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1 **Monitoring SARS-CoV-2 in municipal wastewater to evaluate the success of lockdown**  
2 **measures for controlling COVID-19 in the UK**

3  
4 Luke S. Hillary<sup>a,\*</sup>, Kata Farkas<sup>a,b</sup>, Kathryn H. Maher<sup>c</sup>, Anita Lucaci<sup>d</sup>, Jamie Thorpe<sup>a,b</sup>, Marco  
5 A. Distaso<sup>c</sup>, William H. Gaze<sup>f</sup>, Steve Paterson<sup>d</sup>, Terry Burke<sup>c</sup>, Thomas R. Connor<sup>g,h</sup>, James E.  
6 McDonald<sup>a</sup>, Shelagh K. Malham<sup>b</sup>, David L. Jones<sup>a,i</sup>

7 <sup>a</sup> *School of Natural Sciences, Bangor University, Bangor, Gwynedd, LL57 2UW, UK*

8 <sup>b</sup> *School of Ocean Sciences, Bangor University, Menai Bridge, Anglesey, LL59 5AB, UK*

9 <sup>c</sup> *NERC Environmental Omics Facility, Department of Animal and Plant Sciences, University of*  
10 *Sheffield, Sheffield, S10 2TN, UK*

11 <sup>d</sup> *NERC Environmental Omics Facility, Institute of Infection, Veterinary and Ecological*  
12 *Sciences, University of Liverpool, Liverpool, L69 7ZB, UK*

13 <sup>e</sup> *Centre for Environmental Biotechnology, School of Natural Sciences, Bangor University,*  
14 *Bangor, Gwynedd, LL57 2UW, UK*

15 <sup>f</sup> *European Centre for Environment and Human Health, University of Exeter Medical School,*  
16 *ESI, Penryn Campus, TR10 9FE UK*

17 <sup>g</sup> *Organisms and Environment Division, School of Biosciences, Cardiff University, Cardiff, CF10*  
18 *3AX, UK*

19 <sup>h</sup> *Public Health Wales, University Hospital of Wales, Cardiff, CF14 4XW, UK*

20 <sup>i</sup> *UWA School of Agriculture and Environment, The University of Western Australia, Perth, WA*  
21 *6009, Australia*

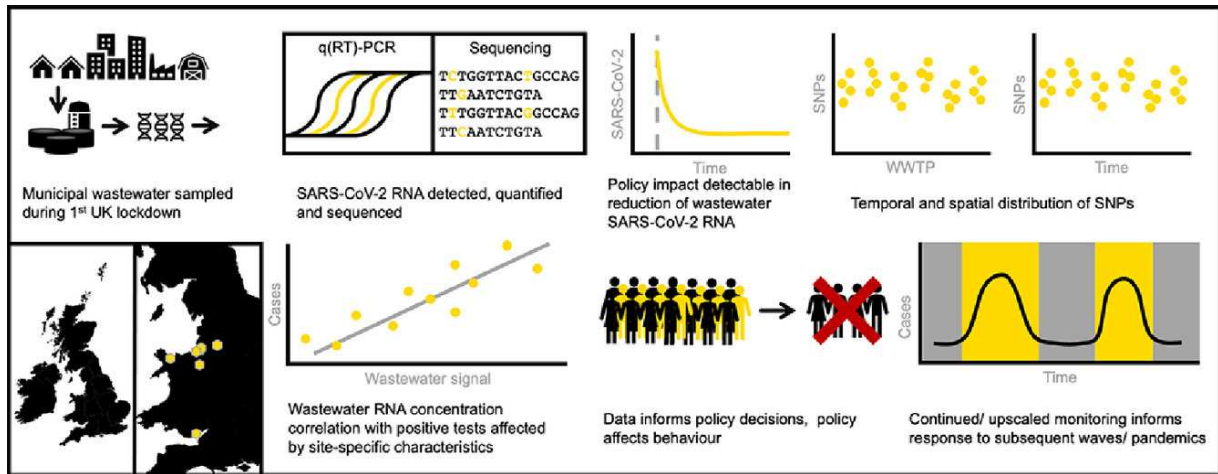
22  
23 \* Corresponding author.

24 *E-mail address: luke.hillary@bangor.ac.uk (Luke S. Hillary).*

25 **Highlights**

- 26 • Wastewater was used to monitor SARS-CoV-2 prevalence and genetic diversity.
- 27 • SARS-CoV-2 RNA abundance and diversity reflects clinical case load and lineages.
- 28 • Temporal analysis of SARS-CoV-2 in sewage tracks the effectiveness of lockdowns.
- 29 • Wastewater-based epidemiology is a useful tool for pandemic response policy.
- 30 • Further research is required to understand factors that affect virus quantification.

31 **Graphical Abstract**



33 **Abstract**

34 SARS-CoV-2 and the resulting COVID-19 pandemic represents one of the greatest recent  
35 threats to human health, wellbeing and economic growth. Wastewater-based epidemiology  
36 (WBE) of human viruses can be a useful tool for population-scale monitoring of SARS-CoV-2  
37 prevalence and epidemiology to help prevent further spread of the disease, particularly within  
38 urban centres. Here we present a longitudinal analysis (March-July, 2020) of SARS-CoV-2 RNA  
39 prevalence in sewage across six major urban centres in the UK (total population equivalent 3  
40 million) by q(RT-)PCR and viral genome sequencing. Our results demonstrate that levels of  
41 SARS-CoV-2 RNA generally correlated with the abundance of clinical cases recorded within

42 the community in large urban centres, with a marked decline in SARS-CoV-2 RNA abundance  
43 following the implementation of lockdown measures. The strength of this association was  
44 weaker in areas with lower confirmed COVID-19 case numbers. Further sequencing analysis of  
45 SARS-CoV-2 from wastewater suggested that multiple genetically distinct clusters were co-  
46 circulating in the local populations covered by our sample sites, and that the genetic variants  
47 observed in wastewater reflected similar SNPs observed in contemporaneous samples from cases  
48 tested in clinical diagnostic laboratories. We demonstrate how WBE can be used for both  
49 community-level detection and tracking of SARS-CoV-2 and other virus' prevalence, and can  
50 inform public health policy decisions. Although, greater understanding of the factors that affect  
51 SARS-CoV-2 RNA concentration in wastewater are needed for the full integration of WBE data  
52 into outbreak surveillance. In conclusion, our results lend support to the use of routine WBE for  
53 monitoring of SARS-CoV-2 and other human pathogenic viruses circulating in the population  
54 and assessment of the effectiveness of disease control measures.

55

56 **Keywords:** coronavirus outbreak, infection control, municipal wastewater, public health, sewage  
57 surveillance.

58

## 59 **1. Introduction**

60 The emergence of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2), and  
61 the resulting global Coronavirus disease 2019 (COVID-19) pandemic has had disastrous socio-  
62 economic and political consequences worldwide (Chakraborty and Maity, 2020). This led to the  
63 World Health Organisation (WHO) declaring the COVID-19 pandemic a global health  
64 emergency (WHO, 2020). In response to this, many countries implemented a range of mitigation  
65 strategies to reduce the spread of disease, including social distancing, restricted movement, use  
66 of personal protection equipment, contact tracing, shielding of vulnerable populations, local or

67 national lockdowns, and community mass testing (Cirrincione et al., 2020; Iacobucci, 2020).  
68 These measures are of particular importance in urbanised areas where the spread of disease is  
69 most likely (Zhang and Schwartz, 2020). These measures proved to be largely effective at  
70 reducing the first wave of COVID-19, albeit not completely eliminating infections (Goscé et al.,  
71 2020; Jarvis et al., 2020). The occurrence of subsequent waves of COVID-19 is of  
72 significant concern, as countries seek to learn from the effectiveness of the mitigation measures  
73 used during the first wave of infection (Aleta et al., 2020).

74 A large proportion of SARS-CoV-2 infections are asymptomatic or result in only a mild  
75 infection (Nishiura et al., 2020). When symptoms do become apparent, this typically occurs 3-7  
76 days after infection (Arons et al., 2020) and severity can vary widely across different sectors of  
77 society, disproportionately affecting the elderly (Wang et al., 2020). Evidence points towards the  
78 fact that individuals can transmit the virus unknowingly prior to developing symptoms.  
79 Furthermore, a- and pre-symptomatic individuals pose challenges to surveillance efforts to  
80 accurately estimate the presence and extent of infection in the community. In a more practical  
81 sense, both asymptomatic and pre-symptomatic individuals also pose a major threat to public  
82 health as they can unknowingly spread the virus to more vulnerable groups (He et al., 2020).

83 Although mass community testing has been instigated in many countries to estimate the  
84 prevalence of COVID-19 in the population, this is costly and the demand for tests frequently  
85 exceeds the capacity of testing facilities (Barasa et al., 2020). Focussing testing solely on  
86 symptomatic cases may also fail to capture asymptomatic and pre-symptomatic infections, and  
87 may focus on populations such as those who are hospitalised, meaning that surveillance is  
88 unavailable for the wider community. In some cases, it can also be difficult to obtain  
89 nasopharyngeal swabs from high-risk parts of the community due to a range of physical,  
90 logistical or cultural issues. Wastewater-based epidemiology (WBE) detects genome fragments

91 of SARS-CoV-2 shed in faeces and urine, and represents an alternative strategy to monitor the  
92 levels of virus circulating at population-level scales (Farkas et al., 2020; Kitajima et al., 2020;  
93 Polo et al., 2020). WBE approaches have previously been successful in evaluating the prevalence  
94 of other viral diseases (e.g. polio-, norovirus) and also for tracking the use of illicit substances,  
95 pharmaceuticals and exposure to xenobiotics (Castiglioni et al., 2014; Ozawa et al., 2019;  
96 Zuccato et al., 2008). Monitoring viruses in wastewater also allows an evaluation of the potential  
97 risk posed by the discharge of treated and untreated wastewater into the wider environment.  
98 Overall, WBE may represent a cost-effective method for determining viral prevalence at the  
99 population-level, and has been used to monitor of SARS-CoV-2 in a range of countries  
100 (Supplementary Table 1).

101 Despite the simplicity of the approach, the quantitative recovery of viruses and viral nucleic  
102 acids from wastewater is notoriously difficult (Farkas et al., 2018a). For example, virus  
103 concentrations in wastewater can be heavily influenced by (i) dilution by rainfall and industrial  
104 inputs, (ii) the presence of compounds that may degrade the virus (e.g. detergents, pH, salt), (iii)  
105 the presence of substances that physically protect the virus (e.g. faecal matter), (iv) loss of viral  
106 RNA during long transit times through the wastewater network due to decay and sorption, (v)  
107 variable shedding rates in the community, and (vi) inhibitory substances in the wastewater that  
108 may interfere with quantitative (reverse transcription)-PCR (q(RT-)PCR ) reactions (Polo et al.,  
109 2020). In addition to these factors, the protocols used to concentrate and purify viral nucleic acids  
110 from wastewater samples can have substantial impacts on recovery, leading to underestimation  
111 of the quantities of the virus present in the wastewater system. Consequently, there is a need to  
112 better understand the factors that influence observable levels of SARS-CoV-2 in wastewater to  
113 allow validation of the approach for surveillance purposes.

114 Largescale efforts to monitor changes in the SARS-CoV-2 genome and track its circulation at  
115 national and global scales have largely relied on the analysis of high-throughput sequencing of  
116 the SARS-CoV-2 genome in symptomatic individuals (Islam et al., 2020; Meredith et al., 2020;  
117 Plessis et al., 2021). As retrospective screening of respiratory samples has detected asymptomatic  
118 cases of COVID-19 (Meredith et al., 2020) it suggests that lineages may appear in wastewater  
119 samples prior to observation in clinical cases. Because wastewater aggregates samples from  
120 across a community/area, sequencing of SARS-CoV-2 RNA recovered from wastewater is likely  
121 to contain multiple lineages and so analysis of this data also has the potential to assess the  
122 proportions of different lineages circulating in the wider population. This potentially enables the  
123 identification of lineages that are known to be present and early warning of new lineages not  
124 previously observed in a catchment.

125 Here, we present a 3.5-month longitudinal analysis of SARS-CoV-2 RNA prevalence and  
126 genetic diversity across six different urban centres during the imposition and gradual lifting of  
127 the first national lockdown period in the UK (March-July 2020). The aims of this study were to  
128 (i) investigate the use of WBE for tracking SARS-CoV-2 after the implementation of national  
129 lockdown measures at six urban centres of varying size within the UK, (ii) determine the  
130 influence of environmental factors (e.g. flow) on levels of SARS-CoV-2 RNA and a human  
131 faecal marker virus (crAssphage) in wastewater, (iii) investigate the impact of wastewater  
132 treatment on the removal of SARS-CoV-2 RNA from wastewater, and (iv) assess the utility of  
133 WBE in understanding SARS-CoV-2 genetic variation through high-throughput sequencing.

## 134 **2. Materials and Methods**

135 All laboratory procedures were carried out in line with Public Health England/ Public Health  
136 Wales advice on the handling of samples suspected of containing SARS-CoV-2.

## 137 2.1 *Sampling sites and wastewater sampling*

138 Untreated influent and treated effluent wastewater were collected from six wastewater  
139 treatment plants (WWTPs) located in Wales and Northwest England. The WWTPs served urban  
140 areas in the local authority areas of Gwynedd, Cardiff, Liverpool, Manchester, the Wirral and  
141 Wrexham, with a total combined population equivalent of ~3 million people (Supplementary Fig.  
142 1). Untreated wastewater influent from the six WWTPs was sampled on a weekly basis between  
143 March and July 2020. Samples were collected in polypropylene bottles as single grab samples  
144 with the exception of the Wirral site, which was collected as a 24-hour composite sample using  
145 an autosampler. Grab samples were collected on weekdays between 08.00 and 09.00 h to ensure  
146 temporal comparability, and treated effluent also collected periodically at the same time as  
147 influent. Samples were transported on either the same day, or overnight on ice, to the laboratory,  
148 stored at 4 °C and processed within 24 h of receipt. Aliquots of wastewater samples (1.5 ml)  
149 were also frozen in polypropylene vials at -80 °C for subsequent physico-chemical analyses and  
150 extraction of pre-concentration viral nucleic acids.

## 151 2.2 *Wastewater physicochemical analyses*

152 Wastewater samples were pasteurised before physicochemical analysis by heating to 60 °C for  
153 90 min. Wastewater ammonium concentrations were determined colorimetrically using the  
154 salicylic acid procedure of Mulvaney (1996). Nitrate was determined colorimetrically using the  
155 vanadate procedure of Miranda et al. (2001) while molybdate-reactive phosphate (MRP) was  
156 determined according to Murphy and Riley (1962). All analysis was performed in a 96-well plate  
157 format using a PowerWave XS Microplate Spectrophotometer (BioTek Instruments Inc.,  
158 Winooski, VT). Wastewater electrical conductivity (EC) was measured using a Jenway 4520  
159 conductivity meter and pH with a Hanna 209 pH meter (Hanna Instruments Ltd., Leighton  
160 Buzzard, UK).

