



Deposited via The University of Sheffield.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/165023/>

Version: Published Version

Article:

Kriston-Pál, É., Haracska, L., Cooper, P. et al. (2020) A regenerative approach to canine osteoarthritis using allogeneic, adipose-derived mesenchymal stem cells. Safety results of a long-term follow-up. *Frontiers in Veterinary Science*, 7. 510.

<https://doi.org/10.3389/fvets.2020.00510>

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:

<https://creativecommons.org/licenses/>

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



A Regenerative Approach to Canine Osteoarthritis Using Allogeneic, Adipose-Derived Mesenchymal Stem Cells. Safety Results of a Long-Term Follow-Up

Éva Kriston-Pál¹, Lajos Haracska², Paul Cooper³, Endre Kiss-Tóth⁴, Valéria Szukacsov² and Éva Monostori^{2*}

¹ Stem CellX Limited, Szeged, Hungary, ² Institute of Genetics, Biological Research Centre, Szeged, Hungary, ³ Assentra Limited, Chelmsford, United Kingdom, ⁴ University of Sheffield, Department of Infection, Immunity & Cardiovascular Disease, Sheffield, United Kingdom

OPEN ACCESS

Edited by:

Mário Ginja,
University of Trás-os-Montes and Alto
Douro, Portugal

Reviewed by:

Fidel San Roman Llorens,
University of Zaragoza, Spain
Sheila Canevese Rahal,
São Paulo State University, Brazil

*Correspondence:

Éva Monostori
monostori.eva@brc.hu

Specialty section:

This article was submitted to
Veterinary Imaging,
a section of the journal
Frontiers in Veterinary Science

Received: 15 March 2020

Accepted: 03 July 2020

Published: 13 August 2020

Citation:

Kriston-Pál É, Haracska L, Cooper P,
Kiss-Tóth E, Szukacsov V and
Monostori É (2020) A Regenerative
Approach to Canine Osteoarthritis
Using Allogeneic, Adipose-Derived
Mesenchymal Stem Cells. Safety
Results of a Long-Term Follow-Up.
Front. Vet. Sci. 7:510.
doi: 10.3389/fvets.2020.00510

Mesenchymal stem cells (MSC) are emerging as an effective therapeutic tool in treating canine osteoarthritis (OA). In this report, we focused on the questions of whether MSC transplantation has long-term beneficial effects for the improvement in motion and also evaluated the safety of MSC injection. Visceral adipose tissue, a surgical waste obtained during routine ovarioectomy served as a source of allogeneic MSCs and used to treat OA. Altogether, fifty-eight dogs were transplanted in the study suffering from OA in the elbow (42 animals), hip (5), knee (8), ankle (2), and hock (1). The effect of MSC transplantation was evaluated by the degree of lameness at a 4-5-years follow-up period based on the owners' subjective observations. The results showed that 83% of the OA patients improved or retained improvement in lameness. Clinical safety of the treatment was assessed by evaluating the coincidence of tumors or other diseases and other adverse reactions (such as local inflammation) after MSC cell therapy. Two incidences of local inflammation for <1 week at the site of injection were reported. No other adverse reactions were detected post-treatment. Sixteen dogs died during the study, 4 due to cancer and 12 due to other diseases, diagnosed by veterinarians. Overall, our survey suggests that MSC transplantation has long-term beneficial effects in reducing lameness. Moreover, no enrichment in a specific cause of death was observed in the transplanted animals, compared to reported literature. Our data suggest that MSC treatment could be an effective and safe long-term therapy for canine OA.

Keywords: dogs, osteoarthritis, mesenchymal stem cell, therapy, safety of therapy, long-term follow-up

INTRODUCTION

Osteoarthritis (OA), in which the integrity of joint cartilage is disrupted, is one of the most prevalent degenerative diseases, both in dogs and humans (1, 2). Due to the low self-regeneration capacity of the cartilage matrix, the disease is irreversible, thus, the quality of life of the affected animal is expected to decrease gradually (3). In the United Kingdom alone, 200,000 new cases of OA are diagnosed in dogs annually, with both external (injury, obesity, age) and genetic factors

contributing (1, 4, 5). At present, the disease is treated symptomatically by regularly administering non-steroidal anti-inflammatory drugs or by repeated injections of hyaluronic acid (6, 7). Surgical solutions such as arthrodesis or excision arthroplasty (e.g., total joint replacement) are also used in more severe cases (8). However, these are invasive procedures, which may also carry risks of complications such as infections, instability, or periprosthetic fracture (2). In view of the above, the development of a safe and long-term regenerative treatment for OA is highly sought after.

