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Surface-induced crystallisation of Sodium Dodecyl Sulfate (SDS) micellar solutions in confinement

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Abstract

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We investigate the role of confinement on the onset of crystallisation in sub-cooled 3 micellar solutions of sodium dodecyl sulfate (SDS), examining the impact of sample 4 volume, substrate surface energy, and surface roughness. Using small angle neutron 5 scattering (SANS) and dynamic light scattering (DLS), we measure the crystallisation 6 temperature upon cooling, and the metastable zone width (MSZW), for bulk 10-30 7 wt% SDS solutions. We then introduce a microdroplet approach to quantify the im-8 pact of surface free energy (18-65 mN/m) and substrate roughness ($R_{\alpha}\simeq$ 0-60 $\mu{\rm m})$ on 9 the kinetics of surface-induced crystallisation through measurements of induction time 10 (t_i) under isothermal conditions. While t_i is found to decrease exponentially with de-11 creasing temperature (increasing sub-cooling) for all tested surfaces, increasing surface 12 energy could cause a significant, further reduction, of up to ${\sim}40$ fold. For substrates 13 with the lowest surface energy and longest t_i , microscale surface roughness is found to 14 enhance crystal nucleation, in particular for $R_{\alpha} \geq 10 \ \mu m$. Finally, we demonstrate that 15

tuning surface energy and microscopic roughness can be effective routes to promote or
delay nucleation in bulk-like volumes, thus greatly impacting the stability of surfactant
solutions at lower temperatures.

¹⁹ Introduction

Solution crystallisation underpins the manufacture of a wide range of materials, such as phar-20 maceutical drugs, food products and personal care formulations.^{1–5} While the focus of some 21 branches of pharmaceutical and food industries is on the promotion of crystal formation and 22 control of their polymorphic structure, the inhibition of crystallisation can also be desirable 23 in the production of soluble drugs and functional stable home care products. As the key 24 component in personal and home care formulations, surfactants are amphiphilic molecules 25 which can spontaneously self-assemble into ordered structures. At low to moderate concen-26 trations (<30-40 wt%), most surfactants form globular micelles, extensively used in cleaning, 27 foaming and encapsulation applications. Inevitable variations in temperature, and partic-28 ularly cooling, can reduce the solubility of surfactants leading to the formation of crystals, 29 hence compromising the long-term stability and overall performance of such micellar sus-30 pensions.^{6–8} Cooling crystallisation is of special concern particularly for systems whose min-31 imum solubility temperature is near room temperature (~ 20 °C) for typical concentrations 32 used in formulated products (5-30 wt%).⁹ Well-known surfactants in this category include 33 CetylTrimethylAmmonium Bromide (CTAB)¹⁰ and Sodium Dodecyl Sulfate (SDS)¹¹ which 34 are widely used in detergent formulations,¹² pharmaceutical products,^{13–17} and bio-^{18–22} and 35 nano-technology²³⁻²⁶ processes. 36

Crystallisation is a first-order phase transition initiated by nucleation events whose thermodynamics is often described in simple terms by the Classic Nucleation Theory (CNT).^{27,28} CNT provides a minimal model for describing various scenarios of homogeneous nucleation in a supersaturated bulk solution. In practice, however, even at high level of supersaturation, bulk crystallisation occurs via a heterogeneous process initiated by nucleation on impurities

(indicated by case 1 in Fig. 1).^{29–32} Additionally, solution crystallisation is often initiated in 42 the vicinity of an interface with another material which may exist in either gas, liquid or 43 solid phases or a combination of these, as illustrated schematically by cases 2-4 in Fig. $1.^{33,34}$ 44 Controlling heterogeneous crystallisation in the bulk is often challenging due to an incom-45 plete knowledge of the structure, size and interfacial properties of impurities. By contrast, 46 surface-induced crystallisation on solid confinement boundaries of prescribed characteris-47 tics offers robust routes to examine the role of the various physico-chemical variables.^{35–41} 48 Additionally, manifestations of confined and substrate-induced crystallisation are expected 49 to be prevalent, and amplified, in common analytical laboratory processes, which employ 50 smaller and geometrically confined sample volumes, such as those encountered in optical mi-51 croscopy analyses, microfluidic platforms and differential scanning calorimetry (DSC).^{42–44} 52 Therefore, understanding the role of commonly-used laboratory substrates on heterogeneous 53 phase change processes is crucial for the realistic interpretation of a range of experimental 54 characterisation measurements. 55

Bulk crystallisation of SDS from aqueous solutions has been the topic of investigations 56 for several decades.^{45–48} Inevitably, most scientific characterisation procedures as well as 57 practical applications that involve manipulation of SDS solutions undergo surface-induced 58 crystallisation which precedes the nucleation within the bulk. In the last two decades, 59 considerable research has been dedicated to analyses of surface-induced crystallisation of 60 a variety of chemical compounds such as salts, 39,49 synthetic and natural polymers $^{41,50-52}$ 61 and complex pharmaceutical formulations.^{37,53} By contrast, heterogeneous crystallisation of 62 surfactants has received comparably little attention in the literature and, to our knowledge, 63 no systematic measurement of the impact of solid surfaces on cooling crystallisation of SDS 64 or similar surfactant solutions has been reported. The main goal of this work is therefore 65 to examine the role of confining solid boundaries, in particular the contribution of surface 66 energy and microscopic surface roughness, on heterogeneous cooling crystallisation of SDS 67 solutions. 68



Figure 1: Schematic of different scenarios of bulk (1) and surface-induced (2, 3, and 4) nucleation considered in this study. R is the radius of the nucleus (in yellow) growing within the sub-cooled liquid phase (in blue). γ_n is the average interfacial energy between the nucleus and the bulk, and θ_n is the contact angle of the nucleus on the solid substrate. γ_{sl} , γ_{ns} and γ_{la} are solid-liquid, solid-nucleus and liquid-air (surface tension of the liquid) interfacial energies. α is the roughness wedge angle and R_a refers to the average surface roughness.

We first examine the bulk crystallisation in micellar solutions of SDS in the range of 10-69 30 wt% in cooling experiments using Small Angle Neutron Scattering (SANS) and Dynamic 70 Light Scattering (DLS), in conditions of negligible impact from the solid boundaries. We 71 measure the solubility boundary of SDS solutions of prescribed concentrations upon cooling 72 and identify the corresponding metastable region in the temperature-concentration phase 73 diagram. Secondly, we focus on surface-induced heterogeneous crystallisation and investigate 74 the effect of free surface energy and roughness of the substrates in contact with μL volumes 75 of micellar solutions of SDS in confined geometry, where interfacial effects are significant. 76 Finally, we validate our findings of surface-induced heterogeneous crystallisation gathered 77 from confined micro-scale geometries in large 'bulk' sample volumes. 78

⁷⁹ Experimental

⁸⁰ Sodium dodecyl sulfate SDS (>99.0% purity) was purchased from Sigma-Aldrich and used ⁸¹ as received. Solutions for DLS and OM measurements were prepared by diluting SDS in ⁸² deionised water and keeping the solutions at room temperature overnight. Surfactant solu-⁸³ tions were filtered (0.2 μ m PTFE syringe filter) before use. For SANS measurements, SDS ⁸⁴ at >99.0% purity was diluted in D₂O. Considering the density difference between H₂O and D_2O , a correction was made when preparing samples in D_2O to keep the molar fraction of surfactant constant.