161 *2.3 Wastewater concentration and nucleic acid extraction*

162 Duplicate samples of 50-100 mL of unpasteurised wastewater influent underwent  
163 centrifugation (10,000 g, 30 min, 4°C) and the supernatant and pellet retained. Supernatants were  
164 concentrated to 500 µL using Centriprep 50 kDa MWCO centrifugal concentrators (Merck  
165 KGaA, Germany). For wastewater effluent samples (see Supplementary Table 5), 1-2 L of each  
166 effluent was initially concentrated using tangential flow ultrafiltration with a 100 kDa PES  
167 membrane (Spectrumlabs, USA) as previously described (Farkas et al., 2018c), followed by  
168 secondary concentration using Centriprep concentrators as described above.

169 Selected wastewater concentrates, centrifugation pellets and unconcentrated wastewater  
170 samples were spiked with approximately  $4 \times 10^5$  genome copies (gc) of murine  
171 norovirus (MNV) as a viral RNA extraction control. Positive and negative nucleic acid control  
172 extractions of nuclease-free water with or without the same quantity of MNV spike-in were used  
173 to quantify MNV recovery by q(RT-)PCR and to check for cross-contamination during the  
174 nucleic acid extraction process or q(RT-)PCR assay setup (described in section 2.4). The MNV  
175 was cultured in BV2 cells in Dulbecco's modified Eagle's minimum essential medium  
176 supplemented with 2% foetal bovine serum (FBS) at 37 °C in 5% CO<sub>2</sub> for two days. Viruses  
177 were harvested by three cycles of freeze-thawing (-20°C/+37°C) followed by centrifugation and  
178 100× dilution of the supernatant in phosphate-buffered saline pH 7.4. Aliquots of MNV stock  
179 were stored at -80°C until use. The MNV and BV2 tissue stocks were kindly provided by Prof  
180 Ian Goodfellow (University of Cambridge, UK).

181 Nucleic acids were extracted using the NucliSENS MiniMag Nucleic Acid Purification  
182 System (BioMérieux SA, Marcy-l'Étoile, France) according to the manufacturer's protocol as  
183 described elsewhere (Farkas et al., 2021) in a final volume of 50 (last week of March 2020) or  
184 100 µL (April-July 2020) of elution buffer. Extracted nucleic acids were stored at -80 °C prior

185 to q(RT-)PCR quantification. The nucleic acid extractions and q(RT-)PCR assay setups were  
186 carried out in separate laboratories inside class II microbiological safety cabinets to minimise the  
187 risk of contamination.

#### 188 *2.4 q(RT-)PCR and qPCR assays*

189 The q(RT-)PCR assays were carried out in a QuantStudio® Flex 6 Real-Time PCR System  
190 (Applied Biosystems, USA) using primers, probes and reaction conditions described in  
191 Supplementary Table 2. SARS-CoV-2 N1 and MNV RNA were quantified using a duplex q(RT-  
192 )PCR assay or in triplex with SARS-CoV-2 E gene, as described in Farkas et al. (2021). The  
193 25 µL reaction mix contained 1×RNA Ultrasense Reaction Mix with 1 µL RNA Ultrasense  
194 Enzyme Mix (Invitrogen, USA), 12.5 pmol of the forward and the reverse primers, 6.25 pmol of  
195 the probe/probes, 0.1× ROX reference dye, 1.25 µg bovine serum albumin (BSA) and 2-5 µL of  
196 the extracted wastewater RNA, molecular grade water as a negative control or virus  
197 standards. Initially, 5 µL of extracted RNA was tested for wastewater samples. If the MNV  
198 recovery was lower than 1%, samples were retested with 2 µL sample/reaction to  
199 assess inhibition of the q(RT-)PCR assay, however this was found to be detrimental to assay  
200 sensitivity. All data-points used in the analysis came from assays of 5 µL of extracted nucleic  
201 acids.

202 CrAssphage was used as a marker of human faecal abundance/loading in the wastewater  
203 (Farkas et al., 2019; Stachler et al., 2018). CrAssphage DNA was quantified using a singleplex  
204 qPCR as described previously (Farkas et al., 2019). The 20 µL reaction mix contained 1× KAPA  
205 Probe Force qPCR mix (KAPA Biosystems, USA) with 10 pmol of the forward, 10 pmol of the  
206 reverse primers, 5 pmol of the probe, 1 µg bovine serum albumin, and 2 µL and 4 µL of the  
207 concentrated and original wastewater nucleic acid extracts or controls.

208 A serial dilution of DNA standards within the range of  $10^5$ - $10^0$  genome copies (gc)  $\mu\text{L}^{-1}$  was used for  
209 quantification. For SARS-CoV-2, commercially available circular plasmids carrying the N gene or E gene  
210 were used (Integrated DNA Technologies Inc., Coralville, IA). Plasmid DNA concentrations were halved  
211 when setting up serial dilutions to account for ssRNA producing half the fluorescence signal of dsDNA  
212 at the same concentration. For MNV and crAssphage, custom-made, single-stranded oligo DNA  
213 sequences carrying the target region were used (Life Technologies, USA). Negative controls  
214 (molecular grade water) were included in each run. All samples, standards and controls were run  
215 in duplicate and the mean value for each extraction replicate used for further analysis.

216 The limit of detection (LoD) and limit of quantification (LoQ) of the triplex q(RT-)PCR assays  
217 were determined previously (Farkas et al., 2021) by running wastewater samples spiked with low  
218 concentrations of SARS-CoV-2 (1-150 gc  $\mu\text{L}^{-1}$  N1 CDC and 1-200 gc  $\mu\text{L}^{-1}$  E Sarbeco) and MNV  
219 RNA (1-80 gc  $\mu\text{L}^{-1}$ ) in ten replicates. The q(RT-PCR) assay LoD (the lowest concentration where  
220 all replicates were positive) were 1.7, 3.8 and 3.1 gc  $\mu\text{L}^{-1}$  for the N gene, E gene and MNV,  
221 respectively. The LoQ (the lowest concentration where the coefficient of variance was below  
222 0.25) were 11.8, 25.1 and 32.1 gc  $\mu\text{L}^{-1}$  for the N gene, E gene and MNV, respectively.

### 223 *2.5 q(RT-)PCR data analysis and visualisation*

224 Data were analysed using QuantStudio™ Real-Time PCR Software, version 1.3 (Applied  
225 Biosystems, USA). The baseline (cycle threshold; Ct) was manually adjusted after each run,  
226 when necessary. Viral concentrations were expressed as mean genome copies (gc)  $100\text{ ml}^{-1}$   
227 wastewater calculated from two q(RT-)PCR duplicates of two extraction duplicates ( $n = 4$ ) per  
228 sampling timepoint. Statistical analyses and data visualisation was performed in R v4.0.2 (R  
229 Core Team, 2020; Wickham, 2016). Supplementary Table 3 contains a full list of packages used  
230 in the data analysis.

231 2.6 SARS-CoV-2 RNA amplicon sequencing and data processing

232 RNA from 84 extraction duplicates from 42 time-points, plus no-template negative controls,  
233 were treated with DNase, and used to generate cDNA (NEB Luna Script). Subsequently, SARS-  
234 CoV-2 cDNA underwent PCR amplification using V3 nCov-2019 primers (ARTIC) generating  
235 400 bp amplicons tiling the viral genome (Quick and Loman, 2020). Amplicon generation was  
236 followed by sequencing library construction (NEB Ultra II DNA), with equimolar pooling of  
237 samples and quantitation. Final library size was assessed on a Bioanalyser high sensitivity DNA  
238 chip, and DNA concentration determined by Qubit double-stranded DNA high sensitivity assay,  
239 and then by qPCR using the Illumina Library Quantification Kit from Kapa (KK4854) on a  
240 Roche Light Cycler LC480II according to the manufacturer's instructions. Libraries were  
241 sequenced on an Illumina MiSeq generating 2×250 bp paired end reads. An average of *ca*  
242 291,000 reads (*ca* 146 Mbp) per sample were mapped using bwa-mem against the SARS-CoV-  
243 2 genome reference (MN908947.3) within the ncov2019-artic-nf v3 pipeline  
244 (<https://github.com/connor-lab/ncov2019-artic-nf>). SNPs and indels were identified using  
245 Varscan v2.4.4 with default settings and summary statistics for coverage and diversity were  
246 generated in R v4.0.2 (R Core Team, 2020; Wickham, 2016). Sites were filtered to remove SNPs  
247 and indels with a coverage of less than 50× and a variant frequency of less than 10% per sample.  
248 The number of SNP and indel sites were calculated per sample.

249 The relationship between SNP and indel site frequency and the proportion of the genome with  
250 coverage at greater than 50× coverage and the  $\log_{10} \text{gc } \mu\text{L}^{-1}$  were examined with Spearman's  
251 correlations. An index of SNP plus indel frequency per sample was calculated by taking the  
252 number of SNP and indel sites and dividing by the proportion of the genome with coverage at  
253 greater than 50 reads. A mean SNP and indel frequency index were then calculated per pair of  
254 wastewater samples to examine the effect of the number of positive tests in the previous 7 days  
255 in the local authority area, sample date and WWTP site on the number of SNPs and indels

256 discovered, using a general linear model using the ‘glm’ function and type II ANOVA using the  
257 R package ‘car’. A Spearman’s correlation was used to examine the relationship between the  
258 index of SNP and indel frequency and the log population equivalent served by each wastewater  
259 treatment plant. Variants at SNP and indel sites were compared to those recorded in clinical  
260 samples using the ‘cov\_glue\_snp\_lineage’ function from R package ‘sars2pack’.

### 261 **3. Results and Discussion**

#### 262 *3.1 Study description and q(RT-)PCR assay development*

263 We monitored the SARS-CoV-2 RNA concentration in influent wastewater at six wastewater  
264 treatment plants (WWTPs) using q(RT-)PCR over a period of 3.5 months during the imposition  
265 and gradual lifting of the first UK-wide lockdown, and compared these data to the numbers of  
266 positive clinical tests and deaths reported by the Office for National Statistics (ONS), UK  
267 Government and Public Health Wales for lower tier local authority areas within which the  
268 WWTPs were located (HM Government, 2020; Office for National Statistics, 2020; Public  
269 Health Wales, 2020). WWTPs represent a range in size (population equivalents from 40 thousand  
270 to 1.1 million) and spatial distribution (see Supplementary Fig. 1) and all implemented combined  
271 stormwater, domestic and trade wastewater collection. Influent wastewater grab samples were  
272 collected at the same time each week with the exception of The Wirral WWTP which was  
273 sampled from a 24 hour composite autosampler. Limits of detection (LoD) and quantification  
274 (LoQ) were determined as described in Farkas et al. (2021).

275 Results for SARS-CoV-2 RNA concentrations from q(RT-)PCR quantification are displayed as  
276 unadjusted mean genome copies/ 100 ml of wastewater rather than normalised by crAssphage  
277 concentrations as factors such as extraction efficiency can vary depending on the virus used  
278 (Medema et al., 2020). Although studies suggest that 24-hour composite sampling is more  
279 representative than grab sampling, it has been shown that grab samples are accurate to within an

280 order of magnitude (Curtis et al., 2020, Ahmed et al., 2021). Further, our previous work has  
281 shown limited diurnal variability, particularly in large wastewater catchments where transit times  
282 can be up to 24 hours and where large amounts of mixing occurs in the network (Farkas et al.,  
283 2018b). Transit times may also influence observable virus quantities due to degradation of viral  
284 nucleic acids as they pass through the sewage system; however, SARS-CoV-2 RNA has been  
285 shown to be relatively stable in wastewater under environmental conditions, with a  $T_{90}$  of 24 or  
286 28 days at 15 or 4 °C (Ahmed et al., 2020b).

287 We compared mean SARS-CoV-2 RNA concentrations to daily flow and influent wastewater  
288 chemistry but found no statistically significant correlations (see supplementary table 4). The  
289 highly abundant bacteriophage crAssphage was used as a human faecal marker. No correlation  
290 was found between crAssphage and SARS-CoV-2 nucleic acid concentrations (Spearman,  $p =$   
291  $0.8341$ ). No effect on crAssphage concentration was observable from sampling week (Kruskal-  
292 Wallis,  $p = 0.9042$ ), but a significant effect was found between crAssphage concentration and  
293 WWTP site (Kruskal-Wallis,  $p = 0.01751$ ). These data indicate that faecal loading was constant  
294 throughout the study period and that different WWTPs have different balances of human waste  
295 and industrial/ other domestic wastewater sources.

### 296 *3.2 Temporal trends in SARS-CoV-2 RNA concentration in wastewater and comparison to* 297 *COVID-19 epidemiology*

298 For each WWTP,  $64\% \pm 6.8$  q(RT-)PCR tests (mean  $\pm$  standard error (SEM), sites = 6,  $n =$   
299 90) detected SARS-CoV-2 in influent wastewater above the LoD, with SARS-CoV-2 RNA  
300 concentrations in wastewater influent having quantities above the LoQ in  $28.9\% \pm 2.2$  of samples  
301 (see Supplementary Fig. 2). No sites showed SARS-CoV-2 concentrations in WWTP effluent  
302 above the LoQ and only one above the LoD (Wrexham, 19/05/20,  $n = 22$ , see supplementary  
303 table 5). Figure 1a shows a drop in wastewater SARS-CoV-2 RNA concentration, new positive

304 clinical tests and COVID-19 related deaths following the imposition of the UK-wide lockdown  
305 beginning in late March 2020. A number of spikes in clinical cases can be observed without  
306 corresponding spikes in wastewater, e.g. Wrexham in late June. These can occur due to surge  
307 testing following local workplace-related outbreaks and changes in testing eligibility during the  
308 study, and highlight the inherent difficulties in comparing wastewater loads to positive tests when  
309 testing is both limited and non-random.

310 WWTPs in Manchester, Liverpool and the Wirral showed strong correlations between SARS-  
311 CoV-2 RNA concentration and daily positive tests (Fig. 1b and Supplementary Fig. 3). Negative  
312 correlations were also observed between viral concentrations in all sites and time following the  
313 implementation of national lockdown, except Cardiff, indicating these measures lowered the  
314 prevalence of the virus in local populations. The Cardiff, Gwynedd and Wrexham WWTPs did  
315 not show the same trends between viral RNA concentrations and tests/ deaths, potentially due to  
316 several different factors such as water chemistry or lower, broader peaks in SARS-CoV-2  
317 prevalence. Gwynedd is also a popular holiday destination and sees regular weekend influxes of  
318 holiday makers from other parts of the UK, which could affect WWTP SARS-CoV-2  
319 concentrations either positively (through visits from asymptomatic/ pre-symptomatic  
320 individuals) or negatively (through people commuting from rural areas outside of the WWTP  
321 catchment area). Additional factors such as transit time within the sewage network, catchment  
322 flow dynamics, and differences between local authority reporting areas for positive tests and  
323 WWTP sewershed coverage could have effects on viral RNA recovery. In contrast to the  
324 Gwynedd site, the Wirral site showed the strongest correlation between SARS-CoV-2 RNA  
325 concentrations and the number of positive clinical tests/ COVID-19 related deaths, and is of a  
326 size inbetween that of the Wrexham and Gwynedd WWTPs (see Supplemental Fig. 1),  
327 suggesting that the use of 24-hour composite sampling may improve the correlation between  
328 SARS-CoV-2 wastewater quantification and local clinical cases.

