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eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/ 1 Bone Metastasis: mechanisms, therapies and biomarkers.

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84 Abstract:

85 Skeletal metastases are frequent complications of many cancers, causing bone complications 86 (fractures, bone pain, disability), which negatively affect the patient's guality of life. Here, we first 87 discuss the burden of skeletal complications in cancer bone metastasis. We then describe the 88 pathophysiology of bone metastasis. Bone metastasis is a multistage process; long before the 89 development of clinically detectable metastases, circulating tumor cells settle and enter a dormant state 90 in normal vascular and endosteal niches present in the bone marrow, which provide immediate 91 attachment and shelter, and only become active years later as they proliferate and alter the functions of 92 bone-resorbing (osteoclasts) and bone-forming (osteoblasts) cells, promoting skeletal destruction. The 93 molecular mechanisms involved in mediating each of these steps are described and we also explain 94 how tumor cells interact with a myriad of interconnected cell populations in the bone marrow, including a 95 rich vascular network, immune cells, adipocytes and nerves. We discuss metabolic programs that tumor 96 cells could engage with to specifically grow in bone. We also describe the progress and future directions 97 of existing bone-targeted agents and report emerging therapies that have arisen from recent advances 98 in our understanding of the pathophysiology of bone metastases. Finally, we discuss the value of bone 99 turnover biomarkers in detection and monitoring of progression and therapeutic effects in patients with 100 bone metastasis.

102 I. INTRODUCTION

103 During metastatic dissemination, cancer cells from the primary tumor must first undergo epithelial-104 to-mesenchymal transition (EMT) to invade the surrounding tissue and enter the microvasculature 105 (intravasation) of the blood and/or lymphatic systems (49, 268). Once in the bloodstream, cancer cells 106 may disseminate to distant organs, exit from blood vessels (extravasation) and settle in the foreign 107 microenvironment where they enter a dormant state or proliferate to subsequently form macroscopic 108 secondary tumors (metastases) (49). It has been estimated that only 0.02% of cancer cells entering the 109 circulation produce clinically detectable metastases (217). Metastasis formation is therefore a highly 110 inefficient process. However, when metastases do occur, they are responsible for 90% of cancer-111 associated mortality (49). There is therefore an urgent need to increase our understanding of the 112 cellular and molecular mechanisms associated with metastasis formation, in order to develop therapies 113 that will improve patient outcome.

114 Bone metastases occur in more than 1.5 million patients with cancer worldwide (361). They are 115 frequent complications of many cancers but are especially common from tumors arising in the breast 116 and prostate. Weakened bones due to skeletal metastases can lead to occurrence of skeletal-related 117 events, such as fractures, spinal cord compression, bone pain and disability, contributing substantially 118 to both morbidity and mortality in patients with advanced cancer (150, 361). In adults, the bone mass is 119 maintained by continuously shaping and reshaping the overall bone structure through a process called 120 bone remodeling, which is a balance between the resorption of mineralized bone by bone-resorbing 121 cells (osteoclasts) and formation of new bone by bone-forming cells (osteoblasts) (76). Bone remodeling 122 is tightly regulated by systemic and local factors in order to maintain this balance at its physiological 123 steady state (76, 150). The late Greg Mundy pioneered the field of cancer and bone, demonstrating that 124 skeletal-related complications associated with bone metastasis were a consequence of a distortion in 125 bone remodeling caused by interactions between cancer cells and cells within the bone 126 microenvironment (236).

127 In this review, we provide a broad overview of the current understanding of cancer-associated 128 bone metastasis. We first review the incidence of bone metastasis in different cancer types and discuss 129 the burden of skeletal complications in cancer bone metastasis. Current knowledge of the 130 pathophysiology of bone metastasis is then described in detail. Bone metastasis is a stepwise sequence 131 of events that include tumor cell colonization of the bone marrow, adaptation to the microenvironment, 132 construction of a cancer niche, disruption of normal bone homeostasis through tumor cell interactions 133 with bone cells (osteoclasts, osteoblasts and osteocytes, the latter being osteoblasts that have 134 undergone a dramatic morphological transformation into stellate cells) and the release of signals from 135 the resorbed bone matrix that promote skeletal tumor growth. We describe molecular mechanisms that 136 are involved in mediating each of these steps and explain how bone marrow cells (e.g. immune cells, 137 endothelial cells, adipocytes, and nerve cells) contribute to tumor development through multiple 138 interactions. We also highlight metabolic adaptations of cancer cells that facilitate tumor progression in 139 bone. Finally, we review current and future therapies for the treatment and prevention of bone 140 metastasis and discuss the clinical utility of bone turnover biomarkers to predict the risk of disease 141 relapse in patients with cancer. Given the vast collection of literature existing on the pathophysiology of 142 bone metastasis we focus here on cellular and molecular mechanisms that are the most relevant to 143 human cancer. However, it is important to note that we also cover emerging research areas where 144 many mechanisms are derived from model systems, which still remain to be validated in human 145 systems but could ultimately yield clues for better understanding and prevention of bone metastases.

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147 II. BONE METASTASIS INCIDENCE AND CONSEQUENCES FOR BONE HEALTH

148

A. Common Ground – Bone Metastasis in Different Cancer Types

Bone is one of the most common sites for metastasis in cancer. Much of the work performed to describe the natural history of bone metastases is based on autopsy studies and large case series 151 from single institutions conducted several decades ago. Although bone is a frequent location for 152 metastases from many malignancies, there are specific types of cancers that have a predilection for 153 metastasis to the skeleton (150, 361). In particular, bone metastases are frequent complications of 154 breast (especially estrogen receptor positive) and prostate cancer. In their retrospective study, Coleman 155 and Rubens found in breast cancer a bone metastasis incidence of around 70% (68). These findings 156 were consistent with the post-mortem examination from Galasko (115), who reported bone metastasis 157 incidences of 73% and 68% of bone metastasis in breast and prostate cancer, respectively. Autopsies 158 allowed the identification of a second group of osteophilic tumors with a postmortem prevalence of bone 159 metastases of 60% in thyroid cancer, 30-40% in lung cancer, 40% in bladder cancer, 20-25% in renal 160 cancer and 14-45% in melanoma (65). Apart from osteoblastic bone metastases in prostate cancer, 161 bone metastases from other cancers are mainly osteolytic or a mix of lytic and blastic changes to the 162 bone structure (Figure 1).

163 With the exception of a few relatively rare malignancies such as high-grade lymphoma or 164 germ cell tumors affecting bone, metastatic bone disease is currently incurable. However, for many 165 patients the median prognosis after development of bone metastasis is measurable in years, especially 166 in those patients with metastatic breast or prostate cancers or multiple myeloma who, with modern 167 treatment approaches, can often be expected to survive more than 5 years after bone involvement is 168 diagnosed (65). Furthermore, new drugs, such as tyrosine kinase inhibitors and immune checkpoint 169 inhibitors, have prolonged primary disease control in patients considerably, resulting in longer survival 170 and consequently living long enough for bone metastasis to become clinically relevant (338). Thus, the 171 epidemiology of bone metastases is evolving. In the coming years we may therefore expect an onset of 172 bone metastases in patients who would have never developed clinically detectable bone metastases 173 some years ago because they would have died from their cancer at a time when they only had (sub-174 clinical) bone marrow micrometastases. As a result, the prevalence of bone metastasis is increasing

and, in many cancers, the dominant site of disease requires specialist expertise and multidisciplinarymanagement (72).

177

178

B. Cancer-Related Skeletal Complications

Bone metastases may be identified when asymptomatic through imaging tests such are computerized tomography (CT), ¹⁸F-fluorodeoxyglucose (FDG)-positron emission tomography (PET) scanning, or radionuclide bone scanning. However, most patients with bone metastasis present with bone pain (150, 236). Usually the onset of pain is insidious, may be localized or multifocal, and is often confused with benign causes such as osteoarthritis. With time the pain typically worsens and becomes persistent, frequently reaching a severe level that may not be relieved by opioids (150, 236).

185 Bone metastasis is associated with impaired quality of life, reduced physical function and 186 loss of autonomy (68, 150). Because of the proximity between bone and neurological structures (spinal 187 cord and nerve roots), bone metastasis often causes neurological pain, such as paresthesia and tingling 188 or burning sensations induced by epiduritis (68). Fractures are major complications of bone metastases 189 and are commonly a result of osteolytic lesions in the vertebrae and weight-bearing bones, such as the 190 proximal femur (68). The humerus is also at risk because of the forces applied through the arm use in 191 daily life. Once pathological fractures have occurred, bone healing is compromised, and surgical 192 intervention often required. Pathological fractures can be devastating complication for cancer patients. 193 typically worsening their quality of life and increasing mortality (68, 150).

Hypercalcemia is an important metabolic complication of bone metastases (337). Symptoms include a wide spectrum of presentations from subtle changes in mood and gastrointestinal symptoms of nausea and constipation to a life-threatening state with vomiting and dehydration, acute renal insufficiency, disordered consciousness and ultimately coma (337). In bone metastases, hypercalcemia usually results from increased osteoclastic bone resorption but may be exacerbated by the paraneoplastic secretion of parathyroid hormone-related peptide (PTHrP) or an abnormal activation of
200 25-OH vitamin D (345). The use of systemic anti-resorptive drugs has considerably reduced the number
of patients with hypercalcemia (61, 345).

202 Most clinical studies use the composite endpoint Skeletal-Related Events (SREs) to establish 203 the efficacy of systemic anti-resorptive drugs (61, 345). SREs are defined as pathologic fractures, spinal 204 cord compression and the requirement for radiation therapy and/or surgery to bone; episodes of 205 hypercalcemia may also be considered within the definition (220). Early placebo-controlled 206 bisphosphonate clinical trials estimated that 50 to 56% of patients with bone metastases from solid 207 tumors suffer from at least one SRE during follow-up on standard anti-cancer treatments without the 208 addition of a bone targeted treatment (157, 279). SREs can occur guite early and indeed can be the 209 presenting event in a patient with bone metastasis. In these trials, the median time to occurrence of the 210 first SRE ranged from 5 to 7 months (157, 279). Moreover, patients with a first SRE are at increased risk 211 for subsequent events, strengthening the importance of primary and secondary SRE prevention in 212 cancer patients with bone metastases (68, 72). In addition to reducing a patient's guality of life and 213 social and functional independence, the management of SREs consumes considerable health care 214 resources (68, 72).

215 Besides analgesics and anticancer treatments, bone metastases benefit from systemic anti-216 resorptive treatments (bisphosphonate or denosumab) and local treatments such as radiotherapy, 217 surgery or interventional radiology (cementoplasty, radiofrequency, ablation, cryotherapy). Optimal care 218 should be discussed in a Bone Metastasis Multidisciplinary Board in order to reach a personalized 219 strategy for every patient (72). Bone-targeted agents such as bisphosphonates and denosumab have 220 been shown to be very effective in preventing and reducing SREs and are now the standard of care for 221 the treatment of patients with malignant bone disease (see sections IX-A.1 and A.2 for further 222 discussion).

224 III. TAKING OVER THE NEIGHBORHOOD - BONE COLONIZATION BY TUMOR

225 **CELLS**

Bone colonization by tumor cells is a stepwise sequence of events that include *(i)* the formation of a pre-metastatic niche in the bone marrow to attract circulating tumor cells, *(ii)* the extravasation of these tumor cells from the circulation and homing to the pre-metastatic niche, and *(iii)*, following tumor cell engraftment, the evolution of this pre-metastatic niche into a metastatic niche, the latter being conducive to the survival of these tumor cells. Each of these events is discussed below (**Figure 2**).

A. Preparing the Soil – the Concept of the Premetastatic Niche

232 The concept of premetastatic niche was first described by Dr Lyden and colleagues showing that 233 vascular endothelial growth factor (VEGF)-A and placental growth factor (PIGF) secreted from primary 234 tumors mobilize bone marrow-derived VEGF receptor 1 (VEGFR-1)-positive hematopoietic cells to the 235 lungs before the arrival of tumor cells (169). Furthermore, an upregulation of fibronectin in resident 236 fibroblasts at these premetastatic sites subsequently supports adhesion of VEGFR-1-positive cells 237 (169). This localized accumulation of bone marrow-derived hematopoietic cells and stromal fibronectin 238 creates docking sites for the future engraftment of tumor cells in lungs (169). Since then, many other 239 tumor-derived factors, including cytokines, chemokines, extracellular matrix components, small 240 noncoding RNAs and tumor-shed extracellular vesicles have been shown to act as systemic signals that 241 trigger the formation of premetastastic niches in lung, liver or lymph nodes in different preclinical models 242 (149, 261). Clinical evidence for the existence of premetastatic tissues comes from patients with 243 meningioma (a benign brain tumor) who later progress with tumor-to-meningioma metastasis of breast, 244 lung or renal cancer (88, 264, 270). It is suggested that the presence of pro-inflammatory macrophages 245 and the high microvascular density in meningioma contribute to metastasis formation (88). Similarly, the 246 existence of a premetastatic tissue in sentinel lymph nodes resected from patients with solid tumors has 247 been reported (302). Thus, there is preclinical and clinical evidence that primary tumors may remotely 248 induce the formation of a permissive environment within distant organs for future metastasis.

249 Multiple molecular mechanisms involved in the formation of a premetastatic niche in bone have 250 been described (98, 137, 151, 216, 247, 248, 250, 251, 261, 271, 327, 336, 341, 371). Some of them 251 already exist in the normal bone marrow (216, 248, 250, 251), whereas others are initiated by systemic 252 signals coming from primary tumors (98, 137, 151, 327, 336, 341, 371). For example, interleukin (IL)-6 253 secreted from senescent osteoblasts promotes osteoclast-mediated bone resorption that, in turn, 254 increases tumor cell seeding and subsequent breast cancer bone metastasis formation in animals 255 (216). Similarly, in the absence of estrogen or androgen, osteoclast activity and bone resorption are 256 increased, which leads to the release of bone-derived factors from resorbed bone that shape a 257 favorable environment for tumor cells to survive and grow (248-250). Additionally, soluble factors 258 secreted from primary tumors can target stromal and/or bone cells to support future metastatic 259 colonization in the bone marrow. For example, in breast cancer models, tumor-derived IL-1B drives 260 bone metastasis formation in vivo (151, 336). Blocking IL-1 β activity with the anti-IL-1 receptor 261 antagonist Anakinra or the IL-1ß specific antibody Canakinumab inhibits tumor cell dissemination from 262 the primary site into the circulation and blocks spontaneous formation of metastases to human bone 263 implants in treated mice, compared to the placebo-treated group (336). Hypoxia-induced lysyl oxidase 264 (LOX) can be secreted from primary tumors into the circulation from which LOX primes distant organs 265 for metastatic colonization, including bone (73, 98, 263, 274). The primary function of LOX is to drive 266 collagen crosslinking and extracellular matrix stiffness (7). In bone, tumor-derived LOX cooperates with 267 receptor activator of nuclear factor kappa-B (RANK) ligand (RANKL) to accelerate osteoclastic bone 268 resorption, and the formation of premetastatic osteolytic lesions in animal models of breast or colon 269 cancer (73, 274, 335). A function-blocking antibody (AB0023) directed against LOXL2 (another member 270 of the LOX family) inhibits breast cancer bone metastasis formation in animals (10), suggesting that 271 LOXL2 could also contribute to the formation of a pre-metastatic niche in the bone marrow. Phase II 272 clinical trials with simtuzumab, a humanized anti-LOXL2 monoclonal antibody, showed however that the 273 addition of this antibody to current treatments of patients with metastatic pancreatic or colorectal cancer

does not improve clinical outcomes (ClinicalTrials.gov identifiers NCT01472198 and NCT01479465, respectively). Conversely, the anti-IL-1 β antibody canakinumab has been shown to significantly reduce the incidence of lung cancer and lung cancer mortality in patients with atherosclerosis (ClinicalTrials.gov identifier NCT01327846). It remains to be established whether blocking IL-1 β , LOX or LOXL2 impedes progression of bone metastasis in breast cancer patients.

279 It also appears that factors contained within cancer cell-derived exosomes can influence bone cell 280 activity before tumor cells arrive at this site. Exosomes are small extracellular vesicles (30-120 nm) 281 containing DNA, RNA [mRNA, miRNA and other noncoding RNAs], lipids and proteins that are released 282 by all types of cells and taken up by recipient cells (368). For example, exosomal amphiregulin secreted 283 by non-small cell lung carcinoma (NSCLC) cells or amphiregulin-containing exosomes released in 284 plasma of NSCLC patients promote the differentiation of human peripheral blood monocytes into 285 osteoclasts (327). In melanoma, the transfer of the MET oncoprotein from tumor-shed exosomes to 286 bone marrow progenitor cells can reprogram these cells towards a prometastatic phenotype in lungs 287 and bone in vivo (261). Similar findings were reported with tumor-derived exosomal miRNAs (miR-21, 288 miR-141, miR-192, and miR-940) (22, 137, 341, 371, 376). In particular, exosomal miR-141 and miR-289 940 produced by prostate cancer cells promote osteoblast differentiation and proliferation, facilitating 290 the formation of bone metastases with an osteoblastic phenotype in mouse models (137, 376). MiRNAs 291 mainly act as negative regulators of gene expression (14). In this respect, tumor-derived exosomal miR-292 141 promotes osteoblast differentiation by inhibiting DLC1 mRNA expression that, in turn, leads to 293 p38MAPKinase activation and increased osteoprotegerin (OPG) expression in osteoblasts (376). 294 Tumor-derived exosomal miR-940 promotes osteogenic differentiation of mesenchymal stem cells by 295 directly inhibiting ARHGAP1 (Rho GTPase Activating Protein 1) and FAM134A (Family with Sequence 296 Similarity 134 Member A) mRNA expression (137).

297 Overall, these experimental findings strongly suggest that, in addition to molecular mechanisms 298 already existing in the normal bone marrow, primary tumors can also remotely control the formation of a premetastatic niche through the release of systemic factors that induce a distortion in bone remodeling. Research designed to determine the mechanisms by which primary tumors promote the formation of pre-metastatic niches in bone is still in its infancy and further investigations using *in vivo* model systems are required to gain a more comprehensive understanding of this process. As tumor cell dissemination into bone is believed to be an early process, likely to occur before the clinical detection of primary tumors, the detection of these molecules in the primary tumor and/or blood may provide useful biomarkers to predict future relapse in bone. Further clinical trials are needed to test this hypothesis.

306

307 The premetastatic niche: current understandings & open questions

- Preclinical and clinical studies support the existence of premetastatic tissues for future
 metastasis.
- The general applicability of these mechanisms associated with the formation of a premetastatic 311 niche remains to be validated *in vivo* for other model systems and for other cancer types.
- Beside the observation that primary tumors can generate systemic changes that modify the
 bone microenvironment, there is also some preclinical evidence suggesting that bone may
 remotely control growth of primary tumors at distant sites (85, 97, 166, 257). These latter
 observations are intriguing and clearly deserve further study.

316

B. Mechanisms of Tumor Cell Extravasation and Homing to the Bone Marrow

In response to pro-migratory and pro-inflammatory molecules produced by the pre-metastatic niche, circulating tumor cells (CTCs) cross the endothelial cell barrier and basement membrane of blood vessels (a process called extravasation) in order to home in the newly invaded parenchyma where they interact with specific extracellular matrix components that facilitate their survival.

322 **1**. Tumor cell extravasation

323 In the bone marrow, the vascular endothelium that constitutes blood vessels (called sinusoids) is 324 predominantly discontinuous and fenestrated, which facilitates the traffic of hematopoietic stem cells 325 (HSCs) (241). Therefore, the sinusoids are likely to be more permissive to CTCs, suggesting there is a 326 limited requirement of extravasation mechanisms for tumor cells to invade the bone marrow (241, 273). 327 Indeed, tumor cells hijack molecular mechanisms that are used by HSCs. In particular, E-selectin 328 (Endothelial selectin) and CXCL-12 are constitutively expressed on sinusoidal endothelial cells, aiding 329 the homing of HSCs in the bone marrow (301, 316). Similarly, E-selectin- and CXCL-12-expressing 330 bone marrow endothelial cells mediate attachment of breast and prostate cancer cells through 331 interaction with E-selectin ligands and CXCR-4, respectively (267, 273). Using high-resolution real-time 332 fluorescence microscopy to track breast cancer cell migration in the calvarial bone marrow in vivo, Price 333 et al. (267) showed that 2 hours after intracardiac injection, tumor cells interacted with endothelial cells 334 in sinusoidal vascular and perisinusoidal vascular regions where expression of E-selectin and CXCL-12 335 is high. Of special interest, the preventive treatment of mice with a selective inhibitor of E-selectin, 336 before tumor cell injection, substantially blocked tumor cell interaction with E-selectin-expressing 337 endothelial cells, whereas pretreatment of animals with a small molecule inhibitor of CXCR-4 338 (AMD3100) did not inhibit breast cancer cell homing to the bone marrow in vivo (267). By contrast, 339 AMD3100 treatment of mice after tumor cell engraftment forced breast cancer cells residing in 340 perivascular niches to migrate from the bone marrow into the peripheral circulation. Overall, these 341 findings demonstrate that E-selectin is critical for allowing breast cancer cells to extravasate in the bone 342 marrow, whereas CXCR-4/CXCL-12 maintains tumor cells in the perivascular environment and controls 343 their exit from the bone marrow (267). The CXCR-4/CXCL-12 axis is the most well-described and 344 prominent mechanism involved in regulating tumor cell entry in the bone marrow environment (235). 345 However, it should be noted that, not all breast cancers that metastasize to bone express CXCR4 (243), 346 and other chemokines produced by the bone microenvironment (CXCL-5, CXCL-10, CXCL-13, CX3CL-347 1, CCL-2) have also been implicated in mediating tumor cell colonization in the bone marrow (158, 159,

193, 213, 232, 275, 300) (Table 1). However, correlations between expression of these chemokines
and relapse in bone, in clinical samples, remains to be established.

350 Another factor that has been implicated in tumor cell extravasation is the cytokine IL-1B whose 351 expression in breast cancer cell lines and primary breast carcinomas is strongly associated with bone 352 metastasis (151, 336). IL-1ß drives metastasis by inducing epithelial-to-mesenchymal transition and 353 increasing dissemination of breast cancer cells into the circulation (151, 336). Once in bone, IL-1B 354 facilitates adhesion of CTCs to sinusoidal endothelial cells by inducing the expression of vascular cell 355 adhesion molecules [intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 356 (VCAM-1), and E-selectin] (273). Then, IL-1 β stimulates expansion of the metastatic niche, increasing 357 proliferation of blood vessels and osteoblasts, thereby promoting tumor cell extravasation and 358 metastatic outgrowth of tumor cells that disseminated in this site (273, 336). Interestingly, IL-1β-359 expressing E0771 primary breast tumors spontaneously metastasize to bones in IL-1β-knockout 360 animals and to an extent similar to that observed in normal mice, indicating that tumor-derived IL-1ß 361 rather than IL-1 β from the bone marrow microenvironment promotes bone metastasis formation (336). 362 Although these mechanisms have all been identified in mouse models it is likely that these, at least in 363 part, explain why increased IL-1 β in primary breast tumors associates with recurrence in bone in cancer 364 patients (151, 336).

365 2. Tumor cell homing

Bone extracellular matrix proteins (*e.g.*, type I collagen, tenascin C, periostin, fibronectin, SIBLINGs) by binding to tumor cell surface integrins play an important role in mediating tumor cell attachment to the bone matrix (17, 57, 221, 255, 260, 285, 311, 317, 333). In particular, type I collagen, which is the most abundant protein among bone extracellular matrix components, mediates attachment of human prostate cancer cells through binding tumor cell $\alpha 2\beta 1$ integrin (311). Fibronectin in the bone marrow mediates survival of human triple negative breast cancer cells by binding to tumor cell surface 372 α5β1 integrin (255). Tenascin C is a hexameric protein that fosters the early colonization of prostate 373 cancer cells in the bone marrow through binding tumor cell surface $\alpha 9\beta 1$ integrin (285). Like tenascin 374 C, periostin is produced by stromal cells and mediates tumor cell adhesion by binding tumor $\alpha\nu\beta\beta$ 375 integrin (221). As such, integrins have been therefore considered as attractive drug targets. For 376 example, $\alpha v\beta 3$ integrin recognizes an Arg-Gly-Asp (RGD) peptide motif expressed by extracellular 377 matrix proteins and the treatment of animals with RGD-based peptide antagonists (PSK104, S247, 378 GLPG0187) of $\alpha\nu\beta3$ integrin suppresses breast and prostate cancer bone metastasis formation, 379 supporting the crucial role played by this integrin in bone metastasis formation (136, 303, 344, 388). 380 However, despite these encouraging experimental studies, ensuing clinical trials that used integrin 381 antagonists have been mainly unsuccessful (132). There may be a need to develop alternative 382 strategies that target specific integrin signaling pathways promoting tumor cell survival and drug 383 resistance (132). Integrins are also emerging as valuable cancer imaging probes probes to identify bone 384 metastases in clinical studies. For example, integrin $\alpha v \beta 3$ is highly expressed in osteotropic tumor cells 385 and osteoclasts and, using PET/CT imaging, this property has been used to show that an RGD peptide-386 containing αvβ3 integrin tracer (99mTc-3P-RGD₂) is superior to 99mTc-bisphosphonates to detect 387 osteolytic bone metastases in patients with advanced lung cancer (297). Similarly, PET/CT imaging with 388 a tracer targeting gastrin-releasing peptide receptor and integrin αvβ3 (68Ga-BBN-RGD) shows a 389 significant uptake in bone metastases from patients with advanced breast cancer (384).

RANK and RANKL have an established role in regulating bone remodeling (183). RANKL secreted by osteoblasts and osteocytes binds to its receptor RANK on osteoclast precursors, leading to the formation of mature osteoclasts and osteoclast-mediated bone resorption (183). Interestingly, tumor cells can also express RANK in breast, prostate, and lung carcinomas and RANKL triggers *in vitro* migration of RANK-expressing breast and prostate cancer and melanoma cells (21, 164, 325, 346). In the case of breast cancer, a high RANK expression in hormone receptor-negative primary tumors is associated with poor relapse-free survival and high risk of bone metastasis in patients (272, 286, 347). It 397 has been therefore suggested that RANK-expressing cancer cells may be specifically attracted to bone 398 where high local concentrations of RANKL exist (346). This contention was supported by the 399 observation that inhibition of RANK/RANKL signaling by soluble decoy receptor OPG, which binds to 400 RANKL, reduces skeletal tumor burden and bone metastasis in a melanoma model that does not 401 activate osteoclasts, whereas metastasis in other organs (ovaries, brain, adrenal glands) remain 402 unchanged (164). However, administration of OPG to a mouse model of breast cancer did not reduce 403 the number of tumor cells that disseminated in bone (249). In clinical studies, the RANKL inhibitor 404 denosumab, when given in patients with early-stage breast cancer or non-metastatic castration-resistant 405 prostate cancer (CRPC), had no effect on disease recurrence in either pre- or postmenopausal women 406 with breast cancer (63) and only modestly increased bone metastasis-free survival in CRPC patients 407 (306), thereby suggesting that RANK/RANKL does not play a major role in tumor cell colonization in 408 bone.

409 <u>Tumor cell extravasation and homing: current understandings & open questions</u>

Recent studies on the bone microvasculature in mouse models have shown that there are particular vessel subtypes within the same vascular bed, termed H (CD31^{hi}Endomucin^{hi}) and L (CD31^{lo}Endomucin^{lo}) vessels, which have different characteristics (182). It will be of particular interest to determine whether disseminated tumor cells preferentially associate with a particular vessel subtype.

Results obtained with PET/CT imaging using integrin-binding tracers to detect skeletal lesions
 and evaluate treatment response in patients with advanced cancer and bone metastases are
 very encouraging and deserve further investigations.

418

419 C. Hiding in Plain Sight –Tumor Cell Dormancy and Dormant Cell Reactivation 420 Mechanisms in Bone Marrow Niches

421 Entering a foreign environment, such as the bone marrow, poses tumor cells with numerous 422 challenges regarding survival and proliferation. It is hypothesized that tumor cells in the bone marrow 423 compete with HSCs for occupancy in the vascular and endosteal niches. Once in the niche, tumor cells 424 can enter a dormant state as a mechanism to help them survive until environmental conditions are 425 sufficiently permissive for proliferation and tumor outgrowth (298, 309). This hypothesis of tumor 426 dormancy is supported by an extensive body of clinical research. First, long before the development of 427 clinically detectable metastases, tumor cells disseminate to the bone marrow, only becoming active 428 following years or decades after primary tumor diagnosis (247). Second, disseminated tumor cells 429 (DTCs) are present in the bone marrow of patients with various types of cancer and are predictive of 430 future relapse, yet some of these cancer types will never develop clinically detectable bone metastases 431 (205). Lastly, for those cancer types that have a high propensity to develop overt bone metastases, 432 such as breast and prostate cancers, the rate of detection of DTCs in the bone marrow is higher than 433 the proportion of patients who subsequently develop skeletal lesions, suggesting the bone marrow 434 microenvironment influences DTC fate (32, 228). Thus, these clinical observations provide strong 435 evidence of tumor dormancy in the bone marrow. However, DTCs isolated from the bone marrow of 436 non-metastatic patients with cancer (breast, prostate, melanoma) fail to generate tumor xenografts in 437 immunodeficient mice (364). Consequently, only tumor cell lines derived from overt metastases from 438 patients with advanced cancer (breast, prostate) have been used in animal models to study tumor 439 dormancy in bone (31, 46, 117, 119, 163, 177, 267, 320, 381, 382). The use of these metastatic cell 440 lines, therefore, poses a major limitation as metastatic cancer cells and DTCs have different genotypic 441 and phenotypic traits (309). Determining the different mechanisms controlling the ability of tumor cells to 442 seed in the bone marrow and those responsible for metastatic outgrowth may require the development 443 of more clinically relevant models. Yet, tumor dormancy is an emerging research area and we believe 444 that unravelling molecular mechanisms associated with tumor dormancy using these existing preclinical 445 models may still help us better understand the earliest stages that precede the clinical development of bone metastases. Below we describe our current understanding of the mechanisms involved in mediating tumor cell interactions with cells from bone marrow niches (Figure 2) and then detail molecular signaling mechanisms proposed to keep these tumor cells in a dormant state (Figure 3). Finally, we describe how osteoclast-mediated bone resorption creates an environment that promotes dormant cell reactivation.

451 1. Tumor cells interactions with cells from bone marrow niches

452 The vascular niche surrounds E-selectin-expressing endothelial cells that form bone marrow 453 sinusoids, and is made up of perivascular cells expressing high levels of CXCL-12 [called CXCL-12-454 abundant reticular (CAR) cells], leptin receptor (Lepr)-expressing perivascular stromal cells, and 455 mesenchymal stem cells (MSCs) (301, 316). This vascular niche regulates HSC quiescence and the 456 supply of lineage-committed progenitors (301). Real-time in vivo microscopy of bone marrow sinusoids 457 in a breast cancer xenograft model has revealed opposing roles of E-selectin and CXCL-12 in tumor cell 458 trafficking (267). Whereas E-selectin interactions are critical for allowing breast cancer cell entry into the 459 bone marrow, CXCL-12/CXCR-4 interactions maintain breast cancer cells dormant in the vascular niche 460 (267). Additional mechanisms are involved in maintaining tumor cells dormant in the vascular niche. For 461 example, endothelium-derived extracellular matrix protein thrombospondin-1 (TSP-1) induces sustained 462 dormancy of breast cancer cells in vivo (119). Conversely, MSCs with both endothelial and pericytic cell 463 surface markers prevent the homing of breast and prostate cancer cells to the bone marrow (282). In 464 model systems, the tumor-suppressive nature of the vascular endothelium is lost when endothelial cells 465 start sprouting, which is characterized by reduced TSP-1 expression and enhanced expression of pro-466 metastatic factors (periostin, tenascin, fibronectin) that promote tumor outgrowth (119). Interestingly, 467 immunohistochemical analysis of the bone marrow from breast cancer patients with micrometastatic 468 disease shows that dormant (Ki67-negative) breast cancer cells are preferentially localized in 469 perisinusoidal, CXCL12-rich vascular regions (267). By contrast, proliferative (Ki67-positive) breast 470 cancer cells in bone marrow biopsies from patients with macrometastatic disease are frequently

471 observed adjacent to the bone surface (267). This observation (267) is in agreement with the fact that 472 calcium levels, which are high at the endosteal mineral surface, can promote breast cancer cell 473 proliferation and bone metastasis formation in animals (354). Thus, it appears that the vascular niche 474 provides a microenvironment supportive of dormancy at least in breast cancer.

475 As the name suggests, the endosteal niche is localized at the inner surface of the bone cavity in 476 the endosteum, and is primarily made up of undifferentiated osteoblastic cells, such as spindle-shaped 477 N-cadherin⁺/CD45⁻ osteoblast (SNO) cells (138). Mature osteoblasts are short-lived cells and, as such, 478 they are unlikely to be part of the endosteal niche (76, 256). Beside SNO cells, CAR cells are also 479 present and are proposed to maintain the guiescent HSC pool through CXCL-12/CXCR-4 interactions 480 (316). The disruption of this connection using CXCR4 antagonists, in mouse models, results in 481 increased mobilization of HSCs from the bone marrow into the circulation (316). Osteoclasts are 482 dispensable for HSC maintenance in the endosteal niche and may function as negative regulators in the 483 hematopoietic system (231). With regard to the homing of tumor cells to the endosteal niche, ER-484 negative (but not ER-positive) breast cancer cells compete with HSC to interact with SNO cells through 485 a specific Jagged-Notch2 interaction that mediates tumor cell dormancy both in vitro and in vivo (46). It 486 must be pointed out however that these particular in vivo experiments were conducted using intratibial 487 tumor cell inoculation, thereby bypassing the blood circulation, which impedes breast cancer cells from 488 homing to the vascular niche. Other studies, using more clinically relevant mouse models in which 489 tumor cells were disseminated into the bone via intra-arterial injection, reported that SNO cells support 490 survival of ER-positive breast cancer cells through specific N-cadherin/E-cadherin interactions and 491 connexin-43 (Cx43) gap junctions that trigger pro-survival mTOR signaling and calcium signaling 492 pathways in tumor cells, respectively, hence promoting micrometastatic progression (354, 355). In 493 addition, independently of the hormone receptor status or breast cancer subtype, CXCL-12 triggers 494 activation of a Src-dependent AKT signaling pathway by binding to CXCR-4, enhancing the survival of 495 breast cancer cells in the bone marrow (387). Of note, Werner-Klein et al. (364) performed single-cell 496 RNA-sequencing analysis of DTCs isolated from the bone marrow of non-metastatic breast cancer 497 patients (n=30 DTCs; 21 patients) and found that mRNA expression of the IL-6 signal transducing unit 498 gp130 (*IL6ST*) is strongly enriched in these cells, whereas the mRNA of the IL-6 binding receptor alpha 499 chain CD126 (*IL6RA*) is absent. In the absence of CD126, the IL-6 signaling pathway can be activated 500 in trans through the binding of IL-6 to the soluble form of CD126 (sIL6RA) prior to binding to gp130. 501 Both IL-6 and sIL6RA are abundant in the bone marrow, and IL-6 trans-signaling through the PI3K/AKT 502 pathway can be activated in tumor cells (364). However, the endosteal niche renders DTCs 503 unresponsive to IL-6 trans-signaling (364). Interestingly, genetic analysis of DTCs revealed that only 504 4.4% (3/68) of nonmetastatic breast cancer patients harbored mutations in the gene for PI3K (PIK3CA), 505 whereas 34.3% (23/67) of metastatic breast cancer patients displayed PIK3CA mutations, indicating 506 that DTCs may undergo further selection to become more independent from their microenvironment 507 during cancer progression. Overall, these results strongly indicate that the endosteal niche provides 508 breast cancer cells with an environment supporting their survival, outgrowth and/or enabling tumor cells 509 to acquire genetic alterations (e.g., PIK3CA mutation) that render them more autonomous (354, 355, 510 364, 387).

511 In prostate cancer, CXCR-4/CXCL-12 and Annexin 2 (ANXA2)/CXCL12 interactions also play a 512 crucial role in the recruitment of tumor cells in the endosteal niche (167, 298, 318, 357). The targeting of 513 CXCR4 in model systems results in increased numbers of prostate cancer cells in the circulation, 514 supporting the notion that these tumor cells inhabit this endosteal niche (298, 357). The current 515 hypothesis is that prostate cancer cells compete with HSCs for space in the endosteal niche (298). 516 However, as opposed to breast cancer, it seems that prostate cancer cells homing in the endosteal 517 niche may benefit from this supportive environment for maintenance of dormancy but not tumor 518 outgrowth (42, 298, 320, 357). Notably, growth arrest-specific 6 (GAS6) is an osteoblast-derived ligand 519 of the MER, TYRO3 and AXL tyrosine kinase receptors that has been shown to induce tumor dormancy 520 in prostate cancer (320). When prostate cancer cells bind to osteoblastic cells in the endosteal niche,

521 they increase their expression level of AXL and consequently GAS6 inhibits tumor cell proliferation by 522 binding to AXL (320). Similarly, high MER expression levels in prostate cancer cells are associated with 523 tumor dormancy in the bone marrow (42). By contrast, when TYRO3 expression levels exceed AXL 524 levels, prostate cancer cells exhibit rapid growth (320). Thus, a balance between expression levels of 525 TYRO3 and AXL/MER may regulate prostate cancer cell dormancy in the endosteal niche. A similar role 526 for AXL in promoting dormancy in models of multiple myeloma has been reported (173). Overall, the 527 relative contribution of these niches/molecules to tumour cell dormancy in these various bone metastatic 528 cancers has yet to be validated in clinical samples.

529 The bone microenvironment is also an immune privileged site, offering protection of HSCs from 530 environmental insults and the resulting immune response. High resolution in vivo imaging shows co-531 localization of HSC and regulatory T cells (Treg) on endosteal surfaces in the trabecular bone marrow 532 areas in mice (111). Treg cells are known to be potent immune suppressors. In addition to vascular and 533 endosteal niches, it has been therefore proposed that Treg cells helped create an immune niche 534 supporting stem cell function whilst providing sanctuary from immune attack (111). The bone marrow 535 also contains very high numbers of myeloid-derived suppressor cells (MDSCs) (254, 370). MDSCs 536 suppress anti-cancer immune activity by inhibiting NK and CD8+ T cells (254, 370). Thus, this type of 537 protected environment would clearly also benefit resident tumor cells, preventing their elimination and 538 promoting their survival in bone. In addition, bone marrow mesenchymal stem cells also promote tumor 539 cell dormancy (247). See section VII for further discussion on the contribution of immune cells to tumor 540 development.

541

542 *2. Tumor cell dormancy*

543 Tumor cell dormancy is defined as the arrest in the cell cycle (also known as mitotic or cellular 544 dormancy). A second mode of dormancy refers to tumor mass dormancy of micrometastases where there is a balance between cell proliferation and cell death, the latter is widely believed to be due to immune surveillance and/or lack of blood supply (309). The signaling pathways through which tumor mass dormancy is controlled are largely unknown, mostly because of the lack of appropriate animal models that reproduce tumor dormancy in bone. Thus, although these two modes of dormancy coexist in the bone marrow, we have concentrated here on molecular mechanisms that regulate tumor cell dormancy in laboratory models.

551 In breast cancer, tumor cell dormancy appears to be determined by a balance between the 552 activities of activated protein kinases ERK1/2 and p38, where a switch towards ERK1/2 phosphorylation 553 favors proliferation whereas activation of p38 leads to guiescence (309). Mitogen- and stress-activated 554 kinase 1 (MSK1) is a downstream effector of the p38 and ERK1/2 signaling pathways (117). Using 555 experimental models of ER-positive human breast cancer (T47D, ZR-75) in which tumor cells form 556 latent micrometastatic bone lesions in vivo. Gawrzak and colleagues (117) showed that p38 depletion in 557 ER-positive human breast cancer cells decreases MSK1 expression. In turn, MSK1 depletion increases 558 the capacity of poorly metastatic ER-positive breast cancer cells to form overt metastasis in animals 559 (117). Thus, MSK1 is a dormancy enforcer and a negative regulator of metastasis initiation.

Another signal that regulates breast cancer dormancy in the bone marrow is leukemia inhibitory factor (LIF) (163). By binding to LIF receptor (LIFR), LIF negatively regulates *STAT3* (signal transducer and activator 3) in breast cancer cells. The loss of LIFR or STAT3 enables otherwise quiescent human MCF-7 breast cancer cells to proliferate and specifically metastasise to bone (163). Indeed, LIFR expression levels in primary tumor of breast cancer patients who are predicted to relapse in bone are significantly lower compared with those with a good prognosis (163), further supporting the observation that LIFR signaling mediates tumor cell dormancy in animal models of bone metastasis.

567 In prostate cancer, bone morphogenetic protein (BMP)-7 secreted from bone marrow stromal cells 568 promotes dormancy of prostate cancer stem-like cells, and an inverse correlation between expression of 569 the BMP7 receptor BMPR2 and occurrence of bone metastasis is found in patients with prostate cancer (177). By binding to BMPR2, BMP7 induces the quiescence of prostate cancer stem-like cells through
p38 activation and increased expression of the cell cycle inhibitor p21 (177). BMP7 also inhibits breast
cancer stem cell population and reduces bone metastasis formation in animals (41).

Bone-derived growth factors TGF β 1 and TGF β 2 exhibit competing functions on the behavior of tumor cells in the bone marrow (309). TGF β 2 promotes tumor cell dormancy, whereas TGF β 1 switches off dormancy, leading to rapid tumor growth *in vivo* (309). In a head and neck squamous cell carcinoma model of bone metastasis, TGF β 2 (but not TGF β 1) activates p38, which up-regulates the metastasis suppressor gene *DEC2* (31). In turn, DEC2 induces p27 and down-regulates cyclin-dependent kinase 4 (CDK4), leading to tumor cell quiescence (31). In model systems of prostate cancer bone metastasis, TGF β 2 induces dormancy through p38 activation and AXL/GAS-6 expression (381, 382).

580 Due to the diversity of the molecular mechanisms that regulate tumor cell dormancy in 581 laboratory models, these processes are difficult to validate in clinical samples. However, future reserch 582 will establish if targeting key drivers of dormancy can be used as a method of retaining tumor cells in 583 this state indefinitely, thereby preventing metastatic outgrowth and symptomatic disease.