⁸⁷ Dynamic Light Scattering (DLS)

Dynamic light scattering (DLS) was performed using a Zetasizer Nano S (Malvern Pana-88 lytical), which operates in back-scattering ($\theta = 173^{\circ}$) with a 633 nm He-Ne laser yielding 89 $q = 0.0026 \text{ Å}^{-1}$. The cuvette temperature was controlled with a Peltier system in the range 90 of 30 °C to 0 °C. All cooling cycles were started from 30 °C to ensure that samples were 91 initially in the isotropic micellar phase, samples were left to reach thermal equilibrium at 92 the final temperature T_f and time-resolved data were acquired over one hour to probe the 93 transition from micellar to crystalline phases. To achieve the largest $\Delta T/\Delta t$ in isothermal 94 DLS experiments, the test environment was stabilised at T_f before inserting the cuvette 95 containing the sample. Data were acquired in triplicate. The raw correlograms were inter-96 preted without any need for cumulant fitting or CONTIN analysis, in order to identify the 97 boundaries of the micellar phase. 98

⁹⁹ Cross-polarised Optical Microscopy (OM)

Cross-polarised optical microscopy was used to detect phase transition from the isotropic 100 micellar phase to the birefringent crystalline phases, following procedures described in a pre-101 vious publication.⁷ Approximately 0.5 μ l of the solution was placed between two thin glass 102 cover slips. Solvent evaporation during the experiments was minimised by sealing the area 103 surrounding the droplet using an adhesive gasket of 120 μ m thickness. Solvent evaporation 104 was further controlled during the image processing step by ensuring that the droplet area 105 remained constant during the experiments. Any measurement showing droplet shrinkage was 106 discarded from the data set. Isothermal experiments were performed by initially stabilising 107 the droplet at 30 °C before a rapid quench (at 80 °C/min) to the final temperature of inter-108 est. Thermal control of the microscopic samples was carried out using a Linkam THMS600 109

temperature control stage. OM images were captured with an Olympus BX41M-LED microscope, using 5X and 10X objectives and a CMOS camera (Basler ac2040-90).

¹¹² Small Angle Neutron Scattering (SANS)

SANS experiments in linear and isothermal cooling cycles were performed on the Larmor 113 diffractometer (ISIS, Harwell, UK), with a polychromatic $\lambda = 0.9-13.3$ Å incident beam and 114 sample-to-detector distance = 4.1 m, yielding a fixed momentum transfer range of approx-115 imately 0.005 < Q < 0.6 Å⁻¹ with the peak flux in the intermediate Q range. Quartz cells 116 (1 mm banjo pathlength, Starna) containing micellar solutions of SDS were installed into 117 a metallic sample changer that was thermally controlled using a liquid bath. Experiments 118 were started at 60 °C, where all solutions were in the isotropic micellar phase, and then a 119 variety of cooling ramps were applied to the samples to reach 0 °C.⁷ The resulting SANS 120 data were reduced, using standard procedures, in MANTID.⁵⁴ 121

¹²² Profilometry and surface roughness

Polydimethylsiloxane (PDMS) surfaces of prescribed roughness were prepared by pouring 123 10:1 (base:cross-linker) solution of Sylgard 184 (Dow Corning) over sandpaper of different 124 roughness, degassed under a vacuum desiccator and baked at 80 °C overnight. Patterned 125 PDMS slabs were then peeled off and used as rough substrates. High fidelity replication of 126 three-dimensional micro- and nano-structure of sandpaper by using PDMS was previously 127 reported in the literature.⁵⁵ A Bruker DektakXT stylus profiler was used to quantify the 128 surface roughness of various PDMS slabs. Stylus force was set at 2 mg and a map of 2 μ m 129 resolution over an area of $4 \times 4 \text{ mm}^2$ was obtained for each sample. The arithmetical 130 mean deviation of the measured profile R_a was used as an estimate of the surface roughness 131 (detailed in SI). 132

¹³³ Surface plasma treatment and contact angle

A series of glass (Fisherbrand borosilicate glass coverslips), polycarbonate (Goodfellow, 325-170-20) and PDMS (1:10, Sylgard 184) substrates were investigated. A Harrick PDC 13.6 MHz oxygen plasma cleaner was employed to tune surface free energy. A power of 18 W and exposure times of 2 min were used, and the resulting contact angles were measured with a digital camera setup.

139 Results and discussion

Due to its wide range of applications, the phase behaviour of aqueous solutions of SDS and 140 its hydrated crystals, at equilibrium and in static conditions, have been extensively inves-141 tigated over the past decades.^{11,46,56–59} Nevertheless, the kinetics of phase transformations 142 upon variations of temperature and concentration, and those occurring in the vicinity of 143 interfaces remain largely unresolved. Here, we focus on transformations from the micellar 144 L_1 to the crystalline C phase that occur upon cooling at fixed surfactant concentration. We 145 first describe signatures of the two phases obtained through conventional analytical meth-146 ods focusing on nucleation of SDS crystals within the bulk solution. Next, we investigate 147 the impact of solid boundaries, particularly surface energy and roughness on heterogeneous 148 nucleation of SDS within micellar solutions in confined settings. 149

150 Bulk crystallisation

Above the Critical Micelle Concentration (CMC) at about 8.2 mM⁶⁰ and below 30-40 wt% of surfactant, aqueous solutions of SDS exist in micellar L₁ phase at temperatures above 25 °C. In this temperature-concentration range, ellipsoidal micelles of SDS elongate and become less negatively charged with decreasing temperature.^{61,62} Upon continuous cooling below room temperature, hydrated crystalline SDS structures (C) are formed within the micellar solutions (L₁).⁶² We employed DLS to monitor the formation of crystals in the solution with

nanoscale resolution in mL volumes of 10-30 wt% SDS. In all experiments, very slow cooling 157 ramps ($\Delta T/\Delta t < 0.5$ °C/min) combined with temperature steps of $\Delta T = 1$ °C and 30 min 158 waiting time at each step is applied to ensure sample thermal equilibrium (Fig. 2a). Without 159 further analysing the correlograms obtained from DLS analyses, nucleation and growth of 160 the crystal within the micellar solutions of SDS can be simply detected by tracking the 161 appearance of a time-dependent slow-decay mode at lower temperatures (Fig. 2b). At 20 162 wt% SDS, no crystalline phase exists at room temperature, while a clear phase transformation 163 was detected at about 13 °C. In parallel, SANS measurements detected formation of larger 164 hydrated SDS crystals at temperatures below 14 °C for 20 wt% SDS, manifested by an 165 upturn in lower-Q region alongside a clear sharp Bragg peak at higher Q, characteristic 166 of the lamellar crystalline structure (Fig. 2c). The Bragg diffraction peaks at 0.192 Å⁻¹ 167 (lamellar spacing, d = 32.7 Å) and 0.378 Å⁻¹ respectively correspond to the first and second 168 lamellar spacing of the SDS-rich crystalline structure.^{44,46} The crystalline lamellar structures 169 are birefringent and clearly observed using cross-polarised optical microscopy OM (Fig. 2d). 170 At a fixed concentration of SDS, the temperature that the crystallisation is first detected 171 upon cooling $T_{C,C}$ is much lower than the equilibrium solubility temperature $T_{C,H}$ since a 172 sufficient level of sub-cooling is required to induce spontaneous nucleation in the solution. 173 The extent of sub-cooling temperature required to obtain crystallisation at a given concen-174 tration is identified as the MetaStable Zone Width (MSZW), see Fig. 2d.^{48,63,64} The MSZW 175 is a strong function of the rate of cooling applied, purity and volume of the sample, and 176 the detection resolution of the experimental technique used in continuous cooling experi-177 ments. In practice, most of these variables cannot be independently controlled since the 178 experimental measurement approach dictates not only the spatial detection resolution, but 179 also the sample volume which in turn impacts the cooling rate, *i.e.* the larger the sample 180 volume the slower the cooling rate that the bulk solution experiences. Moreover, besides the 181 geometry, the experimental approach often limits the choice of sample container material 182 which can significantly affect the observed crystallisation temperature by promoting surface-183

induced heterogeneous crystallisation, especially as the surface-to-volume ratio of the sample
increases. Therefore, any interpretation and comparison of the MSZW measurement reports
must carefully consider the experimental technique and protocols in use.



