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1    **The evolution of structural colour in butterflies**

2    **Victoria J. Lloyd<sup>1\*</sup> and Nicola J. Nadeau<sup>1</sup>**

3    *<sup>1</sup>Animal and Plant Sciences, University of Sheffield, Alfred Denny Building, Western bank,*  
4    *Sheffield S10 2TN, UK*

5

6    \* Corresponding author

7    Email Victoria Lloyd: [vjllloyd1@sheffield.ac.uk](mailto:vjllloyd1@sheffield.ac.uk)

8    Email Nicola Nadeau: [n.nadeau@sheffield.ac.uk](mailto:n.nadeau@sheffield.ac.uk)

9

10    **Abstract**

11    Butterflies display some of the most striking examples of structural colour in nature. These  
12    colours originate from cuticular scales that cover the wing surface, which have evolved a  
13    diverse suite of optical nanostructures capable of manipulating light. In this review we  
14    explore recent advances in the evolution of structural colour in butterflies. We discuss new  
15    insights into the underlying genetics and development of the structural colours in various  
16    nanostructure types. Improvements in -omic and imaging technologies have been paramount  
17    to these new advances and have permitted an increased appreciation of their development and  
18    evolution.

19

20    **Keywords**

21    Structural colour, butterfly, evolution, development, iridescence

## 22 **Introduction**

23 In nature some of the most conspicuous colours come not from pigments but instead from  
24 physical structures within the integument of some animals and plants [1,2]. These structures,  
25 on the order of a few hundred nanometres or less, selectively reflect light to create a vivid  
26 repertoire of colours known as ‘structural colours’ (**box 1**). A diverse range of organisms  
27 produce structural colours, including birds [3], plants [2], fish [4] and invertebrates [5–8].  
28 Structural colours have evolved to fill diverse roles including camouflage [9] and intra- [10]  
29 and interspecific communication [11].

30 Butterflies have long been subjects of study for their pigmentary colours, and they have  
31 also been firmly at the forefront of our understanding of structural colouration. Vivid displays  
32 of structural colours are found across the butterfly phylogeny and utilize diverse optical  
33 mechanisms (**Figure 1**)[12••]. Nevertheless, all butterfly structural colours originate from  
34 tiny (approximately 100µm long) scales which adorn the wing surfaces. Each scale is a  
35 flattened cuticular extension, with an intricate upper lamina (layer) of parallel ridges  
36 connected transversely by crossribs which extend down through a lumen to a flat lower  
37 lamina (**Figure 1a**)[13,14]. Remarkably, each highly-intricate scale is a result of the  
38 expansion of a single cell [14]. Pigments are also crucial for scale colour, with spectral purity  
39 of structurally coloured scales achieved by including pigments, such as melanin, into the  
40 scale structure [15,16••]. Additional mechanisms such as scale stacking can further modify  
41 and enhance the colour [17].

42 Varied ecological pressures and the underlying malleability of the arthropod cuticle has  
43 contributed to the evolution of diverse structural colour mechanisms in butterflies [18,19].  
44 While the vivid blue scales of *Morpho* are a classic example of a complex scale reflector  
45 [20], bright colouration also occurs in other species, such as metalmarks (Riodininae), which

46 have a comparatively simple optical nanostructure [12••]. Indeed, even within a single genus  
47 several divergent optical nanostructures may be found [21], highlighting the flexibility of the  
48 underlying cellular mechanisms governing the evolution and development of structural  
49 colour.

50 In this review we aim to highlight the recent advancements in understanding butterfly  
51 structural colour evolution from a genetic and developmental perspective. We group butterfly  
52 optical nanostructures into three major types based on their location within the scale and  
53 discuss each one in turn. The seminal works of Ghiradella [22–24] have served as a  
54 foundation for understanding both the optics of butterfly structural colours as well as their  
55 development *in vivo*. Following a period of relatively little activity, the last five years has  
56 seen tremendous advancements, including new insights into the genes controlling structural  
57 colour [16••,25••,26]. These insights have been aided by the molecular revolution of the past  
58 decade, with tools such as CRISPR-Cas9 pioneering our understanding of the underlying  
59 genetics. Furthermore, advances in microscopy technologies have reaffirmed our appreciation  
60 for wing scale formation at the nanoscale.

