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Kietsiriroje, N orcid.org/0000-0002-5076-4450, Ariëns, RAS and Ajjan, RA orcid.org/0000-0002-1636-3725 (2021) Fibrinolysis in Acute and Chronic Cardiovascular Disease. Seminars in Thrombosis and Hemostasis. ISSN 0094-6176

https://doi.org/10.1055/s-0040-1718923

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1 Fibrinolysis in acute and chronic cardiovascular disease

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- 17 **ORCID** iD: 0000-0002-1636-3725
- 18 Word count: 5349 words
- 19 Number of Tables: 2
- 20 Number of Figures: 2
- 21 Keywords: fibrinolysis, fibrin, lysis time, cardiovascular disease.
- 22
- **Running title:** Fibrinolysis in acute and chronic CVD
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- 27

30 Abstract

The formation of an obstructive thrombus within an artery remains a major cause of mortality 31 and morbidity worldwide. Despite effective inhibition of platelet function by modern 32 antiplatelet therapies, these agents fail to fully eliminate atherothrombotic risk. This may well 33 34 be related to extensive vascular disease, beyond the protective abilities of the treatment agents used. However, recent evidence suggests that residual vascular risk in those treated with 35 modern antiplatelet therapies is related, at least in part, to impaired fibrin clot lysis. In this 36 review, we attempt to shed more light on the role of hypofibrinolysis in predisposition to 37 arterial vascular events. We provide a brief overview of the coagulation system followed by 38 39 addressing the role of impaired fibrin clot lysis in acute and chronic vascular conditions, including coronary artery, cerebrovascular and peripheral vascular disease. We also discuss the 40 role of combined anticoagulant and antiplatelet therapies to reduce the risk of arterial 41 42 thrombotic events, addressing both efficacy and safety of such an approach. We conclude that impaired fibrin clot lysis appears to contribute to residual thrombosis risk in individuals with 43 arterial disease on antiplatelet therapy, and targeting proteins in the fibrinolytic system 44 represents a viable strategy to improve outcome in this population. Future work is required to 45 refine the antithrombotic approach by modulating pathological abnormalities in the fibrinolytic 46 47 system and tailoring therapy according to the need of each individual.

48

50 List of abbreviations

| A2AP | alpha-2 antiplasmin |
|-------|---|
| ACS | acute coronary syndrome |
| С | complement |
| CVD | cardiovascular disease |
| DAPT | dual-antiplatelet therapy |
| GTT | global thrombosis test |
| Lp(a) | lipoprotein (a) |
| MACE | major adverse cardiovascular events |
| MI | myocardial infarction |
| PAD | peripheral artery disease |
| PAI-1 | plasmin-activator inhibitor 1 |
| PAP | plasmin-antiplasmin complex |
| PAR | protease-activated receptor |
| STEMI | ST-elevated myocardial infarction |
| TAFI | thrombin activatable fibrinolysis inhibitor |
| TEG | thromboelastography |
| TF | tissue factor |
| t-PA | tissue-type plasminogen activator |
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61 **1 Introduction**

Despite significant advances in management, cardiovascular disease (CVD) remains a 62 major cause of mortality worldwide.¹ Myocardial infarction (MI) and cerebrovascular 63 thrombotic events usually follow atherosclerotic plaque rupture, or erosion, which activates the 64 cellular and protein arms of coagulation culminating in thrombus formation. Initially, exposure 65 of subendothelial collagen rapidly mediates platelet adhesion and aggregation through platelet 66 alpha2beta1 and glycoprotein VI interactions with collagen, and glycoprotein Ib interactions 67 68 with von Willebrand factor bound to collagen. Subsequently, activation of coagulation driven by tissue factor (TF) expressed in the exposed core of the plaque generates thrombin for the 69 formation of the fibrin network, that is necessary to provide a scaffold for the blood clot.² 70

71 The molecular processes involved in coagulation activation and fibrinolysis have been reviewed in detail previously³⁻⁵ and only a brief description is provided here (Error! Reference 72 source not found.). Following plaque rupture, TF is exposed, binds factor (F) VII, promoting 73 proteolysis and activation to FVIIa (TF or extrinsic pathway). TF/FVIIa complex subsequently 74 cleaves traces of FIX (contact or intrinsic pathway) and FX (common pathway) into FIXa and 75 76 FXa. The latter is activated by both TF/VIIa and FIXa, and (later together with FVa) converts prothrombin (FII) into thrombin. The slowly accumulated thrombin during the initiation phase 77 further converts several factors including FXI, FVIII and FV, amplifying the activation of 78 79 coagulation cascade, resulting in increased thrombin generation (positive feedback). Sufficient 80 amounts of thrombin are then generated, which is able to convert fibrinogen to form fibrin fibres.³ Finally, thrombin-activated plasma transglutaminase FXIIIa catalyses the formation of 81 82 covalent crosslinks between adjacent D-regions of fibrin monomers, increasing the stability of fibrin clot by tightening its structure and increasing resistance to fibrinolysis.⁶ Additionally, 83 thrombin activates platelets through its protease-activated receptor (PAR) 1 and 4, contributing 84 the formation of the platelet plug.⁷ 85

The formation of fibrin also directly triggers the activation of fibrinolysis by which 86 insoluble fibrin is degraded into small fragments (fibrin degradation products). This interaction 87 is pivotal to ensuring that clotting does not extend within the vessel thus maintaining patency. 88 Upon conversion of fibrinogen to fibrin, conformational changes take place exposing binding 89 sites of plasminogen and tissue-type plasminogen activator (t-PA), while these sites are 90 normally cryptic on fibrinogen.⁸ Plasmin, that is generated through activation of fibrinolytic 91 pathway, degrades fibrin by cleaving the αC-regions first, followed by the coiled-coil region 92 that connects the E- and D-regions in fibrin (Error! Reference source not found.).⁵ 93

94 A balance between clot formation and endogenous fibrinolysis, therefore, is mandatory to restore vascular patency and prevent serious vascular occlusion. Fibrinolysis also plays a key 95 role in wound healing by degrading the blood clot when it is no longer needed, and by 96 97 promoting angiogenesis as appropriate. The impairment of endogenous fibrinolysis, on the other hand, increases the risk of pathological vascular occlusion leading to cardiovascular 98 events. In this review, we delineate the pathological alterations in the fibrinolytic system in 99 CVD and discuss possible therapeutic targets that can be developed to reduce the risk of 100 atherothrombotic events. 101

102 **2 Regulation and alteration of fibrinolysis**

103 The initiation of fibrinolysis commences when fibrinogen is cleaved to form fibrin fibres. 104 The conformational changes that ensue after fibrin formation initially expose t-PA and 105 plasminogen binding sites in the α C-region near the D-region (**Error! Reference source not** 106 **found.**),⁹ consequently leading to the formation of ternary complex between plasminogen, t-107 PA and fibrin, culminating in plasmin activation. When fibrin is degraded, additional binding 108 sites (such as C-terminal lysine residues on α C-region) for plasminogen and t-PA become available, thus accelerating the fibrinolytic process (Figure 2).⁸ Endogenous regulators,
therefore, are necessary to limit this seemingly ceaseless process. However, pathological
alteration of these regulators might lead to impaired fibrinolysis, which increases the risk for
CVD.

Several case-control studies have shown an association between impaired fibrin clot lysis 113 and vascular disease, but it was not until recently that longitudinal studies have been conducted, 114 115 which demonstrated that reduced fibrin clot lysis is an independent predictor of adverse vascular outcome. Sumaya et al have shown, in a large acute coronary syndrome cohort of 116 4354 individuals, that fibrin clot lysis predicts cardiovascular mortality within the first year of 117 the event.¹⁰ This was quickly followed by another publication showing that fibrinolysis can 118 determine major adverse cardiac outcome, 496 patients with ST-elevated myocardial infarction 119 (STEMI).¹¹ Others have shown that fibrin clot lysis does not only predict outcome following 120 arterial occlusion but can also be used to predict outcome following venous occlusive disease.¹² 121 Therefore, fibrin clot lysis appears to be an important modulator of clinical outcome and 122 represents a credible therapeutic target to reduce the risk of vascular occlusion and improve 123 clinical outcome. 124

In this section, we will review the regulation of fibrinolysis and the impact of eachparameter discussed on fibrin degradation.

127 2.1 Clot structure

The thickness of fibrin fibres determines density and pore size of fibrin networks. Thinner fibres are usually associated with denser fibrin clots having smaller pores and this kind of fibrin structure impedes the diffusion of plasminogen and t-PA, thus resulting in delayed fibrinolysis.¹³⁻¹⁵ Two studies exploring clot architecture in coronary artery disease patients

demonstrated that ex vivo clots from these individuals are less permeable with slower lysis time 132 than those from controls.^{16,17} 133

Thrombin generation influences the architecture of clots. Thrombin cleaves short 134 peptides from the N-termini of the A α - and B β -chains, termed FpA and FpB respectively 135 (Error! Reference source not found.), allowing assembly of fibrin through interactions of 136 newly exposed binding sites ("knobs") in the E-region with constitutively expressed binding 137 pockets in the D-region ("holes"). This interaction first generates double stranded protofibrils, 138 which subsequently aggregate laterally to form fibrin fibres that branch into a three-139 dimensional elastic fibrin network (Error! Reference source not found.). An increase or a 140 decrease in thrombin concentration lead to smaller or larger pore sizes, facilitating lower or 141 higher rate of fibrinolysis, respectively.^{18,19} 142

143 Another determinant of fibrin clot structure is plasma levels of fibrinogen with higher levels resulting in more compact clots.²⁰ In addition to quantitative changes in fibrinogen, 144 qualitative alterations in this molecule, such as glycation, phosphorylation, oxidation, and other 145 post-translational modifications lead to changes in clot structure,²¹ which in turn affect 146 resistance to fibrinolysis. 147

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Transglutaminase factor XIII (FXIII) crosslinking and alpha-2 antiplasmin 2.2

149 FXIII is a transglutaminase enzyme that forms a covalent bond between γ - γ , γ - α , and α - α of adjacent fibrin molecules,²² thus stabilizing the fibrin network. Alpha-2 antiplasmin 150 (A2AP), largely produced by the liver, is a strong plasmin inhibitor and is crosslinked by FXIII 151 into the fibrin network, which increases resistance to fibrinolysis by inhibiting plasmin and 152 forming plasmin-antiplasmin complex (PAP) (Error! Reference source not found.).²³ 153

Yet, the association between A2AP level and CVD is still controversial. One study 154 measured plasma levels of fibrinolytic proteins in 555 male survivors of first MI and 635 155 controls, and found that levels of A2AP were independently associated with MI risk.²⁴ Two 156 large cohort studies enrolled 5201 and 6391 patients without baseline CVD also showed that 157 elevated PAP levels were associated with increased risk of MI or death,^{25,26} whereas the study 158 in patients with coronary disease failed to demonstrate an association between PAP levels and 159 cardiovascular death.²⁷ To further add to the confusion, in patients surviving MI, PAP levels 160 were inversely correlated with repeat coronary events.²⁸ However, we need to be cautious in 161 162 our interpretations as plasma levels of PAP may not reflect vascular risk which may instead be related to the amount of A2AP incorporated into fibrin networks. Indeed, an earlier study in 163 type 2 diabetes patients, a high vascular risk group, has shown increased incorporation of A2AP 164 into diabetic clots,²⁹ which correlated with diabetes control measured as HbA1c levels. A 165 subsequent study in type 1 diabetes individuals has again shown increased incorporation of 166 A2AP into diabetes clots,³⁰ and therefore future studies are required to understand the 167 relationship between the amount of plasmin inhibitor present in clots and predisposition to 168 vascular events. 169

170 2.3 Thrombin activatable fibrinolysis inhibitor

Thrombin activatable fibrinolysis inhibitor (TAFI), a zymogen in plasma produced by the liver, can be activated by interaction between thrombin and thrombomodulin complex, located on the endothelial cell surface. Activated TAFI (TAFIa) exerts its fibrinolytic inhibitory effect by cleaving off C-terminal lysine residues from the fibrin surface, thereby limiting the rate of t-PA mediated plasminogen activation (**Error! Reference source not found.**).³¹

The association between TAFI levels and increased risk of CVD is still inconclusive.Case-control studies suggested that high TAFI antigen levels are associated with coronary or

peripheral artery atherosclerosis,^{32,33} whereas other studies proposed that lower levels of TAFI,
due to genetic polymorphisms, are associated with increased risk of MI.³⁴⁻³⁷ In addition, the
AtheroGene study, enrolling patients with coronary disease, demonstrated that a ratio between
activated/inactive TAFI, but not its total level, is associated with cardiovascular death.³⁸ Taken
together, the activation of TAFI rather than its total level may contribute to increased risk of
CVD, but further studies are required to confirm this.