Figure 2: Detection of crystallisation in aqueous micellar solution of SDS upon continuous, stepped, slow cooling. (a) Schematic example of a cooling path starting from T_H in L_1 phase until reaching $T_{C,C}$ in L+C phase. (b) DLS measurements show the appearance of fast-growing larger objects with long decay times, in addition to the primary micellar aggregates, as the 20 wt% SDS solution is cooled below $T = 15^{\circ}$ C. (c) Upon cooling of the SDS solutions below $T < 14^{\circ}$ C, the micellar peak in SANS measurements at intermediate-Q gradually disappears as a Bragg peak at high-Q and upturn at low-Q, both associated with the crystallisation of SDS, are detected. The diffraction Bragg peaks at 0.192 Å⁻¹ and 0.378 Å⁻¹ respectively correspond to the first and second lamellar spacing of SDS-rich crystalline structure. (d) Upon cooling the micellar phase, crystallisation occurs at $T_{C,C}$ (solid line) that is lower than the temperature at which the crystals disappear $T_{C,H}$ (dotted line^{11,57}) in a reverse heating cycle. The L₁ and L₁+C phases are separated by a metastable zone. Cross-polarised optical microscopy images of solutions of 20 wt% SDS show growth of crystals as the solution is cooled below 13°C. The isotropic micellar phase (L₁) appears black, while the crystalline phase C is birefringent.

Owing to the relatively large sample volume and high detection resolution, SANS and DLS measurements allow precise tracking of static nucleation and crystallisation in the bulk based on the assumption of negligible contribution from sample interaction with its surrounding solid boundaries. However, rational comparison of kinetic measurements obtained from different analytical tools, where significant contribution of non-homogeneous nucleation is likely, largely relies on mechanistic understanding of surface-induced heterogeneous nucleation. In the following sections, we use DLS as a facile tool to characterise the bulk crystallisation of SDS solutions at a given concentration (20 wt%) and compare results with those obtained by optical microscopy analyses of microdroplets in order to assess the potential impact of interactions with typical bounding interfaces.



Figure 3: Induction time of crystallisation in isothermal DLS experiments for SDS solutions. (a) Schematic of a typical thermal path followed in isothermal DLS and OM tests. The induction time t_i is measured from the initial time that sample is exposed to T_f . (b) The nucleation and growth of crystals are identified through the appearance a slow-mode decay in the raw DLS correlograms, here after 28 min at 13 °C. (c) Microdroplet experiment: a droplet of 20 wt% SDS solution is confined between two identical transparent substrates that are sealed around the droplet to minimise the evaporation. The sample is placed on top of a thermally controlled stage and is subsequently quenched at 80 °C/min to reach the final temperature T_f . Micro-graphs correspond to OM images of a confined droplet at 4 °C (here using glass substrates), in which crystallisation occurs within the first minute of reaching T_f . Scale bar corresponds to 500 μ m.

¹⁹⁷ Isothermal measurements and induction time

In order to eliminate the effect of cooling rate as a variable in our measurements, here we perform isothermal tests in which the micellar solution is rapidly cooled to the final temperature T_f and held isothermally for 60 min during which continuous time-resolved detection of any possible crystallisation in the solution is performed (Fig. 3a). Under isothermal conditions, the time required for the critical nucleus formation and growth of a detectable crystalline phase in a supersaturated solution is referred to as the induction time t_i .⁶⁵ Induction time provides a convenient measure of the supersaturation level within the solution, while providing an indirect tool for estimating kinetics of nucleation and crystal growth.^{37,65,66}

Miller et al. (2016) measured the induction time for SDS solutions in isothermal tests, 206 however, their measurement showed no crystal formation for $T_f > 5$ °C at 20 wt%, and 207 yielded a wide metastable zone considering the corresponding equilibrium solubility tem-208 perature of 20 $^{\circ}$ C.⁴⁷ Note that the induction time here refers to the time required for the 209 formation of detectable crystalline phase from an equilibrium micellar phase. Using Dif-210 ferential Scanning Calorimetry (DSC), Summerton et al. (2016) observed crystallisation at 211 around 12±1°C for 20 wt% SDS solutions, which is closer to the equilibrium phase transition, 212 but no information on the kinetics of crystallisation and induction time was provided. 44 In 213 our isothermal DLS tests, no detectable change was found within 60 min of reaching $T_f > 14$ 214 °C in micellar solution of 20 wt% SDS. A clear change in the correlation data attributed to 215 the emergence of crystals was found only after 25 min at $T_f = 13$ °C (Fig. 3b). This finding 216 emphasises the sensitivity of MSZW measurements of similar systems to the experimental 217 protocol followed even under identical isothermal test conditions. Given that the induction 218 time is expected to be extremely large near the phase boundary, it is practically difficult 219 to measure accurately and often may be missed if samples are not equilibrated for long 220 enough. In general, in the range of 10-30 wt% of SDS, the cooling crystallisation tempera-221 tures detected here by DLS matched those reported previously based on DSC measurements 222 (Fig. SI1).⁴⁴ 223

²²⁴ Surface-induced crystallisation

While isothermal experiments effectively determine the impact of sub-cooling on the induction time of crystallisation, other conditions can dramatically influence the induction time of crystallisation. Such conditions are often achieved by promoting heterogeneous crystalli-

sation of the solution on additional surfaces and boundaries.^{37,67,68} Here, we characterise the 228 effect of surface energy and topography of solid interfaces on the crystallisation induction 229 time of micellar solutions of SDS through OM analyses of confined microdroplets, where the 230 contribution of the surface effects is maximised. Fig. 3c shows a schematic of the micro-231 droplet experimental setup used in this study. An isothermal quenching test similar to that 232 represented in Fig. 3a is applied to the bottom substrate (here glass) on which the micro-233 droplet of solution of 20 wt% SDS rests. It is practically simpler to impose a well-controlled 234 cooling ramp in the microdroplet experimental setting thanks to the small volume of the 235 sample, large surface area in contact with the cooling device and precise control of thermal 236 ramps imposed by the thermal stage. All isothermal cooling crystallisation tests reported 237 here are thus achieved by initial cooling of the micellar droplet at 80 °C/min. The micro-238 droplet setup allows microscopic detection of various hydration states of SDS in the crystals, 239 which are correlated to their morphology,⁴⁷ and their growth in time thanks to the relatively 240 high temporal resolution of the optical imaging apparatus (here on the order of 10^{-2} s) and 241 fast thermal equilibrium of the sample.⁴⁸ 242

As expected, the induction times t_i measured by DLS and OM significantly decrease 243 upon decreasing the final temperature T_f and reaching higher sub-cooling ΔT in solution 244 (Fig. 4). Considering that the volume of the sample and the detection length-scales have been 245 simultaneously reduced by ${\sim}3$ orders of magnitude in microdroplet experiments compared 246 to DLS, the induction time for detection of bulk crystallisation in the microdroplet setup 247 could be expected to be significantly longer. Note that effects of confinement due to the 248 higher level of supersaturation in the nano-litre environment is negligible here.⁶⁹ However, 249 comparison between microdroplet and DLS measurements at similar conditions presented 250 in Fig. 4 shows a significant reduction in t_i for OM analyses performed on smaller sample 251 volumes. This observation indicates that the crystallisation observed in microdroplets is 252 promoted by the considerably larger contact area of the sample with the bounding surfaces, 253 and thus larger surface-area to volume ratio, SA : V, as shown in Fig. 4 (see SI for more 254

details). Next, we explore the impact of free energy and microscopic roughness of the surface on the SDS crystal nucleation in microdroplet experiments through measurement of t_i .



Figure 4: Comparison between induction time measurements using DLS for 1 mL of solution and microscopy (on glass substrates) for a droplet of $\approx 0.5 \ \mu$ L volume. Solutions contain 20 wt% of SDS. The induction time decreases with reducing the final sub-cooled temperature T_f . Due to the large volume of the samples requiring relatively long thermal equilibrium time and low temporal resolution of DLS, shorter induction times expected at $T_f < 4 \ ^{\circ}$ C are not considered here. ΔT corresponds to $T_{C,C} - T_f$ and Surface-area to volume ratio SA : V is calculated considering all air-liquid interfaces and solid surfaces. Images on the right show examples of SDS crystallisation in aqueous micellar solution in a DLS cuvette (top) and microscopic droplet (bottom).