329 Further exploration of site-specific factors and improved access to higher resolution spatial  
330 distributions of positive test locations is required to improve the accuracy of WBE in predicting  
331 COVID-19 prevalence amongst local populations as part of national monitoring programmes.  
332 Previous studies have corrected SARS-CoV-2 RNA concentration for WWTP flow (Gonzalez et  
333 al., 2020), and adjusted cases or positive tests for differences between local authority populations  
334 and WWTP catchment areas (Medema et al., 2020). Statistically, we found no benefit of  
335 correcting for these factors on Spearman correlation coefficients between WWTP SARS-CoV-2  
336 RNA concentration and positive tests/ COVID-19 related deaths (see Supplemental Fig. 3),  
337 however due to differences between WWTP sites and sewersheds, we would caution against  
338 making extensive quantitative comparisons between sites.

339 Our data confirm that SARS-CoV-2 RNA is readily detectable in wastewater influent across  
340 a range of concentrations from  $<1.2 \times 10^3$  (<LoQ) to the highest recorded concentration of  $1.5 \times$   
341  $10^4$  gc 100 mL<sup>-1</sup>. This highlights how site-specific factors, concentration and quantification  
342 protocols, and sampling strategies can complicate quantitative comparisons between WWTPs  
343 within the same study, and when making comparisons to other international studies. There is a  
344 need to standardise SARS-CoV-2 wastewater quantification and take WWTP site identity into  
345 account when expanding WWTP monitoring programmes to national and international scales  
346 (Chik et al., 2021; Pecson et al., 2021). Nonetheless, this study demonstrates the longitudinal  
347 benefit of using WBE to monitor viral prevalence and the impact of public health interventions,  
348 particularly in the early stages of a novel disease outbreak.

### 349 *3.3 Effect of window size/ offset on correlations*

350 Due to shedding of SARS-CoV-2 from asymptomatic and pre-symptomatic individuals, a key  
351 driver of WBE research is the potential to detect upcoming spikes in infection in wastewater  
352 before increase in positive clinical tests. Consequently, several studies have used modelling

353 approaches to assess if the wastewater concentration of SARS-CoV-2 preceded new spikes in  
354 clinical cases of COVID-19 (Ahmed et al., 2021b; D'Aoust et al., 2021). However, this is  
355 challenging due to variabilities in the point of an infection cycle at which a person gets tested,  
356 the severity and duration of symptoms, and the variability in viral shedding. The effect of varying  
357 the difference between the number of days between wastewater sampling and testing date and  
358 the number of days over which to sum the number of positive tests on the correlation between  
359 wastewater SARS-CoV-2 concentrations and cases was examined (Fig. 2). If only considering  
360 daily clinical testing data, the SARS-CoV-2 wastewater RNA concentration leads testing data  
361 by 2-4 days but this can be extended by approximately 1 day by using a rolling sum of positive  
362 clinical test cases over a series of days leading up to the clinical testing date being considered. It  
363 should be noted that the overall effect of varying these parameters is not large in that the  
364 correlation coefficients stay between 0.8 and 0.9 over a range of permutations.

### 365 *3.4 Sequencing detects mutations in the SARS-CoV-2 genome comparable to those observable* 366 *in clinical cases*

367 WBE can also be used to monitor the genetic diversity SARS-CoV-2 circulating in the wider  
368 population. To this end, SARS-CoV-2 RNA was amplified using the ARTIC protocol primers in  
369 both extraction duplicates, where at least one of which showed q(RT-)PCR amplification were  
370 sequenced. In these samples, between 25–75% of the SARS-CoV-2 genome was recovered (Fig.  
371 3a), with coverage randomly distributed across the genome (Fig. 3b). This included samples that  
372 showed no amplification (8.3%) or amplification below the LoD (3.6%) of the N1 q(RT-)PCR  
373 assay ( $n = 84$ ), suggesting that multi-locus amplicon sequencing based monitoring of wastewater  
374 for WBE may be of significant use in the early stages of future viral outbreaks. The proportion  
375 of the genome sequenced positively correlated with the amount of template (Spearman's  $\rho =$   
376  $0.376, p = 0.0004$ , Fig. 3c).

377 In total, 702 unique SNP sites and 267 indels were detectable across the 84 samples after  
378 filtering to remove sites with less than 50 reads and a variant frequency within a sample of less  
379 than 10%. The number of SNPs found correlated positively with the proportion of the genome  
380 that was sequenced (Spearman's  $\rho = 0.581$ ,  $p < 0.0001$ ; Fig. 3d).

381 Preliminary modelling suggests that the rate of positive tests in the source population and  
382 sampling week did not affect the mean number of SNPs and indels controlled for genome  
383 coverage ( $p > 0.05$ ; Fig. 4a and b), but a reduced model suggested that there was heterogeneity  
384 among sites ( $\chi^2 = 11.57$ ,  $df = 5$ ,  $p = 0.041$ ; Fig. 4c). The index of SNP plus indel frequency was  
385 not related to log population equivalent served by each wastewater treatment plant (Spearman's  
386  $\rho = 0.251$ ,  $p = 0.251$ ; Fig. 4d). This is explained by the presence of multiple viral lineages present  
387 within the sample, corresponding to the diverse infections in the population represented in the  
388 wastewater sample. A substantial fraction of the detected SNPs has previously been identified in  
389 clinical samples across the UK, and has the potential to be informative for distinguishing viral  
390 lineages (Supplementary Table 6).

391 Multiple SARS-CoV-2 lineages can be present within a single wastewater sample. Samples  
392 have the potential to contain viruses from both symptomatic and asymptomatic individuals  
393 within the community, as SARS-CoV-2 has been detected in the faeces of both asymptomatic  
394 and symptomatic individuals (Jones et al., 2020; Tang et al., 2020). Previous studies have  
395 sequenced SARS-CoV-2 genomes from wastewater (Ahmed et al., 2020a; Izquierdo-Lara et al.,  
396 n.d.; Martin et al., 2020; Nemudryi et al., 2020). We have shown not only that viral genome  
397 sequences can be recovered from wastewater samples, but that they exhibit substantial diversity  
398 across dozens of samples. Sequencing the genomes therefore has the potential to assess the  
399 diversity of viral infections in the wastewater catchment population and to identify emerging  
400 genetic variants before they are seen in clinical samples. In support of this, preliminary analysis

401 suggests that the detected SNPs were consistent with those detected previously in clinical  
402 samples (see Supplementary Table 6). However, because the SNPs from wastewater samples are  
403 not phased across the genome, and because the genome coverage is imperfect, assigning viral  
404 lineages to samples will require a bespoke statistical framework to be developed.

### 405 ***3.8. Use of wastewater-based epidemiology in COVID-19 and future pathogen surveillance***

406 Attempting to quantitatively link observed viral RNA concentrations to detectable cases is  
407 challenging (Medema et al., 2020). Many assumptions need to be made regarding the persistence  
408 of SARS-CoV-2 in wastewater, quantities of the virus shed in faeces and the influence of water  
409 chemistry (Ahmed et al., 2020a).

410 Sample processing methodology can also be a substantial source of variability. Concentration  
411 method, qPCR assay design and inter-lab variation can create variation in detectable SARS-CoV-  
412 2 RNA quantities (Pecson et al., 2021; Westhaus et al., 2021). Use of appropriate process controls  
413 is necessary to monitor the effects of these factors when making intra- and inter-laboratory  
414 comparisons. Choice of process control is complex as a closely related surrogate virus should be  
415 used where available and further global collaboration and co-ordination is required to widen  
416 access to WBE technologies (Polo et al., 2020). In addition to this, the effects of SARS-CoV-2  
417 on global supply chains and the need to perform WBE at scale create additional pressures where  
418 sub-optimal protocols may become necessary in the future to achieve testing scale desired for  
419 national monitoring programs.

420 Despite the possible sources of variability mentioned above, we have demonstrated that WBE  
421 is suitable for quantitatively tracking the course of the early stages of the SARS-CoV-2 pandemic  
422 and the effects of public health interventions, even in the early stages of a novel outbreak, where  
423 lack of surge capacity prevents optimal sampling. We highlight how tiled primer array  
424 sequencing complements q(RT-)PCR based detection of SARS-CoV-2 and enhances the

425 sensitivity and usefulness of WBE in detecting the presence of novel mutations in the SARS-  
426 CoV-2 genome. Early detection of viral pathogens by q(RT-)PCR requires a suitable assay and  
427 routine monitoring of WWTPs however alternative technologies such as viral metagenomics  
428 may be more suited to initial detection of emerging and unknown pathogens (Farkas et al., 2020).  
429 Our results suggest that viral amplicon sequencing could be more sensitive than q(RT-)PCR for  
430 detection of known pathogens. In future, monitoring could be targeted towards ports of entry and  
431 major metropolitan centres to maximise the likelihood of detection (Medema et al., 2020).

#### 432 **4. Conclusions**

- 433 • Our results demonstrate that levels of SARS-CoV-2 RNA in wastewater generally  
434 correlated well with the abundance of clinical COVID-19 cases recorded within the  
435 community in large urban centres.
- 436 • At the population level, wastewater-based epidemiology was used to confirm the success  
437 of lockdown measures (i.e. restricted movement and human-to human contact)  
438 implemented at the national scale to control the transmission of SARS-CoV-2.
- 439 • The genetic diversity of SARS-CoV-2 from wastewater suggests that multiple genetically  
440 distinct clusters were co-circulating present in the local populations, and that the genetic  
441 variants observed in wastewater reflect similar SNPs observed in samples from  
442 nasopharyngeal swabs taken contemporaneously at clinical testing centres.
- 443 • A greater understanding of the factors that affect SARS-CoV-2 RNA quantification in  
444 wastewater is still required to enable the full integration of wastewater-based  
445 epidemiology data into wider outbreak surveillance programmes.
- 446 • Our results lend support to the use of routine wastewater-based epidemiology to monitor  
447 SARS-CoV-2 and other human pathogenic viruses circulating in the population and to  
448 assess the effectiveness of disease control measures.

449

450 **Declaration of Competing Interest** The authors declare that they have no known competing  
451 financial interests or personal relationships that could have appeared to influence the work  
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### 463 **Data Availability**

464 q(RT-)PCR and chemical data recorded in this study is available as supplementary information  
465 and from the Environmental Information Data Centre (EIDC, [www.eidc.ceh.uk](http://www.eidc.ceh.uk)).  
466 DOI:10.5285/ce40e62a-21ae-45b9-ba5b-031639a504f7. Sequencing read files analysed in this  
467 study can be accessed from the European Nucleotide Archive (project PRJEB42191).

### 468 **Author contributions**

469 DLJ, LSH, KF, SKM and JEM conceived the project. LSH, JT, MAD and KF undertook the  
470 experimental work. LSH and KF undertook the processing and analysis of the q(RT-)PCR data.  
471 KHM, TB and SP undertook the processing and analysis of the sequencing data. LSH, KF and  
472 DLJ led the data interpretation and writing of the manuscript. All other authors contributed to  
473 the final draft of the article.

- 475 Ahmed, W., Angel, N., Edson, J., Bibby, K., Bivins, A., O'Brien, J.W., Choi, P.M., Kitajima,  
476 M., Simpson, S.L., Li, J., Tschärke, B., Verhagen, R., Smith, W.J.M., Zaugg, J.,  
477 Dierens, L., Hugenholtz, P., Thomas, K.V., Mueller, J.F., 2020a. First confirmed  
478 detection of SARS-CoV-2 in untreated wastewater in Australia: A proof of concept for  
479 the wastewater surveillance of COVID-19 in the community. *Sci. Total Environ.* 728,  
480 138764. <https://doi.org/10.1016/j.scitotenv.2020.138764>
- 481 Ahmed, W., Bertsch, P.M., Bibby, K., Haramoto, E., Hewitt, J., Huygens, F., Gyawali, P.,  
482 Korajkic, A., Riddell, S., Sherchan, S.P., Simpson, S.L., Sirikanchana, K., Symonds,  
483 E.M., Verhagen, R., Vasana, S.S., Kitajima, M., Bivins, A., 2020b. Decay of SARS-  
484 CoV-2 and surrogate murine hepatitis virus RNA in untreated wastewater to inform  
485 application in wastewater-based epidemiology. *Environ. Res.* 191, 110092.  
486 <https://doi.org/10.1016/j.envres.2020.110092>
- 487 Ahmed, W., Bivins, A., Bertsch, P.M., Bibby, K., Gyawali, P., Sherchan, S.P., Simpson, S.L.,  
488 Thomas, K.V., Verhagen, R., Kitajima, M., Mueller, J.F., Korajkic, A., 2021a. Intraday  
489 variability of indicator and pathogenic viruses in 1-h and 24-h composite wastewater  
490 samples: Implications for wastewater-based epidemiology. *Environ. Res.* 193, 110531.  
491 <https://doi.org/10.1016/j.envres.2020.110531>
- 492 Ahmed, W., Tschärke, B., Bertsch, P.M., Bibby, K., Bivins, A., Choi, P., Clarke, L., Dwyer, J.,  
493 Edson, J., Nguyen, T.M.H., O'Brien, J.W., Simpson, S.L., Sherman, P., Thomas, K.V.,  
494 Verhagen, R., Zaugg, J., Mueller, J.F., 2021b. SARS-CoV-2 RNA monitoring in  
495 wastewater as a potential early warning system for COVID-19 transmission in the  
496 community: A temporal case study. *Sci. Total Environ.* 761, 144216.  
497 <https://doi.org/10.1016/j.scitotenv.2020.144216>
- 498 Aleta, A., Martín-Corral, D., Pastore y Piontti, A., Ajelli, M., Litvinova, M., Chinazzi, M.,  
499 Dean, N.E., Halloran, M.E., Longini Jr, I.M., Merler, S., Pentland, A., Vespignani, A.,  
500 Moro, E., Moreno, Y., 2020. Modelling the impact of testing, contact tracing and  
501 household quarantine on second waves of COVID-19. *Nat. Hum. Behav.* 4, 964–971.  
502 <https://doi.org/10.1038/s41562-020-0931-9>
- 503 Arons, M.M., Hatfield, K.M., Reddy, S.C., Kimball, A., James, A., Jacobs, J.R., Taylor, J.,  
504 Spicer, K., Bardossy, A.C., Oakley, L.P., Tanwar, S., Dyal, J.W., Harney, J., Chisty, Z.,  
505 Bell, J.M., Methner, M., Paul, P., Carlson, C.M., McLaughlin, H.P., Thornburg, N.,  
506 Tong, S., Tamin, A., Tao, Y., Uehara, A., Harcourt, J., Clark, S., Brostrom-Smith, C.,  
507 Page, L.C., Kay, M., Lewis, J., Montgomery, P., Stone, N.D., Clark, T.A., Honein,  
508 M.A., Duchin, J.S., Jernigan, J.A., 2020. Presymptomatic SARS-CoV-2 Infections and  
509 Transmission in a Skilled Nursing Facility. *N. Engl. J. Med.* 382, 2081–2090.  
510 <https://doi.org/10.1056/NEJMoa2008457>
- 511 Barasa, E.W., Ouma, P.O., Okiro, E.A., 2020. Assessing the hospital surge capacity of the  
512 Kenyan health system in the face of the COVID-19 pandemic. *PLOS ONE* 15,  
513 e0236308. <https://doi.org/10.1371/journal.pone.0236308>
- 514 Castiglioni, S., Thomas, K.V., Kasprzyk-Hordern, B., Vandam, L., Griffiths, P., 2014. Testing  
515 wastewater to detect illicit drugs: State of the art, potential and research needs. *Sci.*  
516 *Total Environ.* 487, 613–620. <https://doi.org/10.1016/j.scitotenv.2013.10.034>
- 517 Chakraborty, I., Maity, P., 2020. COVID-19 outbreak: Migration, effects on society, global  
518 environment and prevention. *Sci. Total Environ.* 728, 138882.  
519 <https://doi.org/10.1016/j.scitotenv.2020.138882>
- 520 Chik, A.H.S., Glier, M.B., Servos, M., Mangat, C.S., Pang, X.-L., Qiu, Y., D'Aoust, P.M.,  
521 Burnet, J.-B., Delatolla, R., Dorner, S., Geng, Q., Giesy, J.P., McKay, R.M., Mulvey,  
522 M.R., Prystajeky, N., Srikanthan, N., Xie, Y., Conant, B., Hruday, S.E., 2021.

