/ijc.24448
 39. Knaapen AM, Gungor N, Schins RP, Borm PJ, Van Schooten FJ. Neutrophils and respiratory tract DNA damage and mutagenesis: a review. *Mutagenesis* (2006) 21:225–36. doi: 10.1093/mutage/gel032
 40. Gungor N, Knaapen AM, Munia A, Peluso M, Haenen GR, Chiu RK, et al. Genotoxic effects of neutrophils and hypochlorous acid. *Mutagenesis* (2010) 25:149–54. doi: 10.1093/mutage/gep053
 41. Knaapen AM, Schins RP, Polat D, Becker A, Borm PJ. Mechanisms of neutrophil-induced DNA damage in respiratory tract epithelial cells. *Mol Cell Biochem* (2002) 234–235:143–51. doi: 10.1023/A:1015997001657
 42. Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, et al. Neutrophil extracellular traps kill bacteria. *Science* (2004) 303:1532–5. doi: 10.1126/science.1092385
 43. Papayannopoulos V. Neutrophil extracellular traps in immunity and disease. *Nat Rev Immunol* (2018) 18:134–47. doi: 10.1038/nri.2017.105
 44. Demers M, Krause DS, Schatzberg D, Martinod K, Voorhees JR, Fuchs TA, et al. Cancers predispose neutrophils to release extracellular DNA traps that contribute to cancer-associated thrombosis. *Proc Natl Acad Sci U S A* (2012) 109:13076–81. doi: 10.1073/pnas.1200419109
 45. Cools-Lartigue J, Spicer J, Najmeh S, Ferri L. Neutrophil extracellular traps in cancer progression. *Cell Mol Life Sci* (2014) 71:4179–94. doi: 10.1007/s00018-014-1683-3
 46. Albregues J, Shields MA, Ng D, Park CG, Ambrico A, Poindexter ME, et al. Neutrophil extracellular traps produced during inflammation awaken dormant cancer cells in mice. *Science* (2018) 361. doi: 10.1126/science.aao4227
 47. Wilson TJ, Nannuru KC, Futakuchi M, Sadanandam A, Singh RK. Cathepsin G enhances mammary tumor-induced osteolysis by generating soluble receptor activator of nuclear factor-kappaB ligand. *Cancer Res* (2008) 68:5803–11. doi: 10.1158/0008-5472.CAN-07-5889
 48. Berger-Achituv S, Brinkmann V, Abed UA, Kuhn LI, Ben-Ezra J, Elhasid R, et al. A proposed role for neutrophil extracellular traps in cancer immunoediting. *Front Immunol* (2013) 4:48. doi: 10.3389/fimmu.2013.00048
 49. Zhang Y, Chandra V, Riquelme Sanchez E, Dutta P, Quesada PR, Rakoski A, et al. Interleukin-17-induced neutrophil extracellular traps mediate resistance to checkpoint blockade in pancreatic cancer. *J Exp Med* (2020) 217. doi: 10.1084/jem.20190354
 50. Dissemont J, Weimann TK, Schneider LA, Schneeberger A, Scharffetter-Kochanek K, Goos M, et al. Activated neutrophils exert antitumor activity against human melanoma cells: reactive oxygen species-induced mechanisms and their modulation by granulocyte-macrophage-colony-stimulating factor. *J Invest Dermatol* (2003) 121:936–8. doi: 10.1046/j.1523-1747.2003.12475.x
 51. Zivkovic M, Poljak-Blazi M, Egger G, Sunjic SB, Schaur RJ, Zarkovic N. Oxidative burst and anticancer activities of rat neutrophils. *Biofactors* (2005) 24:305–12. doi: 10.1002/biof.5520240136
 52. Clark RA, Klebanoff SJ. Neutrophil-mediated tumor cell cytotoxicity: role of the peroxidase system. *J Exp Med* (1975) 141:1442–7. doi: 10.1084/jem.141.6.1442
 53. Mahidhane K, Blaisdell A, Ma S, Crequer-Grandhomme A, Lowell CA, Erlebacher A. Relief of tumor hypoxia unleashes the tumoricidal potential of neutrophils. *J Clin Invest* (2020) 130:389–403. doi: 10.1172/JCI130952
