

This is a repository copy of *Linking dimensions of data on global marine animal diversity*.

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/165370/

Version: Accepted Version

Article:

Webb, T.J. orcid.org/0000-0003-3183-8116 and Vanhoorne, B. (2020) Linking dimensions of data on global marine animal diversity. Philosophical Transactions of the Royal Society B: Biological Sciences, 375 (1814). ISSN 0962-8436

https://doi.org/10.1098/rstb.2019.0445

© 2020 The Authors. This is an author-produced version of a paper accepted for publication in Philosophical Transactions of the Royal Society B: Biological Sciences. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/

- 1 Linking dimensions of data on global marine animal diversity
- 2 Thomas J. Webb₁, Bart Vanhoorne₂

3 1. Department of Animal and Plant Sciences, University of Sheffield, S10 2TN, UK,

4 t.j.webb@sheffield.ac.uk

5 2. Flanders Marine Institute (VLIZ), Ostend, Belgium

6 Abstract

7 Recent decades have seen an explosion in the amount of data available on all aspects of 8 biodiversity, which has led to data-driven approaches to understand how and why diversity 9 varies in time and space. Global repositories facilitate access to various classes of species-10 level data including biogeography, genetics, and conservation status, which are in turn 11 required to study different dimensions of diversity. Ensuring that these different data sources 12 are interoperable is a challenge as we aim to create synthetic data products to monitor the 13 state of the world's biodiversity. One way to approach this is to link data of different classes, 14 and to inventory the availability of data across multiple sources. Here, we use a 15 comprehensive list of >200,000 marine animal species, and quantify the availability of data 16 on geographic occurrences, genetic sequences, conservation assessments, and DNA 17 barcodes across all phyla and broad functional groups. This reveals a very uneven picture: 18 44% of species are represented by no record other than their taxonomy, but some species 19 are rich in data. Although these data-rich species are concentrated into a few taxonomic and 20 functional groups, especially vertebrates, data is spread widely across marine animals, with 21 members of all 32 phyla represented in at least one database. By highlighting gaps in 22 current knowledge, our census of marine diversity data helps to prioritise future data 23 collection activities, as well as emphasising the importance of ongoing sustained 24 observations and archiving of existing data into global repositories.

25 Introduction

26 The explosion in the availability of data describing the natural world has, in recent decades, 27 transformed the kinds of questions that we can now ask as ecologists. Efforts to reconstruct 28 the evolutionary relationships between all living species (e.g. Open Tree of Life; [1,2]) can 29 draw upon over 200M sequences (https://www.ncbi.nlm.nih.gov/genbank/statistics/) from over 170,000 metazoan species stored in GenBank [3,4]. In 2018, the Global Biodiversity 30 31 Information Facility (GBIF, [5]) passed a billion species occurrence records (https://www.gbif.org/news/5BesWzmwgQ4U84sugWyOQy/big-data-for-biodiversity-gbiforg-32 33 surpasses-1-billion-species-occurrences), providing an unparalleled resource for students of biogeography. The conservation status of >116,000 species has now been formally 34 35 assessed [6]. Significant efforts are underway to collate data biological, physiological, 36 metabolic and thermal traits [7-11] across multiple species, as well as information on animal

movement [12,13] and ecological interactions [14].

38

37

39 Against this background of increased data availability, the oceans are still often 40 characterised as the data-poor relative of the data-rich land. Various autonomous platforms 41 operating throughout the world's oceans do now enable vast quantities of physical and 42 biogeochemical data to be transmitted [15] but marine biodiversity data remain more 43 challenging to collect. In part, the vastness of the oceans precludes routine and casual 44 observation by the citizen scientists who have contributed so much to the collection of 45 terrestrial biodiversity data [16,17], except in some more accessible coastal areas [18-20]. 46 However coordinated global initiatives have made enormous progress in collating existing 47 data and promoting systematic new data collection. The Census of Marine Life [21] drove 48 this effort from 2000-2010, and its legacies include the Ocean Biodiversity Information 49 System (OBIS, [22]), which currently holds nearly 60M occurrence records from over 50 120,000 marine species. Initiatives like this have built on sustained observations of marine 51 ecosystems [23], and continue to be developed to deliver the Essential Biodiversity

52 Variables that we need to monitor progress towards Sustainable Development Goals (e.g. 53 [24]). Application of technologies from satellites and drones to biologgers and molecular 54 methods such as eDNA continue to expand the range of data available to marine biodiversity 55 scientists [25]. Crucially, the accumulation of data has proceeded in parallel with massive 56 improvements in data infrastructure, and much better tools (taking advantage of the 57 improved computing power available even to casual users) with which to access and 58 analyse it [26,27]. This is important because the challenge now is to extract meaning from 59 the sea of data, to deliver effective outcomes for marine conservation and monitoring of the 60 state of the global ocean [19,24].

61

62 Although access to biodiversity data of different types is now much improved, to extract full 63 value from existing data requires linking together different datasets that were often collected 64 for different purposes, by different organisations and at different times. This kind of 65 interoperability of diversity data is central to the vision of a 'macroscope' to sample and 66 monitor the entire biosphere [25], and is a fundamental principle of the Bari Manifesto of best 67 practice in biodiversity informatics [28]. Progress towards such interoperability requires 68 comparable coverage across multiple classes of data and dimensions of diversity, as well as 69 parallel measures of the abiotic environment and of human pressures. An exemplar of 70 successful data integration for terrestrial plant communities is the Botanical Information and 71 Ecology Network [29] which combines standardised information on plant distributions, traits, 72 and evolutionary relationships with the computational tools needed to work with them. An 73 important step towards this kind of model is to fully understand the gaps and biases in 74 available data. In the marine environment, key gaps in the overall knowledge of marine 75 biodiversity have been documented [30-32], including estimates of the extent of unknown 76 biodiversity [33] and undocumented extinction risk [34]. Efforts to quantify these gaps across 77 different key variables and data sources have been limited to the regional scale, but have 78 shown for instance that the species and taxonomic groups that we know most in one 79 dimension (e.g. global occurrences) tend to be those that we also know most about in

another (e.g. biological traits, extinction risk; [34,35]). To date we lack a global overview of
how data (and gaps) are co-distributed across axes of marine diversity, to compare for
example with previous global analyses of terrestrial plants [36].

83

84 Such a task is feasible however, given the availability of a standardised global taxonomy of 85 marine species, the World Register of Marine Species (WoRMS, [37]), which includes links 86 out to other key biodiversity datasets (Table 1). In this paper, we focus on key data sources 87 which, when linked to robust taxonomy, individually or in combination can be used to 88 construct different dimensions of marine diversity. We consider geographic occurrences and 89 nucleotide sequences to be the fundamental building blocks of the spatial and phylogenetic 90 dimensions of diversity, which interact to structure the distribution of key ecological traits 91 across species [38]. A first step to adding the functional dimension of diversity is to classify 92 species into broad ecological guilds, similar to the way in which species can be classified in 93 global theories and models of biodiversity [39,40]. Supplementing these with information on 94 conservation status and molecular taxonomy provides insights into how marine diversity is 95 changing, and how we might efficiently monitor this. Throughout we use open source 96 computational tools to link data across these components of marine diversity to take stock of 97 the current state of data availability, identifying gaps and priorities for future work. In this way 98 we summarise data availability across multiple axes for >200,000 marine animal species 99 from 32 phyla and across broad ecological guilds (e.g. benthos, zooplankton, seabirds), and 100 we assess the extent to which this availability is correlated across different classes of 101 diversity data. Above all, our aim is to highlight the wealth of marine biodiversity data that we 102 have amassed as a community over centuries, and the opportunities that we now have to 103 link different classes of data in order to better understand the dimensions of marine diversity.