523 Comparison of approaches to quantify SARS-CoV-2 in wastewater using RT-qPCR:  
524 Results and implications from a collaborative inter-laboratory study in Canada. *J.*  
525 *Environ. Sci.* 107, 218–229. <https://doi.org/10.1016/j.jes.2021.01.029>

526 Cirrincione, L., Plescia, F., Ledda, C., Rapisarda, V., Martorana, D., Moldovan, R.E.,  
527 Theodoridou, K., Cannizzaro, E., 2020. COVID-19 Pandemic: Prevention and  
528 Protection Measures to Be Adopted at the Workplace. *Sustainability* 12, 3603.  
529 <https://doi.org/10.3390/su12093603>

530 Curtis, K., Keeling, D., Yetka, K., Larson, A., Gonzalez, R., 2020. Wastewater SARS-CoV-2  
531 Concentration and Loading Variability from Grab and 24-Hour Composite Samples.  
532 *medRxiv* 2020.07.10.20150607. <https://doi.org/10.1101/2020.07.10.20150607>

533 D’Aoust, P.M., Graber, T.E., Mercier, E., Montpetit, D., Alexandrov, I., Neault, N., Baig, A.T.,  
534 Mayne, J., Zhang, X., Alain, T., Servos, M.R., Srikanthan, N., MacKenzie, M., Figeys,  
535 D., Manuel, D., Jüni, P., MacKenzie, A.E., Delatolla, R., 2021. Catching a resurgence:  
536 Increase in SARS-CoV-2 viral RNA identified in wastewater 48 h before COVID-19  
537 clinical tests and 96 h before hospitalizations. *Sci. Total Environ.* 770, 145319.  
538 <https://doi.org/10.1016/j.scitotenv.2021.145319>

539 Farkas, K., Adriaenssens, E.M., Walker, D.I., McDonald, J.E., Malham, S.K., Jones, D.L.,  
540 2019. Critical Evaluation of CrAssphage as a Molecular Marker for Human-Derived  
541 Wastewater Contamination in the Aquatic Environment. *Food Environ. Virol.* 11, 113–  
542 119. <https://doi.org/10.1007/s12560-019-09369-1>

543 Farkas, K., Cooper, D.M., McDonald, J.E., Malham, S.K., de Rougemont, A., Jones, D.L.,  
544 Rougemont, A. de, Jones, D.L., de Rougemont, A., Jones, D.L., 2018a. Seasonal and  
545 spatial dynamics of enteric viruses in wastewater and in riverine and estuarine receiving  
546 waters. *Sci. Total Environ.* 634, 1174–1183.  
547 <https://doi.org/10.1016/j.scitotenv.2018.04.038>

548 Farkas, K., Hillary, L.S., Malham, S.K., McDonald, J.E., Jones, D.L., 2020. Wastewater and  
549 public health: the potential of wastewater surveillance for monitoring COVID-19. *Curr.*  
550 *Opin. Environ. Sci. Health* 17, 14–20. <https://doi.org/10.1016/j.coesh.2020.06.001>

551 Farkas, K., Hillary, L.S., Thorpe, J., Walker, D.I., Lowther, J.A., McDonald, J.E., Malham,  
552 S.K., Jones, D.L., 2021. Concentration and Quantification of SARS-CoV-2 RNA in  
553 Wastewater Using Polyethylene Glycol-Based Concentration and qRT-PCR. *Methods*  
554 *Protoc.* 4, 17. <https://doi.org/10.3390/mps4010017>

555 Farkas, K., Marshall, M., Cooper, D., McDonald, J.E., Malham, S.K., Peters, D.E., Maloney,  
556 J.D., Jones, D.L., 2018b. Seasonal and diurnal surveillance of treated and untreated  
557 wastewater for human enteric viruses. *Environ. Sci. Pollut. Res.* 25, 33391–33401.  
558 <https://doi.org/10.1007/s11356-018-3261-y>

559 Farkas, K., McDonald, J.E., Malham, S.K., Jones, D.L., 2018c. Two-Step Concentration of  
560 Complex Water Samples for the Detection of Viruses. *Methods Protoc.* 1.  
561 <https://doi.org/10.3390/mps1030035>

562 Gonzalez, R., Curtis, K., Bivins, A., Bibby, K., Weir, M.H., Yetka, K., Thompson, H., Keeling,  
563 D., Mitchell, J., Gonzalez, D., 2020. COVID-19 surveillance in Southeastern Virginia  
564 using wastewater-based epidemiology. *Water Res.* 186, 116296.  
565 <https://doi.org/10.1016/j.watres.2020.116296>

566 Goscé, L., Phillips, P.A., Spinola, P., Gupta, D.R.K., Abubakar, P.I., 2020. Modelling SARS-  
567 COV2 Spread in London: Approaches to Lift the Lockdown. *J. Infect.* 81, 260–265.  
568 <https://doi.org/10.1016/j.jinf.2020.05.037>

569 He, X., Lau, E.H.Y., Wu, P., Deng, X., Wang, J., Hao, X., Lau, Y.C., Wong, J.Y., Guan, Y.,  
570 Tan, X., Mo, X., Chen, Y., Liao, B., Chen, W., Hu, F., Zhang, Q., Zhong, M., Wu, Y.,  
571 Zhao, L., Zhang, F., Cowling, B.J., Li, F., Leung, G.M., 2020. Temporal dynamics in

572 viral shedding and transmissibility of COVID-19. *Nat. Med.* 26, 672–675.  
573 <https://doi.org/10.1038/s41591-020-0869-5>

574 HM Government, 2020. Coronavirus (COVID-19) in the UK - Cases Legacy CSV download  
575 [WWW Document]. URL [https://coronavirus.data.gov.uk/downloads/csv/coronavirus-](https://coronavirus.data.gov.uk/downloads/csv/coronavirus-cases_latest.csv)  
576 [cases\\_latest.csv](https://coronavirus.data.gov.uk/downloads/csv/coronavirus-cases_latest.csv) (accessed 9.14.20).

577 Iacobucci, G., 2020. Covid-19: UK lockdown is “crucial” to saving lives, say doctors and  
578 scientists. *BMJ* m1204. <https://doi.org/10.1136/bmj.m1204>

579 Islam, M.R., Hoque, M.N., Rahman, M.S., Alam, A.S.M.R.U., Akther, M., Puspo, J.A., Akter,  
580 S., Sultana, M., Crandall, K.A., Hossain, M.A., 2020. Genome-wide analysis of SARS-  
581 CoV-2 virus strains circulating worldwide implicates heterogeneity. *Sci. Rep.* 10,  
582 14004. <https://doi.org/10.1038/s41598-020-70812-6>

583 Izquierdo-Lara, R., Elsinga, G., Heijnen, L., Munnink, B.B.O., Schapendonk, C.M.E.,  
584 Nieuwenhuijse, D., Kon, M., Lu, L., Aarestrup, F.M., Lycett, S., Medema, G.,  
585 Koopmans, M.P.G., Graaf, M. de, n.d. Early Release - Monitoring SARS-CoV-2  
586 Circulation and Diversity through Community Wastewater Sequencing, the Netherlands  
587 and Belgium - Volume 27, Number 5—May 2021 - *Emerging Infectious Diseases*  
588 journal - CDC. <https://doi.org/10.3201/eid2705.204410>

589 Jarvis, C.I., Van Zandvoort, K., Gimma, A., Prem, K., Auzenbergs, M., O’Reilly, K., Medley,  
590 G., Emery, J.C., Houben, R.M.G.J., Davies, N., Nightingale, E.S., Flasche, S., Jombart,  
591 T., Hellewell, J., Abbott, S., Munday, J.D., Bosse, N.I., Funk, S., Sun, F., Endo, A.,  
592 Rosello, A., Procter, S.R., Kucharski, A.J., Russell, T.W., Knight, G., Gibbs, H.,  
593 Leclerc, Q., Quilty, B.J., Diamond, C., Liu, Y., Jit, M., Clifford, S., Pearson, C.A.B.,  
594 Eggo, R.M., Deol, A.K., Klepac, P., Rubin, G.J., Edmunds, W.J., CMMID COVID-19  
595 working group, 2020. Quantifying the impact of physical distance measures on the  
596 transmission of COVID-19 in the UK. *BMC Med.* 18, 124.  
597 <https://doi.org/10.1186/s12916-020-01597-8>

598 Jones, D.L., Baluja, M.Q., Graham, D.W., Corbishley, A., McDonald, J.E., Malham, S.K.,  
599 Hillary, L.S., Connor, T.R., Gaze, W.H., Moura, I.B., Wilcox, M.H., Farkas, K., 2020.  
600 Shedding of SARS-CoV-2 in feces and urine and its potential role in person-to-person  
601 transmission and the environment-based spread of COVID-19. *Sci. Total Environ.* 749,  
602 141364. <https://doi.org/10.1016/j.scitotenv.2020.141364>

603 Martin, J., Klapsa, D., Wilton, T., Zambon, M., Bentley, E., Bujaki, E., Fritzsche, M., Mate, R.,  
604 Majumdar, M., 2020. Tracking SARS-CoV-2 in Sewage: Evidence of Changes in Virus  
605 Variant Predominance during COVID-19 Pandemic. *Viruses* 12.  
606 <https://doi.org/10.3390/v12101144>

607 Medema, G., Heijnen, L., Elsinga, G., Italiaander, R., Brouwer, A., 2020. Presence of SARS-  
608 Coronavirus-2 RNA in Sewage and Correlation with Reported COVID-19 Prevalence  
609 in the Early Stage of the Epidemic in The Netherlands. *Environ. Sci. Technol. Lett.* 7,  
610 511–516. <https://doi.org/10.1021/acs.estlett.0c00357>

611 Meredith, L.W., Hamilton, W.L., Warne, B., Houldcroft, C.J., Hosmillo, M., Jahun, A.S.,  
612 Curran, M.D., Parmar, S., Caller, L.G., Caddy, S.L., Khokhar, F.A., Yakovleva, A.,  
613 Hall, G., Feltwell, T., Forrest, S., Sridhar, S., Weekes, M.P., Baker, S., Brown, N.,  
614 Moore, E., Popay, A., Roddick, I., Reacher, M., Gouliouris, T., Peacock, S.J., Dougan,  
615 G., Török, M.E., Goodfellow, I., 2020. Rapid implementation of SARS-CoV-2  
616 sequencing to investigate cases of health-care associated COVID-19: a prospective  
617 genomic surveillance study. *Lancet Infect. Dis.* 20, 1263–1272.  
618 [https://doi.org/10.1016/S1473-3099\(20\)30562-4](https://doi.org/10.1016/S1473-3099(20)30562-4)

619 Miranda, K.M., Espey, M.G., Wink, D.A., 2001. A Rapid, Simple Spectrophotometric Method  
620 for Simultaneous Detection of Nitrate and Nitrite. *Nitric Oxide* 5, 62–71.  
621 <https://doi.org/10.1006/niox.2000.0319>

622 Mulvaney, R.L., 1996. Nitrogen - Inorganic Forms, in: *Methods of Soil Analysis, Part 3.*  
623 *Chemical Methods* (D.L. Sparks Ed.). SSSA, Madison, WI, USA, pp. 1123–1184.

624 Murphy, J., Riley, J.P., 1962. A modified single solution method for the determination of  
625 phosphate in natural waters. *Anal. Chim. Acta* 27, 31–36.  
626 [https://doi.org/10.1016/S0003-2670\(00\)88444-5](https://doi.org/10.1016/S0003-2670(00)88444-5)

627 Nemudryi, A., Nemudraia, A., Wiegand, T., Surya, K., Buyukyoruk, M., Cicha, C.,  
628 Vanderwood, K.K., Wilkinson, R., Wiedenheft, B., 2020. Temporal Detection and  
629 Phylogenetic Assessment of SARS-CoV-2 in Municipal Wastewater. *Cell Rep. Med.* 1,  
630 100098. <https://doi.org/10.1016/j.xcrm.2020.100098>

631 Nishiura, H., Kobayashi, T., Miyama, T., Suzuki, A., Jung, S., Hayashi, K., Kinoshita, R.,  
632 Yang, Y., Yuan, B., Akhmetzhanov, A.R., Linton, N.M., 2020. Estimation of the  
633 asymptomatic ratio of novel coronavirus infections (COVID-19). *Int. J. Infect. Dis.* 94,  
634 154–155. <https://doi.org/10.1016/j.ijid.2020.03.020>

635 Office for National Statistics, 2020. Deaths registered weekly in England and Wales,  
636 provisional.

637 Ozawa, H., Yoshida, H., Usuku, S., 2019. Environmental Surveillance Can Dynamically Track  
638 Ecological Changes in Enteroviruses. *Appl. Environ. Microbiol.* 85, e01604-19,  
639 [/aem/85/24/AEM.01604-19.atom](https://doi.org/10.1128/AEM.01604-19). <https://doi.org/10.1128/AEM.01604-19>

640 Pecson, B.M., Darby, E., Haas, C.N., Amha, Y.M., Bartolo, M., Danielson, R., Dearborn, Y.,  
641 Giovanni, G.D., Ferguson, C., Fevig, S., Gaddis, E., Gray, D., Lukasik, G., Mull, B.,  
642 Olivas, L., Olivieri, A., Qu, Y., Consortium, S.-C.-2 I., 2021. Reproducibility and  
643 sensitivity of 36 methods to quantify the SARS-CoV-2 genetic signal in raw  
644 wastewater: findings from an interlaboratory methods evaluation in the U.S. *Environ.*  
645 *Sci. Water Res. Technol.* <https://doi.org/10.1039/D0EW00946F>