MSCs are excellent candidates for this purpose with a rapidly expanding published literature demonstrating the effectiveness of these cells in tissue regeneration in various diseases (9). An increasing body of evidence now demonstrates that MSCs administered directly into the joint cavity can reduce the chronic pain caused by cartilage degeneration, can induce hyaline cartilage formation, and the treated dogs can live a highly improved quality of life (10, 11). Though MSCs are present in all tissues, adipose tissue is a major source of therapeutic MSCs due to easy access and their high numbers (12).

Besides being able to differentiate into chondrocytes, MSCs produce bioactive molecules, some of which have chondroprotective activity, while others are immunosuppressive and/or anti-inflammatory, thus enabling the safe injection of MSCs into recipients (13). However, the immunological status of the microenvironment is also critical since differentiation of MSCs is inhibited by inflammation (14).

We previously reported that intra-articular injection of allogeneic, adipose-derived MSCs, combined with hyaluronic acid induced a significant improvement in clinical signs of lameness, lasting for 1 year in dogs suffering from elbow OA (11) and now provide data on the long-term (>4 years) health status of the treated animals. To the best of our knowledge, the current report is the first such long-term survey published. We hypothesized that MSCs are effective in reducing lameness and improving the quality of life of animals and that local MSC transplantation is not associated with an increased prevalence of other diseases or malignancies.

METHODS

Preparation of Visceral Adipose Tissue (AT) Mesenchymal Stem Cells (MSCs) for Therapy

MSCs were extracted from visceral adipose tissue. Adipose tissue was obtained as surgical waste during routine ovarioectomy of healthy, female, mixed-breed dogs (age: between 7 months and 3 years). Donor dogs underwent all routine vaccinations and were regularly surveyed by veterinarians. Stromal vascular fraction (SVF) and then AT-MSC cultures were generated as previously described (11). The purity and differentiation ability of the resulting MSCs were characterized as described previously (11).

Preparation of AT-MSCs for Therapy

Passage 2 AT-MSCs from two different adipose tissue donors were thawed, cultured for 3 days, mixed, and suspended in 0.5%

sodium hyaluronate (TRB Chemedica International S.A. Geneva, Switzerland). Adipose tissue-derived MSCs ($12 \times 10^6 \pm 3.2 \times 10^6$ cells/injection) were transported to the veterinarian clinics in syringes at 4–10°C and injected intra-articularly, within 24 h of dispensing, in a final volume of 1 mL.

Patient Selection and Assessment of the Therapy

This study was approved by the National Scientific Ethical Committee of the National Food Chain Safety Office. All the dog owners signed an informed consent authorizing treatment and were informed of the possible risks of joint injections and potential complications of the procedure.

Fifty-eight dogs suffering from medium or severe osteoarthritis of various joints were included in the study. Severity of OA was evaluated by the participating veterinarians according to a modified method of Black et al. (15) as medium (intermittent but frequent lameness treated with NSAID medication or hyaluronic acid injection) or severe (no cartilage based on the X-ray analysis, continuous pain and lameness, reduction of joint movement, and presence of joint stiffness). The inclusion criteria were recurrent lameness and pain attributed to OA after conventional treatment of dogs (non-steroidal anti-inflammatory drugs, intra-articular injection of hyaluronic acid, arthroscopy, or traditional surgery focusing on debridement and removal of debris from the synovial cavity). Only those dogs were classed as “improved” that did not require further conventional treatment due to their OA during the study period.

The health status, including degree of lameness of the treated dogs was evaluated by the owners using a questionnaire modified from Black et al. (15) (**Supplementary Table 1**), with an occasional participation of the local veterinarian during the first year of follow-up. Further follow-up of up-to 5 years was based on the owners' subjective observations, reporting the degree and frequency of lameness and the possible necessity of pain relief medication since the MSC treatment in personal telephone interviews. The development of other diseases or cause of death was diagnosed by veterinarians during the 5 year follow-up period. The owner-assessed efficacy of the MSC transplantation in OA was evaluated to provide long-term, supportive information on the clinical outcomes following this treatment.