104 Methods

To provide an overview of the state of knowledge of marine animal biodiversity, we mine the
World Register of Marine Species (WoRMS, [37]), the most comprehensive source of

107 taxonomic information on marine species, consisting of over half a million distinct names 108 checked by expert taxonomic editors. We focus our investigation on marine animals, and so 109 filtered the WoRMS database to Kingdom Animalia, retaining only those species considered 110 to be marine by WoRMS (flag isMarine is TRUE), and excluding any species only known 111 from fossils. We consider only taxa identified at the species rank, with a current accepted 112 name and valid WoRMS identifier (Aphia ID).

113

114 In addition to taxonomy, WoRMS has aggregated data on species attributes including broad 115 'functional groups'. In reality these are closer to ecological guilds, defining habitat affinity 116 (e.g. benthos, zooplankton) rather than ecological function, but for transparency we retain 117 the terminology employed by WoRMS. We use these attributes to assign each species to a 118 functional group, using a dedicated R function (https://github.com/tomjwebb/WoRMS-119 functional-groups) which accesses the WoRMS API using the worrms R package [41]. We 120 supplement these functional groups with taxonomic groups to identify fish (using the 121 WoRMS paraphyletic Superclass Pisces; [42]), marine mammals, seabirds, and reptiles. We 122 consolidate functional groups into broad categories for maximum coverage - for example, 123 our 'benthos' group includes all species categorised in WoRMS as endobenthos, 124 epibenthos, hyperbenthos, macrobenthos, meiobenthos, and microbenthos, as well as those 125 originally classified simply as benthos. When separate functional groups are recorded for 126 different life stages, we always use the group for the adult stage. We group together 127 categories with very few species (including meso, macro, neuston) and species with no 128 functional group classification into the single category 'other/unknown'. For fish we include 129 an additional grouping variable based on the broad habitat categories recorded in FishBase 130 [10] accessed using the rfishbase package [43], classifying 17,568 of 18,261 species as 131 bathydemersal, bathypelagic, benthopelagic, demersal, pelagic-oceanic, pelagic-nertitic, or 132 reef-associated.

133

134 The WoRMS database includes links to other major biodiversity databases (table 1), and we 135 exploit these to compare the state of biodiversity information availability across axes of 136 biogeography, genetics, conservation, and molecular taxonomy. Specifically, we record for 137 each species its total number of occurrences in the Ocean Biogeographic Information 138 System (OBIS, [22]), and its total number of nucleotide sequences in GenBank. The 139 taxonomy in OBIS is standardised to WoRMS, making these links straightforward, and 140 GenBank's taxonomic information is generally reliable for marine animals [4] meaning that 141 links between WoRMS and GenBank are likely to robustly associate relevant sequences 142 with the correct taxonomic identifier. We also record for each species its IUCN conservation 143 assessment category (if available), and whether or not it has DNA barcodes listed in the 144 Barcode of Life Data System (BOLD).

145

146 Using our tidy database linking the diversity data sources shown in table 1, we then 147 summarise the availability of biodiversity data across all marine animals as follows. First, we 148 consider the two major quantitative databases, OBIS and GenBank. We calculate the 149 proportion of species within each phylum with records in each of these databases, and the 150 distribution of records between species within each phylum. To derive an indication of 151 relative data availability across functional groups, highlighting groups that are particularly 152 highly likely (or unlikely) to occur in the dataset, and those which tend to have more records 153 when they are present, we model data availability across functional groups. We apply a two-154 step hurdle process, because of the high degree of zero-inflation in our data [44]. To assess 155 whether certain functional groups were better represented in the databases than others, we 156 model presence of species in OBIS or GenBank using a binomial GLM of the form species 157 presence ~ functional group, and we model the distribution of counts (OBIS records or 158 GenBank nucleotides) between functional groups, for those species present in the data 159 source, using a zero-truncated negative binomial GLM. These hurdle models are 160 implemented using the hurdle function in the pscl package [44,45]. For visualisation, we 161 plot the exponentiated binomial coefficients from the zero component of the model, which

shows the ratio of the probability of getting a non-zero to a zero observation within a
functional group. We also plot the predicted counts for the subset of species in each
functional group with non-zero counts.

165

166 To assess whether data availability is correlated across data sources, we use categorical 167 scales of numbers of records per species in both OBIS and GenBank, using categories 168 bounded by upper limits of 0, 1, 10, 100, 1,000, 10,000, 100,000, and a final category of 169 >100,000 records. We use mosaic plots [46], created using the ggmosaic R package [47], 170 to illustrate the distribution of GenBank count categories for each OBIS count category. We 171 also consider how IUCN conservation assessments are distributed across species in 172 different functional groups, and between species present and absent in OBIS, and we 173 compare the number of OBIS occurrence records between species in different IUCN 174 categories. To simplify this analysis, we aggregate to the following IUCN assessment 175 categories: Not Assessed, Data Deficient (i.e., formally assessed but insufficient data to 176 assign the species to a threat category), Threatened (formally assessed as Vulnerable, Endangered, Critically Endangered, Conservation Dependent, Extinct in the Wild, or Extinct), 177 178 and Non-threatened (formally assessed as Near Threatened or Least Concern). We perform 179 a similar analysis comparing species presence or absence in the Barcode of Life database 180 with presence in OBIS and number of OBIS records.

181

All data and links were extracted from WoRMS on 2020-01-11 and the statistics we report are correct as of that date. Manipulation, visualisation, and analysis is performed in R 3.6.2 [48] using RStudio 1.2.5033 [49] and the tidyverse suite of packages [50] as well as worrms [41] to access the WoRMS API and rfishbase {Boettiger:2012bz} to access FishBase, and the plotting packages ggmosaic [47], ggbeeswarm [51] and patchwork [52]. All data used in this article are publicly available via WoRMS. The processed summary data we use for our analysis is openly available under a Creative Commons

- 189 Attribution 4.0 International License in the Marine Data Archive
- 190 (https://doi.org/10.14284/417). R code to replicate our analyses and figures is available
- 191 via https://github.com/tomjwebb/linking_marine_diversity_data and is archived on Figshare
- 192 via the University of Sheffield's Online Research Data repository
- 193 here: https://doi.org/10.15131/shef.data.12833891.
- 194 Results

195 Our final dataset consisted of 206,849 valid marine animal species, from 32 phyla and 89 196 classes. Of these, 106,213 (51%) have at least one occurrence record listed in OBIS (table 197 2). Of these, 18,869 (18% of species in OBIS, 9% of all species) are represented by just a 198 single occurrence record (table 2), while one species (Atlantic Cod, Gadus morhua) has over 199 a million occurrence records (1,108,463). Overall, there are 45,974,726 OBIS occurrence 200 records across all species. 36,094 (17%) of all species have at least one nucleotide 201 recorded in GenBank, while 8 species (five fish, the Antarctic Minke Whale Balaenoptera 202 bonaerensis, the tunicate Ciona intestinalis and the California Sea Hare Aplysia californica) 203 have more than a million. Overall the species in our database total 56,846,294 GenBank 204 nucleotides. Furthermore, 13,179 species have had their conservation status assessed by 205 the IUCN, and 25,272 have at least one DNA barcode in the Barcode of Life database.

