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1 **Changes in groundwater bacterial community during cyclic**  
2 **groundwater-table variations**

3

4 **Running title: Water-table variations affect bacteria**

5

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32 **Abstract**

33 Column experiments containing an aquifer sand were subjected to static and oscillating water  
34 tables to investigate the impact of natural fluctuations and rainfall infiltration on the groundwater  
35 bacterial community just below the phreatic surface, and its association with the geochemistry.  
36 Once the columns were established, the continuously saturated zone was anoxic in all three  
37 columns. The rate of soil organic matter (SOM) mineralization was higher when the water table  
38 varied cyclically than when it was static due to the greater availability of  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$ . Natural  
39 fluctuations in the water table resulted in a similar  $\text{NO}_3^-$  concentration to that observed with a  
40 static water table but the cyclic wetting of the intermittently saturated zone resulted in a higher  
41  $\text{SO}_4^{2-}$  concentration. Rainfall infiltration induced cyclic water-table variations resulted in a  
42 higher  $\text{NO}_3^-$  concentration than those in the other two columns, and a  $\text{SO}_4^{2-}$  concentration  
43 intermediate between those columns. As rainwater infiltration resulted in slow downward  
44 displacement of the groundwater, it is inferred that  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  were being mobilized from  
45 the vadose zone.  $\text{NO}_3^-$  was mainly released by SOM mineralization (which was enhanced by the  
46 infiltration of oxygenated rainwater), but the larger amount of  $\text{SO}_4^{2-}$  release required a second  
47 mechanism (possibly desorption). Different groundwater bacterial communities evolved from  
48 initially similar populations due to the different groundwater histories.

49

50 **Keywords:** Water-table variations; rainfall infiltration; natural fluctuations; SOM mineralization;  
51  $\text{NO}_3^-$ ;  $\text{SO}_4^{2-}$ ; bacterial communities; groundwater histories

52

53 **1. Introduction**

54 Groundwater represents 95% of global freshwater and is thus an essential resource for  
55 drinking water, agriculture and industry (Igor, 1993). The microbial community in an aquifer can  
56 have a profound impact on groundwater quality, as microorganisms break down organic matter,  
57 consume oxygen, change the oxidation state of inorganic compounds, recycle nutrients and break  
58 down pollutants (Kim & Gadd, 2008). Thus, it is important to understand how microbial  
59 community varies as a function of location, and how its metabolic activity varies as a function of  
60 time (Griebler, Malard, & Lefébure, 2014).

61 The groundwater table in an aquifer can fluctuate in short-term, seasonally and from year to  
62 year in response to variations in rainfall infiltration, groundwater flow, groundwater extraction  
63 and recharge, surface water levels and other natural causes (Rühle, von Netzer, Lueders, &  
64 Stumpp, 2015; Haack et al., 2004; Krause, Bronstert, & Zehe, 2007; Dobson, Schroth, & Zeyer,  
65 2007). This produces a zone of intermittent saturation immediately above the continuously  
66 saturated zone where there are cyclical variations in the redox state and geochemistry (Yang et al.,  
67 2017; Stegen et al., 2016). Saturation, redox state and geochemistry are the principal factors that  
68 shape the microbial community present at a location (Shade, Jones, & McMahon, 2008;  
69 Medihala, Lawrence, Swerhone, & Korber, 2012; Zheng et al., 2019). Thus, temporal  
70 heterogeneity associated with groundwater-table fluctuations will impose a selective pressure  
71 that will favor microorganisms possessing metabolic plasticity and redox tolerance mechanisms  
72 (Rosenberg & Freedman, 1994).

73           There have been numerous studies of the effect of groundwater-table variations on bacterial  
74 processes in aquifers, but most have focused on the fate of natural and anthropomorphic  
75 contaminants in the intermittently saturated zone of an aquifer (e.g. Banks, Clennan, Dodds, &  
76 Rice, 1999; Van Driezum et al., 2018). Unsurprisingly, these show that electron donor and  
77 acceptor availability determine both microbial community composition and biogeochemical  
78 processes that the community mediates (e.g. Braun, Schröder, Knecht, & Szewzyk, 2016).  
79 However, it is interesting to note that indigenous microorganisms exhibit greater activity in a  
80 region of groundwater table variation than they would in equivalent zones above a static  
81 groundwater table (Banks et al., 1999), possibly because alternative movements of groundwater  
82 table transport nutrients and air to zone where air- and water-filled pores co-exist creating  
83 microhabitats with optimized conditions for microbial activity (Rainwater, Mayfield, Heintz, &  
84 Claborn, 1993; Banks et al., 1999). Moreover, different causes of a rising groundwater table may  
85 result in different responses of the indigenous microbial populations in an aquifer, due to  
86 difference in water chemistry and water-flow pathway (Zhou, Kellermann, & Griebler, 2012).  
87 Local rainfall infiltration involves downward percolation of water from the atmosphere through  
88 vadose zone, potentially eluting solutes and natural organics, and the upward displacement of  
89 pore air from capillary fringe against the water flow direction (Rainwater et al., 1993). Whereas  
90 regional recharge of an aquifer can result in upward permeation of groundwater from saturated  
91 zone, which will displace the pore air from capillary fringe upwards ahead of the wetting front.  
92 Such differences must lead to different redox conditions, affect the activity of indigenous

93 microorganisms, and impact on the evolution of the groundwater microbial community  
94 (Pett-Ridge & Firestone, 2005).

95 Microbial communities often require time to respond to environmental change (Rezanezhad,  
96 Couture, Kovac, O’Connell, & Van Cappellen, 2014), particularly as metabolic responses to new  
97 environment can themselves cause geochemical changes creating new ecological niches over  
98 time (Broman, Sjöstedt, Pinhassi, & Dopson, 2017; Graham et al., 2016). Therefore, variations in  
99 microbial community with the level of groundwater table depend on the period over which  
100 groundwater table is varying. With short-duration rainfall events, a single cycle may only cause a  
101 small change in microbial population (Steenwerth, Jackson, Calderón, Scow, & Rolston, 2005),  
102 whereas slow seasonal changes in groundwater table can result in significant differences in DNA  
103 “fingerprint” of microbial populations (Zhou, Zhang, Dong, Lin, & Su, 2015). However, with  
104 more rapid wet/dry cycling, the geochemistry can evolve during initial cycles before a relatively  
105 steady state is reached (RoyChowdhury et al., 2018; Park, Yang, Tsang, Alessi, & Baek, 2018),  
106 suggesting that laboratory studies involving a single wet-dry cycle must be interpreted with  
107 caution.

108 This study investigated temporal changes in the groundwater bacterial community during  
109 cyclic groundwater-table variations in laboratory columns. Two different patterns of  
110 groundwater-table variation were simulated and compared with a static groundwater table. These  
111 represented rainfall infiltration (RI), and natural fluctuations (NF) in groundwater table resulting  
112 from variations in regional extraction and recharge. The bacterial communities were determined  
113 over three successive groundwater-table cycles and correlated with geochemical parameters to

114 determine how the bacterial community varied, both within a cycle and between cycles, as a  
115 function of geochemistry.

116

## 117 **2. Materials and methods**

### 118 *2.1. Experimental system and groundwater-table variation procedure*

119 The pilot-scale aquifer columns, consisting of cylindrical acrylic vessels with a length of  
120 120 cm and an internal diameter of 24 cm (Fig. 1), were established in the Water Sciences  
121 laboratory at Beijing Normal University where the ambient temperature was typically  $28 \pm 0.5$   
122 °C. Fine-grained natural river sand was collected from uncontaminated floodplain sediments near  
123 Cihe (Shijiazhuang, China). Prior to use, the sand was washed with tap water, dried at 105 °C for  
124 10 h, and sieved  $< 0.25$  mm. Properties of the sand are reported in Table 1(a). Aquifer columns  
125 were packed with the sand using a wet-packing procedure (see the supplementary material). The  
126 water was completely drained after packing, when the packed height was 110 cm, the compacted  
127 density was  $1.60 \text{ g cm}^{-3}$ , and the effective porosity was 0.35. O<sub>2</sub>-depleted tap water (prepared by  
128 N<sub>2</sub> sparging for about 60 min; see Table 1(b)) was then injected from the column bottom using a  
129 peristaltic pump until the groundwater table reached a position 40 cm above the bottom. In the  
130 column with a static (ST) groundwater table, no further changes in the groundwater table were  
131 imposed. In the other columns, a cyclic variation in the groundwater table was imposed.

132 In both NF and RI experiments, a static groundwater table was maintained for 12 h, then the  
133 cyclic pattern was commenced. Three full groundwater-table cycles were conducted. In the first  
134 step the groundwater table was raised to 80 cm above the bottom of the columns over a period of



135 100 h. With the NF experiments O<sub>2</sub>-depleted tap water was pumped into the bottom of the  
136 column at a rate of 1.09 ml min<sup>-1</sup> (see the supplementary material), whereas in the RI  
137 experiments tap water (see Table 1(b)) was injected into the top of the column using a second  
138 peristaltic pump (at the same flow rate). Subsequent steps were the same in the NF and RI  
139 experiments; the groundwater table was held static for 40 h, and then was lowered to 40 cm  
140 above the base over a period of 100 h by pumping groundwater out of the bottom of the columns  
141 (flow-rate 1.03 ml min<sup>-1</sup>). The groundwater table was held static for 40 h between cycles and for  
142 12 h after the last cycle. The intended pattern of groundwater-table variation during the NF and  
143 RI experiments is shown in Fig. 2. A straight-forward cyclic pattern was used to allow  
144 comparison between the two regimes, and between the cycles of the same column. However, it  
145 had a similar periodicity to variations that occur in the agricultural region of Central Hebei (part  
146 of the North China Plain) in response to (spring) irrigation and periodic heavy summer rainfall  
147 (Hebei has a temperate continental monsoon climate). The rate at which the water table was  
148 increased (0.4 cm/h) was a compromise between that observed in the Central Hebei during  
149 irrigation (~0.04 cm/h) and that anticipated in response to more intense monsoon rainfall. This  
150 created three zones within the columns, a continuously saturated zone from 0-40 cm above the  
151 bottom of the columns, a zone of intermittent saturation from 40-80 cm, and a vadose zone from  
152 80-110 cm.

