

This is a repository copy of *Growth, development, and phenotypic spectrum of individuals with deletions of 2q33.1 involving SATB2.*

White Rose Research Online URL for this paper: https://eprints.whiterose.ac.uk/174966/

Version: Accepted Version

Article:

Zarate, Y.A., Bosanko, K.A., Thomas, M.A. et al. (18 more authors) (2021) Growth, development, and phenotypic spectrum of individuals with deletions of 2q33.1 involving SATB2. Clinical Genetics, 99 (4). pp. 547-557. ISSN 0009-9163

https://doi.org/10.1111/cge.13912

This is the peer reviewed version of the following article: Zarate, YA, Bosanko, KA, Thomas, MA, et al. Growth, development, and phenotypic spectrum of individuals with deletions of 2q33.1 involving SATB2. Clinical Genetics. 2021; 99: 547–557., which has been published in final form at https://doi.org/10.1111/cge.13912. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Use of Self-Archived Versions.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Growth, development, and phenotypic spectrum of individuals with deletions of 2q33.1 involving *SATB2*

Short title: 2q33.1 deletions and SATB2-Associated syndrome

Yuri A. Zarate¹, Katherine A. Bosanko¹, Mary Ann Thomas², David T. Miller³, Kristina Cusmano-Ozog⁴, Antonio Martinez-Monseny⁵, Cynthia J. Curry⁶, John M. Graham Jr⁷, Lea Velsher⁸, Mir Reza Bekheirnia⁹, Veronica Seidel¹⁰, Demitrios Dedousis¹¹, Anna L. Mitchell¹¹, Amy M. DiMarino¹², Angelika Riess¹³, Meena Balasubramanian¹⁴, Jennifer L. Fish¹⁵, Aisling R. Caffrey¹⁶, Nicole Fleischer¹⁷, Tyler Mark Pierson¹⁸, Ronald V. Lacro¹⁹ ¹Section of Genetics and Metabolism, University of Arkansas for Medical Sciences, Little Rock, AR, USA

²Departments of Medical Genetics and Pediatrics, Cumming School of Medicine,

University of Calgary, Calgary, Alberta, Canada

³Division of Genetics and Genomics, Boston Children's Hospital, Boston, MA, USA

⁴Department of Pathology, Stanford University Medical Center, Stanford, CA, USA

⁵Department of Clinical Genetics and Rare Disease Paediatric Unit, Hospital Sant Joan de

Déu, University of Barcelona, Barcelona, Spain

⁶Genetic Medicine, Department of Pediatrics, University of California, San

Francisco/Fresno, Fresno, CA, USA

⁷Medical Genetics, Department of Pediatrics, Cedars-Sinai Medical Center, Los Angeles, CA, USA

⁸North York General, Toronto, Canada

⁹Departments of Pediatrics and Molecular & Human Genetics, Baylor College of Medicine, Houston, TX, USA

¹⁰Clinical Genetics, Department of Pediatrics, HGU Gregorio Marañón, Madrid, Spain

¹¹University Hospitals Center for Human Genetics, Cleveland, OH, USA

¹²UH Rainbow Babies and Children's Hospital, Cleveland, OH, USA

¹³Institute of Medical Genetics and Applied Genomics, Tuebingen, Germany

¹⁴Sheffield Clinical Genetics Service, Sheffield Children's NHS Foundation Trust, UK

¹⁵Department of Biological Sciences, University of Massachusetts Lowell, Lowell,

Massachusetts

¹⁶Health Outcomes, College of Pharmacy, University of Rhode Island, Kingston, RI,

USA

¹⁷FDNA Inc, Boston, MA, USA

¹⁸Departments of Pediatrics and Neurology & The Board of Governors Regenerative

Medicine Institute, Cedars Sinai Medical Center, Los Angeles, CA, USA

¹⁹Department of Cardiology, Boston Children's Hospital and Department of Pediatrics,

Harvard Medical School, Boston, MA, USA

Correspondence To:

Yuri A. Zarate, M.D. Arkansas Children's Hospital 1 Children's Way; Slot 512-22 Little Rock, AR 72202 Telephone: 501-364-2971, <u>yazarate@uams.edu</u> Fax: 501-364-1564

Acknowledgements

The authors are grateful to all participating families. TMP is supported by the Cedars-Sinai institutional funding program, the Cedars-Sinai Diana and Steve Marienhoff Fashion Industries Guild Endowed Fellowship in Pediatric Neuromuscular Diseases and the Fashion Industries Guild Endowed Fellowship for the Undiagnosed Diseases Program.