184 **2.4 Plasminogen-activator inhibitor 1**

Plasminogen activator inhibitor-1 (PAI-1) mitigates fibrinolysis activity, under normal
physiological conditions, by inhibiting t-PA in a 1:1 stoichiometric ratio, preventing plasmin
formation on the fibrin surface (Error! Reference source not found.).

Largely produced and secreted by activated platelets, levels of PAI-1 in thrombi are up to 30 times higher than that in plasma,³⁹ suggesting a key role for PAI-1 in stabilising the clot and making it resistant to lysis. PAI-1 is also synthesized by various cells (including endothelial cells, smooth muscle cells, and adipocytes) and upregulated in different conditions deemed high-risk for CVD such as insulin resistance, obesity and diabetes.^{40,41}

Plasma levels of PAI-1 correlate with clot lysis time in healthy volunteers as well as high vascular risk patients.^{42,43} The role of PAI-1 in the thrombotic process is further highlighted in animal studies showing that PAI-1 inhibition increases recombinant t-PA mediated thrombolysis response and, thus, decreases thrombus extension in experimental thrombosis models.⁴⁴

The prognostic value of PAI-1 as a biomarker for cardiovascular outcomes has been inconsistent, ⁴⁵ related to a number of possible factors including: i) PAI-1 levels are confounded by multiple risk factors (e.g. insulin resistance, obesity, exercise, diet and smoking); ii) circadian variation of PAI-1 levels that is not always taken into account, iii) different PAI-1
 assays and sample handling, and iv) plasma levels of PAI-1 may not necessarily represent
 protein levels near the obstructive thrombus.⁴¹

Recently, Song and colleagues performed a well-conducted systematic meta-analysis of 14 studies and observed an association between PAI-1 and coronary heart disease. From the Mendelian randomization analyses, the authors concluded that plasma PAI-1 levels predict coronary heart disease risk (OR 1.22 per unit increase of log-transformed PAI-1; 95%CI: 1.01, 1.47).⁴⁶

209 2.5 Complement 3 and 5 as a substrate for plasmin

The complement (C) pathway is a complex innate immune system activated in a cascadelike fashion, in which C3 is a central protein for this activation. Plasminogen can directly bind to C3 and C5, thus inhibiting complement activation.⁴⁷ Likewise, C3 and C5 can be a substrate for plasmin therefore, competitively prevent plasmin from cleaving fibrin (**Error! Reference source not found.**).

Our previous study has demonstrated the impact of C3 on fibrinolysis in patients with 215 type 1 diabetes and matched healthy controls.⁴⁸ We found that plasma levels of C3 correlated 216 with clot lysis time, and C3 was directly responsible for prolonging lysis time with an effect 217 that seemed to be enhanced in diabetes. Moreover, C3 plasma levels have shown a correlation 218 219 with fibrin clot lysis in a large cohort of type 2 diabetes patients (n=837), with an effect similar to the classical antifibrinolytic factor PAI-1.43 Interestingly, an *in vitro* study has recently 220 indicated that the activation of C3 and C5 alters clot structure with thinner fibres and prolonged 221 lysis time, and inhibition of C3 and C5 activation can restore both clot density and prolongation 222 of lysis time.49 223

The prognostic value of C3 for prediction of cardiovascular event, however, has not been widely investigated. In one study in patients with stable coronary artery disease, C3 levels were significantly higher than controls, and shown to be an independent predictor of coronary artery disease.⁵⁰

228 **2.6** Lipoprotein (a)

It has been widely accepted that Lipoprotein (a) [Lp(a)] is an independent risk factors for CVD. Abundant evidence derived from meta-analyses, large observational studies, Mendelian randomization studies, and genome-wide association studies supports a relationship between increased circulating Lp(a) concentrations and atherosclerotic CVD.⁵¹

Although the precise pathological mechanisms for this link are not fully understood, one suggested mechanism is related to the sequence homology between Lp(a) and plasminogen, possibly resulting in competitive inhibition of fibrin(ogen) binding thus inhibiting plasmin activation.⁵² Elevation of Lp(a) alters clot structure thus reducing fibrin clot permeability, which in turn affects fibrinolysis.^{16,53} Additionally, Lp(a) stimulates PAI-1 expression and production by endothelial cells, further compromising the fibrinolytic process.^{54,55}

However, this concept has been recently challenged by demonstrating that clot lysis time was not affected by Lp(a) levels using in vitro studies.⁵⁶ Therefore, the precise mechanisms for the association between Lp(a) and fibrinolysis are yet to be determined.

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244 3 Clinical importance of impaired fibrinolysis in patients with CVD

In the second part of this review, we address the clinical relevance of altered fibrin clot 245 lysis in individuals with acute or chronic vascular disease. To ensure adequate coverage of this 246 area, we searched the literature from PubMed®, MEDLINE®, and EMBASE® databases using 247 search terms as follow: "fibrin clot, lysis time or fibrinolysis" and "acute coronary 248 syndrome/myocardial infarction/coronary disease, or stroke/cerebrovascular disorder, or 249 peripheral artery disease". Titles and abstracts were screened to select relevant literature. Only 250 251 those written in English and full text provided were selected. Only case-control or prospective studies investigating a link between altered clot structure or lysis time and CVD or outcomes 252 253 were reviewed. All studies included in this review are summarised in Table 1 and Table 2.

It is worth noting that the heterogeneity of patients in various studies and the different 254 assays carried out make it difficult to compare results across study populations. The 255 measurements of fibrinolytic activity mentioned in this section can be divided into 256 investigating two main blood components; i) platelet rich whole blood clots using 257 258 thromboelastography (TEG) (low shearing force condition) and global thrombosis test (GTT) (high-shearing force condition); and ii) platelet poor plasma clots employing turbidimetric 259 analysis and euglobulin clot lysis time. Additionally, it should be reminded that TEG is a 260 technique measuring mainly clot strength and is not sensitive to assess clot lysis. The details 261 of each test have been concisely reviewed elsewhere.⁵⁷ 262

263 **3.1 Acute CVD**

264 This can be divided into conditions affecting the coronary, cerebrovascular or265 peripheral arteries, each discussed separately below.

266 3.1.1 Acute coronary syndrome

Initial studies investigating fibrinolysis in individuals with coronary artery disease wereobservational and a study with a small number of acute coronary syndrome (ACS) patients

(n=40) showed denser clot structure, lower clot permeability, faster clot polymerization and prolonged lysis time, compared with 40 healthy controls.⁵⁸ Prospective studies followed that have shown that impaired fibrinolysis is associated with adverse vascular outcome in individuals with acute ACS. (Table1).

A study involving 270 patients has shown that increased clot strength measured by TEG can predict recurrent ischaemic events in ACS patients who underwent primary percutaneous coronary intervention.⁵⁹

Three other prospective studies have investigated the effect of prolonged clot lysis time observed by the automated point-of-care GTT in 300, 496 and 82 patients, (Table 1) presenting with ACS or STEMI. These studies clearly demonstrated that prolonged lysis time strongly predicts major adverse cardiovascular events (MACE) or cardiovascular death.^{11,60,61} Besides, STEMI patients with lysis time <1000s had a better chance for spontaneous reperfusion (Table 1).^{11,60}

Analyses from the PLATO substudy which enrolled over 4354 ACS patients also concluded that each 50% increment in plasma clot lysis time, measured by turbidimetric analysis, is associated with 36% increased risk of 12-month cardiovascular death and this association remained significant after adjustment for inflammatory and prognostic biomarkers. Likewise, each 50% increase in maximum turbidity was associated with cardiovascular death but this association lost significance after adjustment for clinical and biochemical vascular markers (Table 1).¹⁰

Hence, results from these studies have delivered an essential message that impaired fibrinolysis represents a residual risk factor for cardiovascular events in patients with ACS, despite modern dual antiplatelet therapy.

292 3.1.2 Acute ischaemic stroke

A case-control study that enrolled 45 patients admitted for acute ischaemic stroke 293 within 72 hours of onset, showed that plasma clots from these patients were less porous and 294 more compact with longer lysis time than those from healthy controls.⁶² A prospective study 295 of 74 patients with acute ischaemic stroke receiving thrombolytic therapy showed that less 296 porous clots with longer lysis time predicted adverse neurological response at 3-month (Table 297 1).⁶³ Generally, acute ischaemic stroke studies have been scarce with limited number of 298 299 individuals and therefore robust conclusions cannot be made.

300

301

3.1.3 Acute limb ischaemia or critical limb ischaemia

A small case-control study included 43 patients with history of acute limb ischaemia, 302 referred for further invasive treatments, demonstrated that plasma clots from the patients had 303 lower clot permeability and higher thrombin generation whereas there were no differences in 304 maximum clot formation or clot lysis time, compared to 43 healthy controls.⁶⁴ Another case-305 control study enrolled 85 critical limb ischaemia patients who underwent endovascular therapy 306 and had symptomatic restenosis in treated segment over 12-month follow-up. The control 307 group in this particular study consisted of 47 age-, sex- and cardiovascular risk-matched 308 patients with peripheral artery disease (PAD). This work demonstrated that plasma clots from 309 critical limb ischaemia patients had lower clot permeability and prolonged clot lysis time 310 (Table 1).⁶⁵ In addition, the authors concluded that critical limb ischaemia patients with 311 restenosis had a 3.3-fold higher rate of the adverse composite events (re-intervention, major 312 amputation and cardiovascular death) compared to controls. 313

3.2 **Chronic stable CVD** 314

315 In chronic stable CVD, an increase in clot strength and prolonged clot lysis time are associated with increased risk of vascular events (summarized in Table 2). However, the lack 316

of large prospective studies makes it difficult to decipher whether altered clot structure
contributes to further events or if it is simply associated with other factors responsible for the
enhance risk.

320 3.2.1 Stable coronary artery disease (CAD)

Patients with stable CAD usually have ex vivo plasma clots that are more compact and 321 display increased resistance to lysis compared with healthy controls.^{66,67} More interestingly, 322 among those with stable CAD, a history of previous MI can be associated with increased clot 323 density and prolonged lysis time (Table 2).⁶⁸ The TRIP study also displayed an association 324 between platelet-fibrin clot strength, measured by TEG, and symptoms of patients with CAD. 325 In this study, 171 patients with CAD including 67 with asymptomatic stable CAD, 71 with 326 stable angina and 33 with unstable disease, showed that patients with unstable angina exhibited 327 328 the strongest clots followed by those with stable angina while individuals with asymptomatic disease had the weakest clots.⁶⁹ 329

Results from two prospective studies have affirmed that increased platelet-rich fibrin 330 clot strength measured by TEG predicts recurrent ischaemic events within 2 years in patient 331 with stable CAD (Table 2).^{70,71} However, the effect of prolonged clot lysis time on the 332 increased risk of cardiovascular events seems to be less profound than in ACS. The study by 333 Neergaard-Petersen and colleagues followed 786 patients with stable CAD (90% had previous 334 MI) over 3.1 years, and found that only area under the curve (AUC) of clot formation and lysis 335 predicted composite cardiovascular outcomes, but not maximum turbidity or lysis time alone 336 (Table 2).⁷² However, the study had relatively small sample size with only 70 (9%) events 337 occurring during the 3.1 years follow-up period. Thus, it was likely that the maximum turbidity 338 or lysis time was underpowered to predict outcomes when analysed separately but the 339

340 combined analysis (that essentially includes these two measures) had enough power to show341 an association.