## 153 2.2. *Groundwater sampling*

154 Triplicate groundwater samples (30 mL) were collected from the ST, NF and RI columns  
155 after 12, 112, 152, 252, 292, 392, 432, 532, 572, 672, 712, 812 and 824 h (Fig. 2) using sampling

156 ports (VICI, USA) 30 cm above the base of the columns (Fig. 1; near the top of continuously  
157 saturated zone). Each sample was separated into two subsamples, which were used for  
158 geochemical and bacterial analysis, respectively.

### 159 2.3. Analytical methods

#### 160 2.3.1 Geochemical analysis

161 The triplicate subsamples of each groundwater sample were filtered ( $< 0.45 \mu\text{m}$  Millipore),  
162 and stored at  $4 \text{ }^\circ\text{C}$  for further analyses. Each subsample was analyzed less than 24 h after being  
163 collected. Dissolved organic carbon (DOC) was measured using a Vario TOC system (Elementar,  
164 Germany).  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  were analyzed by ion chromatography (Dionex, America). Dissolved  
165 oxygen (DO) was measured at a height of 30 cm above the base of each column (near the top of  
166 continuously saturated zone) by an OXY-10 trace SMA technique (PreSens, Germany) with a  
167 DP-Pst3 dipping probe (PreSens, Germany) (details can be found in the supplementary material).

#### 168 2.3.2 Bacterial community analysis

169 The triplicate subsamples of each groundwater sample were pooled for bacterial analysis,  
170 and labelled as ST1-ST13, NF1-NF13 and RI1-RI13 to indicate the experiment and the sample  
171 number (Fig. 2). Total DNA was extracted from 6 ml of groundwater using a TIANamp Bacteria  
172 DNA Kit (TIANGEN, China) following to the manufacturer's instructions (see the  
173 supplementary material for details). The V3-V4 hypervariable region of 16S rRNA gene was  
174 amplified by PCR using the broad specificity primers 338F  
175 ( $5' \text{-ACTCCTACGGGAGGCAGCAG-3'}$ ) and 806R ( $5' \text{-GGACTACHVGGGTWTCTAAT-3'}$ )  
176 (Zhu et al., 2018). PCR conditions are described in the supplementary material. To reduce PCR

177 errors, amplification for each sample was performed in triplicate and mixed together, then the  
178 amplicons were extracted from 2% agarose gels and purified by using the AxyPrep DNA Gel  
179 Extraction Kit (Axygen Biosciences, USA) following the manufacturer's instructions, and  
180 quantified using QuantiFluor™-ST (Promega, USA). Subsequently, all purified amplicons were  
181 pooled in equimolar concentrations and were paired-end sequenced (2 × 300) on an Illumina  
182 MiSeq PE300 sequencer (Illumina, USA) according to standard protocols at Allwegene  
183 Technology Co., Ltd (Beijing, China).

184       The extraction of high-quality sequences was firstly performed with the QIIME package  
185 (version 1.2.1). Raw sequences were selected based on sequence length, quality, primer and tag,  
186 and low-quality sequences were removed (see the supplementary material). The high-quality  
187 sequences were assigned operational taxonomic units (OTUs) under the threshold of 97%  
188 identity using USEARCH (Version 10). Chimeric sequences were identified and removed using  
189 USEARCH (version 10). OTUs with only one sequence (singleton) were not included in  
190 downstream analysis, other OTUs were assigned to taxonomic groups using the Ribosomal  
191 Database Project (RDP) classifier (version 2.2) against the Silva128 16S rRNA database using a  
192 confidence threshold of 70%.

#### 193 *2.4. Data analysis*

194       Analysis of OTU alpha diversity, including rarefaction (Fig. S1), and calculation of  
195 Shannon index, was performed using Mothur package (version 1.34.4). Heat map was generated  
196 from the relative abundances of top 20 OTUs after column-normalization using pheatmap  
197 package in R (version 3.6.1). Based on the detrended correspondence analysis (DCA) result

198 (gradient length < 3.0), redundancy analysis (RDA) was used to investigate multivariate  
199 correlations between the microbial populations and geochemical properties of the columns using  
200 Canoco (version 4.5). The 100 most abundant OTUs across all the samples were identified, and  
201 their relative abundances in each sample were  $\log(x + 1)$  transformed for the RDA.

202

### 203 **3. Results**

#### 204 *3.1. Geochemistry*

205 The DO concentrations near the top of continuously saturated zone showed the same  
206 responses in the three columns, decreasing from an initial value of  $2.5 \text{ mg L}^{-1}$  to essentially zero  
207 over a period of about 252 h. This corresponded with the end of the first groundwater-table cycle  
208 in the NF and RI columns (Fig. 3a).

209 The DOC concentrations responded differently in the three columns. In the ST column,  
210 DOC increased from an initial value of  $3.5 \text{ mg L}^{-1}$  to  $7.1 \text{ mg L}^{-1}$  after 112 h, then decreased to  
211  $3.8 \text{ mg L}^{-1}$  after about 292 h, and finally remained steady at that value until the end of  
212 experiments (824 h). The DOC in the NF column showed a cyclical response, which established  
213 after the first increase in the groundwater table. Initially it exhibited little change over the first  
214 112 h, then it increased while the groundwater table was static and while it was being lowered to  
215  $6.4 \text{ mg L}^{-1}$  at the end of the cycle. During the subsequent groundwater-table cycles, DOC  
216 decreased during the period when the groundwater table was rising and increased again during  
217 the period when the groundwater table was static or falling. The DOC in the RI column exhibited  
218 a slightly different cyclical response. It initially increased from  $3.5 \text{ mg L}^{-1}$  to  $7.2 \text{ mg L}^{-1}$  during

219 the first time when the groundwater table was rising. Thereafter, it increased during the period  
220 when the groundwater table was falling but decreased during the period when the groundwater  
221 table was static or rising (Fig. 3b).

222 The initial  $\text{NO}_3^-$  concentrations were about  $4.0 \text{ mg L}^{-1}$  in the three columns. In the ST  
223 column,  $\text{NO}_3^-$  increased to a value of  $7.5 \text{ mg L}^{-1}$  after 152 h, before decreasing to  $0.4 \text{ mg L}^{-1}$  after  
224 824 h. In the NF column, the overall trend was a decrease in the  $\text{NO}_3^-$  concentration with time  
225 from its initial value to  $0.3 \text{ mg L}^{-1}$  after 824 h. However, there was a cyclic pattern imposed on  
226 this overall trend, the  $\text{NO}_3^-$  concentration decreased during the period when the groundwater  
227 table was static or rising, and increased when the groundwater table was being lowered. In the RI  
228 column, the pattern in the  $\text{NO}_3^-$  concentration was less clear, with largely an increasing trend to  
229  $8.0 \text{ mg L}^{-1}$  during the first two groundwater-table cycles, and a decrease to  $2.1 \text{ mg L}^{-1}$  followed  
230 by recovery to  $9.2 \text{ mg L}^{-1}$  during the final cycle (Fig. 3c).

231 The initial  $\text{SO}_4^{2-}$  concentrations were about  $42.0 \text{ mg L}^{-1}$  in the three columns. In the ST  
232 column, this increased to a value of  $151.3 \text{ mg L}^{-1}$  after 152 h, and then decreased smoothly to  
233  $11.6 \text{ mg L}^{-1}$  after 824 h. In the NF column, the  $\text{SO}_4^{2-}$  concentration exhibited a cyclical response,  
234 which established after the first increase in the groundwater table, where the  $\text{SO}_4^{2-}$  concentration  
235 increased when the groundwater table was lowered and decreased at it was raised, with an  
236 average of about  $70 \text{ mg L}^{-1}$  during the third groundwater cycle. The  $\text{SO}_4^{2-}$  concentration in the RI  
237 column exhibited a similar cyclical response, but with an average of about  $40 \text{ mg L}^{-1}$  during the  
238 third groundwater cycle (Fig. 3d).

239 *3.2. Bacterial community composition*

240 The diversity of the groundwater bacterial communities exhibits broadly same trend in the  
241 three columns (Fig. 4). The Shannon diversity index remained broadly constant for the first 250  
242 h, then decreased slowly over the next 300 h to about 50% of its initial value, before recovering  
243 over ~100 h to broadly the initial value for the last 150 h of the experiments.

244 The initial populations of all three columns (ST1, NF1, RI1) clustered as small group in the  
245 redundancy plot (Fig. 6), suggesting that these populations were similar. The distribution of  
246 readings by phylum also indicates that these populations were similar (Fig. 5a). The initial  
247 populations of the three columns were all dominated by readings within phyla Parcubacteria  
248 (30-49%), Proteobacteria (21-33%), and Bacteroidetes (2-15%). The three most abundant OTUs  
249 in each column contained ~20% of the readings (Table S1). In the ST column these OTUs were  
250 all classified as Parcubacteria (OTU3 and OTU11 were candidate class Jorgensenbacteria,  
251 OTU35 was unclassified class), whereas in the NF column they were OTU66 (genus  
252 Rhodobacter in the class  $\alpha$ -proteobacteria), OTU87 (genus Fluviicola in the phylum  
253 Bacteroidetes) and OTU23 (unclassified class in the phylum Parcubacteria), and in the RI  
254 column they were OTU4 (candidate class Jorgensenbacteria), OTU93 (Family Cryomorphaceae  
255 in the phylum Bacteroidetes), and OTU23 (Table S2). 33 OTUs were abundant in all three  
256 columns, representing typically a third of each population (Table S1).