There were no exclusion criteria except for pre-existing, diagnosed cancer.

RESULTS

Overall, fifty-five out of 58 transplanted animals were available for the long-term follow-up. Forty-two animals suffered from elbow OA (**Table 1** and **Supplementary Table 2**); all suffering from elbow dysplasia, except two dogs (aged 12 and 13 years) which had developed cartilage degeneration probably due to their age. Eleven dogs received MSCs into both elbows, while 31 animals obtained transplant into one joint (**Supplementary Table 2**). Thirty-nine dogs were available for the

TABLE 1 | Summary of long-term follow-up after MSC transplantation.

Site of OA	No. of animals included/No. of animals evaluated	Range of age (average) at the time of MSC injection in years	Average age at the time of follow-up (years)	Results at 2.5, 4, or 5-year follow-up			Joint or spinal diseases other than the transplanted joint	Number and cause of death	
				Improved or sporadic lameness and/or medication during weather fronts or extreme activity	No improvement continuous medication for other joint/spinal problems or death			Other than cancer	Cancer
Elbow OA	39*/31**	0.6–10 (3)	7.3	26 (84%)	5	9	8	3	
Hip OA	5/4&&	1.5–8 (5.5)	9.7	3	1	2	1	1	
Knee OA, dislocation, ligament tear	8/6&&	3–10 (5.6)	10	5	1	3	3	-	
Hock OA	1	0.8	5	1	-	-	-	-	
Ankle OA	2	0.4–1.5 (0.96)	5.5	2	-	-	-	-	
Total	55&/44&&	0.4–10 (3.2)	7.5	37 (84%)#	7 (16%)	14	12	4	

The condition (lameness, usage of drugs) of the transplanted joint of the same animal was followed up for 2.5, 4, or 5 years depending on the date of transplantation.

% = (improved animal number: number of animals in the study) × 100.

* 6 dogs died within 1 year of the survey.

** 3 of them died just before the end of the survey, but their condition was evaluated till death.

& One dog died within 1 year of the survey and 3 died later but before the end of the follow-up.

&& One dog who died before the end of the survey was evaluated till death.

long-term follow-up, by the end of which the average age of the dogs in the study increased from 3 to 7.3 years.

Sixteen dogs that received the MSC treatment into other joints [knee (8), hip (5), ankle (2), and hock (1)] (one joint in one animal, see **Supplementary Table 2**) were also evaluated in the long-term follow-up (**Table 1**). The average age was 4.8 years at the time of transplantation, increasing to 9 years by the end of the study period.

Thirty-seven dogs out of 44 (84%), were reported by the owners to have reduced lameness and an improved quality of life until the end of the follow-up period or the death of the treated animals. Several dogs improved, and although suffered from other joint problems did not require medication. Seven dogs diagnosed with other joint or spinal problems besides elbow OA and hence receiving continuous medication were also included into the survey period of elbow OA (**Table 1**).

Two of 58 dogs showed a short-term local inflammation after MSC transplantation. This inflammatory reaction resolved within a week and did not affect the long-term improvement in the quality of the dogs' life (data not shown).

Sixteen dogs between the ages of 7 and 13 died during the follow-up period, four of them of cancer (between the ages of 11 and 13) and 12 due to various other diseases (aged 7–12 years) (**Table 1**). Cases of cancer included: melanoma, epithelial cancer, metastatic cancer, spleen-liver cancer (a single case for each tumor-type). Other co-morbidities reported by the owners included: epilepsy, pancreatitis, volvulus, heart failure, ulcers, neurological, and musculo-skeletal problems. None of these syndromes were reported more than twice, suggesting that they may not be associated with the MSC treatment.

DISCUSSION

The aim of this study was to explore whether local intra-articular transplantation of MSC had long-term beneficial effects on lameness and hence improving quality of life of the transplanted animals and whether it caused any serious adverse effects or correlated with an increased prevalence of other diseases.

Our data presented in this paper suggest that MSC transplantation results in improvement of motion. In the current study, 60% of the treated animals belonged to 3 breeds: Golden Retriever (9), Labrador Retriever (16), and German Shepherd (8). This is consistent with these breeds being generally recognized as at high risk of OA (1). The sex of the animals did not correlate with either the coincidence of OA or the death occurring subsequently to MSC injection. Our results presented here are in line with the literature suggesting a beneficial effect of MSC transplantation in OA (11, 16).