²⁵⁷ Surface free energy

We investigate the impact of surface energy for substrates that are relevant to laboratoryscale crystallisation measurements (e.g., in sample vials, or microfluidic devices), namely glass, PDMS and polycarbonate (PC). To modify the surface energy of the glass, used in measurements presented in Fig. 3, we use an oxygen plasma treatment. Plasma treatment is a common processing step, used in industrial and laboratory environments, to increase the surface free energy, and enhance molecular interactions and adhesion on the surface,^{75,76} as well as in the surface bonding and sealing of micro-devices.

Our microdroplet experiments on plasma-treated glass show significantly faster induction of crystal nucleation and larger number density of crystals compared to those on untreated glass, as shown in the image sequences in Fig. 5a. At a given temperature (here $T_f = 12$ °C)



Figure 5: Effect of surface energy on surface-induced isothermal crystallisation of 20 wt% micellar SDS solutions. (a) Cross-polarised OM time lapse images of microdroplets sand-wiched between glass cover-slips without (top) and with (bottom) oxygen plasma treatment (for 2 min) at $T_f = 12$ °C. On the native glass substrate, no crystals are formed within the first 20 min of reaching T_f . Plasma treatment of the glass substrate significantly promoted crystal nucleation and detectable crystals were observed within 10 min of reaching the final temperature. The scale bar corresponds to 300 μ m. (b) Induction times t_i measured on glass, oxygen plasma treated glass (P-glass), PDMS and polycarbonate substrates as a function of T_f . The dashed line represent the condition at which measurements in panel (c) are collected. (c) Induction time t_i measurements vs contact angle of water on different surfaces at $T_f = 4$ °C. Average surface free energy values reported in the literature are provided as a reference.⁷⁰⁻⁷⁴ Images show wetting conditions for DI water droplets on the different tested substrates.

no SDS crystals were detected on the surface of untreated glass in the first 20 min, while 268 clear birefringent crystalline structures were visible on plasma-treated glass within 10 min of 269 reaching T_f . Plasma oxidation significantly increases the free surface energy of glass (from 270 $\simeq 60 \pm 7 \text{ mN/m}$ to $70 \pm 7 \text{ mN/m}$), rendering it more hydrophilic. In order to further verify our 271 conclusion that heterogeneous crystallisation of SDS is enhanced by increasing total surface 272 free energy, we measure the crystallisation t_i for 20 wt% micellar solutions for a range of final 273 temperatures 4 °C < T_f < 12 °C on glass, plasma-treated glass (P-glass), and PDMS, see 274 Fig. 5b. For all three surfaces, t_i decreases exponentially with decreasing T_f (Fig. 5b) and 275 becomes closer to the lower boundary metastable zone (Fig. 2d). Considering the anionic 276 nature of SDS, it is expected that the polar contribution of the surface free energy will 277

define the rate of interaction with the surfactant molecules and influence induction time 278 of crystallisation, see Fig. SI3. Therefore, simple water contact angle measurements can 279 adequately describe the impact of surface energy on SDS crystallisation as confirmed in 280 Fig. 5c. For a given final temperature (here 4 °C), we found t_i to increase from ~35 s on 281 plasma-treated glass (contact angle \approx 8°) to ${\sim}80$ s on polycarbonate PC (contact angle \approx 282 75°) and finally to ~650 s on PDMS (contact angle $\approx 107^{\circ}$). The corresponding total surface 283 free energy values reported in the literature are included Fig. 5c as a reference, see Fig. SI3 284 for comparison between impacts of polar and non-polar surface energy contributions.^{75–81} 285

The effect of the substrate free energy on t_i can be rationalised in terms of a lower contact 286 angle of the heterogeneous SDS crystal nuclei forming on the solid substrate (θ_n in Fig. 1) 287 of higher surface energy which in turn yields a lower nucleation energy barrier.³³ We note 288 that crystallisation of aqueous micellar solutions of SDS is mainly controlled by the con-289 tribution of polar surface energy (Fig. SI3), which is clearly reflected in the contact angle 290 of water droplets on the surface, see Fig. 5c. The interfacial energy between the substrate 291 and crystalline phase depends on the absorption and adhesion of the solute molecules on 292 the bounding solid surfaces which is generally governed by the attractive forces arising from 293 chemical bonds, hydrogen bonds and van der Waals interactions,⁸² whose cumulative con-294 tribution is quantified by the surface free energy.^{83–85} Therefore, correlating t_i with surface 295 free energy appears to provide a simple, practical approach to estimate the substrate im-296 pact on heterogeneous nucleation in surfactant crystallisation. More precise identification 297 of molecular aspects of surface-induced crystallisation requires application of time-resolved 298 experimental structural measurements and non-classical two-step theoretical analysis in the 299 vicinity of the interface.^{86–88} 300

³⁰¹ Microscopic surface roughness

Another route for reducing the energy barrier to heterogeneous nucleation on substrates of low free energy is the introduction of roughness on the surface.^{52,89} Roughness can be modelled in terms of geometrical wedges of depth R_a constructed of smooth flat surfaces joined at an angle α , see case 4 in Fig. 1. Heterogeneous crystallisation becomes thus controlled not only by the interfacial energy and contact angle of the nucleus on the surface θ_n , but also by the wedge geometry, namely its depth and angle of the opening.^{90–93} For wedges filled with the liquid phase, which are significantly larger than radius of a critical nuclei R, the energy barrier in CNT disappears when $\theta_n \ll 180^\circ - \alpha$, *i. e.* deep narrow wedge geometries promote spontaneous nucleation.^{34,91}



Figure 6: Promoting surface-induced heterogeneous crystallisation of SDS on PDMS substrates by introducing microscopic surface roughness. (a) Rough PDMS surfaces are fabricated using commercially available sandpaper at different grit numbers. Profilometry measurements on a $4 \times 4 \text{ mm}^2$ are used to correlate the grit size with the quantitative surface roughness amplitude and shape. (b) Surface roughness R_a increases from about 5 μ m for grit size 3000 to around 60 μ m for grit size 60. The wedge angle α is estimated as the average of 10 different measurements performed across the rough PDMS substrates. Examples of typical wedges at different grit numbers are presented with scale bars representing 20 μ m. Optimal rough surfaces for promoting faster heterogeneous nucleation are expected to exhibit larger amplitude of roughness while maintaining wedges of smaller internal angle, α , as highlighted by the blue strip. (c) The induction time is significantly reduced on rough PDMS substrates, especially as roughness in increased to 10 μ m and above, corresponding to sandpaper grit number 1000 and smaller.

While most computational and experimental investigations of heterogeneous nucleation 311 on rough surfaces have been dedicated to understanding the effect of nano-scale surface 312 structures,^{35,52,53,89,92,94,95} our goal here is to identify the impact of larger microscopic sur-313 face roughness,^{96,97} where the wedge dimension is significantly larger than critical radius of 314 the nucleus. To this end, we perform microdroplet experiments on rough PDMS substrates 315 fabricated via a templating approach, using commercially available sandpaper of different 316 grades. Thanks to its high efficacy in replicating surface structures down to sub-micron 317 feature sizes and its low surface energy, PDMS offers a suitable substrate for quantifying the 318 effect of surface roughness on heterogeneous surface-induced crystallisation without modify-319 ing surface chemistry. 320



Figure 7: Time sequence of crystallisation in microdroplets on smooth glass, PDMS and rough PDMS (grit 2000) substrates, at $T_f=3.5\pm0.5$ °C. Crystal formation is initiated at the air-liquid interface around the droplets on smooth surfaces, in contrast to those appearing at the roughness sites on rough PDMS substrates. Scale bar refers to 500 μ m.