206

207 The distribution of OBIS and GenBank records across animal phyla and functional groups is 208 shown in Fig 1. At least one species from every phylum has records in either OBIS or 209 GenBank, with all phyla except Loricifera (which has just 29 species) represented in both 210 databases (Fig 1A). Across all phyla, just over half (55%) of all species are represented in 211 one or other database. Most species that are present in OBIS have only a few occurrence 212 records, with median values of records ranging from 1 to 92 across phyla (Fig 1B). A similar 213 pattern is observed for GenBank nucleotides (fig 1C), with median values between 1 and 94 214 except in phyla Orthonectida and Placozoa, both of which have only two species

215 represented in GenBank, one of which has several thousand nucleotides (in Orthonectida,

216 Intoshia linei has 3,522, in Placozoa, Trichoplax adhaerens has 29,176).

217

218 Data availability is variable across functional groups (fig 1B, C; fig 2). Modelling the presence 219 or absence of species in OBIS in a binomial GLM shows that species of fish, mammal, bird, 220 and reptile are much more likely to have occurrences in OBIS than are benthic or 221 zooplankton species, with nekton falling in between, and species with unknown or other 222 functional group classification the least likely to have occurrence records (fig 2A). A broadly 223 similar pattern holds when modelling the number of occurrence records for those species 224 with at least 1 (fig 2B), with the vertebrate taxa again tending to have most records, although 225 distinctions between vertebrates and other groups are less stark. Benthic invertebrates 226 typically have few OBIS records, but zooplankton that do occur in OBIS tend to have more 227 records than nekton. In GenBank, birds, reptiles and mammals are most likely to be present 228 in the database, followed by fish, nekton, and zooplankton, with benthos and other/unknown 229 functional groups least likely to be represented (fig 2C). The rank order changes somewhat 230 when considering number of nucleotides across species present in GenBank (fig 2D), with 231 most records from mammals and reptiles, followed but birds and fish. Nekton tend to have 232 fewest records, but there is considerable variability within all major groups. Data availability 233 in both major databases is broadly similar across fish habitat groupings (figure S1, S2).

234

Considering the joint distribution of species across OBIS and GenBank categorical scales,
93,519 (45%) species have no records in either database (table 2, fig 3A). In general,
species with more records in OBIS also tend to have more nucleotides in GenBank (table 2,
fig 3), indicating that these different biodiversity data aggregators have similar biases in
terms of the known marine biodiversity that they encompass. There are exceptions though:
in particular several species have many (>100,000) GenBank nucleotides but very few (if
any) OBIS records (table 3).

243 A similar pattern is evident when examining the distribution of OBIS records across different 244 IUCN assessment categories. In general, and across functional groups, the proportion of 245 species with records in OBIS is higher in assessed species (threatened and non-threatened) 246 than it is in unassessed or data-deficient species: overall, 84% of threatened and 94% of 247 non-threatened species have occurrence records in OBIS, compared to 75% of data-248 deficient and 49% of unassessed species (table 4A). Considering only those species with 249 records in OBIS, there is considerable variation within and between IUCN categories in the 250 number of occurrence records per species, but a general tendency is apparent in all 251 functional groups for species in threatened and non-threatened categories to have more 252 occurrence records than those in data-deficient and unassessed categories (fig 4A).

253

254 Species with DNA barcodes are disproportionately likely to also have occurrence records in 255 OBIS: 45% of species with no record in the Barcode of Life database have at least one 256 occurrence record in OBIS, compared to 89% of species with a barcode (table 4B). In 257 addition, in all functional groups, species with barcodes tend to have more OBIS records 258 than those which do not (fig 4B).

259 Discussion

260 Using the taxonomic backbone of the World Register of Marine Species [37] we have 261 summarised data availability across axes of biogeography, genetics, molecular taxonomy, 262 and conservation status for 206.849 marine animal species. This presents a mixed picture. 263 One the one hand, 91,828 (44%) species have no records in any of these databases, and 264 are represented only by their name. This is considerably higher than the 27% of plant 265 species with no information other than their name [36], although of course the marine 266 environment represents far larger habitable volume [53] and marine animals are a much 267 more diverse taxonomic group. Only 6,688 marine animal species (3%) have records in all 268 four of the datasets that we consider - again, rather lower than the 18% of broadly-covered 269 plant species [36]. At the same time, it is important to remember that presence in a dataset

270 does not imply extensive knowledge: among the 106,203 species with records in OBIS, for 271 example, the median number of recorded occurrences is just 7, and 18% of these species 272 (18,869 species) are known from only a single occurrence. Nonetheless, the distribution of 273 biogeographic and genetic information across the animal tree of life is extensive, with all 274 animal phyla represented in at least one database (fig 1). Data availability tends to be biased 275 towards well-known taxa and functional groups (especially vertebrates; figs 1, 2, 4), in 276 agreement with previous assessments (e.g. [32]), but the subset of 225 species with >1,000 277 occurrences in OBIS and >1,000 nucleotides in GenBank is drawn from 10 phyla and 27 278 classes, representing all major functional groups, and most of them have a barcode in BOLD 279 (214 species), and have been assessed by the IUCN as something other than data deficient 280 (102 non-threatened, 23 threatened species). For these diverse marine animal species then, 281 it is reasonable to propose that the information available across multiple sources can be 282 translated into knowledge about their distribution, evolutionary relationships, and 283 conservation status.

284

285 The broad positive correlation between data availability across different sources (table 2, 286 table 4, fig 3) reinforces previous findings that species with good information on one facet of 287 their biology and ecology tend to be well represented in other databases too, both in plants 288 [36] and in marine species [35]. These information-rich species are likely to be those most 289 easily and frequently observed, or those of high economic or cultural value, and so will not 290 be a random subset of all species. However, the consequences of biases towards data 291 availability from these common species will vary depending on the specific question of 292 interest. For instance, ecosystem function may be driven largely by just those common 293 species that tend to be so well known [54]; but rare species will clearly be of great interest to 294 conservationists, and may indeed sometimes contribute unique trait combinations to marine 295 communities [55].