342 It is important to stress that TEG studies that measure clot strength do not necessarily343 reflect fibrinolysis potential, which can be influenced by additional factors.

344

345

5 3.2.2 Previous history of ischaemic stroke

A number of case-control studies have indicated that patients with previous ischaemic 346 stroke produce more compact clots that are resistant to lysis, when compared to healthy controls 347 (Table 2).^{67,73-75} There is only one prospective study observing recurrent ischaemic stroke or 348 transient ischaemic attack events in 218 patients with extra- or intracranial artery stenosis who 349 underwent stenting. Using TEG, 18 patients who developed events showed stronger platelet-350 fibrin clots compared with those who remained free of an event (Table 2).⁷⁶ However, 9 out of 351 18 events occurred within 7 days of stenting, the result therefore should be regarded as a short-352 353 term effect of impaired fibrinolysis on stent re-thrombosis rather than a long-term effect.

354

355 3.2.3 Peripheral artery disease (PAD)

Two small case-control studies have shown that clots produced from plasma of patients 356 357 with peripheral artery disease were more compact and resistant to lysis than matched controls, corresponding to other chronic CVD (Table 2).77,78 Undas and colleagues investigated ex vivo 358 clots from 106 patients with PAD, aged <70 years and found that clots from patients had lower 359 permeability, higher density and prolonged lysis time, compared to matched controls.⁷⁷ 360 Similarly in another study by Okraska-Bylica and colleagues, clots from 31 younger patients 361 with premature CAD, aged \leq 55 years, also showed reduced clot permeability and prolonged 362 lysis time compared with matched controls.⁷⁸ Additionally, our works also demonstrated that 363

ex vivo clots derived from 106 male first-degree relatives and 34 male patients with intermittent
 claudication had higher density and prolonged lysis time, compared to healthy age-matched
 controls (Table 2).^{79,80}

367 4 Potential adjunct therapies to target hypofibrinolysis in patients with CVD

From the evidence presented above, it appears that residual thrombosis risk in individuals 368 with vascular disease may be ameliorated by addressing the hypofibrinolytic environment, in 369 addition to the use of antiplatelet therapies. In this section, we explore the latest in adjunctive 370 antithrombotic therapies, added to existing antiplatelets, to further reduce residual vascular risk 371 372 and improve outcome in individuals with CVD. It should be noted that current chronic therapies for hypofibrinolysis target clot lysis indirectly by making the fibrin network more susceptible 373 to lysis and agents that directly affect one or multiple proteins in the fibrinolytic system are not 374 375 yet available for clinical use.

376

377 4.1 Factor Xa inhibitors

Inhibiting FXa in the coagulation cascade leads to decreased thrombin generation,⁸¹ 378 consequently resulting in reduced clot formation, together with the generation of fibrin 379 networks that are less compact with increased susceptibility to lysis. Moreover, factor Xa 380 inhibitors possibly enhance fibrinolysis by interfering with TAFI activation.⁸² Therefore, the 381 382 use of a FXa inhibitor in addition to standard antiplatelet therapy may offer additional benefits in patients with CVD. However, it remains unclear whether the additional benefits of combined 383 antiplatelet/anticoagulant are related to reduction in fibrin clot formation, enhanced 384 385 fibrinolysis, or a combination of the two.

The EDOC-APT study tested the effects of 30 and 60 mg of edoxaban on clot kinetics in 75 CAD patients taking aspirin and clopidogrel.⁸³ The authors concluded that edoxaban delayed thrombin generation in a dose-dependent manner but did not affect maximum clot firmness measured by TEG. Another ongoing similar study (NCT03775746) on 150 patients with ACS is testing the effects of low-dose rivaroxaban and clopidogrel, in addition to aspirin, on fibrinolytic status measured by GTT.⁸⁴

In the ATLAS ACS 2-TIMI 51 study, a total of 15,526 high-risk ACS patients were 392 randomly assigned to receive low-dose rivaroxaban 2.5 mg twice daily, rivaroxaban 5 mg twice 393 daily, or placebo, on top of standard antiplatelet therapy, and were then followed for a median 394 of 13.1 months.⁸⁵ Compared with placebo, both 2.5 and 5 mg twice daily of rivaroxaban 395 significantly reduced MACE [HR 0.84 (95% CI: 0.72–0.97) and HR 0.85 (95% CI: 0.73–0.98), 396 respectively] at the cost of increased major TIMI bleeding [HR 3.46 (95% CI: 2.08-5.77) and 397 HR 4.47 (95% CI: 2.71–7.36), respectively]. Low-dose rivaroxaban also reduced 398 cardiovascular death whereas rivaroxaban 5 mg twice failed to show an effect. Based on these 399 data, low-dose rivaroxaban in high-risk patients on top of DAPT (aspirin and clopidogrel) or 400 aspirin alone has been approved by the European Medicines Agency. Nevertheless, these 401 results were not reproduced in another similar study using Apixaban 5 mg daily in addition to 402 DAPT or aspirin alone, in patients with high-risk ACS (the APPRAISE-2 study). This trial was 403 404 prematurely terminated because of an increase in major bleeding events with apixaban in the absence of a counterbalancing reduction in recurrent ischemic events, after a median follow-405 up of 241 days.86 406

In chronic stable CVD, the recently published COMPASS study has opened a new opportunity for the use of low-dose rivaroxaban added to aspirin in high-risk patients with stable CVD (older than 65 years, or atherosclerosis involving ≥ 2 vascular beds, or ≥ 2

additional risk factors).⁸⁷ In brief, 27,395 patients with high-risk stable coronary artery or 410 peripheral artery diseases were randomly assigned to low-dose rivaroxaban (2.5g mg twice 411 daily) plus aspirin, rivaroxaban (5 mg twice daily), or aspirin (100 mg daily). During a mean 412 follow-up of 23 months, the composite cardiovascular death, stroke, or MI was significantly 413 lower in low-dose rivaroxaban plus aspirin, compared to aspirin alone [HR 0.76 (95%CI:0.66-414 0.86), p<0.001], with a trade-off from higher major bleeding risk [HR 1.7 (95%CI: 1.4-2.05), 415 p<0.001]. The combination of rivaroxaban and aspirin also offered all-cause mortality 416 reduction by approximately 18% and net clinical benefits by 20%.⁸⁷ Interestingly, in a subgroup 417 418 analysis of PAD patients, combination therapy showed reduction in amputations, demonstrating that the benefits are not only related to reduction of coronary events but also 419 CLI.88 420

Based on the ATLAS ACS 2-TIMI 51 and COMPASS studies, dual pathway inhibition by low-dose rivaroxaban (inhibiting thrombin generation) and antiplatelet therapy synergistically reduces residual cardiovascular risk in patients with high-risk ACS or high-risk stable coronary artery or peripheral artery diseases,⁸⁹ however, the role of this dual combination is not applicable for ischaemic stroke patients, which was one of the exclusion criteria in both studies.

427 **4.2 Direct thrombin inhibitor (dabigatran)**

Dabigatran directly binds to active sites of thrombin, inhibiting its action and also promotes fibrinolysis through reduction in TAFI activation.⁹⁰ In one study measuring *ex vivo* plasma clots from patients with atrial fibrillation by turbidimetric analysis, dabigatran and factor Xa inhibitors delayed clot formation and modestly decreased clot firmness.⁹¹ In contrast, another study scrutinising thrombin generation and clot firmness in whole-blood and plasma by using TEG from 8 heathy volunteers, demonstrated that while both FXa inhibitors and

dabigatran effectively prolonged thrombin generation, neither altered clot firmness.⁹² These 434 findings resembled the results from another a double-blinded, placebo, randomized controlled 435 trial (RCT) that included 35 CAD patients on DAPT with aspirin and clopidogrel. Again, 436 dabigatran significantly decreased thrombin activity and delayed fibrin clot formation, without 437 affecting clot structure or fibrinolysis.⁹³ Importantly, the inconsistency of the dabigatran effect 438 in studies is perhaps related to variation in the methodologies used, particularly when different 439 440 triggers are used to stimulate clot formation (i.e tissue factor vs thrombin) which, in turn, invalidates attempts at comparing results from different studies. 441

Unfortunately, the use of dabigatran with antiplatelet therapy in patients with stable CAD 442 or ACS has been a disappointment. The RE-LY study investigated the use of dabigatran to 443 prevent ischaemic events in patients with atrial fibrillation and unexpectedly showed a possible 444 increased risk of MI, despite the reduction in composite stroke and embolic events.⁹⁴ Results 445 from the phase II RE-DEEM study, dabigatran was associated with a dose-dependent increase 446 in bleeding events without reduction in MACE.⁹⁵ Finally, a meta-analysis included over 30,000 447 patients from 7 trials demonstrated that dabigatran was associated with an increased risk of MI 448 and ACS indicating that this drug is not a viable option for those with coronary artery disease.⁹⁶ 449

450 **4.3** Potential future therapies directly targeting clot lysis

451 As mentioned above, endogenous antifibrinolytic proteins, in pathological alterations, are 452 associated with increased risk of CVD. Modulating their functional activity may, therefore, 453 enable novel therapeutic options to alleviate residual thrombosis risk in patients with CVD.

Inhibition of TAFI was considered as a putative target to enhance fibrinolysis. The discovery of thrombin-thrombomodulin interaction activating TAFIa had drawn attention from pharmaceutical companies to develop compounds inhibiting TAFIa;⁹⁷ however, a limited number of drug candidates (AZD9684 from AstraZeneca and UK-396082 from Pfizer) 458 eventually entered clinical studies which were later discontinued for unknown reasons.⁹⁸
459 Another potential approach of TAFIa inhibitors relied on antibodies or nanobodies, yet *in vivo*460 data are still lacking.⁹⁹

For decades, researchers have been attempting to develop small molecule inhibitors or 461 antibodies against PAI-1. Some of antibodies have previously shown the inhibitory effect in 462 vivo in animal models (TM5275, MA-33H1F7 and MA-MP2D2),^{100,101} whereas most recent 463 nanobodies have been initially tested in an *in vitro* clot lysis assay.¹⁰² Tiplaxtinin (or PAI 039) 464 is one of the most studied small molecule inhibitors, proven the PAI-1 inhibiting efficacy in 465 several animal models.¹⁰³ Nevertheless, none of these has successfully advanced further into 466 clinical studies.^{104,105} Recently, an innovative strategy using heterodimer bi-specifically against 467 TAFI and PAI-1 has been tested in a thrombosis animal model emerging the concept of multi-468 target inhibitors.¹⁰⁶ 469

A2AP is another possible drug target, with a number of different approaches investigated 470 to inhibit protein function including antibodies, plasmin inhibitor mutants, use of synthetic N 471 and C terminal peptides and inhibition of plasmin inhibitor cleavage. However, none of these 472 approaches so far progressed further to clinical trials.⁹⁹ After 20 years of relative inactivity, 473 treatment targeting A2AP has been reactivated via a recent study by Singh et al, showing that 474 the injection of A2AP inactivating antibody in adjunct to recombinant t-PA successfully 475 476 potentiated clot dissolution without increased bleeding in an *in vivo* model of acute pulmonary emboli.¹⁰⁷ Human studies using this approach have been recently undertaken but the work was 477 stopped prematurely and a full report is awaited with interest (NCT03001544). 478

Even though Eculizumab (Soliris®, Alexion Pharma GmbH, Switzerland), a drug inhibiting the terminal (C3 and C5) complement system, has been approved for the treatment of series of inflammatory and autoimmune diseases,¹⁰⁸ the role of C3 or C5 inhibitor in fibrinolysis has not yet been widely tested. The early work from our institute has shown that
high affinity fibrinogen-binding and C3-specific conformational proteins (Affimers®, Avacta,
Cambridge, UK) are able to abolish C3-induced prolongation of clot lysis *ex vivo* by interfering
C3-fibrinogen interaction.¹⁰⁹ Yet it remains to be seen whether this approach can be advanced
further for clinical trials in the future.