In the cohorts included in the long-term survey, most dogs suffered from elbow OA (42 dogs out of 58) of which 40 were diagnosed with elbow dysplasia, hence elbow dysplasia was accepted as the main cause of osteoarthritis in the elbow. Eighty four percentage of the animals maintained an improved condition (no lameness/no medication or sporadic lameness and/or medication during wet weather or with extreme activity) by the subjective assessment of the owners.

Osteoarthritis cases in joints other than the elbow (knee, hip, ankle, and hock) were also evaluated. Eighty four percentage of the evaluated dogs retained their improved condition after the 4-5-year follow-up. It should be noted that the dogs in this mixed group were of higher age than those in the elbow group, and many patients suffered from joint or spinal problems in addition to the treated joint.

MSC transplantation does not appear to be associated with an increase in malignancies or other diseases, and no other adverse effects emerged due to MSC injection. These findings are underpinned by: (1) the published literature that supports that MSC is not a tumorigenic cell type (17, 18); (2) published findings that local injection into the joint results in the adherence of MSCs to the damaged cartilage (19) with no reported evidence for their migration outside of the treated joint. Sixteen dogs out of 58 transplanted animals died during the long-term follow-up: four due to cancer in old age (11–13 years) and 12 from other different causes.

This rate, in spite of the small number of cases overall, is comparable to large-scale disease prevalence statistics in the USA (5.3%) thus does not indicate an association between prevalence of cancer and MSC transplantation (20). Other studies report a prevalence of 3.4–8.63% for melanoma (in dogs the age average was 7, 5 years) in Switzerland and Brazil and 3.4% for internal organ tumors in India over a period of 10 years (21–23). Published literature demonstrates a 4.9% prevalence of death due to heart failure in dogs with a median of 9.9 years in the UK (24), whilst our study reports 3.6%. Of note, the dog population included in our follow-up study is comprised of older dogs, thus, the number of deaths due to tumors is expected to be higher than that in the whole population. In our study, however, the average age of the dogs that died due to tumors was 12.2 years, while in a previous report from the USA involving golden retrievers it was 9.83 years (25).

The potential use of allogeneic MSCs for the treatment of various diseases has also been piloted, including humans and horses, for instance, for indications such as cardiac damage post-myocardial infarction and wound healing (26, 27). However, whilst long-term safety data is still very sparse, evidence is now emerging on a convincing safety profile for the use of allogeneic MSCs in cardiac regeneration (28) and autoimmune disease in man (29) as well as in osteoarthritis in horses (30). Our current report is in alignment with this broader literature and further highlights the potential for the use of allogeneic MSCs in a variety of diseases.

In summary, our data suggest that intra-articular injection of allogeneic MSCs may provide a long-term improvement in lameness secondary to OA, based on subjective reporting of the owners, without the risk of long-term adverse effects on health.

Thus, we conclude that MSC treatment could be an effective and safe long-term therapy for canine OA.

Though the results overall suggest that MSC treatment is beneficial for dogs suffering from osteoarthritis, evaluation is limited by the fact that the improvement was subjectively compared to the initial state by owners. To overcome these limitations, a follow-up with rigorous orthopedic examination performed by veterinarians will be recommended.

DATA AVAILABILITY STATEMENT

All datasets generated for this study are included in the article/ **Supplementary Material**.

ETHICS STATEMENT

The animal study was reviewed and approved by Csongrád County Committee on the Food Chain and Animal Health, Government Office of Csongrád County, Rákóczi tér 1, Szeged 6722 Hungary. Written informed consent was obtained from the owners for the participation of their animals in this study.

AUTHOR CONTRIBUTIONS

EK-P: preparing MSCs for therapy. LH: organization of contacts with vets and owners. PC and EK-T: Evaluating results. VS: keeping contact and interviewing the owners. EM: leading the study. All authors: contributed to writing the manuscript.