 54. Valerius T, Repp R, de Wit TP, Berthold S, Platzer E, Kalden JR, et al. Involvement of the high-affinity receptor for IgG (Fc gamma RI; CD64) in enhanced tumor cell cytotoxicity of neutrophils during granulocyte colony-stimulating factor therapy. *Blood* (1993) 82:931–9. doi: 10.1182/blood.V82.3.931.bloodjournal823931
 55. Kushner BH, Cheung NK. Absolute requirement of CD11/CD18 adhesion molecules, FcRII and the phosphatidylinositol-linked FcRIII for monoclonal antibody-mediated neutrophil antitumor cytotoxicity. *Blood* (1992) 79:1484–90. doi: 10.1182/blood.V79.6.1484.bloodjournal7961484
 56. Feng M, Jiang W, Kim BYS, Zhang CC, Fu YX, Weissman IL. Phagocytosis checkpoints as new targets for cancer immunotherapy. *Nat Rev Cancer* (2019) 19:568–86. doi: 10.1038/s41568-019-0183-z
 57. Ring NG, Herndler-Brandstetter D, Weiskopf K, Shan L, Volkmer JP, George BM, et al. Anti-SIRPalpha antibody immunotherapy enhances neutrophil and macrophage antitumor activity. *Proc Natl Acad Sci U S A* (2017) 114:E10578–E85. doi: 10.1073/pnas.1710877114
 58. Matlung HL, Babes L, Zhao XW, van Houdt M, Treffers LW, van Rees DJ, et al. Neutrophils Kill Antibody-Opsonized Cancer Cells by Trogocytosis. *Cell Rep* (2018) 23:3946–59.e6. doi: 10.1016/j.celrep.2018.05.082
 59. Joly E, Hudrisier D. What is trogocytosis and what is its purpose? *Nat Immunol* (2003) 4:815. doi: 10.1038/ni0903-815
 60. Dance A. Core Concept: Cells nibble one another via the under-appreciated process of trogocytosis. *Proc Natl Acad Sci U S A* (2019) 116:17608–10. doi: 10.1073/pnas.1912252116
 61. Jablonska J, Leschner S, Westphal K, Lienenklaus S, Weiss S. Neutrophils responsive to endogenous IFN-beta regulate tumor angiogenesis and growth in a mouse tumor model. *J Clin Invest* (2010) 120:1151–64. doi: 10.1172/JCI37223
 62. Andzinski L, Wu CF, Lienenklaus S, Kroger A, Weiss S, Jablonska J. Delayed apoptosis of tumor associated neutrophils in the absence of endogenous IFN-beta. *Int J Cancer* (2015) 136:572–83. doi: 10.1002/ijc.28957
 63. Jablonska J, Wu CF, Andzinski L, Leschner S, Weiss S. CXCR2-mediated tumor-associated neutrophil recruitment is regulated by IFN-beta. *Int J Cancer* (2014) 134:1346–58. doi: 10.1002/ijc.28551

64. Ponzetta A, Carriero R, Carnevale S, Barbagallo M, Molgora M, Perucchini C, et al. Neutrophils Driving Unconventional T Cells Mediate Resistance against Murine Sarcomas and Selected Human Tumors. *Cell* (2019) 178:346–60.e24. doi: 10.1016/j.cell.2019.05.047
65. Beauvillain C, Delneste Y, Scotet M, Peres A, Gascan H, Guermontprez P, et al. Neutrophils efficiently cross-prime naive T cells in vivo. *Blood* (2007) 110:2965–73. doi: 10.1182/blood-2006-12-063826
66. Davey MS, Morgan MP, Liuzzi AR, Tyler CJ, Khan MWA, Szakmany T, et al. Microbe-specific unconventional T cells induce human neutrophil differentiation into antigen cross-presenting cells. *J Immunol* (2014) 193:3704–16. doi: 10.4049/jimmunol.1401018
67. Takashima A, Yao Y. Neutrophil plasticity: acquisition of phenotype and functionality of antigen-presenting cell. *J Leukoc Biol* (2015) 98:489–96. doi: 10.1189/jlb.1MR1014-502R
68. Eruslanov EB, Bhojnagarwala PS, Quatromoni JG, Stephen TL, Ranganathan A, Deshpande C, et al. Tumor-associated neutrophils stimulate T cell responses in early-stage human lung cancer. *J Clin Invest* (2014) 124:5466–80. doi: 10.1172/JCI77053