Conflicts of Interest: N.F. is an employee of FDNA Inc. All other authors declare no conflicts of interest.

Data Availability: The data that support the findings of this study are available on request from the corresponding author. The data are not publicly available due to privacy or ethical restrictions.

ABSTRACT

SATB2-Associated syndrome (SAS) is an autosomal dominant, multisystemic, neurodevelopmental disorder due to alterations in SATB2 at 2q33.1. A limited number of individuals with 2q33.1 contiguous deletions encompassing SATB2 (Δ SAS) have been described in the literature. We describe 17 additional individuals with Δ SAS, review the phenotype of 33 previously published individuals with 2q33.1 deletions (n=50, mean age= 8.5 ± 7.8 years), and provide a comprehensive comparison to individuals with other molecular mechanisms that result in SAS (non- Δ SAS). Individuals in the Δ SAS group were often underweight for age (20/41=49%) with a progressive decline in weight (95%)CI=-2.3 to -1.1, p<0.0001) and height (95% CI=-2.3 to -1.0, p<0.0001) Z-score means from birth to last available measurement. Δ SAS individuals were often noted to a broad spectrum of facial dysmorphism. A composite image of Δ SAS individuals generated by automated image analysis was distinct as compared to matched controls and non- ΔSAS individuals. We also present additional genotype-phenotype correlations for individuals in the Δ SAS group such as an increased risk for a rtic root/ascending a rta dilation and primary pulmonary hypertension for those individuals with contiguous gene deletions that include COL3A1/COL5A2 and BMPR2, respectively. Based on these findings, we provide additional care recommendations for individuals with Δ SAS variants.

Key Words: *SATB2*-Associated syndrome, *SATB2*, 2q33.1 deletion, facial recognition technology, genotype-phenotype correlation, Glass syndrome

INTRODUCTION

SATB2-Associated syndrome (SAS; Glass syndrome, OMIM 612313) is a multisystemic disorder caused by variants in *SATB2* at 2q33.1.¹ While numerous coding variants have been reported and are thought to be the most common molecular etiology, other documented molecular mechanisms include intragenic deletions and duplications, translocations interrupting *SATB2* or its regulatory elements, and contiguous deletions.²⁻⁴

Since the original description of a 16-year-old male with an interstitial deletion of 2q32.2-2q33.1⁵, several other individuals with deletions encompassing *SATB2* (henceforth referred to as Δ SAS) have been reported. This has led to the separate designation of 2q33.1 microdeletion syndrome or 2q32-q33 deletion syndrome. Nevertheless, as more individuals with SAS were reported, the most consistent features of SAS continue to consist of neurodevelopmental disabilities, behavioral abnormalities, and craniofacial anomalies, regardless of the molecular mechanism.^{1,2,6,7} A recent review of 17 individuals with Δ SAS revealed a phenotypic spectrum: developmental delay with significantly impaired speech, dental abnormalities (mainly with abnormal teeth shape/size and crowding), behavioral anomalies (hyperactivity and agitation/aggression being most frequent), feeding difficulties, hypotonia, cleft palate, and abnormal neuroimaging.¹ Other features such as facial dysmorphism, clinical seizures, and skeletal anomalies were also reported at variable frequencies.

Despite the limited number of described individuals with Δ SAS, there have been some attempts to identify genotype-phenotype correlations. Pre- or post-natal growth retardation, hypotonia, genitourinary anomalies, congenital cardiac defects, and some ectodermal manifestations such as thin skin or reduced subcutaneous fat were reported to

be more prevalent or exclusively seen in individuals with Δ SAS as opposed to intragenic *SATB2* abnormalities. Interestingly, all have had consistently similar issues with speech development.^{1,6,8}

In this report, we describe 17 additional individuals with 2q33.1 contiguous deletions and provide a review of all previously described cases. We present a comprehensive overview with more detail about the frequency of the main phenotypic features of Δ SAS and how they compare to other molecular mechanisms such as intragenic *SATB2* alterations.