646 Plessis, L. du, McCrone, J.T., Zarebski, A.E., Hill, V., Ruis, C., Gutierrez, B., Raghwani, J.,  
647 Ashworth, J., Colquhoun, R., Connor, T.R., Faria, N.R., Jackson, B., Loman, N.J.,  
648 O’Toole, Á., Nicholls, S.M., Parag, K.V., Scher, E., Vasylyeva, T.I., Volz, E.M., Watts,  
649 A., Bogoch, I.I., Khan, K., Consortium†, C.-19 G.U. (COG-U., Aanensen, D.M.,  
650 Kraemer, M.U.G., Rambaut, A., Pybus, O.G., 2021. Establishment and lineage  
651 dynamics of the SARS-CoV-2 epidemic in the UK. *Science* 371, 708–712.  
652 <https://doi.org/10.1126/science.abf2946>

653 Polo, D., Quintela-Baluja, M., Corbishley, A., Jones, D.L., Singer, A.C., Graham, D.W.,  
654 Romalde, J.L., 2020. Making waves: Wastewater-based epidemiology for COVID-19 –  
655 approaches and challenges for surveillance and prediction. *Water Res.* 186, 116404.  
656 <https://doi.org/10.1016/j.watres.2020.116404>

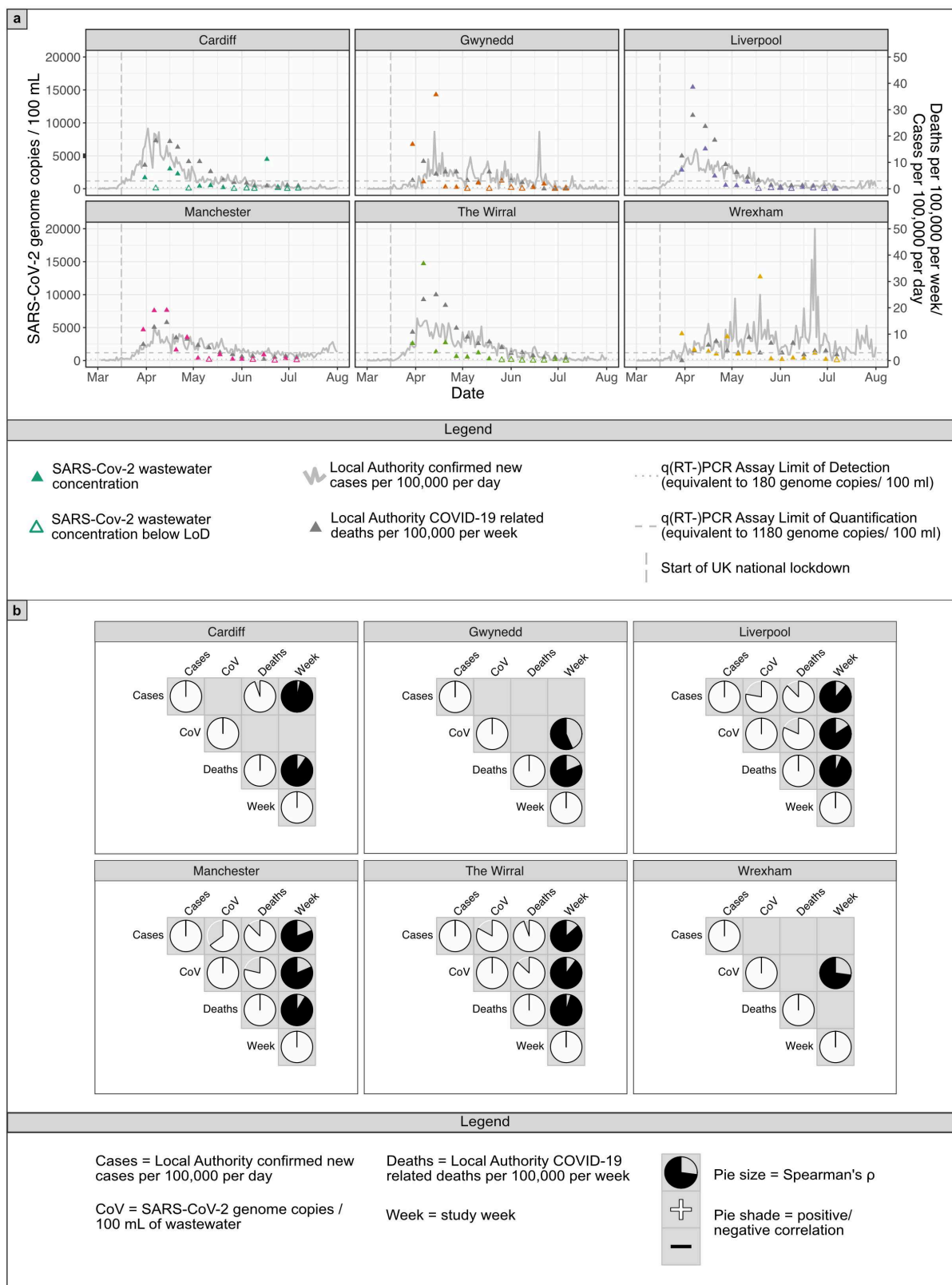
657 Public Health Wales, 2020. Public Health Wales Rapid COVID-19 surveillance [WWW  
658 Document]. URL  
659 [https://public.tableau.com/profile/public.health.wales.health.protection#!/vizhome/Rapi](https://public.tableau.com/profile/public.health.wales.health.protection#!/vizhome/RapidCOVID-19virology-Public/Headlinesummary)  
660 [dCOVID-19virology-Public/Headlinesummary](https://public.tableau.com/profile/public.health.wales.health.protection#!/vizhome/RapidCOVID-19virology-Public/Headlinesummary) (accessed 9.14.20).

661 R Core Team, 2020. R: A Language and Environment for Statistical Computing.

662 Stachler, E., Akyon, B., de Carvalho, N.A., Ference, C., Bibby, K., 2018. Correlation of  
663 crAssphage qPCR Markers with Culturable and Molecular Indicators of Human Fecal  
664 Pollution in an Impacted Urban Watershed. *Environ. Sci. Technol.* 52, 7505–7512.  
665 <https://doi.org/10.1021/acs.est.8b00638>

666 Tang, A., Tong, Z., Wang, H., Dai, Y., Li, K., Liu, J., Wu, W., Yuan, C., Yu, M., Li, P., Yan,  
667 J., 2020. Detection of Novel Coronavirus by RT-PCR in Stool Specimen from  
668 Asymptomatic Child, China. *Emerg. Infect. Dis.* 26, 1337–1339.  
669 <https://doi.org/10.3201/eid2606.200301>

670 Wang, L., He, W., Yu, X., Hu, D., Bao, M., Liu, H., Zhou, J., Jiang, H., 2020. Coronavirus  
671 disease 2019 in elderly patients: Characteristics and prognostic factors based on 4-week  
672 follow-up. *J. Infect.* 80, 639–645. <https://doi.org/10.1016/j.jinf.2020.03.019>  
673 Westhaus, S., Weber, F.-A., Schiwy, S., Linnemann, V., Brinkmann, M., Widera, M., Greve,  
674 C., Janke, A., Hollert, H., Wintgens, T., Ciesek, S., 2021. Detection of SARS-CoV-2 in  
675 raw and treated wastewater in Germany – Suitability for COVID-19 surveillance and  
676 potential transmission risks. *Sci. Total Environ.* 751, 141750.  
677 <https://doi.org/10.1016/j.scitotenv.2020.141750>  
678 Wickham, H., 2016. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York.  
679 Zhang, C.H., Schwartz, G.G., 2020. Spatial Disparities in Coronavirus Incidence and Mortality  
680 in the United States: An Ecological Analysis as of May 2020. *J. Rural Health* 36, 433–  
681 445. <https://doi.org/10.1111/jrh.12476>  
682 Zuccato, E., Chiabrando, C., Castiglioni, S., Bagnati, R., Fanelli, R., 2008. Estimating  
683 Community Drug Abuse by Wastewater Analysis. *Environ. Health Perspect.* 116,  
684 1027–1032. <https://doi.org/10.1289/ehp.11022>  
685

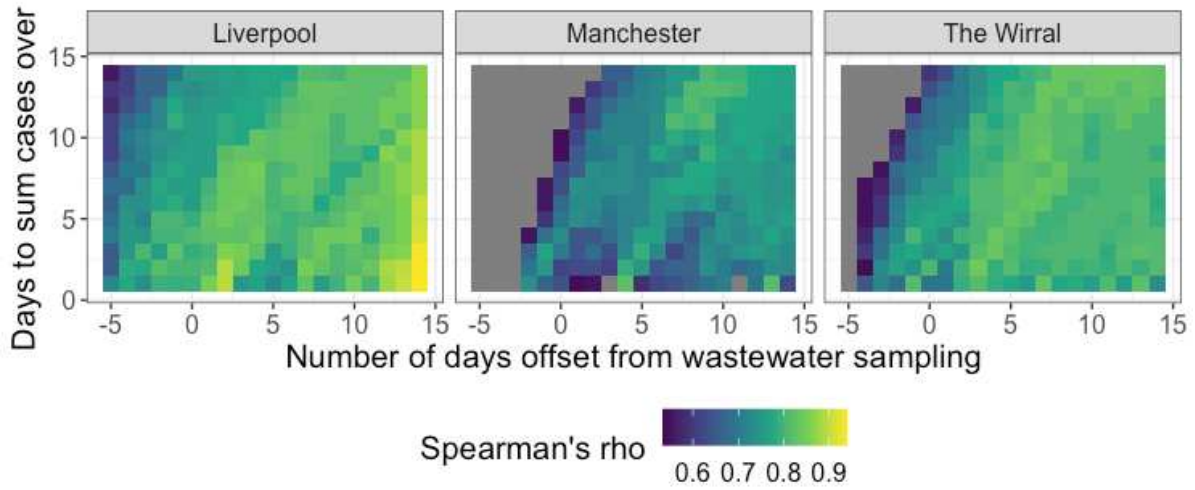


687

688 **Fig. 1(a)** Temporal trend of the recorded number of COVID-19 infections and deaths at six urban

689 centres in the UK and the corresponding levels of SARS-CoV-2 in wastewater. The coloured  
690 triangles represent levels of SARS-CoV-2 in influent wastewater, with open triangles being below  
691 LoD. Grey triangles represent the number of COVID-19 reported deaths and the solid line  
692 represents the number of COVID-19 cases reported in each study region. The dashed and dotted  
693 horizontal lines represent the assay LoQ (scaled to 1180 genome copies/ 100ml) and LoD (180  
694 genome copies/ 100 ml) respectively, scaled for a sample volume of 100 mL. The dashed vertical  
695 line represents the imposition of UK-wide lockdown measures. (b) Correlation of SARS-CoV-2  
696 RNA concentration (CoV) in influent wastewater with COVID-19 related cases and deaths at six  
697 urban centres in the UK. Pie charts represent Spearman correlation  $\rho$  where  $p < 0.05$  with  
698 fullness indicating degree of correlation and colour representing positive (white) or negative  
699 (black) correlations.

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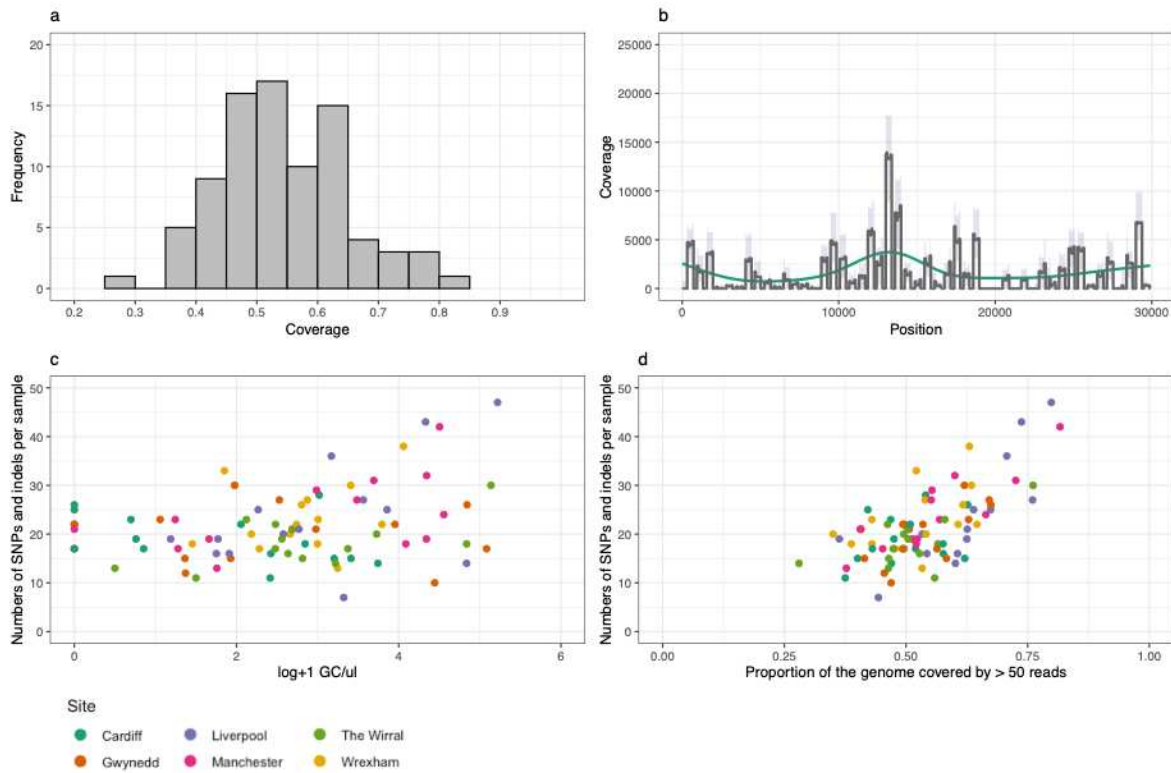


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703 **Fig. 2** Effects of varying the number of days between wastewater sampling date and clinical  
 704 testing date (x axis) and the number of days over which to sum cases over (y axis) on the strength  
 705 of correlation between wastewater SARS-CoV-2 concentration and local authority positive tests.  
 706 Quantities are shown where a false discovery rate corrected p-value was below 0.05.