584 3. Dormant cell reactivation

585 Bone resorption likely creates an environment that promotes tumor cell reactivation. Intravital 586 imaging of the bone microenvironment in murine models of multiple myeloma has shown that tumor 587 cells colonizing endosteal niches are in a dormant state (189). However, these tumor cells are reactived 588 and released from the endosteal niche upon treatment of tumor-bearing animals with a soluble form of 589 RANKL that stimulates osteoclast-mediated resorption (189). By contrast, sRANKL treatment has no 590 effect on tumor cells colonizing soft tissue sites (189). Androgen deprivation by orchidectomy stimulates 591 bone turnover of castrated animals bearing disseminated hormone-insensitive prostate cancer cells in 592 the bones, thereby also increasing the incidence of overt bone metastasis in these animals (251). A 593 similar effect was reported in an animal model of breast cancer, where ovariectomy-induced bone loss

594 triggered growth of disseminated hormone-insensitive breast cancer cells in bone (249,250). Thus, 595 osteoclast-mediated bone resorption plays an important role at an early stage in the establishment of 596 bone metastasis. This contention is also supported by experiments conducted in a mouse model of 597 indolent breast cancer bone metastasis, showing that VCAM-1 overexpression in tumor cells promotes 598 the recruitment of osteoclast precursors by binding to osteoclast integrin α 4 β 1, leading to osteoclast 599 formation and osteoclast-mediated bone resorption (214). In turn, bone-derived growth factorsTGF^{β1} 600 released from resorbed bone switches off dormancy, leading to rapid tumor growth in vivo (309). 601 Furthermore, PTHrP expressed by tumor cells can act in autocrine fashion by reducing pro-dormancy 602 LIFR gene expression (163), suggesting that PTHrP also plays a role in promoting tumor cell exit from 603 dormancy. Thus, changes to the bone environment in favor of bone resorption are sufficient to trigger 604 dormant cell reactivation. This idea is supported by the fact that bisphosphonates, by decreasing bone 605 resorption, improve elimination of DTCs in the bone marrow of breast cancer patients with a minimal 606 residual disease (307), and reduce development of bone metastases when given as a neoadjuvant 607 treatment (66, 123).

608

609

Niches, tumor cell dormancy and reactivation: current understandings & open questions

610 Experimental and clinical studies support the notion that vascular and endosteal niches can be 611 hijacked by arriving tumor cells to provide immediate shelter, thereby preventing their 612 elimination and promoting their survival in the bone marrow. Other existing niches, such as the 613 immune niche, may also support tumor cell survival in the bone marrow. However, many 614 aspects of the interplay between these niches and tumor cells remain elusive. The use of 615 clinically applicable imaging technologies such as PET and SPECT with niche-specific tracers 616 and single cell-omics techniques will certainly help to understand the dynamics of tumor-niche 617 interactions in the future.

• The diversity of the molecular mechanisms associated with tumor dormancy in the bone 619 marrow niches represent both a challenge and an opportunity for therapeutic targeting. How to 620 avoid unwanted effects on normal homeostasis while disrupting interactions that maintain tumor 621 cells in these bone marrow niches remains an open question.

622

623 IV. FITTING IN - ADAPTATION OF TUMOR CELLS TO THE BONE MARROW 624 MICROENVIRONMENT

During the time tumor cells are resident in the bone marrow, exiting and re-entering a dormant state, they rewire their biology to meet the demands of the tissue colonized, thus modifying their primary properties in order to adopt a genetic phenotype similar to bone cells that, in turn, facilitates their survival in the bone microenvironment (16, 291). This process is called osteomimicry (178).

Immunohistochemical analysis of human clinical samples in breast and prostate carcinomas clearly shows that cancer cells metastatic to the bone highly express bone proteins *in situ* (16, 55, 108, 196, 353). In particular, paired immunochemistry on human primary breast tumor samples and matched liver, lung or bone metastases showed that only bone metastatic tumor cells express bone proteins such as cathepsin K (CTSK), osteonectin, cadherin-11 (CDH-11), and Cx-43, which are normally expressed by osteoblasts or osteoclasts (16, 108, 196).

635 The functions of these osteomimicry genes have been studied in animal models of bone 636 metastasis. CDH-11 mediates interactions of breast and prostate cancer cells with osteoblasts in vitro, 637 and its silencing in tumor cells greatly reduces bone metastasis formation in vivo (55, 154, 323). Breast 638 cancer cells can get calcium from the osteogenic niche through Cx43 gap junctions that facilitate 639 calcium influx from osteogenic cells to breast cancer cells and, in turn, calcium promotes tumor cell 640 proliferation (354). Similarly, Cx43 overexpression in human LNCaP prostate cancer cells enhanced 641 their capability to induce bone destruction in vivo following intratibial tumor cell injection, and moderately 642 augmented tumor cell proliferation in vitro, when tumor cells were cocultured with osteoblasts (184).

643 Another example of osteomimicry is the expression of transcription factor RUNX2 (a master regulator of 644 osteoblast differentiation) in osteotropic tumor cells. The disruption of RUNX2 expression in breast 645 cancer cells abolishes their ability to form osteolytic lesions in vivo (266). RUNX2 in osteotropic breast 646 cancer cells promotes expression of metastasis-related factors [MMP-9, MMP-13, VEGF, osteopontin, 647 bone sialoprotein (BSP), ITGA5] and bone-resorbing factors (PTH-rP, IL-8), thereby explaining why 648 RUNX2 inhibition in tumor cells decreases skeletal tumor burden and osteolysis (200, 266). Forkhead 649 box F2 (FOXF2) is another example of master transcription factor that mediates epithelial-to-650 osteomimicry transition, increasing the tendency for breast cancer cells to metastasise to bone. In 651 particular, the orthotopic implantation of murine 4T1 breast cancer cells overexpressing FOXF2 or the 652 intracardiac inoculation of FOXF2-overexpressing human MDA-MB-231 breast cancer cells enhances 653 the formation of osteolytic bone metastases in animals (358). Mechanistically, FOXF2 directly 654 upregulates CTSK that, in turn, increases breast cancer cell invasion (358). Interestingly, high 655 expression levels of transcription factors FOXF2 and RUNX2 in primary mammary carcinomas correlate 656 with bone-specific metastasis in patients with breast cancer (200, 358).

657 It is highly likely that miRNA dysregulation in tumor cells contributes to osteomimicry (36). For 658 instance, the downregulation of miR-30, miR-135, and miR-203 enhances abnormal expression of 659 osteoblast-specific genes (CDH-11, RUNX2, SOST, ITGA5, BSP, OPN), which endows tumor cells with 660 full competence for survival in the bone marrow (75, 320). Other genes associated with osteomimicry 661 (DKK-1), osteoclastogenesis (IL-8, IL-11) and tumour cell invasiveness (CTGF, ITGA5, ITGB3) are 662 direct targets for repression by miR-30 family members, these miR-30s being downregulated in 663 osteotropic breast cancer cells (75). Conversely, miR-218 is highly expressed in human MDA-MB-231 664 breast cancer cells and acts as a promoter of bone metastasis formation through stimulation of the 665 expression of metastasis-related genes (CXCR-4, BSP and OP) that are associated with osteomimicry 666 and production of the bone-resorbing factor PTH-rP (321).

668 Osteomimicry: current understandings & open questions

- In situ expression of bone proteins in tumor cells from human bone metastasis specimens
 unequivocally establishes osteomimicry as a process occurring during the development of bone
 metastases in patients with advanced breast or prostate cancer.
- Experimentally, RUNX2, FOXF2 and some miRNAs (miR-30, miR-135, miR-203, and miR-218)
 function as master regulators of osteomimicry.
- The importance of osteotropic factors as potential biomarkers for the prediction of bone 675 metastasis risk and/or response to bone-targeted agents remains to be investigated.
- 676

677 V. DISRUPTING THE BALANCE - TUMOR-INDUCED BONE DESTRUCTION

The radiographic appearance of bone metastases ranges from typically destructive (osteolytic) to mostly bone-forming (osteoblastic) with most tumors demonstrating a mixture of lesions (**Figure 1**). There is always an imbalance between bone formation and bone resorption during the development of bone metastases. Therefore, predominantly osteolytic lesions are associated with high osteoclast activity and reduced osteoblast activity, whereas predominantly osteoblastic lesions have a high osteoblast activity and variable, but also often increased, osteoclast activity (35, 67).

The different molecular mechanisms associated with the formation of osteolytic lesions are described below (**Figure 4**), whereas tumor-derived factors governing the formation of osteoblastic lesions are described in the next section.

A. Factors Promoting Osteoclast-Mediated Bone Resorption

Several factors secreted by tumor cells stimulate osteoclast activity and bone resorption (PTHrP,
lysophosphatidic acid, macrophage-stimulating protein, prostaglandin E2, IL-8, IL-11, MMP-1, CCN3,
granulocyte macrophage-colony stimulating factor) (18, 26, 126, 252, 329, 361, 362). Among them,

691 PTHrP was the first to be recognized as involved in malignant osteolysis (126, 265). Using 692 immunohistochemistry in a retrospective series of 31 human breast cancer metastasis specimens, 693 PTHrP has been shown to be expressed in 92% of bone metastases (12 out of 13 samples) and 17% of 694 metastases to non-bone sites (3 out of 18 samples) (265). Early investigations showed that preventive 695 treatment of animals with a neutralizing antibody against PTHrP reduced the development of osteolytic 696 lesions caused by human MDA-MB-231 breast cancer cells (126). PTHrP binds to the type 1 697 parathyroid hormone receptor (PTHR1), a seven-transmembrane G protein-coupled receptor expressed 698 by osteoblast, which stimulates the expression of RANKL. In turn, RANKL binds to its receptor RANK on 699 osteoclast precursors, leading to the formation of new osteoclasts and therefore enhanced bone 700 resorption (126, 329). Moreover, tumor-derived PTHrP inhibits OPG production, thus promoting bone 701 metastasis (329). The production of PTHrP by tumor cells is induced by transcription factors RUNX2 702 and Gli2. RUNX2 is upregulated in osteotropic breast cancer cells and directly activates the Indian 703 Hedgehog (IHH) pathway characterized by the upregulation of the Gli family of zinc finger transcription 704 factors (Gli1, Gli2 and Gli3) (266). TGFβ released from resorbed bone also induces Gli2 expression in 705 tumor cells (3). In turn, Gli2 (but not Gli1 and Gli3) induces PTHrP expression in bone metastatic human 706 breast cancer cells and osteolysis in tumor-bearing animals (314). As a result, the blockade of the 707 RUNX2-IHH pathway in MDA-MB-231 breast cancer cells by Runx2 short hairpin RNA inhibition 708 prevents the osteolytic disease in bone metastatic animals (266). Likewise, the transcription factor MAF 709 mediates breast cancer bone metastasis through the control of many factors including PTHrP (259). 710 Interestingly, MAF expression in primary mammary tumors has been shown to predict treatment 711 outomes of the bisphosphonate zoledronic acid in reducing the incidence of bone metastases in early-712 stage breast cancer (64). See section IX for further discussion.

Hypoxia also induces PTHrP expression and secretion by tumor cells through a HIF-dependent mechanism (222). Although bone is highly vascularized, the absolute oxygen tension in the bone marrow is quite low, and there is a moderate oxygen gradient between the peri-sinusoidal regions, 716 which have the lowest levels of oxygen tension (9.9 mmHg), and the endosteal region (13.5 mmHg), 717 which is perfused with small arteries (313). Thus, tumor cells experience hypoxic conditions in the bone 718 marrow. Moreover, tumor cells are also susceptible to hypoxia as they grow in the bone marrow, which 719 is caused by reduced vascular supplies of oxygen and nutrients. The role of HIF-1 α in bone metastasis 720 formation has been therefore tested experimentally (146). The extent of bone destruction and 721 vascularisation of bone metastases in animals injected with MDA-MB-231 cells overexpressing an 722 active form of HIF-1 α was significantly increased compared to mock-transfected cells (146). HIF-1 α 723 also directly regulates the expression of transcription factor TWIST in human breast cancer cells (374), 724 and TWIST overexpression in osteotropic breast cancer cells promotes bone metastasis formation 725 through a mechanism dependent of miR-10b, facilitating tumor cell invasion and cancer-induced bone 726 destruction (74).

727 Platelet-derived lysophosphatidic acid (LPA) supports progression of osteolytic bone metastases in 728 breast cancer (26,27). By binding to its receptor LPA1 at the tumor cell surface, LPA promotes tumor 729 cell proliferation through the stimulation of a Pi3K/ZEB1/miR-21-dependent pathway (284). LPA also 730 induces the production of interleukins IL-6 and IL-8 by human breast cancer cells, which then stimulate 731 osteoclast-mediated bone resorption (26,27). Pharmacological inhibition of LPA action on its receptor, 732 using a LPA1 antagonist, substantially reduces progression of osteolytic bone metastases caused by 733 MDA-MB-231/B02 breast cancer cells in immunodeficient animals (27). Likewise, the treatment of 734 immunocompetent animals with a LPA1 antagonist inhibits spontaneous dissemination of murine 4T1 735 breast cancer cells in distant organs (lungs, liver) with no effect on primary tumor size (225).

Macrophage-stimulating protein (MSP) is produced by tumor cells in breast cancer (362). It binds to the RON receptor tyrosine kinase, which is expressed by osteoclasts but not osteoblasts, and stimulates osteoclast survival and activity (but not osteoclast differentiation) through a RANKindependent, Src phosphorylation-dependent pathway (4). The intratibial injection of MSP-expressing breast cancer cells in syngeneic wild-type mice causes a profound osteolysis (4). Moreover, the

therapeutic targeting of RON with tyrosine kinase inhibitor BMS-777607/ASLAN002 inhibits the formation of osteolytic lesions in tumor-bearing animals (4) and reduces bone resorption in postmenopausal women with advanced cancer (phase-I trial; ClinicalTrials.gov identifier NCT01721148).

745

746 **B.** Factors Suppressing Osteoblast-Mediated Bone Formation

Tumor cells not only stimulate osteoclast activity, but also inhibit osteoblast activity, thereby worsening the imbalance between bone formation and bone resorption, and promoting bone destruction (361). Main factors produced by tumor cells that have been shown to suppress osteoblast differentiation include activin A, the BMP inhibitor noggin, dickkopf-1 (DKK-1), and sclerostin (SOST-1) (198, 293, 326, 330, 395, 396).

752 Activin A is a member of the TGF- β superfamily of growth factors. It binds to activin type IIA 753 (ActRIIA) or type IIB (ActRIIB) receptors and induces the recruitment and phosphorylation of an activin 754 type I receptor (ActRIB), which then phosphorylates Smad2 and Smad3 intracellular signaling proteins 755 (198). In multiple myeloma, it has been reported that activin A secreted by plasma cells inhibits 756 osteoblast differentiation via Smad2-dependent downregulation of DLX (distal-less homeobox)-5 (198). 757 In breast and prostate cancer, activin A might modulate, via Smad signaling, the expression of pro-758 osteoclastic factors (IL-11, CTGF, MMP-1) (198). Interestingly, in animal models of breast cancer bone 759 metastasis and of multiple myeloma with osteolytic lesions, the treatment of mice with a soluble activin 760 receptor type IIA fusion protein (ActRIIA.muFc) blocks bone destruction (51). Specifically, ActRIIA.muFc 761 stimulates osteoblastogenesis and promotes bone formation in tumor-bearing animals, thereby 762 preventing cancer cell-induced suppression of bone formation (51).

Noggin is a BMP antagonist encoded by *NOG*. In breast cancer, *NOG* mRNA expression levels are
 significantly upregulated in bone metastatic lesions, compared to that observed in brain, lung and liver

Resions (326). The silencing of *NOG* in osteotropic breast cancer cell lines substantially reduces bone metastasis formation in animals (326). Similarly, *NOG* is expressed in human prostate cancer cells that metastasize to bone and cause osteolytic lesions in animals (293). Tumor-derived noggin interferes with physiologic bone coupling by inhibiting bone formation, which thereby prevents repair of osteolytic lesions generated by an excess of osteoclast-mediated bone resorption (293).

770 DKK-1 and SOST-1 are two Wnt (Wingless/int) protein antagonists. WNT agonists promote 771 osteoblast proliferation by binding to a receptor complex consisting of a member of the Frizzled 772 transmembrane receptor family and either LRP (low-density lipoprotein receptor-related protein) 5 or 773 LRP 6 (8). Both DKK-1 and SOST-1 exhibit redundant functions by blocking LRP5/6 binding to WNTs. 774 thereby inhibiting WNT signaling (8). High circulating levels of DKK-1 were first reported in multiple 775 myeloma patients with osteolytic lesions (332). Multipe myeloma cells express DKK-1 and the blockade 776 of DKK-1 using neutralizing antibodies results in a decrease of both osteolysis and skeletal tumour 777 growth in murine models of multiple myeloma (76, 372). DKK-1 is also expressed in breast, lung and 778 prostate cancers (39, 76, 130, 131). DKK-1 knockdown in breast and prostate cancer cell lines 779 decreases bone metastasis formation, while DKK-1 overexpression increases bone metastasis and 780 bone destruction in vivo (330, 396). Mechanistically, tumor-derived DKK-1 promotes osteolysis in animal 781 models of multiple myeloma and breast cancer bone metastasis and decreases the formation of 782 osteoblastic lesions in a model of prostate cancer bone metastasis by silencing canonical WNT 783 signaling of osteoblasts (76, 330, 396).

With regard to SOST-1, this WNT inhibitor is expressed in human primary breast tumors and breast cancer cell lines, especially those that are hormone unresponsive (143, 395). An anti-SOST antibody was shown to decrease the extent of osteolytic lesions in mouse models of MDA-MB-231 breast cancer bone metastasis (143, 395). As previously reported for DKK-1 in breast cancer (396), SOST-1 promotes cancer-induced bone destruction by silencing canonical WNT signaling of osteoblasts (143, 395). Furthermore, a treatment with an anti-SOST antibody also protects tumorbearing animals from cancer-induced muscle weakness, which is a debilitating event that can be associated with bone metastases in breast cancer patients (143). Plasma cells in multiple myeloma do not express SOST-1 (227). SOST-1 is however produced by osteocytes and treatment of animals with an anti-SOST antibody reduces osteolytic lesions induced by multiple myeloma, thereby preventing bone destruction (227).

795

796 C. Osteocytes – Silent Partners with a Role to Play

797 Osteocytes are terminally differentiated osteoblast lineage cells that reside in lacunae within the 798 mineralized bone matrix (6, 80). Osteocytes are by far the most abundant cells of the bone. They are 799 stellate cells that communicate with their environment via cytoplasmic projections termed dendrites. 800 Dendrites of osteocytes form Cx43-dependent gap junctions with dendrites of neighbouring osteocytes 801 as well as osteoblasts on the bone surface and cells in the bone marrow and vascular space, which 802 results in the formation of a communication network in the bone matrix (80). Osteocytes modulate bone 803 turnover by regulating osteoblast and osteoclast functions through the secretion of RANKL, SOST and 804 DKK-1, and control calcium homeostasis through remodeling of the osteocytic perilacunar matrix (6, 805 80). They act as mechanosensors to control responses to mechanical loading of the skeleton (6, 80). 806 Moreover, osteocytes regulate phosphate homeostasis through secretion into the circulation of 807 fibroblast growth factor (FGF)-23 (80).

The contribution of osteocytes to bone metastasis is only beginning to be uncovered. This may be explained by the fact that studying osteocytes remains very challenging due to their location within the mineralized bone matrix. Methods used to isolate osteocytes from the bone matrix remain difficult and the phenotype of isolated osteocytes is not necessarily maintained *in vitro* (80). For example, human primary osteocytic cells in 2D culture do not express SOST-1, DKK-1 and FGF-23 (54). Thus, *in vitro* methods that are used to isolate and culture osteocytes may be a limitation to the study of osteocyte 814 functions such as in bone metastasis. Despite the technical challenges, it has been shown that tumor 815 growth in bone induces pressure due to the lack of expansible space, suggesting that physical forces 816 might modulate mechanotransduction properties of osteocytes (310). Indeed, application of hydrostatic 817 pressure to cultures of MLO-Y4 osteocytic cells stimulated the secretion of factors associated with 818 enhanced survival and invasion of prostate cancer cells in vitro (310). Osteocyte-derived CCL-5 and 819 MMPs were among these factors promoting prostate cancer cell invasion in vitro (310). Whether these 820 osteocyte-derived factors promote tumor cell invasion in vivo remains to be determined. Tissue 821 engineered 3D bone models formed by primary human osteocytes have facilitated investigation into 822 osteocyte functions (54). For example, primary human osteocytes in a 3D-culture system produce FGF-823 23, SOST-1, and DKK-1 as opposed to osteocytes in 2D culture (54). Using a 3D model, it has been 824 shown that primary human prostate cancer cells induce a significant increase in the expression of FGF-825 23, RANKL and, to a lower extent, DKK-1 in primary human osteocytes, whereas SOST-1 expression is 826 drastically decreased when compared to that observed with osteocytes in the absence of tumor cells 827 (54). The authors suggested that the greater decrease in SOST-1 could favor the formation of 828 osteoblastic lesions (54). It is however unclear how SOST-1 expression in osteocytes is downregulated 829 by prostate cancer cells. These experimental findings are in contrast with the observation that high 830 circulating levels of SOST-1 are found in prostate cancer patients with osteoblastic bone metastases 831 (6). Further studies are therefore required to better understand the contribution of osteocytic-derived 832 SOST-1 in prostate cancer bone metastasis.

In an *in vivo* model of bone disease caused by human JJN3 multiple myeloma cells, it has been shown that osteocytic dendrites were in direct contact with JJN3 cells in the bone marrow, leading to increases in osteocyte apoptosis and osteocytic RANKL and SOST production (83). *In vitro* cocultures between osteocyte-like MLO-A5 cells and JJN3 myeloma cells showed that cell-to-cell contact activated bidirectional Notch signaling in osteocytes and multiple myeloma cells, which increased multiple myeloma cell proliferation and induced osteocyte apoptosis. In turn, the induction of apoptosis promoted 839 osteocytic RANKL secretion, which then stimulated osteoclast formation (83). Thus, interactions 840 between osteocytes and multiple myeloma cells generate a microenvironment supportive of increased 841 tumor growth and bone destruction (83). These findings are in agreement with the fact that treatment of 842 animals with an anti-SOST antibody prevents bone destruction in different preclinical models of multiple 843 myeloma bone disease (5TGM1, 5T2MM, and MM1.5) (227).

844 In a breast cancer animal model, Cx43 hemichannels in osteocytes have been shown to play a 845 critical role in the suppression of bone metastasis (393). Specifically, Cx43 osteocyte-specific knockout 846 mice and osteocyte-specific Δ 130-136 transgenic mice with impaired Cx43 gap junctions and 847 hemichannels showed increased tumor growth after intra-tibial injection of Py8119 mouse mammary 848 carcinoma cells (393). Additionally, R76W transgenic mice with functional hemichannels but not gap 849 junctions in osteocytes did not display a significant difference (393). Cx43 gap junctions mediate 850 communication between adjacent cells, whereas Cx43 hemichannels serve as a portal for the exit of 851 molecules in the extracellular microenvironment (80). Zhou and colleagues (393) established a specific 852 role for osteocytic Cx43 hemichannels in suppressing breast cancer growth and bone metastasis, 853 whereas osteocytic Cx43 gap junctions did not play such a role. In agreement with this observation 854 (393), ATP is released from osteocytes through Cx43 hemichannels and exerts inhibitory effects on 855 breast cancer cell migration in vitro and tumor growth in vivo (392). However, these findings are 856 contrary to the pro-metastatic role of Cx43 gap junctions between osteoblasts and breast cancer cells, 857

which promote progression of osteolytic lesions in animals (354).

858 Overall, these findings strongly suggest that osteocytes have a role to play in the development of 859 bone metastases. However, a lot of uncertainties remain as to whether osteocytes have bone 860 metastasis suppressor or promoter activities, and whether this activity depends on the cancer cell type 861 that metastasizes to bone. Further studies are therefore warranted to investigate the contribution of 862 osteocytes in bone metastasis formation.
D. The Fertile Soil - Contribution of the Bone Matrix

865 Bone is a riche source of growth factors, including TGF β , IGFs and PDGF (platelet-derived 866 growth factor) (361). For example, while there is no difference in bone marrow TGF- β levels between 867 healthy controls and castration-resistant prostate cancer patients without bone metastases, patients 868 with bone metastases have aberrantly high levels of TGF- β (161). Indeed, when released from the 869 resorbed bone matrix, TGFβ acts on tumor cells, via SMAD- and COX2-dependent signaling pathways, 870 and stimulates the expression of factors such as Gli2, PTHrP, the Notch ligand Jagged-1, IL-11 and 871 PGE2 (3, 168, 295, 361). Jagged-expressing tumor cells are capable of directly activating osteoclasts 872 by activating the Notch signaling pathway and the therapeutic targeting of Jagged-1 with a monoclonal 873 antibody inhibits bone metastasis formation in animals (295, 390). In addition, TGFβ released from the 874 bone matrix during bone destruction contributes to muscle weakness by decreasing Ca²⁺-induced 875 muscle force production (359). Bone-derived IGF-I stimulates growth of breast cancer cells via 876 activation of the IGF type I receptor (IGF-IR)/Akt/NFkB pathway, and IGFR-IR was found to be elevated 877 in 13/15 cases of bone metastases obtained from patients with a range of tumor types, supporting a role 878 for the IGF axis in development of human disease (147). Similarly, bone-derived IGF-II stimulates 879 skeletal outgrowth of prostate cancer cells in vivo (174). Bone-derived PDGF activates the Akt/PKB 880 survival pathway in osteotropic breast and prostate cancer cells, as PDGF receptors in tumor cells 881 growing in bone are highly expressed compared to nonmetastatic cancer cells (89, 199). Thus, 882 evidence from both model systems and clinical samples support that bone-derived growth factors 883 contribute to bone metastasis formation by promoting skeletal tumor outgrowth. To date, therapeutic 884 targeting of growth factors has so far not resulted in patient benefit however trials including anti-growth 885 factor agents as part of combination therapy for patients with bone metastases are ongoing. For 886 example, the XENERA-1 trial (ClinicalTrials.gov Identifier: NCT03659136) aims to assess the antitumor 887 activity of xentuzamab, a monoclonal antibody that binds both IGF-I and IGF-II and inhibits the binding

of these ligands to IGF-R, in patients with ER+/HER2- advanced or metastatic breast cancer and bone
 metastases.

890 Bone is a mineralized tissue, rich in calcium. We previously discussed the contribution of calcium 891 from the osteogenic niche that facilitates tumor cell proliferation through Cx43 gap junctions (354). 892 However, calcium is also released from bone during osteoclastic resorption. It binds on tumor cells 893 (breast, prostate, renal cell carcinoma) via a calcium-sensing receptor (CaSR) and promotes tumor cell 894 proliferation and migration (29, 110, 354). Additionally, calcium stimulates the secretion of PTHrP and 895 epiregulin by tumor cells (29, 361). PTH-rP promotes osteoclast-mediated bone resorption and 896 epiregulin decreases OPG expression in osteoblasts, thereby both contributing to the progression of 897 osteolytic lesions (29, 361).

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- 899 Osteolysis: current understandings & open questions
- Understanding the cancer-associated mechanisms that stimulate osteoclast-mediated bone
 resorption has led to the development of anti-resorptive pharmaceutical agents that have
 become established as a valuable additional approach to the treatment of bone metastases in
 patients with advanced cancer.
- The observation that tumor cells not only stimulate osteoclast activity, but also inhibit osteoblast
 activity, suggests that stimulating osteoblastic bone formation to promote bone repair could be
 a novel alternative approach to treat malignant skeletal lesions.
- The contribution of osteocytes to bone metastasis is only beginning to be uncovered. A better
 understanding of the interplay between osteocytes and tumor cells will represent an opportunity
 for therapeutic targeting.

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911 VI. TOO MUCH OF A GOOD THING - TUMOR-DERIVED FACTORS REGULATING 912 OSTEOSCLEROSIS

913 A number of molecular mechanisms responsible for the formation of osteoblastic lesions have 914 been identified (**Figure 5**), described in the following sections.

915 A. Factors Promoting Osteoblast-Mediated Bone Formation

916 Tumor cells in the bone environment secrete factors that activate osteoblasts, leading to the 917 formation of skeletal lesions with extensive new bone deposition(osteosclerosis) (210, 248). Among 918 them, endothelin-1 (ET-1) was recognized as a major mediator of osteosclerosis; it stimulates 919 osteoblast proliferation and inhibits osteoclast activity and motility (2, 53, 210, 378). In this respect, 920 prostate cancer patients with bone metastases have far higher circulating levels of ET-1, compared to 921 those with localized cancer (276). Interestingly, TMPRSS2-ERG is the most frequent fusion gene 922 expressed in prostate cancer, it is associated with cancer progression, and its expression in PC3c-T1E4 923 prostate cancer cells has been demonstrated to promote ET-1 expression and formation of osteoblastic 924 lesions in animals (84). Human ZR-75-1 breast cancer cells that produce ET-1 stimulate new bone 925 formation and osteoblast proliferation in organ cultures, and osteoblastic metastases in animals (378). 926 Stimulatory effects of ET-1 on osteoblasts are mediated by two receptors, ETAR and ETBR, which 927 activate similar signaling pathways and down-regulate expression of the Wnt signaling inhibitor DKK-1 928 (276). Osteoblast proliferation and bone metastasis are both inhibited by ETAR antagonists atrasentan 929 and zibotentan, as well as by the dual ETAR and ETBR antagonist bosentan, highlighting the prominent 930 role played by ET-1 in the formation of osteoblastic lesions in preclinical settings (276, 378). Despite 931 this, both atrasentan and zibotentan have failed to show benefit in CRPC patients with bone metastases 932 (47, 240).

Breast and prostate cancer cells can produce BMPs, such as BMP-2, BMP-4 or BMP-6, which facilitate the development of osteoblastic bone metastates by stimulating tumor growth and osteogenesis (77, 78, 171, 195). In this respect, prostate cancer cell-derived BMP-4 mediates

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936 conversion of endothelial cells into osteoblasts, thereby promoting aberrant bone formation (204). 937 Analyses of human samples of prostate cancer bone metastases confirmed the presence of cells co-938 expressing endothelial and osteoblastic markers (Tie-2 and osteocalcin, respectively), which together 939 with the detection of increased expression of BMP-4 in bone metastases compared to that of primary 940 prostate tumors, support the hypothesis that endothelial-to-osteoblast conversion could also take place 941 in human disease (204). Overall, these findings (77, 78, 171, 195, 204) illustrate how the dysregulation 942 of BMPs can have deleterious effects on the bone microenvironment. Furthermore, the BMP inhibitor 943 noggin is also secreted by tumor cells and it is the balance between BMPs and noggin that determines, 944 at least in part, the phenotype of breast and prostate cancer bone metastases (293).

945 Prostate cancer cells secrete multiple WNT agonists, including canonical WNTs 3A, 7B and 946 10B, which, by binding to LRP5/6, are known mediators of osteoblast differentiation and mineralization 947 (129, 202, 210, 237). However, the WNT antagonist DKK-1 is also secreted by prostate cancer cells 948 and, as aforementioned for BMPs and noggin (293), it is the relative expression levels of WNT agonists 949 and DKK-1 that determine the phenotype of skeletal lesions (130). For example, C4-2B prostate cancer 950 cells express the WNT agonists WNT7A and WNT8B, but not DKK-1, and they induce mixed 951 osteoblastic/osteolytic lesions in animals (130). DKK-1 overexpression in C4-2B cells antagonizes WNT 952 functions, which leads to the suppression of WNT signaling in osteoblasts and results in the formation of 953 highly osteolytic lesions in animals (130). Among the many proteins downstream of WNT, autocrine 954 WNTs induce BMP-4 and BMP-6 expression in prostate cancer cells that, in turn, promotes osteoblast 955 differentiation (77, 195). WNT expression in tumor cells is itself regulated by many factors. For example, 956 T-box family transcription factor TBX2 is overexpressed in human prostate cancer specimens and bone 957 metastases from xenograft mouse models of human prostate cancer (239). It promotes transcription of 958 WNT3A in prostate cancer cells and the blockade of WNT3A with neutralizing antibodies dramatically 959 reduces experimental bone metastasis formation (239). Similarly, WNT5A and WNT7B are targets for 960 the transcription factor ERR α ("Estrogen Receptor Related Receptor alpha") and the androgen receptor 961 (AR), respectively, which are both highly expressed in castration-resistant prostate cancer cells, and
962 they promote tumor growth and development of osteoblastic lesions in animals (108, 391). Thus, there
963 is evidence that WNT signaling is central to osteoblast-stimulatory activity of metastatic prostate cancer,
964 however therapeutic targeting of this pathway is in its infancy (237).

965 PTHrP can be actively involved in the progression of osteoblastic lesions in prostate cancer by 966 enhancing proliferation of bone marrow stromal cells and early osteoblast differentiation (203). 967 Moreover, prostate-specific antigen (PSA), a serine protease expressed by prostate cancer cells and a 968 well-known marker of cancer progresssion, can cleave IGF binding protein (IGFBP)-5, rendering IGF-I 969 available to bind to its receptor and stimulate osteoblast proliferation (219). PSA also enhances the 970 bioavailability of TGF- β in the bone microenvironment (379). Like PSA, production of urokinase-type 971 plasminogen activator (uPA) by prostate cancer cells can increase IGF-I and TGF-β bioavailibility to the 972 bone microenvironment (113).

Several other osteoblast-regulatory factors expressed by tumor cells have been identified. These include growth factors [PDGF BB (377), FGFs (FGF-8, FGF-9) (201, 342) and VEGF (176)], adrenomedullin (299), TGF β -regulated gene PMEPA1 (107) and prostatic acid phosphatase (PAP) (188). Prostate cancer cells can also secrete neuropeptides, such as substance P (124) and Sema3A (112), which stimulate osteoblast differentiation.

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979 B. Factors Suppressing Osteoclast-Mediated Bone Resorption

Tumor cells that induce osteoblastic lesions not only stimulate osteoblast activity but may sometimes also inhibit osteoclast activity. Among the osteoclast inhibitors produced by cancer cells are ET-1 and OPG (210). Patients with metastatic prostate cancer have high circulating levels of ET-1 and OPG (90, 210, 383). Tumor-derived ET-1 directly inhibits osteoclast-mediated bone resorption by binding to the surface of osteoclasts *via* membrane receptors ETA and ETB (2, 53). Tumor-derived

985 OPG inhibits osteoclast differentiation by binding to RANKL, thereby preventing its interaction with 986 RANK (183). The overexpression of OPG in human C4-2 prostate cancer cells protects these cells from 987 TRAIL (tumor necrosis factor-related apoptosis-inducing ligand)-induced apoptosis, decreases 988 osteoclast formation and promotes the formation of osteoblastic lesions in animals (70). OPG 989 expression in prostate cancer cells is regulated by factors such as PAP, a tumor-derived acid 990 phosphatase that promotes osteoblast differentiation and bone mineralization (175, 188). PAP 991 knockdown in pro-osteoblastic VCAP prostate cancer cells decreases OPG while increasing 992 RANK/RANKL expression (175). Conversely, PAP overexpression in pro-osteolytic PC3M prostate 993 cancer cells has the inverse effect, increasing OPG while decreasing RANK/RANKL expression (175). 994 The transcription factor ERR α induces OPG expression in MDA-MB-231-B02 breast cancer cells, 995 thereby inhibiting osteoclast differentiation (109). Tumor-derived PSA also stimulates OPG production 996 and inhibits RANKL expression in osteoblasts (379). BMP-2 enhances Wnt/beta-catenin-dependent 997 transcriptional activation of the OPG promoter in osteoblasts (287). Thus, tumor-derived ET-1 and OPG 998 produced by both tumor cells and osteoblasts, by means of their ability to inhibit osteoclast activity, 999 contribute substantially to the formation of osteosclerostic bone metastases in model systems. When it 1000 comes to human disease, evidence that tumor-derived OPG contributes to the pathology of bone 1001 lesions is limited, hampered by the lack of bone metastases available for research. A meta-analysis 1002 found that in studies of prostate cancer, patients with bone metastases had higher levels of serum OPG 1003 compared to patients with metastases in other sites or healthy control (383). However, as this study did 1004 not include any information about the lesion types of the patients (sclerotic/lytic/mixed) it was not 1005 possible to link elevated serum OPG levels to decreased osteoclast activity and a shift in the balance 1006 towards osteoblastic bone lesions.

1007

1008 Osteosclerosis: current understandings & open questions

- Despite elevated levels of the osteoblast stimulating factor ET-1 in patients with bone
- 1010 metastases and supportive data from a number of *in vitro* and *in vivo* model systems, drugs 1011 targeting ET receptors have failed to provide benefit in trials of CRPC.
- WNT signalling regulates osteoblast differentiation and is activated in human prostate cancer,
 however the precise role of WNT family members in development and progression of the
 disease remains to be established.
- Studies that combine detailed characterisation of bone lesions with paired measurements of
 factors modifying bone turnover in serum and/or bone marrow are required to provide evidence
 of which molecules are the most promising therapeutic targets in metastatic prostate cancer.
- 1018

1019 VII. CONTRIBUTION OF BONE MARROW CELLS TO TUMOR DEVELOPMENT -

1020 MULTIPLE INTERACTIONS BEYOND THE VICIOUS CYCLE

1021 In addition to the main cell types responsible for bone remodeling described above (osteoblasts, 1022 osteocytes and osteoclasts), the bone microenvironment includes a myriad of interconnected cell 1023 populations, including a rich vascular network, immune cells, adipocytes, nerve cells, and 1024 megakaryocytes. Tumor cells arriving in this environment are proposed to utilize the mechanisms that 1025 regulate normal physiological processes in order to avoid immune surveillance and establish cellular 1026 interactions that support their expansion to overt metastases. As described in earlier sections, 1027 endothelial cells contribute in tumor cell extravasation, tumor cell dormancy and formation of 1028 osteoblastic lesions (119, 204, 267, 273). The role of platelets in stimulating bone metastasis formation 1029 has also been described above (26, 27, 190, 191). With regard to megakaryocytes, the platelet-1030 producing cells, little is known about their role in bone metastasis, with both promoting and inhibitory 1031 roles having been reported (191, 223). In the following sections we chose to cover some of the key 1032 discoveries linking immune cells, nerve cells and adipocytes to the development of bone metastases in 1033 solid tumors.

A. The Immune Cells of the Bone Microenvironment

1035 It has long been clear that the immune system plays an integral part of both normal bone 1036 homeostasis, as well as in a number of pathologies associated with bone loss, mainly through the link 1037 with inflammation. Combining the bone biology and immunology research fields to increase our 1038 understanding of their close connection has resulted in the new discipline of "osteoimmunology" (245). 1039 Initially focused on the bone-destructive effects of immune infiltrates through stimulation of osteoclasts 1040 by pro-inflammatory cytokines, research is expanding to other areas, including bone metastasis (253, 1041 370). Although the inflammatory response undoubtedly contributes to the extent and severity of cancer-1042 induced bone disease, as covered in the following sections, evidence from model systems support that 1043 immune cells may also affect tumor cell colonization and progression in bone (Figure 6).

1044

1045 **1**. Immune cells inhibiting local tumor growth in the bone microenvironment.

1046 CD8+ T cells

1047 CD8+ T cells are central players in controlling infections and cancer, recognised as one of the most 1048 important immune cells associated with tumour destruction (253). By cross-presenting tumor antigens, 1049 dendritic cells (DCs) activate CD8⁺ T cells. In turn, tumor-specific cytotoxic CD8⁺ T cells participate in 1050 the killing of antigen-positive tumor cells (45). The anti-tumor effects of tumor-specific cytotoxic CD8⁺ T 1051 cells is dependent on their ability to produce interferon (IFN)-y. Pioneering work by the Faccio group has 1052 demonstrated that activation of CD8⁺ T cells reduces bone metastasis formation in animals, whereas 1053 depletion of CD8+ T cells enhances it (386). Specifically, using phospholipase C gamma (PLCy) 2-/-1054 mice, which have broadly compromised immune responses and are osteopetrotic due to reduced 1055 osteoclast number and functionality, Zhang and colleagues (386) reported an unexpected increased 1056 tumor growth in bone despite osteoclast dysfunction. This was found to be due to a defective anti-tumor 1057 T cell response in tumor-bearing PLC $\gamma 2^{-/-}$ mice. Similar experiments were then conducted in Lyn^{-/-} mice, 1058 which have enhanced T-cell responses and decreased bone mass due to high number of osteoclasts.

1059 Lyn-⁻ mice had a reduced bone tumor burden despite osteolysis (386). Importantly, injection of antigen-1060 specific wild-type cytotoxic CD8+ T cells in PLCy2-/- mice or depletion of CD8+ T cells in Lyn-/- mice 1061 normalized tumor growth in bone, regardless of osteoclast activity (386). This study is important in that it 1062 used both genetic and pharmacological approaches to demonstrate that the extent of tumor growth in 1063 bone is not only linked to the level of osteoclast activity as stipulated by the vicious cycle of cancer-1064 induced bone destruction. In addition, these findings demonstrate that CD8+ T cells have the potential to 1065 act as regulators of tumor growth in bone. This contention is supported by the observation that 1066 transcription factor ERR α in murine 4T1 breast cancer cells inhibits the progression of bone metastases 1067 by increasing the recruitment of CD8⁺ T cells in the bone marrow (28). However, this remains to be 1068 established for human cancers where our capacity to identify T cell subsets in bone metastatic foci is 1069 limited.

1070

1071 Natural killer (NK) cells

Mature NK cells represent 1% of the lymphocyte population in bone, which is the primary site of murine NK cell development (253). In contrast, human NK cells are shown to differentiate from precursors and located in the secondary lymphoid organs like spleen and lymph nodes, and single cell RNA sequencing of NK cells isolated from both blood and bone marrow of healthy donors has revealed the presence of multiple heterogenous subsets with potentially different functions (373).

NK cells are involved in the nonspecific elimination of tumor cells through the production of IFN γ , release of cytolytic granules or TRAIL/FASL-induced apoptosis (253). Pathways of IFN induction are regulated by IFN regulatory factors (IRF3, IRF5 and IRF7) and NF κ B (20). Bidwell and colleagues (20) found that *irf7* expression was suppressed in murine 4T1.2 mouse breast cancer cells isolated from bone metastases, compared to those of matched primary mammary tumors. Enforced expression of Irf7 in bone metastatic 4T1.2 cells restored an antimetastatic immune response in immunocompetent tumorbearing animals (20). Conversely, the inoculation of Irf7-overexpressing 4T1.2 cells to mice deficient in 1084 NK and CD8⁺ T cell responses led to accelerated development of bone metastases, compared to 1085 immunocompetent mice (20). Similarly, the impairment of NK-cell-mediated anti-tumour immunity with a 1086 JAK/STAT inhibitor enhanced skeletal tumor burden in preclinical models of breast cancer metastasis 1087 (25). Taken together, these data indicated that NK cells (and CD8⁺ T cells), through the production of 1088 IFN-y, contributed to the suppression of bone metastasis and that NK cells are potential therapeutic 1089 targets in this setting. The clinical relevance of these findings was confirmed in over 800 patients in 1090 whom high expression of Irf7-regulated genes in primary tumors was associated with prolonged bone 1091 metastasis-free survival (20). A comprehensive review describing how NK cells may control metastasis 1092 points out that NK cells appear to have a particular role in reducing metastatic dissemination and 1093 speculates that this may be due to their ability to eliminate tumor cells that escape the 1094 immunosuppressive microenvironment of the primary tumor (211). It also includes an overview of 1095 clinical trials with immunotherapy agents boosting NK cell effector functions, the outcomes of which will 1096 provide important information about the impact of NK cells in metastatic disease, including in skeletal 1097 metastasis.