Roughness of sandpaper is commonly quantified by the 'grit size', which refers to the size of the abrasive particles on its surface, and can thus be correlated with the amplitude of surface roughness. Fig. 6a shows surface topography of the PDMS substrates obtained using different grits of sandpaper as templates. A wide range of surface undulations with amplitudes ranging from around 100 μ m down to about 5 μ m were obtained using sandpapers with grit numbers ranging from 60 to 3000, respectively. We characterised the topography of the roughness by measuring both amplitude (R_a) and opening angle of the surface un-

dulation (α) by profilometry, Fig. 6b. Further information on surface roughness analyses is 328 provided in SI. Computational molecular dynamic analyses at the nanoscale^{32,33,93} predict 329 that surfaces of larger roughness with deep narrow wedges offer ideal geometrical settings 330 to promote surface-induced crystallisation. By analogy to our microscale roughness, this 331 range corresponds to PDMS surfaces templated with grit sizes smaller than 1000 (shown in 332 Fig. 6b), where roughness is found to significantly reduce t_i (Fig. 6c). In general, t_i measure-333 ments on PDMS surfaces of various roughness show that adding microscopic roughness to 334 the PDMS surface dramatically reduces the crystallisation induction time (Fig. 6c). Relative 335 to smooth PDMS substrates (towards the right end of the x-axis in Fig. 6c), more than an 336 order of magnitude reduction in t_i is achieved by inducing wedges of depth larger 10 μ m. 337 A boomerang shaped curve with a minimum around grit size 400 describes the trend of t_i 338 vs. surface roughness, similar to the results presented previously in the literature for nucle-339 ation from vapour on microscopically rough glass surfaces.⁹⁶ This observation confirms that 340 effective promotion/inhibition of surface-induced crystallisation can be achieved by careful 341 design of the roughness size and geometry. 342

We note that in the microdroplet experiments on smooth substrates, the initial primary 343 crystallisation sites are typically located at the triple contact line between the liquid droplet, 344 surrounding air and top/bottom solid substrates (first and second columns in Fig. 7). This 345 observation is rationally supported by previous computer simulations by Sear³³ demonstrat-346 ing that the rate of the nucleation is significantly larger at the three-phase contact point 347 of air-water-solid impurities. The nucleation energy barrier at the air-liquid-solid interface 348 (case 3 in Fig. 1) is significantly reduced by the lower contribution of the interfacial tension 349 of nucleus due to the partial interface with the bulk liquid, which minimises the contribution 350 of γ_n . Additionally, nucleation at the triple contact point substitutes the existing interfacial 351 energies γ_{sl} and γ_{la} by the newly generated γ_{sn} and γ_{na} , respectively. Therefore, crystal nu-352 cleation is enhanced at the solid-liquid-air contact point on flat substrates, unless nucleation 353 interfacial energies with the solid γ_{sn} and the gas phase γ_{na} are considerably large. Inter-354

estingly, adding surface roughness not only accelerates the surface crystallisation on PDMS, but also clearly promotes nucleation and crystallisation within the droplet and not on the boundaries at the surrounding air interface, as was observed for flat smooth surfaces and highlighted by comparison of the third with the first two columns in Fig. 7.³³

359 Conclusion

We have investigated the heterogeneous crystallisation of aqueous micellar solutions of SDS 360 upon cooling. Using DLS and SANS we have characterised the 'bulk' metastable zone width 361 MSZW. Comparison with previous reports^{44,47} shows that the detection resolution of the 362 experimental techniques in use, volume of the sample, and cooling rates imposed may sig-363 nificantly impact measurements of the MSZW. We introduced confined microdroplet cooling 364 crystallisation experiments which amplify surface-induced crystallisation, and enable the ex-365 amination of the effects of surface free energy and roughness on heterogeneous crystallisation 366 through measurement on induction time, t_i . In general, heterogeneous nucleation in micro-367 droplets was enhanced by increasing the surface free energy of the substrate and introducing 368 microscopic roughness on the surface. Shortest crystallisation t_i was achieved on surfaces of 369 large roughness amplitude and small roughness wedge angle. 370

As the sample surface-to-volume ratio decreases, one trivially expects the surface-induced 371 heterogeneous nucleation to become less significant compared to the bulk nucleation, espe-372 cially for smooth surfaces of low free energy. As confirmation, our experiments performed 373 in clean untreated glass vials containing approximately 3 mL of 20 wt% SDS solutions show 374 minimal crystallisation within 30 min of reaching the highest level supersaturation tested 375 here at $T_f = 4$ °C, see images of the vials in top row of Fig. 8 for glass and smooth PDMS. 376 For comparison, confined microdroplet experiments on smooth glass and PDMS crystallised 377 within 1 and 16 min, respectively. Nevertheless, we predict that substrates with high free 378 energy and/or microscopic roughness can play significant roles in promoting heterogeneous 379



Figure 8: Crystallisation in glass vials containing ~ 3 mL (bulk-like volume) of 20 wt% SDS solution at $T_f = 4$ °C over 30 min. Samples were cooled from 30 °C, at which SDS solutions are in the micellar L₁ phase. Plasma treatment of the internal surface of the glass vial accelerates the formation of SDS crystals. Exposure to 2 min oxygen plasma significantly reduces the nucleation induction time t_i compared to those exposed to 1 min plasma. Untreated glass vials show minimal crystallisation within 30 min of the test. Placing slabs of PDMS of increasing roughness on the bottom of the containers enhances the heterogeneous nucleation and increases the density of crystallisation relative to vials with no/smooth PDMS. The base diameter of the vials is 20 mm.

nucleation in larger volumes, and are thus important in practical applications. Fig. 8 confirms 380 and quantifies the accelerated surface-induced crystallisation promoted by plasma treatment 381 of the solid boundaries of the container, or by the introduction of rough surfaces. Evidently, 382 understanding the impact of surfaces on heterogeneous crystallisation can be used to con-383 trol the nucleation density and induction time via tuning the surface energy and roughness, 384 even in 'bulk' volumes. The confined microdroplet approach employed here provides a facile 385 yet rigorous tool to examine surface effects on heterogeneous crystallisation which can then 386 be extended to larger volumes often encountered in material processing units, as well as in 387 laboratory analyses. 388

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³⁹⁵ Supporting Information Available

- ³⁹⁶ The following files are available free of charge.
- Filename: SI for Surface-induced crystallisation of SDS micellar solutions in confinement (PDF)

³⁹⁹ References

- (1) Chen, J.; Sarma, B.; Evans, J. M. B.; Myerson, A. S. Pharmaceutical Crystallization.
- 401 Crystal Growth & Design 2011, 11, 887–895.

- 402 (2) Shekunov, B. Y.; York, P. Crystallization processes in pharmaceutical technology and
 403 drug delivery design. *Journal of Crystal Growth* 2000, *211*, 122–136.
- 404 (3) Hartel, R. W. Advances in Food Crystallization. Annual Review of Food Science and
 405 Technology 2013, 4, 277–292.
- (4) Ribeiro, A. P. B.; Masuchi, M. H.; Miyasaki, E. K.; Domingues, M. A. F.; Stroppa, V.
- L. Z.; de Oliveira, G. M.; Kieckbusch, T. G. Crystallization modifiers in lipid systems.
 J Food Sci Technol 2015, 52, 3925–3946.
- (5) Hondoh, H.; Ueno, S.; Sato, K. Fundamental Aspects of Crystallization of Lipids; John
 Wiley Sons, Ltd, 2018; Chapter 4.
- (6) Laughlin, R. G. The aqueous phase behavior of surfactants; Academic Press, 1996.
- (7) Khodaparast, S.; Sharratt, W.; Wang, H.; Robles, E. S.; Dalgliesh, R.; Cabral, J. T.
 Spontaneous formation of multilamellar vesicles from aqueous micellar solutions of sodium linear alkylbenzene sulfonate (NaLAS). *Journal of Colloid and Interface Science*2019, 546, 221 230.
- (8) Summerton, E.; Hollamby, M. J.; Zimbitas, G.; Snow, T.; Smith, A. J.; Sommertune, J.; Bettiol, J.; Jones, C.; Britton, M. M.; Bakalis, S. The impact of N,N-dimethyldodecylamine N-oxide (DDAO) concentration on the crystallisation of sodium dodecyl sulfate (SDS) systems and the resulting changes to crystal structure, shape and the kinetics of crystal growth. J. Colloid Interf. Sci. 2018, 527, 260–266.
- (9) Tadros, T., Ed. Encyclopedia of Colloid and Interface Science; Springer-Verlag Berlin
 Heidelberg, 2013.
- 423 (10) Amos, K. E.; Brooks, N. J.; King, N. C.; Xie, S.; Canales-Vázquez, J.; Danks, M. J.;
 424 Jervis, H. B.; Zhou, W.; Seddon, J. M.; Bruce, D. W. A systematic study of the