257 The populations of the three column experiments at subsequent time points plotted in  
258 different areas of the redundancy plot, suggesting divergence in their populations. After time  
259 point 5 (the start of the second groundwater cycle), the populations of each column experiment

260 formed a loose cluster in slightly different regions of the redundancy plot (populations from the  
261 ST column formed the tightest cluster; Fig. 6). The distribution of readings by phylum exhibited  
262 a less clear trend over the first six time points, with as much variation between time points in the  
263 same column, as between different columns at the same time, but all the populations were  
264 dominated by phyla Parcubacteria and Proteobacteria (typically > 70% of all readings were from  
265 these phyla) (Fig. 5a). Three OTUs were amongst the 20 most abundant OTUs in > 75% of  
266 groundwater samples up to time point 6 (Fig. 5b, they were amongst the 100 most abundant  
267 OTUs in every sample, Table S1), two (OTU3 and OTU4) were classified as Parcubacteria  
268 (candidate class Jorgensenbacteria) and one (OTU1) was classified as Proteobacteria (genus  
269 *Variovorax* belonged to the class  $\beta$ -Proteobacteria).

270 At time point 9 (572 h), when the Shannon diversity index reached its minimum, all three  
271 columns were dominated by readings classified as proteobacteria (see Fig. 5a). At this time, 2/3<sup>rd</sup>  
272 of the readings in all three columns were from OTU2 (genus *Pseudomonas* belonged to the class  
273  $\gamma$ -proteobacteria; 24-59%) and OTU1 (genus *Variovorax*; 11-49 %) mentioned above for its  
274 (albeit more modest) abundance at earlier time points. It should be noted that such increases in  
275 relative abundance may, in part, reflect a drop in overall bacterial abundance.

276 After time point 9, there were fewer common species amongst the abundant OTUs in  
277 groundwater samples from the three columns (Table S1). At time point 13 (824 h), 26 OTUs  
278 typically represented a third of the readings in each column. Further, a small number of highly  
279 abundant OTUs represented a quarter of the readings in each column, and these highly abundant  
280 OTUs were different (Table S1). In the ST column these were OTU3 (candidate class

281 Jorgensenbacteria, phylum Parcubacteria; 10%), OTU10 (order Clostridiales, phylum Firmicutes;  
282 6%), an OTU9 (unclassified bacterium; 3%), OTU28 (genus Opiritatus, phylum Verrucomicrobia;  
283 3%), and OTU36 (family Nitrospiraceae, phylum Nitrospirae; 3%). In the NF column these were  
284 OTU114 (family Comamonadaceae, class  $\beta$ -proteobacteria; 5%), OTU12 (genus Caulobacter,  
285 class  $\alpha$ -proteobacteria; 4%), OTU153 (genus Buchnera, class  $\gamma$ -proteobacteria; 3%), OTU69  
286 (genus Azospirillum, class  $\alpha$ -proteobacteria; 3%), OTU91 (genus Opiritatus, phylum  
287 Verrucomicrobia; 3%), OTU26 (genus Methyloversatilis, class  $\beta$ -proteobacteria; 2%), OTU33  
288 (genus Novosphingobium, class  $\alpha$ -proteobacteria; 2%), OTU138 (order Clostridiales, phylum  
289 Firmicutes; 2%) and OTU1 (genus Variovorax; 2%). In the RI column these were OTU14 and  
290 OTU34 (genus Desulfovirga, class  $\delta$ -proteobacteria; 14% and 3%, respectively) and OTU27  
291 (family Rhodocyclaceae, class  $\beta$ -proteobacteria; 9%).

292 The RDA plot provided correlations between the groundwater bacterial communities and  
293 the geochemical parameters (DO, DOC,  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$ ). At time points 1-3 the bacterial  
294 communities in all three columns were positively correlated with DO, but negatively correlated  
295 thereafter. After time point 4, bacterial communities in the ST column were negatively correlated  
296 with DOC and  $\text{NO}_3^-$ , while bacterial communities in the NF column were positively correlated  
297 with  $\text{SO}_4^{2-}$  and negatively correlated with  $\text{NO}_3^-$  and DOC, and bacterial communities in the RI  
298 column were positively correlated with DOC and  $\text{NO}_3^-$  and negatively correlated with  $\text{SO}_4^{2-}$ .

299



#### 300 **4. Discussion**

301       The initial geochemical response in the continuously saturated soil below the oscillating  
302 zone (or the equivalent zone of the ST column) was similar in the three columns. The DO  
303 concentrations decreased continuously from 2.50 mg/L (~30% oxygen saturation) after column  
304 preparation, to below detection over the first 250 h in the three columns, and did not increase  
305 again despite the potential for air entrapment to occur with NF and RI. This indicates that once  
306 the columns were established the consumption of oxygen within the natural fine-grained river  
307 sand exceeded the O<sub>2</sub> flux to the continuously saturated zone regulated by entrapment, advection  
308 and diffusion (Dutta et al., 2015). The initial decrease in DO is the result of aerobic microbial  
309 metabolism coupled to oxidation of soil organic matter (SOM). The result was that the  
310 biogeochemistry of the continuously saturated zone was essentially anoxic or anaerobic in the  
311 three columns after a time period that corresponded to the end of the first groundwater-table  
312 cycle in the NF and RI columns.

313       In the ST column, DOC increased from ~3.4 mg/L to ~7 mg/L over the first 100 h and then  
314 decreased steadily towards a steady-state value of ~3.8 mg/L after about 250 h. Such a DOC  
315 variation is a footprint of microbial activity (Malik & Gleixner, 2013), as DOC is released by  
316 microbial processing of SOM, although the labile hydrophilic neutral DOC fraction is itself  
317 readily metabolized (Kiikkilä, Kitunen, & Smolander, 2005; Steinbeiss, Temperton, & Gleixner,  
318 2008; Miltner, Bombach, Schmidt-Brücken, & Kästner, 2012). The tap water used in these  
319 experiments contained ~3.4 mg/L DOC, which is likely to be a recalcitrant, hydrophobic acid  
320 DOC fraction that is only degradable on long time scales (Polimene et al., 2018; Kiikkilä et al.,

2005), and thus 3.4 mg/L DOC should be regarded as a baseline for interpreting DOC variations. The difference between the final and initial DOC concentrations may represent an increase in the amount of recalcitrant DOC in the column after the labile fraction of initial DOC pulse has been metabolized, or that there is an equilibrium between continued slow mineralization of SOM and subsequent DOC metabolism. The  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  concentrations initially increased, but both peaked (at 152 h) shortly after the peak in DOC, and then gradually decreased with time as the saturated zone became more reducing. The initial release of  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  from the soil was associated with increased microbial activity as the DO was consumed, whereas their subsequent decrease was the result of their consumption by anaerobic microorganisms as anoxia developed. Although  $\text{SO}_4^{2-}$  reduction coupled to organic matter oxidation is an important process in anaerobic systems (Zhou et al., 2015), it's interesting to find that  $\text{SO}_4^{2-}$  decreased under the presence of  $\text{NO}_3^-$ , which is consistent with the work of Song et al. (2019), who showed that  $\text{SO}_4^{2-}$  and  $\text{NO}_3^-$  were synchronously depleted with DOC decreasing.

In the NF column, DOC concentration decreased when the groundwater table was raised, while it increased when the groundwater table was static at its highest level, when it was lowered, and when it was static at its lowest level. In these experiments water was added or removed from the bottom of the column, displacing the groundwater upwards or downwards as a body. Thus, during an increase in the groundwater table the water at the level of sampling port was replaced by water from the zone just below it, and the displaced water was returned to the vicinity of sampling point when the groundwater table was subsequently lowered (Fig. 2 illustrates how the body of groundwater in the vicinity of the sampling port changes over time). If DOC

342 concentration is taken as an indicator of the rate of SOM mineralization (Song et al., 2018), then  
343 initially there is a decrease in that rate with depth (DOC concentration in the vicinity of sampling  
344 port decreased when the groundwater was displaced upwards and increased when it moved  
345 downwards). The translocation of groundwater into a different region of soil matrix also seems to  
346 increase the rate of DOC mineralization during the subsequent static period. This is probably a  
347 transient effect resulting from the disequilibria associated with the introduction of different  
348 groundwater bacteria and dissolved chemical species to SOM, chemical species and bacteria  
349 associated with the soil matrix. However, it was sufficient to maintain a DOC concentration in  
350 the groundwater near the phreatic surface that was consistently higher than that in the ST column.  
351 The  $\text{NO}_3^-$  concentration in the vicinity of sampling port also decreased when the groundwater  
352 was displaced upwards and increased when it moved downwards, but this pattern diminished  
353 with time. After the first groundwater-table cycle the  $\text{NO}_3^-$  concentration in the NF column  
354 followed a similar trend to that in the ST column, suggesting that anoxia developed at a similar  
355 rate to the ST column and that only a small amount of additional  $\text{NO}_3^-$  was carried from the  
356 intermittently saturated zone into the continuously saturated zone. As a result, most of  $\text{NO}_3^-$  had  
357 been consumed after 800 h at an average rate similar to that during the ST experiments. Like the  
358  $\text{NO}_3^-$  concentration, the  $\text{SO}_4^{2-}$  concentration in the vicinity of sampling port decreased when  
359 groundwater was displaced upwards and increased when it moved downwards, suggesting that  
360  $\text{SO}_4^{2-}$  concentration similarly decreased with depth near the phreatic surface. However, unlike  
361  $\text{NO}_3^-$  concentration, the magnitude of the in-cycle variations in  $\text{SO}_4^{2-}$  concentration was little  
362 changed after three cycles, suggesting the local depth trend in the  $\text{SO}_4^{2-}$  concentration persisted

363 throughout the experiments. Moreover, the transition from  $\text{SO}_4^{2-}$  release when the groundwater  
364 table was static at the lowest level during the first cycle, to  $\text{SO}_4^{2-}$  consumption when the  
365 groundwater table was static at the lowest level during the third cycle, meant the average  $\text{SO}_4^{2-}$   
366 concentration was decreasing slightly with the increasing number of cycles. As in the ST column,  
367 the initial release  $\text{SO}_4^{2-}$  from the soil was associated with increased microbial activity as the DO  
368 was consumed (it's detection at the sampling port was delayed by the position of the  
369 groundwater table). This was probably due to desorption or dissolution of inorganic S from soil  
370 minerals in the intermittently saturated zone, although mineralisation of organic S (either  
371 C-bonded S or ester-bonded sulphates) might also be contributing (Edwards, 1998). The net  
372 decrease in  $\text{SO}_4^{2-}$  concentration from the end of cycle 1 to the end of cycle 3, and particularly the  
373 decrease when the groundwater table was static during cycle 3, were the result of sulphate  
374 reduction by anaerobic microorganisms as anoxia developed.