707



708

709

710 **Fig. 3** Coverage of the SARS-CoV-2 genome from reads recovered from wastewater samples. a)

711 Frequency of the proportion of the genome sequenced at 50× depth or greater. b) Coverage

712 across the genome, median plotted in dark grey, interquartile ranges in purple and a smoothed

713 GAM spline in green. c) Proportion of the genome sequenced relative to the estimated number

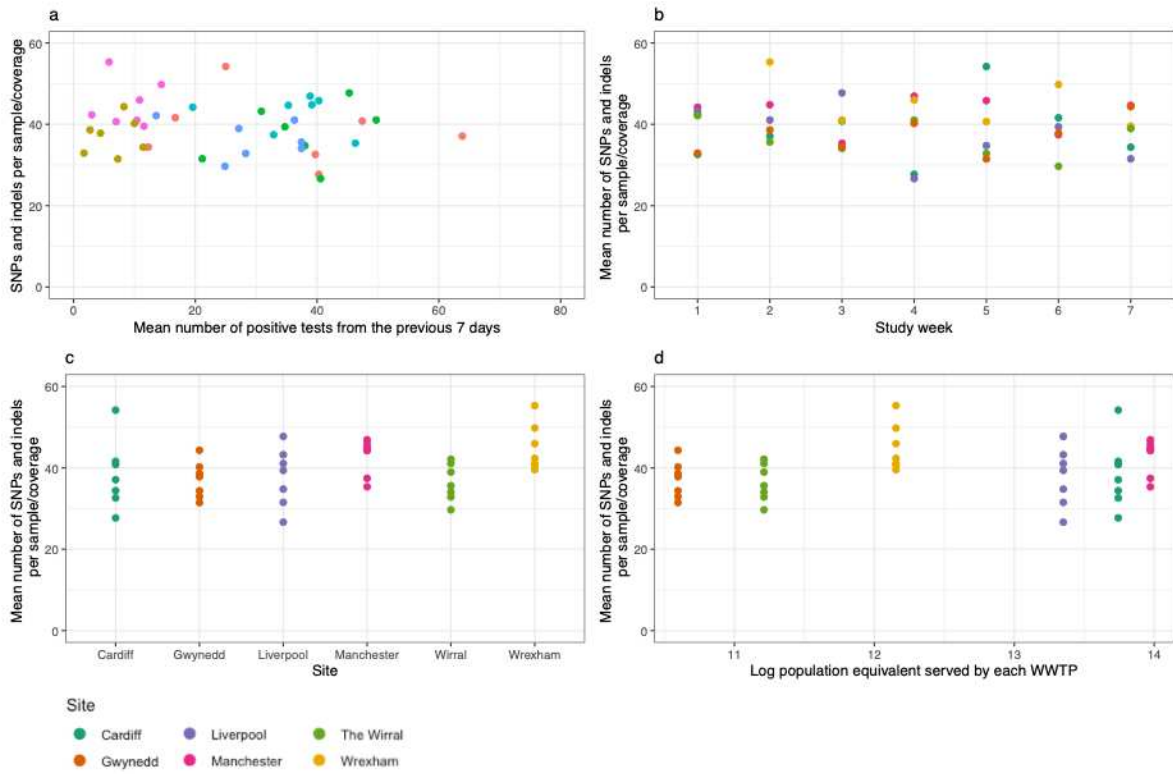
714 of genome copies estimated from (RT)-qPCR. Note that sequence was obtained in several

715 samples where the (RT)-qPCR for this locus was negative, reflecting the ability of the protocol

716 to sequence genomes of low copy number. d) The number of SNP and indel sites detected relative

717 to the proportion of the genome that was sequenced at 50× or higher.

718



719

720

721 **Fig. 4.** Comparison of the mean number of SNP/ INDELs sites divided by genome coverage to

722 (a) positive tests in the previous 7 days in the local authority, (b) sample date, (c) WWTP site

723 and (d)  $\log_{10}$  population equivalent.

724

## 725 **Supplementary Information**

### 726 **Supplementary Results**

#### 727 *4.1 Comparison of N1 CDC and E Sarbeco SARS-CoV-2 q(RT-)PCR assays*

728 Significant correlation was found between SARS-CoV-2 RNA quantified by the N1 CDC and  
729 E Sarbeco gene markers in the same samples (Spearman's  $\rho = 0.56$ ,  $p < 0.0001$ ), however the  
730 LoD and LoQ of the E Sarbeco marker were both 2.1-fold higher than the CDC N1 assay and  
731 three times less likely to detect SARS-CoV-2 in wastewater samples (see Supplementary Fig. 2).  
732 Westhaus et al. (2021) similarly demonstrated varying sensitivity and specificity for SARS-CoV-  
733 2 in commonly used q(RT)-PCR assays and so further comparison, optimisation and  
734 standardisation is required when expanding monitoring programs and making international  
735 comparisons.

736

#### 737 *4.2 Detection of SARS-CoV-2 in WWTP influent suspended solids and effluent*

738 SARS-CoV-2 RNA concentrations were also determined for the pellet from the initial  
739 centrifugation step in the first three weeks of the sampling programme. Only three samples (n =  
740 18) produced quantities above the LoD and consequently, only results for SARS-CoV-2 in  
741 wastewater supernatants were considered in further analysis. It should be noted that SARS-CoV-  
742 2 has been detected in the solid phase in other studies (e.g. primary thickened sludge), however,  
743 the quantity of pelletable solids can be highly variable between samples and between treatment  
744 sites (Peccia et al., 2020; Westhaus et al., 2021).

745 Effluent samples were collected as detailed in Supplementary Table 5 but similarly to  
746 suspended solids, above one sample (Wrexham, 19/05/20) had detectable quantities over the  
747 LoD.

748

**Supplementary Table 1.** Studies reporting SARS-CoV-2 RNA concentration in wastewater influent. p/a = presence/ absence, ct = Ct values only.

749

Site	Peak (gc/ 100 mL)	Reference
<b>Published articles in peer-reviewed journals</b>		
Netherlands (various)	$2.2 \times 10^5$	(Medema et al., 2020)
England/ Wales (various)	$1.5 \times 10^4$	This study
USA (Montana)	$10^5$	(Nemudryi et al., 2020)
Italy (Milan/ Rome)	$5.6 \times 10^3$	(La Rosa et al., 2021)
Australia (Brisbane)	$1.2 \times 10^1$	(Ahmed et al., 2020)
India (Gujarat)	$3.5 \times 10^1$	(Kumar et al., 2020)
USA (Louisiana)	$7.5 \times 10^2$	(Sherchan et al., 2020)
Spain (Mercia)	ct	(Randazzo et al., 2020)
Japan (Yamanashi Prefecture)	$8.2 \times 10^3$	(Haramoto et al., 2020)
France (Montpellier)	$8 \times 10^4$	(Trottier et al., 2020)
Brazil (Rio de Janeiro)	ct	(Prado et al., 2020)
Germany (various)	$2 \times 10^3$	(Westhaus et al., 2021)
USA (Virginia)	$10^4$	(Gonzalez et al., 2020)
<b>Reports hosted on preprint servers</b>		
France (Paris)	$10^6$	(Wurtzer et al., 2020)
India (Jaipur)	pa	(Arora et al., 2020)
Israel (various)	ct	(Bar Or et al., 2020)
Japan (Ishikawa and Toyama)	$4.4 \times 10^3$	(Hata et al., 2020)
Spain (Ourense)	ct	(Balboa et al., 2020)
Spain (Barcelona)	$10^4$	(Chavarria-Miró et al., 2020)
Turkey (Istanbul)	$1.8 \times 10^3$	(Kocamemi et al., 2020)
USA (Massachusetts)	$2.4 \times 10^4$	(Wu et al., 2020)
USA (New York State)	$1.2 \times 10^4$	(Green et al., 2020)

750

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752

**Supplementary Table 2.** q(RT-)PCR and qPCR assay parameters

Primer/Probe	Sequence (5'-3')	Reference	q(RT-)PCR parameters
SARS-CoV-2 (N1) Forward primer	GACCCCAAATCAGCGAAAT	(Centers for Disease Control and Prevention, 2020)	55 °C – 60 min 95 °C – 5 min 45 cycles: 95 °C – 15 s 60 °C – 1 min 65 °C – 1 min
SARS-CoV-2 (N1) Reverse primer	TCTGGTTACTGCCAGTTGAATCTG		
SARS-CoV-2 (N1) Probe*	[FAM]ACCCCGCATTACGTTTGGTGGACC[MGB]		
SARS-CoV-2 (E) Forward primer	ACAGGTACGTTAATAGTTAATAGCGT	(Corman et al., 2020)	
SARS-CoV-2 (E) Reverse primer	ATATTGCAGCAGTACGCACACA		
SARS-CoV-2 (E) Probe*	[VIC]-ACACTAGCCATCCTTACTGCGCTTCG-[QSY]		
MNV Forward primer	CCGCAGGAACGCTCAGCAG	(Kitajima et al., 2010)	
MNV Reverse primer	GGYTGAATGGGACGGCCTG		
MNV Probe*	[ABY]ATGAGTGATGGCGCA[QSY]		
CrAssphage_Q56 Forward primer	CAGAAGTACAACTCCTAAAAACGTAGAG	(Stachler et al., 2017)	98°C – 5 min 40 cycles: 95°C – 15 s 60°C – 1 min
CrAssphage_Q56 CrAssphage Reverse primer	GATGACCAATAAACAAGCCATTAGC		
CrAssphage_Q56 CrAssphage Probe	[FAM]AATAACGATTTACGTGATGTAAC[TAMRA]		

\*Quencher was modified to be compatible with QuantStudio environment.

**Supplementary Table 3.** R packages used in this work

Package Name	Reference
corrplot	(Wei and Simko, 2017)
cowplot	(Wilke, 2020)
data.table	(Dowle and Srinivasan, 2020)
FSA	(Ogle et al., 2020)
ggpubr	(Kassambara, 2020)
ggrepel	(Slowikowski, 2020)
Hmisc	(Harrell, 2020)
plotrix	(Lemon, 2006)
rnaturalearth	(South, 2017)
rworldmap	(South, 2011)
sf	(Pebesma, 2018)
tidyverse	(Wickham et al., 2019)
zoo	(Zeileis and Grothendieck, 2005)

**Supplementary Table 4.** Comparison of SARS-CoV-2 with water quality parameters

Water quality indicator	Site effect (Kruskal-Wallis <i>p</i> -value)	Spearman's correlation with SARS-CoV-2 wastewater concentration ( <i>p</i> -value)
Daily flow/ population equivalent	< 2.2e-16	0.1108
NH <sub>4</sub> <sup>+</sup>	4.402e-05	0.8238
MRP	0.0006494	0.1462
pH	0.004882	0.8141
EC	7.178e-08	0.5206
NO <sub>3</sub> <sup>-</sup>	0.003202	0.06433

761 **Supplementary Table 5.** Genome copies of SARS-CoV-2 in effluent (ND = no detection). All  
 762 values except one were below the LoD (1.7 gc/ $\mu$ l) and all were below the LoQ (11.8 gc/ $\mu$ l).

Site	Sampling Date	Effluent mean SARS-CoV-2 concentration (gc/ $\mu$ L of RNA extract)
Gwynedd	4/5/20	0.51125
Liverpool	11/5/20	ND
Manchester	11/5/20	ND
The Wirral	11/5/20	ND
Gwynedd	18/05/20	ND
Liverpool	18/05/20	ND
Manchester	18/05/29	0.4995
The Wirral	18/05/20	0.492
Wrexham	19/05/20	3.416
Cardiff	27/05/20	1.1365
Liverpool	26/05/20	0.687
Manchester	26/05/20	0.857
The Wirral	26/05/20	ND
Cardiff	4/6/20	0.08925
Liverpool	1/6/20	ND
Manchester	1/6/20	0.405
The Wirral	1/6/20	0.1385
Wrexham	2/6/20	1.4115
Liverpool	8/6/20	0.0965
Manchester	8/6/20	ND
The Wirral	8/6/20	ND
Wrexham	9/6/20	0.198

764 **Supplementary Table 6.** Number of unique SNP/INDEL sites per sample. Of those  
 765 SNP/INDELS we report the number and percentage of sites that match the locations of  
 766 SNP/INDELS found from clinical samples and have the expected variant recorded.

Sample	Number of unique SNP/INDEL sites	Number of sites that match locations in clinical samples	Number of sites that match the expected SNP/INDEL in clinical samples	Percentage of sites that match locations in clinical samples	Percentage of matching sites that match expected base/INDEL from clinical samples
C1WK1	15	11	5	73.33%	45.45%
C2WK1	16	8	5	50.00%	62.50%
D1WK1	42	18	13	42.86%	72.22%
D2WK1	19	8	8	42.11%	100.00%
F1WK1	13	8	8	61.54%	100.00%
F2WK1	37	20	14	54.05%	70.00%
L1WK1	27	15	15	55.56%	100.00%
L2WK1	35	19	11	54.29%	57.89%
M1WK1	17	11	8	64.71%	72.73%
M2WK1	14	8	6	57.14%	75.00%
T1WK1	22	14	9	63.64%	64.29%
T2WK1	10	6	5	60.00%	83.33%
C1WK2	26	14	9	53.85%	64.29%
C2WK2	17	9	9	52.94%	100.00%
D1WK2	19	11	7	57.89%	63.64%
D2WK2	32	20	16	62.50%	80.00%
F1WK2	20	13	7	65.00%	53.85%
F2WK2	23	11	8	47.83%	72.73%
L1WK2	14	9	7	64.29%	77.78%
L2WK2	47	27	22	57.45%	81.48%
M1WK2	29	15	9	51.72%	60.00%
M2WK2	18	8	5	44.44%	62.50%
T1WK2	15	11	5	73.33%	45.45%
T2WK2	21	15	10	71.43%	66.67%
C1WK3	14	10	8	71.43%	80.00%
C2WK3	28	13	8	46.43%	61.54%
D1WK3	24	13	12	54.17%	92.31%
D2WK3	18	10	6	55.56%	60.00%
F1WK3	16	10	7	62.50%	70.00%
F2WK3	18	9	5	50.00%	55.56%
L1WK3	25	17	11	68.00%	64.71%
L2WK3	43	24	16	55.81%	66.67%
M1WK3	17	4	4	23.53%	100.00%