296

297 In terrestrial conservation, considerable concern has been expressed over the likely 298 conservation status of species too poorly known to formally assess, as they tend to have 299 characteristics (rarity, small ranges, occurring in poorly studied regions) which will 300 predispose them to be at risk [56]. For some marine taxa this appears to be the case too. 301 with high rates of extinction risk predicted for European sharks and rays formally assessed 302 as Data Deficient [57], and low levels of conservation assessment in poorly-known marine 303 groups may contribute to low overall documented levels of extinction risk [58]. On the other 304 hand, the fact that the biggest data gaps in marine biodiversity tend to be in remote habitats 305 largely inaccessible to humans (e.g. the deep pelagic ocean; [59]), and the highest rates of 306 discoveries of new species and habitats are also in the deep sea [60,61], provides some 307 contrast with the terrestrial situation, and may insulate these poorly-known species 308 somewhat from human pressures. However, some patterns still hold in the deep sea, such 309 as the tendency for widespread species to be encountered and described first [62], meaning 310 that many of the species not yet present in major databases may be genuinely rare. Given 311 the acceleration of human activities into previously unexploited regions of the oceans [63]. 312 with new threats including deep sea mining [64] and exploitation of the mesopelagic [65], it 313 seems unwise to assume that the large fraction of marine biodiversity that remains poorly 314 known is not at risk. Given the fact that Data Deficient conservation assessments are twice 315 as frequent in marine versus non-marine taxa [34], data-driven predictive conservation 316 assessments [57,66,67] which rely on some of the kinds of data we consider here (spatial 317 distribution, evolutionary relationships, ecological guilds) combined with biological traits may 318 prove to be especially valuable tools.

319

An aim of this study was to flag priorities for future work. One important point is that the major publicly available databases on which we draw do not constitute the sum total of data ever collected on marine species. This is particularly the case for occurrence data, as globally researchers have yet to adopt the routine deposition of species occurrences in OBIS as a cultural norm, in the way that genetic sequence data is deposited in GenBank. To this

325 end, improving incentives for researchers to add their data to global repositories in an 326 important goal [25], while data archaeology and rescue initiatives can help to ensure that 327 historical data are captured [68]. Equally, it remains vital that ongoing survey schemes are 328 properly valued [69], even as novel exploration is planned. At the same time, our 329 quantification exercise can help to identify groups of species where a little additional 330 research effort in one area would quickly result in a more valuable dataset. One candidate 331 set of species might be those that are frequently observed but poorly represented in other 332 databases. For instance, 1,216 species have >1,000 OBIS records but <10 GenBank 333 nucleotides; and over half of the 3,533 species with >1,000 OBIS occurrences are either not 334 assessed by the IUCN (1,876 species) or data-deficient (82 species). The fact that almost 335 90% (3,163) of these species have DNA barcodes in BOLD is encouraging, however, 336 suggesting considerable potential for an increasing role for molecular studies to address a 337 wide range of questions in marine ecology [70].

338

339 Mining the spatial information already present in other databases also has potential for 340 supplementing existing occurrence datasets. In this study we relied on existing links between 341 WoRMS and GenBank and BOLD, which simply summarise the number of nucleotides or 342 barcodes present for each species. The spatial meta-data stored in the sequence databases 343 provides an additional source of information, although in GenBank this data is relatively 344 unstructured. Searching the GenBank nucleotide database, we found just 1,437 records for animals which contained a lat-lon field; matching this to our list of marine animals reduced 345 346 this further to 183 records from 42 species. Nonetheless, even from this small set of species, 347 21 do not have occurrence records in OBIS, suggesting that mining GenBank for spatial data 348 would likely add valuable information for a small number of species. Various methods have 349 been developed to attempt this, based around mining spatial information from the full text of 350 associated publications [71,72] with initiatives such as the Genomic Observatories 351 MetaDatabase (GEOME, https://geome-db.org) also seeking to simplify access to meta-data 352 from sequence datasets.

353

354 BOLD typically does store spatial data for individual specimens in a well-structured manner, 355 only some of which has been harvested by OBIS. In our dataset, 3,117 species have BOLD 356 barcodes but no OBIS records. Several of these are parasites, which we know are not well 357 recorded in OBIS (e.g. Schistocephalus solidus, 718 barcodes; Anguillicoloides crassus, 508 358 barcodes) but there are free-living marine species too, such as the Gastropod mollusc 359 Littoraria sinensis (257 barcodes) and the Copepod Calanoides natalis (183 barcodes). 360 Accessing the specimen data from BOLD using the bold R package [73] for these two 361 species reveals that none of the L. sinensis have information in the latitude and longitude 362 fields, but full geographic information is available for 227 specimens for Calanoides natalis. 363 Although none of these locations are currently recorded in OBIS, some are in GBIF, 364 highlighting the often complex pipelines from data providers to global data aggregators. 365 Improving pipelines from genetic databases to occurrence databases is currently a priority 366 for OBIS (W. Appeltans, OBIS Project Manager, pers. comm.).

367

368 Finally, the dimensions of diversity that we summarise in this study are somewhat limited. 369 We did not consider the traits of species, for instance, beyond functional groups that indicate 370 habitat affiliation in very broad terms (e.g. benthic vs planktonic). These groupings are 371 already useful as global patterns of diversity are known to differ between them [39], and they 372 can also be used to refine methods of matching species occurrences to global sea 373 temperature datasets [74], helping to predict species responses to climate change [75]. 374 Beyond these coarse functional groups, however, traits data remain scarce even in 375 reasonably common marine species in well-studied regions [35], and despite many efforts at 376 collating traits - including within WoRMS; [76] - there is still no widely-adopted central 377 standard [77]. Certain groups are well covered by existing initatives (e.g. FishBase [10], the 378 Coral Trait Database [11]), and whether a single overarching portal to cover the immense 379 diversity of marine lifeforms is possible - or even desirable - remains open for discussion. 380 However, it is certainly the case that multiple smaller-scale projects collect valuable traits

381 data for a subset of species which is typically made available (if at all) via supplementary 382 material or bespoke web portals, at risk of being lost to the community. A wider adoption of 383 principles embedded in initiatives like the Open Traits Network [7] would ensure 384 interoperability of these small, project-specific traits datasets, maximising the availability of 385 information on key traits for the largest possible fraction of marine diversity. Readily 386 availabilie information on even just a few traits (e.g. body size, longevity, fecundity, 387 planktonic larval duration) would help to test predictions from biodiversity models, embed life 388 history theory into marine conservation, and predict the consequences of human activities on 389 marine diversity [39,78-80].

390

391 The stocktake of marine biodiversity data availability that we have undertaken here adds to 392 previous efforts focused on occurrence data [19,32,81]. While we reveal a similar story of 393 gaps and biases across other data sources, there is considerable overlap in coverage too, 394 and overall the potential to link dimensions of marine animal diversity is now high. The 395 priority now should be to build on the substantial community-built foundations and to improve 396 the pipeline from raw data to interoperable data products, both as a resource for 397 fundamental macroecological research and to facilitate effective stewardship of our blue 398 planet.

399 Acknowledgements

400 The ideas for this study were conceived while TJW was a Royal Society University Research 401 Fellow, and developed over the course of the Natural Environment Research Council and 402 Department for Environment, Food and Rural Affairs Marine Ecosystems Research 403 Programme (grant number NE/L003279/1) and EMODnet Biology. Thanks to Helmut 404 Hillebrand for the invitation to present this work at the 2019 HIFMB Symposium on 405 Functional Marine Biodiversity and for the opportunity to contribute to this volume. 406 Constructive and insightful comments from two anonymous reviewers have helped to greatly 407 improve this work.

