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1 Supplementary Figures

HELIX 8

Xenopus	YNPVIYIVLNKQFR NCL ITTLCCGKNPFGDEDGSS-AATSKTEASSVSSSQVSPA
Frog	$\verb"YNPVIYIMLNKQFRncmittlccgknpfgdddass-aatskteatsvstsqvspa"$
Shark	YNPLIYILLNKQFR NCMITTLCCGKHPFEEDESTS-AAASKTEASSVSSSQVSPA
Pufferfish	YNPLIYICMNKQFRHCMITTLCCGKNPFEEEEGAS-TT-SKTEASSVSSSSVSPA
Ayu	$\verb"YNPLIYVCMNKQFRHCMITTLCCGKNPFEEEEGAS-TTASKTEASSVSSSSVAPA"$
Carp	$\verb"YNPCIYICMNKQFRHCMITTLCCGKNPFEEEEGAS-TTASKTEASSVSSSSVSPA"$
Zebrafish	$\verb"YNPCIYICMNKQFRHCMITTLCCGKNPFEEEEGAS-TTASKTEASSVSSSSVSPA"$
Guppy	$\verb"YNPLIYICMNKQFRHCMITTLCCGKNPFEEEEGAS-TTASKTEASSVSSSSVSPA"$
Medaka	YNPAIYICMNKQFRNCMITTLCCGKNPFEEEEGAS-TTASKTEASSVSSSSVSPA
Turtle	YNPIIYVLMNKQFR NCMITTICCGKNPFGDDDVSSTVSQSKTEVSSVSSSQVSPA
Alligator	$\verb"YNPVIYIVMNKQFRncmittlccgknplgddeta-tgsktetssvstsqvspa"$
Chick	$\verb"YNPVIYIVMNKQFRncmittlccgknplgdedts-ag-ktetssvstsqvspa"$
Lamprey	$\verb"YNPVIYILMNKQFRncmittlccgknplgddesgastsktevssvstspvspa"$
Platypus	YNPVIYIMMNKQFR NCMLTTICCGKNPLGDDEASATASKTEQSSVSTSQVSPA
Guinea Pig	YNPVIYIMMNKQFR NCMLTTICCGKNPLGDDEASTTVSKTETSQVAPA
Human	YNPVIY IMMNKQFR NCMLTTICCGKNPLGDDEASATVSKTETSQVAPA
Cat	YNPVIYIMMNKQFR NCMLTTLCCGKNPLGDDEASTTGSKTETSQVAPA
Pig	YNPVIYIMMNKQFR NCMLTTLCCGKNPLGDDEASTTTSKTETSQVAPA
Bovine	YNPVIYIMMNKQFR NCMVTTLCCGKNPLGDDEASTTVSKTETSQVAPA
Dog	YNPVIYIMMNKQFR NCMITTLCCGKNPLGDDEASASASKTETSQVAPA
Mouse	YNPVIYIMLNKQFR NCMLTTLCCGKNPLGDDDASATASKTETSQVAPA
Rat	YNPIIYIMMNKQFR NCMLTTLCCGKNPLGDDEASATASKTETSQVAPA
	*** **: :*****:*::**:*: ::: *** :**:*:

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3 Fig. S1 The alignment of the CT44 C-terminal sequences from vertebrate species as 4 indicated. The methionine at position 317 is highlighted in red. In Xenopus, it is substituted 5 by leucine. Alligator, Alligator mississippiensis; Ayu, Plecoglossus altivelis; Bovine, Bos 6 taurus; Carp, Cyprinus carpio; Cat, Felis catus; Chick, Gallus gallus; Dog, Canis lupus 7 familiaris; Frog, Lithobates pipiens; Guinea pig, Cavia porcellus; Guppy, Poecilia 8 reticulate; Human, Homo sapiens; Lamprey, Lethenteron camtschaticum; Medaka, Oryzias 9 latipes; Mouse, Mus musculus; Platypus, Ornithorhynchus anatinus; Pig, Sus scrofa; 10 Pufferfish, Tetraodon nigroviridis; Rat, Rattus norvegicus; Shark, Callorhinchus milii; 11 Turtle, Chelonia mydas; Xenopus, Xenopus laevis; Zebrafish, Danio rerio.



13 Fig. S2 Comparison of OS-targeting efficiency of Xenopus and human opsin C-terminal tails 14 (CT44). (A) Alignment of rod opsin C terminal amino acids from human and Xenopus and the AAR sequence applied in this assay. Palmitoylated cysteines are highlighted in red. (B-F) 15 16 Confocal images of cryosections through the photoreceptor cell layer of wild-type zebrafish 17 retinae that transiently express the following variants of the EGFP-CT44 construct at 5 dpf: (B) 18 XCT44; (C) HCT38; (D) H-M317L; (E) AAR-X; (F) AAR(F>L)-X. Confocal image (left) and 19 a heat map (right) of GFP signal intensity are juxtaposed. (G) Quantification of fluorescence 20 intensity in photoreceptor cell bodies. Each dot represents a measurement from a single 21 photoreceptor cell. Data are from 3 independent experiments. Mean and 95% confidence 22 interval are indicated. Sample sizes are provided in italics below the horizontal axis. In 23 schematic drawings above panels (B-F), human, Xenopus and AAR sequences are color-coded in light blue, dark blue and magenta, respectively. Red bars indicate conserved cysteine 24 25 residues in opsin C terminus. Data are log transformed for statistical analysis as described in 26 methods.

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28 Fig. S3 Transport efficiency of opsin/melanopsin hybrid GPCRs into the photoreceptor ciliary 29 compartment. (A) Images of cryosections through zebrafish retinae, expressing wild-type or 30 hybrid GPCRs, schematically shown to the left of each image. In each panel, a confocal image 31 (left) and a heat map (right) of GFP signal intensity are shown. (B) Quantification of the GFP 32 signal intensity in photoreceptor cell bodies for wild-type and hybrid GPCRs shown in (A). 33 Data from 4 independent experiments are shown. Sample sizes are provided in italics below 34 the horizontal axis. (C-D) Confocal images of PAC2 cells showing the expression of wild-type 35 opsin (C) and melanopsin-No3 (ML-NO.3) (D) (green) in relation to the ER marker GRP78/Bip 36 (red). Merged (left), GFP (middle) and ER (right) images are provided. Nuclei are stained with 37 DAPI (blue). (E) Counts of GFP-positive cells in retinae expressing four opsin/Sstr5 hybrid 38 constructs at 5 and 12 hours after heat-shock as indicated. Each data point represents a single section. Data are from 4 independent experiments. Mean and 95% confidence interval are 39 40 shown. Mann-Whitney test is used. Scale bars represent 10µm.



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Fig. S4 Confocal images of PAC2 cells showing the expression of wild-type opsin and deletion
variants (green) in relation to the ER (GRP78/Bip) (left, red) and the plasma membrane (4D2)
(right, red). Nuclei are stained with DAPI (blue). Scale bars represent 10μm.

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46 Table S1 Amino acid sequences of Sstr5, rod opsin, opsin/Sstr5 hybrids, melanopsin and 47 opsin/melanopsin hybrid appended with GFP. The Sstr5 sequence is in pink, rod opsin 48 sequence is in blue, melanopsin sequence is in purple and GFP sequence is in black. For Sstr5 49 and opsins, letters in upper case indicate transmembrane regions. Cytoplasmic and extracellular 50 sequences are in lower case. Cytoplasmic sequences are underlined.