375 The DOC response in the RI column was initially indistinguishable from the ST column,  
376 but subsequently exhibited a clear cyclic pattern from the point where the groundwater table was  
377 first lowered from its highest level. This pattern was an increase in DOC concentration during  
378 the period when the groundwater table was being lowered, a rapid decrease when the  
379 groundwater table was static at its lowest level, and a slower decrease when the groundwater  
380 table was being increased or static at its highest level. Tap water was injected into the top of the  
381 column, but groundwater was removed from the bottom of column. Thus, during an increase in  
382 the groundwater table, the groundwater in the vicinity of sampling port remained static, but was  
383 replaced by the simulated rainfall from the intermittently saturated zone when the groundwater

384 table was subsequently lowered (Fig. 2). Thus, the increase in DOC when the groundwater table  
385 was lowered is an indication that the rate of SOM mineralization in the intermittently saturated  
386 zone was higher than that in the continuously saturated zone in ~100 h after it had been  
387 inundated with the simulated rainfall. This is associated with the transport of electron acceptors,  
388 such as DO in the simulated rainfall and  $\text{NO}_3^-$  eluted from the vadose zone to the intermittently  
389 saturated zone. The rate of decrease in DOC concentration through the subsequent stages of the  
390 groundwater-table cycle (when groundwater in the vicinity of sampling port was static) reflects a  
391 steady decrease in the rate of SOM mineralization with time (rainfall infiltration into the  
392 intermittently saturated zone later in the cycle may have had a second order effect through  
393 diffusive transport of electron acceptors from the recently saturated zone). The cyclic variations  
394 in  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$  concentrations differed slightly between cycles, but the dominant pattern was  
395 an increase in concentration when the groundwater table was lowered, and predominantly of  
396 consumption when the groundwater table was static or increasing. Thus  $\text{NO}_3^-$  was consumed in  
397 the continuously saturated zone when the groundwater table was static and replenished when the  
398 simulated rainfall was drawn down into the continuously saturated zone. The mechanism of  $\text{NO}_3^-$   
399 replenishment might involve eluting soluble  $\text{NO}_3^-$  from the vadose zone or the intermittently  
400 saturated zone (Huebsch et al., 2014), but almost certainly also involved mineralization of  
401 organic-N. Mineralisation of nitrogen is most rapid when soil is warm, moist and well aerated  
402 (Johnson, Albrecht, Kettrings, Beckman, & Stockin, 2005), so is likely to be enhanced by  
403 periodic inundation by DO containing rainfall. The  $\text{SO}_4^{2-}$  concentration decreased slightly with  
404 successive groundwater-table cycles. Like the other columns, the initial increase of  $\text{SO}_4^{2-}$

405 concentration was associated with increased microbial activity as the DO was consumed.  
406 Similarly, the mechanism was probably desorption or dissolution of inorganic S from soil  
407 minerals, although mineralisation of organic S might also be contributing (Edwards, 1998).  
408 However, the variation in the  $\text{SO}_4^{2-}$  concentration with time differed from the ST column because  
409 each rainfall event mobilised further  $\text{SO}_4^{2-}$  from the vadose zone, but the downward movement  
410 of the groundwater with each cycle prevented  $\text{SO}_4^{2-}$  accumulating near phreatic surface as it did  
411 in the NF columns.

412 The RDA ordination indicates that the groundwater bacterial communities diverged with  
413 time from initially similar populations due to groundwater-table variations (waterborne bacteria  
414 are subset of the bacteria in the columns, but bacteria attached to the soil particles were not  
415 analysed). The initial populations were dominated by phyla Parcubacteria, Proteobacteria and  
416 Bacteroidetes, which are widely found in marine and terrestrial environments (Sun et al., 2019;  
417 León-Zayas et al., 2017). The gradual decrease in bacterial community diversity over the first  
418 two groundwater cycles (time points 1-9) was the result of the selective pressure of rapidly  
419 developing anoxia (Humbert & Dorigo, 2005) and the dominance of OTUs from the initial  
420 bacterial communities immediately being able to exploit the resulting ecological niche. At time  
421 point 9, the dominant populations of all three columns were from the genera *Pseudomonas* and  
422 *Variovorax*. *Pseudomonas* are facultative anaerobes capable of heterotrophic denitrification using  
423 a variety of carbon substrates (Wu et al., 2019; Dolan et al., 2020), and *Variovorax* genus  
424 includes species capable of denitrification and sulphate reduction (Crevecoeur, Vincent, Comte,  
425 & Lovejoy, 2015). The subsequent increase in diversity of each column and the further

426 divergence in their populations are then due to differences in relative competitiveness of the  
427 species present as they adapt to the evolving geochemistry of each column by metabolic  
428 regulation (Ayuso, Acebes, López-Archilla, Montes, & Guerrero, 2009). The final bacterial  
429 population of the ST column was dominated by OTU3 (10% of all readings) and OTU10 (6% of  
430 all readings). The first, which was also abundant in the initial population, was classified as a  
431 Parcubacteria, a phylum of poorly characterised fermentative anaerobes (León-Zayas et al.,  
432 2017). The second, which had low abundance in any of the columns until the start of the third  
433 groundwater cycle, was classified as a Clostridiales, an order of fermentative obligate anaerobes  
434 (Stackebrandt, 2014). Other highly abundant OTUs in the ST13 (together making up 25% of the  
435 population) were closely related to anaerobic nitrate reducers (opitutus; Chin, Liesack, & Janssen,  
436 2001) or sulphate reducers (currently known anaerobes within the Nitrospiraceae family are all  
437 within the genus *Thermodesulfovibrio*; Daims, 2014).

438         The NF column had the most diversity amongst the highly abundant OTUs at time point 13,  
439 with three different  $\alpha$ -proteobacteria, three different  $\beta$ -proteobacteria, a clostridia, a  
440  $\gamma$ -proteobacteria and a Verrucomicrobia representing 25% of all readings. Based on their  
441 similarity to well-characterised species, this population is likely to contain both facultative and  
442 obligate anaerobes, including species capable of nitrate and sulphate reduction (Willems, 2014;  
443 Chin et al., 2001; Smalley et al., 2015; Kaksonen, Spring, Schumann, Kroppenstedt, & Puhakka,  
444 2007). What is really notable about these OTUs is that most were not abundant in the NF and RI  
445 columns until the third groundwater cycle and not at all in the ST column (the exceptions were  
446 OTU1, *Variovorax*, which was ubiquitous throughout all three column experiments, and OTU114,

447 Comamonadaceae family, which was briefly abundant in the ST5). This suggests that it can take  
448 several groundwater cycles for the bacterial populations to evolve to fully exploit the  
449 geochemical conditions produced by a varying water-table, possibly because the geochemistry  
450 itself varies during the cycles.

451 The final bacterial population of the RI column had the least diversity amongst the highly  
452 abundant OTUs, with just three OTUs representing 25% of the population. Two were classified  
453 as *Desulfovirga* (a sulfate-reducing strict anaerobe; Kaksonen et al., 2007) within the class  
454  $\delta$ -proteobacteria (OTU14 and OTU34; 14% and 3%, respectively), and the other was classified  
455 to the family Rhodocyclaceae (OTU27; 9%) within the class  $\beta$ -proteobacteria (a disparate class  
456 of mainly facultative anaerobic bacteria that includes many nitrate reducers; Oren, 2014). Other  
457 abundant OTUs (OTU117 and OTU28, each representing ~2% of the population) were closely  
458 related to anaerobic nitrate reducers (Thrash, Ahmadi, Torok, & Coates, 2010; Chin et al., 2001).

459 In summary, the RDA indicates that the bacterial populations of all three columns evolved  
460 slightly differently over the course of the experiments, presumably as each community adapted  
461 to its specific geochemical environment. However, the principal species in time point 13 suggest  
462 that the population of the ST column has evolved the least (the most abundant OTU was  
463 abundant in the initial population), and the high abundance of a probable fermentative species  
464 (OTU10) may reflect the lower concentration of electron acceptors, such as sulphate and nitrate,  
465 that support anaerobic respiration. The RDA indicates that populations of the NF and RI columns  
466 differed from each other at time point 13, which may reflect the low nitrate, but elevated sulphate  
467 concentrations in the NF column, and elevated nitrate, but moderate sulphate concentrations in



468 the RI column. However, nitrate and sulphate reducers appeared to be abundant in both bacterial  
469 populations, perhaps indicating that even the modest nitrate release from the intermittently  
470 saturated influenced the bacterial population of the NF column.