69. Psaila B, Lyden D. The metastatic niche: adapting the foreign soil. *Nat Rev Cancer* (2009) 9:285–93. doi: 10.1038/nrc2621
70. Mehlen P, Puisieux A. Metastasis: a question of life or death. *Nat Rev Cancer* (2006) 6:449–58. doi: 10.1038/nrc1886
71. Dillekas H, Rogers MS, Straume O. Are 90% of deaths from cancer caused by metastases? *Cancer Med* (2019) 8:5574–76. doi: 10.1002/cam4.2474
72. Neutrophils Help Circulating Tumor Cells Metastasize. *Cancer Discov* (2019) 9:458–59. doi: 10.1158/2159-8290.CD-NB2019-025
73. Tuting T, de Visser KE. CANCER. How neutrophils promote metastasis. *Science* (2016) 352:145–6. doi: 10.1126/science.aaf7300
74. Liang W, Ferrara N. The Complex Role of Neutrophils in Tumor Angiogenesis and Metastasis. *Cancer Immunol Res* (2016) 4:83–91. doi: 10.1158/2326-6066.CIR-15-0313
75. Coffelt SB, Kersten K, Doornebal CW, Weiden J, Vrijland K, Hau CS, et al. IL-17-producing gamma delta T cells and neutrophils conspire to promote breast cancer metastasis. *Nature* (2015) 522:345–48. doi: 10.1038/nature14282
76. Kowanetz M, Wu X, Lee J, Tan M, Hagenbeek T, Qu X, et al. Granulocyte-colony stimulating factor promotes lung metastasis through mobilization of Ly6G+Ly6C+ granulocytes. *Proc Natl Acad Sci U S A* (2010) 107:21248–55. doi: 10.1073/pnas.1015855107
77. Wculek SK, Malanchi I. Neutrophils support lung colonization of metastasis-initiating breast cancer cells. *Nature* (2015) 528:413–7. doi: 10.1038/nature16140
78. Casbon AJ, Reynaud D, Park C, Khuc E, Gan DD, Schepers K, et al. Invasive breast cancer reprograms early myeloid differentiation in the bone marrow to generate immunosuppressive neutrophils. *Proc Natl Acad Sci U S A* (2015) 112:E566–75. doi: 10.1073/pnas.1424927112
79. Qu X, Zhuang G, Yu L, Meng G, Ferrara N. Induction of Bv8 expression by granulocyte colony-stimulating factor in CD11b+Gr1+ cells: key role of Stat3 signaling. *J Biol Chem* (2012) 287:19574–84. doi: 10.1074/jbc.M111.326801
80. Shojaei F, Singh M, Thompson JD, Ferrara N. Role of Bv8 in neutrophil-dependent angiogenesis in a transgenic model of cancer progression. *Proc Natl Acad Sci U S A* (2008) 105:2640–5. doi: 10.1073/pnas.0712185105
81. Shojaei F, Wu X, Zhong C, Yu L, Liang XH, Yao J, et al. Bv8 regulates myeloid-cell-dependent tumour angiogenesis. *Nature* (2007) 450:825–31. doi: 10.1038/nature06348
82. Bald T, Quast T, Landsberg J, Rogava M, Glodde N, Lopez-Ramos D, et al. Ultraviolet-radiation-induced inflammation promotes angiogenesis and metastasis in melanoma. *Nature* (2014) 507:109–13. doi: 10.1038/nature13111
83. Antonio N, Bonnelykke-Behrndtz ML, Ward LC, Collin J, Christensen IJ, Steiniche T, et al. The wound inflammatory response exacerbates growth of pre-neoplastic cells and progression to cancer. *EMBO J* (2015) 34:2219–36. doi: 10.15252/embj.201490147
84. Wu QD, Wang JH, Condon C, Bouchier-Hayes D, Redmond HP. Human neutrophils facilitate tumor cell transendothelial migration. *Am J Physiol Cell Physiol* (2001) 280:C814–22. doi: 10.1152/ajpcell.2001.280.4.C814
85. Strell C, Lang K, Niggemann B, Zaenker KS, Entschladen F. Neutrophil granulocytes promote the migratory activity of MDA-MB-468 human breast carcinoma cells via ICAM-1. *Exp Cell Res* (2010) 316:138–48. doi: 10.1016/j.yexcr.2009.09.003