409 1. 1. Redelings, B. D. & Holder, M. T. 2017 A supertree pipeline for summarizing 410 phylogenetic and taxonomic information for millions of species. PeerJ 5, e3058. 411 (doi:10.7717/peerj.3058) 412 2. Hinchliff, C. E. et al. 2015 Synthesis of phylogeny and taxonomy into a comprehensive tree of life. P Natl Acad Sci USA 112. 12764-12769. 413 (doi:10.1073/pnas.1423041112) 414 415 Clark, K., Karsch-Mizrachi, I., Lipman, D. J., Ostell, J. & Sayers, E. W. 2016 3. GenBank. Nucleic Acids Res. 44, D67–D72. (doi:10.1093/nar/gkv1276) 416 417 4. Leray, M., Knowlton, N., Ho, S.-L., Nguyen, B. N. & Machida, R. J. 2019 GenBank is a reliable resource for 21st century biodiversity research. P Natl 418 419 Acad Sci USA 116, 22651–22656. (doi:10.1073/pnas.1911714116) 420 GBIF 2020 GBIF Home Page. https://www.gbif.org. 5. 421 IUCN 2020 The IUCN Red List of Threatened Species. Version 2020-1. 6. 422 httpswww.iucnredlist.org. 423 Gallagher, R. et al. 2019 The Open Traits Network: Using Open Science 7. 424 principles to accelerate trait-based science across the Tree of Life. 425 ecoevorxiv.org. (doi:https://doi.org/10.32942/osf.io/kac45) 426 8. Bennett, J. M. et al. 2018 GlobTherm, a global database on thermal tolerances 427 for aquatic and terrestrial organisms. Sci. Data 5, 180022. 428 (doi:10.1038/sdata.2018.22) 429 9. Makarieva, A., Gorshkov, V. & LI, B. 2005 Biochemical universality of living 430 matter and its metabolic implications. Funct Ecol 431 10. Froese, R. & Pauly, D. 2019 FishBase. World Wide Web electronic publication. 432 version (12/2019). www.fishbase.org. 433 11. Madin, J. S. et al. 2016 The Coral Trait Database, a curated database of trait information for coral species from the global oceans. Sci. Data 3, 178-22. 434 435 (doi:10.1038/sdata.2016.17) 436 Kranstauber, B., Cameron, A., Weinzerl, R., Fountain, T., Tilak, S., Wikelski, M. 12. & Kays, R. 2011 The Movebank data model for animal tracking. Environmental 437 Modelling & Software 26, 834-835. (doi:10.1016/j.envsoft.2010.12.005) 438 439 13. Wikelski, M., Davidson, S. C. & Kays, R. 2020 Movebank: archive, analysis and 440 sharing of animal movement data. www.movebank.org. 441 Poelen, J. H., Simons, J. D. & Mungall, C. J. 2014 Global biotic interactions: An 14. open infrastructure to share and analyze species-interaction datasets. 442 443 Ecological Informatics 24, 148–159. (doi:10.1016/j.ecoinf.2014.08.005) 444 15. Tanhua, T. et al. 2019 Ocean FAIR Data Services. Front. Mar. Sci. 6, 92. (doi:10.3389/fmars.2019.00440) 445

448 Chandler, M. et al. 2017 Contribution of citizen science towards international 17. biodiversity monitoring. Biological Conservation 213, 280-294. 449 (doi:10.1016/j.biocon.2016.09.004) 450 451 18. Hyder, K., Townhill, B., Anderson, L. G., Delany, J. & Pinnegar, J. K. 2015 Can 452 citizen science contribute to the evidence-base that underpins marine policy? 453 Marine Policy 59, 112-120. (doi:10.1016/j.marpol.2015.04.022) 454 Edgar, G. J., Bates, A. E., Bird, T. J., Jones, A. H., Kininmonth, S., Stuart-Smith, 19. 455 R. D. & Webb, T. J. 2015 New Approaches to Marine Conservation Through Scaling Up of Ecological Data. Annual Review of Marine Science 8, 456 150807173619006. (doi:10.1146/annurev-marine-122414-033921) 457 458 20. Edgar, G. J. & Stuart-Smith, R. D. 2014 Systematic global assessment of reef fish communities by the Reef Life Survey program. Sci. Data 1, 1-8. 459 460 (doi:10.1038/sdata.2014.7) 461 21. Snelgrove, P. V. R. 2010 Discoveries of the Census of Marine Life: Making 462 Ocean Life Count. 1st edn. Cambridge University Press. 463 22. OBIS 2020 Ocean Biodiversity Information System. www.iobis.org. 464 23. Mieszkowska, N., Sugden, H., Firth, L. B. & Hawkins, S. J. 2014 The role of sustained observations in tracking impacts of environmental change on marine 465 466 biodiversity and ecosystems. Philos T R Soc A 372, 20130339–20130339. (doi:10.1098/rsta.2013.0339) 467 468 Miloslavich, P. et al. 2018 Essential ocean variables for global sustained 24. observations of biodiversity and ecosystem changes. Global Change Biol 105, 469 470 10456. (doi:10.1111/gcb.14108) 471 25. Dornelas, M. et al. 2019 Towards a macroscope: Leveraging technology to 472 transform the breadth, scale and resolution of macroecological data. Global Ecol 473 Biogeogr 28, 1937–1948. (doi:10.1111/geb.13025) 474 26. Basset, A. & Los, W. 2012 Biodiversity e-Science: LifeWatch, the European infrastructure on biodiversity and ecosystem research. Plant Biosystems - An 475 476 International Journal Dealing with all Aspects of Plant Biology 146, 780–782. (doi:10.1080/11263504.2012.740091) 477 478 27. La Salle, J., Williams, K. J. & Moritz, C. 2016 Biodiversity analysis in the digital era. Philos T R Soc B **371**. (doi:10.1098/rstb.2015.0337) 479 480 Hardisty, A. R. et al. 2019 The Bari Manifesto: An interoperability framework for 28. 481 essential biodiversity variables. Ecological Informatics 49, 22-31. 482 (doi:10.1016/j.ecoinf.2018.11.003) 483 29. Maitner, B. S. et al. 2018 The bien r package: A tool to access the Botanical 484 Information and Ecology Network (BIEN) database. Methods in Ecology and Evolution 9, 373-379. (doi:10.1111/2041-210X.12861) 485

Silvertown, J. 2009 A new dawn for citizen science. Trends Ecol Evol 24, 467-

471. (doi:10.1016/j.tree.2009.03.017)

446

447

16.