61

## 62 **Surface nanostructures – ridge reflectors**

63 Several features of the upper scale surface can be modified into reflective structures (**Figure**  
64 **1**). The parallel ridges, which all butterfly scales possess, can become elaborated into a series  
65 of layered lamellae, creating an intense reflection through constructive interference (**Box 1**)  
66 [24]. Remarkably, this can result in around 75% of the light being reflected in cases such as  
67 the blue *Morpho* scale [27]. However, the light reflected by these structures is not solely  
68 limited to the human visible spectrum, for example UV reflectance is present in species such  
69 as *Eurema lisa* [24]. Ridge reflectors have evolved independently in multiple lineages and in

70 some cases several times within a lineage, for example in *Heliconius*, driven by convergent  
71 evolution for mimicry [28•].

72 Ridge reflectors form during wing scale development, with alternating air and chitin layers  
73 attaining optically precise spacing through drying post-eclosion [19]. Electron micrographs  
74 by Ghiradella [23] led her to propose that ridge multilayers form by elastic buckling of the  
75 cuticle in response to intracellular stresses. Notably though, this hypothesis draws on a  
76 purportedly similar process Locke [29] proposed for the formation of taenidial folds in the  
77 tracheae of insects; where cuticle folds formed through mechanical buckling [23,29].  
78 Subsequent re-evaluation of taenidial fold formation now suggests a more active role of the  
79 actin cytoskeleton and chitin synthases, rather than mechanical buckling [30,31]. While much  
80 work is still needed on the development of ridge reflectors it is plausible that these  
81 nanostructures also require an active role for the actin cytoskeleton and chitin synthases in  
82 their formation. Indeed, recent studies have highlighted the importance of the actin  
83 cytoskeleton in wing scale development [32,33].

84 Brien *et al.*, [25••] present some of the first insights into the genetic basis of ridge reflector  
85 evolution. Using phenotypic analyses of crosses between iridescent and non-iridescent races  
86 of *Heliconius erato*, they show that structural colour is a quantitative trait not associated with  
87 any of the major effect loci previously linked to pigmentary wing patterning in *Heliconius*.  
88 Interestingly, a moderate effect locus was detected on the sex chromosome, reminiscent of  
89 much older work [34] showing that differences in ridge-reflector-based UV colouration  
90 between *Colias* species is due to a sex-linked locus.

91 Perhaps unsurprisingly, given the precise nanostructures involved, ridge reflector  
92 formation shows sensitivity to developmental conditions in many species [35,36•]. Together,  
93 with a sex-linked genetic architecture, this has interesting implications for the evolution of

94 ridge reflector colour as a sexual signal. Sex-linkage would aid the evolution of sexually  
95 dimorphic colour, important if the sexes have different evolutionary optima; while condition  
96 dependence could increase the information content of these colour signals to potential mates.

97

### 98 **Lower lamina reflectors**

99 The scale's lower-most cuticle layer can function as a thin-film capable of scattering light  
100 (**Box 1**). Often this nanostructure is accompanied by pigments which function as 'optical  
101 filters' by absorbing wavelengths of light to enhance the structural colour [15]. Lower lamina  
102 reflectors are often considered an optically and developmentally simple mechanism of  
103 achieving structural coloration. From an optical perspective, the lower lamina is a simple  
104 interface of contrasting refractive indexes which can scatter light [16••]. This optical process  
105 is reminiscent of that present in ancient Lepidopteran structurally coloured scales from the  
106 Jurassic (albeit possessing a slightly different scale morphology of a fused lower and upper  
107 lamina) [37•]. Lower lamina reflectance is an "evolutionarily accessible" optical phenotype  
108 because all scales possess a flat lower lamina. As such, this removes the need to evolve  
109 complex ridge or crossrib structures, which may impact on other intrinsic scale properties  
110 including thermoregulation, aerodynamics, hydrophobicity or self-cleaning [38–40].