471

## 472 **5. Conclusions**

473 A cyclically varying groundwater table in an uncontaminated fine-grained natural river sand  
474 representative of aquifer soils increased the rate of SOM mineralization in the continuously  
475 saturated zone immediately below the intermittently saturated zone in comparison with a control  
476 experiment with a static groundwater table. This was an anoxic zone, and enhanced  
477 mineralization appeared to be associated with greater availability of electron accepting  
478 compounds, particularly  $\text{NO}_3^-$  and  $\text{SO}_4^{2-}$ . When variations in the groundwater table resulted from  
479 natural fluctuations,  $\text{NO}_3^-$  was consumed at similar rate to that observed with a static  
480 groundwater table.  $\text{SO}_4^{2-}$  was also consumed but was replenished by periodic wetting of the  
481 intermittently saturated zone. However, when the varying groundwater table resulted from  
482 rainfall infiltration,  $\text{NO}_3^-$  was consumed when the groundwater table was static, but was  
483 replenished when the simulated rainfall percolated down into the continuously saturated zone.  
484 The mechanism of  $\text{NO}_3^-$  replenishment might involve eluting soluble  $\text{NO}_3^-$  from the vadose zone,  
485 but almost certainly also involved mineralization of organic-N, which was likely to be enhanced  
486 by periodic inundation by DO containing rainfall.

487 The RDA ordination indicated that the groundwater bacterial communities at the top of  
488 continuously saturated zone of the NF and RI columns diverged from the ST column with time

489 from initially similar populations due to groundwater-table variations. However obvious  
490 differences in the most abundant OTUs in the three experiments only emerged after two  
491 complete groundwater cycles (~500 h). In conclusion, variations in the water table and,  
492 furthermore, the local flow direction during recharge have a strong influence on the  
493 geochemistry and microbiology of the groundwater bacterial community just below the phreatic  
494 surface of an aquifer.

495

#### 496 **Declarations of competing interest**

497 None.

498

#### 499 **Data availability statement**

500 The data used to support the findings of this study are available from the corresponding  
501 authors upon reasonable request.

502

#### 503 **Supporting file legend**

504 Supplementary Material.

505

#### 506 **References**

507 Ayuso, S. V., Acebes, P., López-Archilla, A. I., Montes, C., & Guerrero, M. C. (2009).

508 Environmental factors controlling the spatiotemporal distribution of microbial communities  
509 in a coastal, sandy aquifer system (Doñana, southwest Spain). *Hydrogeology Journal*, 17(4),

510 767-780.

511 Banks, M. K., Clennan, C., Dodds, W., & Rice, C. (1999). Variations in microbial activity due to  
512 fluctuations in soil water content at the water table interface. *Journal of Environmental*  
513 *Science & Health Part A*, 34(3), 479-505.

514 Braun, B., Schröder, J., Knecht, H., & Szewzyk, U. (2016). Unraveling the microbial community  
515 of a cold groundwater catchment system. *Water Research*, 107, 113-126.

516 Broman, E., Sjöstedt, J., Pinhassi, J., & Dopson, M. (2017). Shifts in coastal sediment  
517 oxygenation cause pronounced changes in microbial community composition and  
518 associated metabolism. *Microbiome*, 5, 96.

519 Chin, K. J., Liesack, W., & Janssen, P. H. (2001). *Opiritatus terrae* gen. nov., sp. nov., to  
520 accomodate novel strains of the division 'Verrucomicrobia' isolated from rice paddy soil.  
521 *International Journal of Systematic and Evolutionary Microbiology*, 51, 1965-1968.

522 Crevecoeur, S., Vincent, W. F., Comte, J., & Lovejoy C. (2015). Bacterial community structure  
523 across environmental gradients in permafrost thaw ponds: methanotroph-rich ecosystems.  
524 *Frontiers in Microbiology*, 6, 192.

525 Daims, H. (2014). The Family Nitrospiraceae. In: E. Rosenberg, E. F. DeLong, S. Lory, E.  
526 Stackebrandt, & F. Thompson (Eds.), *The Prokaryotes* (pp. 733-749). Heidelberg, Berlin:  
527 Springer.

528 Dobson, R., Schroth, M. H., & Zeyer, J. (2007). Effect of water-table fluctuation on dissolution  
529 and biodegradation of a multi-component, light nonaqueous-phase liquid. *Journal of*  
530 *Contaminant Hydrology*, 94(3-4), 235-248.

531 Dolan, S. K., Kohlstedt, M., Trigg, S., Vallejo Ramirez, P., Kaminski, C. F., Wittmann, C., &  
532 Welch, M. (2020). Contextual flexibility in *Pseudomonas aeruginosa* central carbon  
533 metabolism during growth in single carbon sources. *mBio*, *11*(2), e02684-19.

534 Dutta, T., Carles-Brangarí, A., Fernández-García, D., Rubol, S., Tirado-Conde, J., &  
535 Sanchez-Vila, X. (2015). Vadose zone oxygen (O<sub>2</sub>) dynamics during drying and wetting  
536 cycles: An artificial recharge laboratory experiment. *Journal of Hydrology*, *527*, 151-159.

537 Edwards, P. J. (1998). Sulfur cycling, retention, and mobility in soils: A review. *USDA, General*  
538 *Technical Report NE-250*.

539 Graham, E., Crump, A. R., Resch, C. T., Fansler, S., Arntzen, E., Kennedy, D., ... Stegen, J. C.  
540 (2016). Coupling spatiotemporal community assembly processes to changes in microbial  
541 metabolism. *Frontiers in Microbiology*, *7*, 1949.

542 Griebler, C., Malard, F., & Lefébure, T. (2014). Current developments in groundwater  
543 ecology-from biodiversity to ecosystem functioning and services. *Current Opinion in*  
544 *Biotechnology*, *27*, 159-167.

545 Haack, S. K., Fogarty, L. R., West, T. G, Alm, E. W., Mcguire, J. T., Long, D. T., ... Forney, L. J.  
546 (2004). Spatial and temporal changes in microbial community structure associated with  
547 recharge-influenced chemical gradients in a contaminated aquifer. *Environmental*  
548 *Microbiology*, *6*(5), 438-48.

549 Huebsch, M., Fenton, O., Horan, B., Hennessy, D., Richards, K. G., Jordan, P., ... Blum, P.  
550 (2014). Mobilisation or dilution? Nitrate response of karst springs to high rainfall events.  
551 *Hydrology and Earth System Sciences*, *11*(4), 4131-4161.

552 Humbert, J. F., & Dorigo, U. (2005). Biodiversity and aquatic ecosystem functioning: A mini  
553 review. *Aquatic Ecosystem Health & Management*, 8(4), 367-374.

554 Igor, S. (1993). World fresh water resources. In: P. H. Gleick (Ed.) *Water in crisis: A guide to the*  
555 *world's fresh water resources*. Oxford, New York: Oxford University Press.

556 Johnson, C., Albrecht, G., Kettrings, Q., Beckman, J., & Stockin, K. (2005). Nitrogen basics-the  
557 nitrogen cycle. *Agronomy Fact Sheet Series, Fact Sheet 2*. Cornell University, Cooperative  
558 Extension.

559 Kaksonen, A. H., Spring, S., Schumann, P., Kroppenstedt, R. M., & Puhakka, J. A. (2007).  
560 *Desulfurispora thermophila* gen. nov., sp. nov., a thermophilic, spore-forming  
561 sulfate-reducer isolated from a sulfidogenic fluidized-bed reactor. *International Journal of*  
562 *Systematic and Evolutionary Microbiology*, 57(5), 1089-1094.

563 Kim, B. H., & Gadd, G. M. (2008). *Bacterial Physiology and Metabolism*. New York: Cambridge  
564 University Press.

565 Kiikkilä, O., Kitunen, V., & Smolander, A. (2005). Degradability of dissolved soil organic carbon  
566 and nitrogen in relation to tree species. *FEMS Microbiology Ecology*, 53(1), 33-40.

567 Krause, S., Bronstert, A., & Zehe, E. (2007). Groundwater-surface water interactions in a North  
568 German lowland floodplain-Implications for the river discharge dynamics and riparian  
569 water balance. *Journal of Hydrology*, 347(3-4), 404-417.

570 León-Zayas, R., Peoples, L., Biddle, J. F., Podell, S., Novotny, M., Cameron, J., ... Bartlett, D. H.  
571 (2017). The metabolic potential of the single cell genomes obtained from the Challenger  
572 Deep, Mariana Trench within the candidate superphylum Parcubacteria (OD1).

573 *Environmental Microbiology*, 19(7), 2769-2784.

574 Malik, A., & Gleixner, G. (2013). Importance of microbial soil organic matter processing in  
575 dissolved organic carbon production. *FEMS Microbiology Ecology*, 86(1), 139-148.

576 Medihala, P. G., Lawrence, J. R., Swerhone, G. D. W., & Korber, D. R. (2012). Effect of  
577 pumping on the spatio-temporal distribution of microbial communities in a water well field.  
578 *Water Research*, 46(4), 1286-1300.

579 Miltner, A., Bombach, P., Schmidt-Brücken, B., & Kästner, M. (2012). SOM genesis: Microbial  
580 biomass as a significant source. *Biogeochemistry*, 111(1-3), 41-55.

581 Oren, A. (2014). The Family Rhodocyclaceae. In: E. Rosenberg, E. F. DeLong, S. Lory, E.  
582 Stackebrandt, & F. Thompson (Eds.), *The Prokaryotes* (pp. 975-998). Heidelberg, Berlin:  
583 Springer.