- formation of mesostructured silica using surfactant ruthenium complexes in high- and low-concentration regimes. J. Mater. Chem. 2008, 18, 5282–5292.
- (11) Kékicheff, P. Phase diagram of sodium dodecyl sulfate-water system: 2. Complementary
 isoplethal and isothermal phase studies. Journal of Colloid and Interface Science 1989,
 131, 133–152.
- (12) Johansson I., S. P. Handbook for cleaning/decontamination of surfaces; Elsevier, 2007.
- (13) Balakrishnan, A.; Rege, B. D.; Amidon, G. L.; Polli, J. E. Surfactant-mediated dissolution: Contributions of solubility enhancement and relatively low micelle diffusivity. *Journal of Pharmaceutical Sciences* 2004, *93*, 2064–2075.
- (14) Ito, E.; Yip, K. W.; Katz, D.; Fonseca, S. B.; Hedley, D. W.; Chow, S.; Xu, G. W.;
 Wood, T. E.; Bastianutto, C.; Schimmer, A. D.; Kelley, S. O.; Liu, F.-F. Potential Use
 of Cetrimonium Bromide as an Apoptosis-Promoting Anticancer Agent for Head and
 Neck Cancer. *Molecular Pharmacology* 2009, *76*, 969–983.
- (15) Knöös, P.; Onder, S.; Pedersen, L.; Piculell, L.; Ulvenlund, S.; Wahlgren, M. Surfactants
 modify the release from tablets made of hydrophobically modified poly (acrylic acid). *Result Pharm. Sci.* 2013, *3*, 7–14.
- (16) Wang, G.; Wang, J.; Li, F.; To, S. T. Development and evaluation of a novel drug
 delivery: pluronics/SDS mixed micelle loaded with myricetin in vitro and in vivo. J. *Pharm. Sci.* 2016, 105, 1535–1543.
- (17) Sharma, G.; Naushad, M.; Thakur, B.; Kumar, A.; Negi, P.; Saini, R.; Chahal, A.; Kumar, A.; Stadler, F. J.; Aqil, U. M. H. Sodium Dodecyl Sulphate-supported nanocomposite as drug carrier system for controlled delivery of ondansetron. *Int. J. Environ. Res.* 2018, 15, 414.

- (18) Del Sal, G.; Manfioletti, G.; Schneider, C. The CTAB-DNA precipitation method: a
 common mini-scale preparation of template DNA from phagemids, phages or plasmids
 suitable for sequencing. *BioTechniques* 1989, 7, 514–520.
- (19) Nielsen, M. M.; Andersen, K. K.; Westh, P.; Otzen, D. E. Unfolding of beta-sheet
 proteins in SDS. *Biophys. J.* 2007, *92*, 3674–3685.
- (20) Azmat, M. A.; Khan, I. A.; Cheema, H. M. N.; Rajwana, I. A.; Khan, A. S.; Khan, A. A.
 Extraction of DNA suitable for PCR applications from mature leaves of Mangifera
 indica L. Journal of Zhejiang University. Science. B 2012, 13, 239–243.
- (21) Otzen, D. E. Proteins in a brave new surfactant world. Curr. Opin Colloid In. 2015,
 20, 161–169.
- 458 (22) Jafari, M.; Mehrnejad, F.; Rahimi, F.; Asghari, S. M. The Molecular Basis of the
 Sodium Dodecyl Sulfate Effect on Human Ubiquitin Structure: A Molecular Dynamics
 Simulation Study. Sci. Rep. 2018, 8, 2150.
- (23) Wu, S.-H.; Chen, D.-H. Synthesis of high-concentration Cu nanoparticles in aqueous
 CTAB solutions. Journal of Colloid and Interface Science 2004, 273, 165–169.
- (24) Sun, X. M.; Chen, X.; Deng, Z. X.; Li, Y. D. A CTAB-assisted hydrothermal orientation
 growth of ZnO nanorods. *Materials Chemistry and Physics* 2003, 78, 99–104.
- (25) Smith, D. K.; Korgel, B. A. The Importance of the CTAB Surfactant on the Colloidal
 Seed-Mediated Synthesis of Gold Nanorods. *Langmuir* 2008, 24, 644–649.
- 467 (26) Moon, S. Y.; Kusunose, T.; Sekino, T. CTAB-Assisted Synthesis of Size- and Shape 468 Controlled Gold Nanoparticles in SDS Aqueous Solution. *Materials Letters* 2009, 63,
 469 2038–2040.
- 470 (27) Metastable liquids: concepts and principles; Princeton University Press, Princeton, NJ,
 471 1996.

- 472 (28) Vekilov, P. G. Nucleation. Crys. Growth Des. 2010, 10, 5007–5019.
- (29) Perepezko, J. H. Nucleation in undercooled liquids. Materials Science and Engineer-*ing* 1984, 65, 125 135, Solidification Microstructure: 30 Years after Constitutional
 Supercooling.
- (30) Greenwood, G. W.; Greer, A. L.; Herlach, D. M.; Kelton, K. F.; Cantor, B. Heterogeneous nucleation and adsorption. *Philosophical Transactions of the Royal Society of London. Series A: Mathematical, Physical and Engineering Sciences* 2003, 361, 409–
 417.
- (31) Greenwood, G. W.; Greer, A. L.; Herlach, D. M.; Kelton, K. F.; Perepezko, J. H.;
 Tong, W. S. Nucleation-catalysis-kinetics analysis under dynamic conditions. *Philosophical Transactions of the Royal Society of London. Series A: Mathematical, Physical and Engineering Sciences* 2003, 361, 447–461.
- (32) Sear, R. P. Heterogeneous and homogeneous nucleation compared: rapid nucleation on
 microscopic impurities. *The Journal of Physical Chemistry B* 2006, *110*, 4985–4989.
- (33) Sear, R. P. Nucleation at contact lines where fluid-fluid interfaces meet solid surfaces.
 Journal of Physics: Condensed Matter 2007, 19, 466106.
- (34) Sear, R. P. Nucleation: theory and applications to protein solutions and colloidal suspensions. *Journal of Physics: Condensed Matter* 2007, 19, 033101.
- (35) Page, A. J.; Sear, R. P. Heterogeneous nucleation in and out of pores. *Phys. Rev. Lett.* **2006**, *97*, 065701.
- (36) Frenkel, D. Seeds of phase change. *Nature* **2006**, *443*, 641–641.
- (37) Diao, Y.; Myerson, A. S.; Hatton, T. A.; Trout, B. L. Surface Design for Controlled
 Crystallization: The Role of Surface Chemistry and Nanoscale Pores in Heterogeneous
 Nucleation. Langmuir 2011, 27, 5324–5334.