111 Artificial selection experiments in both *Bicyclus anyana* [41] and *Junonia coenia* [16••]  
112 have demonstrated that within a short period of time (6 generations and 12 generations,  
113 respectively) the colour produced by lower lamina reflectors can be considerably modified  
114 (**Figure 2**). Selection in both these species resulted in changes in thickness of the lower  
115 lamina, demonstrating the ease with which this scale element can be sculpted. Indeed, in both  
116 *Bicyclus* and *Junonia*, between-species variation in structural colour appears to have evolved  
117 through tuning the lower lamina thickness. This reinforces suggestions that the lower lamina

118 is a common evolutionary target for selection in diverse species and that relatively small,  
119 quantitative changes to just one part of the scale architecture can have profound micro- and  
120 macroevolutionary consequences [16••,42].

121 Zhang *et al.* [26] demonstrated that knockouts of the developmental patterning gene *optix*  
122 also results in a brown to blue colour change in *J. coenia*. Thayer *et al.* [16••] showed this  
123 switch in colouration through *optix* deletion was the result of lower lamina thickening,  
124 recapitulating what occurs in both the artificial selection experiments and within natural  
125 populations (**Figure 2**). This hints at the possibility of *optix* and its associated gene regulatory  
126 networks being the target for selection in naturally evolving, structurally-coloured  
127 populations. Interestingly, Thayer *et al.*, [16••] also showed that in other coloured scales of  
128 *Junonia*, such as gold scales, the lower lamina was tuned to a thickness to produce a  
129 complementary reflected wavelength. By regulating lamina thickness, *optix* and its  
130 downstream targets could control the range of wavelengths produced through thin film  
131 interference [16••]. Future work on the downstream targets of *optix* should aid our  
132 understanding of the regulatory networks and cellular control of structural colouration.

133 *Optix* also plays a conserved role in pigmentation and scale structure in butterflies  
134 [26,43]. This suggests that the evolution of pigments and some nanostructures may be more  
135 intertwined than previously thought and may be controlled by a few ‘adaptive hotspots’ such  
136 as *optix* [26]. Indeed, several recent studies have highlighted a regulatory link between both  
137 scale structure and pigments [16••,26,44,45••,46]. Future studies should continue to address  
138 the link between scale structure, pigments and nanostructures.

### 139 **Internal nanostructures – lumen multilayers and photonic crystals**

140 Diverse nanostructures may be present within the scale lumen, ranging from simple  
141 multilayers to complex 3D photonic crystals (**Figure 1**) [21]. While often considered as

142 highly-ordered structures, lumen nanostructures may also encompass a degree of disorder, for  
143 instance in the lumen multilayers of *Ornithoptera* (birdwing) scales [47].

144 Focusing on photonic crystal formation, Ghiradella *et al.* [22] provided the first insights  
145 into the role of the smooth endoplasmic reticulum (SER) as a templating network for highly  
146 ordered chitin deposition within the developing scale. Subsequently, electron microscopy  
147 deduced a gyroid structure of the photonic crystals, consistent with a process of templating by  
148 intracellular membranes (**Box 1**)[48]. Recently, techniques such as SAXS and X-ray  
149 tomography have permitted deeper understanding of not just the optical properties of such  
150 highly-ordered structures but also the underlying cellular processes guiding their formation  
151 [49,50]. Indeed, recent insights by Wilts *et al.* [50] showed that the photonic crystals in the  
152 scales of *Thecla opisena* were arranged in a size gradient over the scale's proximo-distal axis.  
153 Investigations over this gradient demonstrated a time-dependent growth process of crystal  
154 formation and that crystals do not form in the same orientation. The authors concluded that  
155 rather than a pre-folding template of SER, gyroid formation more likely involves  
156 simultaneous membrane templating and chitin deposition.

157 Characterising the optical properties of diverse luminal structures while considering the  
158 underlying phylogenetic relationships has provided deeper insights into the evolution of such  
159 structures. For example, gyroid structures and multilayers are found in closely-related species  
160 of Cattlehearts (*Parides*). This may suggest an underlying commonality in the developmental  
161 pathways and cellular effectors governing such structures. Wilts *et al.* [21] suggest that minor  
162 deviations in developmental parameters may shift scale cell fate between multilayers or  
163 gyroid structures within the scale lumen.