MATERIALS AND METHODS

Individuals with 2q33.1 deletions that include *SATB2* entirely or with breakpoints near the 3' or 5' end of *SATB2* were included. All new reported individuals agreed to share clinical information and were enrolled under a research clinical registry protocol approved by the Institutional Review Board of the University of Arkansas for Medical Sciences and comforming with the Declaration of Helsinki. For all individuals, medical records including laboratory results were reviewed. Supplementary information was also obtained by parental report through a REDCapTM questionnaire. All families who shared photographs signed consent for publication. Individuals SATB2-85 and SATB2-168 were briefly presented in previous publications.^{9,10}

Molecular cytogenetic studies

Several different chromosomal microarray platforms were used over time to analyze genomic DNA extracted from peripheral blood over time. Earlier patients were tested with oligonucleotide-based platforms (Agilent Human Genome 44K and 180K oligonucleotide arrays, Agilent Technologies, Santa Clara, CA; Oxford Gene Technology

CytoSure[™] 60K Oligonucleotide array, Oxford, UK), while more recent cases were evaluated using the whole genome SNP arrays (Affymetrix, Santa Clara, CA). Furthermore, microarray findings were confirmed with FISH analysis whenever possible. Coordinates for older studies were converted to the GRCh37/h19 build.

Anthropometric values and growth charts

Available height, weight, and head circumference measurements were extracted from medical records. Reference data and Z-scores for height, weight, and head circumference were obtained from 2006 World Health Organization (WHO) international growth standards for individuals aged 0 to 24 months and adjusted by gestational age accordingly, and the 2000 CDC growth curves for children and adolescents from 2 to 20 years of age.¹¹

Statistical analysis

Individuals' demographic and clinical milestones were characterized using medians and means according to the genetic alteration type. For genotype-phenotype correlation, the categorical analysis was performed using Fisher's exact test. A paired t-test was used for prospective analysis of anthropometric measurements over time. Analyses were performed using SAS 9.4 software (SAS Institute, Cary NC).

RESULTS

Detailed clinical information from 17 novel individuals with deletions that included *SATB2* entirely (6 males and 11 females, mean age of 6.8 years, age range of 6 months to 34 years), along with the cumulative prevalence of clinical features of all 50 individuals (17 from this cohort and 33 individuals previously reported in the literature) are presented in Table 1.^{6,12-21} Two individuals from the literature with 2q33.1 deletions that did not

include *SATB2* but had breakpoints near it were included in the Δ SAS group given the known presence of regulatory elements at least on the 3' end of *SATB2* and their overlapping phenotypic features with those reported in SAS.¹² Clinical information for all 50 individuals is described in the following sections with particular emphasis on phenotypic features that are unique or have not been explored in detail for individuals with Δ SAS.

Cytogenetic alterations

Through a variety of different molecular cytogenetic platforms utilized in clinical settings, the mean deletion size for all individuals was 11Mb (range 600kb to 26.3Mb), with 16 individuals having deletions greater than 15Mb (Figure 1). Patient 1 and patient 6 reported by Balasubramanian et al. had deletions falling 20kb and 220kb from the 5' and 3' end of *SATB2* respectively.¹² Segregation analysis in several families revealed *de novo* status in 17/18 (94%) individuals, with a single case resulting from a balanced maternal chromosomal rearrangement involving chromosomes 2 and 21.