487

488 **5** Conclusion and future directions

489 It is now without doubt that prolonged fibrinolysis represents a risk factor for adverse vascular outcome, even when aggressive antiplatelet therapies are in place. A number of factors 490 are responsible for altered fibrin clot lysis, including quantitative and qualitative changes in 491 different coagulation proteins. These can affect fibrinolysis indirectly by altering structure of 492 the clot, secondary to raised fibrinogen levels or presence of post-translational modification of 493 the protein.^{20,21} Fibrinolysis can also be directly affected due to raised levels of antifibrinolytic 494 proteins such as PAI-1,^{42,43} increased incorporation of A2AP into the clot,²⁹ or increased 495 glycation of plasminogen.¹¹⁰ 496

Importantly, recent evidence indicates that impaired fibrinolysis is a key factor responsible 497 for atherothrombotic events in high risk patients, even when adequately covered with modern 498 antiplatelet therapies. Therefore, the hotly-debated area of "residual thrombosis risk" appears 499 to be related, in large part, to impaired fibrin clot lysis. Consequently, targeting compromised 500 fibrin clot lysis in these individuals represents a legitimate therapeutic approach. Indeed, 501 studies have generally shown that combining antiplatelet with anticoagulant therapies reduces 502 the risk of arterial vascular occlusion but usually at the expense of increased bleeding events, 503 making widespread use of this strategy problematic. However, studies have used combination 504

treatment without an attempt to "individualise" such dual therapy and limit to those at the 505 highest risk. This is due to the fact that tangible strategies to stratify patients at particular risk 506 of impaired fibrinolysis have not made it into clinical practice. Therefore, developing and 507 validating an assay that can be routinely used to determine fibrinolytic efficiency will help to 508 facilitate patient stratification and tailor anti-thrombotic therapy accordingly. It must be 509 acknowledged that current chronic antithrombotic therapies that target the protein phase of 510 511 coagulation do not directly affect the fibrinolytic system but exert their effects indirectly, which may explain the limited clinical benefit, which is largely related to increased bleeding risk. 512 513 Therefore, more work is needed to develop therapies that target the fibrinolytic system with a particular focus on modulating the pathological abnormalities thus maximising anti-thrombotic 514 benefits while minimising bleeding risks. For example, inhibiting A2AP and/or PAI-1 in 515 individuals with diabetes and vascular disease is likely to be more effective that targeting FX 516 activity by offering a more favourable benefit:risk ratio, given the focus on known pathological 517 abnormalities in this metabolic condition. Finally, we will also need a reliable test that can 518 monitor response to a particular antithrombotic agent, akin to anti-hypertensive and anti-519 hyperlipidaemic therapies, which will further refine our approach and allow a more structured 520 strategy to the management of thrombosis risk. 521

Anti-thrombotic therapies have made great progress over the past few decades and this trend is likely to continue. The next steps will require the development of more targeted therapies and expansion of individualised strategies of patients care, in order to ensure the best clinical outcome in population with high vascular risk.

526 6 Conflicts of interest

527 All authors have no conflict of interest to be declared.

528 **7 Funding**

529 This work receives no specific funding. NK is funded by the Faculty of Medicine, Prince of

530 Songkla University, Thailand. RAA research work is supported by grants from the NIHR,

531 Diabetes UK, the British Heart Foundation, Avacta Life Sciences and Abbott Diabetes Care.

532 RASA is supported by the British Heart Foundation (RG/18/11/34036) and the Wellcome Trust

533 (204951/B/16/Z).

534 8 Author contributions

535 NK was responsible for drafting and writing of the manuscript, searching of literature and 536 interpreting of data. RASA and RAA was responsible for the drafting and writing the 537 manuscript and critical revision of important intellectual content. All authors approved the 538 version to be published.

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540 9 References

- Roth GA, Johnson C, Abajobir A, et al. Global, Regional, and National Burden of Cardiovascular Diseases for 10 Causes, 1990 to 2015. *Journal of the American College of Cardiology*. 2017;70:1-25.
 Reininger AJ, Bernlochner I, Penz SM, et al. A 2-step mechanism of arterial thrombus formation induced by human atherosclerotic plaques. *Journal of the American College of Cardiology*. 2010;55:1147-1158.
- Versteeg HH, Heemskerk JW, Levi M, Reitsma PH. New fundamentals in hemostasis.
 Physiological reviews. 2013;93:327-358.
- 549 4. Okafor ON, Gorog DA. Endogenous Fibrinolysis: An Important Mediator of
 550 Thrombus Formation and Cardiovascular Risk. *Journal of the American College of*551 *Cardiology*. 2015;65:1683-1699.
- 552 5. Longstaff C, Kolev K. Basic mechanisms and regulation of fibrinolysis. *Journal of* 553 *thrombosis and haemostasis : JTH*. 2015;13 Suppl 1:S98-105.
- 6. Hethershaw EL, Cilia La Corte AL, Duval C, et al. The effect of blood coagulation
 factor XIII on fibrin clot structure and fibrinolysis. *Journal of thrombosis and haemostasis : JTH*. 2014;12:197-205.

- 557 7. Stegner D, Nieswandt B. Platelet receptor signaling in thrombus formation. *Journal of molecular medicine*. 2011;89:109-121.
- 8. Medved L, Nieuwenhuizen W. Molecular mechanisms of initiation of fibrinolysis by
 fibrin. *Thrombosis and haemostasis*. 2003;89:409-419.
- 9. Yakovlev S, Makogonenko E, Kurochkina N, Nieuwenhuizen W, Ingham K, Medved
 L. Conversion of fibrinogen to fibrin: mechanism of exposure of tPA- and
 plasminogen-binding sites. *Biochemistry*. 2000;39:15730-15741.
- Sumaya W, Wallentin L, James SK, et al. Fibrin clot properties independently predict adverse clinical outcome following acute coronary syndrome: a PLATO substudy. *European heart journal*. 2018;39:1078-1085.
- Farag M, Spinthakis N, Gue YX, et al. Impaired endogenous fibrinolysis in STsegment elevation myocardial infarction patients undergoing primary percutaneous
 coronary intervention is a predictor of recurrent cardiovascular events: the RISK PPCI
 study. *European heart journal*. 2019;40:295-305.
- 571 12. Cieslik J, Mrozinska S, Broniatowska E, Undas A. Altered plasma clot properties
 572 increase the risk of recurrent deep vein thrombosis: a cohort study. *Blood*.
 573 2018;131:797-807.
- 13. Carr ME, Jr., Alving BM. Effect of fibrin structure on plasmin-mediated dissolution
 of plasma clots. *Blood coagulation & fibrinolysis : an international journal in haemostasis and thrombosis.* 1995;6:567-573.
- 577 14. Collet JP, Park D, Lesty C, et al. Influence of fibrin network conformation and fibrin
 578 fiber diameter on fibrinolysis speed: dynamic and structural approaches by confocal
 579 microscopy. *Arteriosclerosis, thrombosis, and vascular biology*. 2000;20:1354-1361.
- 580 15. Gabriel DA, Muga K, Boothroyd EM. The effect of fibrin structure on fibrinolysis.
 581 *The Journal of biological chemistry*. 1992;267:24259-24263.
- Undas A, Plicner D, Stepien E, Drwila R, Sadowski J. Altered fibrin clot structure in patients with advanced coronary artery disease: a role of C-reactive protein, lipoprotein(a) and homocysteine. *Journal of thrombosis and haemostasis : JTH*.
 2007;5:1988-1990.
- 586 17. Collet JP, Allali Y, Lesty C, et al. Altered fibrin architecture is associated with
 hypofibrinolysis and premature coronary atherothrombosis. *Arteriosclerosis, thrombosis, and vascular biology.* 2006;26:2567-2573.
- 18. Wolberg AS, Monroe DM, Roberts HR, Hoffman M. Elevated prothrombin results in clots with an altered fiber structure: a possible mechanism of the increased thrombotic risk. *Blood*. 2003;101:3008-3013.
- He S, Blomback M, Bark N, Johnsson H, Wallen NH. The direct thrombin inhibitors
 (argatroban, bivalirudin and lepirudin) and the indirect Xa-inhibitor (danaparoid)
 increase fibrin network porosity and thus facilitate fibrinolysis. *Thrombosis and haemostasis*. 2010;103:1076-1084.
- Ariens RA. Fibrin(ogen) and thrombotic disease. *Journal of thrombosis and haemostasis : JTH*. 2013;11 Suppl 1:294-305.
- de Vries JJ, Snoek CJM, Rijken DC, de Maat MPM. Effects of Post-Translational
 Modifications of Fibrinogen on Clot Formation, Clot Structure, and Fibrinolysis: A
 Systematic Review. *Arteriosclerosis, thrombosis, and vascular biology*. 2020;40:554569.
- Ariens RA, Lai TS, Weisel JW, Greenberg CS, Grant PJ. Role of factor XIII in fibrin
 clot formation and effects of genetic polymorphisms. *Blood.* 2002;100:743-754.
- Aoki N. Discovery of alpha2-plasmin inhibitor and its congenital deficiency. *Journal of thrombosis and haemostasis : JTH*. 2005;3:623-631.

24. Meltzer ME, Doggen CJ, de Groot PG, Rosendaal FR, Lisman T. Plasma levels of 606 fibrinolytic proteins and the risk of myocardial infarction in men. Blood. 607 2010:116:529-536. 608 25. Cushman M, Lemaitre RN, Kuller LH, et al. Fibrinolytic activation markers predict 609 myocardial infarction in the elderly. The Cardiovascular Health Study. 610 Arteriosclerosis, thrombosis, and vascular biology. 1999;19:493-498. 611 26. Folsom AR, Delaney JA, Lutsey PL, et al. Associations of factor VIIIc, D-dimer, and 612 plasmin-antiplasmin with incident cardiovascular disease and all-cause mortality. 613 American journal of hematology. 2009;84:349-353. 614 27. Morange PE, Bickel C, Nicaud V, et al. Haemostatic factors and the risk of 615 cardiovascular death in patients with coronary artery disease: the AtheroGene study. 616 Arteriosclerosis, thrombosis, and vascular biology. 2006;26:2793-2799. 617 618 28. Redondo M, Carroll VA, Mauron T, et al. Hemostatic and fibrinolytic parameters in 619 survivors of myocardial infarction: a low plasma level of plasmin-alpha2-antiplasmin complex is an independent predictor of coronary re-events. Blood coagulation & 620 fibrinolysis : an international journal in haemostasis and thrombosis. 2001;12:17-24. 621 622 29. Dunn EJ, Philippou H, Ariens RA, Grant PJ. Molecular mechanisms involved in the resistance of fibrin to clot lysis by plasmin in subjects with type 2 diabetes mellitus. 623 Diabetologia. 2006;49:1071-1080. 624 30. Agren A, Jorneskog G, Elgue G, Henriksson P, Wallen H, Wiman B. Increased 625 incorporation of antiplasmin into the fibrin network in patients with type 1 diabetes. 626 Diabetes care. 2014:37:2007-2014. 627 31. Foley JH, Kim PY, Mutch NJ, Gils A. Insights into thrombin activatable fibrinolysis 628 inhibitor function and regulation. Journal of thrombosis and haemostasis : JTH. 629 2013;11 Suppl 1:306-315. 630 32. Schroeder V, Chatterjee T, Mehta H, et al. Thrombin activatable fibrinolysis inhibitor 631 (TAFI) levels in patients with coronary artery disease investigated by angiography. 632 Thrombosis and haemostasis. 2002;88:1020-1025. 633 33. de Bruijne EL, Gils A, Rijken DC, et al. High thrombin activatable fibrinolysis 634 inhibitor levels are associated with an increased risk of premature peripheral arterial 635 disease. Thrombosis research. 2011;127:254-258. 636 de Bruijne EL, Gils A, Guimaraes AH, et al. The role of thrombin activatable 637 34. fibrinolysis inhibitor in arterial thrombosis at a young age: the ATTAC study. Journal 638 of thrombosis and haemostasis : JTH. 2009;7:919-927. 639 Meltzer ME, Doggen CJ, de Groot PG, Meijers JC, Rosendaal FR, Lisman T. Low 35. 640 641 thrombin activatable fibrinolysis inhibitor activity levels are associated with an increased risk of a first myocardial infarction in men. Haematologica. 2009;94:811-642 818. 643 36. Juhan-Vague I, Morange PE, Aubert H, et al. Plasma thrombin-activatable fibrinolysis 644 inhibitor antigen concentration and genotype in relation to myocardial infarction in 645 the north and south of Europe. Arteriosclerosis, thrombosis, and vascular biology. 646 2002;22:867-873. 647 Juhan-Vague I, Morange PE, Group PS. Very high TAFI antigen levels are associated 648 37. with a lower risk of hard coronary events: the PRIME Study. Journal of thrombosis 649 and haemostasis : JTH. 2003;1:2243-2244. 650 38. Tregouet DA, Schnabel R, Alessi MC, et al. Activated thrombin activatable 651 fibrinolysis inhibitor levels are associated with the risk of cardiovascular death in 652 patients with coronary artery disease: the AtheroGene study. Journal of thrombosis 653 and haemostasis : JTH. 2009;7:49-57. 654