- (38) Carter, P. W.; Ward, M. D. Directing polymorph selectivity during nucleation of anthranilic acid on molecular substrates. *Journal of the American Chemical Society* 1994,
 116, 769–770.
- (39) D'Souza, S. M.; Alexander, C.; Carr, S. W.; Waller, A. M.; Whitcombe, M. J.; Vulfson, E. N. Directed nucleation of calcite at a crystal-imprinted polymer surface. *Nature* **1999**, *398*, 312–316.
- ⁵⁰² (40) Lee, A. Y.; Lee, I. S.; Dette, S. S.; Boerner, J.; Myerson, A. S. Crystallization on Con⁵⁰³ fined Engineered Surfaces: A Method to Control Crystal Size and Generate Different
 ⁵⁰⁴ Polymorphs. Journal of the American Chemical Society 2005, 127, 14982–14983.
- (41) Lee, T.; Hung, S. T.; Kuo, C. S. Polymorph farming of acetaminophen and sulfathiazole
 on a chip. *Pharm Res* 2006, *23*, 2542–2555.
- ⁵⁰⁷ (42) Wang, H.; Khodaparast, S.; Carroll, J.; Kelly, C.; Robles, E. S. J.; Cabral, J. T. A
 ⁵⁰⁸ microfluidic-multiwell platform for rapid phase mapping of surfactant solutions. *Review* ⁵⁰⁹ of Scientific Instruments 2020, 91, 045109.
- (43) Adamo, M.; Poulos, A. S.; G. Lopez, C.; Martel, A.; Porcar, L.; Cabral, J. T. Droplet
 microfluidic SANS. Soft Matter 2018, 14, 1759–1770.
- (44) Summerton, E.; Zimbitas, G.; Britton, M.; Bakalis, S. Crystallisation of sodium dodecyl
 sulfate and the corresponding effect of 1-dodecanol addition. *Journal of Crystal Growth*2016, 455, 111–116.
- (45) Smith, L. A.; Hammond, R. B.; Roberts, K. J.; Machin, D.; McLeod, G. Determination
 of the crystal structure of anhydrous sodium dodecyl sulphate using a combination of
 synchrotron radiation powder diffraction and molecular modelling techniques. J. Molec.
 Struct. 2000, 554, 173 –182.

522		<i>Growth</i> 2004 , <i>263</i> , 480–490.
521		morphology, surface chemistry and kinetic interface roughening. Journal of Crystal
520		sation of sodium dodecyl sulphate from aqueous solution: phase identification, crystal
519	(46)	Smith, L.; Duncan, A.; Thomson, G.; Roberts, K.; Machin, D.; McLeod, G. Crystalli-

- ⁵²³ (47) Miller, R. M.; Poulos, A. S.; Robles, E. S. J.; Brooks, N. J.; Ces, O.; Cabral, J. T.
 ⁵²⁴ Isothermal Crystallization Kinetics of Sodium Dodecyl Sulfate–Water Micellar Solutions. *Crystal Growth & Design* 2016, *16*, 3379–3388.
- (48) Miller, R. M.; Ces, O.; Brooks, N. J.; Robles, E. S. J.; Cabral, J. T. Crystallization of
 Sodium Dodecyl Sulfate-Water Micellar Solutions under Linear Cooling. *Cryst. Growth Des.* 2017, 17, 2428–2437.
- (49) Lioliou, M. G.; Paraskeva, C. A.; Koutsoukos, P. G.; Payatakes, A. C. Heterogeneous
 nucleation and growth of calcium carbonate on calcite and quartz. *Journal of Colloid and Interface Science* 2007, 308, 421 –428.
- (50) Chatterjee, A. M.; Price, F. P.; Newman, S. Heterogeneous nucleation of crystallization
 of high polymers from the melt. I. Substrate-induced morphologies. *Journal of Polymer Science: Polymer Physics Edition* 1975, 13, 2369–2383.
- ⁵³⁵ (51) Schonhorn, H. Heterogeneous Nucleation of Polymer Melts on High-Energy Surfaces. II.
 ⁵³⁶ Effect of Substrate on Morphology and Wettability. *Macromolecules* 1968, 1, 145–151.
- ⁵³⁷ (52) Chayen, N. E.; Saridakis, E.; Sear, R. P. Experiment and theory for heterogeneous
 ⁵³⁸ nucleation of protein crystals in a porous medium. *Proceedings of the National Academy* ⁵³⁹ of Sciences 2006, 103, 597–601.
- (53) Diao, Y.; Harada, T.; Myerson, A. S.; Hatton, T. A.; Trout, B. L. The role of nanopore
 shape in surface-induced crystallization. *Nat Mater* 2011, 10, 867–871.

- ⁵⁴² (54) Arnold, O. et al. Mantid—Data analysis and visualization package for neutron scatter-⁵⁴³ ing and μ SR experiments. *Nucl. Instrum. Meth. A* **2014**, *764*, 156–166.
- ⁵⁴⁴ (55) Rasel, M. S. U.; Park, J.-Y. A sandpaper assisted micro-structured polydimethylsilox⁵⁴⁵ ane fabrication for human skin based triboelectric energy harvesting application. Ap⁵⁴⁶ plied Energy 2017, 206, 150 158.
- ⁵⁴⁷ (56) Leigh, I. D.; McDonald, M. P.; Wood, R. M.; Tiddy, G. J. T.; Trevethan, M. A.
 ⁵⁴⁸ Structure of liquid-crystalline phases formed by sodium dodecyl sulphate and water as
 ⁵⁴⁹ determined by optical microscopy, X-ray diffraction and nuclear magnetic resonance
 ⁵⁵⁰ spectroscopy. J. Chem. Soc., Faraday Trans. 1 1981, 77, 2867–2876.
- ⁵⁵¹ (57) Kekicheff, P.; Grabielle, M. C.; Ollivon, M. Phase diagram of sodium dodecyl sulfate⁵⁵² water system: 1. A calorimetric study. J. Colloid Interf. Sci. 1989, 131, 112 132.
- (58) Itri, R.; Amaral, L. Q.; Mariani, P. Structure of the hexagonal phase of the sodium
 dodecyl sulfate and water system. *Phys. Rev. E* 1996, 54, 5211–5216.
- (59) Lee, T.; Yeh, K. L.; You, J. X.; Fan, Y. C.; Cheng, Y. S.; Pratama, D. E. Reproducible
 Crystallization of Sodium Dodecyl Sulfate 1/8 Hydrate by Evaporation, Antisolvent
 Addition, and Cooling. ACS Omega 2020, 5, 1068–1079.
- (60) Moroi, Y.; Motomura, K.; Matuura, R. The critical micelle concentration of sodium
 dodecyl sulfate-bivalent metal dodecyl sulfate mixtures in aqueous solutions. *Journal*of Colloid and Interface Science 1974, 46, 111–117.
- (61) Hammouda, B. Temperature effect on the nanostructure of SDS micelles in water. J. *Res. Natl. Inst. Stan.* 2013, 118, 151–167.
- (62) Khodaparast, S.; Sharratt, W. N.; Tyagi, G.; Dalgliesh, R. M.; Robles, E. S.;
 Cabral, J. T. Pure and mixed aqueous micellar solutions of Sodium Dodecyl sulfate