Phenotypes

Individuals with Δ SAS, similar to non- Δ SAS, had the core clinical features of developmental delay with severe speech delay, behavioral abnormalities, and craniofacial anomalies (Table 1, Table 2). Developmental delay was present in all individuals with delayed acquisition of early milestones and speech limited to 10 words or less in 69% (22/32 with reported data) of individuals 4 years or older. Speech ability was not correlated with the deletion size (data not shown). The frequency of other features was in concordance with the reported prevalence in non- Δ SAS individuals: behavioral abnormalities (85%), hypotonia (67%), abnormal neuroimaging (57%) ophthalmologic

anomalies (56%), cleft palate (50%), and epilepsy (28%).³ Dental anomalies in the Δ SAS cohort were frequently documented (96%) of en with distinctive large and abnormally shaped upper central incisors. Several subjects underwent bone density scans with low bone density reported in 44% (4/9).

Perinatal history, growth, and nutrition

The mean gestational age was 38 weeks (range 32 to 41 weeks). The mean Z-scores (adjusted for gestational age) for birth weight, length, and head circumference were: -0.47 (n=49; -4.0 to +2.9), +0.22 (n=33, -2.4 to +4.4), and -0.32 (n=20, -3.8 to +2.83), respectively. Feeding difficulties at any age were reported in 80% (40/50) of individuals, with 24% (8/33) requiring a gastrostomy tube. At their most recent visit, almost half of the individuals were underweight for age (49%; 20/41), often with short stature (41%; 15/37) and microcephaly (36%; 12/33). Detailed analysis of postnatal growth revealed that for individuals with birth and postnatal values available for review (at least 6 months apart), there was a tendency for decreased Z-score values with age for weight and height. From birth until the latest measurement, there was a decrease in Z-score means for weight and height of -1.7 (95% CI=-2.3 to -1.1, p<0.0001) and -1.7 (95% CI=-2.3 to -1.0, p<0.0001), respectively (Figure 2). A higher frequency of low weight was seen for individuals with deletions larger than 5Mb (62% vs 21%, p=0.02). Only one individual (SATB2-147) received growth hormone supplementation briefly with no improvement noted before discontinuation.

Cardiovascular and respiratory

Several individuals had cardiovascular defects (26%; 11/42) with five individuals having septal defects alone or in combination with other findings. Other less common findings

included patent ductus arteriosus persisting beyond the neonatal period, vascular ring, and valve dysplasias that were documented beyond the neonatal period.

Deletions involving BMPR2

Fifty percent (11/22) of Δ SAS individuals with deletions that included *BMPR2* had echocardiograms performed and 27% (3/11) had pulmonary hypertension diagnosed during the first few weeks of life (one individual had a ventricular septal defect that required surgical repair and it is unclear whether pulmonary hypertension persisted after repair of the septal defect, the other two lacked related severe congenital heart defects) and were diagnosed with pulmonary hypertension. We highlight two individuals from the new cohort below.

Individual SATB2-099 is a 2-year-old female diagnosed with a 2q32.1q33.3 deletion at 2 weeks of age. Her large deletion includes *COL3A1*, *COL5A2*, and *BMPR2*. After an initial normal echocardiogram at 1 month of age, a repeat study at 9 months of age revealed pulmonary hypertension that was persistent on her most recent evaluation at 2 years of age (moderate based on echocardiogram). To date, she has not required medical or surgical treatment for her pulmonary hypertension. Because bronchiectasis has not been reported to be associated with SAS, further clinical genetic testing was conducted identifying a single variant (p.Phe508del) in *CFTR* thought to be insufficient to explain the pulmonary phenotype.

Individual SATB2-193 is a 7-month old male, former 36-week infant who presented with desaturation, cyanosis, hypoglycemia, and feeding difficulty. His deletion includes *BMPR2*. Postnatal echocardiogram on day of life 1 showed right ventricular pressures estimated to be about 2/3 systemic consistent with pulmonary hypertension. He

required ventilator support, dopamine for persistently low systemic blood pressures, and veno-arterial extracorporeal membrane oxygenation (ECMO) for the first 9 days of life. At 12 days of age, despite his clinical improvement, his persistent pulmonary hypertension required treatment with sildenafil that continued until his discharge from the NICU at 2 months of age. Over the following months, he had persistent episodes of cyanosis, arching, and desaturation consistent with pulmonary hypertensive spells and confirmed by right heart catheterization. Bosentan and amlodipine were added to his pulmonary hypertension management plan. His pulmonary hypertension is persistent to his current age.