| 655 | 39. | Robbie LA, Bennett B, Croll AM, Brown PA, Booth NA. Proteins of the fibrinolytic |
|------------|-----|--|
| 656 | | system in human thrombi. Thrombosis and haemostasis. 1996;75:127-133. |
| 657 | 40. | Kohler HP, Grant PJ. Plasminogen-activator inhibitor type 1 and coronary artery |
| 658 | | disease. The New England journal of medicine. 2000;342:1792-1801. |
| 659 660 | 41. | Jung RG, Simard T, Labinaz A, et al. Role of plasminogen activator inhibitor-1 in coronary pathophysiology. <i>Thrombosis research</i> . 2018;164:54-62. |
| 661 | 42. | Urano T, Sakakibara K, Rydzewski A, Urano S, Takada Y, Takada A. Relationships |
| 662 | 12. | between euglobulin clot lysis time and the plasma levels of tissue plasminogen |
| 663 | | activator and plasminogen activator inhibitor 1. <i>Thrombosis and haemostasis</i> . |
| 664 | | 1990;63:82-86. |
| 665 | 43. | Hess K, Alzahrani SH, Price JF, et al. Hypofibrinolysis in type 2 diabetes: the role of |
| 666 | 13. | the inflammatory pathway and complement C3. <i>Diabetologia</i> . 2014;57:1737-1741. |
| 667 | 44. | Levi M, Biemond BJ, van Zonneveld AJ, ten Cate JW, Pannekoek H. Inhibition of |
| 668 | | plasminogen activator inhibitor-1 activity results in promotion of endogenous |
| 669 | | thrombolysis and inhibition of thrombus extension in models of experimental |
| 670 | | thrombosis. <i>Circulation</i> . 1992;85:305-312. |
| 671 | 45. | Gorog DA. Prognostic value of plasma fibrinolysis activation markers in |
| 672 | 10. | cardiovascular disease. Journal of the American College of Cardiology. |
| 673 | | 2010;55:2701-2709. |
| 674 | 46. | Song C, Burgess S, Eicher JD, O'Donnell CJ, Johnson AD. Causal Effect of |
| 675 | 10. | Plasminogen Activator Inhibitor Type 1 on Coronary Heart Disease. <i>Journal of the</i> |
| 676 | | American Heart Association. 2017;6. |
| 677 | 47. | Barthel D, Schindler S, Zipfel PF. Plasminogen is a complement inhibitor. <i>The</i> |
| 678 | .,. | Journal of biological chemistry. 2012;287:18831-18842. |
| 679 | 48. | Hess K, Alzahrani SH, Mathai M, et al. A novel mechanism for hypofibrinolysis in |
| 680 | 10. | diabetes: the role of complement C3. <i>Diabetologia</i> . 2012;55:1103-1113. |
| 681 | 49. | Schutt KA, Maxeiner S, Lysaja K, et al. Complement activation leads to C3 and C5 |
| 682 | | dependent prothrombotic alterations of fibrin clots. <i>European heart journal</i> . 2019;40. |
| 683 | 50. | Ajjan R, Grant PJ, Futers TS, et al. Complement C3 and C-reactive protein levels in |
| 684 | | patients with stable coronary artery disease. <i>Thrombosis and haemostasis</i> . |
| 685 | | 2005;94:1048-1053. |
| 686 | 51. | Wilson DP, Jacobson TA, Jones PH, et al. Use of Lipoprotein(a) in clinical practice: |
| 687 | | A biomarker whose time has come. A scientific statement from the National Lipid |
| 688 | | Association. Journal of clinical lipidology. 2019;13:374-392. |
| 689 | 52. | Boffa MB, Koschinsky ML. Lipoprotein (a): truly a direct prothrombotic factor in |
| 690 | | cardiovascular disease? Journal of lipid research. 2016;57:745-757. |
| 691 | 53. | Undas A, Stepien E, Tracz W, Szczeklik A. Lipoprotein(a) as a modifier of fibrin clot |
| 692 | | permeability and susceptibility to lysis. Journal of thrombosis and haemostasis : JTH. |
| 693 | | 2006;4:973-975. |
| 694 | 54. | Etingin OR, Hajjar DP, Hajjar KA, Harpel PC, Nachman RL. Lipoprotein (a) |
| 695 | | regulates plasminogen activator inhibitor-1 expression in endothelial cells. A potential |
| 696 | | mechanism in thrombogenesis. The Journal of biological chemistry. 1991;266:2459- |
| 697 | | 2465. |
| 698 | 55. | Ren S, Man RY, Angel A, Shen GX. Oxidative modification enhances lipoprotein(a)- |
| 699 | | induced overproduction of plasminogen activator inhibitor-1 in cultured vascular |
| 700 | | endothelial cells. Atherosclerosis. 1997;128:1-10. |
| 701 | 56. | Boffa MB, Marar TT, Yeang C, et al. Potent reduction of plasma lipoprotein (a) with |
| 702 | | an antisense oligonucleotide in human subjects does not affect ex vivo fibrinolysis. |
| 703 | | Journal of lipid research. 2019;60:2082-2089. |

57. Ilich A, Bokarev I, Key NS. Global assays of fibrinolysis. International journal of 704 laboratory hematology. 2017;39:441-447. 705 Undas A, Szuldrzynski K, Stepien E, et al. Reduced clot permeability and 706 58. susceptibility to lysis in patients with acute coronary syndrome: effects of 707 inflammation and oxidative stress. Atherosclerosis. 2008;196:551-557. 708 59. Kreutz RP, Schmeisser G, Maatman B, et al. Fibrin clot strength measured by 709 thrombelastography and outcomes after percutaneous coronary intervention. 710 Thrombosis and haemostasis. 2017;117:426-428. 711 60. Christopoulos C, Farag M, Sullivan K, Wellsted D, Gorog DA. Impaired thrombolytic 712 status predicts adverse cardiac events in patients undergoing primary percutaneous 713 coronary intervention. Thrombosis and haemostasis. 2017;117:457-470. 714 Saraf S, Christopoulos C, Salha IB, Stott DJ, Gorog DA. Impaired endogenous 715 61. 716 thrombolysis in acute coronary syndrome patients predicts cardiovascular death and 717 nonfatal myocardial infarction. Journal of the American College of Cardiology. 2010;55:2107-2115. 718 Undas A, Slowik A, Wolkow P, Szczudlik A, Tracz W. Fibrin clot properties in acute 62. 719 720 ischemic stroke: relation to neurological deficit. Thrombosis research. 2010;125:357-361. 721 63. Bembenek JP, Niewada M, Siudut J, Plens K, Czlonkowska A, Undas A. Fibrin clot 722 723 characteristics in acute ischaemic stroke patients treated with thrombolysis: the impact on clinical outcome. Thrombosis and haemostasis. 2017;117:1440-1447. 724 725 64. Karpinska IA, Nowakowski T, Wypasek E, Plens K, Undas A. A prothrombotic state and denser clot formation in patients following acute limb ischemia of unknown 726 cause. Thrombosis research. 2020;187:32-38. 727 65. Nowakowski T, Malinowski KP, Nizankowski R, Iwaniec T, Undas A. Restenosis is 728 729 associated with prothrombotic plasma fibrin clot characteristics in endovascularly treated patients with critical limb ischemia. Journal of thrombosis and thrombolysis. 730 2019;47:540-549. 731 66. Reddel CJ, Curnow JL, Voitl J, et al. Detection of hypofibrinolysis in stable coronary 732 artery disease using the overall haemostatic potential assay. Thrombosis research. 733 2013;131:457-462. 734 735 67. Siegerink B, Meltzer ME, de Groot PG, Algra A, Lisman T, Rosendaal FR. Clot lysis time and the risk of myocardial infarction and ischaemic stroke in young women; 736 results from the RATIO case-control study. British journal of haematology. 737 2012;156:252-258. 738 739 68. Neergaard-Petersen S, Ajjan R, Hvas AM, et al. Fibrin clot structure and platelet aggregation in patients with aspirin treatment failure. PloS one. 2013;8:e71150. 740 Tantry US, Bliden KP, Suarez TA, Kreutz RP, Dichiara J, Gurbel PA. 69. 741 742 Hypercoagulability, platelet function, inflammation and coronary artery disease acuity: results of the Thrombotic RIsk Progression (TRIP) study. Platelets. 743 2010;21:360-367. 744 70. Gurbel PA, Bliden KP, Kreutz RP, Dichiara J, Antonino MJ, Tantry US. The link 745 between heightened thrombogenicity and inflammation: pre-procedure 746 characterization of the patient at high risk for recurrent events after stenting. Platelets. 747 748 2009:20:97-104. 71. Hou X, Han W, Gan Q, Liu Y, Fang W. Relationship between thromboelastography 749 750 and long-term ischemic events as gauged by the response to clopidogrel in patients 751 undergoing elective percutaneous coronary intervention. Bioscience trends. 2017;11:209-213. 752