- (SDS) and Dimethyldodecyl Amine Oxide (DDAO): Role of temperature and composition. Journal of Colloid and Interface Science 2021, 582, 1116 1127.
- Kadam, S. S.; Kulkarni, S. A.; Ribera, R. C.; Stankiewicz, A. I.; ter Horst, J. H.;
 Kramer, H. J. M. A new view on the metastable zone width during cooling crystallization. *Chem. Eng. Sci.* 2012, 72, 10–19.
- ⁵⁷⁰ (64) van Gelder, R.; Roberts, K.; Chambers, J.; Instone, T. Nucleation of single and mixed
 ⁵⁷¹ straight chain surfactants from dilute aqueous solutions. *Journal of Crystal Growth*⁵⁷² 1996, 166, 189 194.
- ⁵⁷³ (65) Söhnel, O.; Mullin, J. W. Interpretation of crystallization induction periods. *Journal of*⁵⁷⁴ Colloid and Interface Science 1988, 123, 43–50.
- ⁵⁷⁵ (66) Cardew, P. T.; Davey, R. J.; Garside, J. Evaluation of supersaturation in crystal growth
 ⁵⁷⁶ from solution. *Journal of Crystal Growth* 1979, *46*, 534–538.
- ⁵⁷⁷ (67) Artusio, F.; Pisano, R. Surface-induced crystallization of pharmaceuticals and biophar⁵⁷⁸ maceuticals: A review. Int J Pharm 2018, 547, 190–208.
- ⁵⁷⁹ (68) Archer, A. J.; Malijevský, A. Crystallization of soft matter under confinement at inter⁵⁸⁰ faces and in wedges. *Journal of Physics: Condensed Matter* 2016, 28, 244017.
- (69) Grossier, R.; Magnaldo, A.; Veesler, S. Ultra-fast crystallization due to confinement.
 Journal of Crystal Growth 2010, 312, 487 489.
- ⁵⁸³ (70) Kirby, B. J.; Hasselbrink Jr., E. F. Zeta potential of microfluidic substrates: 2. Data
 ⁵⁸⁴ for polymers. *ELECTROPHORESIS* 2004, 25, 203–213.
- ⁵⁸⁵ (71) Naseh, N.; Mohseni, M.; Ramezanzadeh, B. Role of surface active additives on reduction
 of surface free energy and enhancing the mechanical Attributes of easy-to-clean automotive clearcoats: Investigating resistance against simulated tree gum. Int. J. Adhes.
 Adhes. 2013, 44, 209–219.

- ⁵⁸⁹ (72) Zhang, R.; Somasundaran, P. Advances in adsorption of surfactants and their mixtures
 ⁵⁹⁰ at solid/solution interfaces. Adv. Colloid Interface Sci. 2006, 123-126, 213-229.
- (73) McKechnie, D.; Anker, S.; Zahid, S.; Mulheran, P. A.; Sefcik, J.; Johnston, K. Interfacial
 Concentration Effect Facilitates Heterogeneous Nucleation from Solution. *The Journal of Physical Chemistry Letters* 2020, *11*, 2263–2271.
- (74) Vitha, M. F.; Carr, P. W. Study of the Polarity and Hydrogen-Bond Ability of Dodecyltrimethylammonium Bromide Micelles by the KamletTaft Solvatochromic Comparison Method. J. Phys. Chem. B 1998, 102, 1888–1895.
- ⁵⁹⁷ (75) Abenojar, J.; Martínez, M.; Encinas, N.; Velasco, F. Modification of glass surfaces
 ⁵⁹⁸ adhesion properties by atmospheric pressure plasma torch. *International Journal of*⁵⁹⁹ Adhesion and Adhesives 2013, 44, 1–8.
- (76) Terpilowski, K.; Rymuszka, D. Surface properties of glass plates activated by air, oxygen, nitrogen and argon plasma. *Glass Physics and Chemistry* 2016, 42, 535–541.
- ⁶⁰² (77) Rymuszka, D.; Terpiłowski, K.; Hołysz, L. Influence of Volume Drop on Surface Free
 ⁶⁰³ Energy of Glass. Annales UMCS, Chemia 2014, 68, 121–132.
- (78) Kim, Y. G.; Lim, N.; Kim, J.; Kim, C.; Lee, J.; Kwon, K.-H. Study on the surface
 energy characteristics of polydimethylsiloxane (PDMS) films modified by C4F8/O2/Ar
 plasma treatment. Applied Surface Science 2019, 477, 198–203.
- (79) Vicente, C.; André, P.; Ferreira, R. Simple measurement of surface free energy using a
 web cam. *Revista Brasileira de Ensino de Física* 2012, 34, 1–5.
- (80) Chibowski, E.; Terpilowski, K. Surface free energy of polypropylene and polycarbonate
 solidifying at different solid surfaces. *Applied Surface Science* 2009, 256, 1573–1581.
- (81) Surface tension values of some common test liquids for surface energy analysis. http:
 //www.surface-tension.de/solid-surface-energy.htm.

- (82) Owens, D. K.; Wendt, R. C. Estimation of the surface free energy of polymers. Journal
 of Applied Polymer Science 1969, 13, 1741–1747.
- (83) Sirringhaus, H.; Brown, P. J.; Friend, R. H.; Nielsen, M. M.; Bechgaard, K.; LangeveldVoss, B. M. W.; Spiering, A. J. H.; Janssen, R. A. J.; Meijer, E. W.; Herwig, P.;
 de Leeuw, D. M. Two-dimensional charge transport in self-organized, high-mobility
 conjugated polymers. *Nature* 1999, 401, 685–688.
- (84) Afzali, A.; Dimitrakopoulos, C. D.; Breen, T. L. High-Performance, Solution-Processed
 Organic Thin Film Transistors from a Novel Pentacene Precursor. *Journal of the Amer- ican Chemical Society* 2002, *124*, 8812–8813.
- (85) Kim, N.; Kee, S.; Lee, S. H.; Lee, B. H.; Kahng, Y. H.; Jo, Y.-R.; Kim, B.-J.; Lee, K.
 Highly Conductive PEDOT:PSS Nanofibrils Induced by Solution-Processed Crystallization. Advanced Materials 2014, 26, 2268–2272.
- (86) Schniepp, H. C.; Shum, H. C.; Saville, D. A.; Aksay, I. A. Orientational Order of
 Molecular Assemblies on Rough Surfaces. *The Journal of Physical Chemistry C* 2008, *112*, 14902–14906.
- (87) Ma, X.; Zhang, S.; Jiao, F.; Newcomb, C. J.; Zhang, Y.; Prakash, A.; Liao, Z.;
 Baer, M. D.; Mundy, C. J.; Pfaendtner, J.; Noy, A.; Chen, C.-L.; De Yoreo, J. J.
 Tuning crystallization pathways through sequence engineering of biomimetic polymers. *Nature Materials* 2017, 16, 767–774.
- (88) Striolo, A.; Grady, B. P. Surfactant Assemblies on Selected Nanostructured Surfaces:
 Evidence, Driving Forces, and Applications. *Langmuir* 2017, 33, 8099–8113.
- (89) Yan, D.; Zeng, Q.; Xu, S.; Zhang, Q.; Wang, J. Heterogeneous Nucleation on Concave
 Rough Surfaces: Thermodynamic Analysis and Implications for Nucleation Design. *The Journal of Physical Chemistry C* 2016, *120*, 10368–10380.

31

- (90) Campbell, J. M. On topography and crysatl nucleation. Ph.D. thesis, University of
 Leeds, 2014.
- (91) Sholl, C.; Fletcher, N. Decoration criteria for surface steps. Acta Metallurgica 1970,
 18, 1083–1086.
- (92) Page, A. J.; Sear, R. P. Crystallization Controlled by the Geometry of a Surface. Journal
 of the American Chemical Society 2009, 131, 17550–17551.
- (93) Bi, Y.; Cao, B.; Li, T. Enhanced heterogeneous ice nucleation by special surface geometry. *Nature Communications* 2017, *8*, 15372.
- (94) Walker, C.; Lerch, S.; Reininger, M.; Eghlidi, H.; Milionis, A.; Schutzius, T. M.;
 Poulikakos, D. Desublimation Frosting on Nanoengineered Surfaces. ACS Nano 2018,
 12, 8288–8296.
- (95) Zeng, Q.; Xu, S. Thermodynamics and Characteristics of Heterogeneous Nucleation on
 Fractal Surfaces. *The Journal of Physical Chemistry C* 2015, 119, 27426–27433.
- (96) Holbrough, J. L.; Campbell, J. M.; Meldrum, F. C.; Christenson, H. K. Topographical
 Control of Crystal Nucleation. *Crystal Growth & Design* 2012, *12*, 750–755.
- (97) Zhang, Y.; Wang, M.; Lin, X.; Huang, W. Effect of Substrate Surface Microstructure on
 Heterogeneous Nucleation Behavior. *Journal of Materials Science Technology* 2012,
 28, 67–72.
- (98) ISIS Neutron and Muon Source experiments RB1820374. https://doi.org/10.5286/
 ISIS.E.RB1820374.

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