164 Similarly, Ren *et al.*, [12••] used comparative studies across butterfly families to  
165 understand the diversity of scale ultrastructures underlying metallic reflectance. Scales of

166 Lycaenids were able to produce metallic, silver scales through an internal multilayer-type  
167 architecture. The authors suggested that differences in the number of chitin layers and  
168 perforation of the upper lamina could have led to a transition from blue to silver scales. The  
169 next major breakthrough will be in identifying the molecular switches involved in these  
170 evolutionary transitions between nanostructure types.

171

## 172 **Beyond butterflies – evolutionary insights from other systems**

173 We are beginning to gain an appreciation of the underlying evolutionary development of  
174 structural colours in many systems. Knowledge from these systems may present unique  
175 insights into the evolution of structural colour in butterflies. For example, photonic crystals  
176 within weevils evolved only once, involving a transition from a hollow scale to one in which  
177 the spongy network within the lumen becomes increasingly ordered [51•]. This bears  
178 similarity to the evolution of photonic crystals in butterflies, which likely required a transition  
179 from a hollow lumen to one with an increasingly ordered multilayer [21]. In weevils,  
180 photonic crystal evolution was associated with shifts in feeding strategy and the need for  
181 crypsis [51•], raising the question as to whether similar ecological pressures also shaped  
182 butterfly structural colour evolution.

183 While much focus has been placed on structural colouration in animals, diverse structural  
184 colours are also present in the petals, leaves and fruits of plants [2]. Remarkably, despite  
185 disparity in cell types and cuticular materials used for nanostructures, many of the underlying  
186 cellular principles are likely conserved between animals and plants. For example, multilayer  
187 reflectors in fruits form through microtubule cytoskeleton guided deposition of cellulose into  
188 layered, helicoidal structures [2]. In butterfly scales, the actin cytoskeleton guides chitin  
189 deposition on the ridges and may also play a role in nanostructure formation [32].

190 Additionally, light-reflecting ridges on the epidermal layer of petals form through buckling of  
191 the cuticle [52], similar to the suggested mechanism of ridge reflector formation in  
192 butterflies, which could involve stress-mediated buckling of the chitin cuticle [24]. Overall,  
193 different systems may give universal insights into the underlying principles governing  
194 structural colour evolution.

195

## 196 **Conclusions**

197 Butterflies have evolved a diverse suite of optical nanostructures to produce vivid displays of  
198 structural colour. Advances in molecular genetic tools, such as CRISPR, in addition to much  
199 improved resolution of microscopy techniques have provided tantalising new insights into the  
200 evolution and development of optical nanostructures in butterflies. Whilst much of the  
201 underlying cellular dynamics and developmental pathways remain unknown, the next few  
202 years is set to see major advances in our knowledge of structural colour evolution. By its  
203 very nature, the study of structural colours is a highly interdisciplinary topic, involving  
204 collaborations between physicists, material scientists and biologists alike. Such collaborations  
205 will not just give unique insights into evolutionary processes governing structural colouration  
206 but will open the doors to a whole range of biomimetic technologies, taking inspiration from  
207 the photonic structures which evolution has been finely sculpting over millennia.

208

209

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### 217 **Conflict of interest statement**

218 The authors declare no conflict of interest.

219

### 220 **References and recommended reading**

221 Papers of particular interest, published within the period of review, have been highlighted as:

222 • of special interest

223 •• of outstanding interest

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381

## 382 **Special interest**

### 383 **•• Ren et al., 2020**

384 This study provides insights into how metallic colouration in diverse butterfly families has  
385 convergently evolved. Using analyses of spectral properties as well as investigations of scale  
386 ultrastructure the authors gain insights into the underlying mechanisms governing metallic  
387 colour production, demonstrating a convergence in optical principles such as spatial colour  
388 mixing and an unperforated upper lamina. They describe several different nanostructure  
389 modifications resulting in the production of metallic colouration, including a potential  
390 transition from a blue producing multilayer to a silver producing multilayer in Lycaenidae.