1098

1099 2. Immunosuppressive cells promoting local tumor growth in the bone 1100 microenvironment

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1102 Myeloid Derived Suppressor Cells (MDSCs)

1103 MDSCs describe a heterogeneous collective of immature progenitor populations for the myeloid cells, 1104 for which a variety of roles in tumor progression have been reported (172). Bone marrow accumulation 1105 of MDSCs is found in many cancer types, indicating pathological disruption of myeloid cell maturation 1106 (114). An important role of MDSCs in the metastatic process is their immune-suppressive functions, 1107 which include induction of oxidative stress, interference with lymphocyte trafficking and expansion of 1108 Treg cells (172). MDSCs are proposed to increase the number of Treg cells and modify tumor growth in 1109 bone independent of osteoclast activation through modification of T cell responses (45). For instance, 1110 PLC $\gamma 2^{-/-}$ mice are osteopetrotic due to reduced osteoclast number and functionality (45). Interestingly,

1111 despite osteoclast dysfunction, tumor growth in bone of PLC $\gamma 2^{-/-}$ mice was significantly higher than that 1112 observed in their wild-type counterparts due to an aberrant increased percentage of MDSCs in the bone 1113 marrow that, in turn, inhibited anti-tumor T cell response in tumor-bearing PLC $\gamma 2^{-/-}$ mice (45, 386).

1114

Few studies have investigated the potential role of MDSCs in human bone metastases, but a recent report compared polymorphonuclear (PN-) MDSC distribution in primary prostate tumours (n=90) and their corresponding lymph node metastases (n=37) to that of bone metastases (n=35) (363). PN-MDSCs were found to mainly infiltrate the stroma (rather than the epithelial areas), and that this was more prominent in the metastases compared to the primary tumour. The authors propose that this stromal location would facilitate better suppression of infiltrating T cells by the PN-MDSCs and that the high levels of CXCL5 in bone may drive MDSC infiltration and ultimately metastatic progression (363).

1122

A combination of the CXCR4 antagonist AMD3465 and the IDO1 inhibitor D1MT has been shown to delay the progression of breast cancer bone metastases in mice through activation of CD8+ T-cells and inhibition of Treg cells and MDSCs, supporting that suppression of MDSCs could potentially reduce metastatic progression in bone (385).

1127

1128 MDSCs isolated from tumor bearing mice have been also shown to be able to differentiate into 1129 functional osteoclasts in vitro and in vivo (81, 288). Interestingly, only MDSCs from mice with confirmed 1130 tumor growth in bone had osteoclastic potential, whereas those isolated from mice with peripheral 1131 tumors or control mice did not (288). The increased understanding of the inter-connectivity between 1132 cells residing in the bone marrow has resulted in studies exploring the effects of anti-resorptive agents 1133 beyond their traditional osteoclast targets. For example, in mouse models, a single, clinically relevant 1134 dose of the osteoclast inhibitor zoledronic acid has widespread effects on a number of cell types in the 1135 bone marrow, including hematopoietic stem cells, myeloid-biased progenitor cells and lymphoid-biased 1136 cells (340). Importantly, bone marrow cells isolated from zoledronic acid treated animals, but not from 1137 control, were able to suppress tumor growth in vivo when co-injected with tumor cells, supporting the 1138 finding that anti-resorptive agents could support the generation of tumor-suppressing myeloid cells 1139 (340). In follow-up studies, these findings were confirmed, demonstrating that even a single dose of 1140 zoledronic acid skews myeloid progenitor cells to enter the macrophage, rather than the osteoclast 1141 lineage (339). This exemplifies the potential for unexpected (both beneficial and harmful) effects of anti-1142 cancer therapies on bone marrow cell populations with implications for tumour progression, generally 1143 not considered when assessing the clinical benefits of cancer treatment. As the anti-resorptive 1144 bisphosphonates are increasingly used as adjuvant therapies in post-menopausal breast cancer without 1145 the precise mechanism conveying their positive effects on survival (66), it will be interesting to see if 1146 additional patient benefit could be linked to effects of these agents on a range of bone marrow cell 1147 populations.

1148

1149 Macrophages

1150 Macrophages develop from circulating monocytes within tissues and are heterogeneous and highly 1151 plastic cells, which can polarize into pro- or anti-inflammatory sub-types (M1 and M2, respectively) 1152 depending on signaling cues (45, 118). However, there are many other discrete sub-populations across 1153 the M1/M2 spectrum determined by the location and activation status of macrophages (45, 118). 1154 Macrophages are consistently found in bone metastases from patients with prostate cancer (369). In 1155 breast cancer, tumor-associated macrophages are also significantly increased in bone metastases 1156 compared to matched primary mammary tumors (394). Experimentally, tumor-associated macrophages 1157 were found to promote breast and lung cancer bone metastasis formation (102, 148). Additionally, a population of specialist osteal tissue macrophages termed 'osteomacs', whose normal function is to 1158 1159 regulate osteoblast differentiation (50), were found to facilitate formation of osteoblastic lesions in an 1160 animal model of prostate cancer (369). Nonetheless, their specific role in bone metastasis has proven 1161 elusive, in part because of ablation techniques that remove a number of myeloid related populations, 1162 including the closely related osteoclast precursors that are established as major drivers of bone 1163 metastasis. For example, treatment of animals with clodronate-encapsulated liposomes markedly 1164 reduced the number of monocytes in peripheral blood, and the formation of bone metastasis when 1165 HARA-B lung cancer cells were injected intracardiacally to mice (148). However, in this study, 1166 clodronate-encapsulated liposomes not only reduced macrophages within tumors, but also osteoclasts 1167 in metastastic bone lesions, thereby explaining the reduction of bone destruction (148). Similarly, the 1168 use of an anti-mouse CD115 monoclonal antibody, which specifically targets monocytic cells, inhibited 1169 breast cancer bone metastasis formation in animals by blocking osteoclast activity (102). Beside the use 1170 of techniques that can directly interfere with osteoclast function (102, 148), tumor cells can recruit 1171 macrophages and osteoclasts through the same mechanism of action. For example, breast cancer-1172 derived CCL2, which is a ligand for the chemokine receptor CCR2 expressed by myelomonocytic 1173 progenitors such as macrophages and osteoclast precursors, can stimulate through the same molecular 1174 mechanism the migration of macrophages in lung parenchyma and the differentiation of osteoclasts in 1175 the bone marrow, which ultimately aids metastasis to lungs and bone (214). Similar findings were 1176 reported with CCL2 produced by human prostate cancer cells, which promoted recruitment of 1177 macrophages within subcutaneous tumor xenografts and osteoclast-mediated bone destruction in 1178 animals bearing bone metastases (232). Thus, despite some suggestions that macrophages and 1179 osteomacs can contribute to bone metastasis (45, 369), further studies, including of samples human 1180 bone metastases, are needed to establish the precise mechanisms that could regulate these highly 1181 adaptable cells in the context of tumour growth in bone.

1182

1183

1184 Dendritic cells

1185 Dendritic cells are specialised antigen-presenting cells that are derived from hematopoietic bone 1186 marrow progenitor cells that differentiate into 2 subsets: conventional or myeloid dendritic cells (mDCs, 1187 similar to monocytes, produce IL-12) and plasmacytoid dendritic cells (pDCs, resembling plasma cells, 1188 produce IFN- α). These cells are responsible for presentation of antigens on their surface to induce T 1189 cell activation and prime CD8⁺ T cells (253). However, DCs in tumors can have limited antigen-1190 presenting function, thereby affecting the generation of anti-tumor immune responses (218). 1191 Additionally, DCs in cancer may exhibit immunosuppressive properties under certain circumstances 1192 (218). For example, using different syngeneic breast cancer models, Sawant and colleagues (289) 1193 observed an increased number of pDCs with increased bone metastasis in animals. Conversely, 1194 depletion of pDCs following treatment of animals with PDCA1 antibody prevented breast cancer bone 1195 metastasis formation (289). Furthermore, isolated CD8⁺ T cells from pDC-depleted mice exhibited 1196 enhanced cytotoxic activity compared to those from untreated animals, indicating that in bone-1197 metastatic disease pDCs exhibit immunosuppressive properties on CD8+ T cells (289). Thus, there is 1198 some evidence to support that pDCs may be critical regulators of bone metastasis, however this is one 1199 of the least investigated immune cell types in this context, with a paucity of informative clinical studies to 1200 provide solid data to allow a conclusion regarding their importance to be drawn at this point.

1201

1202 Regulatory T (Treg) cells)

Treg cells are potent immune suppressors, impairing CD8⁺ cell proliferation (45). The role of Treg cells in bone remodeling has not been extensively studied, with a few examples of inhibition of osteoclast maturation by Treg cells due to their production of IL-10, IL-4 and TGF β (30). In prostate cancer, Treg cells are significantly increased in the bone marrow of patients with bone metastasis compared to those without (389). Furthermore, the intravenous injection of activated Treg cells to immunodeficient NOD/SCID mice bearing human PC3 prostate cancer skeletal lesions leads to a reduction of bone destruction, due to the osteoclast-inhibitory effect of Treg cells (389). 1210 A study by Jiao et al (161) has shed light on why a combination of anti-CTLA-4 (ipilimumab) and anti-1211 PD-1 (nivolumab) checkpoint inhibitors that reduce primary prostate tumour growth in patients are 1212 largely ineffective in reducing bone metastatic disease. The study compared levels of TGFB in bone 1213 from patients with and without bone metastases and mapped T cell subsets in primary tumors and bone 1214 metastases from patients treated with ipilimumab. Results showed that the increased levels of TGFB 1215 released during cancer-induced bone resorption caused helper T cells to polarise into Th17 CD4 cells 1216 instead of the Th1 CD4 effector cells required to trigger an anti-tumour immune response (161). 1217 Combining anti-TGF β and anti-CTLA-4 therapy in a mouse model resulted in reduced bone metastasis, 1218 a strategy that the researchers now will take forward in clinical trials of patients with metastatic prostate 1219 cancer (161). This study (161) is an example of 'reverse translation', where a clinical observation is 1220 explored in model systems to identify mechanisms responsible for the observed effects and how they 1221 can be overcome. It also highlights that the specific immune tumor environment in bone presents a 1222 particular challenge when considering immunotherapy approaches in patients with bone metastases.

1223

1224 Immune cells and bone metastasis: current understandings & open questions

Depletion of CD8+ T cells results in increased bone metastasis in model systems, but direct
 evidence for a role of CD8+ T cells in development of human skeletal metastases is missing.

- The anti-metastatic functions of NK cells are not yet established in human disease, but evidence from models of bone metastasis support their role in suppressing tumour growth.
- Myeloid Derived Suppressor Cells (MDSCs) have been identified in bone metastases in model
 systems and in human samples, where they are proposed to increase Tregs and inhibit immune
 elimination of tumour cells.
- Macrophages are highly plastic cells with context-specific functions, their role in the different
 stages of human bone metastasis remains to be defined.

Plasmacytoid dendritic cells (pDCs) may be critical regulators of bone metastasis in model
 systems, however there is a paucity of informative clinical studies to draw any conclusion at this
 point.

The specific immune tumour environment in bone presents a particular challenge when
 considering immunotherapy approaches in patients with bone metastases. Differences between
 murine and human immune cell populations must be considered when translating findings from
 in vivo model systems to human disease.

1241

1242 **B. Nerve cells**

Bone is highly innervated, and a recent extensive review describes how bone homeostasis is influenced by sympathetic, parasympathetic, and sensory nerves (95). How bone remodelling is regulated by nerve cells is illustrated by the example of norepinephrine, released by sympathetic nerves, which activates β 2-adrenergic receptors expressed by osteoblasts, stimulating synthesis of RANKL that in turn modifies both osteoblast and osteoclast activity (95).

1248 As a cancer diagnosis is associated with increased levels of stress and depression, a number of 1249 studies have investigated whether stress-induced activation of the sympathetic nervous system (SNS) 1250 results in increased bone metastasis. In a model of learned helplessness (chronic immobilization 1251 stress), intracardiac injection of MDA-MB-231 breast cancer cells in mice that had undergone 2 weeks 1252 of SNS activation resulted in increased number of bone metastatic foci associated with larger lytic bone 1253 lesions, compared to control (44). Similar results were obtained when injecting MDA-MB-231 breast 1254 cancer cells in mice pre-treated for 3 weeks with isoproterenol (ISO), which is used as a surrogate of 1255 SNS activation through stimulation of the β 2-adrenergic receptor (44, 234). Importantly, these effects 1256 observed in mice under chronic stress or treated with ISO were inhibited by administration of the B2-1257 blocker isopropranolol (44, 234), supporting that sympathetic nerve activity mediates the 'pro-metastatic' 1258 effect of chronic stress. Furthermore, breast cancer bone metastasis in animals that are under chronic

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1259 stress is inhibited by knocking down RANK in MDA-MB-231 cells (44). Mechanistically, it is proposed 1260 that stress-induced activation of the β2-adrenergic receptor in osteoblasts stimulates RANKL production 1261 that, in turn, promotes MDA-MB-231 cell migration to bone in a RANK-dependent manner (44). ISO pre-1262 treatment of bone marrow stromal cells also induce the expression of pro-inflammatory cytokines (IL-1), 1263 IL-6) that increase the expression of E- and P-selectins by endothelial cells and the subsequent 1264 adhesion of MDA-MB-231 breast cancer cells to these cells under static and dynamic conditions in vitro 1265 (56). It has also been suggested that DTCs residing in endosteal niches could be affected by 1266 norepinephrine (82). Specifically, Decker et al. (82) found that the binding of norepinephrine to β 2-1267 adrenergic receptors affects osteoblasts in vitro through downregulation of the dormancy-inducing 1268 molecule GAS6, thereby re-activating proliferation of dormant disseminated prostate cancer cells that 1269 interacted with these osteoblasts. Taken together, these experimental data (44, 56, 82) suggest that 1270 stress-induced activation of the SNS prior to tumor cells arriving in bone alters the microenvironment to 1271 become more supportive of tumor outgrowth. Whether this also applies in human disease is difficult to 1272 investigate. A study including over 100,000 women in UK did not find any evidence of increased risk of 1273 breast cancer in those who reported high levels of stress (292). The current experimental evidence for 1274 neuronal involvement therefore relates to progression of established disease, and great care should be 1275 taken not to suggest that patients are in any way responsible for their disease progression through their 1276 ability to manage the inevitable stress associated with a cancer diagnosis and treatment.

Once bone metastases are established and progressing, their interaction with the nervous system is obvious. Pain is one of the most common and difficult to treat complications associated with skeletal metastases (229). Tumor cells, their associated stromal cells and osteoclasts can generate pain by releasing algogenic substances including protons (create acidosis), bradykinin, endothelins, prostaglandins, proteases, and tyrosine kinase activators such as nerve growth factor (NGF) (100). Sensory fibers in the bone marrow express acid-sensing nociceptor TRPV1 (transient receptor potential vanilloid 1) and the NGF tyrosine kinase receptor type 1 (TrkA) (23, 351). In murine models of intra1284 femoral or intra-tibial injection of NGF-expressing breast, prostate or Lewis lung cancer cells, tumour 1285 growth in bone is associated with induced sprouting of sensory nerve fibers and lytic bone lesions (23, 1286 162, 226, 351). SNS activation and bone pain (as judged by hyperalgesia and flinching) caused by 1287 Lewis lung cancer cells are substantially reduced in TRPV1-/- mice compared to wild-type animals with 1288 comparable tumor burden (351). Similarly, systemic administration of a neutralizing anti-NGF antibody 1289 to animals bearing breast or prostate cancer bone metastasis reduces ectopic nerve fiber sprouting and 1290 attenuates nociceptive behaviors (spontaneous guarding and flinching) (23,162, 226). Overall these 1291 findings suggest that targeting NGF and/or TRPV1 are potential strategies to treat bone pain.

1292

1293 Nerve cells and bone metastasis: current understandings & open questions

- Bone homeostasis is influenced by sympathetic, parasympathetic, and sensory nerves. These
 interactions are affected by numerous factors released by tumor cells.
- In model systems, stress-induced activation of the sympathetic nervous system alters the bone
 microenvironment to become more supportive of tumor outgrowth. However, there is no
 evidence that it also applies in human disease.
- 1299

1300 **C. Adipocytes**

The adipose content of the bone marrow increases with age, obesity levels and metabolic conditions (134, 230). A large body of research underpins the current view that bone marrow fat is a hormone-sensitive endocrine tissue with the capacity to modify bone mass, and hence could contribute to skeletal tumour growth through a range of mechanisms, including provision of energy and prosurvival factors for tumor cells (134). However, the majority of these studies are from model systems, often involving injection of large numbers of tumor cells directly into bone of immunocompromised mice fed a high fat diet. Due to suitable clinical material being difficult to obtain, the relevance of the information generated in model systems described in this section remains to be validated in studies ofhuman samples.

1310 Adipocytes can be drivers of chronic inflammation, resulting in immune cell infiltration and release 1311 of high levels of pro-inflammatory cytokines, including CXCL-1, CXCL-2, IL-1 β , IL-6 and TNF α , 1312 molecules known to stimulate bone resorption and bone metastasis (134, 135, 140, 151). Using an in 1313 vivo diet-induced obesity model. Herroon et al. (142) have demonstrated a direct link between 1314 adipocytes and prostate cancer growth in bone. Specifically, the intratibial injection of PC3 prostate 1315 cancer cells into high-fat-diet-fed mice led to larger tumors than those observed in mice on normal diet 1316 (142). In vitro, prostate cancer cells exposed to lipids supplied by bone marrow adipocytes displayed 1317 increased invasive and proliferative capacity compared to control, which was associated with induction 1318 of lipid chaperone FABP4 (fatty acid binding protein 4) and IL-1β in tumor cells (142). Although FABP4 1319 is known for its expression in adipocytes, it was also expressed by PC3 cells co-cultured with 1320 adipocytes, and its inhibition with a selective inhibitor (BMS309403) blocked PC3 cell invasion in vitro 1321 (142). Immunohistochemical staining showed that in the small number of human prostate cancer bone 1322 metastasis samples analysed (n=5), FABP4 positivity was more pronounced compared to benign 1323 prostate lesions and primary tumor tissues (142). The authors acknowledge that further studies are 1324 however required to establish whether FABP4 acts as a mediator between adipocytes and tumor cells 1325 to stimulate tumor growth in human bone metastases.

In addition to FABP4, the expression of oxidative stress enzyme HO-1 (heme oxygenase 1) was also found to be significantly upregulated in prostate cancer bone metastases from high-fat-diet-fed mice (141). *In vitro*, bone marrow adipocytes induced the upregulation of HO-1 in prostate cancer cells, whereas, *in vivo*, HO-1 overexpression in human prostate cancer cells promoted skeletal tumor growth and osteolysis (141). Importantly, a link to human prostate cancer was established by analysis of five datasets of patients with metastatic disease (n=89), showing significant upregulation of HO-1 in metastatic foci (including in bone) compared to primary tumors. HO-1 expression was identified by immunohistochemical staining in two samples from prostate cancer bone metastasis biopsies, providing
some limited support that HO-1 is associated with tumor progression in bone, which requires validation
in a larger sample set.

Bone marrow adipocytes also promote metabolic reprogramming of prostate cancer cells in bone towards a glycolytic phenotype (87) (*see section VII for further discussion*). Additional studies utilizing these diet-induced models of increased bone marrow adiposity demonstrated that the adipose-derived chemokines CXCL1 and CXCL2 cause accelerated osteoclastogenesis *in vitro*, leading to enhanced prostate cancer-associated bone degradation *in vivo* (135) (**Table 1**).

1341 Adipocytes have also been linked to bone metastasis in a number of cancer types other than 1342 prostate cancer. In multiple myeloma, bone marrow adipocytes support tumor cell proliferation and 1343 migration *in vitro* through mechanisms that are, at least in part, mediated by leptin (43). Furthermore, 1344 myeloma cells promote MSC differentiation into adipocytes at the expense of osteoblasts by inhibiting 1345 expression of the ubiquitin ligase MURF1, thereby suppressing osteoblast-mediated bone formation in 1346 tumor-bearing animals and in cells from patients with myeloma (208). In a model of breast cancer bone 1347 metastasis, using conditioned medium generated from cultured adult human bone fragments, migration 1348 of MDA-MB-231 cells was found to correlate with increasing levels of the adipokine leptin and IL-18 1349 (328). Direct co-cultures demonstrated high numbers of breast cancer cells associated with the marrow 1350 adipose tissue within the bone fragments, supporting that tumour cells colonise areas of bone with high 1351 adiposity (328). Similar findings were reported from studies of melanoma models of bone metastasis, 1352 showing that melanoma cells were located in close proximity to adipocytes when colonising bone (52, 1353 356). Furthermore, the intra-tibial injection of B16F10 melanoma cells resulted in larger tumors and 1354 increased osteoclast numbers in mice fed a high-fat diet, compared to that observed in mice fed a 1355 normal diet (52). In agreement with these findings, melanoma cell-adipocyte co-culture experiments 1356 showed an increase in pro-inflammatory and pro-osteoclastic production of cytokines and chemokines 1357 (IL-6, IL-1β, CXCL-1, CXCL-2, and CXCL-5) by melanoma cells (52). Wang and colleagues (356)

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1358 reported that following intra-cardiac injection of B16F10 melanoma cells into mice, regardless of diet, 1359 there was a transient, highly significant increase in bone marrow adipocity and serum leptin levels 1360 compared to that of age-matched controls. In vitro, conditioned medium from melanoma cells promoted 1361 differentiation of adipocytes; conversely melanoma cell proliferation was stimulated by exposure to 1362 adipocyte-conditioned medium. The authors conclude that adipocytes may contribute to the lytic bone 1363 disease caused by melanoma cells (356). However, as tumours grow and induce lytic bone disease the 1364 number of adipocytes is rapidly reduced (356). This decrease in the number of adipocytes may result 1365 from a lipolysis that fuels tumor cells with adipocyte-derived fatty acids, thereby promoting tumor growth 1366 (140).

1367 Collectively, the experimental studies described above represent the increasing volume of data 1368 providing a strong link between bone marrow adipocytes and the progression of bone metastasis, 1369 however evidence from human disease to support this remains surprisingly limited. A relatively small 1370 retrospective study of 2,731 patients with early breast cancer found no significant link between body 1371 mass index and the subsequent pattern of metastasis, but in agreement with other reports, obese and 1372 owerweight patients had significantly shorter survival compared to the normal weight group (375). As 1373 levels of obesity are rising, including in young people, there are concerns that we are facing an increase 1374 in almost all cancers, as well as the potential for more aggressive metastatic disease, including in bone, 1375 driven by some of the mechanisms described in this section.

1376

1377 Adipocytes and bone metastasis: current understandings & open questions

Bone marrow fat is a hormone-sensitive endocrine tissue with the potential to modify bone
 mass and to influence skeletal tumour growth through provision of energy and tumour cell
 survival factors, as well as through generation of a pro-inflammatory environment.

Solid evidence from clinical studies for a link between obesity and bone metastasis is lacking,
 this important area should be the focus of retrospective analyses of large datasets that include
 detailed information about patient BMI and metastatic sites.

1384

1385VIII.FUELING EXPANSION - REPROGRAMMING ENERGY METABOLISM TO1386FACILITATE BONE METASTASIS PROGRESSION

1387 Metastatic tumor cells colonizing distant organs must rewire their biology in order to grow in the 1388 colonized organ (1,133, 271, 291). Most cancer cells use glycolysis (an oxygen-independent metabolic 1389 pathway) for glucose metabolism even when oxygen is sufficient (133). This phenomenon is called "the 1390 Warburg effect" or "aerobic glycolysis". Consequently, glucose is utilized for ATP generation through 1391 lactate production and via the pentose phosphate pathway (PPP) for nucleotide synthesis that is 1392 essential for cell proliferation (Figure 7). Not only does the Warburg effect allow cells to maintain ATP 1393 levels but also reduce oxidative stress and generation of ROS, enabling cancer cells to survive at the 1394 metastatic site (291). The existence of these metabolic adaptation mechanisms in cancer cells has been 1395 observed in situ in patients with bone metastasis (262), as visualized by ¹⁸F-FDG-PET scanning of 1396 breast cancer bone metastases (Figure 8).

1397 In order to increase glucose uptake, cancer cells upregulate glucose transporters, notably glucose 1398 transporter 1 (GLUT1), phosphoglycerate kinase and lactate dehydrogenase A (133). These enzymes 1399 of the glycolytic pathway, including phosphoglycerate kinase and PPP-associated proteins, such as 6-1400 phosphogluconolactonase, were observed to be highly expressed in osteotropic breast cancer cells and 1401 bone metastases from patients with breast cancer (48). Moreover, osteotropic MDA-MB-231 breast 1402 cancer cells produce large amounts of lactate, compared to non-osteotropic ones (197). Lactate is 1403 released from MDA-MB-231 cells by monocarboxylate transporter 4 (MCT4) and uptaken by osteoclasts 1404 through the transporter MCT1, which then fuels their oxidative metabolism and promotes osteoclast-1405 mediated bone resorption (197) (Figure 7). These experimental findings are supported by 1406 immunohistochemical analysis of a small number of human breast cancer bone metastasis specimens

1407 (n = 4) showing a strong staining for MCT4 in tumor cells at the bone metastatic site (197). In prostate 1408 cancer bone metastasis, bone marrow adipocytes promote aerobic glycolysis in tumor cells in vitro and 1409 *in vivo* by up-regulating HIF-1 α (87). In turn, HIF-1 α stimulates the expression of proteins involved in 1410 glucose uptake, such as GLUT1, and glycolytic genes, such as phosphoglycerate kinase and lactate 1411 dehydrogenase A (87). Thus, aerobic glycolysis seems to be key in supporting skeletal tumor growth 1412 and osteolysis, at least experimentally. This metabolic adaptation system of cancer cells in bone also 1413 occurs in breast cancer patients with bone metastasis (262), as judged by FDG-PET (Figure 8). 1414 Although some clinical studies suggest that FDG has a higher sensitivity for detecting bone metastasis 1415 than primary lesions in prostate cancer, FDG remains however of limited use in this cancer type when 1416 compared to imaging agent ¹⁸F-fluorocholine (FCH), thereby suggesting prostate cancer cells also use 1417 nonglucose metabolic pathways to thrive in colonized organs (352).

1418 Beside aerobic glycolysis, cancer cells in bone can use additional sources of energy. In the 1419 previous section we have discussed the contribution of bone marrow adipocytes, which can provide free 1420 fatty acids as an energy source for tumor cell survival and growth. In addition, the cell membrane 1421 phospholipid choline can be abnormally metabolized and internalized in tumor cells overexpressing 1422 choline kinase (352). This abnormal regulation of the phospholipid metabolism has been observed in 1423 situ in cancer cells metastatic to bone (290, 348), as visualized by FCH-PET scanning of prostate 1424 cancer bone metastases (Figure 8). Autophagy could be another potential source of energy for tumor 1425 cells in bone (233). For example, the small-GTPase Rab5a and Runx2 stimulate autophagosome 1426 trafficking in human osteotropic metastatic breast cancer cells (224, 324). However, further studies are 1427 clearly needed to understand how autophagy facilitates bone metastasis formation. Of note, cancer 1428 cells transport a significant portion of glucose-derived pyruvate into mitochondria where it serves as an 1429 anaplerotic substrate to replenish tricarboxylic acid (TCA) cycle intermediates used for the biosynthesis 1430 of fatty acids and cholesterol as well as protein acetylation (1) (Figure 7). In this respect, there is 1431 experimental evidence that this mitochondrial metabolism can be used as a source of energy for prostate cancer cells in bone (331). High levels of cholesterol in human prostate cancer bone metastasis specimens were observed, compared to normal bone (331). In addition, immunohistochemistry shows intense staining of the low-density lipoprotein receptor and variable levels of the scavenger receptor class B type 1 and 3-hydroxy-3-methylglutaryl-coenzyme reductase in prostate cancer cells that are metastatic to bone, thereby indicating possibilities for influx and *de novo* synthesis of cholesterol (331).

Taken together, these studies provide evidence that aerobic glycolysis and/or abnormal phospholipid and mitochondrial metabolism in tumor cells may contribute to skeletal tumor burden and bone destruction *in vivo*.

1441

1442 Reprogramming energy metabolism: facts & open questions

1443	•	There is substantial evidence from model systems and human studies that energy metabolism
1444		is disrupted to favour glycolysis in breast cancer bone metastases. However, further work is
1445		required to validate the importance of aerobic glycolysis in other model systems of cancer and
1446		bone metastasis, and in patients with advanced cancer and bone metastasis other than breast
1447		cancer.

The importance of other mechanisms (autophagy, increased cholesterol synthesis) and the
 potential for therapeutic targeting of energy metabolism to inhibit bone metastasis needs to be
 established in model systems in order to determine the implications for human disease.

1451

1452IX.BLOCKING BONE DECONSTRUCTION - CURRENT THERAPIES FOR THE1453TREATMENT OF BONE METASTASIS

In general, the treatment of bone metastases is aimed at palliating morbidity associated with skeletal lesions. It cures only rarely (*e.g.*, in lymphoma) and treatment varies depending on the tumor type. The treatment of bone metastasis includes external beam radiotherapy, systemic therapy with

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1457 cytotoxic antineoplastic drugs (chemotherapy) and endocrine agents, targeted therapies and targeted 1458 radionucleotide therapy. In addition, orthopedic intervention may be necessary for impending 1459 pathological fractures. Optimal management of skeletal metastases requires a multimodality approach 1460 that involves the combined expertise of medical and radiation oncologists, interventional radiologists, 1461 nuclear medicine and orthopedic oncologists, general physicians, palliative medicine specialists and the 1462 symptom control team (69, 72). Treatment decisions depend on whether the bone disease is localized 1463 or widespread, the presence or absence of extra-skeletal metastases, and the nature of the underlying 1464 malignancy. Systemic therapy for bone metastases can be directed against the tumor cell to reduce 1465 both cell proliferation and, in consequence, the production of cytokines and growth factors influencing 1466 bone cell function. Alternatively, systemic treatment is directed toward blocking the effect of these 1467 substances on host cells. Chemotherapy, biologically targeted agents, and endocrine treatments have direct antitumor effects, whereas bone targeted agents (BTA) such as the bisphosphonates and 1468 1469 denosumab are effective by preventing host cells (primarily osteoclasts) from reacting to tumor 1470 products.

1471 In the past two decades BTA have become established as a valuable additional approach to the 1472 range of current treatments (60, 345). Biochemical data indicate that osteoclast-mediated bone 1473 resorption is of importance not only in osteolytic bone metastases such as in breast and lung cancer but 1474 also in prostate cancer osteoblastic lesions, with values of resorption markers in the latter at least as 1475 high as those seen in breast cancer and other solid tumors (*see section X for further discussion*). As a 1476 result, the osteoclast is a key therapeutic target for skeletal metastases irrespective of the tissue of 1477 origin.

BTA provide an additional treatment approach for the relief of bone pain across a range of tumor types, with effects that seem to be independent of the nature of the underlying tumor or radiographic appearance of metastases (367). Approved BTA for use in oncology include the bisphosphonates, the RANK ligand inhibitor denosumab and bone seeking radionuclides including radium-223, strontium-89 and samarium-153.

As our understanding of the signaling mechanisms between bone cells and tumor cells increases, several new, targeted agents have entered clinical development. These agents include inhibitors of cathepsin K and Src kinase (both key regulators of osteoclast function), mammalian target of rapamycin (mTOR) inhibitors such as everolimus, endothelin antagonists such as atrasentan, several agents targeting TGF beta and various anabolic agents including inhibitors of the WNT signaling pathway.

Below, we describe the progress and future directions of existing bone-targeted therapies and report emerging therapies that have arisen from advances in our understanding of the biology of bone metastases (**Figure 9**).

1491

1492 A. Inhibiting Bone Resorption by Targeting Osteoclasts

1493 **1**. Bisphosphonates

1494 The bisphosphonates are pyrophosphate analogs, characterized by a P-C-P-containing central 1495 structure rather than the P-O-P of pyrophosphate and a variable R' chain that determines the relative 1496 potency, adverse effects, and precise mechanism of action (58). The P-C-P backbone renders 1497 bisphosphonates resistant to phosphatase activity and promotes binding to the mineralized bone matrix. 1498 The absorption of oral bisphosphonates from the gut is poor, variable, and dramatically inhibited by food 1499 intake. After intravenous administration of a bisphosphonate, the kidney rapidly excretes approximately 1500 25% to 40% of the absorbed dose and the remainder binds avidly to exposed bone around resorbing 1501 osteoclasts, leading to high local concentrations of bisphosphonate in the resorption lacunae.

During bone resorption, bisphosphonates are internalized by the osteoclast, where they cause disruption of several biochemical processes involved in osteoclast function, ultimately leading to apoptotic cell death. Nitrogen-containing bisphosphonates inhibit farnesyl pyrophosphate (FPP) synthase within the mevalonate pathway that is responsible for events that catalyse post-translational

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modification of a number of proteins, including the small guanosine triphosphatases such as Ras and Rho (58). Bisphosphonates that do not contain nitrogen, such as clodronate, induce osteoclast apoptosis through the generation of cytotoxic adenosine triphosphate analogs. The biological half-life of bisphosphonates is long with effects after a single dose still detectable several years later (34, 58).

Based on the results of large randomized controlled trials (see below), BTA have become the standard of care for the treatment and prevention of skeletal complications associated with bone metastases in patients with solid tumors (60, 345). The primary end point of these studies was the influence of bone-targeted treatment on the number of patients experiencing SREs, as well as the time to the first SRE and the rate of SREs as determined by either a simple annual rate or more complex multiple event analysis techniques.

1516

1517 Bisphosphonates for metastatic bone disease

The greatest experience with BTAs from the use of bisphosphonates in the management of bone metastases from breast cancer, where the value of the agents is undisputed. Placebo controlled randomized trials have shown that both the oral agents, clodronate and ibandronate, and the intravenous formulations of pamidronate, ibandronate and zoledronic acid all have useful clinical efficacy.

1523 Pamidronate was the first intravenous bisphosphonate to be systematically evaluated and has 1524 clinically important efficacy on skeletal morbidity, quality of life, pain and analgesic use in patients with 1525 breast cancer (153, 207). Zoledronic acid is the most potent bisphosphonate available and has been the 1526 bisphosphonate of choice in most clinical settings and health care systems around the world for more 1527 than a decade. In a placebo-controlled trial of zoledronic acid, the percentage of patients with at least 1528 one SRE after one year was significantly reduced from 50% in the placebo group to 30% with zoledronic 1529 acid (P = .003) (179). In comparison with placebo, zoledronic acid also significantly delayed the time to 1530 first SRE and reduced the overall risk of SREs by 41% (179). Zoledronic acid is somewhat more effective than pamidronate and oral ibandronate in preventing SREs (9, 278). In a multiple event analysis, zoledronic acid reduced the risk of developing skeletal complications by an additional 20% compared with pamidronate (P = .025) (277). In a randomized comparison of oral ibandronate and intravenous zoledronic acid, the two agents had broadly similar activity although ibandronate did not meet the strict statistical criteria for non-inferiority defined in the study protocol (9).

Bisphosphonates have been shown to reduce bone pain and biochemical markers of bone resorption in patients with osteoblastic bone lesions that are associated with advanced prostate cancer. Despite somewhat disappointing preliminary results with other bisphosphonates, zoledronic acid was investigated in patients with CRPC and bone metastases and showed that the increased potency of this compound translated into improved clinical benefit (283). Treatment with zoledronic acid reduced the overall risk of skeletal complications by 36% and extended the time to first skeletal complication by more than 4 months. Bone pain was also reduced at all time points (283).

1543 The pathophysiology of bone metastases is broadly similar in all tumor types, and BTA could thus 1544 be expected to be of value in preventing skeletal morbidity across the range of tumors involving the 1545 skeleton, especially if metastatic bone disease was a patient's dominant site of disease. As part of the 1546 development program for zoledronic acid, a phase-III randomized, placebo-controlled trial was 1547 performed in patients with bone metastases from a wide range of solid tumours other than breast or 1548 prostate cancer; more than half of the persons recruited had lung cancer (279, 280). Zoledronic acid 1549 significantly reduced the proportion of patients with at least one SRE, almost doubled the time to the 1550 first SRE compared with placebo and reduced the overall risk for SRE(s) by about 30% compared with 1551 placebo.

There remains uncertainty regarding the most appropriate duration and schedule of treatment. Bone targeted therapy should certainly not be stopped following the development of a first skeletal related event whilst on treatment; this should not be considered a failure of treatment, as the trials demonstrate a significant reduction in second and subsequent complications with continued treatment.

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Several trials have investigated the schedule of bisphosphonate treatment and suggested that the efficacy of 3 monthly and monthly administration of zoledronic acid is similar. For example, the CALGB 70604 (Alliance) trial, randomized patients with bone metastases from a range of different primary tumor types to zoledronic acid on a monthly or three-monthly schedule from the outset of treatment for two years (145). This study demonstrated non-inferiority of less frequent administration; in both arms, 29% of patients developed \geq 1 SRE and suggests that three monthly administration of zoledronic acid is a reasonable choice for most patients (145).

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- 1564

1565 Bisphosphonates for prevention of cancer treatment induced bone loss

1566 There are now increasing numbers of long-term survivors from cancer who have received 1567 combination chemotherapy, radiotherapy, and hormonal cancer treatments. Many of these survivors are 1568 at increased risk of osteoporosis, largely because of the endocrine changes induced by treatment. 1569 Cancer treatment-induced bone loss (CTIBL) is a particularly important long-term problem for women 1570 with breast cancer and men receiving androgen deprivation therapy (ADT). For example, the fracture 1571 incidence in women with breast cancer on an aromatase inhibitor was found to be around 18-20% after 5 years follow-up suggesting that about one in five women on an aromatase inhibitor without a bone 1572 1573 protective agent will sustain a fracture (121).

More than half of cases of prostate cancer occur in men over the age of 70 and thus many men with prostate cancer are at risk of osteoporosis, exacerbated by the ADT many will receive as treatment for their cancer. ADT reduces serum concentrations of testosterone to less than 5% of normal level and estrogen to less than 20% of normal level with consequent adverse effects on bone turnover and an increase in fracture rate, as clearly demonstrated by large retrospective epidemiological studies (296). In addition, ADT affects muscle mass, making falls more likely.

1580 The effects of both bisphosphonates and denosumab (*see next section*) on CTIBL have been

1581 studied in multiple randomized clinical trials. These studies have used dosing regimens that are similar, 1582 but not necessarily identical to those used for the treatment of age-related osteoporosis. In breast 1583 cancer, zoledronic acid is the most comprehensively studied bisphosphonate. In premenopausal 1584 women, zoledronic acid (4 mg IV every 6 months) prevented the bone loss associated with ovarian 1585 function suppression (OFS) and tamoxifen or an aromatase inhibitor whereas in the control group 1586 reductions in BMD at 3 years were around 5% and 10% with OFS plus tamoxifen and anastrazole 1587 respectively (123). In postmenopausal women, three companion trials (Z-FAST, ZO-FAST, E-ZO-FAST) 1588 compared the efficacy of a similar dosing schedule of zoledronic acid given either in conjunction with 1589 initiation of an aromatase inhibitor (immediate group), or if required due to a decline in BMD during 1590 adjuvant aromatase inhibitor therapy to a T-score<-2.0 at any site or a non-traumatic fracture (delayed 1591 group). At 5 years all three trials reported similar results with 7-9% and 4-6% differences in lumbar spine 1592 and hip BMD respectively between the two treatment arms in favor of zoledronic acid (38, 62, 209). 1593 None of these studies were designed to show a significant difference in fracture incidence between the 1594 treatment arms. Nevertheless, the BMD effects are similar to those seen in trials performed in 1595 postmenopausal osteoporosis in which bisphosphonates confer a relative risk reduction (RRR) of 45% 1596 for vertebral fractures and approximately 16% RRR for non-vertebral fractures (94).

1597 Several other randomized clinical trials have investigated the efficacy of oral bisphosphonates for 1598 preventing CTIBL in breast cancer (127). The numbers of patients included in each study is somewhat 1599 less than for the zoledronic acid studies and thus, unlike for other forms of osteoporosis, the evidence 1600 for efficacy of oral bisphosphonates in this specific setting is less robust. Indirect cross trial comparisons 1601 suggest the increases in BMD with oral regimens are somewhat less than with zoledronic acid or 1602 denosumab, especially in younger women receiving ovarian function suppression (OFS) or experiencing 1603 chemotherapy induced menopause. Again, none of the trials with oral agents were designed to assess 1604 reliably the impact of oral bisphosphonates on fracture risk.

1605 Alendronate, risedronate, pamidronate, and zoledronic acid have all been shown to prevent loss

in BMD in patients with prostate cancer but the studies have been small and, while the preservation of
BMD would argue for a favorable effect on fractures, the magnitude of effect cannot be reliably
assessed (144).

Overall, current guidelines for preventing CTIBL suggest that patients having adjuvant endocrine treatment should be managed according to risk of fracture (60, 127). Patients with a T-score of greater than -2 and no additional risk factors for fracture are advised to exercise and receive calcium and vitamin D, with risks and BMD monitored every one-two years. If the T-score is less than -2, or there are two or more risk factors for fracture, patients should receive the same advice and supplements plus a bisphosphonate or denosumab (*see next section*). Guidelines recommend continuing anti-resorptive therapy for as long as the patient is receiving endocrine treatment.