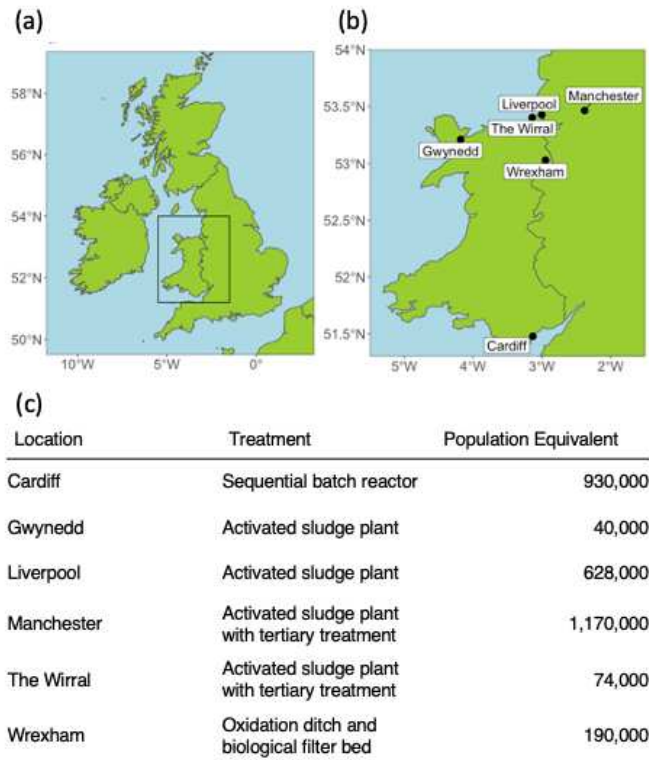
M2WK3	15	8	6	53.33%	75.00%
T1WK3	17	9	6	52.94%	66.67%
T2WK3	26	16	14	61.54%	87.50%
C1WK4	18	10	7	55.56%	70.00%
C2WK4	15	12	8	80.00%	66.67%
D1WK4	21	11	8	52.38%	72.73%
D2WK4	29	18	15	62.07%	83.33%
F1WK4	27	15	13	55.56%	86.67%
F2WK4	18	10	10	55.56%	100.00%
L1WK4	7	3	3	42.86%	100.00%
L2WK4	20	8	5	40.00%	62.50%
M1WK4	20	11	10	55.00%	90.91%
M2WK4	21	13	8	61.90%	61.54%
T1WK4	15	9	5	60.00%	55.56%
T2WK4	22	15	11	68.18%	73.33%
C1WK5	23	12	10	52.17%	83.33%
C2WK5	25	14	10	56.00%	71.43%
D1WK5	27	14	12	51.85%	85.71%
D2WK5	31	16	11	51.61%	68.75%
F1WK5	30	13	9	43.33%	69.23%
F2WK5	22	12	9	54.55%	75.00%
L1WK5	25	16	14	64.00%	87.50%
L2WK5	19	11	10	57.89%	90.91%
M1WK5	19	12	8	63.16%	66.67%
M2WK5	13	10	8	76.92%	80.00%
T1WK5	23	12	10	52.17%	83.33%
T2WK5	12	4	3	33.33%	75.00%
C1WK6	22	7	5	31.82%	71.43%
C2WK6	19	12	10	63.16%	83.33%
D1WK6	13	6	5	46.15%	83.33%
D2WK6	23	13	8	56.52%	61.54%
F1WK6	22	6	5	27.27%	83.33%
F2WK6	33	16	15	48.48%	93.75%
L1WK6	19	11	9	57.89%	81.82%
L2WK6	15	7	6	46.67%	85.71%
M1WK6	23	13	12	56.52%	92.31%
M2WK6	11	7	5	63.64%	71.43%
T1WK6	22	10	9	45.45%	90.00%
T2WK6	17	9	7	52.94%	77.78%
C1WK7	11	6	6	54.55%	100.00%
C2WK7	17	7	6	41.18%	85.71%
D1WK7	17	8	6	47.06%	75.00%
D2WK7	21	8	6	38.10%	75.00%

F1WK7	20	12	10	60.00%	83.33%
F2WK7	26	12	9	46.15%	75.00%
L1WK7	16	13	12	81.25%	92.31%
L2WK7	21	14	13	66.67%	92.86%
M1WK7	16	12	8	75.00%	66.67%
M2WK7	22	12	10	54.55%	83.33%
T1WK7	30	16	11	53.33%	68.75%
T2WK7	27	10	7	37.04%	70.00%
F1WK13	23	15	11	65.22%	73.33%
F2WK13	43	27	20	62.79%	74.07%
G1WK13	15	11	8	73.33%	72.73%
G2WK13	11	5	5	45.45%	100.00%
H1WK13	8	4	4	50.00%	100.00%
H2WK13	16	11	9	68.75%	81.82%
T1WK13	14	5	4	35.71%	80.00%
T2WK13	7	3	3	42.86%	100.00%

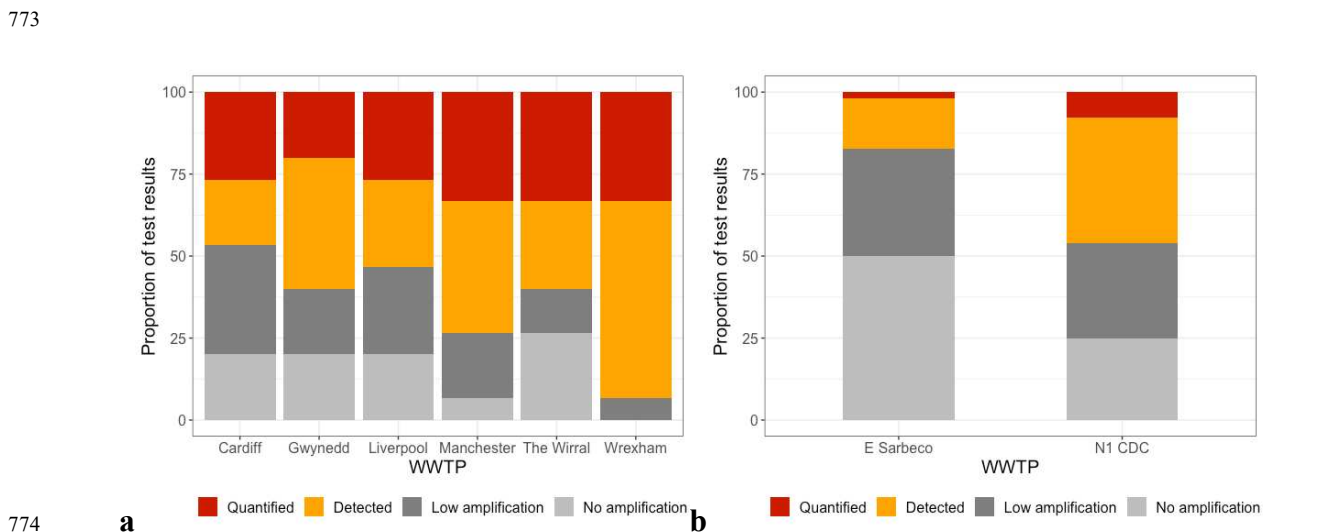
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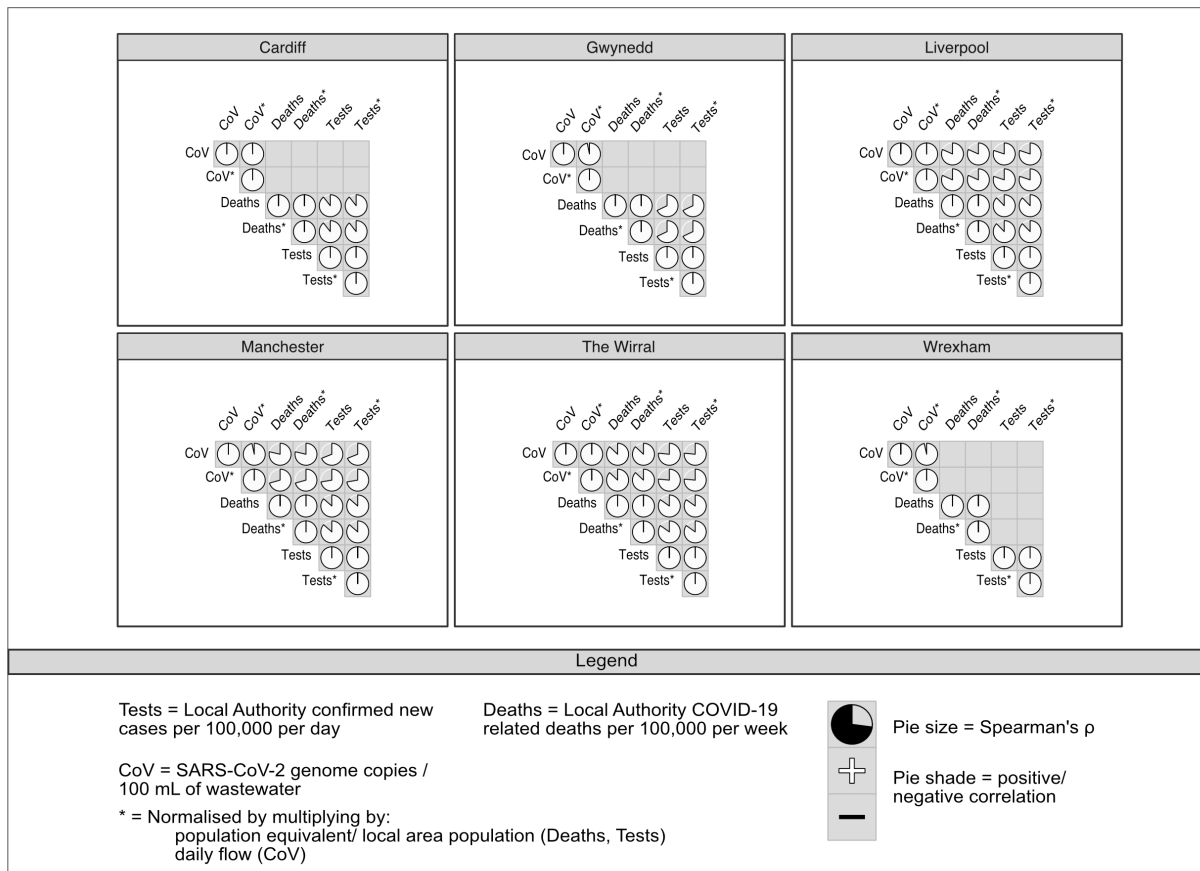
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769  
 770 **Supplementary Fig. 1** General (a) and specific (b) locations of wastewater treatment sites  
 771 surveyed in this study and (c) the equivalent population sizes served. All WWTPs combine  
 772 domestic, trade and stormwater



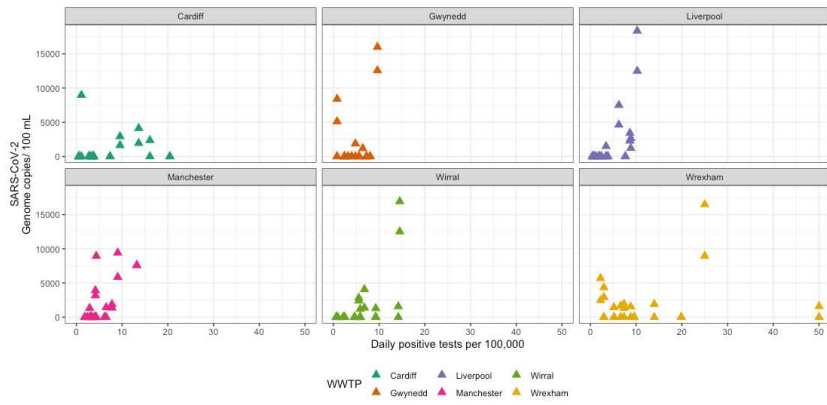
775 **Supplementary Fig. 2.** Proportion of tests that were above LoQ and LoD for (a) SARS-CoV-2  
 776 N1-gene q(RT-)PCR assay split by site and (b) samples assayed with both N1 and E gene  
 777 markers.



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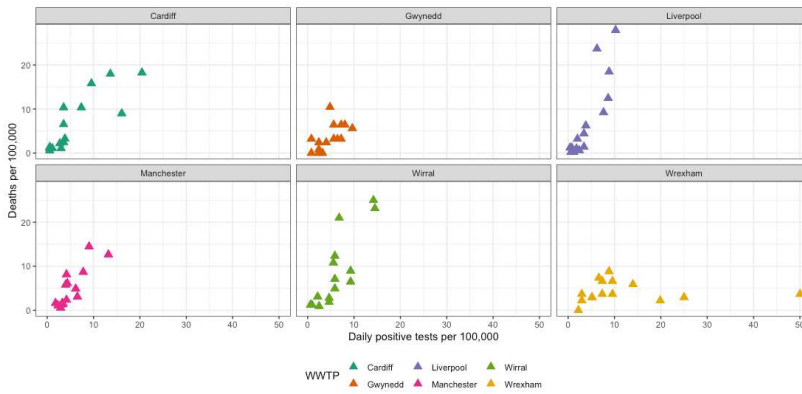
780 **Supplementary Fig. 3** Correlations of SARS-CoV-2 genome copies  $100 \text{ ml}^{-1}$  of wastewater with  
 781 local authority daily positive tests and COVID-19 related deaths per 100,000. SARS-CoV-2  
 782 wastewater concentrations were also normalised by daily flow (\*) and tests/ cases adjusted to  
 783 take account of differences between sewershed population equivalents and local authority  
 784 populations. These corrections had no substantial effect on correlations with only Manchester  
 785 seeing a slight decrease in correlation between SARS-CoV-2 wastewater concentrations and  
 786 tests/ deaths when corrected for the population size mismatch.

787 **a**



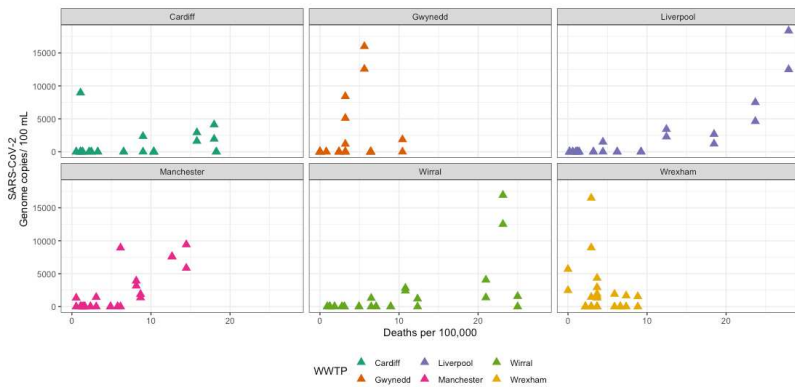
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789 **b**



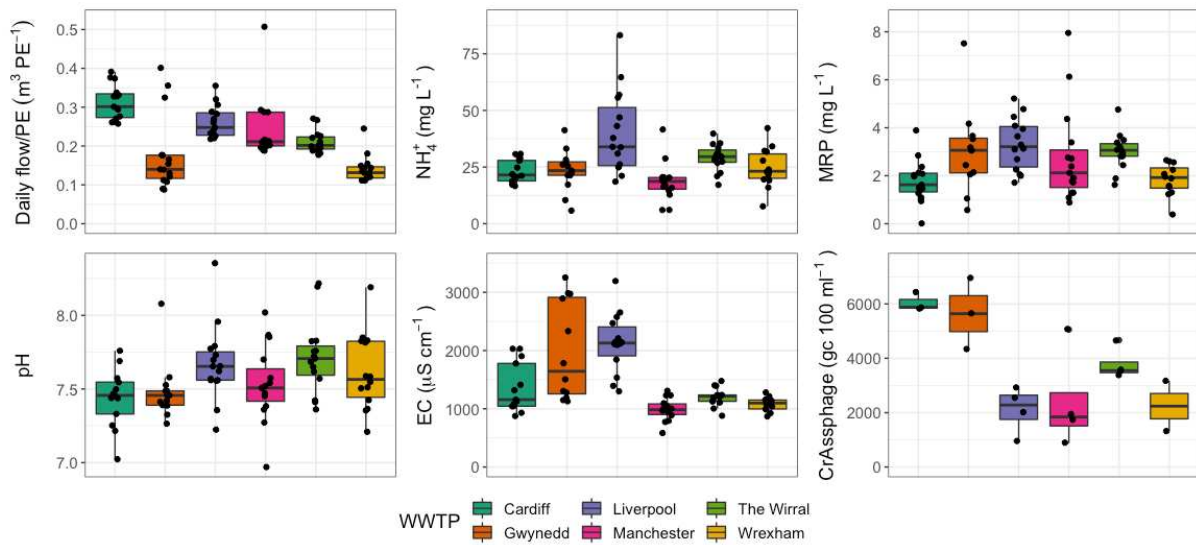
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791 **c**



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793 **Supplementary Fig. 4.** Comparisons of SARS-CoV-2 RNA wastewater concentration with  
 794 daily positive tests (a), COVID-19 related deaths (b) and between tests and deaths (c).