### 391 **•• Thayer et al., 2020**

392 This article provides insights into the structural and genetic control of lower lamina  
393 reflectance. Using investigations of scale architecture in artificially-selected *Junonia coenia*  
394 the authors show changes to lower lamina thickness is responsible for dramatic shifts from  
395 brown to blue colour. The authors further show that CRISPR-Cas9 deletion of *optix* results in

396 changes to lower lamina thickness, also resulting in blue colouration. They further show that  
397 natural variation in colour throughout the *Junonia* genus is controlled by lower lamina  
398 thickness, linking scale structure alterations and evolution.

399 •• **Brien et al., 2018**

400 The authors use phenotypic crosses of iridescent and non-iridescent races of *Heliconius erato*  
401 to determine the underlying genetic control of structural colour. They show that in *Heliconius*  
402 *erato* structural colour is a quantitative trait, controlled by a moderate number of loci. In  
403 addition, they show evidence for structural colour being sex-linked.

404 • **Parnell et al., 2018**

405 This paper characterises the ultrastructures underpinning structural colouration in *Heliconius*  
406 butterflies. Within the genus, ridge reflectors are responsible for producing structural colour  
407 with differences in ridge overlap, curvature and density controlling the brightness.

408 • **Fenner et al., 2019**

409 This study explores UV structural colouration in two closely related species of Dogface  
410 butterfly (Pieridae). Using changes to larval diet the authors explore the impacts on the  
411 ultrastructures responsible for structural colouration and find plasticity in structures  
412 governing UV colour production.

413 • **Zhang et al., 2018**

414 This study describes the architecture of fossil wing scales from the Jurassic period, which  
415 have an overall scale morphology similar to extant basal Lepidopterans. Through scale  
416 ultrastructure measurements and optical modelling, the authors determine that the scales  
417 would have produced a metallic colouration, similar to those produced by many extant basal  
418 Lepidopterans today.

419 •• **Matsuoka and Monteiro, 2018**

420 This article demonstrates a link between scale colour and morphology. By harnessing  
421 CRISPR-Cas9 the authors show that knockouts of melanin pathway genes in the butterfly  
422 *Bicyclus anyana* not only cause effects on pigmentary colour but also various effects on scale  
423 structure.

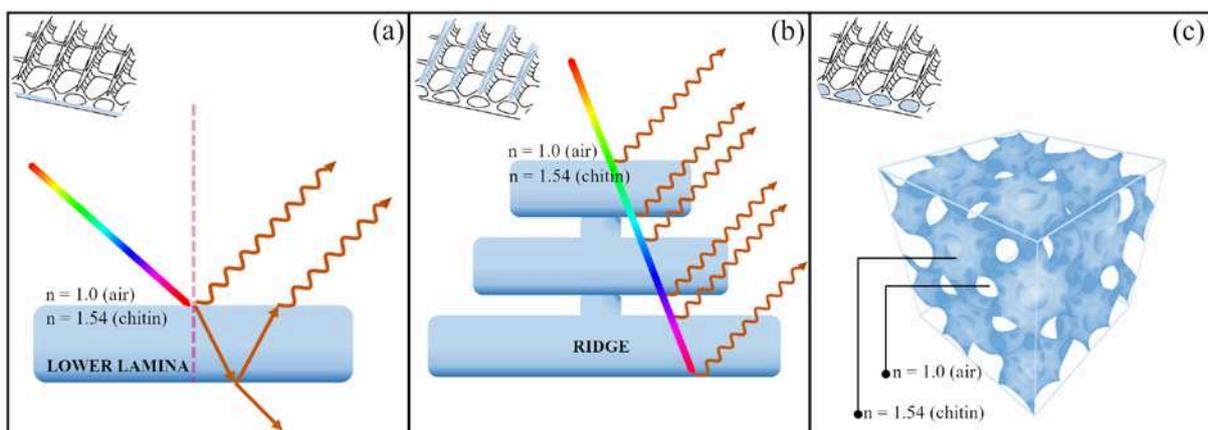
424 • **Seago et al., 2019**

425 This study analyses the diversity of photonic nanostructures in weevils to gain an  
426 understanding of the evolution of photonic crystals. The authors predict that structural colour  
427 evolved only once in weevils and required a transition from a hollow scale with a spongy  
428 network to a photonic crystal with gyroid symmetries.