Deletions involving COL3A1 and COL5A2

Of the 10 individuals reviewed in the present study with deletions that included both of these collagen genes, six had documented echocardiograms performed and two had aortic dilation. Individual SATB2-184 was noted to have mild aortic dilation at 2 years of age (Z-score +2.13) with no other structural heart defect. Individual SATB2-118 is a 9-year old female who was born full-term to a 26-year-old healthy mother. An echocardiogram on day 6 of life showed a patent foramen ovale versus small secundum atrial septal defect, a trivial, physiologic patent ductus arteriosus, aortic root dilation (Zscore +3.82), and normal ascending aorta dimension. Echocardiogram at age 16 months revealed aortic root dilation (Z-score +4.0) and normal ascending aorta dimension. Over time, she was diagnosed with tortuosity of craniocervical vasculature, severe aortic root dilation, and moderate ascending aorta dilation, with stable Z-scores over several years (+5 - 6 for the aortic root and +4 – 5 for the ascending aorta). She is currently on losartan and atenolol therapy without evidence of pulmonary artery hypertension.

Other systemic manifestations

Four individuals were reported to have pectus type deformity (3 excavatum, 1 carinatum) and three others had scoliosis (only one of these seven individuals had a deletion encompassing *COL3A1* and *COL5A2*). In addition to five individuals with inguinal hernias, no other consistent genitourinary anomalies were described. Single instances of colobomas, cataracts, metopic synostosis, and congenital diaphragmatic hernia were also reported. Ectodermal manifestations such as thin skin, reduced subcutaneous fat, and fine/sparse hair were reported in earlier cases but not consistently found in other recently described individuals, including the 17 novel cases presented here.

Facial phenotype

Facial dysmorphic features were described in 90% of individuals (45/50) but with more variability than previously reported in SAS.^{3,6,7}Besides a thin vermilion of the upper lip, long and flat philtrum, and deeply set eyes, other features noted include a prominent forehead with a receding anterior hairline, a pointed chin, and triangular appearance of the lower face with a narrow jaw (Figure 3). Of note, overlapping facial features to those described in Sotos syndrome were seen in 2 inviduals (individual SATB2-093 and individual SATB2-147). For additional objectivity, 2D photographs were analyzed using Face2Gene image analysis (FDNA Inc., USA, v19.1.12), a tool that has been shown to add value to phenotypic evaluations in clinical genetics.²² Images from 24 individuals with Δ SAS were noted to be significantly different from 24 age-, ethnicity-, and gendermatched controls (area under the curve 0.89, p-value = 0.008). When compared to similarly matched non- Δ SAS group (N=59), the Δ SAS cohort were very close to being

different in a statistically significant manner (area under the curve 0.706, p-value = 0.06) (Figure 3).

DISCUSSION

SAS caused by contiguous gene deletions as compared to other molecular mechanisms contributes to approximately 20% of SAS cases (https://satb2gene.com/pros-molecular-data/). Through the comprehensive analysis of 50 Δ SAS individuals (17 that were previously unreported), we determined that the cardinal findings include severe speech delay, behavioral issues, cleft palate, dental anomalies, hypotonia, ophthalmologic abnormalities, and epilepsy. Of note, we present additional clinical features that were more prevalent or unique to our - Δ SAS cohort. These features are important in that they require additional counseling, surveillance, or interventions.

Detailed anthropometric measurement analysis has not previously been conducted in SAS. Approximately 2/3 of individuals with Δ SAS were found to have growth retardation, which was twice as common when compared to non- Δ SAS individuals. This growth retardation occurred despite the lack of any difference in the frequency of cleft palate or reported feeding difficulties. Indeed, Z-score values for weight and height showed a significant decrease over time in the Δ SAS cohort with individuals with larger deletions having a higher frequency of low weight. This suggests that other genes in the contiguous deleted segment may be influencing growth parameters in the Δ SAS cohort. Generation of dedicated SAS growth charts may assist in predicting the expected growth patterns in the Δ SAS population as compared to non- Δ SAS individuals. This information will assist in determining whether aggressive feeding interventions (including

gastrostomy tubes) may be required to assist the child to attain a normalized weight for better growth and for airway protection.