584 Park, S. M., Yang, J. S., Tsang, D. C. W., Alessi, D. S., & Baek, K. (2018). Enhanced irreversible  
585 fixation of cesium by wetting and drying cycles in soil. *Environmental Geochemistry &*  
586 *Health*, 41(1), 149-157.

587 Pett-Ridge, J., & Firestone, M. K. (2005). Redox fluctuation structures microbial communities in  
588 a wet tropical soil. *Applied and Environmental Microbiology*, 71(11), 6998-7007.

589 Polimene, L., Rivkin, R. B., Luo, Y. W., Kwon, E. Y., Gehlen, M., Peña, M. A., ... Jiao, N. Z.  
590 (2018). Modelling marine DOC degradation time scales. *National Science Review*, 5(4),  
591 468-474.

592 Rainwater, K., Mayfield, M. P., Heintz, C., & Claborn, B. J. (1993). Enhanced in situ  
593 biodegradation of diesel fuel by cyclic vertical water table movement: Preliminary studies.

594 *Water Environment Research*, 65(6), 717-725.

595 Rezanezhad, F., Couture, R. M., Kovac, R., O'Connell, D., & Van Cappellen, P. (2014). Water  
596 table fluctuations and soil biogeochemistry: An experimental approach using an automated  
597 soil column system. *Journal of Hydrology*, 509, 245-256.

598 Rosenberg, D. B., & Freedman, S. M. (1994). Temporal heterogeneity and ecological community  
599 structure. *International Journal of Environmental Studies*, 46(2-3), 97-102.

600 RoyChowdhury, T., Bramer, L., Hoyt, D. W., Kim, Y. M., Metz, T. O., McCue, L. A., ... Bailey,  
601 V. (2018). Temporal dynamics of CO<sub>2</sub> and CH<sub>4</sub> loss potentials in response to rapid  
602 hydrological shifts in tidal freshwater wetland soils. *Ecological Engineering*, 114, 104-114.

603 Rühle, F. A., von Netzer, F., Lueders, T., & Stumpp, C. (2015). Response of transport parameters  
604 and sediment microbiota to water table fluctuations in laboratory columns. *Vadose Zone*  
605 *Journal*, 14(5), 12.

606 Shade, A., Jones, S. E., & McMahon, K. (2008). The influence of habitat heterogeneity on  
607 freshwater bacterial community composition and dynamics. *Environmental Microbiology*,  
608 10(4), 1057-1067.

609 Smalley, N. E., Taipale, S., De Marco, P., Doronina, N. V., Kyrpides, N., Shapiro, N., ...  
610 Kalyuzhnaya, M. G. (2015). Functional and genomic diversity of methylotrophic  
611 Rhodocyclaceae: Description of *Methyloversatilis discipulorum* sp. nov. *International*  
612 *Journal of Systematic & Evolutionary Microbiology*, 65(7), 2227-2233.

613 Song, N., Xu, H., Yan, Z., Yang, T., Wang, C., & Jiang, H. L. (2019). Improved lignin  
614 degradation through distinct microbial community in subsurface sediments of one eutrophic

615 lake. *Renewable Energy*, 138, 861-869.

616 Song, Y. Y., Song, C. C., Hou, A. X., Ren, J. S., Wang, X. W., Cui, Q., & Wang, M. Q. (2018).  
617 Effects of temperature and root additions on soil carbon and nitrogen mineralization in a  
618 predominantly permafrost peatland. *Catena*, 165, 381-389.

619 Stackebrandt, E. (2014). The Family Clostridiaceae, Other Genera. In: E. Rosenberg, E. F.  
620 DeLong, S. Lory, E. Stackebrandt, & F. Thompson (Eds.), *The Prokaryotes* (pp. 67-73).  
621 Heidelberg, Berlin: Springer.

622 Steenwerth, K. L., Jackson, L. E., Calderón, F. J., Scow, K. M., & Rolston, D. E. (2005).  
623 Response of microbial community composition and activity in agricultural and grassland  
624 soils after a simulated rainfall. *Soil Biology & Biochemistry*, 37(12), 2249-2262.

625 Steinbeiss, S., Temperton, V. M., & Gleixner, G. (2008). Mechanisms of short-term soil carbon  
626 storage in experimental grasslands. *Soil Biology & Biochemistry*, 40(10), 2634-2642.

627 Stegen, J. C., Fredrickson, J. K., Wilkins, M. J., Konopka, A. E., Nelson, W. C., Arntzen, E.  
628 V., ... Tfaily, M. (2016). Groundwater-surface water mixing shifts ecological assembly  
629 processes and stimulates organic carbon turnover. *Nature Communications*, 7, 11237.

630 Sun, Y., Li, X., Liu, J. J., Yao, Q., Jin, J., Liu, X. B., & Wang, G. H. (2019). Comparative  
631 analysis of bacterial community compositions between sediment and water in different  
632 types of wetlands of northeast china. *Journal of Soils and Sediments*, 19, 3083-3097.

633 Thrash, J. C., Ahmadi, S., Torok, T., & Coates, J. D. (2010). *Magnetospirillum bellicus* sp. nov.,  
634 a novel dissimilatory perchlorate-reducing alphaproteobacterium isolated from a  
635 bioelectrical reactor. *Applied and Environmental Microbiology*, 76(14), 4730-4737.



636 Van Driezum, I. H., Chik, A. H. S., Jakwerth, S., Lindner, G., Farnleitner, A. H., Sommer, R., ...  
637 Kirschner, A. K. T. (2018). Spatiotemporal analysis of bacterial biomass and activity to  
638 understand surface and groundwater interactions in a highly dynamic riverbank filtration  
639 system. *Science of the Total Environment*, 627, 450-461.

640 Willems, A. (2014). The Family Comamonadaceae. In: E. Rosenberg, E. F. DeLong, S. Lory, E.  
641 Stackebrandt, & F. Thompson (Eds.), *The Prokaryotes* (pp. 771-851). Heidelberg, Berlin:  
642 Springer.

643 Wu, Z. S., Xu, F., Yang, C., Su, X. X., Guo, F. C., Xu, Q. Y., ... Chen, Y. (2019). Highly efficient  
644 nitrate removal in a heterotrophic denitrification system amended with redox-active biochar:  
645 A molecular and electrochemical mechanism. *Bioresource Technology*, 275, 297-306.

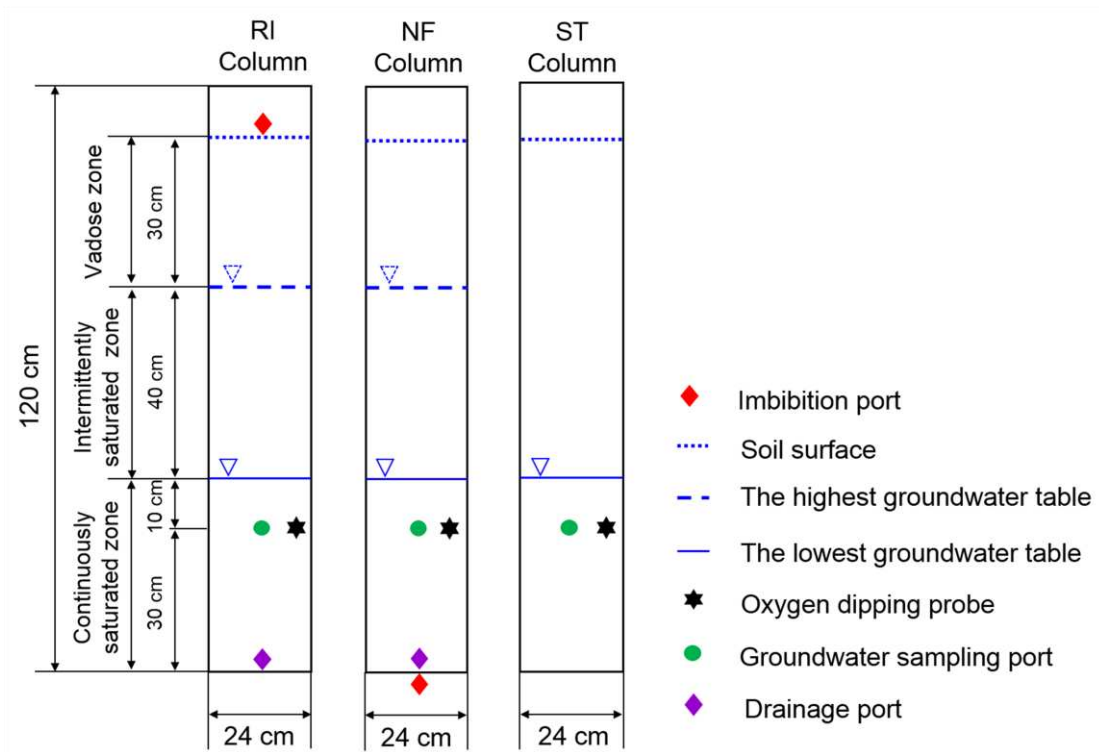
646 Yang, Y. S., Li, P. P., Zhang, X., Li, M. J., Lu, Y., Xu, B., & Yu, T. (2017). Lab-based  
647 investigation of enhanced BTEX attenuation driven by groundwater table fluctuation.  
648 *Chemosphere*, 169, 678-684.

649 Zheng, T. L., Deng, Y. M., Wang, Y. X., Jiang, H. C., O'Loughlin, E. J., Flynn, T. M., ... Ma, T.  
650 (2019). Seasonal microbial variation accounts for arsenic dynamics in shallow alluvial  
651 aquifer systems. *Journal of Hazardous Materials*, 367, 109-119.