86. Huh SJ, Liang S, Sharma A, Dong C, Robertson GP. Transiently entrapped circulating tumor cells interact with neutrophils to facilitate lung metastasis development. *Cancer Res* (2010) 70:6071–82. doi: 10.1158/0008-5472.CAN-09-4442
87. Queen MM, Ryan RE, Holzer RG, Keller-Peck CR, Jorczyk CL. Breast cancer cells stimulate neutrophils to produce oncostatin M: potential implications for tumor progression. *Cancer Res* (2005) 65:8896–904. doi: 10.1158/0008-5472.CAN-05-1734
88. Wu Y, Zhao Q, Peng C, Sun L, Li XF, Kuang DM. Neutrophils promote motility of cancer cells via a hyaluronan-mediated TLR4/PI3K activation loop. *J Pathol* (2011) 225:438–47. doi: 10.1002/path.2947
89. Dumitru CA, Gholaman H, Trellakis S, Bruderek K, Dominas N, Gu X, et al. Tumor-derived macrophage migration inhibitory factor modulates the biology of head and neck cancer cells via neutrophil activation. *Int J Cancer* (2011) 129:859–69. doi: 10.1002/ijc.25991
90. Song W, Yeh CR, He D, Wang Y, Xie H, Pang ST, et al. Infiltrating neutrophils promote renal cell carcinoma progression via VEGFa/HIF2alpha and estrogen receptor beta signals. *Oncotarget* (2015) 6:19290–304. doi: 10.18632/oncotarget.4478
91. Lin C, Lin W, Yeh S, Li L, Chang C. Infiltrating neutrophils increase bladder cancer cell invasion via modulation of androgen receptor (AR)/MMP13 signals. *Oncotarget* (2015) 6:43081–9. doi: 10.18632/oncotarget.5638
92. Cools-Lartigue J, Spicer J, McDonald B, Gowing S, Chow S, Giannias B, et al. Neutrophil extracellular traps sequester circulating tumor cells and promote metastasis. *J Clin Invest* (2013) 123:3446–58. doi: 10.1172/JCI67484
93. Yang L, Liu Q, Zhang X, Liu X, Zhou B, Chen J, et al. DNA of neutrophil extracellular traps promotes cancer metastasis via CCDC25. *Nature* (2020) 583:133–8. doi: 10.1038/s41586-020-2394-6
94. Granot Z, Henke E, Comen EA, King TA, Norton L, Benezra R. Tumor entrained neutrophils inhibit seeding in the premetastatic lung. *Cancer Cell* (2011) 20:300–14. doi: 10.1016/j.ccr.2011.08.012
95. Finisguerra V, Di Conza G, Di Matteo M, Serneels J, Costa S, Thompson AA, et al. MET is required for the recruitment of anti-tumoural neutrophils. *Nature* (2015) 522:349–53. doi: 10.1038/nature14407
96. El Rayes T, Catena R, Lee S, Stawowczyk M, Joshi N, Fischbach C, et al. Lung inflammation promotes metastasis through neutrophil protease-mediated degradation of Tsp-1. *Proc Natl Acad Sci U S A* (2015) 112:16000–5. doi: 10.1073/pnas.1507294112
97. Schupp J, Krebs FK, Zimmer N, Trzeciak E, Schuppan D, Tuettenberg A. Targeting myeloid cells in the tumor sustaining microenvironment. *Cell Immunol* (2019) 343:103713. doi: 10.1016/j.cellimm.2017.10.013
98. McAllister SS, Weinberg RA. The tumour-induced systemic environment as a critical regulator of cancer progression and metastasis. *Nat Cell Biol* (2014) 16:717–27. doi: 10.1038/ncb3015
99. Messmer MN, Netherby CS, Banik D, Abrams SI. Tumor-induced myeloid dysfunction and its implications for cancer immunotherapy. *Cancer Immunol Immunother* (2015) 64:1–13. doi: 10.1007/s00262-014-1639-3
100. Wu WC, Sun HW, Chen HT, Liang J, Yu XJ, Wu C, et al. Circulating hematopoietic stem and progenitor cells are myeloid-biased in cancer patients. *Proc Natl Acad Sci U S A* (2014) 111:4221–6. doi: 10.1073/pnas.1320753111