Individuals with Δ SAS had variable facial dysmorphisms beyond the previously described features seen in non- Δ SAS. Some individuals in the Δ SAS cohort displayed changes in the lower face configuration which could be related to other contiguous genes included in their deletions or could be a reflection of a true *SATB2*-dosage effect that supports haploinsufficiency in Δ SAS individuals. In mice, jaw development is *Satb2*dosage sensitive and although the loss of *Satb2* affects both upper and lower jaws, previous studies have shown a predominant compromise of the distal part of the mandible independently of the proximal region.^{23,24}

The deletions encompassing the *SATB2* gene and involving the 2q32-q33 region were quite diverse in size and makeup. Some of the genes located in this region that are linked to potentially relevant dominant phenotypes and with high probability of being loss-of-function intolerant: *HSPD1* (pLI=0.99), *HECW2* (pLI=1), *COL3A1* (pLI=1), *COL5A2* (pLI=1), and *BMPR2* (pLI=1). The full impact that results from the haploinsufficiency of these genes in Δ SAS individuals, genes remains to be elucidated.

Hemizygous deletion of *COL3A1* with resulting haploinsufficiency has been associated with cardiovascular complications. Meienberg et al. described a family with a 3.4Mb deletion at 2q31.1q32.3 and that included both *COL3A1* and *COL5A2* (but not *SATB2*). All seven individuals displayed features of a connective tissue disorder characterized by some overlapping clinical signs of vascular and classic Ehlers-Danlos syndromes. In two of these subjects, fatal aortic dissections (thoracic and abdominal) occurred during adulthood and there was evidence of involvement of medium-sized

arteries.²⁵ Based on the individuals described in this report and the previously described family by Meienberg et al., we recommend baseline echocardiographic evaluation for those individuals with deletions that include the *COL3A1* and/or the *COL5A2*, with consideration for ongoing yearly surveillance of the thoracic and abdominal aorta for aortic aneurysms and risk of dissection. Furthermore, we recommend having a low threshold to obtain detailed imaging of the vascular tree depending on echocardiographic results, as well as clinical symptoms such as headache, stroke, or abdominal pain, or physical findings such as decreased peripheral pulses, asymmetric blood pressure measurements, or abdominal bruits.

Primary pulmonary hypertension (PPH) is a rare and often fatal disease characterized by pulmonary vascular cell proliferation and a sustained elevation in mean pulmonary artery pressure. Heterozygous mutations (including whole gene deletions) resulting in haploinsufficiency in *BMPR2* are often found in children and adults with idiopathic and familial cases of PPH.²⁶⁻²⁹ Considering the known reduced penetrance and variable expressivity of BMPR2-related PPH and based on the cases here described, we suggest that Δ SAS individuals with deletions containing *BMPR2* should also have a baseline echocardiogram at diagnosis and should then continue to follow published expert guidelines to accurately identify PPH.³⁰

In summary, in addition to the core features of SAS such as developmental delay, behavioral issues, and craniofacial anomalies, Individuals with Δ SAS also displayed a higher tendency for growth retardation, variable facial dysmorphism, and increased risk for pulmonary hypertension and aortic dilation/dissection. Larger studies that look at the

impact of contiguous genes deleted are needed to better delineate the phenotypic

differences among Δ SAS individuals.