1616

1617 Disease modifying effects of bisphosphonate treatments

1618 The potential benefits of bone-targeted treatments on the clinical course of breast cancer in terms 1619 of prevention of recurrence and death from breast cancer have been an area of intense study over the 1620 past 20 years. In breast cancer patients with no sign of distant metastases, but having a minimal 1621 residual disease in the bone marrow, CTIBL leads to the release of bone-derived growth factors from 1622 resorbed bone that, in turn, may activate DTCs from a dormant to a proliferative state and trigger bone 1623 relapses. Of note, adjuvant zoledronic acid treatment of patients with early breast cancer improves 1624 elimination of DTCs (307). Bisphosphonates exert, at least experimentally, a variety of direct and 1625 indirect anticancer activities (58, 312). Moreover, bisphosphonates, by decreasing bone resorption, may 1626 also make the bone microenvironment less hospitable for tumor cells, thereby explaining the elimination 1627 of DTCs. These findings suggested a greater role for the use of bisphosphonates than has previously been considered. Individual trials provided varying results that suggested benefits were restricted to 1628 1629 women who had low levels of reproductive hormones due to either natural age-related menopause or 1630 ovarian function suppression. This hypothesis was confirmed by the Early Breast Cancer Trialists'

1631 Collaborative Group (EBCTCG) meta-analysis of individual patient data from >18,000 breast cancer 1632 patients included in randomized trials of adjuvant bisphosphonates. The meta-analysis showed that 1633 adjuvant bisphosphonates (intravenous zoledronic acid, oral clodronate and oral ibandronate) only 1634 reduced breast cancer recurrences and breast cancer deaths in postmenopausal women (93). Overall, 1635 across all age and menopausal groups, despite a reduction in bone metastases, adjuvant use of a 1636 bisphosphonate had no significant effect on breast cancer recurrence (rate ratio (RR)=0.94) and the 1637 effect on breast cancer mortality, though statistically significant, was small (RR=0.91) (93). However, in 1638 postmenopausal women or those receiving ovarian suppression with goserelin, clinically important 1639 benefits were seen with statistically significant improvements in overall breast cancer recurrence 1640 (RR=0.86), distant recurrence at any site (RR=0.82), bone recurrence (RR=0.72) and breast cancer-1641 specific mortality (RR=0.82) (123). This equates to prevention of more that 1 in 6 breast cancer deaths 1642 at 10 years. Several international guidelines now recommend the use of adjuvant bisphosphonates in 1643 postmenopausal early breast cancer, especially for those at moderate to high for recurrence (86, 128).

1644 Understanding why the benefits of adjuvant bisphosphonates appear restricted to 1645 postmenopausal women is a priority area for further research. There does not appear to be a link 1646 between bone resorption rates and treatment efficacy (33). On the other hand, more detailed evaluation 1647 on the primary tumor may help identify patients who will benefit from an adjuvant bisphosphonate. For 1648 example, a study demonstrated that patients with a MAF negative tumour (79% of all patients), 1649 evaluated using a FISH assay for the transcription factor, had significantly improved survival at 10 years 1650 and a lower relapse rate with the use of adjuvant zoledronic acid (64). On the other hand, in the 21% of 1651 women with tumors that over-express MAF no benefits were seen in this subset of patients treated with 1652 an adjuvant bisphosphonate and, in younger patients, disease outcomes were significantly worse (64). 1653 For reasons that remain unclear, the disease modifying effects of bisphosphonates in

postmenopausal breast cancer have not been seen in men with prostate cancer treated with ADT. In therandomized controlled STAMPEDE trial, the addition of zoledronic acid alone or in combination with

docetaxel chemotherapy to ADT for men with advanced prostate cancer did not improve survival, despite extending the time to first skeletal complication (157). By contrast, docetaxel showed evidence of survival improvement when combined to ADT (157).

1659

1660 2. Anti-RANKL antibody: Denosumab

1661 Therapeutic candidates to inhibit RANK/RANKL interaction were developed. Fusion proteins were 1662 initially engineered. These are recombinant proteins comprising the Fc portion of human IgG1 fused with 1663 the N-terminal ligand binding cysteine-rich domain (CRD) of OPG (OPG-Fc/AMGN-007 and Fc-OPG) or the 1664 four extracellular CRDs of RANK (RANK-Fc) (183). Both Fc-OPG and RANK-Fc potently inhibit bone 1665 resorption in preclinical models of osteoporosis and of cancer and bone metastasis (183, 312). However, 1666 following repeat dosing of human RANK-Fc in primates, autoantibodies were detected (183). This 1667 highlighted the potential risk of an immune response against endogeneous RANK or OPG in patients, when 1668 using RANK-Fc or AMGN-007, respectively. An anti-RANKL antibody approach was therefore preferred, 1669 which led to the development of denosumab.

Denosumab is a fully human, synthetic antibody that binds to RANKL with high affinity, thereby preventing its interaction with RANK in a way similar to that of OPG (183). Denosumab is administered by subcutaneous injection. The biological half-life of denosumab is only weeks compared to the months or years seen with bisphosphonates (34). Rebound osteolysis may occur following discontinuation of denosumab with accelerated bone loss and, in a few patients, an increased incidence of vertebral fractures 12-36 months after treatment cessation (334).

1676

1677 Denosumab for metastatic bone disease

1678 Denosumab has been shown to be superior to zoledronic acid for the prevention of SREs from 1679 breast cancer. 2046 patients were randomly assigned to receive four weekly subcutaneous injections of 1680 denosumab (120 mg) or intravenous zoledronic acid (4 mg), with supplements of calcium and vitamin D. 1681 Denosumab was statistically superior to zoledronic acid in delaying the first SRE (315). Overall, 1682 denosumab treatment delayed the occurrence of all types of SREs. However, no differences in survival 1683 or investigator-reported disease progression were found between the two treatment groups.

Denosumab was investigated in patients with CRPC and bone metastases and showed that in a randomized trial versus zoledronic acid, this compound was statistically superior to the bisphosphonate in delaying the first SRE and the overall risk of SREs (105).

1687 Denosumab has also been studied in advanced solid tumors other than breast and prostate 1688 cancers. Non-inferiority to zoledronic acid was demonstrated with a trend to better control of skeletal 1689 morbidity (139).

1690

1691 Denosumab for prevention of cancer treatment induced bone loss

1692 Denosumab is the only agent to have a specific license for CTIBL following large randomised 1693 trials in postmenopausal women with breast cancer receiving an aromatase inhibitor and in men with 1694 prostate cancer receiving ADT (121, 305). In both studies, fracture incidence was the primary endpoint. 1695 The ABCSG-18 trial compared adjuvant denosumab (60 mg by subcutaneous injection given twice a 1696 year) with placebo (both with calcium and Vitamin D supplements) in 3425 postmenopausal women 1697 receiving adjuvant aromatase inhibitor treatment (121). Women treated with denosumab had a 50% 1698 (95% CI 39–65%, p<0.0001) risk reduction for any clinical fracture. The fracture risk reduction appeared 1699 to be irrespective of age and baseline BMD. Furthermore, the disease-free survival (secondary 1700 endpoint) was significantly improved in the denosumab group (HR=0.82, 95% Cl 0.69 - 0.98, p = 0.026) 1701 compared to placebo group (122). In a placebo-controlled trial of denosumab in 1468 men receiving 1702 ADT for non-metastatic prostate cancer, 36 months of denosumab treatment was associated with a 1703 significantly reduced incidence of new vertebral fractures (1.5% with denosumab vs. 3.9% with placebo; 1704 relative risk [RR] 0.38; 95% CI 0.19-0.78). BMD increased from baseline at all sites in the denosumab 1705 group, whereas it declined in the placebo group (305).

As aforementioned for bisphosphonates, current guidelines for preventing CTIBL recommend continuing anti-resorptive therapy for as long as the patient is receiving endocrine treatment. Patients treated with denosumab may need additional bone protection with a bisphosphonate when denosumab is discontinued to prevent rebound osteolysis and the increased risk of multiple vertebral fractures associated with treatment withdrawal (334).

- 1711
- 1712

Disease modifying effects of denosumab treatment

1713 The disease modifying effects of denosumab have also been assessed in early breast cancer but 1714 this agent, at least when given in the intensive schedule selected in the adjuvant D-CARE study, had no 1715 effect on disease recurrence in either pre- or postmenopausal women (63). The osteoporosis schedule 1716 of denosumab has a beneficial effect on the underlying disease, as observed in the ABCSG-18 study 1717 (121,122). However, no survival benefits have been yet seen and the agent is therefore only 1718 recommended for fracture prevention (122). Of note, the presence of RANK-positive CTCs in the 1719 bloodstream of metastatic breast cancer patients (n = 20/42) is associated with a better response to 1720 denosumab therapy with respect to time to first SRE [HR=0.25 (0.1 - 10.62), P = 0.0012)], compared to 1721 metastatic patients with RANK-negative CTCs (n = 22/42) (254). It would be interesting to determine if 1722 RANK expression in CTCs could help identify high-risk early-stage breast cancer patients who might 1723 benefit from adjuvant denosumab. Similarly, a *post-hoc* analysis of the D-CARE trial will be conducted 1724 to determine if the expression of transcription factor MAF in primary tumors will help clarify the potential 1725 anticancer mechanism of denosumab (122) (see section X-C for further discussion).

Denosumab may have some disease modifying effects in prostate cancer: 1432 men with nonmetastatic CRPC who were at high risk for bone metastasis by virtue of either a PSA of \geq 8.0 ng/mL and/or PSA doubling time \leq 10.0 months were randomized to receive monthly denosumab, 120 mg, or placebo in addition to continuation of ADT. Denosumab significantly increased bone metastasis-free survival by a median of 4.2 months compared with placebo and delayed the time to symptomatic first bone metastases (306). However, this effect on the disease process did not translate into an improvement in overall survival and, in light of the relatively high cumulative incidence of ONJ (4% at 3 years), the marginal benefits were not considered sufficient to change clinical practice.

1734 The RANK/RANKL pathway has been shown to play a crucial role in the initiation and 1735 progression of inherited breast cancer caused by mutation in the tumor-suppressor gene breast cancer 1736 1 (BRCA1) (269). BRCA1 mutation carriers have a greater propensity to generate cancer stem cells, 1737 whose expansion is RANKL/RANK dependent, and denosumab inhibits the expansion of these cancer 1738 stem cells in vitro (269). Therefore, these findings strongly suggest that targeting the RANK/RANKL 1739 pathway could be beneficial for the prevention of breast cancer in BRCA1 mutation carriers. Two pilot 1740 studies are currently evaluating the biological effects of denosumab on Ki67 proliferation index (primary 1741 endpoint) in normal breast and fallopian tube fimbrial tissues from BRCA1 and BRCA2 mutation carriers 1742 (ACTRN12614000694617 and ClinicalTrials.gov NCT03382574 studies). Furthermore, a randomized, 1743 double-blind, placebo-controlled, multi-center, international phase 3 study will determine if denosumab 1744 can prevent breast cancer development in women carrying a BRCA1 germline mutation (ABCSG-50, 1745 EudraCT number 2017-002505-35; estimated number of subjects to be enrolled in the study: 2,918).

1746

1747 *3. Novel antiresorptive agents*

1748 LGR4/RANKL and small-molecule RANKL inhibitors

The leucine-rich repeat-containing G-protein-coupled receptor 4 (LGR4) has recently been identified as a RANKL receptor that negatively regulates osteoclast differentiation (215). LGR4 competes with RANK to bind RANKL and suppresses canonical RANK signaling during osteoclast differentiation (215). A soluble LGR4 extracellular domain (ECD), which binds to RANKL, was examined in animal models of osteoporosis. LGR4-ECD notably increased bone mass and inhibits osteoclast differentiation *in vivo* (215). Interestingly, LGR4-ECD had little physiological effect on osteoclast
differentiation in normal mice, which suggests that LGR4-ECD could antagonize excessive RANKL in
benign and malignant osteoclast-related diseases with few side effects.

The efficacy of a small-molecule RANKL inhibitor (AS2676293) has been tested in an animal model of bone metastasis (238). Oral administration of AS2676293 to animals inhibits formation of osteolytic lesions caused by MDA-MB-231 breast cancer cells. *In vitro*, AS2676293 inhibits osteoclastogenesis.

1761 These antiresorptive agents are still at a preclinical stage of development.

1762 Cathepsin K inhibitors

1763 Cathepsin K is a lysosomal cysteine protease highly expressed in osteoclasts, which degrades 1764 collagen during bone resorption. Cathepsin K inhibitors (AFG-495, L-235) reduce bone destruction and 1765 skeletal tumor burden in animal models of breast cancer bone metastasis (92, 196), providing a direct 1766 proof for causal role of cathepsin K in bone metastasis formation. Additionally, metastatic tumor cells in 1767 bone express cathepsin K and AFG-495 dose-dependently inhibits breast cancer cell invasion in vitro, 1768 but not tumor growth in vivo. These results suggest that cathepsin K inhibitors in the treatment of bone 1769 metastasis could potentially have a dual effect, inhibiting both osteoclast-mediated bone resorption and, 1770 to a less extent, tumor burden (196). Interestingly, a phase II trial in women with breast cancer and bone 1771 metastases showed that cathepsin K inhibitor odanacatib (which is structurally related to L-235) 1772 successfully reduced circulating levels of bone resorption markers after 4 weeks of treatment (160). 1773 Similarly, a phase-II trial in postmenopausal women with osteoporosis showed that odanacatib therapy 1774 was effective at inhibiting bone resorption and increasing bone mineral density (186). A large phase III 1775 trial, Long-Term Odanacatib Fracture Trial (LOFT), enrolling 16,713 participants with osteoporosis from 387 centres was therefore conducted (24). However, development of the agent has been discontinued 1776 1777 due to possible cardiovascular adverse events. A phase III trial assessing the efficacy of odanacatib in reducing the risk of bone metastasis in women with breast cancer (ClinicalTrials.gov identifier
 NCT00692458) has also been withdrawn for undisclosed commercial reasons.

1780

1781 Mammalian target of rapamycin (mTOR) inhibitors

1782 In bone physiology, RANKL and M-CSF promote osteoclast survival by signalling through 1783 mTOR (19). Rapamycin and everolimus (a rapamycin analog) are both mTOR inhibitors that block 1784 osteoclast differentiation in vitro and suppress bone resorption in an animal model of bone loss caused 1785 by ovariectomy (19, 36). In addition to their antiresorptive effects, mTOR inhibitors exhibit anti-cancer 1786 effects. Everolimus and temsirolimus were the first mTOR inhibitors to be approved in the treatment of 1787 advanced breast and renal cell cancers. Interestingly, everolimus and temsirolimus inhibit skeletal tumor 1788 burden and osteolysis in animal models of bone metastasis caused by breast and renal cell carcinoma, 1789 respectively (36, 343). Everolimus combined with aromatase inhibitor exemestane has been approved 1790 for patients with advanced hormone receptor-positive/HER2-negative breast cancer who progress on 1791 prior nonsteroidal aromatase inhibitor therapy with either letrozole or anastrozole (BOLERO-2 study). In 1792 this study, the progression-free survival was significantly improved in the everolimus plus exemestane 1793 arm, compared to the placebo plus exemestane arm (11). Because exemestane is known to increase 1794 bone turnover, an exploratory analysis in the BOLERO-2 study has been conducted in patients with or 1795 without bone-only disease (120, 152). Compared to placebo plus exemestane, everolimus plus 1796 exemestane increased the median progression-free survival (5.3 and 12.9 months, respectively), 1797 regardless of bisphosphonate use and presence of bone metastases at baseline, indicating a 67% 1798 reduction in the risk of progression (120, 152). In addition, bone marker levels increased with 1799 exemestane monotherapy, but decreased when used in combination therapy with everolimus (120). 1800 Overall, these results suggest that everolimus plus exemestane decreases disease progression in the 1801 bone, probably by suppressing increased bone turnover observed with exemestane monotherapy in 1802 addition to the greater antitumor effects of the combination.

1803 Src non-receptor tyrosine kinase inhibitors

1804 Src is one of eleven members of a family of non-receptor tyrosine kinases that interact with 1805 several protein-tyrosine kinase receptors, G-protein-coupled receptors and integrins, which are 1806 expressed at the plasma membrane (281). c-Src plays multiple roles in regulating cell proliferation, 1807 survival, adhesion, migration, invasion, metastasis, and angiogenesis (281). Although Src is 1808 ubiguitously expressed in vertebrate cells, much higher protein levels are found in osteoclasts, platelets 1809 and neurons than most other cells. Of note, the most noticeable phenotype of Src knock-out mice is 1810 osteopetrosis (enhanced bone mass) as a result of osteoclast dysfunction (281). In fact, following 1811 integrin $\alpha v \beta 3$ activation, Src phosphorylation regulates the organization of osteoclast's actin 1812 cytoskeleton, which enables the osteoclast to attach and spread to bone and optimally resorb bone. 1813 Additionally, following RANK/RANKL interaction, Src activation triggers signaling through the 1814 PI3K/AKT/mTOR pathway and promotes osteoclast survival (312). As exemplified in experimental bone 1815 metastasis, the injection of cancer cells in Src knock-out mice shows that these animals are protected 1816 from tumor-associated bone destruction because Src-defective osteoclasts do not resorb bone (361). 1817 Thus, Src plays a central role in osteoclast function. In addition, elevated expression and activity of c-1818 Src have been reported in a variety of cancers. Three Src inhibitors (dasatinib, saracatinib and 1819 bosutinib) underwent clinical studies in patients with cancer and (bone) metastases (312). To date 1820 clinical development has yielded disappointing results in the setting of solid tumors and bone 1821 metastases.

1822

RON receptor tyrosine kinase inhibitor.

1823 RON is a receptor tyrosine kinase receptor expressed by osteoclasts (4). Tumor-derived MSP 1824 promotes the formation of osteolytic lesions in an animal model of breast cancer bone metastasis, 1825 whose extent is inhibited upon treatment of metastatic animals with RON tyrosine kinase inhibitor BMS-1826 777607/ASLAN002 (4). A phase-I trial in postmenopausal women with advanced cancer shows that 1827 BMS-777607/ASLAN002 reduced bone resorption after 4 weeks of treatment (ClinicalTrials.gov 1828 identifier NCT01721148).

1829 Dual c-MET and VEGFR2 receptor tyrosine kinase inhibitor (cabozantinib)

1830 Receptor tyrosine kinases c-MET and vascular endothelial growth factor (VEGF) receptor 2 1831 (VEGFR2) and their respective ligands, hepatocyte growth factor (HGF) and VEGF, are expressed by 1832 both osteoblasts and osteoclasts, enabling the activation of autocrine and paracrine HGF/c-MET and 1833 VEGF/VEGFR2 signaling pathways for the regulation of osteoblast and osteoclast activities (194). 1834 Additionally, c-MET and VEGFR signaling facilitates tumor progression through the activation of multiple 1835 pathways (PI3K/AKT/mTOR, SRC, MAPKinases) (194). Dual tyrosine kinase inhibitors that target both 1836 VEGFR2 and c-MET (cabozantinib and TAS-115) were therefore developed and investigated in pre-1837 clinical and clinical settings (194).

Cabozantinib inhibits human osteoclast differentiation and osteoclast-mediated bone resorption and increases OPG production by human osteoblasts *in vitro* (104). In animal models of prostate cancer, cabozantinib decreased skeletal tumor burden and formation of osteoblastic lesions, indicating that this compound suppresses bone metastasis formation, at least in part, through inhibition of bone remodeling (79, 192). Similarly, TAS-115 inhibits human PC-3 prostate cancer bone metastasis formation and suppresses bone destruction (360). The effect of cabozantinib was therefore investigated in the treatment of prostate cancer patients with bone metastasis.

In phase-II studies, cabozantinib treatment of metastatic CRPC patients with bone metastases showed a remarkable 68% rate of normalization of bone scans and suggested disease benefits with prolongation of progression-free survival (194). However in the phase-III trial COMET-1, comparing cabozantinib with prednisone in patients with metastatic CRPC and bone metastases following prior treatment with docetaxel and either abiraterone or enzalutamide, although cabozantinib improved progression-free survival and time to first SRE, it failed to improve overall survival, the primary end point of the trial (304). In the light of these data, further development of cabozantinib in prostate cancer has been halted and a second trial (COMET-2) comparing cabozantinib with mitoxantrone plus prednisone
in a similar patient population as COMET-1 closed prematurely. It is likely that the dramatic effects on
bone scan appearances seen in the initial studies reflected the direct effects of cabozantinib on bone
cell function and skeletal blood flow rather than effects on the underlying malignant disease.

Cabozantinib is approved for the treatment of patients with advanced renal cell carcinoma after previous antiangiogenic therapy on the basis of significant improvements in progression-free survival and overall survival when compared with everolimus (phase-3 METEOR trial). Pre-specified analyses of progression-free survival and overall survival were conducted in a sub-group of patients with bone metastasis from the METEOR trial (99). Compared to everolimus, cabozantinib treatment was associated with a significant improvement of progression-free survival and overall survival in patients with bone metastases, indicating it is a good treatment option for these patients.

1863 *miR-34a mimic (MRX34)*

1864 MiR-34a is a critical suppressor of osteoclastogenesis and bone resorption by directly targeting 1865 the pro-osteoclastic factor Tgif2 (transforming growth factor- β -induced factor 2) (181). Its expression is 1866 therefore downregulated during osteoclast differentiation. The pharmacological administration of a 1867 miR-34a mimic delivered in nanoparticles (whose aim is to replenish the lost miRNA expression) can 1868 attenuate bone metastases in animals bearing breast or skin tumours (181). A phase I, open-label, 1869 multicenter, dose-escalation study investigated the safety, pharmacokinetics and pharmacodynamics 1870 of a miR-34a mimic (MRX34) encapsulated in lipid nanoparticles, in patients with unresectable primary 1871 liver cancer or advanced or metastatic cancer with or without liver involvement or hematologic 1872 malignancies (15). However, the clinical trial was terminated prematurely due to cases of immune-1873 related serious adverse events (ClinicalTrials.gov identifier NCT01829971).

1874 BET inhibitor

1875 The bromodomain and extraterminal (BET) protein family (BRD2, BRD3, BRD4 and BRDT) is an 1876 important class of chromatin readers, regulating chromatin accessibility to transcription factors and RNA 1877 polymerase. JQ1, a thienotriazolo-1,4-diazapine that binds selectively to BET bromodomain proteins, 1878 inhibits osteoclast differentiation by interfering with BRD4-dependent RANKL activation of *NFATC1* 1879 transcription (185). Moreover, JQ1 inhibits bone resorption in experimental models of malignant 1880 osteolytic lesions and osteoporosis (12, 185). JQ1 is still at a preclinical stage of development.

1881 Dock5 inhibitor

Dock5 (Dedicator of cytokinesis 5), a guanine nucleotide exchange factor for the small GTPase Rac, participates to the formation of the sealing zone in osteoclasts. C21, a chemical inhibitor of Dock5, reduces osteoclast-mediated bone resorption *in vitro* and blocks bone destruction in a melanoma model of bone metastasis *in vivo* (349). C21 is still at a preclinical stage of development.

1886 Jagged/Notch inhibitor

In bone metastasis, tumor-derived Jagged1 activate Notch signaling in osteoclast precursors, promoting osteoclast differentiation and bone resorption (295). Tumor-derived Jaddged1 also engages Notch signaling in osteoblasts, stimulating IL-6 production. In turn, IL-6 secreted from osteoblasts stimulates tumor growth (295). Therefore, a fully human monoclonal antibody (15D11) against Jagged-1 has been developed. 15D11 inhibits bone metastasis formation in animals and sensitizes bone metastases to chemotherapy (390). 15D11 is still at a preclinical stage of development.

1893

1894 **B.** Promoting Bone Formation by Targeting Osteoblasts

1895 1. Agents blocking WNT inhibitors

Anti-DKK1 (BHQ880 and DKN-01) and anti-SOST (blosozumab, BPS804 and romosozumab) antibodies have been developed to block the inhibitory effect of Wnt antagonists DKK-1 and SOST on osteoblast-mediated bone formation (71, 170, 227, 395). BHQ880 and DKN-01 are in phase I/II clinical 1899 trials for patients with multiple myeloma and other solid tumors, such as cholangiocarcinoma, 1900 esophageal cancer and gastric cancer, whereas romosozumab, blosozumab and BPS804 are in phase 1901 II/III clinical trials for osteoporosis (170). In a phase III trial romosozumab decreased the risk of vertebral 1902 fractures in postmenopausal women with osteoporosis (36% lower risk with romosozumab than 1903 placebo) (71). Experimentally, an anti-SOST antibody decreased the extent of osteolytic lesions in a 1904 mouse model of breast cancer bone metastasis and multiple myeloma (227, 395). However, up to know, 1905 there is no clinical study investigating the effect of anti-SOST antibodies in cancer-induced bone 1906 diseases.

Of note, the inhibition of one of these two WNT inhibitors (DKK1 and SOST) may engender a compensatory response in order to return to a steady state (106). By contrast, a bispecific antibody targeting SOST and DKK-1 (Hetero-DS) leads to synergistic bone formation in rodents and non-human primates (106). Thus, Hetero-DS could have a valuable role in increasing bone mass and improving healing of lytic bone lesions associated with bone metastasis.

1912 2. Endothelin-1 receptor inhibitors

Preclinical studies have uncovered a prominent role for ET-1 in the formation of osteosclerostic lesions (276, 378). However, phase 3 trials of the inhibitors, atrasentan and zibotentan in combination with docetaxel failed to improve overall survival in patients with metastatic castration-resistant prostate cancer compared with docetaxel alone and this treatment approach seems unlikely to reach the clinic (47, 240).

1918 *3.* Androgen Inhibitors

Abiraterone acetate is an orally administered selective androgen biosynthesis inhibitor derived from the structure of pregnenolone. It is an irreversible inhibitor of cytochrome CYP17A, resulting in virtually undetectable serum and intratumoral androgen levels (244). Abiraterone was evaluated in chemotherapy-naïve and chemotherapy-treated men with metastatic castration-resistant prostate cancer (COU-AA-301 and COU-AA-302 trials). The data from these two phase-III trials showed that abiraterone treatment significantly improves overall survival and skeletal outcomes (delay of symptomatic progression and reduction of time to first SRE) (116). These benefits of abiraterone treatment on metastatic bone disease may not only be related to a systemic control of the disease but also associated with a direct effect in the bone. Indeed, abiraterone exhibits direct bone anabolic and anti-resorptive effects (156).

Another promising inhibitor is the androgen receptor antagonist enzalutamide. Enzalutamide was evaluated in chemotherapy-naïve and chemotherapy-treated men with metastatic castration-resistant prostate cancer patients (13, 116). In these two phase-III trials (AFFIRM and PREVAIL), enzalutamide significantly decreased the risk of death and improved skeletal outcomes (time to first SREs and radiographic progression, respectively).

However, despite approval of abiraterone and enzalutamide in metastatic castration-resistant prostate cancer, virtually all patients eventually acquire secondary resistance. One plausible explanation for resistance may involve the presence of the androgen-receptor isoform encoded by splice variant 7 (AR.V7), which lacks the ligand-binding domain, but remains constitutively active as a transcription factor (5).

1939 4. Activin A inhibitors

1940 Activin A binds to activin type IIA (ActRIIA) or type IIB (ActRIIB) receptors and induces the 1941 recruitment and phosphorylation of an activin type I receptor (ActRIB), which then phosphorylates 1942 Smad2 and Smad3 intracellular signaling proteins (312). The treatment of non-human primates with a 1943 soluble chimeric protein composed of the extracellular domain of ActRIIA fused to human IgG-Fc 1944 (sotatercept, formerly called ACE-011) increased bone volume by decreasing bone resorption and 1945 increasing bone formation (212). In animal models of multiple myeloma with osteolytic lesions, the 1946 treatment of mice with a soluble activin receptor type IIA fusion protein (ActRIIA.muFc) blocked bone 1947 destruction (51). Specifically, ActRIA.muFc treatment significantly stimulated osteoblastogenesis, 1948 prevented myeloma-induced suppression of bone formation, blocked the development of osteolytic

1949 bone lesions and increased survival (51). In the clinic, sotatercept improved bone mineral density and 1950 bone formation in multiple myeloma patients (312). These pre-clinical and clinical findings suggest that 1951 stimulating osteoblastic bone formation to facilitate bone repair might be an alternative or additional 1952 therapeutic approach to the use of antiresorptive agents to treat osteolytic lesions. Although higher 1953 serum levels of activin A were reported in breast or prostate cancer patients with bone metastases. 1954 compared with those of patients without bone metastases, there are currently no ongoing trials in breast 1955 or prostate cancer with bone metastases (312).

1956

1957 C. Targeting the Bone Matrix and the Microenvironment

1958 1. Bone targeted radiopharmaceuticals

1959 The therapeutic use of radioactive-labeled tracer molecules is currently an area of considerable 1960 interest and research. Targeted radiotherapy has potential advantages over external beam radiotherapy 1961 in that the radiation dose may be delivered more specifically to the tumor and normal tissues may 1962 partially be spared unnecessary irradiation (242). Theoretically, it should also be possible to administer 1963 high doses of radiation to the tumor on a recurrent basis if necessary.

1964 The α -emitting radiopharmaceutical ²²³radium dichloride (radium-223, a calcium-mimetic 1965 radioisotope) and the β-emitting radiopharmaceuticals 89 strontium (strontium-89, a calcium-mimetic 1966 radioisotope) and ethylene diamine tetramethylene phosphonate-153 samarium (samarium-153, a 1967 bisphosphonate-conjugated radioisotope) bind to bone mineral and preferentially to newly formed bone 1968 matrix, such as areas of osteosclerostic bone metastatic lesions (59, 312). These radiopharmaceuticals 1969 emit radiation causing DNA damage and cell death. Radium-223 almost exclusively produces alpha 1970 particles that produce a high-linear energy transfer (LET) with ultra-short penetration (< 100 µm; 2-10 1971 cell diameters) resulting in a highly localized antitumor effect on adjacent bone metastases while limiting 1972 damage to the surrounding normal tissue (242). In contrast to radium-223, strontium-89 and samarium-

1973 153 have a low-LET with a penetration range of 3 to 8 millimetres, which results in considerably more
1974 dose to normal tissues, notably the bone marrow and this limits the use of these agents and the ability
1975 to combine with other treatments (59, 312).

1976 Strontium-89 and samarium-153 are approved for palliation of bone pain (103), but only 1977 occasionally used and it is the bone-seeking, alpha particle emitting, radium-223 that is of most 1978 relevance to current practice. Radium-223 is now approved for the treatment of bone metastases from 1979 CRPC following a placebo-controlled randomized phase-III trial (ALSYMPCA) in which radium-223 1980 increased the survival of patients by 3.6 months and further reduced skeletal morbidity over and above 1981 a bisphosphonate (258). Treatment was well tolerated and improved guality of life with no significant 1982 long-term toxicities identified (258, 348). Radium-223 has subsequently been studied earlier in the 1983 course of metastatic prostate cancer and in combination with other therapies. However, a double-1984 blinded, placebo-controlled randomized phase-III trial (ERA 223; NCT02043678) investigating the 1985 efficacy and safety of radium-223 and abiraterone versus abiraterone alone in chemotherapy-naïve 1986 CRPC patients with bone metastases showed that the combination of radium-223 and abiraterone did 1987 not improve either disease or skeletal outcomes compared with abiraterone alone. Furthermore, more 1988 bone fractures were observed in the combined treatment arm, particularly in patients not receiving 1989 concomitant antiresorptive agents (zoledronic acid, denosumab). In breast cancer, experimental 1990 findings showed that radium-223 inhibits skeletal tumor burden and bone destruction in a mouse model 1991 of breast cancer bone metastasis (319). A phase IIa study was conducted in breast cancer patients with 1992 bone-dominant disease. Radium-223 induced metabolic changes, as judged by a 25% decrease of ¹⁸F-1993 fluorodeoxyglucose uptake in osteosclerostic lesions using positron emission tomography and 1994 computed tomography (59).

1995

1996 *2.* Agents targeting nerve- or bone-derived growth factors.

1997 The release of algogenic factors by cancer/stromal cells and osteoclasts can induce sensitization 1998 and activation of sensory fibers that innervate the bone. Bisphosphonates and denosumab have been approved for the treatment of bone pain (100). A phase II study evaluated the safety and efficacy of the anti-NGF antibody tanezumab in patients with painful bone metastases taking daily opioids (308). The data are encouraging and suggest that tanezumab treatment results in sustained analgesic improvements.

2003 The TGF β signaling pathway plays a critical and dual role in cancer progression. Several inhibitors 2004 of TGF β signaling, such as neutralizing antibodies, antisense oligonucleotides, and receptor kinase 2005 inhibitors, have been developed and shown to have inhibitory effects on bone metastases in animal 2006 models (40, 101).

2007

2009

2008 X. THE VALUE OF BONE TURNOVER BIOMARKERS IN BONE METASTASIS

As previously discussed in *section I-C*, clinical presentations of bone metastases are highly diverse, and many locations remain asymptomatic. Nowadays, plain radiography is insufficient to correctly identify bone metastases since more than 50% of an affected bone is required to be detected. As a consequence, bone metastases are often diagnosed at the time symptoms occur, increasing the risk of developing SREs, which significantly impair patients' quality of life (64).

2015 In adults, the bone mass is maintained by a continuous bone remodeling, which is a balance 2016 between osteoclast-mediated bone resorption and osteoblast-mediated bone formation (76). The 2017 turnover between resorption and formation leads to the release of bone-derived molecules that are 2018 amenable to measurement in blood and urine (350). Some of these molecules have been used as 2019 biochemical biomarkers of bone turnover, reflecting ongoing rates of bone formation or bone resorption 2020 (Table 2). Since cancer cells cause a distortion of bone turnover, several clinical studies examined 2021 whether variations in the expression levels of these bone biomarkers in blood or urine were associated 2022 with malignant bone disease progression (61, 90). However, at present, the high inter- and intra-2023 individual variability represents a limitation to the routine use of these biomarkers (90). Clinical 2024 applications of these biomarkers for the detection and monitoring of bone metastasis and response to 2025 antiresorptive therapies have recently been reviewed (90) and are outlined in the following sections.

2026

A. Bone Formation Markers

Biochemical markers of bone formation include bone alkaline phosphatase (BALP), which is an enzyme localized at the plasma membrane of osteoblasts, and procollagen I carboxyl-terminal and amino-terminal propeptides (PICP and PINP, respectively), which are cleaved during the processing of type I collagen (**Table 2**) (350). Their clinical applications in the management of malignant bone diseases are summarized below.

2033 Diagnosis of bone metastasis

2034 In breast and prostate cancer, serum concentrations of PINP were found significantly increased in 2035 patients with bone metastases (90). In a retrospective analysis, PINP was measured in the serum of 2036 prostate cancer patients with different clinical outcomes (N0/M0: no metastases, N1/M0: lymph node 2037 metastases only, and M1: bone metastases) (180). Increased PINP levels in the M1 group were 2038 detectable 8 months before the first positive bone scintigraph (180). PICP and BALP were also 2039 investigated in the prostate cancer setting. In particular, serum BALP concentrations significantly 2040 correlated with the extent of bone involvement (90). Furthermore, a meta-analysis including 19 trials and 2041 3,628 patients with solid tumors showed that serum BALP levels in patients with bone metastases were 2042 2.9-fold higher (P < 0.05) than in patients without bone lesions (91). Another meta-analysis in lung 2043 cancer including 16 trials and 1,720 patients with or without bone metastases showed that high 2044 concentrations of BALP were also associated with bone metastasis (155).

2045 Prognosis of bone metastasis

2046 BALP levels were assessed in patients with bone metastases from breast cancer (n = 1,648), 2047 castration-resistant prostate cancer (n = 643) and lung cancer and other solid tumors (n = 773) treated 2048 with a bisphosphonate (zoledronic acid or pamidronate). High serum levels of BALP at baseline and on-2049 study were associated with increased risks of SREs, disease progression and death in patients who did 2050 not receive bisphosphonate therapy (61, 90). Similar findings were reported in a retrospective analysis 2051 involving 5,543 patients who received zoledronic acid or denosumab for bone metastasis treatment 2052 (206). Furthermore, after 3 months of treatment with either denosumab or zoledronic acid, patients with 2053 serum BALP levels \geq median at month 3 had significantly reduced overall survival compared with those 2054 who had serum BALP levels < median (HR = 2.44; P < 0.0001) (206).

The prognosis value of PINP for bone metastasis was investigated in breast cancer (33). Specifically, PINP was measured at baseline in the serum from 872 patients from a large randomized trial of adjuvant zoledronic acid (AZURE) in early breast cancer (33). High baseline PINP was prognostic for future bone recurrence at any time (P < 0.006), but was not predictive for distant metastasis taken as a whole, demonstrating the bone metastasis specificity of PINP (33).

2060 Response to bone-targeted therapies

2061 The predictive value of BALP and PICP has been evaluated in castration-resistant prostate cancer 2062 patients with bone metastases (n = 778) treated on a placebo-controlled phase III trial of docetaxel with 2063 or without atrasentan (SWOG S0421) (187). High baseline serum levels of BALP and PICP were 2064 associated with poor overall survival (HR = 1.23; P < 0.001 and HR = 1.38; P < 0.001, respectively). 2065 Increasing BALP and PICP levels by week 9 of therapy were associated with a significantly increased 2066 risk of death (HR = 1.28; P < 0.001 and HR = 1.35; P < 0.001, respectively). For patients with the 2067 highest biomarker levels (upper 25th percentile), improved survival was observed in the atrasentan arm compared with placebo arm (HR = 0.65; P < 0.04 and HR = 0.61; P < 0.02 for BALP and PICP, 2068 2069 respectively).

B. Bone Resorption Markers

2072 Biochemical markers of bone resorption include (1) the carboxyterminal telopeptide of type I 2073 collagen (ICTP), which is a degradation product of mature type I collagen cleaved by MMP, (2) C- and 2074 N-telopeptides (CTX and NTX, respectively), which are proteolytic fragments generated by cathepsin K 2075 cleavage of type I collagen, (3) pyridinoline (PYD) and deoxypyridinoline (DPD), which are nonreducible 2076 pyridinium cross-links present in the mature form of type I collagen, and (4) tartrate resistant acid 2077 phosphatase 5b (TRACP), which is an osteoclast-derived enzyme (Table 2) (350). Additional potential 2078 bone resorption markers are RANKL and OPG, the RANK-L/OPG ratio being used to estimate the 2079 osteolysis rate, and miRNAs (Table 2), the latter being significantly upregulated in the serum of patients 2080 with osteoporotic fractures and breast cancer patients with osteolytic bone metastases (96, 294). BSP 2081 and osteopontin, which are osteoblast-derived bone matrix components, have been also investigated as 2082 potential bone markers associated with osteolysis (165). ICTP, TRACP, serum CTX, and urinary NTX 2083 are the most common resorption markers used in clinical practice. Clinical applications for these bone 2084 resorption markers in the management of malignant bone diseases are summarized below.

2085 Diagnosis of bone metastasis

Serum concentrations of TRACP were found increased in patients with bone metastases from breast cancer, but not lung cancer (90, 155). In prostate cancer, high NTX and CTX levels were associated with bone metastases (35). In lung cancer, a meta-analysis including 16 trials and 1,720 patients with or without bone metastasis showed that high concentrations of NTX and ICTP (but not CTX) were associated with bone metastasis (155).

2091 Increased serum levels of BSP and OPN have been associated with bone metastases from breast,
2092 lung and prostate cancer (165). However, these proteins are also expressed by tumor cells, suggesting
2093 they can be considered tumor markers rather than bone biomarkers (165).

Increased serum levels of RANKL and/or OPG have been associated with bone metastases from prostate cancer (165). Similarly, the RANKL/OPG ratio is increased in severe osteolysis associated with primary bone tumors and bone metastasis from lung, renal and breast cancer (125). However, the low sensitivity of the assays to measure circulating levels of RANKL and OPG and the observation that RANK, RANKL and OPG are expressed in a wide variety of different cell types including tumors cells have so far limited the routine measurement of these molecules (165, 183).

2100 Circulating miRNAs originating from tissues, being remarkably stable in blood, may be able to 2101 serve as biomarkers. For example, Seeliger et al. (294) identified 5 miRNAs (miR-21, miR-23a, miR-25, 2102 miR-100 and miR-125b) that were upregulated in both the serum and the bone tissue of osteoporotic 2103 patients with bone fractures. Although these miRNAs are not bone tissue-specific, they have been 2104 reported to play a role in bone remodeling when they are expressed by osteoblasts (294). Similarly, Ell 2105 et al. (96) identified a series of 4 miRNAs (miR-16, miR-211, miR-378 and Let-7a) that were specifically 2106 upregulated during osteoclast differentiation. The authors then thought to investigate this series of 2107 miRNAs as potential biomarkers for osteolytic bone metastases. They found that miR-16 and miR-378 2108 were consistently increased in the serum from breast cancer patients with bone metastases (n = 38), 2109 compared to healthy female donors (n = 21) (96).

2110 Prognosis of bone metastasis

NTX levels were assessed in patients with bone metastases from breast cancer (n = 1,648), castration-resistant prostate cancer (n = 643) and lung cancer and other solid tumors (n = 773) treated with a bisphosphonate (zoledronic acid or pamidronate). High serum levels of NTX at baseline and onstudy were associated with increased risks of SREs, disease progression and death in patients who did not receive bisphosphonate therapy (61, 90). Similar findings were reported in a retrospective analysis involving 5,543 patients who received zoledronic acid or anti-RANKL antibody denosumab for bone metastasis treatment (206). Furthermore, after 3 months of treatment with either denosumab or 2118 zoledronic acid, patients with urinary NTX levels \geq median at month 3 had significantly reduced overall

survival compared with those who had urinary NTX levels < median (HR = 1.85; P < 0.0001) (206).

The prognosis value of CTX and ICTP for bone metastasis was investigated in breast cancer (33). These bone resorption markers were measured at baseline in the serum from 872 patients from a large randomized trial of adjuvant zoledronic acid (AZURE) in early breast cancer (33). High baseline CTX or ICTP was prognostic for future bone recurrence at any time (P < 0.009 and 0.008, respectively), but were not predictive for overall distant recurrence, demonstrating the bone metastasis specificity of CTX and ICTP (33).

2126 Response to bone-targeted therapies

The value of measuring NTX levels to assess response to bisphosphonate therapy was investigated by exploring databases from phase III trials of zoledronic acid in solid tumors and multiple myeloma. The analysis revealed that patients with high NTX levels at baseline that normalize during zoledronic acid treatment have improved survival as compared to patients with persistent elevated NTX levels (61).

2132 The predictive value of NTX and PYD has been evaluated in castration-resistant prostate cancer 2133 patients with bone metastases (n = 778) treated on a placebo-controlled phase III trial of docetaxel with 2134 or without atrasentan (SWOG S0421) (187). As for bone formation markers BALP and PICP, high 2135 baseline levels of NTX and PYD were associated with poor overall survival (HR = 1.40: P < 0.001 and 2136 HR = 1.52; P < 0.001, respectively). Increasing bone resorption marker levels by week 9 of therapy 2137 were associated with a significantly increased risk of death (HR = 1.36; P = 0.002 and HR = 1.36; P = 2138 0.002 for NTX and PYD, respectively). In contrast to what was observed for patients with the highest 2139 bone formation marker levels, there was however no survival benefit from atrasentan when using NTX 2140 or PYD in the upper 25th percentile. Nonetheless, when combining all four biomarkers in the highest 2141 quartile, there was clear evidence that patients had a survival benefit from atrasentan (HR = 0.33; 2142 median survival = 13 [atrasentan] vs 5 months [placebo]; P = .005) (187).