486 30. Costello, M., Coll, M., Danovaro, R., Halpin, P. & Ojaveer, H. 2010 A Census of Marine Biodiversity Knowledge, Resources, and Future Challenges. PLoS ONE 487 488 Snelgrove, P. et al. 2016 Global Patterns in Marine Biodiversity. In The First 31. 489 Global Integrated Marine Assessment World Ocean Assessment, un.org. 490 Miloslavich, P. et al. 2016 Extent of Assessment of Marine Biological Diversity. 32. 491 In The First Global Integrated Marine Assessment World Ocean Assessment 492 (eds L. Inniss & A. Simcock), United Nations. 493 33. Appeltans, W. et al. 2012 The magnitude of global marine species diversity. 494 Curr. Biol. 22, 2189–2202. (doi:10.1016/j.cub.2012.09.036) 495 Webb, T. J. & Mindel, B. L. 2015 Global Patterns of Extinction Risk in Marine 34. 496 and Non-marine Systems. Current Biology 25, 506-511. 497 (doi:10.1016/j.cub.2014.12.023) 498 Tyler, E. H. M., Somerfield, P. J., Berghe, E. V., Bremner, J., Jackson, E., 35. Langmead, O., Palomares, M. L. D. & Webb, T. J. 2012 Extensive gaps and 499 500 biases in our knowledge of a well-known fauna: implications for integrating 501 biological traits into macroecology. Global Ecol Biogeogr 21, 922-934. (doi:10.1111/j.1466-8238.2011.00726.x) 502 503 36. Cornwell, W. K., Pearse, W. D., Dalrymple, R. L. & Zanne, A. E. 2019 What we 504 (don't) know about global plant diversity. Ecography, ecog.04481. 505 (doi:10.1111/ecog.04481) 506 37. WoRMS Editorial Board 2020 World Register of Marine Species. 507 http://www.marinespecies.org. 508 Freckleton, R. P. & Jetz, W. 2009 Space versus phylogeny: disentangling 38. phylogenetic and spatial signals in comparative data. P R Soc B 276, 21-30. 509 (doi:10.1098/rspb.2008.0905) 510 Worm, B. & Tittensor, D. P. 2018 A Theory of Global Biodiversity. Princeton and 511 39. 512 Oxford: Princeton University Press. (doi:10.2307/j.ctt1zkjz6q) 513 40. Harfoot, M. B. J., Newbold, T., Tittensor, D. P., Emmott, S., Hutton, J., 514 Lvutsarev, V., Smith, M. J., Scharlemann, J. P. W. & Purves, D. W. 2014 Emergent Global Patterns of Ecosystem Structure and Function from a 515 Mechanistic General Ecosystem Model. PLoS Biol 12, e1001841. 516 (doi:10.1371/journal.pbio.1001841) 517 Chamberlain, S. 2019 World Register of Marine Species (WoRMS) Client. R 518 41. package worrms version 0.4.0. httpsCRAN.R-project.orgpackageworrms. 519 520 42. WoRMS 2020 Pisces. https://www.marinespecies.orgaphia.phpptaxdetailsid. 521 43. Boettiger, C., Lang, D. T. & Wainwright, P. C. 2012 rfishbase: exploring, manipulating and visualizing FishBase data from R. Journal of Fish Biology 81, 522 2030–2039. (doi:10.1111/j.1095-8649.2012.03464.x) 523 524 44. Zeileis, A., Kleiber, C. & Jackman, S. 2008 Regression models for count data in 525 R. Journal of Statistical Software 27, 1–25.

Jackman, S. 2020 pscl: Classes and Methods for R Developed in the Political 526 45. Science Computational Laboratory. United States Studies Centre, University of 527 Sydney, Sydney, New South Wales, Australia. R package version 1.5.5. 528 529 httpsgithub.comatahkpscl. 530 Hofmann, H. 2008 Mosaic Plots and Their Variants. In Handbook of Data 46. 531 Visualisation (eds C.-H. Chen W. Härdle & A. Unwin), Berlin Heidelberg. Jeppson, H., Hofmann, H. & Cook, D. 2018 ggmosaic: Mosaic Plots in the 532 47. 533 ggplot2' Framework. R package version 0.2.0. https://CRAN.Rproject.orgpackageggmosaic. 534 535 48. R Core Team 2019 R: A language and environment for statistical computing. 536 https://www.R-project.org. 537 49. RStudio Team 2019 RStudio: Integrated Development for R. 538 httpwww.rstudio.com. Wickham, H. et al. 2019 Welcome to the Tidyverse. Journal of Open Source 539 50. 540 Software 4, 1686. (doi:10.21105/joss.01686) Clarke, E. & Sherrill-Mix, S. 2017 ggbeeswarm: Categorical Scatter (Violin 541 51. Point) Plots. R package version 0.6.0. httpsCRAN.R-542 543 project.orgpackageggbeeswarm. 544 52. Pedersen, T. L. 2019 patchwork: The Composer of Plots, R package version 1.0.0. httpsCRAN.R-project.orgpackagepatchwork. 545 546 Dawson, M. N. 2012 Species richness, habitable volume, and species densities 53. in freshwater, the sea, and on land. Frontiers of Biogeography 4, fb 12675. 547 548 54. Gaston, K. & Fuller, R. 2008 Commonness, population depletion and conservation biology. Trends Ecol Evol 23, 14-19. 549 550 55. Mouillot, D. et al. 2013 Rare Species Support Vulnerable Functions in High-551 Diversity Ecosystems. PLoS Biol 11, e1001569. 552 (doi:10.1371/journal.pbio.1001569) 553 56. Scheffers, B. R., Joppa, L. N., Pimm, S. L. & Laurance, W. F. 2012 What we know and don't know about Earth's missing biodiversity. Trends Ecol Evol 27, 554 555 501-510. (doi:10.1016/j.tree.2012.05.008) Walls, R. H. L. & Dulvy, N. K. 2019 Predicting the conservation status of 556 57. 557 Europe's Data Deficient sharks and rays. bioRxiv 276, 614776. 558 (doi:10.1101/614776) 559 58. Mindel, B. L., Webb, T. J., Neat, F. C. & Blanchard, J. L. 2016 A trait-based metric sheds new light on the nature of the body size-depth relationship in the 560 deep sea. J Anim Ecol 85, 427-436. (doi:10.1111/1365-2656.12471) 561 59. 562 Webb, T. J., Vanden Berghe, E. & O'Dor, R. 2010 Biodiversity's big wet secret: 563 the global distribution of marine biological records reveals chronic under-564 exploration of the deep pelagic ocean. PLoS ONE 5, e10223. (doi:10.1371/journal.pone.0010223) 565