RFERENCES

- 1. Zarate YA, Fish JL. SATB2-associated syndrome: Mechanisms, phenotype, and practical recommendations. *Am J Med Genet A*. 2017;173:327-337.
- Zarate YA, Kaylor J, Fish J. SATB2-Associated Syndrome. In: Adam MP, Ardinger HH, Pagon RA, et al., eds. *GeneReviews(R)*. Seattle (WA)1993.
- Zarate YA, Bosanko KA, Caffrey AR, et al. Mutation update for the SATB2 gene. *Hum Mutat.* 2019;40:1013-1029.
- Rainger JK, Bhatia S, Bengani H, et al. Disruption of SATB2 or its long-range cis-regulation by SOX9 causes a syndromic form of Pierre Robin sequence. *Hum Mol Genet.* 2014;23:2569-2579.
- Glass IA, Swindlehurst CA, Aitken DA, McCrea W, Boyd E. Interstitial deletion of the long arm of chromosome 2 with normal levels of isocitrate dehydrogenase. *J Med Genet.* 1989;26:127-130.
- Zarate YA, Smith-Hicks CL, Greene C, et al. Natural history and genotypephenotype correlations in 72 individuals with SATB2-associated syndrome. *Am J Med Genet A*. 2018;176:925-935.
- Bengani H, Handley M, Alvi M, et al. Clinical and molecular consequences of disease-associated de novo mutations in SATB2. *Genet Med.* 2017;19:900-908.
- Thomason A, Pankey E, Nutt B, Caffrey AR, Zarate YA. Speech, language, and feeding phenotypes of SATB2-associated syndrome. *Clin Genet*. 2019;96:485-492.

- Cotton AP, Gokarakonda S, Caffrey AR, Zarate YA, Kumar N. Behavioral phenotype and sleep problems in SATB2-associated syndrome. *Dev Med Child Neurol.* 2019.
- Lewis H, Samanta D, Orsell JL, et al. Epilepsy and Electroencephalographic Abnormalities in SATB2-Associated Syndrome. *Pediatr Neurol.* 2020.
- Chou JH, Roumiantsev S, Singh R. PediTools Electronic Growth Chart Calculators: Applications in Clinical Care, Research, and Quality Improvement. J Med Internet Res. 2020;22:e16204.
- Balasubramanian M, Smith K, Basel-Vanagaite L, et al. Case series: 2q33.1 microdeletion syndrome--further delineation of the phenotype. *J Med Genet*. 2011;48:290-298.
- Gregoric Kumperscak H, Krgovic D, Vokac NK. Specific behavioural phenotype and secondary cognitive decline as a result of an 8.6 Mb deletion of 2q32.2q33.1. *J Int Med Res.* 2016;44:395-402.
- Mc Cormack A, Taylor J, Gregersen N, George AM, Love DR. Delineation of 2q32q35 deletion phenotypes: two apparent "proximal" and "distal" syndromes. *Case Rep Genet.* 2013;2013:823451.
- 15. Rifai L, Port-Lis M, Tabet AC, et al. Ectodermal dysplasia-like syndrome with mental retardation due to contiguous gene deletion: further clinical and molecular delineation of del(2q32) syndrome. *Am J Med Genet A*. 2010;152A:111-117.
- 16. Tomaszewska A, Podbiol-Palenta A, Boter M, et al. Deletion of 14.7 Mb
 2q32.3q33.3 with a marfanoid phenotype and hypothyroidism. *Am J Med Genet A.* 2013;161A:2347-2351.

- Urquhart J, Black GC, Clayton-Smith J. 4.5 Mb microdeletion in chromosome band 2q33.1 associated with learning disability and cleft palate. *Eur J Med Genet*. 2009;52:454-457.
- Van Buggenhout G, Van Ravenswaaij-Arts C, Mc Maas N, et al. The del(2)(q32.2q33) deletion syndrome defined by clinical and molecular characterization of four patients. *Eur J Med Genet*. 2005;48:276-289.
- Yamada M, Uehara T, Suzuki H, et al. SATB2-associated syndrome in patients from Japan: Linguistic profiles. *Am J Med Genet A*. 2019.
- Yu N, Shin S, Lee KA. First Korean case of SATB2-associated 2q32-q33 microdeletion syndrome. *Ann Lab Med.* 2015;35:275-278.
- de Ravel TJ, Balikova I, Thiry P, Vermeesch JR, Frijns JP. Another patient with a de novo deletion further delineates the 2q33.1 microdeletion syndrome. *Eur J Med Genet.* 2009;52:120-122.
- 22. Gurovich Y, Hanani Y, Bar O, et al. Identifying facial phenotypes of genetic disorders using deep learning. *Nat Med.* 2019;25:60-64.
- Britanova O, Depew MJ, Schwark M, et al. Satb2 haploinsufficiency phenocopies
 2q32-q33 deletions, whereas loss suggests a fundamental role in the coordination
 of jaw development. *Am J Hum Genet.* 2006;79:668-678.
- 24. Fish JL, Villmoare B, Kobernick K, et al. Satb2, modularity, and the evolvability of the vertebrate jaw. *Evol Dev.* 2011;13:549-564.
- 25. Meienberg J, Rohrbach M, Neuenschwander S, et al. Hemizygous deletion of COL3A1, COL5A2, and MSTN causes a complex phenotype with aortic