2144 **C.** Insights from markers not associated with bone turnover

2145 Westbrook and colleagues (365) have identified two proteins [macrophage-capping protein (CAPG) 2146 and PDZ domain-containing protein (GIPC1)] from proteomic analysis of osteotropic human breast 2147 cancer cell lines whose expression in tumor cells was subsequently validated by immunohistochemistry 2148 using tumor tissue microarrays (TMAs) from breast cancer patients (n = 364) of the AZURE trial. Clinical 2149 validation of these two markers showed that patients who did not receive a bisphosphonate therapy 2150 were more likely to develop a first distant recurrence in bone (HR = 4.5; P < 0.001) and die (HR = 1.8; P 2151 = 0.045) if CAPG and GIPC1 were highly expressed in the primary tumor. Moreover, patients with high 2152 expression of CAPG and GIPC1 had a 10-fold increase in treatment benefit, compared with patients on 2153 standard therapy (365).

2154 Dedicator of cytokinesis protein 4 (DOCK4) is another protein specifically expressed in osteotropic 2155 tumor cells (366). DOCK4 expression in primary tumors was validated by immunohistochemistry, using 2156 TMAs from breast cancer patients (n = 345) of the AZURE trial (366). Adjusted Cox regression analyses 2157 showed that patients who did not receive a bisphosphonate therapy were more likely to develop a first 2158 distant recurrence in bone (HR = 2.13; P = 0.034) if DOCK4 was highly expressed in the primary tumor 2159 (366).

2160 MAF is a transcription factor of the AP-1 family shown to mediate breast cancer bone metastasis 2161 (260). The value of MAF expression in primary tumors to predict the treatment outcomes of adjuvant 2162 zoledronic acid in breast cancer patients from the AZURE trial (n = 1,739) has been investigated (64). In 2163 patients with MAF-negative tumors (79% of all patients), there was a lower relapse rate with the use of 2164 zoledronic acid (HR = 0.74), but not in patients who had MAF-positive tumors (64). Additionally, MAF 2165 positivity was associated with increased extraskeletal recurrence in the zoledronic acid group (HR = 2166 6.92) (64). Data from ABCSG-18 trial are also being used in a *post-hoc* analysis addressing MAF to 2167 help clarify the anticancer mechanism of denosumab (122).

2169 XI. CONCLUSION

2170 Bone is one of the most common sites for metastasis, especially from breast, prostate and lung 2171 cancer. These skeletal metastases contribute substantially to morbidity and mortality in patients with 2172 advanced cancer. It is therefore essential to better understand the pathophysiology of bone metastasis 2173 in order to improve therapies for the treatment and prevention of bone metastasis and predict the risk of 2174 disease relapse. In this review, we described the importance of the systemic effect of primary tumors in 2175 preparing a pre-metastatic niche to facilitate the arrival of tumor cells in the bone marrow and we 2176 highlighted the prominence of metastatic niches in mediating dormancy of tumor cells. We also 2177 discussed the key role of the environment for reactivation of dormant tumor cells, which then undergo 2178 further selection to acquire a full complement of metastasis-colonization functions that dormant tumor 2179 cells did not express before. We also explained how, at a later stage, tumor cells induce osteolytic or 2180 osteoblastic lesions. These findings provide the rationale for the use of bone-targeted agents such as 2181 the bisphosphonates, the RANK ligand inhibitor denosumab and bone seeking radiopharmaceuticals. 2182 However, as our understanding of the signaling mechanisms between tumor cells and cells in the bone 2183 marrow microenvironment increases, several new, targeted agents have entered clinical development. 2184 They could be used in combination with anti-resorptive agents to efficiently block the development of 2185 skeletal lesions. Another attractive avenue of research would be to reconstruct bone lesions by restoring 2186 osteoblast anabolic functions. Finally, we showed that bone markers potentially provide important 2187 insight for predicting the risk of disease relapse in patients with cancer and evaluating a patient's risk of 2188 worsening skeletal health.

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2194

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2203 XII. REFERENCES

- Ahn CS, Metallo CM. Mitochondria as biosynthetic factories for cancer proliferation. *Cancer Metab.* 3:1, 2015. doi: 10.1186/s40170-015-0128-2.
- Alam AS, Gallagher A, Shankar V, Ghatei MA, Datta HK, Huang CL, Moonga BS, Chambers
 TJ, Bloom SR, Zaidi M. Endothelin inhibits osteoclastic bone resorption by a direct effect on
 cell motility: implications for the vascular control of bone resorption. *Endocrinology*. 130:3617 3624, 1992.
- 2210 3. Alexaki VI, Javelaud D, Van Kempen LC, Mohammad KS, Dennler S, Luciani F, Hoek KS,
- 2211 Juàrez P, Goydos JS, Fournier PJ, Sibon C, Bertolotto C, Verrecchia F, Saule S, Delmas V,
- 2212 Ballotti R, Larue L, Saiag P, Guise TA, Mauviel A. GLI2-mediated melanoma invasion and 2213 metastasis. *J Natl Cancer Inst.* 102:1148-1159, 2010. doi: 10.1093/jnci/djg257.
- Andrade K, Fornetti J, Zhao L, Miller SC, Randall RL, Anderson N, Waltz SE, McHale M, Welm
 AL. RON kinase: A target for treatment of cancer-induced bone destruction and osteoporosis.
 Sci Transl Med. 9. pii: eaai9338, 2017. doi: 10.1126/scitranslmed.aai9338.
- 2217 5. Antonarakis ES, Lu C, Luber B, Wang H, Chen Y, Zhu Y, Silberstein JL, Taylor MN, Maughan
- 2218 BL, Denmeade SR, Pienta KJ, Paller CJ, Carducci MA, Eisenberger MA, Luo J. Clinical
- 2219 Significance of Androgen Receptor Splice Variant-7 mRNA Detection in Circulating Tumor
- 2220 Cells of Men With Metastatic Castration-Resistant Prostate Cancer Treated With First- and
- 2221 Second-Line Abiraterone and Enzalutamide. *J Clin Oncol.* 35:2149-2156, 2017. doi:
 2222 10.1200/JCO.2016.70.1961.
- Atkinson EG, Delgado-Calle J. The Emerging Role of Osteocytes in Cancer in Bone. *JBMR Plus* 3:e10186, 2019. doi: 10.1002/jbm4.10186.
- Barker HE, Cox TR, Erler JT. The rationale for targeting the LOX family in cancer. *Nat Rev Cancer* 12: 540-552, 2012. doi: 10.1038/nrc3319.

- Baron R, Kneissel M. WNT signaling in bone homeostasis and disease: from human mutations
 to treatments. *Nat Med.* 19:179-192, 2013. doi: 10.1038/nm.3074.
- Barrett-Lee P, Casbard A, Abraham J, Hood K, Coleman R, Simmonds P, Timmins H,
 Wheatley D, Grieve R, Griffiths G, Murray N. Oral ibandronic acid versus intravenous
 zoledronic acid in treatment of bone metastases from breast cancer: a randomised, open label,
 non-inferiority phase 3 trial. *Lancet Oncol.* 15:114-122, 2014. doi: 10.1016/S1470 2045(13)70539-4.
- Barry-Hamilton V, Spangler R, Marshall D, McCauley S, Rodriguez HM, Oyasu M, Mikels A,
 Vaysberg M, Ghermazien H, Wai C, Garcia CA, Velayo AC, Jorgensen B, Biermann D, Tsai D,
 Green J, Zaffryar-Eilot S, Holzer A, Ogg S, Thai D, Neufeld G, Van Vlasselaer P, Smith V.
 Allosteric inhibition of lysyl oxidase-like-2 impedes the development of a pathologic
 microenvironment. *Nat Med* 16: 1009-1017, 2010. doi: 10.1038/nm.2208.
- Baselga J, Campone M, Piccart M, Burris HA 3rd, Rugo HS, Sahmoud T, Noguchi S, Gnant M,
 Pritchard KI, Lebrun F, Beck JT, Ito Y, Yardley D, Deleu I, Perez A, Bachelot T, Vittori L, Xu Z,
 Mukhopadhyay P, Lebwohl D, Hortobagyi GN. Everolimus in postmenopausal hormonereceptor-positive advanced breast cancer. *N Engl J Med.* 366:520-529, 2012. doi:
 10.1056/NEJMoa1109653.
- Baud'huin M, Lamoureux F, Jacques C, Rodriguez Calleja L, Quillard T, Charrier C, Amiaud J,
 Berreur M, Brounais-LeRoyer B, Owen R, Reilly GC, Bradner JE, Heymann D, Ory B. Inhibition
 of BET proteins and epigenetic signaling as a potential treatment for osteoporosis. *Bone*.
 94:10-21, 2017. doi: 10.1016/j.bone.2016.09.020.
- Beer TM, Armstrong AJ, Rathkopf DE, Loriot Y, Sternberg CN, Higano CS, Iversen P,
 Bhattacharya S, Carles J, Chowdhury S, Davis ID, de Bono JS, Evans CP, Fizazi K, Joshua
 AM, Kim CS, Kimura G, Mainwaring P, Mansbach H, Miller K, Noonberg SB, Perabo F, Phung
 D, Saad F, Scher HI, Taplin ME, Venner PM, Tombal B; PREVAIL Investigators. Enzalutamide

- in metastatic prostate cancer before chemotherapy. *N Engl J Med.* 371:424-433, 2014. doi:
 10.1056/NEJMoa1405095.
- Beermann J, Piccoli MT, Viereck J, Thum T. Non-coding RNAs in development and disease:
 background, mechanisms, and therapeutic approaches. *Physiol Rev* 96: 1297-1325, 2016. doi:
 10.1152/physrev.00041.2015.
- Beg MS, Brenner AJ, Sachdev J, Borad M, Kang YK, Stoudemire J, Smith S, Bader AG, Kim
 S, Hong DS. Phase I study of MRX34, a liposomal miR-34a mimic, administered twice weekly
 in patients with advanced solid tumors. *Invest New Drugs.* 35:180-188, 2017. doi:
 10.1007/s10637-016-0407-y.
- Bellahcène A, Bachelier R, Detry C, Lidereau R, Clézardin P, Castronovo V. Transcriptome
 analysis reveals an osteoblast-like phenotype for human osteotropic breast cancer cells.
 Breast Cancer Res Treat 101: 135-148, 2007. doi: 10.1007/s10549-006-9279-8.
- 17. Bellahcène A, Castronovo V, Ogbureke KU, Fisher LW, Fedarko NS. Small integrin-binding
 ligand N-linked glycoproteins (SIBLINGs): multifunctional proteins in cancer. *Nat Rev Cancer*8: 212-226, 2008. doi: 10.1038/nrc2345.
- Bendre MS, Gaddy-Kurten D, Mon-Foote T, Akel NS, Skinner RA, Nicholas RW, Suva LJ.
 Expression of interleukin 8 and not parathyroid hormone-related protein by human breast
 cancer cells correlates with bone metastasis in vivo. *Cancer Res.* 62: 5571-5579, 2002.
- Bertoldo F, Silvestris F, Ibrahim T, Cognetti F, Generali D, Ripamonti CI, Amadori D, Colleoni
 MA, Conte P, Del Mastro L, De Placido S, Ortega C, Santini D. Targeting bone metastatic
 cancer: Role of the mTOR pathway. *Biochim Biophys Acta* 1845:248-254, 2014. doi:
 10.1016/j.bbcan.2014.01.009.
- 2274 20. Bidwell BN, Slaney CY, Withana NP, Forster S, Cao Y, Loi S, Andrews D, Mikeska T, Mangan
 2275 NE, Samarajiwa SA, de Weerd NA, Gould J, Argani P, Möller A, Smyth MJ, Anderson RL,

Hertzog PJ, Parker BS. Silencing of Irf7 pathways in breast cancer cells promotes bone metastasis through immune escape. *Nat Med.* 18:1224-1231, 2012. doi: 10.1038/nm.2830.

- 2278 21. Blake ML, Tometsko M, Miller R, Jones JC, Dougall WC. RANK expression on breast cancer
 2279 cells promotes skeletal metastasis. *Clin Exp Metastasis* 31:233-245, 2014. doi:
 2280 10.1007/s10585-013-9624-3.
- Bliss SA, Sinha G, Sandiford OA, Williams LM, Engelberth DJ, Guiro K, Isenalumhe LL, Greco
 SJ, Ayer S, Bryan M, Kumar R, Ponzio NM, Rameshwar P. Mesenchymal Stem Cell-Derived
 Exosomes Stimulate Cycling Quiescence and Early Breast Cancer Dormancy in Bone Marrow.
 Cancer Res 76: 5832-5844, 2016. doi: 10.1158/0008-5472.CAN-16-1092.
- 2285 23. Bloom AP, Jimenez-Andrade JM, Taylor RN, Castañeda-Corral G, Kaczmarska MJ, Freeman
 KT, Coughlin KA, Ghilardi JR, Kuskowski MA, Mantyh PW. Breast cancer-induced bone
 remodeling, skeletal pain, and sprouting of sensory nerve fibers. *J Pain.* 12:698-711, 2011. doi:
 10.1016/j.jpain.2010.12.016.
- 2289 24. Bone HG, Dempster DW, Eisman JA, Greenspan SL, McClung MR, Nakamura T, Papapoulos
 S, Shih WJ, Rybak-Feiglin A, Santora AC, Verbruggen N, Leung AT, Lombardi A. Odanacatib
 for the treatment of postmenopausal osteoporosis: development history and design and
 participant characteristics of LOFT, the Long-Term Odanacatib Fracture Trial. *Osteoporos Int.*2293 26:699-712, 2015. doi: 10.1007/s00198-014-2944-6.
- 2294 25. Bottos A, Gotthardt D, Gill JW, Gattelli A, Frei A, Tzankov A, Sexl V, Wodnar-Filipowicz A,
 Hynes NE. Decreased NK-cell tumour immunosurveillance consequent to JAK inhibition
 enhances metastasis in breast cancer models. *Nat Commun.* 7:12258, 2016. doi:
 10.1038/ncomms12258.
- 2298 26. Boucharaba A, Serre CM, Grès S, Saulnier-Blache JS, Bordet JC, Guglielmi J, Clézardin P,
 2299 Peyruchaud O. Platelet-derived lysophosphatidic acid supports the progression of osteolytic
 2300 bone metastases in breast cancer. *J Clin Invest* 114; 1714-1725, 2004. doi: 10.1172/JCl22123.

2301 27. Boucharaba A, Serre CM, Guglielmi J, Bordet JC, Clézardin P, Peyruchaud O. The type 1
2302 lysophosphatidic acid receptor is a target for therapy in bone metastases. *Proc Natl Acad Sci*2303 USA 103; 9643-9648, 2006. doi: 10.1073/pnas.0600979103.

- 2304 28. Bouchet M, Lainé A, Boyault C, Proponnet-Guerault M, Meugnier E, Bouazza L, Kan CWS,
 2305 Geraci S, El-Moghrabi S, Hernandez-Vargas H, Benetollo C, Yoshiko Y, Duterque-Coquillaud
 2306 M, Clézardin P, Marie JC, Bonnelye E. ERRα Expression in Bone Metastases Leads to an
 2307 Exacerbated Antitumor Immune Response. *Cancer Res* 80; 2914-2926, 2020. doi:
 2308 10.1158/0008-5472.CAN-19-3584.
- 2309 29. Boudot C, Hénaut L, Thiem U, Geraci S, Galante M, Saldanha P, Saidak Z, Six I, Clézardin P,
 2310 Kamel S, Mentaverri R. Overexpression of a functional calcium-sensing receptor dramatically
 2311 increases osteolytic potential of MDA-MB-231 cells in a mouse model of bone metastasis
 2312 through epiregulin-mediated osteoprotegerin downregulation. *Oncotarget* 8:56460-56472,
 2313 2017. doi: 10.18632/oncotarget.16999.
- 2314 30. Bozec A, Zaiss MM. T regulatory cells in bone remodelling. *Curr Osteoporos Rep* 15:121-125,
 2017. doi: 10.1007/s11914-017-0356-1.
- 31. Bragado P, Estrada Y, Parikh F, Krause S, Capobianco C, Farina HG, Schewe DM, AguirreGhiso JA. TGF-β2 dictates disseminated tumour cell fate in target organs through TGF-β-RIII
 and p38α/β signalling. *Nat Cell Biol* 15: 1351-1361, 2013. doi: 10.1038/ncb2861.
- 2319 32. Braun S, Vogl FD, Naume B, Janni W, Osborne MP, Coombes RC, Schlimok G, Diel IJ,
- 2320 Gerber B, Gebauer G, Pierga JY, Marth C, Oruzio D, Wiedswang G, Solomayer EF, Kundt G,
- Strobl B, Fehm T, Wong GY, Bliss J, Vincent-Salomon A, Pantel K. A pooled analysis of bone
 marrow micrometastasis in breast cancer. *N Engl J Med.* 353:793-802, 2005.
 doi:10.1056/NEJMoa050434.
- 33. Brown J, Rathbone E, Hinsley S, Gregory W, Gossiel F, Marshall H, Burkinshaw R, Shulver H,
 Thandar H, Bertelli G, Maccon K, Bowman A, Hanby A, Bell R, Cameron D, Coleman R.

- Associations Between Serum Bone Biomarkers in Early Breast Cancer and Development of Bone Metastasis: Results From the AZURE (BIG01/04) Trial. *J Natl Cancer Inst.* 110:871-879, 2018. doi: 10.1093/jnci/djx280.
- Brown JE, Coleman RE. Denosumab in patients with cancer-a surgical strike against the
 osteoclast. *Nat Rev Clin Oncol.* 9:110-118, 2012. doi: 10.1038/nrclinonc.2011.197.
- 35. Brown JE, Cook RJ, Major P, Lipton A, Saad F, Smith M, Lee KA, Zheng M, Hei YJ, Coleman
 RE. Bone turnover markers as predictors of skeletal complications in prostate cancer, lung
 cancer, and other solid tumors. *J Natl Cancer Inst.* 97:59-69, 2005. DOI: 10.1093/jnci/dji002.
- 36. Browne AJ, Kubasch ML, Göbel A, Hadji P, Chen D, Rauner M, Stölzel F, Hofbauer LC,
 Rachner TD. Concurrent antitumor and bone-protective effects of everolimus in osteotropic
 breast cancer. *Breast Cancer Res.* 19:92, 2017. doi: 10.1186/s13058-017-0885-7.
- 37. Browne G, Taipaleenmäki H, Stein GS, Stein JL, Lian JB. MicroRNAs in the control of
 metastatic bone disease. *Trends Endocrinol Metab* 25: 320-327, 2014. doi:
 10.1016/j.tem.2014.03.014.
- 38. Brufsky AM, Harker WG, Beck JT, Bosserman L, Vogel C, Seidler C, Jin L, Warsi G, ArgonzaAviles E, Hohneker J, Ericson SG, Perez EA. Final 5-year results of Z-FAST trial: adjuvant
 zoledronic acid maintains bone mass in postmenopausal breast cancer patients receiving
 letrozole. *Cancer.* 118:1192-1201, 2012. doi: 10.1002/cncr.26313.
- Bu G, Lu W, Liu CC, Selander K, Yoneda T, Hall C, Keller ET, Li Y. Breast cancer-derived
 Dickkopf1 inhibits osteoblast differentiation and osteoprotegerin expression: implication for
 breast cancer osteolytic bone metastases. *Int J Cancer.* 123:1034-1042, 2008. doi:
 10.1002/ijc.23625.
- 40. Buijs JT, Stayrook KR, Guise TA. The role of TGF-β in bone metastasis: novel therapeutic
 perspectives. *Bonekey Rep.* 1:96, 2012. doi: 10.1038/bonekey.2012.96.

- Buijs JT, van der Horst G, van den Hoogen C, Cheung H, de Rooij B, Kroon J, Petersen M,
 van Overveld PG, Pelger RC, van der Pluijm G. The BMP2/7 heterodimer inhibits the human
 breast cancer stem cell subpopulation and bone metastases formation. *Oncogene* 31: 21642174, 2012. doi: 10.1038/onc.2011.400.
- 2354 42. Cackowski FC, Eber MR, Rhee J, Decker AM, Yumoto K, Berry JE, Lee E, Shiozawa Y, Jung
 2355 Y, Aguirre-Ghiso JA, Taichman RS. Mer Tyrosine Kinase Regulates Disseminated Prostate
 2356 Cancer Cellular Dormancy. *J Cell Biochem* 118: 891-902, 2017. doi: 10.1002/jcb.25768.
- Caers J, Deleu S, Belaid Z, De Raeve H, Van Valckenborgh E, De Bruyne E, Defresne MP,
 Van Riet I, Van Camp B, Vanderkerken K. Neighboring adipocytes participate in the bone
 marrow microenvironment of multiple myeloma cells. *Leukemia*. 21:1580-1584, 2007. doi:
 10.1038/sj.leu.2404658
- 44. Campbell JP, Karolak MR, Ma Y, Perrien DS, Masood-Campbell SK, Penner NL, Munoz SA,
 Zijlstra A, Yang X, Sterling JA, Elefteriou F. Stimulation of host bone marrow stromal cells by
 sympathetic nerves promotes breast cancer bone metastasis in mice. *PLoS Biol.*10:e1001363, 2012. doi: 10.1371/journal.pbio.1001363.
- 2365 45. Capietto AH, Faccio R. Immune regulation of bone metastasis. *Bonekey Rep.* 3:600, 2014. doi:
 10.1038/bonekey.2014.95.
- Capulli M, Hristova D, Valbret Z, Carys K, Arjan R, Maurizi A, Masedu F, Cappariello A, Rucci
 N, Teti A. Notch2 pathway mediates breast cancer cellular dormancy and mobilisation in bone
 and contributes to haematopoietic stem cell mimicry. *Br J Cancer*. 121:157-171, 2019. doi:
 10.1038/s41416-019-0501-y.
- 2371 47. Carducci MA, Saad F, Abrahamsson PA, Dearnaley DP, Schulman CC, North SA, Sleep DJ,
 2372 Isaacson JD, Nelson JB. A phase 3 randomized controlled trial of the efficacy and safety of
 2373 atrasentan in men with metastatic hormone-refractory prostate cancer.; Atrasentan Phase III
- 2374 Study Group Institutions. *Cancer.* 110:1959-66, 2007. doi: 10.1002/cncr.22996.

- 2375 48. Cha YJ, Jung WH, Koo JS. Differential Site-Based Expression of Pentose Phosphate
 2376 Pathway-Related Proteins among Breast Cancer Metastases. *Dis Markers* 2017:7062517,
 2377 2017. doi: 10.1155/2017/7062517.
- 2378 49. Chaffer CL, San Juan BP, Lim E, Weinberg RA. EMT, cell plasticity and metastasis. *Cancer* 2379 *Metastasis Rev* 35: 645-654, 2016. doi: 10.1007/s10555-016-9648-7.
- Chang MK, Raggatt LJ, Alexander KA, Kuliwaba JS, Fazzalari NL, Schroder K, Maylin ER,
 Ripoll VM, Hume DA, Pettit AR. Osteal tissue macrophages are intercalated throughout human
 and mouse bone lining tissues and regulate osteoblast function in vitro and in vivo. *J Immunol*.181:1232-1244, 2008.
- Chantry AD, Heath D, Mulivor AW, Pearsall S, Baud'huin M, Coulton L, Evans H, Abdul N,
 Werner ED, Bouxsein ML, Key ML, Seehra J, Arnett TR, Vanderkerken K, Croucher P.
 Inhibiting activin-A signaling stimulates bone formation and prevents cancer-induced bone
 destruction in vivo. *J Bone Miner Res.* 25:2633-2646, 2010. doi: 10.1002/jbmr.142.
- 52. Chen GL, Luo Y, Eriksson D, Meng X, Qian C, Bäuerle T, Chen XX, Schett G, Bozec A. High
 fat diet increases melanoma cell growth in the bone marrow by inducing osteopontin and
 interleukin 6. *Oncotarget.* 7:26653-26669, 2016. doi: 10.18632/oncotarget.8474.
- 53. Chiao JW, Moonga BS, Yang YM, Kancherla R, Mittelman A, Wu-Wong JR, Ahmed T.
 Endothelin-1 from prostate cancer cells is enhanced by bone contact which blocks osteoclastic
 bone resorption. *Br J Cancer* 83:360-365, 2000. doi: 10.1054/bjoc.2000.1261.
- 54. Choudhary S, Ramasundaram P, Dziopa E, Mannion C, Kissin Y, Tricoli L, Albanese C, Lee
 W, Zilberberg J. Human ex vivo 3D bone model recapitulates osteocyte response to metastatic
 prostate cancer. *Sci Rep.* 8:17975, 2018. doi: 10.1038/s41598-018-36424-x.
- 2397 55. Chu K, Cheng CJ, Ye X, Lee YC, Zurita AJ, Chen DT, Yu-Lee LY, Zhang S, Yeh ET, Hu MC,
- Logothetis CJ, Lin SH. Cadherin-11 promotes the metastasis of prostate cancer cells to bone.
- 2399 Mol Cancer Res 6:1259-67, 2008. doi: 10.1158/1541-7786.MCR-08-0077.

2400	56.	Clément-Demange L, Mulcrone PL, Tabarestani TQ, Sterling JA, Elefteriou F. β2ARs
2401		stimulation in osteoblasts promotes breast cancer cell adhesion to bone marrow endothelial
2402		cells in an IL-1 β and selectin-dependent manner. J Bone Oncol. 13:1-10, 2018. doi:
2403		10.1016/j.jbo.2018.09.002.

- 2404 57. Clézardin P. Integrins in bone metastasis formation and potential therapeutic implications. *Curr* 2405 *Cancer Drug Targets* 9: 801-806, 2009. DOI: 10.2174/156800909789760348.
- 2406 58. Clézardin P, Benzaïd I, Croucher PI. Bisphosphonates in preclinical bone oncology. *Bone*.
 2407 49:66-70, 2011. doi: 10.1016/j.bone.2010.11.017.
- 2408 59. Coleman R. Treatment of Metastatic Bone Disease and the Emerging Role of Radium-223.
 2409 Semin Nucl Med. 46:99-104, 2016. doi: 10.1053/j.semnuclmed.2015.10.012.
- 2410 60. Coleman R, Body JJ, Aapro M, Hadji P, Herrstedt J. Bone health in cancer patients: ESMO
 2411 Clinical Practice Guidelines. *Ann Oncol.* 25 Suppl 3, iii124-iii137, 2014.
- Coleman R, Costa L, Saad F, Cook R, Hadji P, Terpos E, Garnero P, Brown J, Body JJ, Smith
 M, Lee KA, Major P, Dimopoulos M, Lipton A. Consensus on the utility of bone markers in the
 malignant bone disease setting. *Crit Rev Oncol Hematol.* 80:411-432, 2011. doi:
 10.1016/j.critrevonc.2011.02.005.
- 2416 62. Coleman R, de Boer R, Eidtmann H, Llombart A, Davidson N, Neven P, von Minckwitz G,
- 2417 Sleeboom HP, Forbes J, Barrios C, Frassoldati A, Campbell I, Paija O, Martin N, Modi A,
- 2418 Bundred N. Zoledronic acid (zoledronate) for postmenopausal women with early breast cancer
- receiving adjuvant letrozole (ZO-FAST study): final 60-month results. Ann Oncol. 24:398-405,
- 2420 2013. doi: 10.1093/annonc/mds277.

- Coleman R, Finkelstein DM, Barrios C, Martin M, Iwata H, Hegg R, Glaspy J, Periañez AM,
 Tonkin K, Deleu I, Sohn J, Crown J, Delaloge S, Dai T, Zhou Y, Jandial D, Chan A. Adjuvant
- 2424 controlled, phase 3 trial. *Lancet Oncol.* 21:60-72, 2020. doi: 10.1016/S1470-2045(19)30687-4.

99

denosumab in early breast cancer (D-CARE): an internatinal, multicentre, randomised,

Coleman R, Hall A, Albanell J, Hanby A, Bell R, Cameron D, Dodwell D, Marshall H, JeanMairet J, Tercero JC, Rojo F, Gregory W, Gomis RR. Effect of MAF amplification on treatment
outcomes with adjuvant zoledronic acid in early breast cancer: a secondary analysis of the
international, open-label, randomised, controlled, phase 3 AZURE (BIG 01/04) trial. *Lancet Oncol.* 18:1543-1552, 2017. doi: 10.1016/S1470-2045(17)30603-4.

- Coleman RE. Clinical features of metastatic bone disease and risk of skeletal morbidity. *Clin Cancer Res.* 12:6243s-6249s, 2006. doi:10.1158/1078-0432.CCR-06-0931.
- 2432 66. Coleman RE, Collinson M, Gregory W, Marshall H, Bell R, Dodwell D, Keane M, Gil M, Barrett-
- Lee P, Ritchie D, Bowman A, Liversedge V, De Boer RH, Passos-Coelho JL, O'Reilly S,
- 2434 Bertelli G, Joffe J, Brown JE, Wilson C, Tercero JC, Jean-Mairet J, Gomis R, Cameron D.
- 2435 Benefits and risks of adjuvant treatment with zoledronic acid in stage II/III breast cancer. 10
- 2436 years follow-up of the AZURE randomized clinical trial (BIG 01/04). *J Bone Oncol.* 13:123-135,
- 2437 2018. doi: 10.1016/j.jbo.2018.09.008.
- Coleman RE, Major P, Lipton A, Brown JE, Lee KA, Smith M, Saad F, Zheng M, Hei YJ,
 Seaman J, Cook R. Predictive value of bone resorption and formation markers in cancer
 patients with bone metastases receiving the bisphosphonate zoledronic acid. J Clin Oncol.
 23:4925-4935, 2005. doi: 10.1200/JCO.2005.06.091.
- 2442 68. Coleman RE, Rubens RD. The clinical course of bone metastases from breast cancer. Br J
 2443 Cancer. 55:61–66, 1987. doi: 10.1038/bjc.1987.13.
- Confavreux CB, Pialat JB, Bellière A, Brevet M, Decroisette C, Tescaru A, Wegrzyn J, Barrey
 C, Mornex F, Souquet PJ, Girard N. Bone metastases from lung cancer: A paradigm for
 multidisciplinary onco-rheumatology management. *Joint Bone Spine*. pii: S1297319X(18)30048-4, 2018. doi: 10.1016/j.jbspin.2018.03.005.

- Corey E, Brown LG, Kiefer JA, Quinn JE, Pitts TE, Blair JM, Vessella RL. Osteoprotegerin in
 prostate cancer bone metastasis. *Cancer Res.* 65:1710-1718, 2005. doi: 10.1158/00085472.CAN-04-2033.
- 2451 71. Cosman F, Crittenden DB, Grauer A. Romosozumab Treatment in Postmenopausal
 2452 Osteoporosis. *N Engl J Med.* 376:396-397, 2017. doi: 10.1056/NEJMc1615367.
- 2453 72. Costa L, Badia X, Chow E, Lipton A, Wardley A. Impact of skeletal complications on patients'
 2454 quality of life, mobility, and functional independence. *Support Care Cancer* 16:879–889, 2008.
 2455 doi:10.1007/s00520-008-0418-0.
- Cox TR, Rumney RMH, Schoof EM, Perryman L, Høye AM, Agrawal A, Bird D, Latif NA,
 Forrest H, Evans HR, Huggins ID, Lang G, Linding R, Gartland A, Erler JT. The hypoxic cancer
 secretome induces pre-metastatic bone lesions through lysyl oxidase. *Nature* 522: 106-110,
 2015. doi: 10.1038/nature14492.
- 2460 74. Croset M, Goehrig D, Frackowiak A, Bonnelye E, Ansieau S, Puisieux A, Clézardin P. TWIST1
 2461 expression in breast cancer cells facilitates bone metastasis formation. J Bone Miner Res.
 2462 29:1886-1899, 2014. doi: 10.1002/jbmr.2215.
- 75. Croset M, Pantano F, Kan CWS, Bonnelye E, Descotes F, Alix-Panabières C, Lecellier CH,
 Bachelier R, Allioli N, Hong SS, Bartkowiak K, Pantel K, Clézardin P. MicroRNA-30 family
 members inhibit breast cancer invasion, osteomimicry, and bone destruction by directly
 targeting multiple bone metastasis-associated genes. *Cancer Res* 78: 5260-5273, 2018. doi:
 10.1158/0008-5472.CAN-17-3058.
- 2468 76. Croucher PI, McDonald MM, Martin TJ. Bone metastasis: the importance of the 2469 neighbourhood. *Nat Rev Cancer* 16: 373-386, 2016. doi: 10.1038/nrc.2016.44.
- 2470 77. Dai J, Hall CL, Escara-Wilke J, Mizokami A, Keller JM, Keller ET. Prostate cancer induces
 2471 bone metastasis through Wnt-induced bone morphogenetic protein-dependent and

independent mechanisms. *Cancer Res* 68: 5785-5794, 2008. doi: 10.1158/0008-5472.CAN07-6541.

- 2474 78. Dai J, Keller J, Zhang J, Lu Y, Yao Z, Keller ET. Bone morphogenetic protein-6 promotes
 2475 osteoblastic prostate cancer bone metastases through a dual mechanism. *Cancer Res*2476 65:8274-8285, 2005. doi: 10.1158/0008-5472.CAN-05-1891.
- 2477 79. Dai J, Zhang H, Karatsinides A, Keller JM, Kozloff KM, Aftab DT, Schimmoller F, Keller ET.
 2478 Cabozantinib inhibits prostate cancer growth and prevents tumor-induced bone lesions. *Clin*2479 *Cancer Res.* 20:617-630, 2014. doi: 10.1158/1078-0432.CCR-13-0839.
- 2480 80. Dallas SL, Prideaux M, Bonewald LF. The osteocyte: an endocrine cell ... and more. *Endocr*2481 *Rev.* 34:658-690, 2013. doi: 10.1210/er.2012-1026.
- 2482 81. Danilin S, Merkel AR, Johnson JR, Johnson RW, Edwards JR, Sterling JA. Myeloid-derived
 2483 suppressor cells expand during breast cancer progression and promote tumor-induced bone
 2484 destruction. *Oncoimmunology*. 1:1484-1494, 2012. doi: 10.4161/onci.21990.
- 2485 82. Decker AM, Jung Y, Cackowski FC, Yumoto K, Wang J, Taichman RS. Sympathetic signaling
 2486 re-activates proliferation of dormant disseminated prostate cancer cells in the bone marrow.
 2487 *Mol Cancer Res.* 15:1644-1655, 2017. doi: 10.1158/1541-7786.MCR-17-0132.
- 2488 83. Delgado-Calle J, Anderson J, Cregor MD, Hiasa M, Chirgwin JM, Carlesso N, Yoneda T,
 2489 Mohammad KS, Plotkin LI, Roodman GD, Bellido T. Bidirectional Notch Signaling and
 2490 Osteocyte-Derived Factors in the Bone Marrow Microenvironment Promote Tumor Cell
 2491 Proliferation and Bone Destruction in Multiple Myeloma. *Cancer Res.* 76:1089-1100, 2016. doi:
 2492 10.1158/0008-5472.CAN-15-1703.
- 2493 84. Delliaux C, Tian TV, Bouchet M, Fradet A, Vanpouille N, Flourens A, Deplus R, Villers A, Leroy
- 2494 X, Clézardin P, de Launoit Y, Bonnelye E, Duterque-Coquillaud M. *TMPRSS2:ERG* gene 2495 fusion expression regulates bone markers and enhances the osteoblastic phenotype of

2496 prostate cancer bone metastases. *Cancer Lett.* 438:32-43, 2018. doi:
2497 10.1016/j.canlet.2018.08.027.

- 2498 85. Devignes CS, Aslan Y, Brenot A, Devillers A, Schepers K, Fabre S, Chou J, Casbon AJ, Werb
 2499 Z, Provot S. HIF signaling in osteoblast-lineage cells promotes systemic breast cancer growth
 and metastasis in mice. *Proc Natl Acad Sci U S A* 115: E992-E1001, 2018. doi:
 10.1073/pnas.1718009115.
- 2502 86. Dhesy-Thind S, Fletcher GG, Blanchette PS, Clemons MJ, Dillmon MS, Frank ES, Gandhi S,
 2503 Gupta R, Mates M, Moy B, Vandenberg T, Van Poznak CH. Use of Adjuvant Bisphosphonates
 2504 and Other Bone-Modifying Agents in Breast Cancer: A Cancer Care Ontario and American
 2505 Society of Clinical Oncology Clinical Practice Guideline. *J Clin Oncol.* 35:2062-2081, 2017. doi:
 2506 10.1200/JCO.2016.70.7257.
- 2507 87. Diedrich JD, Rajagurubandara E, Herroon MK, Mahapatra G, Hüttemann M, Podgorski I. Bone
 2508 marrow adipocytes promote the Warburg phenotype in metastatic prostate tumors via HIF-1α
 2509 activation. *Oncotarget*. 7:64854-64877, 2016. doi: 10.18632/oncotarget.11712.
- 251088.Dietterle J, Frydrychowicz C, Müller W, Hoffmann KT, Jähne K, Meixensberger J. Tumor-to-2511Tumor Metastasis of Multiple Meningiomas and Clear Cell Renal Cell Carcinoma Metastasis as
- 2512 First Clinical Appearance of Kidney Cancer: A Case Report and Analysis. *J Neurol Surg Rep.*
- 2513 81(1):e10-e14, 2020. doi: 10.1055/s-0040-1708846.
- 2514 89. Dolloff NG, Shulby SS, Nelson AV, Stearns ME, Johannes GJ, Thomas JD, Meucci O, Fatatis
 2515 A. Bone-metastatic potential of human prostate cancer cells correlates with Akt/PKB activation
 2516 by alpha platelet-derived growth factor receptor. *Oncogene*. 24:6848-6854, 2015. doi:
 2517 10.1038/sj.onc.1208815.
- D'Oronzo S, Brown J, Coleman R. The role of biomarkers in the management of bone-homing
 malignancies. *J Bone Oncol.* 9:1-9, 2017. doi: 10.1016/j.jbo.2017.09.001.

- Du WX, Duan SF, Chen JJ, Huang JF, Yin LM, Tong PJ. Serum bone-specific alkaline
 phosphatase as a biomarker for osseous metastases in patients with malignant carcinomas: a
 systematic review and meta-analysis. *J Cancer Res Ther.* 10 Suppl:C140-133, 2014. doi:
 10.4103/0973-1482.145842.
- Duong LT, Wesolowski GA, Leung P, Oballa R, Pickarski M. Efficacy of a cathepsin K inhibitor
 in a preclinical model for prevention and treatment of breast cancer bone metastasis. Mol
 Cancer Ther. 13:2898-2909, 2014. doi: 10.1158/1535-7163.MCT-14-0253.
- 2527 93. Early Breast Cancer Trialists Cooperative Group. Adjuvant bisphosphonate treatment in early
 breast cancer: meta-analyses of individual patient data from randomized trials. *Lancet.* 386:
 1353-1361, 2015. doi: 10.1016/S0140-6736(15)60908-4.
- 2530 94. Eastell R, O'Neill TW, Hofbauer LC, Langdahl B, Reid IR, Gold DT, Cummings SR.
 2531 Postmenopausal osteoporosis. *Nat Rev Dis Primers*. 2:16069, 2016. doi:
 2532 10.1038/nrdp.2016.69.
- 2533 95. Elefteriou F. Impact of the Autonomic Nervous System on the Skeleton. *Physiol Rev.* 98: 10831112, 2018. doi: 10.1152/physrev.00014.2017.
- 2535 96. Ell B, Mercatali L, Ibrahim T, Campbell N, Schwarzenbach H, Pantel K, Amadori D, Kang Y.
 2536 Tumor-induced osteoclast miRNA changes as regulators and biomarkers of osteolytic bone
 2537 metastasis. *Cancer Cell.* 24: 542-556, 2013. doi: 10.1016/j.ccr.2013.09.008.
- Prischke C, Zilionis R, Da Silva Martins J, Bos SA, Courties G, Rickelt S, Severe
 N, Baryawno N, Faget J, Savova V, Zemmour D, Kline J, Siwicki M, Garris C, Pucci F, Liao
 HW, Lin YJ, Newton A, Yaghi OK, Iwamoto Y, Tricot B, Wojtkiewicz GR, Nahrendorf M,
 Cortez-Retamozo V, Meylan E, Hynes RO, Demay M, Klein A, Bredella MA, Scadden DT,
 Weissleder R, Pittet MJ. Osteoblasts remotely supply lung tumors with cancer-promoting
- 2543 SiglecFhigh neutrophils. *Science* 358, 2017. pii: eaal5081. doi: 10.1126/science.aal5081.

- 2544 98. Erler JT, Bennewith KL, Cox TR, Lang G, Bird D, Koong A, Le QT, Giaccia AJ. Hypoxia-2545 induced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the 2546 premetastatic niche. *Cancer Cell.* 15: 35-44, 2009. doi: 10.1016/j.ccr.2008.11.012.
- 2547 99. Escudier B, Powles T, Motzer RJ, Olencki T, Arén Frontera O, Oudard S, Rolland F, Tomczak
- 2548 P, Castellano D, Appleman LJ, Drabkin H, Vaena D, Milwee S, Youkstetter J, Lougheed JC,
- Bracarda S, Choueiri TK. Cabozantinib, a New Standard of Care for Patients With Advanced
- Renal Cell Carcinoma and Bone Metastases? Subgroup Analysis of the METEOR Trial. J Clin
 Oncol. 36:765-772, 2018. doi: 10.1200/JCO.2017.74.7352.
- 2552 100. Falk S, Bannister K, Dickenson AH. Cancer pain physiology. *Br J Pain.* 8:154-162, 2014. doi:
 10.1177/2049463714545136.
- 101. Fang Y, Chen Y, Yu L, Zheng C, Qi Y, Li Z, Yang Z, Zhang Y, Shi T, Luo J, Liu M. Inhibition of
 breast cancer metastases by a novel inhibitor of TGFbeta receptor 1. *J Natl Cancer Inst* 105:
 47-58, 2013. doi: 10.1093/jnci/djs485.
- 102. Fend L, Accart N, Kintz J, Cochin S, Reymann C, Le Pogam F, Marchand JB, Menguy T, Slos 2557 2558 P, Rooke R, Fournel S, Bonnefoy JY, Préville X, Haegel H. Therapeutic effects of anti-CD115 2559 monoclonal antibody in mouse cancer models through dual inhibition of tumor-associated 2560 macrophages and osteoclasts. PLoS One. 8:e73310, 2013. doi: 2561 10.1371/journal.pone.0073310.
- Finlay IG, Mason MD, Shelley M. Radioisotopes for the palliation of metastatic bone cancer: a
 systematic review. *Lancet Oncol.* 6: 392-400, 2005. doi: 10.1016/S1470-2045(05)70206-0.
- 2564 104. Fioramonti M, Santini D, Iuliani M, Ribelli G, Manca P, Papapietro N, Spiezia F, Vincenzi B,
- 2565Denaro V, Russo A, Tonini G, Pantano F. Cabozantinib targets bone microenvironment2566modulating human osteoclast and osteoblast functions. *Oncotarget*. 8:20113-20121, 2017. doi:
- 2567 10.18632/oncotarget.15390.