| 753 | 72. | Neergaard-Petersen S, Larsen SB, Grove EL, Kristensen SD, Ajjan RA, Hvas AM. |
|-----|-----|---|
| 754 | | Imbalance between Fibrin Clot Formation and Fibrinolysis Predicts Cardiovascular |
| 755 | | Events in Patients with Stable Coronary Artery Disease. Thrombosis and haemostasis. |
| 756 | | 2020;120:75-82. |
| 757 | 73. | Anzej S, Bozic M, Antovic A, et al. Evidence of hypercoagulability and inflammation |
| 758 | | in young patients long after acute cerebral ischaemia. Thrombosis research. |
| 759 | | 2007;120:39-46. |
| 760 | 74. | Undas A, Podolec P, Zawilska K, et al. Altered fibrin clot structure/function in |
| 761 | | patients with cryptogenic ischemic stroke. Stroke. 2009;40:1499-1501. |
| 762 | 75. | Vuckovic BA, Djeric MJ, Ilic TA, et al. Fibrinolytic parameters, lipid status and |
| 763 | | lipoprotein(a) in ischemic stroke patients. Srpski arhiv za celokupno lekarstvo. |
| 764 | | 2010;138 Suppl 1:12-17. |
| 765 | 76. | Wang B, Li XQ, Ma N, et al. Association of thrombelastographic parameters with |
| 766 | | post-stenting ischemic events. Journal of neurointerventional surgery. 2017;9:192- |
| 767 | | 195. |
| 768 | 77. | Undas A, Nowakowski T, Ciesla-Dul M, Sadowski J. Abnormal plasma fibrin clot |
| 769 | | characteristics are associated with worse clinical outcome in patients with peripheral |
| 770 | | arterial disease and thromboangiitis obliterans. <i>Atherosclerosis</i> . 2011;215:481-486. |
| 771 | 78. | Okraska-Bylica A, Wilkosz T, Slowik L, Bazanek M, Konieczynska M, Undas A. |
| 772 | | Altered fibrin clot properties in patients with premature peripheral artery disease. |
| 773 | | Polskie Archiwum Medycyny Wewnetrznej. 2012;122:608-615. |
| 774 | 79. | Bhasin N, Ariens RA, West RM, Parry DJ, Grant PJ, Scott DJ. Altered fibrin clot |
| 775 | | structure and function in the healthy first-degree relatives of subjects with intermittent |
| 776 | | claudication. Journal of vascular surgery. 2008;48:1497-1503, 1503 e1491. |
| 777 | 80. | Bhasin N, Parry DJ, Scott DJ, Ariens RA, Grant PJ, West RM. Regarding "Altered |
| 778 | | fibrin clot structure and function in individuals with intermittent claudication". |
| 779 | | Journal of vascular surgery. 2009;49:1088-1089. |
| 780 | 81. | Orfeo T, Gissel M, Butenas S, Undas A, Brummel-Ziedins KE, Mann KG. |
| 781 | | Anticoagulants and the propagation phase of thrombin generation. <i>PloS one</i> . |
| 782 | | 2011;6:e27852. |
| 783 | 82. | Morishima Y, Honda Y. A direct oral anticoagulant edoxaban accelerated fibrinolysis |
| 784 | | via enhancement of plasmin generation in human plasma: dependent on thrombin- |
| 785 | | activatable fibrinolysis inhibitor. Journal of thrombosis and thrombolysis. |
| 786 | | 2019;48:103-110. |
| 787 | 83. | Franchi F, Rollini F, Garcia E, et al. Effects of Edoxaban on the Cellular and Protein |
| 788 | | Phase of Coagulation in Patients with Coronary Artery Disease on Dual Antiplatelet |
| 789 | | Therapy with Aspirin and Clopidogrel: Results of the EDOX-APT Study. <i>Thrombosis</i> |
| 790 | | and haemostasis. 2020;120:83-93. |
| 791 | 84. | Gue YX, Kanji R, Wellsted DM, Srinivasan M, Wyatt S, Gorog DA. Rationale and |
| 792 | | design of "Can Very Low Dose Rivaroxaban (VLDR) in addition to dual antiplatelet |
| 793 | | therapy improve thrombotic status in acute coronary syndrome (VaLiDate-R)" study : |
| 794 | | A randomised trial modulating endogenous fibrinolysis in patients with acute |
| 795 | | coronary syndrome. Journal of thrombosis and thrombolysis. 2020;49:192-198. |
| 796 | 85. | Mega JL, Braunwald E, Wiviott SD, et al. Rivaroxaban in patients with a recent acute |
| 797 | | coronary syndrome. The New England journal of medicine. 2012;366:9-19. |
| 798 | 86. | Alexander JH, Lopes RD, James S, et al. Apixaban with antiplatelet therapy after |
| 799 | | acute coronary syndrome. The New England journal of medicine. 2011;365:699-708. |

87. Eikelboom JW, Connolly SJ, Bosch J, et al. Rivaroxaban with or without Aspirin in 800 Stable Cardiovascular Disease. The New England journal of medicine. 801 2017:377:1319-1330. 802 Anand SS, Caron F, Eikelboom JW, et al. Major Adverse Limb Events and Mortality 803 88. in Patients With Peripheral Artery Disease: The COMPASS Trial. Journal of the 804 American College of Cardiology. 2018;71:2306-2315. 805 89. Coppens M, Weitz JI, Eikelboom JWA. Synergy of Dual Pathway Inhibition in 806 Chronic Cardiovascular Disease. Circulation research. 2019;124:416-425. 807 90. Ammollo CT, Semeraro F, Incampo F, Semeraro N, Colucci M. Dabigatran enhances 808 clot susceptibility to fibrinolysis by mechanisms dependent on and independent of 809 thrombin-activatable fibrinolysis inhibitor. Journal of thrombosis and haemostasis : 810 JTH. 2010;8:790-798. 811 812 91. Konigsbrugge O, Weigel G, Quehenberger P, Pabinger I, Ay C. Plasma clot formation and clot lysis to compare effects of different anticoagulation treatments on hemostasis 813 in patients with atrial fibrillation. Clinical and experimental medicine. 2018;18:325-814 336. 815 816 92. Salta S, Papageorgiou L, Larsen AK, et al. Comparison of antithrombin-dependent and direct inhibitors of factor Xa or thrombin on the kinetics and qualitative 817 characteristics of blood clots. Research and practice in thrombosis and haemostasis. 818 2018;2:696-707. 819 93. Franchi F, Rollini F, Cho JR, et al. Effects of dabigatran on the cellular and protein 820 phase of coagulation in patients with coronary artery disease on dual antiplatelet 821 therapy with aspirin and clopidogrel. Results from a prospective, randomised, double-822 blind, placebo-controlled study. Thrombosis and haemostasis. 2016;115:622-631. 823 94. Connolly SJ, Ezekowitz MD, Yusuf S, et al. Dabigatran versus warfarin in patients 824 with atrial fibrillation. The New England journal of medicine. 2009;361:1139-1151. 825 95. Oldgren J, Budaj A, Granger CB, et al. Dabigatran vs. placebo in patients with acute 826 coronary syndromes on dual antiplatelet therapy: a randomized, double-blind, phase II 827 trial. European heart journal. 2011;32:2781-2789. 828 Uchino K, Hernandez AV. Dabigatran association with higher risk of acute coronary 829 96. events: meta-analysis of noninferiority randomized controlled trials. Archives of 830 internal medicine. 2012;172:397-402. 831 97. Willemse JL, Heylen E, Nesheim ME, Hendriks DF. Carboxypeptidase U (TAFIa): a 832 new drug target for fibrinolytic therapy? Journal of thrombosis and haemostasis : 833 JTH. 2009;7:1962-1971. 834 835 98. Wyseure T, Declerck PJ. Novel or expanding current targets in fibrinolysis. Drug discovery today. 2014;19:1476-1482. 836 99. Kearney K, Tomlinson D, Smith K, Ajjan R. Hypofibrinolysis in diabetes: a 837 therapeutic target for the reduction of cardiovascular risk. Cardiovascular 838 diabetology. 2017;16:34. 839 100. Van De Craen B, Scroyen I, Vranckx C, et al. Maximal PAI-1 inhibition in vivo 840 requires neutralizing antibodies that recognize and inhibit glycosylated PAI-1. 841 Thrombosis research. 2012;129:e126-133. 842 101. Izuhara Y, Yamaoka N, Kodama H, et al. A novel inhibitor of plasminogen activator 843 inhibitor-1 provides antithrombotic benefits devoid of bleeding effect in nonhuman 844 primates. Journal of cerebral blood flow and metabolism : official journal of the 845 International Society of Cerebral Blood Flow and Metabolism. 2010;30:904-912. 846

| 847 | 102. | Zhou X, Hendrickx ML, Hassanzadeh-Ghassabeh G, Muyldermans S, Declerck PJ. |
|-----|------|---|
| 848 | | Generation and in vitro characterisation of inhibitory nanobodies towards |
| 849 | | plasminogen activator inhibitor 1. Thrombosis and haemostasis. 2016;116:1032-1040. |
| 850 | 103. | Elokdah H, Abou-Gharbia M, Hennan JK, et al. Tiplaxtinin, a novel, orally |
| 851 | | efficacious inhibitor of plasminogen activator inhibitor-1: design, synthesis, and |
| 852 | | preclinical characterization. Journal of medicinal chemistry. 2004;47:3491-3494. |
| 853 | 104. | Fortenberry YM. Plasminogen activator inhibitor-1 inhibitors: a patent review (2006- |
| 854 | | present). Expert opinion on therapeutic patents. 2013;23:801-815. |
| 855 | 105. | Rouch A, Vanucci-Bacque C, Bedos-Belval F, Baltas M. Small molecules inhibitors |
| 856 | | of plasminogen activator inhibitor-1 - an overview. European journal of medicinal |
| 857 | | chemistry. 2015;92:619-636. |
| 858 | 106. | Wyseure T, Rubio M, Denorme F, et al. Innovative thrombolytic strategy using a |
| 859 | | heterodimer diabody against TAFI and PAI-1 in mouse models of thrombosis and |
| 860 | | stroke. Blood. 2015;125:1325-1332. |
| 861 | 107. | Singh S, Houng A, Reed GL. Releasing the Brakes on the Fibrinolytic System in |
| 862 | | Pulmonary Emboli: Unique Effects of Plasminogen Activation and alpha2- |
| 863 | | Antiplasmin Inactivation. Circulation. 2017;135:1011-1020. |
| 864 | 108. | Ricklin D, Lambris JD. Complement-targeted therapeutics. <i>Nature biotechnology</i> . |
| 865 | | 2007;25:1265-1275. |
| 866 | 109. | King R, Tiede C, Simmons K, Fishwick C, Tomlinson D, Ajjan R. Inhibition of |
| 867 | | complement C3 and fibrinogen interaction: a potential novel therapeutic target to |
| 868 | | reduce cardiovascular disease in diabetes. Lancet. 2015;385 Suppl 1:S57. |
| 869 | 110. | Ajjan RA, Gamlen T, Standeven KF, et al. Diabetes is associated with |
| 870 | | posttranslational modifications in plasminogen resulting in reduced plasmin |
| 871 | | generation and enzyme-specific activity. <i>Blood</i> . 2013;122:134-142. |
| 872 | 111. | Meltzer ME, Doggen CJ, de Groot PG, Rosendaal FR, Lisman T. Reduced plasma |
| 873 | | fibrinolytic capacity as a potential risk factor for a first myocardial infarction in young |
| 874 | | men. British journal of haematology. 2009;145:121-127. |
| 875 | | |

876 **10 Figure legends**

Figure 1 Overview of coagulation and fibrinolysis activation. Tissue injury exposes 877 collagen enabling platelet adhesion and activation through platelet alpha2beta1 (GPIa/IIa), 878 glycoprotein VI (GPVI) and glycoprotein Ib (GPIb) interacting with von Willebrand factor 879 (vWF). Simultaneously, tissue factor (TF) is exposed activating factor (F) VII to form VIIa/TF 880 complex (extrinsic pathway of coagulation). The VIIa/TF complex subsequently activates a 881 number of proteins in the contact ('intrinsic') and common pathways of coagulation. The 882 activation of these pathways ultimately leads to the accumulation of thrombin which, in turn, 883 stimulates platelet activation via protease-activated receptor 1 or 4 (PAR1/4) and further 884 amplifies activation of coagulation factors in the intrinsic and common pathways. An adequate 885 amount of thrombin finally converts fibrinogen into fibrin and forms double stranded 886 protofibrils. Thrombin also activates FXIII to stabilise the fibrin by crosslinking. The 887 protofibrils then aggregate further into fibrin fibres that generate a network, trapping blood 888 components, before forming a mature thrombus. The fibrinolysis starts after, and even during, 889 fibrin network formation which uncovers binding sites for plasminogen and tissue-890 plasminogen activator (t-PA), enabling plasmin activation and start of the fibrinolytic process. 891

Figure 2 Regulation and alteration of fibrinolysis. Plasmin degradation of fibrin exposes C-892 terminal lysine (lys) residue – the docking site for plasminogen (Plg) and tissue plasminogen 893 activator (t-PA) on α chain of fibrinogen, commencing fibrinolysis. This plasmin-induced 894 proteolysis is seemingly endless and requires other endogenous regulators to limit this process. 895 The interaction between thrombin and thrombomodulin (TM), located on endothelial cells, 896 897 activates thrombin activatable fibrinolysis inhibitor (TAFI) which cleaves lys residue off the fibrin surface, compromising plasminogen binding and fibrinolysis. Activated platelets release 898 plasmin-activator inhibitor 1 (PAI-1) to inhibit t-PA. Moreover, alpha 2 antiplasmin (a2AP) is 899 900 crosslinked into fibrin and inhibits Plg. Complement (C) 3 is a substrate for plasmin which competitively prevent plasmin from cleaving fibrin. Lipoprotein (a) [Lp(a)] has similar 901 homology to plasminogen, competitively binding to fibrinogen, thus, preventing plasmin 902 903 activation. Lp(a) also stimulates PAI-1 secretion by endothelial cells.