dissection: a lesson for and from true haploinsufficiency. *Eur J Hum Genet*. 2010;18:1315-1321.

- Jang AY, Kim BG, Kwon S, et al. Prevalence and clinical features of bone morphogenetic protein receptor type 2 mutation in Korean idiopathic pulmonary arterial hypertension patients: The PILGRIM explorative cohort. *PLoS One*. 2020;15:e0238698.
- 27. Momose Y, Aimi Y, Hirayama T, et al. De novo mutations in the BMPR2 gene in patients with heritable pulmonary arterial hypertension. *Ann Hum Genet*. 2015;79:85-91.
- 28. Aldred MA, Vijayakrishnan J, James V, et al. BMPR2 gene rearrangements account for a significant proportion of mutations in familial and idiopathic pulmonary arterial hypertension. *Hum Mutat.* 2006;27:212-213.
- Zhu N, Gonzaga-Jauregui C, Welch CL, et al. Exome Sequencing in Children With Pulmonary Arterial Hypertension Demonstrates Differences Compared With Adults. *Circ Genom Precis Med.* 2018;11:e001887.
- Galiè N, Humbert M, Vachiery JL, et al. 2015 ESC/ERS Guidelines for the
 Diagnosis and Treatment of Pulmonary Hypertension. *Rev Esp Cardiol (Engl Ed)*.
 2016;69:177.

FIGURE LEGENDS

Figure 1. Schematic representation and main phenotypic features of individuals with chromosomal deletions at 2q33.1 including (48 individuals) or near (2 individuals) *SATB2* based on GRCh37/hg19 assembly (Dashed lines only to help to identify individuals with their corresponding deletion). IDs/references are listed on the left side with "*" depicting novel individuals from this report. Stars next to ID on left side represent those individuals with current weight below 3rd centile. Individuals ID#11, #12, #13, #17, #19, #21, #30, #32, #37, #40, #44, #45, #50, #72, #74, and #77 were previously published.⁶ Only some of the OMIM genes with associated autosomal dominant phenotypes are represented. Boxes highlight overlapping deletions with deleted *COL3A1*, *COL5A2*, *SATB2* or *BMPR2*.

Figure 2. Anthropometric changes over time for individuals with deletions involving *SATB2*. Birth and last available measurement pairs for weight (**A**., 34 individuals) and height (**B**., 21 individuals) were analyzed using paired t-test. Mean Z scores and standard deviations for each group are provided.

Figure 3.Facial features in individuals with deletions encompassing *SATB2* were variable. **A.** Individual SATB2-085. **B.** Individual SATB2-093. **C.** Individual SATB2-099. **D.** Individual SATB2-118. **E.** Individual SATB2-141. **F.** Individual SATB2-147. **G.** Individual SATB2-156. **H.** Individual SATB2-176. **I.** Individual SATB2-180. Note thin vermilion of the upper lip with a flat and/or long philtrum in individuals A, B, C, E, and G; deeply-set eyes in individuals A and C; prominent forehead with a receding anterior hairline in individuals C, D, F, G and I; pointed chin in individuals E, F, G;

narrow jaw in individuals B and F; abnormal appearance of upper central incisors in individuals D and F; and subtle myopathic face in individual H.

J., K., L. Composite facial features as generated by facial recognition technology as provided by Face2Gene vs.19.1.2 for individuals with Δ SAS (24 individuals), Control cohort (24 individuals), and non- Δ SAS (59 individuals).