429

430 **Additional elements**

431 **Box 1. Optical nanostructures in butterflies.**



432

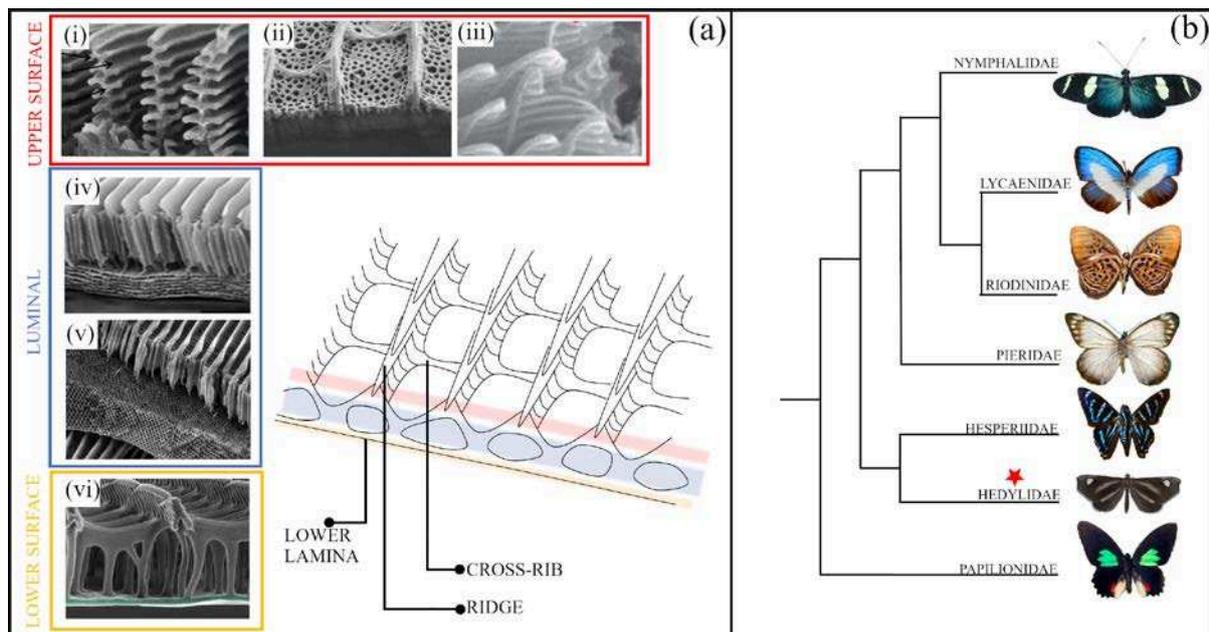
433 Diverse optical mechanisms in butterflies are governed by the same basic principle – to  
434 generate structural colour light must pass through materials of differing refractive indexes  
435 ( $n$ ). The cuticle of butterflies is composed of chitin, which has a refractive index of 1.54 (at

436 590 nm). To attain a contrasting refractive index, butterfly nanostructures also encompass air  
437 spaces which have a refractive index of 1.0 [3].

438 Thin films are the simplest optical mechanism present in butterfly scales. The lower  
439 lamina of the scale can form an optically precise thin-film reflector (a). Light waves are  
440 reflected by both the upper and lower surface of the lamina. When these reflected light waves  
441 are in phase, so that they peak at the same time, they can interfere constructively to  
442 accentuate particular wavelengths of light. The colour produced depends on the thickness of  
443 the chitin layer as well as the viewing and incidence angles [16••].