11 Tables

Table 1 Summary of studies investigated the impacts of altered fibrinolysis in patients with acute cardiovascular disease.

| Study | Population | Patients | Controls | Lab technique | Outcome | Main findings | | Other |
|---------------------------|------------------|----------|-----------|-----------------|------------|------------------|-------------------|----------|
| | | | | | | Clot strength | Lysis time | comments |
| ACS: Case-control studies | | | | | | | | |
| Undas A. | 40 patients | 40 ACS | 40 stable | 1.Clot | Clot | Patients with | Patients with | |
| (2008) ⁵⁸ | with ACS | patients | angina | permeability | properties | ACS had denser | ACS had | |
| | admitted to | | patients | 2.Turbidimetric | | clot structure | prolonged | |
| | the coronary | | | analysis | | observed by | fibrinolysis time | |
| | care unit | | | 3.Plasma clot | | SEM, lower clot | (p < 0.0001) | |
| | within the | | | lysis assay | | permeability (p | | |
| | first 12 h after | | | 4.SEM | | = 0.001), faster | | |
| | the onset | | | | | fibrin | | |

| | | | | | | polymerization | | |
|-----------------------|----------------|--------------|------------|---------------|------------------------|----------------|---------------|-------------|
| | | | | | | (p = 0.008), | | |
| Mirjam E. | 426 men | 426 men | 646 men | Clot LT | Risk of | NA | overall | OR adjusted |
| (2008) ¹¹¹ | surviving the | surviving | without a | measured by | MI of the | | OR 1.0 | for |
| | first MI, aged | the first MI | history of | turbidimetric | 4 th | | (95%CI: 0.6- | age,BMI, |
| | below 70 | | MI | analysis | quartile | | 1.5) | smoking |
| | years old, | | | | of clot | | | status, |
| | excluded the | | | | LT, | | Age <50 years | presence of |
| | use of | | | | compared | | OR 1.8 | diabetes, |
| | anticoagulants | | | | to the 1 st | | (95%CI: 0.7- | blood |
| | | | | | quartile | | 4.8) | pressures, |
| | | | | | | | | lipid |
| | | | | | | | Age ≥50 years | parameters, |
| | | | | | | | | and C- |

| | | | | | | | OR 0.7 | reactive |
|----------------------|----------------|----------------------|-----------|-------------|-----------|------------------|--------------|----------|
| | | | | | | | (95%CI: 0.4- | protein. |
| | | | | | | | 1.1) | |
| ACS: Prospect | tive studies | | | | | | | |
| Kreutz RP. | 270 Patients | 142 patients | 128 | Whole blood | First | HR 3.8 | NA | |
| (2017) ⁵⁹ | underwent | with TEG- | patients | TEG | recurrent | (95%CI: 1.7-8.3, | | |
| | PCI | MA <u>></u> 35.35 | with TEG- | | MI or | p=0.001) | | |
| | (15.6% | mm | MA | | CVD | | | |
| | STEMI, | | <35.35 | | death, | | | |
| | 22.2% | | mm | | mean | | | |
| | NSTEMI, | | | | follow-up | | | |
| | 24.8% | | | | 2.9 years | | | |
| | unstable | | | | | | | |
| | angina, 35.9% | | | | | | | |
| | stable angina) | | | | | | | |

| Saraf S. | 300 patients | 69 baseline | 231 | GTT | 12-month | NA | HR 2.52 | No |
|----------------------|--------------|----------------------|----------|-----|----------|----|-----------------|-------------|
| $(2010)^{61}$ | with | LT <u>></u> 3000s | baseline | | MACE | | (95%CI: 1.34- | association |
| | hospitalized | | LT<3000s | | | | 4.71, p=0.004) | between OT |
| | ACS | | | | 12-month | NA | HR 4.2 | and MACE |
| | | | | | CV death | | (95%CI: 1.13- | |
| | | | | | | | 15.62, p=0.033) | |
| Farag M. | 496 patients | 70 baseline | 426 | GTT | 12-month | NA | HR 9.1 | LT was not |
| (2019) ¹¹ | presenting | LT <u>></u> 2500s | baseline | | MACE | | (95%CI: 1.34- | altered by |
| | with STEMI | | LT<2500s | | | | 4.71, p<0.001) | standard of |
| | for primary | | | | 12-month | NA | HR 18.5 | care |
| | PCI | | | | CV death | | (95%CI: 7.69- | treatment |
| | | | | | | | 44.31, p<0.001) | including |
| | | | | | | | | DAPT and |
| | | | | | | | | was |

| | | | | | | | | unchanged |
|-------------------------|--------------|----------------------|------------|---------------|----------|----|-----------------|-------------|
| | | | | | | | | at 30 days |
| Christopoulos | 82 patients | 11 baseline | 71 | GTT | 12-month | NA | HR 3.31 | LT<1000 s |
| C. (2017) ⁶⁰ | presenting | LT <u>></u> 3000s | baseline | | MACE | | (95%CI: 1.02- | was |
| | with STEMI | | LT<3000s | | | | 10.78, p=0.045) | associated |
| | for primary | | | | 12-month | NA | HR 4.17 | with |
| | PCI | | | | CV death | | (95%CI: 0.99- | spontaneous |
| | | | | | | | 17.51, p=0.05) | reperfusion |
| Sumaya S. | 4354 | 1082 in | 1098 in | Turbidimetric | 12-month | NS | HR 1.48 | 50% |
| (2018) ¹⁰ | moderate- to | Quartile 4 | Quartile 1 | analysis | CV death | | (95%CI: 1.06- | increase in |
| | high-risk | (LT >888s) | (LT<564s) | | or MI | | 2.06, p=0.027) | LT |
| | ACS patients | | | | 12-month | NS | HR 1.92 | was |
| | | | | | CV death | | (95%CI: 1.19- | associated |
| | | | | | | | 3.1, p<0.001) | with 17% |
| | | | | | | | | increased |

| | | | | | | | | risk of CV |
|---------------|------------------|-------------|----------|-----------------|------------|------------------|------------------|-------------|
| | | | | | | | | death/MI |
| | | | | | | | | and 36% CV |
| | | | | | | | | death alone |
| | | | | | | | | |
| AIS: Case-con | trol studies | | | | | | | |
| Undas A. | 45 patients | 45 patients | 45 age- | 1.Clot | Clot | AIS patients had | AIS patients had | |
| $(2010)^{62}$ | with AIS | | and sex- | permeability | properties | clots with | clots with | |
| | admitted | | matched | 2.Turbidimetric | | 30.5% less | 10.8% longer | |
| | within the | | healthy | analysis | | porous | lysis time | |
| | first 72 h after | | control | | | (p<0.0001), | (p=0.001) | |
| | the onset | | | | | 20.5% more | | |
| | | | | | | compact | | |
| | | | | | | (p<0.0001), | | |

| | | | | | | 17.1% higher | | |
|--------------------------|----------------|-------------|-----------|-----------------|------------|-----------------|----------------|--|
| | | | | | | clot mass | | |
| | | | | | | (p<0.0001), | | |
| | | | | | | 10.2% increased | | |
| | | | | | | overall fiber | | |
| | | | | | | thickness | | |
| | | | | | | (p<0.0001) | | |
| | | | | | | | | |
| AIS: Prospecti | ive studies | | | | | | | |
| Bembenek | 74 Patients | 44 patients | 30 | 1.Clot | Baseline | Patients with | Patients with | |
| JP. (2017) ⁶³ | admitted up to | with good | patients | permeability | clot | poor outcome | poor outcome | |
| | 4.5 h since | outcome at | with poor | 2.Turbidimetric | properties | had clots with | had clots with | |
| | AIS onset, | 3-month | outcome | analysis | | lower baseline | Prolonged | |
| | eligible | (mRS 0-2) | at 3- | | | permeability | baseline LT | |
| | | | month | | | (p<0.01) | (p=0.01) | |

| | for | | | | | | | |
|-------------------|----------------|-------------|------------|-----------------|------------|-----------------|-----------------|-------------|
| | thrombolysis | | | | | | | |
| ALI/CLI: case | e-control | | | | | | | |
| studies | | | | | | | | |
| Karpińska | 43 patients | 43 patients | 43 | 1.Clot | Clots | Patients with | NS | |
| IA. ⁶⁴ | referred for | | controls | permeability | properties | ALI | | |
| | ALI | | without | 2.Turbidimetric | | exhibited 13.4% | | |
| | treatment, off | | history of | analysis | | lower | | |
| | anticoagulant | | ALI or | | | permeability, | | |
| | at least | | vascular | | | p=.001) | | |
| | 3months since | | event | | | | | |
| | the onset | | | | | | | |
| Nowakowski | 85 patients | 85 patients | 47 PAD | 1.Clot | Clots | CLI patients | CLI patients | During a 24 |
| T. ⁶⁵ | with CLI and | with CLI | patients, | permeability | properties | with restenosis | with restenosis | months |
| | symptomatic | and | age-, sex- | | | had clots with | had clots with | follow-up |

| restenosis | symptomatic | and CV | 2.Turbidimetric | 9.5% lower | 12.4% | the |
|--------------|-------------|-----------|-----------------|--------------|--------------|---------------|
| after | restenosis | risk | analysis | permeability | prolonged LT | composite |
| receiving | | matched | | (p<0.001) | | re- |
| percutaneous | | controls | | | | intervention, |
| angioplasty | | (with CLI | | | | amputation |
| within 12 | | =32, | | | | or CV death |
| months | | without | | | | occurred in |
| | | CLI =15) | | | | 63.5% of |
| | | | | | | CLI patients |
| | | | | | | with |
| | | | | | | restenosis |
| | | | | | | and 19.1% |
| | | | | | | of controls |
| | | | | | | (p<0.001) |
| | | | | | | |

ACS = acute coronary syndrome; AIS = acute ischaemic stroke; ALI = acute limb ischaemia; CLI = critical limb ischaemia; mRS = modified

Rankin Scale; GTT = global thrombosis test; TEG = thromboelastography; TEG-MA = TEG maximum amplitude; LT = lysis time; OT =

occlusion time; MACE = major adverse cardiovascular event; CV= cardiovascular; *NA* = not applicable; *NS* = not significant; MI = myocardial infarction; STEMI= ST-elevated myocardial infarction; NSTEMI = ; non ST-elevated myocardial infarction PCI=percutaneous coronary intervention; SEM = scanning electron microscopy;

| Table 2 Summary of studies investigated the impacts of altered fibrinolysis in patients with chronic stable cardiovascular of | disease. |
|---|----------|
| | |

| Study | Population | Patients | Controls | Lab | Outcome | Main fin | ndings | Other |
|----------------------|-------------|-------------|------------|---------------|-------------|---------------|------------|----------|
| | | | | technique | | Clot strength | lysis time | comments |
| Stable CAD: Co | ase-control | | | | | | | |
| studies | | | | | | | | |
| Reddel CJ. | 56 patients | 56 patients | 73 heathy | Turbidimetric | Overall | CAD patients | CAD | |
| (2013) ⁶⁶ | with stable | | volunteers | analysis | coagulatio | had higher | patients | |
| | angina | | | | n property | OCP | had lower | |
| | | | | | (OCP) | (p<0.001) | OFP and | |
| | | | | | Overall | | LT | |
| | | | | | fibrinolysi | | (p<0.001) | |
| | | | | | s property | | | |
| | | | | | (OFP) | | | |

| Siegerink B. | 380 young | 205 MI | 638 age- | Turbidimetric | LT | NA | 75.2±25 | The study |
|--------------|---------------|----------|----------|---------------|----|----|------------|----------------|
| (2012) 67 | women age | patients | matched | analysis | | | min vs | contained |
| | 18-50 years, | | women | | | | 64.4±14 | several biases |
| | diagnosed | | | | | | min (no p- | from |
| | with arterial | | | | | | value | confounders, |
| | thrombosis | | | | | | provided) | drop-out rate |
| | (205 with | | | | | | | and blood |
| | MI and 175 | | | | | | | sample |
| | with | | | | | | | collection |
| | ischaemic | | | | | | | |
| | stroke) | | | | | | | |
| | | | | | | | | |