101. Wu C, Ning H, Liu M, Lin J, Luo S, Zhu W, et al. Spleen mediates a distinct hematopoietic progenitor response supporting tumor-promoting myelopoiesis. *J Clin Invest* (2018) 128:3425–38. doi: 10.1172/JCI97973
102. Abu-Shawar O, Abu-Shawar M, Hirmas N, Alhourri A, Massad A, Alsibai B, et al. Hematologic markers of distant metastases and poor prognosis in gynecological cancers. *BMC Cancer* (2019) 19:141. doi: 10.1186/s12885-019-5326-9
103. Zhao G, Liu N, Wang S, Guo J, Song X, Qi Y, et al. Prognostic significance of the neutrophil-to-lymphocyte and platelet-to-lymphocyte ratio in patients with metastatic gastric cancer. *Medicine (Baltimore)* (2020) 99:e19405. doi: 10.1097/MD.00000000000019405
104. Bayne LJ, Beatty GL, Jhala N, Clark CE, Rhim AD, Stanger BZ, et al. Tumor-derived granulocyte-macrophage colony-stimulating factor regulates

- myeloid inflammation and T cell immunity in pancreatic cancer. *Cancer Cell* (2012) 21:822–35. doi: 10.1016/j.ccr.2012.04.025
105. Pylayeva-Gupta Y, Lee KE, Hajdu CH, Miller G, Bar-Sagi D. Oncogenic Kras-induced GM-CSF production promotes the development of pancreatic neoplasia. *Cancer Cell* (2012) 21:836–47. doi: 10.1016/j.ccr.2012.04.024
 106. Al Sayed MF, Amrein MA, Buhner ED, Huguenin AL, Radpour R, Riether C, et al. T-cell-Secreted TNF α Induces Emergency Myelopoiesis and Myeloid-Derived Suppressor Cell Differentiation in Cancer. *Cancer Res* (2019) 79:346–59. doi: 10.1158/0008-5472.CAN-17-3026
 107. Zhu YP, Padgett L, Dinh HQ, Marcovecchio P, Blatchley A, Wu R, et al. Identification of an Early Unipotent Neutrophil Progenitor with Pro-tumoral Activity in Mouse and Human Bone Marrow. *Cell Rep* (2018) 24:2329–41.e8. doi: 10.1016/j.celrep.2018.07.097
 108. Evrard M, Kwok IWH, Chong SZ, Teng KWW, Becht E, Chen J, et al. Developmental Analysis of Bone Marrow Neutrophils Reveals Populations Specialized in Expansion, Trafficking, and Effector Functions. *Immunity* (2018) 48:364–79.e8. doi: 10.1016/j.immuni.2018.02.002
 109. Uribe-Querol E, Rosales C. Neutrophils in Cancer: Two Sides of the Same Coin. *J Immunol Res* (2015) 2015:983698. doi: 10.1155/2015/983698
 110. Huang S, Mills L, Mian B, Tellez C, McCarty M, Yang XD, et al. Fully humanized neutralizing antibodies to interleukin-8 (ABX-IL8) inhibit angiogenesis, tumor growth, and metastasis of human melanoma. *Am J Pathol* (2002) 161:125–34. doi: 10.1016/S0002-9440(10)64164-8
 111. Chao T, Furth EE, Vonderheide RH. CXCR2-Dependent Accumulation of Tumor-Associated Neutrophils Regulates T-cell Immunity in Pancreatic Ductal Adenocarcinoma. *Cancer Immunol Res* (2016) 4:968–82. doi: 10.1158/2326-6066.CIR-16-0188
 112. Steele CW, Karim SA, Leach JDG, Bailey P, Upstill-Goddard R, Rishi L, et al. CXCR2 Inhibition Profoundly Suppresses Metastases and Augments Immunotherapy in Pancreatic Ductal Adenocarcinoma. *Cancer Cell* (2016) 29:832–45. doi: 10.1016/j.ccell.2016.04.014
 113. Acharyya S, Oskarsson T, Vanharanta S, Malladi S, Kim J, Morris PG, et al. A CXCL1 paracrine network links cancer chemoresistance and metastasis. *Cell* (2012) 150:165–78. doi: 10.1016/j.cell.2012.04.042
 114. Wisdom AJ, Hong CS, Lin AJ, Xiang Y, Cooper DE, Zhang J, et al. Neutrophils promote tumor resistance to radiation therapy. *Proc Natl Acad Sci U S A* (2019) 116:18584–89. doi: 10.1073/pnas.1901562116