444 Thin film reflectors may also be stacked upon each other to form a multilayer-type  
445 nanostructure [3]. Examples of multilayers in butterfly scales may be found on both the upper  
446 surface as well in the lumen. The same principle of differences in refractive indexes applies,  
447 with the alternating air and chitin layers forming the contrasting indexes necessary to  
448 generate the phase change required for light interference. In the example of the ridge reflector  
449 multilayer (b) light is reflected and transmitted by each layer, producing a colour through  
450 constructive interference. Changing the number of layers governs the intensity of reflection  
451 while modifying the spacing of the air and chitin layers changes the colour produced [1].

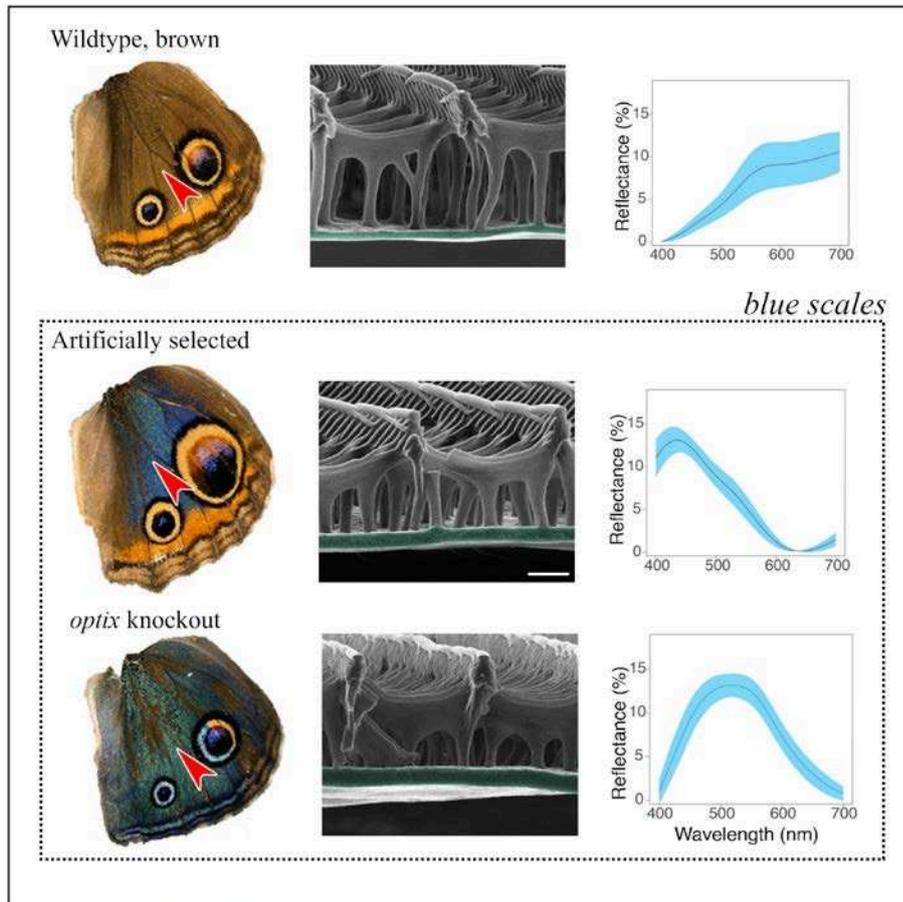
452 3D photonic crystals in butterflies are highly-ordered, repeating structures on the order of  
453 the wavelength of light (c). As the light waves enter the structure, only certain wavelengths of  
454 light may propagate, producing a specific colour [3]. This reflectance of a certain wavelength  
455 occurs regardless of the angle of light entering. Such structures are found in the lumen of  
456 butterfly scales, as highly periodic 3D crystals of chitin. One particularly interesting type is  
457 the ‘gyroid’ structure, having a nanoscale bicontinuous structure surrounding air spaces,  
458 which can be described by a mathematical concept in which a curved structure is maximally  
459 connected through the smallest surface area possible [1].