652 Zhou, A. X., Zhang, Y. L., Dong, T. Z., Lin, X. Y., & Su, X. S. (2015). Response of the microbial  
653 community to seasonal groundwater level fluctuations in petroleum  
654 hydrocarbon-contaminated groundwater. *Environmental Science and Pollution Research*,  
655 22(13), 10094-10106.

656 Zhou, Y. X., Kellermann, C., & Griebler, C. (2012). Spatio-temporal patterns of microbial  
657 communities in a hydrologically dynamic pristine aquifer. *FEMS Microbiology Ecology*,  
658 81(1), 230-242.

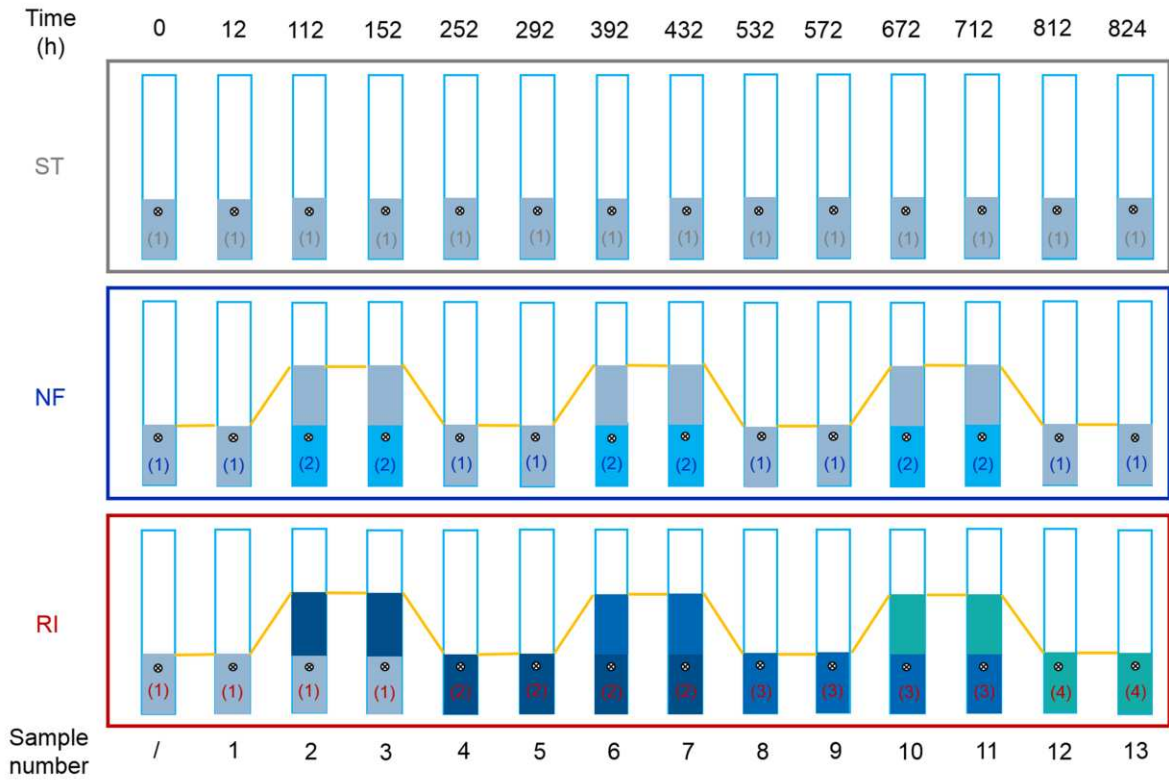
659 Zhu, F. X., Zhu, C. Y., Doyle, E., Liu, H. L., Zhou, D. M., & Gao, J. (2018). Fate of di (2  
660 ethylhexyl) phthalate in different soils and associated bacterial community changes. *Science*  
661 *of the Total Environment*, 637-638, 460-469.



662

663 **Fig. 1.** Schematic of the experimental system showing the static (ST), natural fluctuations (NF),  
 664 and rainfall infiltration (RI) columns.

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666

667 **Fig. 2.** Schematic representation of the experimental timelines showing the sampling points. The

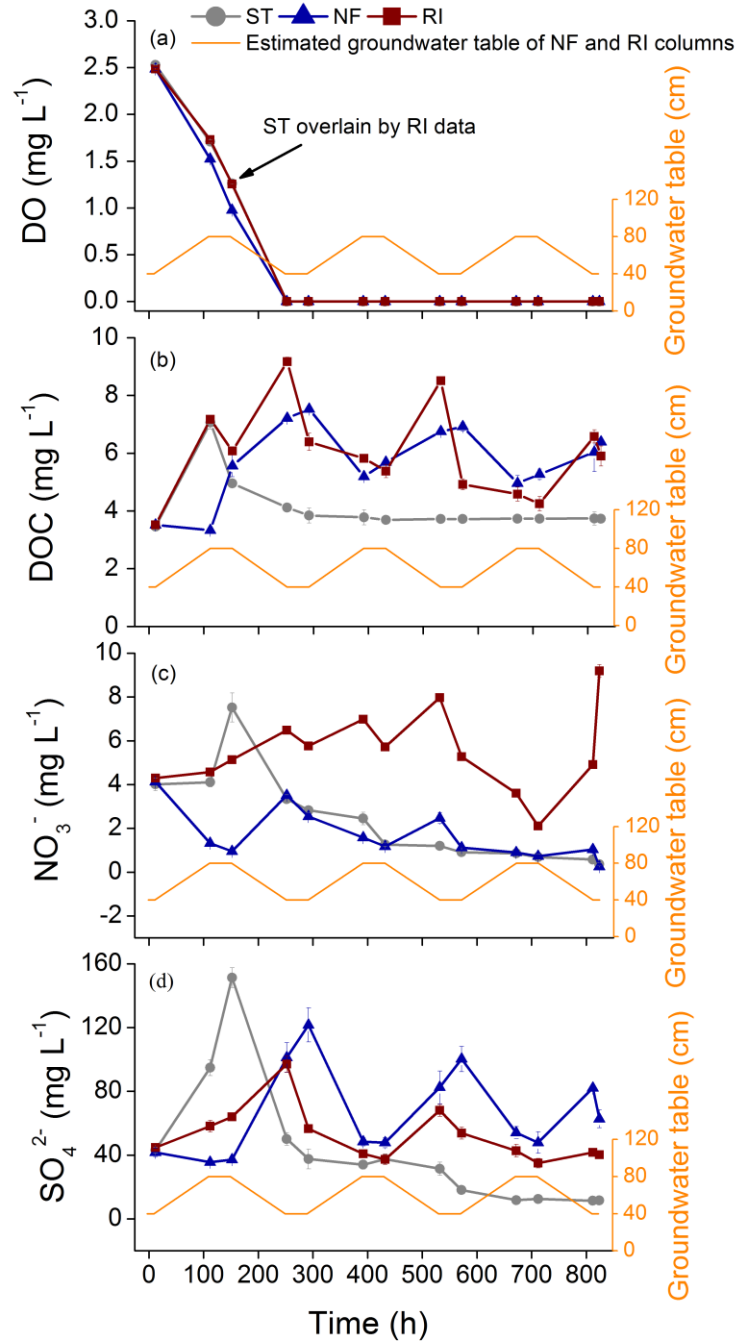
668 grey (1), light blue (2), mid-blue (3) and dark blue (4) shading represents the movement of the

669 groundwater body in the static (ST), natural fluctuations (NF) and rainfall infiltration (RI)

670 columns. Orange lines represent the intended pattern of groundwater-table in the NF and RI

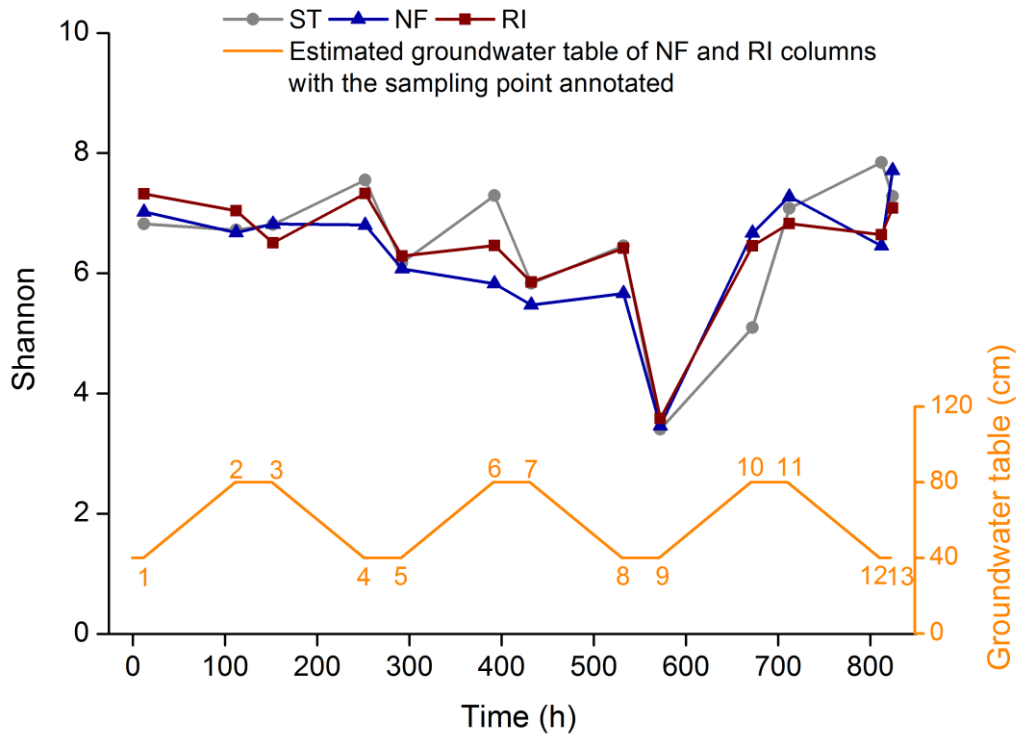
671 columns.