Fizazi K, Carducci M, Smith M, Damião R, Brown J, Karsh L, Milecki P, Shore N, Rader M,
Wang H, Jiang Q, Tadros S, Dansey R, Goessl C. Denosumab versus zoledronic acid for
treatment of bone metastases in men with castration-resistant prostate cancer: a randomised,
double-blind study. *Lancet.* 377:813-822, 2011. doi: 10.1016/S0140-6736(10)62344-6.

- Florio M, Gunasekaran K, Stolina M, Li X, Liu L, Tipton B, Salimi-Moosavi H, Asuncion FJ, Li
 C, Sun B, Tan HL, Zhang L, Han CY, Case R, Duguay AN, Grisanti M, Stevens J, Pretorius
 JK, Pacheco E, Jones H, Chen Q, Soriano BD, Wen J, Heron B, Jacobsen FW, Brisan E,
 Richards WG, Ke HZ, Ominsky MS. A bispecific antibody targeting sclerostin and DKK-1
 promotes bone mass accrual and fracture repair. *Nat Commun.* 7:11505, 2016. doi:
 10.1038/ncomms11505.
- Fournier PG, Juárez P, Jiang G, Clines GA, Niewolna M, Kim HS, Walton HW, Peng XH, Liu Y,
 Mohammad KS, Wells CD, Chirgwin JM, Guise TA. The TGF-β signaling regulator PMEPA1
 suppresses prostate cancer metastases to bone. *Cancer Cell* 27:809-821, 2015. doi:
 10.1016/j.ccell.2015.04.009.
- 108. Fradet A, Bouchet M, Delliaux C, Gervais M, Kan C, Benetollo C, Pantano F, Vargas G,
 Bouazza L, Croset M, Bala Y, Leroy X, Rosol TJ, Rieusset J, Bellahcène A, Castronovo V,
 Aubin JE, Clézardin P, Duterque-Coquillaud M, Bonnelye E. Estrogen related receptor alpha in
 castration-resistant prostate cancer cells promotes tumor progression in bone. *Oncotarget* 7:
 77071-77086, 2016. doi: 10.18632/oncotarget.12787.
- Fradet A, Sorel H, Bouazza L, Goehrig D, Dépalle B, Bellahcène A, Castronovo V, Follet H,
 Descotes F, Aubin JE, Clézardin P, Bonnelye E. Dual function of ERRα in breast cancer and
 bone metastasis formation: implication of VEGF and osteoprotegerin. *Cancer Res* 71: 57285738, 2011. doi: 10.1158/0008-5472.CAN-11-1431.
- 110. Frees S, Breuksch I, Haber T, Bauer HK, Chavez-Munoz C, Raven P, Moskalev I, D Costa N,
 Tan Z, Daugaard M, Thüroff JW, Haferkamp A, Prawitt D, So A, Brenner W. Calcium-sensing

- receptor (CaSR) promotes development of bone metastasis in renal cell carcinoma.
 Oncotarget. 9:15766-15779, 2018. doi: 10.18632/oncotarget.24607.
- 2595111. Fujisaki J, Wu J, Carlson AL, Silberstein L, Putheti P, Larocca R, Gao W, Saito TI, Lo Celso C,2596Tsuyuzaki H, Sato T, Côté D, Sykes M, Strom TB, Scadden DT, Lin CP. In vivo imaging of2597Treg cells providing immune privilege to the haematopoietic stem-cell niche. Nature 474: 216-
- 2598 219, 2011. doi: 10.1038/nature10160.
- Fukuda T, Takeda S, Xu R, Ochi H, Sunamura S, Sato T, Shibata S, Yoshida Y, Gu Z, Kimura
 A, Ma C, Xu C, Bando W, Fujita K, Shinomiya K, Hirai T, Asou Y, Enomoto M, Okano H,
 Okawa A, Itoh H. Sema3A regulates bone-mass accrual through sensory innervations. *Nature*497: 490-493, 2013. doi: 10.1038/nature12115.
- Furlan F, Galbiati C, Jorgensen NR, Jensen JE, Mrak E, Rubinacci A, Talotta F, Verde P, Blasi
 F. Urokinase plasminogen activator receptor affects bone homeostasis by regulating
 osteoblast and osteoclast function. *J Bone Miner Res* 22:1387-96, 2007. doi:
 10.1359/jbmr.070516.
- 2607 114. Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune
 2608 system. *Nat Rev Immunol.* 9: 162-174, 2009. doi: 10.1038/nri2506.
- 2609 115. Galasko CS. Monitoring of bone metastases. *Schweiz Med Wochenschr.* 111:1873-1875,
 2610 1981.
- 2611 116. Gartrell BA, Saad F. Managing bone metastases and reducing skeletal related events in
 2612 prostate cancer. *Nat Rev Clin Oncol.* 11:335-345, 2014. doi: 10.1038/nrclinonc.2014.70.
- 2613 117. Gawrzak S, Rinaldi L, Gregorio S, Arenas EJ, Salvador F, Urosevic J, Figueras-Puig C, Rojo
- 2614 F, Del Barco Barrantes I, Cejalvo JM, Palafox M, Guiu M, Berenguer-Llergo A, Symeonidi A,
- 2615 Bellmunt A, Kalafatovic D, Arnal-Estapé A, Fernández E, Müllauer B, Groeneveld R,
- 2616 Slobodnyuk K, Stephan-Otto Attolini C, Saura C, Arribas J, Cortes J, Rovira A, Muñoz M, Lluch
- A, Serra V, Albanell J, Prat A, Nebreda AR, Benitah SA, Gomis RR. MSK1 regulates luminal
2618 cell differentiation and metastatic dormancy in ER+ breast cancer. Nat Cell Biol. 20: 211-221,

2619 2018. doi: 10.1038/s41556-017-0021-z.

- 2620 118. Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M, Ley K. Development of monocytes,
 2621 macrophages, and dendritic cells. *Science.* 327: 656-61, 2010. doi: 10.1126/science.
- 2622 119. Ghajar CM, Peinado H, Mori H, Matei IR, Evason KJ, Brazier H, Almeida D, Koller A, Hajjar
 2623 KA, Stainier DY, Chen EI, Lyden D, Bissell MJ. The perivascular niche regulates breast tumour
 2624 dormancy. *Nat Cell Biol* 15: 807-817, 2013. doi: 10.1038/ncb2767.
- 2625 120. Gnant M, Baselga J, Rugo HS, Noguchi S, Burris HA, Piccart M, Hortobagyi GN, Eakle J,
 2626 Mukai H, Iwata H, Geberth M, Hart LL, Hadji P, El-Hashimy M, Rao S, Taran T, Sahmoud T,
 2627 Lebwohl D, Campone M, Pritchard KI. Effect of everolimus on bone marker levels and
 2628 progressive disease in bone in BOLERO-2. *J Natl Cancer Inst* 105(9):654-663, 2013. doi:
 2629 10.1093/jnci/djt026.
- 2630 121. Gnant M, Pfeiler G, Dubsky PC, Hubalek M, Greil R, Jakesz R, Wette V, Balic M, Haslbauer F,
- 2631 Melbinger E, Bjelic-Radisic V, Artner-Matuschek S, Fitzal F, Marth C, Sevelda P, Mlineritsch B,
- 2632 Steger GG, Manfreda D, Exner R, Egle D, Bergh J, Kainberger F, Talbot S, Warner D, Fesl C,
- 2633 Singer CF; Austrian Breast and Colorectal Cancer Study Group. Adjuvant denosumab in
- breast cancer (ABCSG-18): a multicentre, randomised, double-blind, placebo-controlled trial.
- 2635 *Lancet.* 386:433-443, 2015. doi: 10.1016/S0140-6736(15)60995-3.
- 2636 122. Gnant M, Pfeiler G, Steger GG, Egle D, Greil R, Fitzal F, Wette V, Balic M, Haslbauer F,
 2637 Melbinger-Zeinitzer E, Bjelic-Radisic V, Jakesz R, Marth C, Sevelda P, Mlineritsch B, Exner R,
 2638 Fesl C, Frantal S, Singer CF, on behalf of Austrian Breast and Colorectal Cancer Study Group.
 2639 Adjuvant denosumab in postmenopausal patients with hormone receptor-positive breast
 2640 cancer (ABCSG-18): disease-free survival results from a randomized, double-blind, placebo2641 controlled, phase 3 trial. *Lancet Oncol.* 20: 339-351, 2019. doi: 10.1016/S14702642 2045(18)30862-3.

2643 123. Gnant MF, Mlineritsch B, Luschin-Ebengreuth G, Grampp S, Kaessmann H, Schmid M,
2644 Menzel C, Piswanger-Soelkner JC, Galid A, Mittlboeck M, Hausmaninger H, Jakesz R;
2645 Austrian Breast and Colorectal Cancer Study Group. Zoledronic acid prevents cancer
2646 treatment-induced bone loss in premenopausal women receiving adjuvant endocrine therapy
2647 for hormone-responsive breast cancer: a report from the Austrian Breast and Colorectal
2648 Cancer Study Group. J Clin Oncol. 25:820-828, 2007. doi: 10.1200/JCO.2005.02.7102.

- 2649 124. Goto T, Nakao K, Gunjigake KK, Kido MA, Kobayashi S, Tanaka T. Substance P stimulates
 2650 late-stage rat osteoblastic bone formation through neurokinin-1 receptors. *Neuropeptides* 41:
 2651 25-31, 2007. doi: 10.1016/j.npep.2006.11.002.
- 2652 125. Grimaud E, Soubigou L, Couillaud S, Coipeau P, Moreau A, Passuti N, Gouin F, Redini F,
 2653 Heymann D. Receptor activator of nuclear factor kappaB ligand (RANKL)/osteoprotegerin
 2654 (OPG) ratio is increased in severe osteolysis. *Am. J Pathol.* 163: 2021-2031, 2003. doi:
 2655 10.1016/s0002-9440(10)63560-2.
- 2656 126. Guise TA, Yin JJ, Taylor SD, Kumagai Y, Dallas M, Boyce BF, Yoneda T, Mundy GR.
 2657 Evidence for a causal role of parathyroid hormone-related protein in the pathogenesis of
 2658 human breast cancer-mediated osteolysis. *J Clin Invest.* 98:1544-1549, 1996. DOI:
 2659 10.1172/JCI118947.
- 127. Hadji P, Aapro MS, Body JJ, Gnant M, Brandi ML, Reginster JY, Zillikens MC, Glüer CC, de
 Villiers T, Baber R, Roodman GD, Cooper C, Langdahl B, Palacios S, Kanis J, Al-Daghri N,
 Nogues X, Eriksen EF, Kurth A, Rizzoli R, Coleman RE. Management of Aromatase InhibitorAssociated Bone Loss (AIBL) in postmenopausal women with hormone sensitive breast
 cancer: Joint position statement of the IOF, CABS, ECTS, IEG, ESCEO IMS, and SIOG. J *Bone Oncol.* 7:1-12, 2017. doi: 10.1016/j.jbo.2017.03.001.
- 2666 128. Hadji P, Coleman RE, Wilson C, Powles TJ, Clézardin P, Aapro M, Costa L, Body JJ,
 2667 Markopoulos C, Santini D, Diel I, Di Leo A, Cameron D, Dodwell D, Smith I, Gnant M, Gray R,

- Harbeck N, Thurlimann B, Untch M, Cortes J, Martin M, Albert US, Conte PF, Ejlertsen B,
 Bergh J, Kaufmann M, Holen I. Adjuvant bisphosphonates in early breast cancer: consensus
 guidance for clinical practice from a European Panel. *Ann Oncol.* 27:379-390, 2016. doi:
 10.1093/annonc/mdv617.
- 2672 129. Hall CL, Bafico A, Dai J, Aaronson SA, Keller ET. Prostate cancer cells promote osteoblastic
 2673 bone metastases through Wnts. *Cancer Res.* 65:7554-7560, 2005. doi: 10.1158/00082674 5472.CAN-05-1317.
- 130. Hall CL, Daignault SD, Shah RB, Pienta KJ, Keller ET. Dickkopf-1 expression increases early
 in prostate cancer development and decreases during progression from primary tumor to
 metastasis. *Prostate*. 68:1396-1404, 2008. doi: 10.1002/pros.20805.
- Hall CL, Zhang H, Baile S, Ljungman M, Kuhstoss S, Keller ET. p21CIP-1/WAF-1 induction is
 required to inhibit prostate cancer growth elicited by deficient expression of the Wnt inhibitor
 Dickkopf-1. Cancer Res. 70:9916-9926, 2010. doi: 10.1158/0008-5472.CAN-10-0440.
- Hamidi H, Ivaska J. Every step of the way: integrins in cancer progression and metastasis. *Nat Rev Cancer* 2018. doi: 10.1038/s41568-018-0038-z.
- 2683 133. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 144: 646-674, 2011.
 2684 doi: 10.1016/j.cell.2011.02.013.
- 134. Hardaway AL, Herroon MK, Rajagurubandara E, Podgorski I. Bone marrow fat: linking
 adipocyte-induced inflammation with skeletal metastases. *Cancer Metastasis Rev.* 33:527-453,
 2687 2014. doi: 10.1007/s10555-013-9484-y.
- 2688135.Hardaway AL, Herroon MK, Rajagurubandara E, Podgorski I. Marrow adipocyte-derived2689CXCL1 and CXCL2 contribute to osteolysis in metastatic prostate cancer. *Clin Exp Metastasis.*
- 2690 32:353-368, 2015. doi: 10.1007/s10585-015-9714-5.
- 2691 136. Harms JF, Welch DR, Samant RS, Shevde LA, Miele ME, Babu GR, Goldberg SF, Gilman VR,
 2692 Sosnowski DM, Campo DA, Gay CV, Budgeon LR, Mercer R, Jewell J, Mastro AM, Donahue

HJ, Erin N, Debies MT, Meehan WJ, Jones AL, Mbalaviele G, Nickols A, Christensen ND,
Melly R, Beck LN, Kent J, Rader RK, Kotyk JJ, Pagel MD, Westlin WF, Griggs DW. A small
molecule antagonist of the alpha(v)beta3 integrin suppresses MDA-MB-435 skeletal
metastasis. *Clin Exp Metastasis* 21:119-128, 2004.

- 137. Hashimoto K, Ochi H, Sunamura S, Kosaka N, Mabuchi Y, Fukuda T, Yao K, Kanda H, Ae K,
 Okawa A, Akazawa C, Ochiya T, Futakuchi M, Takeda S, Sato S. Cancer-secreted hsa-miR940 induces an osteoblastic phenotype in the bone metastatic microenvironment via targeting
 ARHGAP1 and FAM134A. *Proc Natl Acad Sci USA* 115: 2204-2209, 2018. doi:
 10.1073/pnas.1717363115.
- Haug JS, He XC, Grindley JC, Wunderlich JP, Gaudenz K, Ross JT, Paulson A, Wagner KP,
 Xie Y, Zhu R, Yin T, Perry JM, Hembree MJ, Redenbaugh EP, Radice GL, Seidel C, Li L. Ncadherin expression level distinguishes reserved versus primed states of hematopoietic stem
 cells. Cell Stem Cell 2:367-379, 2008. doi: 10.1016/j.stem.2008.01.017.
- Henry DH, Costa L, Goldwasser F, Hirsh V, Hungria V, Prausova J, Scagliotti GV, Sleeboom
 H, Spencer A, Vadhan-Raj S, von Moos R, Willenbacher W, Woll PJ, Wang J, Jiang Q, Jun S,
 Dansey R, Yeh H. Randomized, double-blind study of denosumab versus zoledronic acid in
 the treatment of bone metastases in patients with advanced cancer (excluding breast and
 prostate cancer) or multiple myeloma. *J Clin Oncol.* 29:1125-1132, 2011. doi:
 10.1200/JCO.2010.31.3304.
- 140. Herroon MK, Diedrich JD, Rajagurubandara E, Martin C, Maddipati KR, Kim S, Heath El,
 Granneman J, Podgorski I. Prostate Tumor Cell-Derived IL1β Induces an Inflammatory
 Phenotype in Bone Marrow Adipocytes and Reduces Sensitivity to Docetaxel via LipolysisDependent Mechanisms. *Mol Cancer Res.* 17:2508-2521, 2019. doi: 10.1158/15417786.MCR-19-0540.

- 141. Herroon MK, Rajagurubandara E, Diedrich JD, Heath El, Podgorski I. Adipocyte-activated
 oxidative and ER stress pathways promote tumor survival in bone via upregulation of Heme
 Oxygenase 1 and Survivin. *Sci Rep.* 8:40, 2018. doi: 10.1038/s41598-017-17800-5.
- 2720 142. Herroon MK, Rajagurubandara E, Hardaway AL, Powell K, Turchick A, Feldmann D, Podgorski
- I. Bone marrow adipocytes promote tumor growth in bone via FABP4-dependent mechanisms.
 Oncotarget. 4:2108-2123, 2013. doi: 10.18632/oncotarget.1482.
- 143. Hesse E, Schröder S, Brandt D, Pamperin J, Saito H, Taipaleenmäki H. Sclerostin inhibition
 alleviates breast cancer-induced bone metastases and muscle weakness. *JCI Insight*. pii:
 125543, 2019. doi: 10.1172/jci.insight.125543.
- 144. Higano CS. Understanding treatments for bone loss and bone metastases in patients with
 prostate cancer: a practical review and guide for the clinician. *Urol Clin North Am.* 3: 331-352,
 2004. 10.1016/j.ucl.2004.01.001.
- 2729 145. Himelstein AL, Foster JC, Khatcheressian JL, Roberts JD, Seisler DK, Novotny PJ, Qin R, Go
- 2730 RS, Grubbs SS, O'Connor T, Velasco MR Jr, Weckstein D, O'Mara A, Loprinzi CL, Shapiro CL.
- 2731 Effect of Longer-Interval vs Standard Dosing of Zoledronic Acid on Skeletal Events in Patients
- With Bone Metastases: A Randomized Clinical Trial. JAMA. 317:48-58, 2017. doi:
 10.1001/jama.2016.19425.
- 146. Hiraga T, Kizaka-Kondoh S, Hirota K, Hiraoka M, Yoneda T. Hypoxia and hypoxia-inducible
 factor-1 expression enhance osteolytic bone metastases of breast cancer. *Cancer Res.* 67:
 4157-4163, 2007. doi: 10.1158/0008-5472.CAN-06-2355.
- 2737 147. Hiraga T, Myoui A, Hashimoto N, Sasaki A, Hata K, Morita Y, Yoshikawa H, Rosen CJ, Mundy
- 2738 GR, Yoneda T. Bone-derived IGF mediates crosstalk between bone and breast cancer cells in 2739 bony metastases. *Cancer Res.* 72:4238-4249, 2012. doi: 10.1158/0008-5472.CAN-11-3061.
- 2740 148. Hiraoka K, Zenmyo M, Watari K, Iguchi H, Fotovati A, Kimura YN, Hosoi F, Shoda T, Nagata
- 2741 K, Osada H, Ono M, Kuwano M. Inhibition of bone and muscle metastases of lung cancer cells

- by a decrease in the number of monocytes/ macrophages. *Cancer Sci.* 99:1595-602, 2008.
 doi: 10.1111/j.1349-7006.2008.00880.x.
- 149. Hiratsuka S, Watanabe A, Aburatani H, Maru Y. Tumour-mediated upregulation of
 chemoattractants and recruitment of myeloid cells predetermines lung metastasis. *Nat Cell Biol.* 8:1369-1375, 2006. doi: 10.1038/ncb1507.
- 150. Hofbauer LC, Rachner TD, Coleman RE, Jakob F. Endocrine aspects of bone metastases. *Lancet Diabetes Endocrinol.* 2:500-512, 2014. doi: 10.1016/S2213-8587(13)70203-1.
- 151. Holen I, Lefley DV, Francis SE, Rennicks S, Bradbury S, Coleman RE, Ottewell P. IL-1 drives
 breast cancer growth and bone metastasis in vivo. *Oncotarget* 7: 75571-75584, 2016. doi:
 10.18632/oncotarget.12289.
- Hortobagyi GN. Everolimus plus exemestane for the treatment of advanced breast cancer: a
 review of subanalyses from BOLERO-2. *Neoplasia* 17:279-288, 2015. doi:
 10.1016/j.neo.2015.01.005.
- Hortobagyi GN, Theriault RL, Porter L, Blayney D, Lipton A, Sinoff C, Wheeler H, Simeone JF,
 Seaman J, Knight RD. Efficacy of pamidronate in reducing skeletal complications in patients
 with breast cancer and lytic bone metastases. Protocol 19 Aredia Breast Cancer Study Group. *N Engl J Med.* 335:1785-1791, 1996. doi: 10.1056/NEJM199612123352401.
- Huang CF, Lira C, Chu K, Bilen MA, Lee YC, Ye X, Kim SM, Ortiz A, Wu FL, Logothetis CJ,
 Yu-Lee LY, Lin SH. Cadherin-11 increases migration and invasion of prostate cancer cells and
 enhances their interaction with osteoblasts. Cancer Res 70: 4580-4589, 2010. doi:
 10.1158/0008-5472.CAN-09-3016.
- 155. Huang J, Gu T, Ying J. A meta-analysis survey of appropriate bone turnover markers in the
 detection of bone metastasis in lung cancer. *Int J Clin Oncol.* 22:1015-1025, 2017. doi:
 10.1007/s10147-017-1159-1.

- 156. Iuliani M, Pantano F, Buttigliero C, Fioramonti M, Bertaglia V, Vincenzi B, Zoccoli A, Ribelli G,
 Tucci M, Vignani F, Berruti A, Scagliotti GV, Tonini G, Santini D. Biological and clinical effects
 of abiraterone on anti-resorptive and anabolic activity in bone microenvironment. *Oncotarget*.
 6:12520-12528, 2015. doi: 10.18632/oncotarget.3724.
- 2770 157. James ND, Sydes MR, Clarke NW, Mason MD, Dearnaley DP, Spears MR, Ritchie AW, Parker 2771 CC, Russell JM, Attard G, de Bono J, Cross W, Jones RJ, Thalmann G, Amos C, Matheson D, 2772 Millman R, Alzouebi M, Beesley S, Birtle AJ, Brock S, Cathomas R, Chakraborti P, Chowdhury 2773 S, Cook A, Elliott T, Gale J, Gibbs S, Graham JD, Hetherington J, Hughes R, Laing R, McKinna F, McLaren DB, O'Sullivan JM, Parikh O, Peedell C, Protheroe A, Robinson AJ, 2774 2775 Srihari N, Srinivasan R, Staffurth J, Sundar S, Tolan S, Tsang D, Wagstaff J, Parmar MK; 2776 STAMPEDE investigators. Addition of docetaxel, zoledronic acid, or both to first-line long-term 2777 hormone therapy in prostate cancer (STAMPEDE): survival results from an adaptive, multiarm, multistage, platform randomised controlled trial. Lancet. 387:1163-1177, 2016. doi: 2778 2779 10.1016/S0140-6736(15)01037-5.
- Jamieson WL, Shimizu S, D'Ambrosio JA, Meucci O, Fatatis A. CX3CR1 is expressed by
 prostate epithelial cells and androgens regulate the levels of CX3CL1/fractalkine in the bone
 marrow: potential role in prostate cancer bone tropism. *Cancer Res.* 68:1715-22, 2008. doi:
 10.1158/0008-5472.CAN-07-1315.
- Jamieson-Gladney WL, Zhang Y, Fong AM, Meucci O, Fatatis A. The chemokine receptor
 CX₃CR1 is directly involved in the arrest of breast cancer cells to the skeleton. *Breast Cancer Res.* 13:R91, 2011. doi: 10.1186/bcr3016.
- 2787160.Jensen AB, Wynne C, Ramirez G, He W, Song Y, Berd Y, Wang H, Mehta A, Lombardi A. The2788cathepsin K inhibitor odanacatib suppresses bone resorption in women with breast cancer and2789established bone metastases: results of a 4-week, double-blind, randomized, controlled trial.
- 2790 *Clin Breast Cancer.* 10:452-458, 2010. doi: 10.3816/CBC.2010.n.059.

Jiao S, Subudhi SK, Aparicio A, Ge Z, Guan B, Miura Y, Sharma P. Differences in Tumor
 Microenvironment Dictate T Helper Lineage Polarization and Response to Immune Checkpoint
 Therapy. *Cell.* 179:1177-1190.e13, 2019. doi: 10.1016/j.cell.2019.10.029.

- 2794 162. Jimenez-Andrade JM, Ghilardi JR, Castañeda-Corral G, Kuskowski MA, Mantyh PW.
 2795 Preventive or late administration of anti-NGF therapy attenuates tumor-induced nerve
 2796 sprouting, neuroma formation, and cancer pain. *Pain.* 152:2564-2574, 2011. doi:
 2797 10.1016/j.pain.2011.07.020.
- Indexto 163. Johnson RW, Finger EC, Olcina MM, Vilalta M, Aguilera T, Miao Y, Merkel AR, Johnson JR,
 Sterling JA, Wu JY, Giaccia AJ. Induction of LIFR confers a dormancy phenotype in breast
 cancer cells disseminated to the bone marrow. *Nat Cell Biol* 18: 1078-1089, 2016. doi:
 10.1038/ncb3408.
- Interpretation 2802
 Interpretation 2803
 Interpretation 2804
 Interpretation 2805
 Interpretation 2805
 Interpretation 2806
 Interpretation 2807
 Interpr
- 165. Jung K, Lein M. Bone turnover markers in serum and urine as diagnostic, prognostic and
 monitoring biomarkers of bone metastasis. *Biochim Biophys Acta*. 1846:425-438, 2014. doi:
 10.1016/j.bbcan.2014.09.001.
- Jung Y, Kim JK, Shiozawa Y, Wang J, Mishra A, Joseph J, Berry JE, McGee S, Lee E, Sun H,
 Wang J, Jin T, Zhang H, Dai J, Krebsbach PH, Keller ET, Pienta KJ, Taichman RS.
 Recruitment of mesenchymal stem cells into prostate tumours promotes metastasis. *Nat Commun.* 4:1795, 2013. doi: 10.1038/ncomms2766.
- 167. Jung Y, Wang J, Lee E, McGee S, Berry JE, Yumoto K, Dai J, Keller ET, Shiozawa Y,
 Taichman RS. Annexin 2-CXCL12 interactions regulate metastatic cell targeting and growth in
 the bone marrow. *Mol Cancer Res.* 13:197-207, 2015. doi: 10.1158/1541-7786.MCR-14-0118.

168. Kakonen SM, Selander KS, Chirgwin JM, Yin JJ, Burns S, Rankin WA, Grubbs BG, Dallas M,
Cui Y, Guise TA. Transforming growth factor-beta stimulates parathyroid hormone-related
protein and osteolytic metastases via Smad and mitogen-activated protein kinase signaling
pathways. *J Biol Chem* 277:24571-24578, 2002. doi: 10.1074/jbc.M202561200.

- 2820 169. Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, MacDonald DD, Jin DK,
- 2821 Shido K, Kerns SA, Zhu Z, Hicklin D, Wu Y, Port JL, Altorki N, Port ER, Ruggero D, Shmelkov
- 2822 SV, Jensen KK, Rafii S, Lyden D. VEGFR1-positive haematopoietic bone marrow progenitors 2823 initiate the pre-metastatic niche. *Nature*. 438:820-7, 2005. doi: 10.1038/nature04186.
- 170. Katoh M, Katoh M. Molecular genetics and targeted therapy of WNT-related human diseases.
 Int J Mol Med 40 :587-606, 2017. doi: 10.3892/ijmm.2017.3071.
- 171. Katsuno Y, Hanyu A, Kanda H, Ishikawa Y, Akiyama F, Iwase T, Ogata E, Ehata S, Miyazono
 K, Imamura T. Bone morphogenetic protein signaling enhances invasion and bone metastasis
 of breast cancer cells through Smad pathway. *Oncogene*. 27:6322-6333, 2008. doi:
 10.1038/onc.2008.232.
- 172. Keskinov AA, Shurin MR. Myeloid regulatory cells in tumor spreading and metastasis.
 Immunobiology 220: 236-242, 2015. doi: 10.1016/j.imbio.
- 2832 173. Khoo WH, Ledergor G, Weiner A, Roden DL, Terry RL, McDonald MM, Chai RC, De Veirman
- 2833 K, Owen KL, Opperman KS, Vandyke K, Clark JR, Seckinger A, Kovacic N, Nguyen A,
- 2834 Mohanty ST, Pettitt JA, Xiao Y, Corr AP, Seeliger C, Novotny M, Lasken RS, Nguyen TV,
- 2835 Oyajobi BO, Aftab D, Swarbrick A, Parker B, Hewett DR, Hose D, Vanderkerken K, Zannettino
- 2836 ACW, Amit I, Phan TG, Croucher PI. A niche-dependent myeloid transcriptome signature
- 2837 defines dormant myeloma cells. *Blood* 134:30-43, 2019. doi: 10.1182/blood.2018880930.
- 2838 174. Kimura T, Kuwata T, Ashimine S, Yamazaki M, Yamauchi C, Nagai K, Ikehara A, Feng Y,
 2839 Dimitrov DS, Saito S, Ochiai A. Targeting of bone-derived insulin-like growth factor-II by a

- human neutralizing antibody suppresses the growth of prostate cancer cells in a human bone environment. *Clin Cancer Res.* 16:121-129, 2010. doi: 10.1158/1078-0432.CCR-09-0982.
- Kirschenbaum A, Izadmehr S, Yao S, O'Connor-Chapman KL, Huang A, Gregoriades EM,
 Yakar S, Levine AC. Prostatic Acid Phosphatase Alters the RANKL/OPG System and Induces
 Osteoblastic Prostate Cancer Bone Metastases. *Endocrinology* 157:4526-4533, 2016. doi:
- 2845 <u>10.1210/en.2016-1606</u>.
- 176. Kitagawa Y, Dai J, Zhang J, Keller JM, Nor J, Yao Z, Keller ET. Vascular endothelial growth
 factor contributes to prostate cancer-mediated osteoblastic activity. *Cancer Res* 65:1092110929, 2005. doi: 10.1158/0008-5472.CAN-05-1809.
- 177. Kobayashi A, Okuda H, Xing F, Pandey PR, Watabe M, Hirota S, Pai SK, Liu W, Fukuda K,
 Chambers C, Wilber A, Watabe K. Bone morphogenetic protein 7 in dormancy and metastasis
 of prostate cancer stem-like cells in bone. *J Exp Med* 208: 2641-2655, 2011. doi:
 10.1084/jem.20110840.
- 178. Koeneman KS, Yeung F, Chung LW. Osteomimetic properties of prostate cancer cells: a
 hypothesis supporting the predilection of prostate cancer metastasis and growth in the bone
 environment. Prostate 39:246-261, 1999. doi: <u>10.1002/(sici)1097-</u>
 0045(19990601)39:4<246::aid-pros5>3.0.co;2-u.
- Kohno N, Aogi K, Minami H, Nakamura S, Asaga T, Iino Y, Watanabe T, Goessl C, Ohashi Y,
 Takashima S. Zoledronic acid significantly reduces skeletal complications compared with
 placebo in Japanese women with bone metastases from breast cancer: a randomized,
 placebo-controlled trial. *J Clin Oncol.* 23:3314-3321, 2005. doi: 10.1200/JCO.2005.05.116.
- 180. Koopmans N, de Jong IJ, Breeuwsma AJ, van der Veer E. Serum bone turnover markers
 (PINP and ICTP) for the early detection of bone metastases in patients with prostate cancer: a
 longitudinal approach. *J Urol.* 178: 849-853, 2007. doi: 10.1016/j.juro.2007.05.029.

- 2864 181. Krzeszinski JY, Wei W, Huynh H, Jin Z, Wang X, Chang TC, Xie XJ, He L, Mangala LS, Lopez-2865 Berestein G, Sood AK, Mendell JT, Wan Y. miR-34a blocks osteoporosis and bone metastasis 2866 inhibiting osteoclastogenesis and Tgif2. Nature. 512:431-435, 2014. doi: by 2867 10.1038/nature13375.
- 182. Kusumbe AP, Ramasamy SK, Adams RH. Coupling of angiogenesis and osteogenesis by a
 specific vessel subtype in bone. *Nature* 507:323-328, 2014. doi: 10.1038/nature13145.
- Lacey DL, Boyle WJ, Simonet WS, Kostenuik PJ, Dougall WC, Sullivan JK, San Martin J,
 Dansey R. Bench to bedside: elucidation of the OPG-RANK-RANKL pathway and the
 development of denosumab. *Nat Rev Drug Discov* 11: 401-419, 2012. doi: 10.1038/nrd3705.
- 184. Lamiche C, Clarhaut J, Strale PO, Crespin S, Pedretti N, Bernard FX, Naus CC, Chen VC,
 Foster LJ, Defamie N, Mesnil M, Debiais F, Cronier L. The gap junction protein Cx43 is
 involved in the bone-targeted metastatic behaviour of human prostate cancer cells. *Clin Exp Metastasis*. 29:111-122, 2012. doi: 10.1007/s10585-011-9434-4.
- Lamoureux F, Baud'huin M, Rodriguez Calleja L, Jacques C, Berreur M, Rédini F, Lecanda F,
 Bradner JE, Heymann D, Ory B. Selective inhibition of BET bromodomain epigenetic signalling
 interferes with the bone-associated tumour vicious cycle. *Nat Commun.* 5:3511, 2014. doi:
 10.1038/ncomms4511.
- 186. Langdahl B, Binkley N, Bone H, Gilchrist N, Resch H, Rodriguez Portales J, Denker A,
 Lombardi A, Le Bailly De Tilleghem C, Dasilva C, Rosenberg E, Leung A. Odanacatib in the
 treatment of postmenopausal women with low bone mineral density: five years of continued
 therapy in a phase 2 study. *J Bone Miner Res.* 27:2251-2258, 2012. doi: 10.1002/jbmr.1695.
- Lara PN Jr, Ely B, Quinn DI, Mack PC, Tangen C, Gertz E, Twardowski PW, Goldkorn A,
 Hussain M, Vogelzang NJ, Thompson IM, Van Loan MD. Serum biomarkers of bone
 metabolism in castration-resistant prostate cancer patients with skeletal metastases: results
 from SWOG 0421. *J Natl Cancer Inst.* 106:dju013, 2014. doi: 10.1093/jnci/dju013.

- Larson SR, Chin J, Zhang X, Brown LG, Coleman IM, Lakely B, Tenniswood M, Corey E,
 Nelson PS, Vessella RL, Morrissey C. Prostate cancer derived prostatic acid phosphatase
 promotes an osteoblastic response in the bone microenvironment. *Clin Exp Metastasis* 31:
 247-256, 2014. doi: 10.1007/s10585-013-9625-2.
- 2893 189. Lawson MA, McDonald MM, Kovacic N, Hua Khoo W, Terry RL, Down J, Kaplan W, Paton-
- Hough J, Fellows C, Pettitt JA, Neil Dear T, Van Valckenborgh E, Baldock PA, Rogers MJ,
- Eaton CL, Vanderkerken K, Pettit AR, Quinn JM, Zannettino AC, Phan TG, Croucher PI.
 Osteoclasts control reactivation of dormant myeloma cells by remodelling the endosteal niche.
 Nat Commun 6:8983, 2015. doi: 10.1038/ncomms9983.
- Leblanc R, Lee SC, David M, Bordet JC, Norman DD, Patil R, Miller D, Sahay D, Ribeiro J,
 Clézardin P, Tigyi GJ, Peyruchaud O. Interaction of platelet-derived autotaxin with tumor
 integrin αVβ3 controls metastasis of breast cancer cells to bone. *Blood* 124:3141-3150, 2014.
 doi: 10.1182/blood-2014-04-56868.
- 2902 191. Leblanc R, Peyruchaud O. Metastasis: new functional implications of platelets and
 2903 megakaryocytes. *Blood* 128:24-31, 2016. doi: 10.1182/blood-2016-01-636399.
- Lee C, Whang YM, Campbell P, Mulcrone PL, Elefteriou F, Cho SW, Park SI. Dual targeting cmet and VEGFR2 in osteoblasts suppresses growth and osteolysis of prostate cancer bone
 metastasis. *Cancer Lett.* 414:205-213, 2018. doi: 10.1016/j.canlet.2017.11.016.
- Lee JH, Kim HN, Kim KO, Jin WJ, Lee S, Kim HH, Ha H, Lee ZH. CXCL10 promotes osteolytic
 bone metastasis by enhancing cancer outgrowth and osteoclastogenesis. *Cancer Res* 72:
 3175-3186, 2012. doi: 10.1158/0008-5472.CAN-12-0481.
- 2910 194. Lee RJ, Smith MR. Targeting MET and vascular endothelial growth factor receptor signaling in
 2911 castration-resistant prostate cancer. *Cancer J.* 19:90-98, 2013. doi:
 2912 10.1097/PPO.0b013e318281e280.

- 2913 195. Lee YC, Cheng CJ, Bilen MA, Lu JF, Satcher RL, Yu-Lee LY, Gallick GE, Maity SN, Lin SH.
 2914 BMP4 promotes prostate tumor growth in bone through osteogenesis. *Cancer Res* 71: 51942915 5203, 2011. doi: 10.1158/0008-5472.CAN-10-4374.
- 196. Le Gall C, Bellahcène A, Bonnelye E, Gasser JA, Castronovo V, Green J, Zimmermann J,
 Clézardin P. A cathepsin K inhibitor reduces breast cancer induced osteolysis and skeletal
 tumor burden. *Cancer Res.* 67:9894-9902, 2007. doi: 10.1158/0008-5472.CAN-06-3940.
- Lemma S, Di Pompo G, Porporato PE, Sboarina M, Russell S, Gillies RJ, Baldini N, Sonveaux
 P, Avnet S. MDA-MB-231 breast cancer cells fuel osteoclast metabolism and activity: A new
 rationale for the pathogenesis of osteolytic bone metastases. *Biochim Biophys Acta Mol Basis Dis.* 1863:3254-3264, 2017. doi: 10.1016/j.bbadis.2017.08.030.
- 2923 198. Leto G. Activin A and bone metastasis. *J Cell Physiol.* 225:302-309, 2010. doi:
 2924 10.1002/jcp.22272.
- Lev DC, Kim SJ, Onn A, Stone V, Nam DH, Yazici S, Fidler IJ, Price JE. Inhibition of plateletderived growth factor receptor signaling restricts the growth of human breast cancer in the
 bone of nude mice. *Clin Cancer Res.* 11:306-314, 2005.
- 2928 200. Li XQ, Lu JT, Tan CC, Wang QS, Feng YM. RUNX2 promotes breast cancer bone metastasis
 2929 by increasing integrin α5-mediated colonization. *Cancer Lett* 380: 78-86, 2016. doi:
 2930 10.1016/j.canlet.2016.06.007.
- 201. Li ZG, Mathew P, Yang J, Starbuck MW, Zurita AJ, Liu J, Sikes C, Multani AS, Efstathiou E,
 Lopez A, Wang J, Fanning TV, Prieto VG, Kundra V, Vazquez ES, Troncoso P, Raymond AK,
 Logothetis CJ, Lin SH, Maity S, Navone NM. Androgen receptor-negative human prostate
 cancer cells induce osteogenesis in mice through FGF9-mediated mechanisms. *J Clin Invest*118: 2697-2710, 2008. doi: 10.1172/JCI33093.
- 2936 202. Li ZG, Yang J, Vazquez ES, Rose D, Vakar-Lopez F, Mathew P, Lopez A, Logothetis CJ, Lin
 2937 SH, Navone NM. Low-density lipoprotein receptor-related protein 5 (LRP5) mediates the

- 2938 prostate cancer-induced formation of new bone. *Oncogene* 27:596-603, 2008. doi:
 2939 10.1038/sj.onc.1210694.
- 203. Liao J, Li X, Koh AJ, Berry JE, Thudi N, Rosol TJ, Pienta KJ, McCauley LK. Tumor expressed
 PTHrP facilitates prostate cancer-induced osteoblastic lesions. *Int J Cancer* 123: 2267-78,
 2008. doi: 10.1002/ijc.23602.
- 204. Lin SC, Lee YC, Yu G, Cheng CJ, Zhou X, Chu K, Murshed M, Le NT, Baseler L, Abe JI,
 Fujiwara K, deCrombrugghe B, Logothetis CJ, Gallick GE, Yu-Lee LY, Maity SN, Lin SH.
 Endothelial-to-osteoblast conversion generates osteoblastic metastasis of prostate cancer. *Dev Cell* 41: 467-480, 2017. doi: 10.1016/j.devcel.2017.05.005.
- 2047 205. Lindemann F, Schlimok G, Dirschedl P, Witte J, Riethmüller G. Prognostic significance of
 micrometastatic tumour cells in bone marrow of colorectal cancer patients. Lancet. 340:685689, 1992. doi: 10.1016/0140-6736(92)92230-d.
- 2950 206. Lipton A, Smith MR, Fizazi K, Stopeck AT, Henry D, Brown JE, Shore ND, Saad F, Spencer A,
 2951 Zhu L, Warner DJ. Changes in Bone Turnover Marker Levels and Clinical Outcomes in
 2952 Patients with Advanced Cancer and Bone Metastases Treated with Bone Antiresorptive
 2953 Agents. *Clin Cancer Res.* 22:5713-5721, 2016. doi: 10.1158/1078-0432.CCR-15-3086.
- 207. Lipton A, Theriault RL, Hortobagyi GN, Simeone J, Knight RD, Mellars K, Reitsma DJ,
 Heffernan M, Seaman JJ. Pamidronate prevents skeletal complications and is effective
 palliative treatment in women with breast carcinoma and osteolytic bone metastases: long term
 follow-up of two randomized, placebo-controlled trials. *Cancer.* 88:1082-1090, 2000. doi:
 10.1002/(sici)1097-0142(20000301)88:5<1082::aid-cncr20>3.0.co;2-z.
- 2059 208. Liu Z, Liu H, He J, Lin P, Tong Q, Yang J. Myeloma cells shift osteoblastogenesis to
 adipogenesis by inhibiting the ubiquitin ligase MURF1 in mesenchymal stem cells. *Sci Signal.*13:eaay8203, 2020. doi: 10.1126/scisignal.aay8203.
- 209. Llombart A, Frassoldati A, Paija O, Sleeboom HP, Jerusalem G, Mebis J, Deleu I, Miller J,
 Schenk N, Neven P. Immediate Administration of Zoledronic Acid Reduces Aromatase

- Inhibitor-Associated Bone Loss in Postmenopausal Women With Early Breast Cancer: 12 month analysis of the E-ZO-FAST trial. *Clin Breast Cancer*. 12:40-48, 2012. doi:
 10.1016/j.clbc.2011.08.002.
- 2967 210. Logothetis CJ, Lin SH. Osteoblasts in prostate cancer metastasis to bone. *Nat Rev Cancer*2968 5:21-28, 2005. doi: 10.1038/nrc1528.
- 2969 211. López-Soto A, Gonzalez S, Smyth MJ, Galluzzi L. Control of Metastasis by NK Cells. Cancer
 2970 Cell. 32:135-154, 2017. doi:10.1016/j.ccell.2017.06.009
- 2971 212. Lotinun S, Pearsall RS, Davies MV, Marvell TH, Monnell TE, Ucran J, Fajardo RJ, Kumar R,
 2972 Underwood KW, Seehra J, Bouxsein ML, Baron R. A soluble activin receptor Type IIA fusion
 2973 protein (ACE-011) increases bone mass via a dual anabolic-antiresorptive effect in
 2974 Cynomolgus monkeys. *Bone.* 46:1082-1088, 2010. doi: 10.1016/j.bone.2010.01.370.
- 2975 213. Lu X, Kang Y. Chemokine (C-C motif) ligand 2 engages CCR2+ stromal cells of monocytic 2976 origin to promote breast cancer metastasis to lung and bone. *J Biol Chem.* 284:29087-29096,

2977 2009. doi: 10.1074/jbc.M109.035899.