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797 **Supplementary Fig. 5** Site-specific variation in daily wastewater flow-rate (normalised by  
 798 population equivalent), chemical indicators [ $\text{NH}_4^+$ , molybdate-reactive phosphate (MRP), pH,  
 799 Electrical Conductivity (EC)] and a marker virus for human faecal loading (crAssphage) at six  
 800 urban wastewater treatment facilities over the course of the study. Boxes are bounded on the  
 801 first and third quartiles; horizontal lines denote medians. Black dots are outliers beyond the  
 802 whiskers, which denote  $1.5 \times$  the interquartile range.

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## 804 Supplemental References

- 805 Ahmed, W., Angel, N., Edson, J., Bibby, K., Bivins, A., O'Brien, J.W., Choi, P.M., Kitajima,  
 806 M., Simpson, S.L., Li, J., Tschärke, B., Verhagen, R., Smith, W.J.M., Zaugg, J.,  
 807 Dierens, L., Hugenholtz, P., Thomas, K.V., Mueller, J.F., 2020. First confirmed  
 808 detection of SARS-CoV-2 in untreated wastewater in Australia: A proof of concept for  
 809 the wastewater surveillance of COVID-19 in the community. *Sci. Total Environ.* 728,  
 810 138764. <https://doi.org/10.1016/j.scitotenv.2020.138764>  
 811 Arora, S., Nag, A., Sethi, J., Rajvanshi, J., Saxena, S., Shrivastava, S.K., Gupta, A.B., 2020.  
 812 Sewage surveillance for the presence of SARS-CoV-2 genome as a useful wastewater  
 813 based epidemiology (WBE) tracking tool in India. *medRxiv* 2020.06.18.20135277.  
 814 <https://doi.org/10.1101/2020.06.18.20135277>  
 815 Balboa, S., Mauricio-Iglesias, M., Rodríguez, S., Martínez-Lamas, L., Vasallo, F.J., Regueiro,  
 816 B., Lema, J.M., 2020. The fate of SARS-CoV-2 in wastewater treatment plants

817 out the sludge line as a suitable spot for incidence monitoring. medRxiv.  
818 <https://doi.org/10.1101/2020.05.25.20112706>

819 Bar Or, I., Yaniv, K., Shagan, M., Ozer, E., Erster, O., Mendelson, E., Mannasse, B., Shirazi,  
820 R., Kramarsky-Winter, E., Nir, O., Abu-Ali, H., Ronen, Z., Rinott, E., Lewis, Y.E.,  
821 Friedler, E.F., Paitan, Y., Bitkover, E., Berchenko, Y., Kushmaro, A., 2020. Regressing  
822 SARS-CoV-2 sewage measurements onto COVID-19 burden in the population: a proof-  
823 of-concept for quantitative environmental surveillance. medRxiv.  
824 <https://doi.org/10.1101/2020.04.26.20073569>

825 Centers for Disease Control and Prevention, 2020. Research Use Only 2019-Novel  
826 Coronavirus (2019-nCoV) Real-time RT-PCR Primers and Probes [WWW Document].  
827 URL <https://www.cdc.gov/coronavirus/2019-ncov/lab/rt-pcr-panel-primer-probes.html>  
828 (accessed 10.14.20).

829 Chavarria-Miró, G., Anfruns-Estrada, E., Guix, S., Paraira, M., Galofré, B., Sáanchez, G.,  
830 Pintó, R., Bosch, A., 2020. Sentinel surveillance of SARS-CoV-2 in wastewater  
831 anticipates the occurrence of COVID-19 cases. medRxiv.  
832 <https://doi.org/10.1101/2020.06.13.20129627>

833 Corman, V.M., Landt, O., Kaiser, M., Molenkamp, R., Meijer, A., Chu, D.K., Bleicker, T.,  
834 Brünink, S., Schneider, J., Schmidt, M.L., Mulders, D.G., Haagmans, B.L., Veer, B.,  
835 van der, Brink, S. van den, Wijsman, L., Goderski, G., Romette, J.-L., Ellis, J.,  
836 Zambon, M., Peiris, M., Goossens, H., Reusken, C., Koopmans, M.P., Drosten, C.,  
837 2020. Detection of 2019 novel coronavirus (2019-nCoV) by real-time RT-PCR.  
838 Eurosurveillance 25, 2000045. [https://doi.org/10.2807/1560-  
839 7917.ES.2020.25.3.2000045](https://doi.org/10.2807/1560-7917.ES.2020.25.3.2000045)

840 Dowle, M., Srinivasan, A., 2020. data.table: Extension of `data.frame`.

841 Gonzalez, R., Curtis, K., Bivins, A., Bibby, K., Weir, M.H., Yetka, K., Thompson, H., Keeling,  
842 D., Mitchell, J., Gonzalez, D., 2020. COVID-19 surveillance in Southeastern Virginia  
843 using wastewater-based epidemiology. Water Res. 186, 116296.  
844 <https://doi.org/10.1016/j.watres.2020.116296>

845 Green, H., Wilder, M., Middleton, F.A., Collins, M., Fenty, A., Gentile, K., Kmush, B., Zeng,  
846 T., Larsen, D.A., 2020. Quantification of SARS-CoV-2 and cross-assembly phage  
847 (crAssphage) from wastewater to monitor coronavirus transmission within  
848 communities. medRxiv. <https://doi.org/10.1101/2020.05.21.20109181>

849 Haramoto, E., Malla, B., Thakali, O., Kitajima, M., 2020. First environmental surveillance for  
850 the presence of SARS-CoV-2 RNA in wastewater and river water in Japan. Sci. Total  
851 Environ. 737, 140405. <https://doi.org/10.1016/j.scitotenv.2020.140405>

852 Harrell, F.E.Jr., 2020. Hmisc: Harrell Miscellaneous.

853 Hata, A., Honda, R., Hara-Yamamura, H., Meuchi, Y., 2020. Detection of SARS-CoV-2 in  
854 wastewater in Japan by multiple molecular assays-implication for wastewater-based  
855 epidemiology (WBE). medRxiv. <https://doi.org/10.1101/2020.06.09.20126417>

856 Kassambara, A., 2020. ggpubr: “ggplot2” Based Publication Ready Plots.

857 Kitajima, M., Oka, T., Takagi, H., Tohya, Y., Katayama, H., Takeda, N., Katayama, K., 2010.  
858 Development and application of a broadly reactive real-time reverse transcription-PCR  
859 assay for detection of murine noroviruses. J. Virol. Methods 169, 269–273.  
860 <https://doi.org/10.1016/j.jviromet.2010.07.018>

861 Kocamemi, B.A., Kurt, H., Hacioglu, S., Yarali, C., Saatci, A.M., Pakdemirli, B., 2020. First  
862 Data-Set on SARS-CoV-2 Detection for Istanbul Wastewaters in Turkey. medRxiv  
863 2020.05.03.20089417. <https://doi.org/10.1101/2020.05.03.20089417>

864 Kumar, M., Patel, A.K., Shah, A.V., Raval, J., Rajpara, N., Joshi, M., Joshi, C.G., 2020. First  
865 proof of the capability of wastewater surveillance for COVID-19 in India through  
866 detection of genetic material of SARS-CoV-2. *Sci. Total Environ.* 746, 141326.  
867 <https://doi.org/10.1016/j.scitotenv.2020.141326>

868 La Rosa, G., Mancini, P., Bonanno Ferraro, G., Veneri, C., Iaconelli, M., Bonadonna, L.,  
869 Lucentini, L., Suffredini, E., 2021. SARS-CoV-2 has been circulating in northern Italy  
870 since December 2019: Evidence from environmental monitoring. *Sci. Total Environ.*  
871 750, 141711. <https://doi.org/10.1016/j.scitotenv.2020.141711>

872 Lemon, J., 2006. Plotrix: a package in the red light district of R. *R-News* 6, 8–12.

873 Medema, G., Heijnen, L., Elsinga, G., Italiaander, R., Brouwer, A., 2020. Presence of SARS-  
874 Coronavirus-2 RNA in Sewage and Correlation with Reported COVID-19 Prevalence  
875 in the Early Stage of the Epidemic in The Netherlands. *Environ. Sci. Technol. Lett.* 7,  
876 511–516. <https://doi.org/10.1021/acs.estlett.0c00357>

877 Nemudryi, A., Nemudraia, A., Wiegand, T., Surya, K., Buyukyoruk, M., Cicha, C.,  
878 Vanderwood, K.K., Wilkinson, R., Wiedenheft, B., 2020. Temporal Detection and  
879 Phylogenetic Assessment of SARS-CoV-2 in Municipal Wastewater. *Cell Rep. Med.* 1,  
880 100098. <https://doi.org/10.1016/j.xcrm.2020.100098>

881 Ogle, D.H., Wheeler, P., Dinno, A., 2020. FSA: Fisheries Stock Analysis.

882 Pebesma, E., 2018. Simple Features for R: Standardized Support for Spatial Vector Data. *R J.*  
883 10, 439–446. <https://doi.org/10.32614/RJ-2018-009>

884 Peccia, J., Zulli, A., Brackney, D.E., Grubaugh, N.D., Kaplan, E.H., Casanovas-Massana, A.,  
885 Ko, A.I., Malik, A.A., Wang, D., Wang, M., Warren, J.L., Weinberger, D.M., Arnold,  
886 W., Omer, S.B., 2020. Measurement of SARS-CoV-2 RNA in wastewater tracks  
887 community infection dynamics. *Nat. Biotechnol.* 38, 1164–1167.  
888 <https://doi.org/10.1038/s41587-020-0684-z>

889 Prado, T., Fumian, T.M., Mannarino, C.F., Maranhão, A.G., Siqueira, M.M., Miagostovich,  
890 M.P., Prado, T., Fumian, T.M., Mannarino, C.F., Maranhão, A.G., Siqueira, M.M.,  
891 Miagostovich, M.P., 2020. Preliminary results of SARS-CoV-2 detection in sewerage  
892 system in Niterói municipality, Rio de Janeiro, Brazil. *Mem. Inst. Oswaldo Cruz* 115.  
893 <https://doi.org/10.1590/0074-02760200196>

894 Randazzo, W., Truchado, P., Cuevas-Ferrando, E., Simón, P., Allende, A., Sánchez, G., 2020.  
895 SARS-CoV-2 RNA in wastewater anticipated COVID-19 occurrence in a low  
896 prevalence area. *Water Res.* 181, 115942. <https://doi.org/10.1016/j.watres.2020.115942>

897 Rimoldi, S.G., Stefani, F., Gigantiello, A., Polesello, S., Comandatore, F., Mileto, D., Maresca,  
898 M., Longobardi, C., Mancon, A., Romeri, F., Pagani, C., Cappelli, F., Roscioli, C.,  
899 Moja, L., Gismondo, M.R., Salerno, F., 2020. Presence and infectivity of SARS-CoV-2  
900 virus in wastewaters and rivers. *Sci. Total Environ.* 744, 140911.  
901 <https://doi.org/10.1016/j.scitotenv.2020.140911>

902 Sherchan, S.P., Shahin, S., Ward, L.M., Tandukar, S., Aw, T.G., Schmitz, B., Ahmed, W.,  
903 Kitajima, M., 2020. First detection of SARS-CoV-2 RNA in wastewater in North  
904 America: A study in Louisiana, USA. *Sci. Total Environ.* 743, 140621.  
905 <https://doi.org/10.1016/j.scitotenv.2020.140621>

906 Slowikowski, K., 2020. ggrepel: Automatically Position Non-Overlapping Text Labels with  
907 “ggplot2.”

908 South, A., 2017. rnatualearth: World Map Data from Natural Earth.

909 South, A., 2011. rworldmap: A New R package for Mapping Global Data. *R J.* 3, 35–43.

910 Stachler, E., Kelty, C., Sivaganesan, M., Li, X., Bibby, K., Shanks, O.C., 2017. Quantitative  
911 CrAssphage PCR Assays for Human Fecal Pollution Measurement. *Environ. Sci.*  
912 *Technol.* 51, 9146–9154. <https://doi.org/10.1021/acs.est.7b02703>

- 913 Trottier, J., Darques, R., Ait Mouheb, N., Partiot, E., Bakhache, W., Deffieu, M.S., Gaudin, R.,  
914 2020. Post-lockdown detection of SARS-CoV-2 RNA in the wastewater of Montpellier,  
915 France. *One Health* 10, 100157. <https://doi.org/10.1016/j.onehlt.2020.100157>
- 916 Wei, T., Simko, V., 2017. R package “corrplot”: Visualization of a Correlation Matrix.
- 917 Westhaus, S., Weber, F.-A., Schiwy, S., Linnemann, V., Brinkmann, M., Widera, M., Greve,  
918 C., Janke, A., Hollert, H., Wintgens, T., Ciesek, S., 2021. Detection of SARS-CoV-2 in  
919 raw and treated wastewater in Germany – Suitability for COVID-19 surveillance and  
920 potential transmission risks. *Sci. Total Environ.* 751, 141750.  
921 <https://doi.org/10.1016/j.scitotenv.2020.141750>
- 922 Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L.D., François, R., Golemund,  
923 G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T.L., Miller, E., Bache, S.M.,  
924 Müller, K., Ooms, J., Robinson, D., Seidel, D.P., Spinu, V., Takahashi, K., Vaughan,  
925 D., Wilke, C., Woo, K., Yutani, H., 2019. Welcome to the tidyverse. *J. Open Source*  
926 *Softw.* 4, 1686. <https://doi.org/10.21105/joss.01686>
- 927 Wilke, C.O., 2020. cowplot: Streamlined Plot Theme and Plot Annotations for “ggplot2.”
- 928 Wu, F., Xiao, A., Zhang, J., Gu, X., Lee, W.L., Kauffman, K., Hanage, W., Matus, M., Ghaeli,  
929 N., Endo, N., Duvallet, C., Moniz, K., Erickson, T., Chai, P., Thompson, J., Alm, E.,  
930 2020. SARS-CoV-2 titers in wastewater are higher than expected from clinically  
931 confirmed cases. *medRxiv* 2020.04.05.20051540.  
932 <https://doi.org/10.1101/2020.04.05.20051540>
- 933 Wurtzer, S., Marechal, V., Mouchel, J.-M., Maday, Y., Teyssou, R., Richard, E., Almayrac,  
934 J.L., Moulin, L., 2020. Evaluation of lockdown impact on SARS-CoV-2 dynamics  
935 through viral genome quantification in Paris wastewaters.  
936 <https://doi.org/10.1101/2020.04.12.20062679>
- 937 Zeileis, A., Grothendieck, G., 2005. zoo: S3 Infrastructure for Regular and Irregular Time  
938 Series. *J. Stat. Softw.* 14, 1–27. <https://doi.org/10.18637/jss.v014.i06>

939

940

941

942

943

944