461

462 **Figure 1. The diversity of structural colour in butterflies.** (a) Morphologically diverse  
 463 optical nanostructures have evolved through modifications to a basic scale ‘ground plan’.  
 464 Modifications can be grouped depending on their location within a scale (upper surface,  
 465 lumen, lower surface). Several features of the upper surface may be modified, including the  
 466 ridges (i), inter-ridge cuticle (ii), and microribs (iii). The hollow lumen may also contain  
 467 nanostructures, including multilayers (iv) and highly-ordered photonic crystals (v). Changes  
 468 in cuticle thickness of the lower lamina can produce lower lamina reflectors (vi). (b)  
 469 Examples of structural colour in the major families of Papilionoidea (butterflies). Structural  
 470 colour remains undescribed for the Hedyliidae (star), but are presumed present due to the  
 471 existence of white scales in some species. Phylogeny drawn from [53]. SEM images in (a)  
 472 reprinted from: (i) Potyrailo et al., [54] (iv, v) Wilts et al., [21], (vi) Thayer et al., [16••]. All  
 473 made available under a CC-BY 4.0 license (<https://creativecommons.org/licenses/by/4.0/>).  
 474 Image (ii) reprinted with permission from Trzeciak et al., [55] © The Optical Society. Image  
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 476 permission conveyed through Copyright Clearance Center, Inc. Images of Lycaenidae

477 ([https://commons.wikimedia.org/wiki/File:Lycaenidae - Danis species.JPG](https://commons.wikimedia.org/wiki/File:Lycaenidae_-_Danis_species.JPG)) , Riodininae  
478 ([https://commons.wikimedia.org/wiki/File:Riodinidae - Paralaxita tesia-001.JPG](https://commons.wikimedia.org/wiki/File:Riodinidae_-_Paralaxita_tesia-001.JPG)) ,  
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480 Hesperiiidae ([https://commons.wikimedia.org/wiki/File:Hesperidae -](https://commons.wikimedia.org/wiki/File:Hesperidae_-_)  
481 [Jemadia menechmus.JPG](https://commons.wikimedia.org/wiki/File:Hesperidae_-_Jemadia_menechmus.JPG)) and Papilionidae  
482 ([https://commons.wikimedia.org/wiki/File:Papilionidae - Parides sesostris zestos.JPG](https://commons.wikimedia.org/wiki/File:Papilionidae_-_Parides_sesostris_zestos.JPG))  
483 obtained from Wikimedia, where they were made available by Hectonichus under a CC-BY-  
484 SA-3.0 (<https://creativecommons.org/licenses/by-sa/3.0/legalcode>) license. Image of  
485 Hedyliidae obtained from Wikimedia  
486 ([https://commons.wikimedia.org/wiki/File:Hedylid\\_Moth\\_\(Macrosoma\\_lucivittata\)\\_254659](https://commons.wikimedia.org/wiki/File:Hedylid_Moth_(Macrosoma_lucivittata)_254659)  
487 [99377\).jpg](https://commons.wikimedia.org/wiki/File:Hedylid_Moth_(Macrosoma_lucivittata)_254659_99377.jpg)) , where it was made available by Bernard DUPONT under a CC-BY-SA-2.0  
488 license (<https://creativecommons.org/licenses/by-sa/2.0/legalcode>).  
489



490

491 **Figure 2. Insights into the genetic and structural control of lower lamina structural**  
 492 **colour in *Junonia coenia* through artificial selection experiments and *optix* knockouts.**

493 Wildtype *J. coenia* hindwings are predominantly brown in colour. A blue colouration is  
 494 observed in wings which have been artificially selected for blue scales and in CRISPR-Cas9  
 495 knockouts of the developmental patterning gene *optix*. Changes in the observed colour of  
 496 artificially selected and *optix* knockout wings can be seen in reflectance plots, with a shift to  
 497 the blue end of the spectrum compared to wildtype brown scales. Helium ion microscopy  
 498 (HIM) cross-sections showing scale morphology, with the lower lamina false coloured in  
 499 green. The lower lamina is considerably thicker in *optix* knockout wings and artificially  
 500 selected wings compared to wildtype wings. Images reproduced from Thayer *et al.*, [16••]

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 502 <https://creativecommons.org/licenses/by/4.0/> ).