- 56660.Ramirez-Llodra, E. et al. 2010 Deep, diverse and definitely different: unique567attributes of the world's largest ecosystem. Biogeosciences 7, 2851–2899.568(doi:10.5194/bg-7-2851-2010)
- 56961.Danovaro, R., Snelgrove, P. V. R. & Tyler, P. 2014 Challenging the paradigms570of deep-sea ecology. Trends Ecol Evol 29, 465–475.571(doi:10.1016/j.tree.2014.06.002)
- 572 62. Higgs, N. D. & Attrill, M. 2015 Biases in biodiversity: wide-ranging species are
 573 discovered first in the deep sea. Front. Mar. Sci. 2, 717.
 574 (doi:10.3389/fmars.2015.00061)
- 57563.Jouffray, J. B., Blasiak, R., Norström, A. V., Österblom, H. & Nyström, M. 2020576The Blue Acceleration: The Trajectory of Human Expansion into the Ocean. One577Earth 2, 43–54. (doi:10.1016/j.oneear.2019.12.016)
- 57864.Jones, D. O. B., Amon, D. J. & Chapman, A. S. A. 2018 Mining Deep-Ocean579Mineral Deposits: What are the Ecological Risks? Elements 14, 325–330.580(doi:10.2138/gselements.14.5.325)
- 58165.Hidalgo, M. & Browman, H. I. 2019 Developing the knowledge base needed to582sustainably manage mesopelagic resources. ICES Journal of Marine Science583**76**, 609–615. (doi:10.1093/icesjms/fsz067)
- 58466.Jetz, W. & Freckleton, R. P. 2015 Towards a general framework for predicting585threat status of data-deficient species from phylogenetic, spatial and586environmental information. Philos T Roy Soc B **370**, 20140016–20140016.587(doi:10.1098/rstb.2014.0016)
- 58867.González-del-Pliego, P., Freckleton, R. P., Edwards, D. P., Koo, M. S.,589Scheffers, B. R., Pyron, R. A. & Jetz, W. 2019 Phylogenetic and Trait-Based590Prediction of Extinction Risk for Data-Deficient Amphibians. Current Biology 29,5911557–1563.e3. (doi:10.1016/j.cub.2019.04.005)
- 59268.Faulwetter, S. et al. 2016 EMODnet Workshop on mechanisms and guidelines593to mobilise historical data into biogeographic databases. RIO 2, e9774–28.594(doi:10.3897/rio.2.e9774)
- 59569.Mieszkowska, N., Sugden, H., Firth, L. B. & Hawkins, S. J. 2014 The role of596sustained observations in tracking impacts of environmental change on marine597biodiversity and ecosystems. Philos T R Soc A **372**, 20130339.598(doi:10.1098/rsta.2013.0339)
- 59970.Goodwin, K. D., Thompson, L. R., Duarte, B., Kahlke, T., Thompson, A. R.,600Marques, J. C. & Caçador, I. 2017 DNA Sequencing as a Tool to Monitor Marine601Ecological Status. Front. Mar. Sci. 4, e1002358.602(doi:10.3389/fmars.2017.00107)
- 60371.Tahsin, T., Weissenbacher, D., Rivera, R., Beard, R., Firago, M., Wallstrom, G.,604Scotch, M. & Gonzalez, G. 2016 A high-precision rule-based extraction system605for expanding geospatial metadata in GenBank records. J Am Med Inform606Assoc 23, 934–941. (doi:10.1093/jamia/ocv172)
- 60772.Tahsin, T., Weissenbacher, D., O'Connor, K., Magge, A., Scotch, M. &608Gonzalez-Hernandez, G. 2017 GeoBoost: accelerating research involving the

- 609 geospatial metadata of virus GenBank records. Bioinformatics 34, 1606–1608. (doi:10.1093/bioinformatics/btx799) 610 611 Chamberlain, S. In press. bold: Interface to Bold Systems API. R package 73. version 1.1.0. httpsCRAN.R-project.orgpackagebold. 612 613 74. Webb, T. J., Lines, A. & Howarth, L. M. 2020 Occupancy-derived thermal 614 affinities reflect known physiological thermal limits of marine species. Ecology and Evolution 75, 209. (doi:10.1002/ece3.6407) 615 75. Pinsky, M. L., Selden, R. L. & Kitchel, Z. J. 2020 Climate-Driven Shifts in Marine 616 Species Ranges: Scaling from Organisms to Communities. Annual Review of 617 618 Marine Science 12, 153–179. (doi:10.1146/annurev-marine-010419-010916) Costello, M. J., Claus, S., Dekeyzer, S., Vandepitte, L., Tuama, É. Ó., Lear, D. & 619 76. 620 Tyler-Walters, H. 2015 Biological and ecological traits of marine species. PeerJ 621 3, e1201. (doi:10.7717/peerj.1201) 622 77. Beauchard, O., Veríssimo, H., Queirós, A. M. & Herman, P. M. J. 2017 The use 623 of multiple biological traits in marine community ecology and its potential in 624 ecological indicator development. Ecol Indic 76, 81-96. 625 (doi:10.1016/j.ecolind.2017.01.011) 626 78. Kindsvater, H. K., Mangel, M., Reynolds, J. D. & Dulvy, N. K. 2016 Ten 627 principles from evolutionary ecology essential for effective marine conservation. Ecology and Evolution 6, 2125-2138. (doi:10.1002/ece3.2012) 628 629 79. Hiddink, J. G. et al. 2019 Assessing bottom trawling impacts based on the longevity of benthic invertebrates. J Appl Ecol 56, 1075–1084. 630 631 (doi:10.1111/1365-2664.13278) 632 Álvarez-Noriega, M., Burgess, S. C., Byers, J. E., Pringle, J. M., Wares, J. P. & 80. Marshall, D. J. 2020 Global biogeography of marine dispersal potential. Nature 633 634 Ecology & Evolution 2017 1:9 4, 1–8. (doi:10.1038/s41559-020-1238-y) Menegotto, A. & Rangel, T. F. 2018 Mapping knowledge gaps in marine 635 81. 636 diversity reveals a latitudinal gradient of missing species richness. Nature Communications 9, 4713. (doi:10.1038/s41467-018-07217-7) 637 Ratnasingham, S. & Hebert, P. 2007 BOLD: The Barcode of Life Data System 638 82. 639 (http://www.barcodinglife.org). Mol. Ecol. Notes (doi:doi: 10.1111/j.1471-640 8286.2006.01678.x) 641 642
 - 643

Table 1. Data sources used to link different dimensions of diversity across all marine645 animals.

Dimension of diversity	Data source	Data type	Reference
Taxonomy WoRMS		Authoritative classification and catalogue of marine taxonomic names	[37]
Functional Groups	WoRMS	Classification of marine species into broad ecological groups	[37]
Biogeography	OBIS	Global database of marine species occurrence records	[22]
Genetics	GenBank	The NIH genetic sequence database, an annotated collection of all publicly available DNA sequences	[3]
Molecular taxonomy	BOLD	Barcode of Life Data System for DNA barcodes	[82]
Conservation status	IUCN Red List	The IUCN Red List of threatened species	[6]

Number of	Number of GenBank nucleotides										
OBIS	0	1	2-10	11-100	101-	1,001-	10,001-	100,001-	>1,000,000	Totals	In OBIS
records					1,000	10,000	100,000	1,000,000			
0	93,519	1,312	4,484	1,164	116	13	15	13	0	100,636	
1	16,905	356	1,253	314	33	3	3	2	0	18,869	
2-10	35,613	1,086	3,714	990	122	17	11	8	0	41,561	
11-100	19,998	1,392	5,931	2,733	351	32	30	26	2	30,495	
101-1,000	4,274	594	3,334	2,917	512	51	35	37	1	11,755	
1,001-	402	86	630	1,113	315	33	53	33	4	2,669	
10,000											106,213
10,001-	42	4	107	406	167	31	22	31	1	811	
100,000											
100,001-	2	0	0	14	20	5	3	8	0	52	
1,000,000											
>1,000,000	0	0	0	0	0	0	1	0	0	1	
Totals	170,755	4,830	19,453	9,651	1,636	185	173	158	8	206,849	
In GenBank	36,094										

Table 2. Breakdown of 206,849 marine animal species by number of global occurrence records in OBIS, and numbers of nucleotide sequences in GenBank.