| Tantry US. | 171 patients | 33 unstable | 71 stable | TEG | TEG- | Unstable | NA | Data |
|----------------------|--------------|-----------------|-----------------|-----|-------------------|----------------|----|--------------|
| (2010) ⁶⁹ | with | angina patients | angina patients | | MA _{ADP} | angina | | presented in |
| | asymptomati | | and | | | patients | | figures |
| | c stable | | 67 | | | produced the | | |
| | CAD, stable | | asymptomatic | | | strongest | | |
| | angina, and | | CAD patients | | | clots compare | | |
| | unstable | | | | | to | | |
| | angina | | | | | asymptomatic | | |
| | | | | | | and stable | | |
| | | | | | | angina groups | | |
| | | | | | | (p<0.001 and | | |
| | | | | | | 0.02, | | |
| | | | | | | respectively) | | |
| | | | | | | Stable angina | | |
| | | | | | | patients had | | |
| | | | | | | stronger clots | | |

| | | | | | | than | | |
|---------------|--------------|--------------|------------|---------------|------------|--------------|-----------|---------------|
| | | | | | | asymptomatic | | |
| | | | | | | one (p=0.02) | | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | | | | | |
| Neergaard- | 177 patients | 116 CAD with | 61 CAD | Turbidimetric | Clot | MA | LT | CAD with MI |
| Neergaaru- | 177 patients | 110 CAD with | 01 CAD | Turbianneuric | Clot | MA | | CAD with MI |
| Petersen S. | with stable | MI | without MI | analysis | properties | [median | [median | patients were |
| $(2013)^{68}$ | CAD with at | | | | | (IQR)] | (IQR)] | older and had |
| | least 50% | | | | | 0.48 (0.41; | 552 (498; | lower LDL |
| | stenosis | | | | | 0.52) | 756) | cholesterol |
| | | | | | | vs. | VS. | |
| | | | | | | 0.42 (0.38; | 519 (468; | |
| | | | | | | 0.50) | 633) | |
| | | | | | | p = 0.02 | | |

| | | | | | | | seconds | |
|----------------|--------------|-----------------|-----------------|---------------|-----------|----------------|----------|----------------|
| | | | | | | | p = 0.02 | |
| | | | | | | | | |
| | | | | | | | | |
| | | | | SEM | clot | Thinner fibres | NA | |
| | | | | | structure | (mean±SD) | | |
| | | | | | | 119.7±27.5 | | |
| | | | | | | vs. | | |
| | | | | | | 127.8±31.1 | | |
| | | | | | | nm | | |
| | | | | | | p = 0.003 | | |
| Stable CAD: Pr | ospective | | | | | | | |
| studies | | | | | | | | |
| Neergaard- | 786 patients | 197 in quartile | 196 in quartile | Turbidimetric | MACE | Clot A | UC | Clot AUC |
| Petersen S. | with stable | 1 (Clot AUC | 4 (Clot AUC= | analysis | | Adjusted I | HR: 2.4 | represents the |
| $(2020)^{72}$ | CAD with at | =121-195) | 585-2005) | | | (1.2–4.8), | p=0.01 | net effect |

| | least 50% | | | | | MA | LT | between OCP |
|----------------------|---------------|----------------|----------------|-----|------------|-----------------------|------------|------------------------|
| | stenosis, | | | | | Adjusted HR: | Adjusted | and OFP |
| | 90% had | | | | | 1.8 | HR: 1.6 | |
| | prior MI | | | | | (0.9-3.7), | (0.8-3.0), | |
| | | | | | | p=NS | p=NS | |
| Hou X. | 759 patients | 58 patients | 701 patients | TEG | Whole | TEG-MA _{ADP} | NA | TEG-MA >34 |
| (2017) ⁷¹ | underwent | with an | without | | blood clot | 40.8 ± 10.1 | | mm predicted |
| | elective PCI, | ischaemic | ischaemic | | strength | mm vs 26.7 ± | | ischaemic |
| | excluded | event in 2 | event | | | 13.7 mm, | | events after |
| | Acute MI | years | | | | p<0.001 | | PCI |
| | within 48h | | | | | | | |
| | | | | | | | | |
| Gurbel PA. | 84 patients | 21 patients in | 63 patients in | TEG | Ischaemic | Ischaemic | NA | TEG- |
| $(2009)^{70}$ | underwent | quartile 4 | quartile 1-3 | | events | events | | MA _{ADP} ≥68m |
| | elective PCI, | | | | | Q4 = 48% | | m (quartile3- |

| | excluded | (TEG- | (TEG- | | within 2 | VS | | 4) probably |
|----------------------|---------------|-------------------------|-------------------------|---------------|-------------|--------------|-----------|-------------|
| | Acute MI | MA _{ADP} ≥71mm | MA _{ADP} <71mm | | years | Q1 = 13%, | | predicted |
| | within 48h |) |) | | (MACE | p=0.02 | | ischaemic |
| | | | | | and | Q2 = 15%, | | events |
| | | | | | hospitalise | p=0.03 | | |
| | | | | | d angina) | Q3 = 30%, | | |
| | | | | | | p=0.24 | | |
| Previous ischae | emic stroke: | | | | | | | |
| Case-control st | udies | | | | | | | |
| Anžej S. | Patients with | 44 patients | 46 Healthy | Turbidimetric | OCP and | NS | Patients | |
| $(2007)^{73}$ | previous | | control | analysis | OFP | | had lower | |
| | AIS, age <45 | | | | | | OFP | |
| | years old | | | | | | (p<0.001) | |
| Undas A. | 147 patients | 147 patients | 120 healthy | 1.Clot | Clot | Patients had | Patients | |
| (2009) ⁷⁴ | with or | | controls | permeability | properties | clots with | had clots | |

| | without | | | 2.Turbidimetri | | lower | with | |
|----------------------|--------------|-------------|-----------------|-----------------|----|----------------------|------------|--|
| | patent | | | c analysis | | permeability | prolonged | |
| | foramen | | | 3.SEM | | (<i>p</i> <0.0001), | clot LT | |
| | ovale and a | | | | | faster fibrin | (p<0.0001) | |
| | history of | | | | | polymerizatio | | |
| | first-ever | | | | | n (p<0.0001), | | |
| | ischemic | | | | | and increased | | |
| | stroke | | | | | fibre diameter | | |
| | | | | | | and | | |
| | | | | | | density | | |
| Vučković BA. | 60 ischaemic | 60 patients | 30 age- and | Euglobulin | LT | NA | 219.7±78.8 | |
| (2010) ⁷⁵ | stroke | | sex-matched | clot lysis time | | | min vs | |
| | patients | | health controls | | | | 183.5±58.2 | |
| | | | | | | | min | |
| | | | | | | | (p=0.005) | |

| | (90% had | | | | | | | |
|--------------|---------------|-----------------|----------|---------------|----|----|------------|----------------|
| | stroke within | | | | | | | |
| | 1year) | | | | | | | |
| Siegerink B. | 380 young | 175 ischaemic | 638 age- | Turbidimetric | LT | NA | 68.1.2±36. | The study |
| (2012) 67 | women age | stroke patients | matched | analysis | | | 3 min vs | contained |
| | 18-50 years, | | women | | | | 64.4±14 | several biases |
| | diagnosed | | | | | | min (no p- | from |
| | with arterial | | | | | | value | confounders, |
| | thrombosis | | | | | | provided) | drop-out rate |
| | (205 with | | | | | | | and blood |
| | MI and 175 | | | | | | | sample |
| | with | | | | | | | collection |
| | ischaemic | | | | | | | |
| | stroke) | | | | | | | |
| | | | | | | | | |

| Previous isch | aemic stroke: | | | | | | | |
|----------------------|---------------|----------------|--------------|-----|-------------------|-------------|----|-------------------|
| prospective studies | | | | | | | | |
| Wang B. | 218 patients | 18 patients | 200 patients | TEG | Baseline | 41.57±15.10 | NA | The incidence |
| (2017) ⁷⁶ | who | with recurrent | without | | TEG- | VS | | of ischemic |
| | underwent | ischaemic | ischaemic | | MA _{ADP} | 33.50±13.86 | | events in |
| | stenting for | stroke or TIA | event | | | mm | | patients with |
| | extracranial | events | | | | (p=0.020) | | MA _{ADP} |
| | or | (9 events | | | | | | >49.95 mm |
| | intracranial | occurred | | | | | | was markedly |
| | artery (70- | within 7 days) | | | | | | higher than in |
| | 99%) | | | | | | | patients with |
| | stenosis | | | | | | | MAADP |
| | | | | | | | | ≤49.95 mm |
| | | | | | | | | (20.8% vs |

| | | | | | | | | 6.7%, |
|----------------------|---------------------------|--------------|----------------|-----------------|------------|---------------|------------|-----------|
| | | | | | | | | p=0.018). |
| PAD: Case-cont | PAD: Case-control studies | | | | | | | |
| Undas A. | 106 patients | 106 patients | 106 age-, sex- | 1.turbidimetric | Clot | Patients had | Patients | |
| (2011) ⁷⁷ | with PAD | | and CV risk | analysis | properties | clots with | had clots | |
| | (ABI <u>≤</u> 0.9) | | matched | 2. clot | | 18.8% lower | with | |
| | aged <u><</u> 70 | | controls | permeability | | clot | 30.6% | |
| | years | | | | | permeability | prolonged | |
| | | | | | | (p = 0.005), | clot lysis | |
| | | | | | | 35.3% faster | time (p = | |
| | | | | | | fibrin | 0.003) | |
| | | | | | | polymerizatio | | |
| | | | | | | n (p<0.001), | | |

| | | | | | | 22.4% higher | |
|---------------------------|----------------------|-----------------|-------------|-----------------|------------|---------------|-------------|
| | | | | | | МА | |
| | | | | | | (p<0.001) | |
| Okraska-Bylic | 31 patients | 31 patients | 40 age- and | 1.turbidimetric | Clot | Patients had | Patients |
| a A. (2012) ⁷⁸ | with | | sex-matched | analysis | properties | clots with | had clots |
| | symptomatic | | controls | 2. clot | | 32% lower | with 7% |
| | premature | | | permeability | | clot | longer clot |
| | PAD | | | | | permeability | lysis time |
| | (ABI <u>≺</u> 0.9) | | | | | (p<0.001) | (p = 0.004) |
| | patients aged | | | | | | |
| | <u><</u> 55 years | | | | | | |
| | old, | | | | | | |
| Bhasin N. | 106 male | 106 male first- | 107 healthy | 1. | Clot | Male first- | Male first- |
| (2008) ⁷⁹ | first-degree | degree | male, age- | turbidimetric | properties | degree | degree |
| | relatives of | relatives | | analysis | | relatives had | relatives |

| male patients | matched | 2. clot | clots with | had slower | |
|---------------|----------|--------------|--------------|-------------|--|
| with | controls | permeability | higher MA | clot lysis | |
| intermittent | | 3. LSCM | and fibre | velocity on | |
| claudication | | | thickness on | LSCM | |
| | | | LSCM | (p=0.018) | |
| | | | (p<0.001)_ | | |
| | | | | | |

CAD = coronary artery disease; PAD = peripheral artery disease; TIA = transient ischaemic attack; ABI = ankle-brachial index; OCP = overall coagulation property; OFP = overall fibrinolysis property; MA = maximum absorbance; AUC = area under the curve; LT = lysis time; TEG = thromboelastography; TEG-MA_{ADP} = TEG-maximum amplitude of ADP-induced platelet-fibrin clot strength; LSCM, laser scanning confocal microscopy; MACE = major adverse cardiovascular event; CV= cardiovascular;*NA*= not applicable;*NS*= not significant; MI = myocardial infarction; SEM = scanning electron microscopy.