2978 214. Lu X, Mu E, Wei Y, Riethdorf S, Yang Q, Yuan M, Yan J, Hua Y, Tiede BJ, Lu X, Haffty BG,
 2979 Pantel K, Massagué J, Kang Y. VCAM-1 promotes osteolytic expansion of indolent bone
 2980 micrometastasis of breast cancer by engaging α4β1-positive osteoclast progenitors. *Cancer* 2981 *Cell* 20: 701-714, 2011. doi: 10.1016/j.ccr.2011.11.002.

- 2982 215. Luo J, Yang Z, Ma Y, Yue Z, Lin H, Qu G, Huang J, Dai W, Li C, Zheng C, Xu L, Chen H,
 2983 Wang J, Li D, Siwko S, Penninger JM, Ning G, Xiao J, Liu M. LGR4 is a receptor for RANKL
 2984 and negatively regulates osteoclast differentiation and bone resorption. *Nat Med.* 22:539-546,
 2985 2016. doi: 10.1038/nm.4076.
- 2986 216. Luo X, Fu Y, Loza AJ, Murali B, Leahy KM, Ruhland MK, Gang M, Su X, Zamani A, Shi Y,
 2987 Lavine KJ, Ornitz DM, Weilbaecher KN, Long F, Novack DV, Faccio R, Longmore GD, Stewart

- SA. Stromal-Initiated Changes in the Bone Promote Metastatic Niche Development. *Cell Rep.*14:82-92, 2016. doi: 10.1016/j.celrep.2015.12.016.
- 2990 217. Luzzi KJ, MacDonald IC, Schmidt EE, Kerkvliet N, Morris VL, Chambers AF, Groom AC.
 2991 Multistep nature of metastatic inefficiency: dormancy of solitary cells after successful
 2992 extravasation and limited survival of early micrometastases. *Am J Pathol.* 153:865-873, 1998.
 2993 10.1016/S0002-9440(10)65628-3.
- 2994 218. Ma Y, Shurin GV, Peiyuan Z, Shurin MR. Dendritic cells in the cancer microenvironment. J
 2995 Cancer. 4:36-44, 2013. doi: 10.7150/jca.5046.
- 2996 219. Maeda H, Yonou H, Yano K, Ishii G, Saito S, Ochiai A. Prostate-specific antigen enhances
 2997 bioavailability of insulin-like growth factor by degrading insulin-like growth factor binding protein
 2998 5. *Biochem Biophys Res Commun* 381: 311-316, 2009. doi: 10.1016/j.bbrc.2009.01.096.
- 2999 220. Major PP, Cook RJ, Chen BL, Zheng M. Survival-adjusted multiple-event analysis for the 3000 evaluation of treatment effects of zoledronic Acid in patients with bone metastases from solid
- 3001 tumors. *Support Cancer Ther*. 2:234-240, 2005. doi: 10.3816/SCT.2005.n.017.
- 3002 221. Malanchi I, Santamaria-Martínez A, Susanto E, Peng H, Lehr HA, Delaloye JF, Huelsken J.
 3003 Interactions between cancer stem cells and their niche govern metastatic colonization. *Nature*
- 3004 481: 85-89, 2011. doi: 10.1038/nature10694.
- 3005 222. Manisterski M, Golan M, Amir S, Weisman Y, Mabjeesh NJ. Hypoxia induces PTHrP gene
 3006 transcription in human cancer cells through the HIF-2α. *Cell Cycle*. 9:3723-3729, 2010.
- 3007 223. Maroni P. Megakaryocytes in Bone Metastasis: Protection or Progression? Cells. 8: 2019. pii:
 3008 E134. doi: 10.3390/cells8020134.
- 3009 224. Maroni P, Bendinelli P, Matteucci E, Locatelli A, Nakamura T, Scita G, Desiderio MA.
- 3010 Osteolytic bone metastasis is hampered by impinging on the interplay among autophagy,
- 3011 anoikis and ossification. *Cell Death Dis* 5:e1005, 2014. doi: 10.1038/cddis.2013.465.

3012 225. Marshall JC, Collins JW, Nakayama J, Horak CE, Liewehr DJ, Steinberg SM, Albaugh M,
3013 Vidal-Vanaclocha F, Palmieri D, Barbier M, Murone M, Steeg PS. Effect of inhibition of the
3014 lysophosphatidic acid receptor 1 on metastasis and metastatic dormancy in breast cancer. J
3015 Natl Cancer Inst 104: 1306-1319, 2012. doi: 10.1093/jnci/djs319.

- 3016 226. McCaffrey G, Thompson ML, Majuta L, Fealk MN, Chartier S, Longo G, Mantyh PW. NGF
 3017 blockade at early times during bone cancer development attenuates bone destruction and
 3018 increases limb use. *Cancer Res.* 74:7014-7023, 2014. doi: 10.1158/0008-5472.CAN-14-1220.
- 3019 227. McDonald MM, Reagan MR, Youlten SE, Mohanty ST, Seckinger A, Terry RL, Pettitt JA, Simic
- 3020 MK, Cheng TL, Morse A, Le LMT, Abi-Hanna D, Kramer I, Falank C, Fairfield H, Ghobrial IM,
- 3021 Baldock PA, Little DG, Kneissel M, Vanderkerken K, Bassett JHD, Williams GR, Oyajobi BO,
- 3022 Hose D, Phan TG, Croucher PI. Inhibiting the osteocyte-specific protein sclerostin increases
- 3023 bone mass and fracture resistance in multiple myeloma. *Blood*. 129:3452-3464, 2017. doi:
 3024 10.1182/blood-2017-03-773341.
- 3025 228. Melchior SW, Corey E, Ellis WJ, Ross AA, Layton TJ, Oswin MM, Lange PH, Vessella RL.
 3026 Early tumor cell dissemination in patients with clinically localized carcinoma of the prostate.
 3027 *Clin Cancer Res.* 3:249-256, 1997.
- Mercadante S, Arcuri E. Breakthrough pain in cancer patients: pathophysiology and treatment.
 Cancer Treat Rev. 24:425-432, 1998. doi: 10.1016/s0305-7372(98)90005-6.
- 3030230.Meunier P, Aaron J, Edouard C, Vignon G. Osteoporosis and the replacement of cell3031populations of the marrow by adipose tissue. A quantitative study of 84 iliac bone biopsies.
- 3032 *Clin Orthop Relat Res.* 80:147-154, 1971. doi: 10.1097/00003086-197110000-00021.
- 2033 231. Miyamoto K, Yoshida S, Kawasumi M, Hashimoto K, Kimura T, Sato Y, Kobayashi T, Miyauchi
- 3034 Y, Hoshi H, Iwasaki R, Miyamoto H, Hao W, Morioka H, Chiba K, Kobayashi T, Yasuda H,
- 3035 Penninger JM, Toyama Y, Suda T, Miyamoto T. Osteoclasts are dispensable for hematopoietic

3036 stem cell maintenance and mobilization. J Exp Med 208: 2175-2181, 2011. doi:
 3037 10.1084/jem.20101890.

- 3038 232. Mizutani K, Sud S, McGregor NA, Martinovski G, Rice BT, Craig MJ, Varsos ZS, Roca H,
 3039 Pienta KJ. The chemokine CCL2 increases prostate tumor growth and bone metastasis
 3040 through macrophage and osteoclast recruitment. *Neoplasia*. 11:1235-1242, 2009. doi:
 3041 10.1593/neo.09988.
- 3042 233. Mowers EE, Sharifi MN, Macleod KF. Autophagy in cancer metastasis. *Oncogene*. 2017
 3043 36:1619-1630, 2017. doi: 10.1038/onc.2016.333.
- 3044 234. Mulcrone PL, Campbell JP, Clément-Demange L, Anbinder AL, Merkel AR, Brekken RA,
 3045 Sterling JA, Elefteriou F. Skeletal Colonization by Breast Cancer Cells Is Stimulated by an
 3046 Osteoblast and β2AR-Dependent Neo-Angiogenic Switch. *J Bone Miner Res.* 32:1442-1454,
 3047 2017. doi: 10.1002/jbmr.3133.
- 3048 235. Müller A, Homey B, Soto H, Ge N, Catron D, Buchanan ME, McClanahan T, Murphy E, Yuan
 3049 W, Wagner SN, Barrera JL, Mohar A, Verástegui E, Zlotnik A. Involvement of chemokine
 3050 receptors in breast cancer metastasis. *Nature* 410: 50-56, 2001. doi: 10.1038/35065016.
- 3051 236. Mundy GR. Metastasis to bone: causes, consequences and therapeutic opportunities. *Nat Rev* 3052 *Cancer.* 2:584-593, 2002. DOI: <u>10.1038/nrc867.</u>
- 3053 237. Murillo-Garzón V, Kypta R. WNT signalling in prostate cancer. *Nat Rev Urol.* 14:683-696,
 3054 2017. doi: 10.1038/nrurol.2017.144.
- 3055 238. Nakai Y, Okamoto K, Terashima A, Ehata S, Nishida J, Imamura T, Ono T, Takayanagi H.
- 3056 Efficacy of an orally active small-molecule inhibitor of RANKL in bone metastasis. *Bone Res.*
- 3057 7:1, 2019. doi: 10.1038/s41413-018-0036-5.
- 3058 239. Nandana S, Tripathi M, Duan P, Chu CY, Mishra R, Liu C, Jin R, Yamashita H, Zayzafoon M,
 3059 Bhowmick NA, Zhau HE, Matusik RJ, Chung LW. Bone Metastasis of Prostate Cancer Can Be

3060 Therapeutically Targeted at the TBX2-WNT Signaling Axis. *Cancer Res* 77:1331-1344, 2017.

3061 doi: 10.1158/0008-5472.CAN-16-0497.

- 240. Nelson JB, Fizazi K, Miller K, Higano C, Moul JW, Akaza H, Morris T, McIntosh S, Pemberton
 K, Gleave M. Phase 3, randomized, placebo-controlled study of zibotentan (ZD4054) in
 patients with castration-resistant prostate cancer metastatic to bone. *Cancer.* 118:5709-18,
- 3065 2012. doi: 10.1002/cncr.27674.
- 3066 241. Nguyen DX, Bos PD, Massagué J. Metastasis: from dissemination to organ-specific
 3067 colonization. *Nat Rev Cancer* 9: 274-284, 2009. doi: 10.1038/nrc2622.
- Nilsson S, Larsen RH, Fossa SD. First clinical experience with a-emitting radium-223 in the
 treatment of skeletal metastases. *Clin Cancer Res.* 11:4451-4459, 2005.
- 3070 243. Nutter F, Holen I, Brown HK, Cross SS, Evans CA, Walker M, Coleman RE, Westbrook JA,
- 3071 Selby PJ, Brown JE, Ottewell PD. Different molecular profiles are associated with breast 3072 cancer cell homing compared with colonisation of bone: evidence using a novel bone-seeking 3073 cell line. *Endocr Relat Cancer.* 21:327-341, 2014. doi: 10.1530/ERC-13-0158.
- 3074 244. O'Donnell A, Judson I, Dowsett M, Raynaud F, Dearnaley D, Mason M, Harland S, Robbins A,
- 3075 Halbert G, Nutley B, Jarman M. Hormonal impact of the 17alpha-hydroxylase/C(17,20)-lyase
- 3076 inhibitor abiraterone acetate (CB7630) in patients with prostate cancer. Br J Cancer. 90:2317-
- 3077 2325, 2004. doi: 10.1038/sj.bjc.6601879.
- 3078 245. Okamoto K, Nakashima T, Shinohara M, Negishi-Koga T, Komatsu N, Terashima A, Sawa S,
- 3079 Nitta T, Takayanagi H. Osteoimmunology: The Conceptual Framework Unifying the Immune
 3080 and Skeletal Systems. *Physiol Rev* 97: 1295-1349, 2017. doi: 10.1152/physrev.00036.
- 3081 246. Ono M, Kosaka N, Tominaga N, Yoshioka Y, Takeshita F, Takahashi RU, Yoshida M, Tsuda
 3082 H, Tamura K, Ochiya T. Exosomes from bone marrow mesenchymal stem cells contain a
- 3083 microRNA that promotes dormancy in metastatic breast cancer cells. *Sci Signal*. 7:ra63, 2014.
- doi: 10.1126/scisignal.2005231.

3085	247.	Oskarsson	Τ,	Batlle	Ε,	Massagué	J.	Metastatic	stem	cells:	sources,	niches,	and	vital
3086	pathways. Cell Stem Cell 14: 306-321, 2014. doi: 10.1016/j.stem.2014.02.002.													

- 3087 248. Ottewell PD. The role of osteoblasts in bone metastasis. *J Bone Oncol* 5: 124-127, 2016. doi:
 3088 10.1016/j.jbo.2016.03.007.
- Ottewell PD, Wang N, Brown HK, Fowles CA, Croucher PI, Eaton CL, Holen I. OPG-Fc inhibits
 ovariectomy-induced growth of disseminated breast cancer cells in bone. *Int J Cancer*137:968-977, 2015. doi: 10.1002/ijc.29439.
- 3092 250. Ottewell PD, Wang N, Brown HK, Reeves KJ, Fowles CA, Croucher PI, Eaton CL, Holen I.
 3093 Zoledronic acid has differential antitumor activity in the pre- and postmenopausal bone
 3094 microenvironment in vivo. *Clin Cancer Res* 20: 2922-2932, 2014. doi: 10.1158/1078 3095 0432.CCR-13-1246.
- 3096 251. Ottewell PD, Wang N, Meek J, Fowles CA, Croucher PI, Eaton CL, Holen I. Castration-induced
 3097 bone loss triggers growth of disseminated prostate cancer cells in bone. *Endocr Relat Cancer*3098 21: 769-781, 2014. doi: 10.1530/ERC-14-0199.
- 2099 252. Ouellet V, Tiedemann K, Mourskaia A, Fong JE, Tran-Thanh D, Amir E, Clemons M, Perbal B,

3100 Komarova SV, Siegel PM. CCN3 impairs osteoblast and stimulates osteoclast differentiation to

- favor breast cancer metastasis to bone. Am J Pathol 178: 2377-2388, 2011. doi:
- 3102 10.1016/j.ajpath.2011.01.033.
- 3103 253. Owen KL, Parker BS. Beyond the vicious cycle: The role of innate osteoimmunity, automimicry
 and tumor-inherent changes in dictating bone metastasis. *Mol Immunol.* 110: 57-68, 2019. doi:
 10.1016/j.molimm.2017.11.023.
- Pantano F, Rossi E, Iuliani M, Facchinetti A, Simonetti S, Ribelli G, Zoccoli A, Vincenzi B,
 Tonini G, Zamarchi R, Santini D. Dynamic changes of Receptor activator of nuclear factor-κB
 expression in Circulating Tumor Cells during Denosumab predict treatment effectiveness in
- 3109 Metastatic Breast Cancer. *Sci Rep.* 10:1288, 2020. doi: 10.1038/s41598-020-58339-2.

3110 255. Pantano F, Croset M, Driouch K, Bednarz-Knoll N, Iuliani M, Ribelli G, Bonnelye E, Wilkman H,

3111 Geraci S, Bonin F, Simonetti S, Vincenzi B, Hong SS, Sousa S, Pantel K, Tonini G, Santini D,

3112 Clézardin P. Integrin alpha-5 in human breast cancer is a mediator of bone metastasis and a 3113 therapeutic target for the treatment of osteolytic lesions. *Oncogene* (accepted for publication).

- 256. Parfitt AM, Mundy GR, Roodman GD, Hughes DE, Boyce BF. A new model for the regulation
 of bone resorption with particular reference to the effects of bisphosphonates. *J Bone Min Res.*11:150-159, 1996.
- 257. Park SI, Lee C, Sadler WD, Koh AJ, Jones J, Seo JW, Soki FN, Cho SW, Daignault SD,
 McCauley LK. Parathyroid hormone-related protein drives a CD11b+Gr1+ cell-mediated
 positive feedback loop to support prostate cancer growth. Cancer Res. 73:6574-6583, 2013.
 doi: 10.1158/0008-5472.CAN-12-4692.
- 258. Parker C, Nilsson S, Heinrich D, Helle SI, O'Sullivan JM, Fosså SD, Chodacki A, Wiechno P,
 Logue J, Seke M, Widmark A, Johannessen DC, Hoskin P, Bottomley D, James ND, Solberg
 A, Syndikus I, Kliment J, Wedel S, Boehmer S, Dall'Oglio M, Franzén L, Coleman R,
 Vogelzang NJ, O'Bryan-Tear CG, Staudacher K, Garcia-Vargas J, Shan M, Bruland ØS, Sartor
 O; ALSYMPCA Investigators. Alpha emitter radium-223 and survival in metastatic prostate
 cancer. *N Engl J Med.* 369:213-223, 2013. doi: 10.1056/NEJMoa1213755.
- 3127259.Pavlovic M, Arnal-Estapé A, Rojo F, Bellmunt A, Tarragona M, Guiu M, Planet E, Garcia-3128Albéniz X, Morales M, Urosevic J, Gawrzak S, Rovira A, Prat A, Nonell L, Lluch A, Jean-Mairet3129J, Coleman R, Albanell J, Gomis RR. Enhanced MAF Oncogene Expression and Breast
- 3130 Cancer Bone Metastasis. J Natl Cancer Inst. 107:djv256, 2015. doi: 10.1093/jnci/djv256.
- 260. Pécheur I, Peyruchaud O, Serre CM, Guglielmi J, Voland C, Bourre F, Margue C, Cohen-Solal
 M, Buffet A, Kieffer N, Clézardin P. Integrin alpha(v)beta3 expression confers on tumor cells a
- 3133 greater propensity to metastasize to bone. FASEB J 16:1266-1268, 2002. doi: 10.1096/fj.01-
- 3134 0911fje.

- 3135 261. Peinado H, Zhang H, Matei IR, Costa-Silva B, Hoshino A, Rodrigues G, Psaila B, Kaplan RN, 3136 Bromberg JF, Kang Y, Bissell MJ, Cox TR, Giaccia AJ, Erler JT, Hiratsuka S, Ghajar CM, 3137 Lyden D. Pre-metastatic niches: organ-specific homes for metastases. Nat Rev Cancer 17: 3138 302-317, 2017. doi: 10.1038/nrc.2017.6.
- 3139 262. Peterson LM, O'Sullivan J, Wu QV, Novakova-Jiresova A, Jenkins I, Lee JH, Shields A, 3140 Montgomery S, Linden HM, Gralow JR, Gadi VK, Muzi M, Kinahan PE, Mankoff DA, Specht 3141 JM. Prospective study of serial 18F-FDG PET and 18F-fluoride (18F-NaF) PET to predict time 3142 to skeletal related events, time-to-progression, and survival in patients with bone-dominant 3143 metastatic breast cancer. J Nucl Med 59: 1823-1830, 2018.doi: 10.2967/jnumed.118.211102.
- 3144 263. Pez F, Dayan F, Durivault J, Kaniewski B, Aimond G, Le Provost GS, Deux B, Clézardin P, 3145 Sommer P, Pouysségur J, Reynaud C. The HIF-1-inducible lysyl oxidase activates HIF-1 via 3146 the Akt pathway in a positive regulation loop and synergizes with HIF-1 in promoting tumor cell 3147 growth. Cancer Res 71: 1647-1657, 2011. doi: 10.1158/0008-5472.CAN-10-1516.
- 3148 264. Pham JT, Kim RC, Nguyen A, Bota D, Kong XT, Vadera S, Hsu F, Carrillo JA. Intracranial 3149 meningioma with carcinoma tumor-to-tumor metastasis: two case reports. CNS Oncol. 3150
- 7:CNS09, 2018. doi: 10.2217/cns-2017-0022.
- 3151 265. Powell GJ, Southby J, Danks JA, Stillwell RG, Hayman JA, Henderson MA, Bennett RC, Martin
- 3152 TJ. Localization of parathyroid hormone-related protein in breast cancer metastases: increased 3153 incidence in bone compared with other sites. *Cancer Res* 51:3059-3061, 1991.
- 3154 266. Pratap J, Lian JB, Stein GS. Metastatic bone disease: role of transcription factors and future 3155 targets. Bone 48: 30-36, 2011. doi: 10.1016/j.bone.2010.05.035.
- 3156 267. Price TT, Burness ML, Sivan A, Warner MJ, Cheng R, Lee CH, Olivere L, Comatas K, Magnani
- 3157 J, Kim Lyerly H, Cheng Q, McCall CM, Sipkins DA. Dormant breast cancer micrometastases
- 3158 reside in specific bone marrow niches that regulate their transit to and from bone. Sci Transl
- 3159 Med 8: 340ra73, 2016. doi: 10.1126/scitranslmed.aad4059.

- 268. Puisieux A, Brabletz T, Caramel J. Oncogenic roles of EMT-inducing transcription factors. *Nat Cell Biol* 16:488-494, 2014. doi: 10.1038/ncb2976.
- Rao S, Cronin SJF, Sigl V, Penninger JM. RANKL and RANK: From Mammalian Physiology to
 Cancer Treatment. *Trends Cell Biol.* 28:213-223, 2018. doi: 10.1016/j.tcb.2017.11.001.
- 270. Ravnik J, Ravnik M, Bunc G, Glumbic I, Tobi-Veres E, Velnar T. Metastasis of an occult
 pulmonary carcinoma into meningioma: a case report. World J Surg Oncol 13:292, 2015. doi:
 10.1186/s12957-015-0714-3
- Reagan MR, Rosen CJ. Navigating the bone marrow niche: translational insights and cancerdriven dysfunction. *Nature Rev Rheum* 12: 154-168, 2016. doi: 10.1038/nrrheum.2015.160.
- 3169 272. Reyes ME, Fujii T, Branstetter D, Krishnamurthy S, Masuda H, Wang X, Reuben JM,
- 3170 Woodward WA, Edwards BJ, Hortobagyi GN, Tripathy D, Dougall WC, Eckhardt BL, Ueno NT.
- 3171 Poor prognosis of patients with triple-negative breast cancer can be stratified by RANK and
- 3172 RANKL dual expression. Breast Cancer Res Treat 164: 57-67, 2017. doi: 10.1007/s10549-
- 3173 017-4233-5.
- Reymond N, d'Água BB, Ridley AJ. Crossing the endothelial barrier during metastasis. *Nat Rev Cancer* 13: 858-870, 2013. doi: 10.1038/nrc3628.
- 3176 274. Reynaud C, Ferreras L, Di Mauro P, Kan C, Croset M, Bonnelye E, Pez F, Thomas C, Aimond
 3177 G, Karnoub AE, Brevet M, Clézardin P. Lysyl Oxidase Is a Strong Determinant of Tumor Cell
- 3178 Colonization in Bone. *Cancer Res* 77: 268-278, 2017. doi: 10.1158/0008-5472.CAN-15-2621.
- Romero-Moreno R, Curtis KJ, Coughlin TR, Miranda-Vergara MC, Dutta S, Natarajan A,
 Facchine BA, Jackson KM, Nystrom L, Li J, Kaliney W, Niebur GL, Littlepage LE. The
 CXCL5/CXCR2 axis is sufficient to promote breast cancer colonization during bone
 metastasis. *Nat Commun* 10: 4404, 2019. doi: 10.1038/s41467-019-12108-6.
- 3183 276. Rosanò L, Spinella F, Bagnato A. Endothelin 1 in cancer: biological implications and
 3184 therapeutic opportunities. *Nat Rev Cancer*. 13:637-651, 2013. doi: 10.1038/nrc3546.

Rosen LS, Gordon D, Kaminski M, Howell A, Belch A, Mackey J, Apffelstaedt J, Hussein MA,
Coleman RE, Reitsma DJ, Chen BL, Seaman JJ. Long-term efficacy and safety of zoledronic
acid compared with pamidronate disodium in the treatment of skeletal complications in patients
with advanced multiple myeloma or breast carcinoma: a randomized, double-blind, multicenter,
comparative trial. *Cancer.* 98:1735-1744, 2003. doi: 10.1002/cncr.11701.

- 278. Rosen LS, Gordon D, Kaminski M, Howell A, Belch A, Mackey J, Apffelstaedt J, Hussein M,
 Coleman RE, Reitsma DJ, Seaman JJ, Chen BL, Ambros Y. Zoledronic acid versus
 pamidronate in the treatment of skeletal metastases in patients with breast cancer or osteolytic
 lesions of multiple myeloma: a phase III, double-blind, comparative trial. *Cancer J.* 7:377-387,
 2001.
- Rosen LS, Gordon D, Tchekmedyian NS, Yanagihara R, Hirsh V, Krzakowski M, Pawlicki M,
 De Souza P, Zheng M, Urbanowitz G, Reitsma D, Seaman J. Long-term efficacy and safety of
 zoledronic acid in the treatment of skeletal metastases in patients with nonsmall cell lung
 carcinoma and other solid tumors: a randomized, Phase III, double-blind, placebo-controlled
 trial. *Cancer.* 100:2613-2621, 2004. doi: 10.1002/cncr.20308.
- Rosen LS, Gordon D, Tchekmedyian S, Yanagihara R, Hirsh V, Krzakowski M, Pawlicki M, de
 Souza P, Zheng M, Urbanowitz G, Reitsma D, Seaman JJ. Zoledronic acid versus placebo in
 the treatment of skeletal metastases in patients with lung cancer and other solid tumors: a
 phase III, double-blind, randomized trial--the Zoledronic Acid Lung Cancer and Other Solid
 Tumors Study Group. *J Clin Oncol.* 21:3150-3157, 2003. DOI: 10.1200/JCO.2003.04.105.
- 3205 281. Roskoski R Jr. Src protein-tyrosine kinase structure, mechanism, and small molecule
 inhibitors. *Pharmacol Res.* 94:9-25, 2015. doi: 10.1016/j.phrs.2015.01.003.
- Rossnagl S, Ghura H, Groth C, Altrock E, Jakob F, Schott S, Wimberger P, Link T, Kuhlmann
 JD, Stenzl A, Hennenlotter J, Todenhöfer T, Rojewski M, Bieback K, Nakchbandi IA. A

- 3209 Subpopulation of Stromal Cells Controls Cancer Cell Homing to the Bone Marrow. Cancer Res
 3210 78: 129-142, 2018. doi: 10.1158/0008-5472.CAN-16-3507.
- 3211 283. Saad F, Gleason DM, Murray R, Tchekmedyian S, Venner P, Lacombe L, Chin JL, Vinholes
 3212 JJ, Goas JA, Zheng M; Zoledronic Acid Prostate Cancer Study Group. Long-term efficacy of
 3213 zoledronic acid for the prevention of skeletal complications in patients with metastatic
 3214 hormone-refractory prostate cancer. *J Natl Cancer Inst.* 96:879-882, 2004. doi:
 3215 10.1093/jnci/djh141.
- 3216 284. Sahay D, Leblanc R, Grunewald TG, Ambatipudi S, Ribeiro J, Clézardin P, Peyruchaud O. The
 3217 LPA1/ZEB1/miR-21-activation pathway regulates metastasis in basal breast cancer.
 3218 Oncotarget 6:20604-20620, 2015. doi: 10.18632/oncotarget.3774.
- 3219 285. San Martin R, Pathak R, Jain A, Jung SY, Hilsenbeck SG, Piña-Barba MC, Sikora AG, Pienta
 3220 KJ, Rowley DR. Tenascin-C and Integrin α9 Mediate Interactions of Prostate Cancer with the
 Bone Microenvironment. *Cancer Res* 77: 5977-5988, 2017. doi: 10.1158/0008-5472.CAN-173222 0064.
- 3223 286. Santini D, Schiavon G, Vincenzi B, Gaeta L, Pantano F, Russo A, Ortega C, Porta C, Galluzzo
 S, Armento G, La Verde N, Caroti C, Treilleux I, Ruggiero A, Perrone G, Addeo R, Clézardin P,
 Muda AO, Tonini G. Receptor activator of NF-kB (RANK) expression in primary tumors
 associates with bone metastasis occurrence in breast cancer patients. *PLoS One* 6:e19234,
- 3227 2011. doi: 10.1371/journal.pone.0019234.
- 3228 287. Sato MM, Nakashima A, Nashimoto M, Yawaka Y, Tamura M. Bone morphogenetic protein-2
 enhances Wnt/beta-catenin signaling-induced osteoprotegerin expression. *Genes Cells* 14:
 141-153, 2009. doi: 10.1111/j.1365-2443.2008.01258.x.
- 3231 288. Sawant A, Deshane J, Jules J, Lee CM, Harris BA, Feng X, Ponnazhagan S. Myeloid-derived
 suppressor cells function as novel osteoclast progenitors enhancing bone loss in breast
 cancer. *Cancer Res* 73: 672-82, 2013. doi: 10.1158/0008-5472.

3234 289. Sawant A, Hensel JA, Chanda D, Harris BA, Siegal GP, Maheshwari A, Ponnazhagan S.
3235 Depletion of plasmacytoid dendritic cells inhibits tumor growth and prevents bone metastasis
3236 of breast cancer cells. *J Immunol.* 189:4258-4265, 2012. doi: 10.4049/jimmunol.1101855.

- Scalzi P, Baiocco C, Genovese S, Trevisan A, Sirotova Z, Poti C. Evaluation of bone
 metastases by 18F-choline PET/CT in a patient with castration-resistant prostate cancer
 treated with radium-223. *Urologia*. 84:61-64, 2017. doi: 10.5301/uro.5000206.
- Schild T, Low V, Blenis J, Gomes AP. Unique Metabolic Adaptations Dictate Distal OrganSpecific Metastatic Colonization. *Cancer Cell.* 33:347-354, 2018. doi:
 10.1016/j.ccell.2018.02.001.
- Schoemaker MJ, Jones ME, Wright LB, Griffin J2, McFadden E, Ashworth A, Swerdlow AJ.
 Psychological stress, adverse life events and breast cancer incidence: a cohort investigation in
 106,000 women in the United Kingdom. *Breast Cancer Res.* 18:72, 2016. doi:
 10.1186/s13058-016-0733-1.
- 3247 293. Secondini C, Wetterwald A, Schwaninger R, Thalmann GN, Cecchini MG. The role of the BMP
 3248 signaling antagonist noggin in the development of prostate cancer osteolytic bone metastasis.
- 3249 *PLoS One*. 6:e16078, 2011. doi: 10.1371/journal.pone.0016078.
- Seeliger C, Karpinski K, Haug AT, Vester H, Schmitt A, Bauer JS, van Griensven M. Five freely
 circulating miRNAs and bone tissue miRNAs are associated with osteoporotic fractures. *J Bone Miner Res.* 29:1718-1728, 2014. doi: 10.1002/jbmr.2175.
- 295. Sethi N, Dai X, Winter CG, Kang Y. Tumor-derived JAGGED1 promotes osteolytic bone metastasis of breast cancer by engaging notch signaling in bone cells. *Cancer Cell.* 19: 192-
- 3255 205, 2011. doi: 10.1016/j.ccr.2010.12.022.
- Shahinian VB, Kuo YF, Freeman JL, Goodwin JS. Risk of fracture after androgen deprivation
 for prostate cancer. *N Engl J Med.* 352:154-164, 2005. doi: 10.1056/NEJMoa041943.

Shao G, Gu W, Guo M, Zang S, Fu J, Liu S, Wang F, Wang Z. Clinical study of (99m)Tc-3PRGD2 peptide imaging in osteolytic bone metastasis. *Oncotarget*. 8:75587-75596, 2017. doi:
10.18632/oncotarget.17486.

- Shiozawa Y, Pedersen EA, Havens AM, Jung Y, Mishra A, Joseph J, Kim JK, Patel LR, Ying
 C, Ziegler AM, Pienta MJ, Song J, Wang J, Loberg RD, Krebsbach PH, Pienta KJ, Taichman
 RS. Human prostate cancer metastases target the hematopoietic stem cell niche to establish
 footholds in mouse bone marrow. *J Clin Invest* 121: 1298-1312, 2011. doi: 10.1172/JCl43414.
- 3265 299. Siclari VA, Mohammad KS, Tompkins DR, Davis H, McKenna CR, Peng X, Wessner LL,
 3266 Niewolna M, Guise TA, Suvannasankha A, Chirgwin JM. Tumor-expressed adrenomedullin
 3267 accelerates breast cancer bone metastasis. *Breast Cancer Res.* 16: 458, 2014. doi:
 3268 10.1186/s13058-014-0458-y.
- 3269 300. Singh S, Singh R, Sharma PK, Singh UP, Rai SN, Chung LW, Cooper CR, Novakovic KR, 3270 Grizzle WE, Lillard JW Jr. Serum CXCL13 positively correlates with prostatic disease, prostate-3271 specific antigen and mediates prostate cancer cell invasion, integrin clustering and cell 3272 adhesion. *Cancer Lett.* 283:29-35, 2009. doi: 10.1016/j.canlet.2009.03.022.
- 3273 301. Sipkins DA, Wei X, Wu JW, Runnels JM, Côté D, Means TK, Luster AD, Scadden DT, Lin CP.
- In vivo imaging of specialized bone marrow endothelial microdomains for tumour engraftment.
 Nature 435: 969-973, 2005. doi: 10.1038/nature03703.
- 3276 302. Sleeman JP. The lymph node pre-metastatic niche. *J Mol Med (Berl*). 93:1173-84, 2015. doi:
 3277 10.1007/s00109-015-1351-6.
- 303. Sloan EK, Pouliot N, Stanley KL, Chia J, Moseley JM, Hards DK, Anderson RL. Tumor-specific
 expression of alphavbeta3 integrin promotes spontaneous metastasis of breast cancer to
 bone. *Breast Cancer Res* 8: R20, 2006. doi: 10.1186/bcr1398.
- 304. Smith M, De Bono J, Sternberg C, Le Moulec S, Oudard S, De Giorgi U, Krainer M, Bergman
 A, Hoelzer W, De Wit R, Bögemann M, Saad F, Cruciani G, Thiery-Vuillemin A, Feyerabend S,

Miller K, Houédé N, Hussain S, Lam E, Polikoff J, Stenzl A, Mainwaring P, Ramies D, Hessel
C, Weitzman A, Fizazi K. Phase III Study of Cabozantinib in Previously Treated Metastatic
Castration-Resistant Prostate Cancer: COMET-1. *J Clin Oncol.* 34:3005-3013, 2016. doi:
10.1200/JCO.2015.65.5597.

- 305. Smith MR, Egerdie B, Hernández Toriz N, Feldman R, Tammela TL, Saad F, Heracek J,
 Szwedowski M, Ke C, Kupic A, Leder BZ, Goessl C; Denosumab HALT Prostate Cancer Study
 Group. Denosumab in men receiving androgen-deprivation therapy for prostate cancer. *N Engl J Med.* 361:745-755, 2009. doi: 10.1056/NEJMoa0809003.
- 306. Smith MR, Saad F, Coleman R, Shore N, Fizazi K, Tombal B, Miller K, Sieber P, Karsh L,
 Damião R, Tammela TL, Egerdie B, Van Poppel H, Chin J, Morote J, Gómez-Veiga F,
 Borkowski T, Ye Z, Kupic A, Dansey R, Goessl C. Denosumab and bone-metastasis-free
 survival in men with castration-resistant prostate cancer: results of a phase 3, randomised,
 placebo-controlled trial. *Lancet.* 379:39-46, 2012. doi: 10.1016/S0140-6736(11)61226-9.
- 3296 307. Solomayer EF, Gebauer G, Hirnle P, Janni W, Lück HJ, Becker S, Huober J, Krämer B,
 3297 Wackwitz B, Wallwiener D, Fehm T. Influence of zoledronic acid on disseminated tumor cells in

3298 primary breast cancer patients. *Ann Oncol.* 23:2271-2277, 2012. doi: 10.1093/annonc/mdr612.

- 3299 308. Sopata M, Katz N, Carey W, Smith MD, Keller D, Verburg KM, West CR, Wolfram G, Brown
- MT. Efficacy and safety of tanezumab in the treatment of pain from bone metastases. *Pain*.
 156:1703-1713, 2015. doi: 10.1097/j.pain.0000000000211.
- 309. Sosa MS, Bragado P, Aguirre-Ghiso JA. Mechanisms of disseminated cancer cell dormancy:
 an awakening field. *Nat Rev Cancer* 14:611-622, 2014. doi: 10.1038/nrc3793.
- 3304 310. Sottnik JL, Dai J, Zhang H, Campbell B, Keller ET. Tumor-induced pressure in the bone
 microenvironment causes osteocytes to promote the growth of prostate cancer bone
 metastases. *Cancer Res.* 75:2151-2158, 2015. doi: 10.1158/0008-5472.CAN-14-2493.

- 3307 311. Sottnik JL, Daignault-Newton S, Zhang X, Morrissey C, Hussain MH, Keller ET, Hall CL.
 3308 Integrin alpha2beta 1 (α2β1) promotes prostate cancer skeletal metastasis. *Clin Exp* 3309 *Metastasis* 30:569-578, 2013. doi: 10.1007/s10585-012-9561-6.
- 3310 312. Sousa S, Clézardin P. Bone-Targeted Therapies in Cancer-Induced Bone Disease. *Calcif* 3311 *Tissue Int.* 102:227-250, 2018. doi: 10.1007/s00223-017-0353-5.
- 3312 313. Spencer JA, Ferraro F, Roussakis E, Klein A, Wu J, Runnels JM, Zaher W, Mortensen LJ, Alt
- C, Turcotte R, Yusuf R, Côté D, Vinogradov SA, Scadden DT, Lin CP. Direct measurement of
 local oxygen concentration in the bone marrow of live animals. *Nature*. 508:269-273, 2014. doi:
 10.1038/nature13034.
- 3316 314. Sterling JA, Oyajobi BO, Grubbs B, Padalecki SS, Munoz SA, Gupta A, Story B, Zhao M,
 3317 Mundy GR. The hedgehog signaling molecule Gli2 induces parathyroid hormone-related
 3318 peptide expression and osteolysis in metastatic human breast cancer cells. *Cancer Res.*3319 66:7548-7553, 2006. doi: 10.1158/0008-5472.CAN-06-0452.
- 3320 315. Stopeck AT, Lipton A, Body JJ, Steger GG, Tonkin K, de Boer RH, Lichinitser M, Fujiwara Y,
 3321 Yardley DA, Viniegra M, Fan M, Jiang Q, Dansey R, Jun S, Braun A. Denosumab compared
 3322 with zoledronic acid for the treatment of bone metastases in patients with advanced breast
 3323 cancer: a randomized, double-blind study. *J Clin Oncol.* 28:5132-5139, 2010. doi:
 3324 10.1200/JCO.2010.29.7101.
- 3325 316. Sugiyama T, H. Kohara, M. Noda, T. Nagasawa. Maintenance of the hematopoietic stem cell
 pool by CXCL12-CXCR4 chemokine signaling in bone marrow stromal cell niches *Immunity* 25:
 977-988, 2006. doi: 10.1016/j.immuni.2006.10.016.
- 317. Sun YX, Fang M, Wang J, Cooper CR, Pienta KJ, Taichman RS. Expression and activation of
 alpha v beta 3 integrins by SDF-1/CXC12 increases the aggressiveness of prostate cancer
 cells. *Prostate* 67: 61-73, 2007. doi: 10.1002/pros.20500.

- 318. Sun YX, Schneider A, Jung Y, Wang J, Dai J, Wang J, Cook K, Osman NI, Koh-Paige AJ,
 Shim H, Pienta KJ, Keller ET, McCauley LK, Taichman RS. Skeletal localization and
 neutralization of the SDF-1(CXCL12)/CXCR4 axis blocks prostate cancer metastasis and
 growth in osseous sites in vivo. *J Bone Miner Res* 20: 318-329, 2005. doi:
 10.1359/JBMR.041109.
- 319. Suominen MI, Fagerlund KM, Rissanen JP, Konkol YM, Morko JP, Peng Z, Alhoniemi EJ,
 Laine SK, Corey E, Mumberg D, Ziegelbauer K, Käkönen SM, Halleen JM, Vessella RL,
 Scholz A. Radium-223 Inhibits Osseous Prostate Cancer Growth by Dual Targeting of Cancer
 Cells and Bone Microenvironment in Mouse Models. *Clin Cancer Res.* 23:4335-4346, 2017.
 doi: 10.1158/1078-0432.CCR-16-2955.
- 3341 320. Taichman RS, Patel LR, Bedenis R, Wang J, Weidner S, Schumann T, Yumoto K, Berry JE,
 3342 Shiozawa Y, Pienta KJ. GAS6 receptor status is associated with dormancy and bone
 3343 metastatic tumor formation. *PLoS One* 8:e61873, 2013. doi: 10.1371/journal.pone.0061873.
- 321. Taipaleenmäki H, Browne G, Akech J, Zustin J, van Wijnen AJ, Stein JL, Hesse E, Stein GS,
 Lian JB. Targeting of Runx2 by miR-135 and miR-203 impairs progression of breast cancer
 and metastatic bone disease. *Cancer Res* 75:1433-1444, 2015. doi: 10.1158/0008-5472.CAN14-1026.
- 3348 322. Taipaleenmäki H, Farina NH, van Wijnen AJ, Stein JL, Hesse E, Stein GS, Lian JB.
 Antagonizing miR-218-5p attenuates Wnt signaling and reduces metastatic bone disease of
 triple negative breast cancer cells. *Oncotarget* 7: 79032-79046, 2016. doi:
 10.18632/oncotarget.12593.
- 3352 323. Tamura D, Hiraga T, Myoui A, Yoshikawa H, Yoneda T. Cadherin-11-mediated interactions
 with bone marrow stromal/osteoblastic cells support selective colonization of breast cancer
 cells in bone. *Int J Oncol* 33: 17-24, 2008.