ACKNOWLEDGMENTS

We thank Mrs. András Gercsó the excellent laboratory assistance, Ms. Gabriella Tick for the extensive help in writing the manuscript, Prof. Péter Vajdovich (Chairman of Animal Oncology at Veterinary University, Budapest) the important professional input, and Ms. Karolina Korom the assistance in interviewing the owners. We are also grateful to the veterinarians who treated the dogs included into this study: Drs. Ottó Sebo (Small Animal Clinic, Makó), Péter Pál Muka (Profivet Veterinary Surgery Center and Animal Hospital, Göd), Péter Makai (MiniVet Small Animal Clinic, Budapest), György Pelle (Teaching Animal Hospital, Nyíregyháza), and András Bánfi (PrimaVet Small Animal Clinic, Budapest).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fvets.2020.00510/full#supplementary-material>

REFERENCES

- Anderson KL, O'Neill DG, Brodbelt DC, Church DB, Meeson RL, Sargan D, et al. Prevalence, duration and risk factors for appendicular osteoarthritis in a UK dog population under primary veterinary care. *Sci Rep.* (2018) 8:5641. doi: 10.1038/s41598-018-23940-z
- Kwak JM, Koh KH, Jeon IH. Total elbow arthroplasty: clinical outcomes, complications, and revision surgery. *Clin Orthop Surg.* (2019) 11:369. doi: 10.4055/cios.2019.11.4.369