 115. Kargl J, Zhu X, Zhang H, Yang GHY, Friesen TJ, Shipley M, et al. Neutrophil content predicts lymphocyte depletion and anti-PD1 treatment failure in NSCLC. *JCI Insight* (2019) 4. doi: 10.1172/jci.insight.130850
 116. Shojaei F, Wu X, Qu X, Kowanetz M, Yu L, Tan M, et al. G-CSF-initiated myeloid cell mobilization and angiogenesis mediate tumor refractoriness to anti-VEGF therapy in mouse models. *Proc Natl Acad Sci U S A* (2009) 106:6742–7. doi: 10.1073/pnas.0902280106
 117. Phan VT, Wu X, Cheng JH, Sheng RX, Chung AS, Zhuang G, et al. Oncogenic RAS pathway activation promotes resistance to anti-VEGF therapy through G-CSF-induced neutrophil recruitment. *Proc Natl Acad Sci U S A* (2013) 110:6079–84. doi: 10.1073/pnas.1303302110
 118. Shojaei F, Wu X, Malik AK, Zhong C, Baldwin ME, Schanz S, et al. Tumor refractoriness to anti-VEGF treatment is mediated by CD11b+Gr1+ myeloid cells. *Nat Biotechnol* (2007) 25:911–20. doi: 10.1038/nbt1323
 119. Yan J, Kloecker G, Fleming C, Bousamra M, Hansen R, Hu X, et al. Human polymorphonuclear neutrophils specifically recognize and kill cancerous cells. *Oncoimmunology* (2014) 3:e950163. doi: 10.4161/15384101.2014.950163
 120. Takeshima T, Pop LM, Laine A, Iyengar P, Vitetta ES, Hannan R. Key role for neutrophils in radiation-induced antitumor immune responses: Potentiation with G-CSF. *Proc Natl Acad Sci U S A* (2016) 113:11300–05. doi: 10.1073/pnas.1613187113
 121. Pang Y, Gara SK, Achyut BR, Li Z, Yan HH, Day CP, et al. TGF- β signaling in myeloid cells is required for tumor metastasis. *Cancer Discov* (2013) 3:936–51. doi: 10.1158/2159-8290.CD-12-0527
 122. Sun R, Luo J, Li D, Shu Y, Luo C, Wang SS, et al. Neutrophils with protumor potential could efficiently suppress tumor growth after cytokine priming and in presence of normal NK cells. *Oncotarget* (2014) 5:12621–34. doi: 10.18632/oncotarget.2181
 123. Netea MG, Dominguez-Andres J, Barreiro LB, Chavakis T, Divangahi M, Fuchs E, et al. Defining trained immunity and its role in health and disease. *Nat Rev Immunol* (2020) 20:375–88. doi: 10.1038/s41577-020-0285-6
 124. Netea MG, Schlitzer A, Placek K, Joosten LAB, Schultze JL. Innate and Adaptive Immune Memory: an Evolutionary Continuum in the Host's Response to Pathogens. *Cell Host Microbe* (2019) 25:13–26. doi: 10.1016/j.chom.2018.12.006
 125. Mitroulis I, Ruppova K, Wang B, Chen LS, Grzybek M, Grinenko T, et al. Modulation of Myelopoiesis Progenitors Is an Integral Component of Trained Immunity. *Cell* (2018) 172:147–61.e12. doi: 10.1016/j.cell.2017.11.034
 126. Bekkering S, Arts RJW, Novakovic B, Kourtzelis I, van der Heijden C, Li Y, et al. Metabolic Induction of Trained Immunity through the Mevalonate Pathway. *Cell* (2018) 172:135–46.e9. doi: 10.1016/j.cell.2017.11.025
 127. Kaufmann E, Sanz J, Dunn JL, Khan N, Mendonca LE, Pacis A, et al. BCG Educates Hematopoietic Stem Cells to Generate Protective Innate Immunity against Tuberculosis. *Cell* (2018) 172:176–90.e19. doi: 10.1016/j.cell.2017.12.031
 128. Kalafati L, Kourtzelis I, Schulte-Schrepping J, Li X, Hatzioannou A, Grinenko T, et al. Innate Immune Training of Granulopoiesis Promotes Anti-tumor Activity. *Cell* (2020) 183:771–85. doi: 10.1016/j.cell.2020.09.058

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Kalafati, Mitroulis, Verginis, Chavakis and Kourtzelis. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.