- 3355 324. Tandon M, Othman AH, Ashok V, Stein GS, Pratap J. The role of Runx2 in facilitating
 autophagy in metastatic breast cancer cells. *J Cell Physiol.* 233:559-571, 2018. doi:
 10.1002/jcp.25916.
- 3358 325. Tang ZN, Zhang F, Tang P, Qi XW, Jiang J. Hypoxia induces RANK and RANKL expression
 by activating HIF-1α in breast cancer cells. *Biochem Biophys Res Commun.* 408:411-416,
 2011. doi: 10.1016/j.bbrc.2011.04.035.
- 3361 326. Tarragona M, Pavlovic M, Arnal-Estapé A, Urosevic J, Morales M, Guiu M, Planet E,
 3362 González-Suárez E, Gomis RR. Identification of NOG as a specific breast cancer bone
 3363 metastasis-supporting gene. *J Biol Chem.* 287(25):21346-21355, 2012. doi:
 3364 10.1074/jbc.M112.355834.
- 3365 327. Taverna S, Pucci M, Giallombardo M, Di Bella MA, Santarpia M, Reclusa P, Gil-Bazo I, Rolfo
 C, Alessandro R. Amphiregulin contained in NSCLC-exosomes induces osteoclast
 differentiation through the activation of EGFR pathway. *Sci Rep.* 7:3170, 2017. doi:
 10.1038/s41598-017-03460-y.
- 3369 328. Templeton ZS, Lie WR, Wang W, Rosenberg-Hasson Y, Alluri RV, Tamaresis JS, Bachmann 3370 MH, Lee K, Maloney WJ, Contag CH, King BL. Breast Cancer Cell Colonization of the Human 3371 Bone Marrow Adipose Tissue Niche. Neoplasia. 17:849-861. 2015. doi: 10.1016/i.neo.2015.11.005. 3372
- 3373 329. Thomas RJ, Guise TA, Yin JJ, Elliott J, Horwood NJ, Martin TJ, Gillespie MT. Breast cancer
 cells interact with osteoblasts to support osteoclast formation. *Endocrinology* 140: 4451-4458,
 1999. doi: 10.1210/endo.140.10.7037.
- 3376 330. Thudi NK, Martin CK, Murahari S, Shu ST, Lanigan LG, Werbeck JL, Keller ET, McCauley LK,
- 3377 Pinzone JJ, Rosol TJ. Dickkopf-1 (DKK-1) stimulated prostate cancer growth and metastasis
- 3378 and inhibited bone formation in osteoblastic bone metastases. *Prostate*. 71:615-625, 2011. doi:
- 3379 10.1002/pros.21277.

- 3380 331. Thysell E, Surowiec I, Hörnberg E, Crnalic S, Widmark A, Johansson AI, Stattin P, Bergh A,
 3381 Moritz T, Antti H, Wikström P. Metabolomic characterization of human prostate cancer bone
 3382 metastases reveals increased levels of cholesterol. *PLoS One.* 5:e14175, 2010. doi:
 3383 10.1371/journal.pone.0014175.
- 3384 332. Tian E, Zhan F, Walker R, Rasmussen E, Ma Y, Barlogie B, Shaughnessy JD Jr. The role of
 the Wnt-signaling antagonist DKK1 in the development of osteolytic lesions in multiple
 myeloma. *N Engl J Med.* 349:2483-2494, 2003. doi: 10.1056/NEJMoa030847.
- 3387 333. Trotter TN, Yang Y. Matricellular proteins as regulators of cancer metastasis to bone. Matrix
 Biol 52-54:301-314, 2016. doi: 10.1016/j.matbio.2016.01.006.
- 3389 334. Tsourdi E, Langdahl B, Cohen-Solal M, Aubry-Rozier B, Eriksen EF, Guañabens N,
 Obermayer-Pietsch B, Ralston SH, Eastell R, Zillikens MC. Discontinuation of Denosumab
 therapy for osteoporosis: A systematic review and position statement by ECTS. *Bone* 105:11-
- 3392 17, 2017. doi: 10.1016/j.bone.2017.08.003.
- 3393 335. Tsukasaki M, Hamada K, Okamoto K, Nagashima K, Terashima A, Komatsu N, Win SJ,
 Okamura T, Nitta T, Yasuda H, Penninger JM, Takayanagi H. LOX Fails to Substitute for
- 3395 RANKL in Osteoclastogenesis. *J Bone Miner Res* 32:434-439, 2017. doi: 10.1002/jbmr.2990.
- 3396 336. Tulotta C, Lefley DV, Freeman K, Gregory WM, Hanby AM, Heath PR, Nutter-Howard F,
- 3397 Wilkinson JM, Spicer-Hadlington AR, Liu X, Bradbury S, Hambley L, Cookson V, Allocca G,
- 3398 Kruithof-de-Julio M, Coleman RE, Brown JE, Holen I, Ottewell PD. Endogenous production of
- 3399 IL-1B by breast cancer cells drives metastasis and colonisation of the bone microenvironment.

3400 *Clin Cancer* Res. 25:2769-2782, 2019. doi: 10.1158/1078-0432.CCR-18-2202.

- 3401 337. Turner JJO. Hypercalcaemia presentation and management. *Clin Med (Lond).* 17:270-273,
 2017. doi: 10.7861/clinmedicine.
- 3403 338. Turpin A, Duterque-Coquillaud M, Vieillard MH. Bone Metastasis: Current State of Play. *Transl* 3404 *Oncol.* 13: 308-320, 2019. doi: 10.1016/j.tranon.2019.10.012.

3405 339. Ubellacker JM, Baryawno N, Severe N, DeCristo MJ, Sceneay J, Hutchinson JN, Haider MT,
3406 Rhee CS, Qin Y, Gregory WM, Garrido-Castro AC, Holen I, Brown JE, Coleman RE, Scadden
3407 DT, McAllister SS. Modulating bone marrow hematopoietic lineage potential to prevent bone
3408 metastasis in breast cancer. *Cancer Res.* 78:5300-5314, 2018. doi: 10.1158/0008-5472.CAN3409 18-0548.

- 3410 340. Ubellacker JM, Haider MT, DeCristo MJ, Allocca G, Brown NJ, Silver DP, Holen I, McAllister
 3411 SS. Zoledronic acid alters hematopoiesis and generates breast tumor-suppressive bone
 3412 marrow cells. *Breast Cancer Res* 19:23, 2017. doi: 10.1186/s13058-017-0815-8.
- 3413 341. Valencia K, Luis-Ravelo D, Bovy N, Antón I, Martínez-Canarias S, Zandueta C, Ormazábal C,
- Struman I, Tabruyn S, Rebmann V, De Las Rivas J, Guruceaga E, Bandrés E, Lecanda F.
 miRNA cargo within exosome-like vesicle transfer influences metastatic bone colonization. *Mol Oncol.* 8:689-703, 2014. doi: 10.1016/j.molonc.2014.01.012.
- 3417 342. Valta MP, Tuomela J, Bjartell A, Valve E, Väänänen HK, Härkönen P. FGF-8 is involved in 3418 bone metastasis of prostate cancer. *Int J Cancer* 123: 22-31, 2008. doi: 10.1002/ijc.23422.
- 3419 343. Valta MP, Zhao H, Ingels A, Thong AE, Nolley R, Saar M, Peehl DM. Development of a
 realistic in vivo bone metastasis model of human renal cell carcinoma. Clin Exp Metastasis.
 3421 31:573-584, 2014.
- 3422 344. van der Horst G, van den Hoogen C, Buijs JT, Cheung H, Bloys H, Pelger RC, Lorenzon G,
 3423 Heckmann B, Feyen J, Pujuguet P, Blanque R, Clément-Lacroix P, van der Pluijm G.
 3424 Targeting of α(v)-integrins in stem/progenitor cells and supportive microenvironment impairs
 3425 bone metastasis in human prostate cancer. *Neoplasia* 13: 516-525, 2011.
- 3426345. Van Poznak C, Somerfield MR, Barlow WE, Biermann JS, Bosserman LD, Clemons MJ,3427Dhesy-Thind SK, Dillmon MS, Eisen A, Frank ES, Jagsi R, Jimenez R, Theriault RL,
- 3428 Vandenberg TA, Yee GC, Moy B. Role of Bone-Modifying Agents in Metastatic Breast Cancer:

- An American Society of Clinical Oncology-Cancer Care Ontario Focused Guideline Update. J
 Clin Oncol. 35:3978-3986, 2017. doi: 10.1200/JCO.2017.75.4614.
- 3431 346. Vargas G, Bouchet M, Bouazza L, Reboul P, Boyault C, Gervais M, Kan C, Benetollo C,
 3432 Brevet M, Croset M, Mazel M, Cayrefourcq L, Geraci S, Vacher S, Pantano F, Filipits M,
 3433 Driouch K, Bieche I, Gnant M, Jacot W, Aubin JE, Duterque-Coquillaud M, Alix-Panabières C,
 3434 Clézardin P, Bonnelye E. ERRα promotes breast cancer cell dissemination to bone by
 increasing RANK expression in primary breast tumors. *Oncogene*. 38:950-964, 2019. doi:
 10.1038/s41388-018-0579-3.
- 3437 347. Vidula N, Yau C, Li J, Esserman LJ, Rugo HS. Receptor activator of nuclear factor kappa B
 3438 (RANK) expression in primary breast cancer correlates with recurrence-free survival and
 3439 development of bone metastases in I-SPY1 (CALGB 150007/150012; ACRIN 6657). *Breast*3440 *Cancer Res Treat* 165: 129-138, 2017. doi: 10.1007/s10549-017-4318-1.
- 3441 348. Vija Racaru L, Sinigaglia M, Kanoun S, Ben Bouallègue F, Tal I, Brillouet S, Bauriaud-Mallet
 M, Zerdoud S, Dierickx L, Vallot D, Caselles O, Gabiache E, Pascal P, Courbon F. Fluorine18-fluorocholine PET/CT parameters predictive for hematological toxicity to radium-223
 therapy in castrate-resistant prostate cancer patients with bone metastases: a pilot study. *Nucl Med Commun.* 39:672-679, 2018. doi: 10.1097/MNM.00000000000850.
- 3446 349. Vives V, Cres G, Richard C, Busson M, Ferrandez Y, Planson AG, Zeghouf M, Cherfils J,
 3447 Malaval L, Blangy A. Pharmacological inhibition of Dock5 prevents osteolysis by affecting
 3448 osteoclast podosome organization while preserving bone formation. *Nat Commun.* 6:6218,
- 3449 2015. doi: 10.1038/ncomms7218.
- 3450 350. Voorzanger-Rousselot N, Garnero P. Biochemical markers in oncology. Part I: molecular
 basis. Part II: clinical uses. *Cancer Treat Rev.* 33:230-283, 2007. doi:
 10.1016/j.ctrv.2007.01.008.

- 3453 351. Wakabayashi H, Wakisaka S, Hiraga T, Hata K, Nishimura R, Tominaga M, Yoneda T.
 3454 Decreased sensory nerve excitation and bone pain associated with mouse Lewis lung cancer
 3455 in TRPV1-deficient mice. *J Bone Miner Metab*.36:274-285, 2018. doi: 10.1007/s00774-0173456 0842-7.
- 3457 352. Wallitt KL, Khan SR, Dubash S, Tam HH, Khan S, Barwick TD. Clinical PET Imaging in 3458 Prostate Cancer. *Radiographics*. 37:1512-1536, 2017. doi: 10.1148/rg.2017170035.
- 3459 353. Waltregny D, Bellahcène A, Van Riet I, Fisher LW, Young M, Fernandez P, Dewé W, de Leval
 J, Castronovo V. Prognostic value of bone sialoprotein expression in clinically localized human
 prostate cancer. *J Natl Cancer Inst.* 90:1000-1008, 1998. doi: 10.1093/jnci/90.13.1000
- 3462 354. Wang H, Tian L, Liu J, Goldstein A, Bado I, Zhang W, Arenkiel BR, Li Z, Yang M, Du S, Zhao
 3463 H, Rowley DR, Wong STC, Gugala Z, Zhang XH. The Osteogenic Niche Is a Calcium
 3464 Reservoir of Bone Micrometastases and Confers Unexpected Therapeutic Vulnerability.
 3465 *Cancer Cell.* 34:823-839, 2018.e7. doi: 10.1016/j.ccell.2018.10.002.
- 3466 355. Wang H, Yu C, Gao X, Welte T, Muscarella AM, Tian L, Zhao H, Zhao Z, Du S, Tao J, Lee B,
 Westbrook TF, Wong ST, Jin X, Rosen JM, Osborne CK, Zhang XH. The osteogenic niche
 promotes early-stage bone colonization of disseminated breast cancer cells. *Cancer Cell*27:193-210, 2015. doi: 10.1016/j.ccell.2014.11.017.
- 3470 356. Wang J, Chen GL, Cao S, Zhao MC, Liu YQ, Chen XX, Qian C. Adipogenic niches for
 3471 melanoma cell colonization and growth in bone marrow. *Lab Invest.* 97:737-745, 2017. doi:
 3472 10.1038/labinvest.2017.14.
- 3473 357. Wang N, Docherty FE, Brown HK, Reeves KJ, Fowles AC, Ottewell PD, Dear TN, Holen I,
- 3474 Croucher PI, Eaton CL. Prostate cancer cells preferentially home to osteoblast-rich areas in
- 3475 the early stages of bone metastasis: evidence from in vivo models. *J Bone Miner Res* 29:2688-
- 3476 2696, 2014. doi: 10.1002/jbmr.2300.

3477 358. Wang S, Li GX, Tan CC, He R, Kang LJ, Lu JT, Li XQ, Wang QS, Liu PF, Zhai QL, Feng YM.
FOXF2 reprograms breast cancer cells into bone metastasis seeds. *Nat Commun.* 10:2707,
2019. doi: 10.1038/s41467-019-10379-7.

3480 359. Waning DL, Mohammad KS, Reiken S, Xie W, Andersson DC, John S, Chiechi A, Wright LE,
3481 Umanskaya A, Niewolna M, Trivedi T, Charkhzarrin S, Khatiwada P, Wronska A, Haynes A,

3482

Excess TGF-β mediates muscle weakness associated with bone metastases in mice. *Nat Med.*21:1262-1271, 2015. doi: 10.1038/nm.3961.

Benassi MS, Witzmann FA, Zhen G, Wang X, Cao X, Roodman GD, Marks AR, Guise TA.

- 3485 360. Watanabe K, Hirata M, Tominari T, Matsumoto C, Fujita H, Yonekura K, Murphy G, Nagase H,
- 3486 Miyaura C, Inada M. The MET/Vascular Endothelial Growth Factor Receptor (VEGFR)-
- targeted Tyrosine Kinase Inhibitor Also Attenuates FMS-dependent Osteoclast Differentiation
 and Bone Destruction Induced by Prostate Cancer. *J Biol Chem.* 291:20891-20899, 2016.
- 3489 361. Weilbaecher KN, Guise TA, McCauley LK. Cancer to bone: a fatal attraction. *Nat Rev Cancer*3490 11: 411-425, 2011. doi: 10.1038/nrc3055.
- 3491 362. Welm AL, Sneddon JB, Taylor C, Nuyten DS, van de Vijver MJ, Hasegawa BH, Bishop JM.
 The macrophage-stimulating protein pathway promotes metastasis in a mouse model for
 breast cancer and predicts poor prognosis in humans. *Proc Natl Acad Sci U S A*. 104:75707575, 2007.
- 3495 363. Wen J, Huang G, Liu S, Wan J, Wang X, Zhu Y, Kaliney W, Zhang C, Cheng L, Wen X, Lu X.
 Polymorphonuclear MDSCs are enriched in the stroma and expanded in metastases of
 prostate cancer. *J Pathol Clin Res.* 6:171-177, 2020. doi: 10.1002/cjp2.160.
- 3498 364. Werner-Klein M, Grujovic A, Irlbeck C, Obradović M, Hoffmann M, Koerkel-Qu H, Lu X, 3499 Treitschke S, Köstler C, Botteron C, Weidele K, Werno C, Polzer B, Kirsch S, Gužvić M,
- 3500 Warfsmann J, Honarnejad K, Czyz Z, Feliciello G, Blochberger I, Grunewald S, Schneider E,
- Haunschild G, Patwary N, Guetter S, Huber S, Rack B, Harbeck N, Buchholz S, Rümmele P,
- Heine N, Rose-John S, Klein CA. Interleukin-6 trans-signaling is a candidate mechanism to
 drive progression of human DCCs during clinical latency. *Nat Commun* 11:4977, 2020. doi:
 10.1038/s41467-020-18701-4.
- 3505 365. Westbrook JA, Cairns DA, Peng J, Speirs V, Hanby AM, Holen I, Wood SL, Ottewell PD,
 3506 Marshall H, Banks RE, Selby PJ, Coleman RE, Brown JE. CAPG and GIPC1: Breast Cancer
 3507 Biomarkers for Bone Metastasis Development and Treatment. *J Natl Cancer Inst.* 108(4),
 2016. doi: 10.1093/jnci/djv360.
- 3509 366. Westbrook JA, Wood SL, Cairns DA, McMahon K, Gahlaut R, Thygesen H, Shires M, Roberts
- 3510 S, Marshall H, Oliva MR, Dunning MJ, Hanby AM, Selby PJ, Speirs V, Mavria G, Coleman RE,
- Brown JE. Identification and validation of DOCK4 as a potential biomarker for risk of bone
- metastasis development in patients with early breast cancer. *J Pathol.* 247:381-391, 2019. doi:
 10.1002/path.5197.
- 3514 367. Wong R, Wiffen PJ. Bisphosphonates for the relief of pain secondary to bone metastases.
 3515 Cochrane *Database Syst Rev.* CD002068, 2002.
- 3516 368. Wortzel I, Dror S, Kenific CM, Lyden D. Exosome-Mediated Metastasis: Communication from a
 3517 Distance. *Dev Cell.* 49:347-360, 2019. doi: 10.1016/j.devcel.2019.04.011.
- 3518 369. Wu AC, He Y, Broomfield A, Paatan NJ, Harrington BS, Tseng HW, Beaven EA, Kiernan DM,
- 3519 Swindle P, Clubb AB, Levesque JP, Winkler IG, Ling MT, Srinivasan B, Hooper JD, Pettit AR.
- 3520 CD169(+) macrophages mediate pathological formation of woven bone in skeletal lesions of
 prostate cancer. *J Pathol.* 239:218-230, 2016. doi: 10.1002/path.4718.
- 3522 370. Xiang L, Gilkes DM. The Contribution of the Immune System in Bone Metastasis
 3523 Pathogenesis. *Int J Mol Sci.* 20(4). pii: E999, 2019. doi: 10.3390/ijms20040999.
- 3524 371. Xu Z, Liu X, Wang H, Li J, Dai L, Li J, Dong C. Lung adenocarcinoma cell-derived exosomal
 3525 miR-21 facilitates osteoclastogenesis. *Gene* 666: 116-122, 2018. doi:
 3526 10.1016/j.gene.2018.05.008.
 - 144

- 3527 372. Yaccoby S, Ling W, Zhan F, Walker R, Barlogie B, Shaughnessy JD Jr. Antibody-based
 inhibition of DKK1 suppresses tumor-induced bone resorption and multiple myeloma growth in
 vivo. *Blood.* 109:2106-2111, 2007. doi: 10.1182/blood-2006-09-047712.
- 3530 373. Yang C, Siebert JR, Burns R, Gerbec ZJ, Bonacci B, Rymaszewski A, Rau M, Riese MJ, Rao
 3531 S, Carlson KS, Routes JM, Verbsky JW, Thakar MS, Malarkannan S. Heterogeneity of human
 bone marrow and blood natural killer cells defined by single-cell transcriptome. *Nat Commun.*10:3931, 2019. doi: 10.1038/s41467-019-11947-7.
- 3534 374. Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, Teng SC, Wu KJ. Direct regulation
 of TWIST by HIF-1alpha promotes metastasis. *Nat Cell Biol.* 10:295-305, 2008. doi:
 10.1038/ncb1691.
- 3537 375. Yazici O, Aksoy S, Sendur MA, Babacan T, Ozdemir N, Ozisik Y, Zengin N, Altundag K. The
 effect of obesity on recurrence pattern in early breast cancer patients. J BUON. 20:954-962,
 2015.
- 3540 376. Ye Y, Li SL, Ma YY, Diao YJ, Yang L, Su MQ, Li Z, Ji Y, Wang J, Lei L, Fan WX, Li LX, Xu Y, 3541 Hao XK. Exosomal miR-141-3p regulates osteoblast activity to promote the osteoblastic 3542 prostate cancer. Oncotarget 8: 94834-94849, 2017. metastasis of doi: 3543 10.18632/oncotarget.22014.
- 3544 377. Yi B, Williams PJ, Niewolna M, Wang Y, Yoneda T. Tumor-derived platelet-derived growth
 actor-BB plays a critical role in osteosclerotic bone metastasis in an animal model of human
 breast cancer. *Cancer Res* 62: 917-923, 2002.
- 3547 378. Yin JJ, Mohammad KS, Käkönen SM, Harris S, Wu-Wong JR, Wessale JL, Padley RJ, Garrett
 3548 IR, Chirgwin JM, Guise TA. A causal role for endothelin-1 in the pathogenesis of osteoblastic
 3549 bone metastases. *Proc Natl Acad Sci USA* 100: 10954-10959, 2003. doi:
 3550 10.1073/pnas.1830978100.

- 3551 379. Yonou H, Aoyagi Y, Kanomata N, Kamijo T, Oda T, Yokose T, Hasebe T, Nagai K, Hatano T, 3552 Ogawa Y, Ochiai A. Prostate-specific antigen induces osteoplastic changes by an autonomous 3553 mechanism. Biochem Biophys Res Commun 289: 1082-1087, 2001. doi: 3554 10.1006/bbrc.2001.6129.
- 3555 380. Yonou H, Horiguchi Y, Ohno Y, Namiki K, Yoshioka K, Ohori M, Hatano T, Tachibana M.
 3556 Prostate-specific antigen stimulates osteoprotegerin production and inhibits receptor activator
 3557 of nuclear factor-kappaB ligand expression by human osteoblasts. *Prostate* 67:840-848, 2007.
 3558 doi: 10.1002/pros.20574.
- 3559 381. Yu-Lee LY, Yu G, Lee YC, Lin SC, Pan J, Pan T, Yu KJ, Liu B, Creighton CJ, Rodriguez3560 Canales J, Villalobos PA, Wistuba II, de Nadal E, Posas F, Gallick GE, Lin SH. Osteoblast3561 Secreted Factors Mediate Dormancy of Metastatic Prostate Cancer in the Bone via Activation
 3562 of the TGFβRIII-p38MAPK-pS249/T252RB Pathway. *Cancer Res.* 78: 2911-2924, 2018. doi:
 3563 10.1158/0008-5472.CAN-17-1051.
- 3564 382. Yumoto K, Eber MR, Wang J, Cackowski FC, Decker AM, Lee E, Nobre AR, Aguirre-Ghiso JA,
 3565 Jung Y, Taichman RS. Axl is required for TGF-β2-induced dormancy of prostate cancer cells in
 the bone marrow. *Sci Rep.* 6:36520, 2016. doi: 10.1038/srep36520.
- 3567 383. Zang L, Ma M, Hu J, Qiu H, Huang B, Chu T. The effects of lung and prostate cancer bone
 metastasis on serum osteoprotegerin levels: a meta-analysis. *Sci Rep* 5:18324, 2015. doi:
 10.1038/srep18324.
- 3570 384. Zhang J, Mao F, Niu G, Peng L, Lang L, Li F, Ying H, Wu H, Pan B, Zhu Z, Chen X. (68)Ga3571 BBN-RGD PET/CT for GRPR and Integrin α(v)β(3) Imaging in Patients with Breast Cancer.
 3572 *Theranostics*. 8:1121-1130, 2018. doi: 10.7150/thno.22601.
- 3573 385. Zhang J, Pang Y, Xie T, Zhu L. CXCR4 antagonism in combination with IDO1 inhibition 3574 weakens immune suppression and inhibits tumor growth in mouse breast cancer bone 3575 metastases. *Onco Targets Ther.* 12:4985-4992, 2019. doi:10.2147/OTT.S200643

- 3576 386. Zhang K, Kim S, Cremasco V, Hirbe AC, Collins L, Piwnica-Worms D, Novack DV,
 3577 Weilbaecher K, Faccio R. CD8+ T cells regulate bone tumor burden independent of osteoclast
 3578 resorption. *Cancer Res.* 71:4799-4808, 2011. doi: 10.1158/0008-5472.CAN-10-3922.
- 3579 387. Zhang XH, Wang Q, Gerald W, Hudis CA, Norton L, Smid M, Foekens JA, Massagué J. Latent
 bone metastasis in breast cancer tied to Src-dependent survival signals. *Cancer Cell*. 16: 6778, 2009. doi: 10.1016/j.ccr.2009.05.017.
- 3582 388. Zhao Y, Bachelier R, Treilleux I, Pujuguet P, Peyruchaud O, Baron R, Clément-Lacroix P,
 S583 Clézardin P. Tumor alphavbeta3 integrin is a therapeutic target for breast cancer bone
 metastases. *Cancer Res* 67: 5821-5830, 2007. DOI: 10.1158/0008-5472.CAN-06-4499.
- 3585 389. Zhao E, Wang L, Dai J, Kryczek I, Wei S, Vatan L, Altuwaijri S, Sparwasser T, Wang G, Keller
 ET, Zou W. Regulatory T cells in the bone marrow microenvironment in patients with prostate
 cancer. *Oncoimmunology*. 1:152-161, 2012. doi: 10.4161/onci.1.2.18480.
- 3588 390. Zheng H, Bae Y, Kasimir-Bauer S, Tang R, Chen J, Ren G, Yuan M, Esposito M, Li W, Wei Y,
- Shen M, Zhang L, Tupitsyn N, Pantel K, King C, Sun J, Moriguchi J, Jun HT, Coxon A, Lee B,
 Kang Y. Therapeutic Antibody Targeting Tumor- and Osteoblastic Niche-Derived Jagged1
 Sensitizes Bone Metastasis to Chemotherapy. *Cancer Cell.* 32:731-747, 2017. doi:
- 3592 10.1016/j.ccell.2017.11.002.
- 3593 391. Zheng D, Decker KF, Zhou T, Chen J, Qi Z, Jacobs K, Weilbaecher KN, Corey E, Long F, Jia
 L. Role of WNT7B-induced noncanonical pathway in advanced prostate cancer. *Mol Cancer Res* 11: 482-493, 2013. doi: 10.1158/1541-7786.MCR-12-0520.
- 3596 392. Zhou JZ, Riquelme MA, Gao X, Ellies LG, Sun LZ, Jiang JX. Differential impact of adenosine
 nucleotides released by osteocytes on breast cancer growth and bone metastasis. *Oncogene*3598 34: 1831-42, 2015. doi: 10.1038/onc.2014.113.

3599	393.	Zhou JZ, Riquelme MA, Gu S, Kar R, Gao X, Sun L, Jiang JX. Osteocytic connexin
3600		hemichannels suppress breast cancer growth and bone metastasis. Oncogene. 35:5597-5607,
3601		2016. doi: 10.1038/onc.2016.101.

- 3602 394. Zhu L, Narloch JL, Onkar S, Joy M, Broadwater G, Luedke C, Hall A, Kim R, Pogue-Geile K,
 3603 Sammons S, Nayyar N, Chukwueke U, Brastianos PK, Anders CK, Soloff AC, Vignali DAA,
 3604 Tseng GC, Emens LA, Lucas PC, Blackwell KL, Oesterreich S, Lee AV. Metastatic breast
 3605 cancers have reduced immune cell recruitment but harbor increased macrophages relative to
 3606 their matched primary tumors. J Immunother Cancer. 7:265, 2019. doi: 10.1186/s40425-0193607 0755-1.
- 3608 395. Zhu M, Liu C, Li S, Zhang S, Yao Q, Song Q. Sclerostin induced tumor growth, bone
 metastasis and osteolysis in breast cancer. *Sci Rep.* 7:11399, 2017. doi: 10.1038/s41598-01711913-7.
- 3611 396. Zhuang X, Zhang H, Li X, Li X, Cong M, Peng F, Yu J, Zhang X, Yang Q, Hu G. Differential
 a612 effects on lung and bone metastasis of breast cancer by Wnt signalling inhibitor DKK1. *Nat*a613 *Cell Biol* 19:1274-1285, 2017. doi: 10.1038/ncb3613.

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Figure legends:

Figure 1: Patterns of bone metastases from solid tumors ranging from mostly destructive (osteolytic) to mostly bone-forming (osteoblastic). Representative radiographs and histology of bone metastases with osteolytic (white arrow) or osteoblastic (black arrow) lesions are shown. For bone tumor sections, mineralized bone is stained green, whereas bone marrow and tumor cells (*) are stained red. Of note, in osteoblastic lesions, extensive new woven bone (stained dark red) can be observed, leading to the formation of new trabecular bone that fills the bone marrow cavity (white arrow).

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3624 Figure 2: Bone colonization by tumor cells is a stepwise sequence of events that include (i) the 3625 formation of a pre-metastatic niche to attract circulating tumor cells (CTCs) in bone, (ii) the 3626 extravasation and homing of CTCs within the pre-metastatic niche where they bind to bone extracellular 3627 matrix proteins, and *(iii)* the maintenance of tumor cells in the vascular niche and the osteoblastic niche 3628 where tumor cells become quiescent through specific adhesive interactions with host cells. ANXA2: 3629 annexin A2; AXL: tyrosine kinase receptor; CAR cells: CXCL-12-abundant reticular cells; CDH: 3630 cadherin; Cx43: connexin 43; CXCL: chemokine; CXCR: chemokine receptor; ECM: extracellular matrix; 3631 GAS-6: growth arrest-specific 6; IL-1: interleukin-1; IL-6: interleukin-6; LOX: lysyl oxidase; RANK-L: 3632 receptor activator of nuclear factor kappa-B (RANK) ligand; SNO: spindle-shaped N-cadherin+ 3633 osteoblast; TSP-1: thrombospondin-1.

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Figure 3: The fate of bone-resident tumor cells is determined by a balance between activities of multiple activated protein kinases (MAPK) ERK1/2 and p38, where a switch towards ERK1/2 phosphorylation favors proliferation whereas activation of p38 leads to quiescence. This balance between ERK1/2 and p38 activities is governed by several factors that either promote dormancy (boxes in green), helping tumor cells to survive in the vascular and osteoblastic niches, or enhance tumor cell reactivation and proliferation (boxes in red). However, proliferative tumor cells become vulnerable to immune

3641 surveillance, leading to tumor cell killing by CD8+ T cells and NK cells. The bone microenvironment also 3642 contains immunosuppressive cells (MDSCs, Treg, pDC) that help tumor cells to escape from adaptive 3643 immunity. AXL: tyrosine kinase receptor; BMP-7: bone morphogenetic protein-7; C CAR cells: CXCL-3644 12-abundant reticular cells; CXCL-12: chemokine;: GAS-6: growth arrest-specific 6; IFN-y: interferon 3645 γ ; LIF: leukemia inhibitory factor; MDSCs: myeloid-derived suppressor cells; MSK1: mitogen- and 3646 stress-activated kinase 1; NK cell: natural killer cell; POSTN: periostin; pDC: plasmacytoid dendritic cell; 3647 PTHrP: parathyroid hormone-related peptide; RANK-L: receptor activator of nuclear factor kappa-B 3648 (RANK) ligand; SNO: spindle-shaped N-cadherin+ osteoblast; TGFB: transforming growth factor beta; 3649 Treg: regulatory T cells; TSP-1: thrombospondin-1; TYRO3: tyrosine kinase receptor; VCAM-1: vascular 3650 cell adhesion protein 1; VEGF: vascular endothelial growth factor.

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3652 Figure 4: Mechanisms governing the formation of osteolytic bone metastases. Several factors secreted 3653 by tumor cells enhance osteoclast-mediated bone resorption, either directly (e.g., IL-8) or indirectly 3654 (e.g., PTHrP, IL-6) via stimulation of RANK-L secretion and inhibition of OPG production by osteoblasts. 3655 In turn, the binding of RANK-L to RANK on osteoclast precursors leads to the formation of new 3656 osteoclasts. LPA by binding to its receptor LPA1 at the tumor cell surface promotes tumor cell 3657 proliferation and the production of IL-6 and IL-8, further enhancing osteoclast-mediated bone resorption. 3658 In addition, tumor-derived LOX and IL-1beta accelerate RANKL-induced osteoclastogenesis. 3659 Consequently, growth factors (TGF β , IGFs, PDGF) and calcium are released from the resorbed bone 3660 matrix. TGFB acts on tumor cells and stimulates the expression of factors such as PTHrP and Notch 3661 ligand Jagged-1. In turn, Jagged/Notch signaling promotes osteoclast differentiation. IGFs and calcium 3662 promote tumor cell proliferation. Calcium also stimulates the secretion of PTHrP and epiregulin by tumor 3663 cells. Tumor-derived epiregulin decreases OPG expression in osteoblasts. Thus, there is a vicious cycle 3664 where tumor cells stimulate bone destruction and factors released from resorbed bone stimulate tumor 3665 growth. This cycle is enhanced by the secretion of tumor-derived factors (DKK-1, SOST-1, noggin,

activin A) that inhibit osteoblast activity, thereby worsening the imbalance between bone formation and
 bone resorption, and promoting bone destruction.

3668 DKK-1: dickkopf-1; IGF: insulin-like growth factor; IL-6: interleukin-6; LOX: lysyl oxidase; LPA: 3669 lysophosphatidic acid; OPG: osteoprotegerin; PDGF: platelet-derived growth factor; PTHrP: parathyroid

3670 hormone-related peptide; RANK-L: receptor activator of nuclear factor kappa-B (RANK) ligand; SOST-1:

- 3671 sclerostin; TGF β : transforming growth factor beta.
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3673 Figure 5: Mechanisms governing the formation of osteoblastic bone metastases. Several factors 3674 secreted by tumor cells directly enhance osteoblast differentiation (ET-1, BMP-2, BMP-6, Whts), BMP-4 3675 mediates conversion of endothelial cells into osteoblasts. The stimulation of osteoblast differentiation is 3676 associated with increased OPG production, whereas RANK-L secretion is decreased. Tumor cells also 3677 produce OPG. Tumor-derived ET-1 directly acts onto mature osteoclasts to inhibit osteoclast activity. 3678 Therefore, there is a strong imbalance between bone formation and bone resorption, leading to aberrant 3679 bone formation. In addition, tumor-derived PSA and uPA increase the bioavailability of tumor growth-3680 promoting factors to the bone microenvironment, such as IGF-I and TGF- β .

BMP: bone morphogenetic protein; ET-1: endothelin-1; IGF: insulin-like growth factor; OPG:
 osteoprotegerin; PSA: prostate specific antigen; TGFβ: transforming growth factor beta; uPA:
 urokinase.

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Figure 6: Contribution of immune cells to bone metastasis formation. The innate and adaptive immune cells in the bone tissue microenvironment harbor both tumor-promoting and tumor-suppressing activities. CD8⁺ T cells and natural killer (NK) cells eliminate tumor cells through the production of interferon (IFN)- γ or TRAIL/FASL-induced apoptosis. However, these tumor cells may escape to the cytotoxic activity of immune cells (*e.g.*, CD8⁺ T cells), by inducing the recruitment of myeloid derived suppressor cells (MDSC), plasmacytoid dendritic cells (pDC) and regulatory T cells (Treg) that induce an immunosuppressive state within the bone tissue microenvironment. Beside tumor-suppressing activities, MDSCs can differentiate into functional osteoclasts. Furthermore, tumor-associated macrophages and a population of specialist osteal tissue macrophages termed osteomacs facilitate bone metastasis formation. RANK-L: receptor activator of nuclear factor kappa-B (RANK) ligand.

3695

3696 Figure 7: Metabolic pathways associated with bone metastasis progression. In order to increase 3697 glucose uptake, cancer cells up-regulate glucose transporters, notably glucose transporter 1 (GLUT1). 3698 Glucose is then utilized for ATP generation through lactate production (aerobic glycolysis), via glucose-3699 6-phosphate (G6P) and the pentose phosphate pathway (PPP) for nucleotide synthesis and through the 3700 tricarboxylic acid (TCA) cycle for lipid biosynthesis and protein acetylation. Lactate is released from 3701 tumor cells by monocarboxylate transporter 4 (MCT4) and then uptaken by osteoclasts through the 3702 transporter MCT1. Lactate stimulates osteoclast-mediated bone resorption, whereas fatty acids, 3703 cholesterol and nucleotides stimulate tumor cell proliferation.

3704

3705 Figure 8: (A) Clinical presentation of a solitary asymptomatic bone metastasis in a 54-year-old patient 3706 with breast cancer. A hot spot localized on L1 left pedicle (black arrow) was initially detected using 3707 technetium-99m (99m Tc)-bisphosphonate (BP) planar whole-body scintigraphy. Further analysis was 3708 conducted using fluorodeoxyglucose (FDG) positron emission tomography (PET) imaging. FDG/PET 3709 imaging displayed a hypermetabolic focus (white arrow) congruent to 99mTc-BP scintigraphy. (B) 3710 Clinical presentation of a solitary asymptomatic bone metastasis in a 67-year-old patient with prostate 3711 cancer. A hot spot localized on L3 vertebral body (black arrow) was initially detected using 99mTc-BP 3712 planar whole-body scintigraphy. Further analysis was conducted using fluorocholine (FCH)/PET 3713 imaging, displaying a hypermetabolic focus (white arrow) on L3, which was congruent to 99mTc-BP 3714 scintigraphy.

- **Figure 9:** Current and emerging bone-targeted therapies. Summary of cellular and molecular targets
- 3717 and corresponding bone-targeted agents that are approved by the FDA and EMA for use in oncology or
- evaluated in phase trials. NCT: ClinicalTrials.gov identifier (ID) number.



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PROGRESSION OF BONE COLONIZATION





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PROGRESSION OF OSTEOLYTIC BONE METASTASES



PROGRESSION OF OSTEOBLASTIC BONE METASTASES









В

Matrix, Cell	Molecular target	Drug	Phase / Trial ID (NCT)
	Calcium	Bisphosphonates, Radium-223, Samarium-223, Strontium-89	Approved
Bone matrix	TGFβ	Galinusertib	II / NCT02452008
	FPPS	Bisphosphonates	Approved
	RANKL	Denosumab	Approved
	mTOR	Everolimus	Approved
Osteoclast	VEGFR, c-MET	Cabozantinib	Approved
	RON	BMS777607/ASLAN002	I / NCT01721148, NCT00605618
	Androgen receptor	Enzalutamide	Approved
	CYP17A	Abiraterone	Approved
Osteoblast	Activin A	Sotatercept	II / NCT01562405, NCT00747123
	DKK-1	BHQ880	II / NCT01302886, NCT01337752, NCT00741377
the second se			
AFT	NGF	Tanezumab	III / NCT00545129, NCT02609828
Nerve cell Downloaded from journals physiology org/journal/physray at INSEPM (103.054.110.061) on December 20. 2020			nher 29, 2020

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Chemokine receptor	Chemokine Ligand	Tumor type	Function in bone metastasis	Reference
CXCR-2	CXCL-1, CXCL-2	Prostate	Marrow adipocyte-derived CXCL-1 and CXCL-2 contribute to osteolysis in metastatic prostate cancer	135
CXCR-2	CXCL-5	Breast	The CXCR-2/CXCL-5 axis promotes tumor cell colonization in the bone marrow	275
CXCR-2	CXCL-8 (IL8)	Breast	Tumor-derived IL8 contributes to osteoclastic bone destruction in bone metastasis	18, 26, 27, 361
CXCR-3	CXCL-10	Breast, melanoma	CXCL-10 facilitates trafficking of CXCR-3-expressing cancer cells to bone	193
CXCR-4	CXCL-12	Breast, prostate	The CXCR-4/CXCL-12 axis regulates tumor cell entry in the bone marrow	235, 267, 273
CXCR-5	CXCL-13	Prostate	CXCL-13 mediates attachment of CXCR-5-expressing tumor cells to bone marrow endothelial cells <i>in</i> <i>vitro</i>	300
CX3CR-1	CX3CL-1	Breast, prostate	CX3CR-1 and CX3CL-1 interactions promote tumor cell extravasation in the bone marrow	158,159
CCR-2	CCL-2	Breast, prostate	CCL2-expressing tumor cells engage CCR-2-expressing macrophages and pre-osteoclasts to facilitate colonization in bone	213, 232

 Table 1. Chemokines and their receptors involved in the formation of bone metastasis.

Table 2. Potential clinical utility of bone turnover biomarkers.

Biomarker	Abbreviation	Clinical application	Reference
Bone formation marker			
Bone alkaline phosphatase	BALP	Diagnosis of bone metastasis in solid tumors. Prognosis of bone metastasis in solid tumors. Risk of skeletal related events. Prognosis during anti-resorptive therapy. Prediction of response to treatment.	61, 67, 90, 91, 155, 187, 206
procollagen I carboxyl-terminal propeptide	PICP	Diagnosis of bone metastasis in prostate cancer. Prediction of response to atrasentan in prostate cancer.	187
Procollagen I amino-terminal propeptide	PINP	Diagnosis of bone metastasis in breast and prostate cancer.	35, 90, 180
Bone resorption marker			
C-telopeptide	СТХ	Diagnosis of bone metastasis in prostate cancer. Prognosis of bone metastasis in breast cancer.	33, 35
N-telopeptide	NTX	Diagnosis of bone metastasis in prostate and lung cancer. Prognosis of bone metastasis in solid tumors. Risk of skeletal related events. Prognosis during anti-resorptive therapy. Prediction	35, 61, 67,90, 155, 187, 206

of response to treatment.

carboxyterminal telopeptide of type I collagen	ICTP	Diagnosis of bone metastasis in lung cancer. Prognosis of bone metastasis in breast cancer.	33, 155
Tartrate resistant acid phosphatase 5b	TRACP	Diagnosis of bone metastasis in breast cancer.	90, 155
pyridinoline	PYD	Prediction of response to atrasentan in prostate cancer.	186
Receptor activator of nuclear factor κB -ligand/osteoprotegerin	RANKL/OPG	Diagnosis of bone metastasis in solid tumors	125, 165, 183
microRNAs	miRNAs	Diagnosis of bone metastasis in breast cancer.	96, 294