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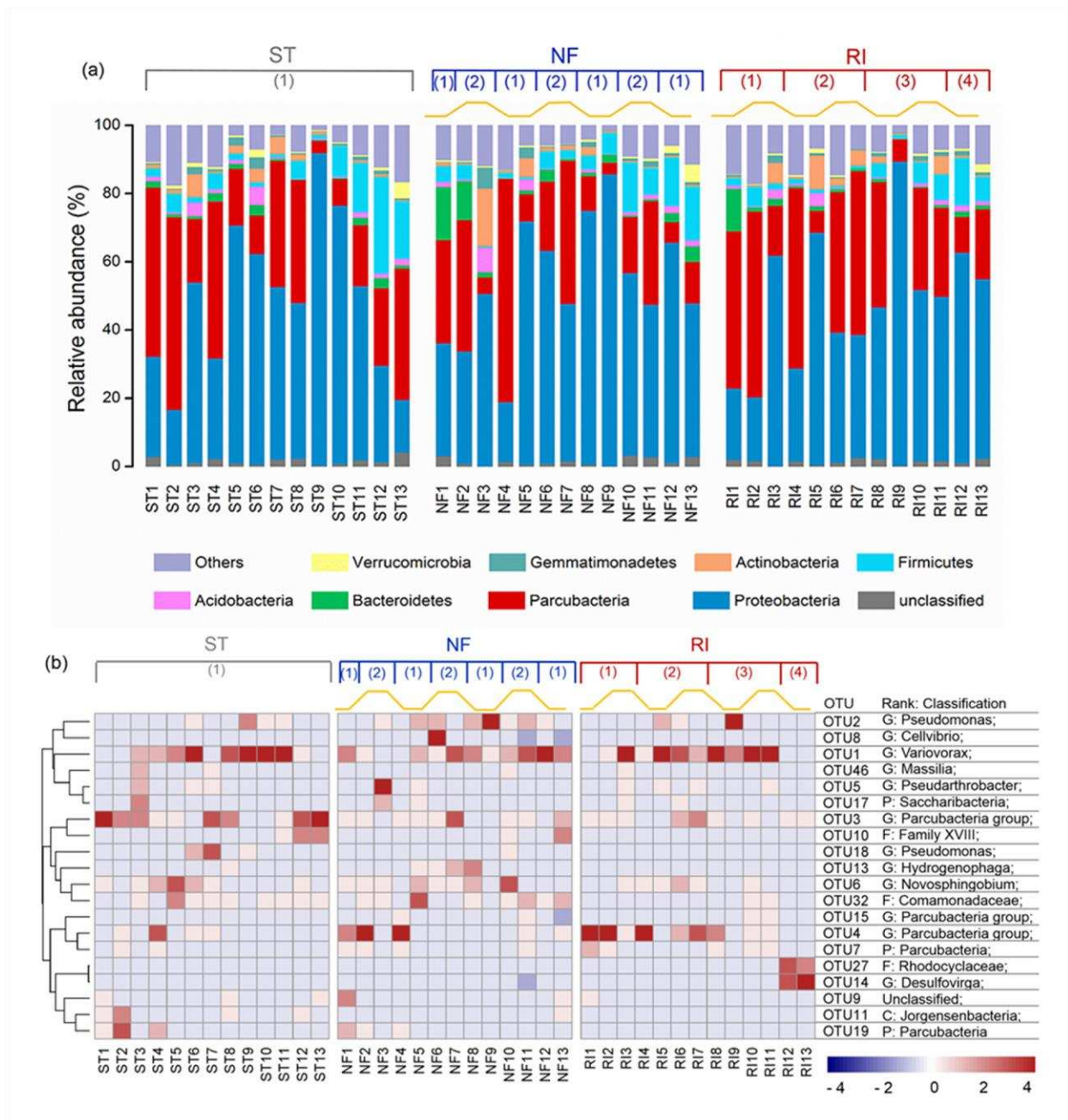
674 **Fig. 3.** Variations of (a) dissolved oxygen (DO); (b) DOC; (c)  $\text{NO}_3^-$  and (d)  $\text{SO}_4^{2-}$  in  
 675 groundwater samples from 10 cm below the surface of continuously saturated zone when the  
 676 water-table is static (ST), and when natural fluctuations (NF) and the rainfall infiltration (RI)  
 677 cause cyclic variations (the error bars represent the standard deviations of the mean values from  
 678 triplicate measurements).



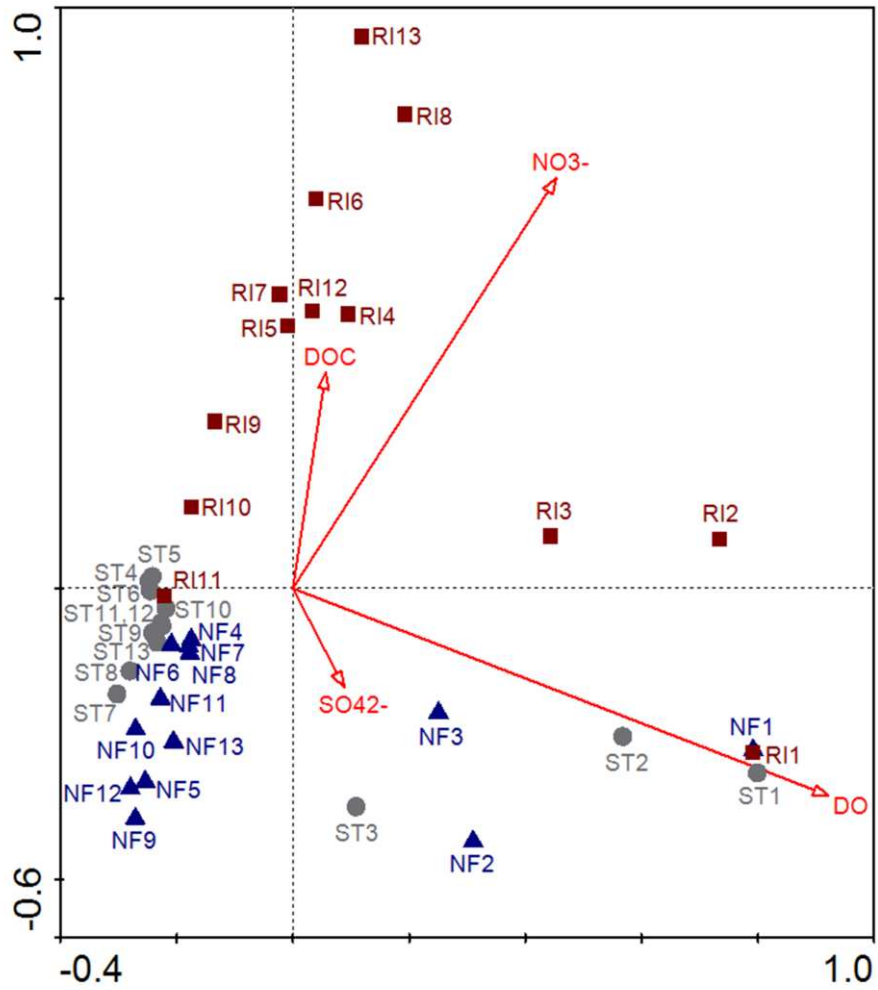
680

681 **Fig. 4.** Shannon index of bacterial community diversity in groundwater samples from 10 cm  
 682 below the surface of continuously saturated zone when the water-table is static (ST), and when  
 683 natural fluctuations (NF) and the rainfall infiltration (RI) cause cyclic variations.

684



685  
 686 **Fig. 5.** (a) Relative abundance of the main phyla (relative abundance > 0.1 %) and (b) heat map  
 687 of the 20 most abundant OTUs in groundwater samples from 10 cm below the surface of  
 688 continuously saturated zone when the water-table is static (ST), and when natural fluctuations  
 689 (NF) and the rainfall infiltration (RI) cause cyclic variations (heat map: red indicates high relative  
 690 abundance and blue indicates low relative abundance). Orange lines at the top of (a) and (b)  
 691 represent the intended pattern of groundwater-table in the NF and RI columns.



693

694 **Fig. 6.** Redundancy analysis (RDA) showing correlations between bacterial community structure  
695 and key geochemical parameters (bacterial community structure was represented by the relative  
696 abundance of the 100 most abundant OTUs across all samples). Grey circles, blue triangles and  
697 dark red squares represent the static (ST), natural fluctuations (NF) and rainfall infiltration (RI)  
698 columns, respectively.



699 Table 1:

700 (a) Basic properties of the fine-grained natural river sand.

<b>Parameter</b>		<b>Value</b>
	Water saturation (%)	0.10
	Bulk density (g cm <sup>-3</sup> )	1.32
	TOC (g kg <sup>-1</sup> )	1.46
	TN (g kg <sup>-1</sup> )	0.20
	TS (g kg <sup>-1</sup> )	0.11
Size distribution (%)	< 0.02 mm	4.36
	0.02-0.1 mm	11.14
	0.1-0.25 mm	13.84
	0.25-1 mm	64.81
	> 1 mm	5.85

701 (b) Basic properties of the water.

<b>Parameter</b>		<b>Value</b>
DO (mg L <sup>-1</sup> )	oxygen-depleted tap water	1 ± 0.2
	tap water	8 ± 0.5
	DOC (mg L <sup>-1</sup> )	3.39
	NO <sub>3</sub> <sup>-</sup> (mg L <sup>-1</sup> )	3.71
	SO <sub>4</sub> <sup>2-</sup> (mg L <sup>-1</sup> )	30.44

702