Species	Phylum	Class	Functional	GenBank	OBIS
	,		Group	Nucleotides	Records
Olavius algarvensis	Annelida	Clitellata	benthos	173,609	0
Capitella teleta	Annelida	Polychaeta	benthos	208,794	1
Platynothrus peltifer	Arthropoda	Arachnida	other/unknown	106,099	0
Caligus rogercresseyi	Arthropoda	Hexanauplia	other/unknown	628,843	0
Proasellus racovitzai	Arthropoda	Malacostraca	benthos	127,716	0
Proasellus ibericus	Arthropoda	Malacostraca	benthos	150,798	0
Bragasellus molinai	Arthropoda	Malacostraca	benthos	209,419	0
Proasellus beticus	Arthropoda	Malacostraca	benthos	228,033	0
Seriola quinqueradiata	Chordata	Actinopterygii	fish	105,911	6
Theragra finnmarchica	Chordata	Actinopterygii	fish	130,916	0
Takifugu flavidus	Chordata	Actinopterygii	fish	138,301	0
Takifugu rubripes	Chordata	Actinopterygii	fish	466,790	5
Molgula tectiformis	Chordata	Ascidiacea	benthos	106,904	0
Halocynthia roretzi	Chordata	Ascidiacea	benthos	116,123	4
Pelecanus crispus	Chordata	Aves	birds	231,775	0
Balaenoptera acutorostrata	Chordata	Mammalia	mammals	238,976	0
scammoni					
Emydocephalus ijimae	Chordata	Reptilia	reptiles	157,876	0
Hemicentrotus pulcherrimus	Echinodermata	Echinoidea	benthos	153,541	3
Apostichopus parvimensis	Echinodermata	Holothuroidea	benthos	166,764	1
Apostichopus japonicus	Echinodermata	Holothuroidea	benthos	401,310	4
Cumia reticulata	Mollusca	Gastropoda	benthos	144,517	2
Amphimedon queenslandica	Porifera	Demospongiae	benthos	142,554	9
Thunnus alalunga	Chordata	Actinopterygii	fish	0	114,485
Chrysophrys auratus	Chordata	Actinopterygii	fish	0	104,066

Table 3. Species with high numbers of GenBank nucleotide records but few OBIS occurrences, or species with large numbers of OBIS occurrences but few GenBank nucleotides.

		IUCN Assessment Status											
	N	ot assesse	d	Da	ata Deficier	ıt	Т	Threatened			Non-threatened		
Functional	N(species	N(specie	%	N(species	N(specie	%	N(species	N(specie	%	N(species	N(specie	%	
Group)	s in	species)	s in	species)	s in	species)	s in	specie	
*		OBIS)	in OBIS		OBIS)	in OBIS		OBIS)	in OBIS		OBIS)	s in	
											1.0.0.6	ORI2	
Benthos	144,097	73,610	51%	749	530	71%	305	258	85%	1,400	1,206	86%	
Zooplankto	5,742	3,027	53%	0	0	-	4	2	50%	2	2	100%	
n													
Nekton	3,076	1,878	61%	160	127	79%	7	2	29%	156	151	97%	
Fish	8,599	6,161	72%	1,780	1,350	76%	523	457	87%	7,359	6,997	95%	
Mammals	66	26	39%	20	17	85%	36	29	81%	70	68	97%	
Birds	179	71	40%	1	0	0%	125	92	74%	382	340	89%	
Reptiles	20	9	45%	21	14	67%	11	11	100%	44	37	84%	
Other /	31,891	9,725	31%	3	3	100%	10	4	40%	11	9	82%	
Unknown													
Totals	193,670	94,507	49%	2,734	2,041	75%	1,021	855	84%	9,424	8,810	94%	

Table 4A Breakdown of marine animal species by functional group and IUCN Assessment status. Listed for each IUCN assessment status are the total number of species per functional group, the number of these species with occurrences in OBIS, and the associated percentage.

Table 4B Breakdown of marine animal species by functional group and presence in the BOLD DNA Barcode database. Listed for species absente from or present in BOLD are the total number of species per functional group, the number of these species with occurrences in OBIS, and the associated percentage.

	In Barcode of Life Database?								
		No		Yes					
Functional Group	N(species)	N(species in OBIS)	% species in OBIS	N(species)	N(species in OBIS)	% species in OBIS			
Benthos	131,390	62,316	47%	15,161	13,288	88%			
Zooplankton	4,768	2,117	44%	980	914	93%			
Nekton	2,506	1,355	54%	893	803	90%			
Fish	8,683	5,842	67%	9,578	9,123	95%			
Mammals	85	37	44%	107	103	96%			
Birds	238	108	45%	449	395	88%			

Reptiles	80	59	70%	16	15	94%
Other / Unknown	30,292	8,692	29%	1,623	1,049	65%
Totals	178,042	80,523	45%	28,807	25,690	89%

Figure Legends

Figure 1. Availability of biogeographic (>45M OBIS occurrence records) and genetic (>56M GenBank nucleotides) data across 206,849 marine animal species, summarised by phylum and by broad functional group. **(A)** Proportion of species in each phylum with data in either database, both databases, or neither. Bar width is proportional to the number of species in each phylum. Number of **(B)** OBIS occurrence records and **(C)** Genbank nucleotide sequences are shown for species that occur in the respective database. Each point represents a species, coloured by functional group. Box plots are superimposed with X marking the median number of records within each phylum. Phylum size varies from 2 species (Cycliophora) to 57,336 species (Arthropoda).

Figure 2. Coefficients from the hurdle models of data availability across functional groups, first modelling presence in a database with a binomial model, and then non-zero counts of records in a database as a negative binomial model. Species presence in OBIS (**A**) or GenBank nucleotide database (**C**) across functional groups is indicated with binomial coefficients (with 95% confidence intervals) on the response scale, representing the ratio of the probabilities of species within a group having records in the database versus not having records in the database. For the subset of species present in (**B**) OBIS or (**D**) GenBank, the empirical mean number of records per species is plotted together with bootstrapped 95% confidence intervals. For each group, the predicted non-zero count from the hurdle model is indicated with an X. Point size is scaled to the total number of species in each functional group (A, C, ranging from 96 reptiles to 146,551 benthos) and to the number of species in each group with records in OBIS (B, 71 reptiles to 75,604 benthos) or GenBank (D, 78 reptiles to 19,235 benthos).

Figure 3. Mosaic plot showing the joint distribution of species between categories of OBIS records and GenBank nucleotides. (A) shows all species, and is dominated by species with no records in either database. (B) zooms in on species with high numbers (>100) of OBIS records, and (C) reverses the axes and zooms in on species with high numbers (>100) of GenBank nucleotides. Axis labels indicate the number of records at the right-hand bound of each category.

Figure 4. Distribution of occurrence records across 106,213 marine animal species present in OBIS by functional group and by **(A)** IUCN assessment status and **(B)** presence in the Barcode of Life Data System. Each point represents a species.



no data

OBIS only

OBIS and GenBank

GenBank only







functional group nekton reptiles birds benthos other/unknown fish zooplankton mammals

C















⊳