3. Hunziker EB, Lippuner K, Keel MJB, Shintani N. An educational review of cartilage repair: precepts & practice - myths & misconceptions - progress & prospects. *Osteoarthr Cartil.* (2015) 23:334–50. doi: 10.1016/j.joca.2014.12.011
4. Poulet B, Staines KA. New developments in osteoarthritis and cartilage biology. *Curr Opin Pharmacol.* (2016) 28:8–13. doi: 10.1016/j.coph.2016.02.009
5. Anderson KL, Zulch H, O'Neill DG, Meeson RL, Collins LM. Risk factors for canine osteoarthritis and its predisposing arthropathies: a systematic review. *Front Vet Sci.* (2020) 7:220. doi: 10.3389/fvets.2020.00220
6. Zhang W, Ouyang H, Dass CR, Xu J. Current research on pharmacologic and regenerative therapies for osteoarthritis. *Bone Res.* (2016) 4:15040. doi: 10.1038/boneres.2015.40
7. Silva Júnior JIS, Rahal SC, Santos IFC, Martins DJC, Michelon F, Mamprim MJ, et al. Use of reticulated hyaluronic acid alone or associated with ozone gas in the treatment of osteoarthritis due to hip dysplasia in dogs. *Front Vet Sci.* (2020) 7:265. doi: 10.3389/fvets.2020.00265
8. Montasell X, Dupuis J, Huneault L, Ragetly GR. Short- and long-term outcomes after shoulder excision arthroplasty in 7 small breed dogs. *Can Vet J.* (2018) 59:277–83.
9. Rajabzadeh N, Fathi E, Farahzadi R. Stem cell-based regenerative medicine. *Stem Cell Investig.* (2019) 6:19. doi: 10.21037/sci.2019.06.04
10. Shah K, Drury T, Roic I, Hansen P, Malin M, Boyd R, et al. Outcome of allogeneic adult stem cell therapy in dogs suffering from osteoarthritis and other joint defects. *Stem Cells Int.* (2018) 2018:7309201. doi: 10.1155/2018/7309201
11. Kriston-Pál É, Czibula Á, Gyuris Z, Balka G, Seregi A, Sükösd F, et al. Characterization and therapeutic application of canine adipose mesenchymal stem cells to treat elbow osteoarthritis. *Can J Vet Res.* (2017) 81:73–8.
12. da Silva Meirelles L, Chagastelles PC, Nardi NB. Mesenchymal stem cells reside in virtually all post-natal organs and tissues. *J Cell Sci.* (2006) 119:2204–13. doi: 10.1242/jcs.02932
13. Sun Q, Huang Z, Han F, Zhao M, Cao R, Zhao D, et al. Allogeneic mesenchymal stem cells as induction therapy are safe and feasible in renal allografts: pilot results of a multicenter randomized controlled trial. *J Transl Med.* (2018) 16:52. doi: 10.1186/s12967-018-1422-x
14. Szabó E, Fajka-Boja R, Kriston-Pál É, Hornung Á, Makra I, Kudlik G, et al. Licensing by inflammatory cytokines abolishes heterogeneity of immunosuppressive function of mesenchymal stem cell population. *Stem Cells Dev.* (2015) 24:2171–80. doi: 10.1089/scd.2014.0581
15. Black LL, Gaynor J, Gahring D, Adams C, Aron D, Harman S, et al. Effect of adipose-derived mesenchymal stem and regenerative cells on lameness in dogs with chronic osteoarthritis of the coxofemoral joints: a randomized, double-blinded, multicenter, controlled trial. *Vet Ther.* (2007) 8:272–84.
16. Cabon Q, Febre M, Gomez N, Cachon T, Pillard P, Carozzo C, et al. Long-term safety and efficacy of single or repeated intra-articular injection of allogeneic neonatal mesenchymal stromal cells for managing pain and lameness in moderate to severe canine osteoarthritis without anti-inflammatory pharmacological support: pilot clinical study. *Front Vet Sci.* (2019) 6:10. doi: 10.3389/fvets.2019.00010
17. Park SJ, Kim HJ, Kim W, Kim OS, Lee S, Han SY, et al. Tumorigenicity evaluation of umbilical cord blood-derived mesenchymal stem cells. *Toxicol Res.* (2016) 32:251–8. doi: 10.5487/TR.2016.32.3.251
18. Sivanathan KN, Coates PT. Improving human kidney function in renovascular disease with mesenchymal stem cell therapy. *Kidney Int.* (2020) 97:655–6. doi: 10.1016/j.kint.2019.12.020
19. Pak J, Lee JH, Pak N, Pak Y, Park KS, Jeon JH, et al. Cartilage regeneration in humans with adipose tissue-derived stem cells and adipose stromal vascular fraction cells: updated status. *Int J Mol Sci.* (2018) 19:2146. doi: 10.3390/ijms19072146
20. Schiffman JD, Breen M. Comparative oncology: what dogs and other species can teach us about humans with cancer. *Philos Trans R Soc B Biol Sci.* (2015) 370:20140231. doi: 10.1098/rstb.2014.0231
21. Tostes RA, Branco A, Cestari FK, Caleffo T, De Marco Viott A. Retrospective study of canine cutaneous neoplasia. *Arch Vet Sci.* (2017) 22:71–80. doi: 10.5380/avs.v22i1.49290
22. Graf R, Pospischil A, Guscetti F, Meier D, Welle M, Dettwiler M. Cutaneous tumors in swiss dogs: retrospective data from the swiss canine cancer registry, 2008–2013. *Vet Pathol.* (2018) 55:809–20. doi: 10.1177/0300985818789466
23. Lather D, Gupta RP, Sharma S. Retrospective studies on tumor conditions in dogs over a period of ten years (2005–2014). *Haryana Vet.* (2017) 56:47–49.
24. Lewis TW, Wiles BM, Llewellyn-Zaidi AM, Evans KM, O'Neill DG. Longevity and mortality in Kennel Club registered dog breeds in the UK in 2014. *Canine Genet Epidemiol.* (2018) 5:10. doi: 10.1186/s40575-018-0066-8
25. Kent MS, Burton JH, Dank G, Bannasch DL, Rebhun RB. Association of cancer-related mortality, age and gonadectomy in golden retriever dogs at a veterinary academic center (1989–2016). *PLoS ONE.* (2018) 13:e0192578. doi: 10.1371/journal.pone.0192578
26. Poh KK, Sperry E, Young RG, Freyman T, Barringhaus KG, Thompson CA. Repeated direct endomyocardial transplantation of allogeneic mesenchymal stem cells: safety of a high dose, “off-the-shelf”, cellular cardiomyoplasty strategy. *Int J Cardiol.* (2007) 117:360–4. doi: 10.1016/j.ijcard.2006.04.092
27. Textor JA, Clark KC, Walker NJ, Aristizobal FA, Kol A, LeJeune SS, et al. Allogeneic stem cells alter gene expression and improve healing of distal limb wounds in horses. *Stem Cells Transl Med.* (2018) 7:98–108. doi: 10.1002/sctm.17-0071
28. Natsumeda M, Florea V, Rieger AC, Tompkins BA, Banerjee MN, Golpanian S, et al. A combination of allogeneic stem cells promotes cardiac regeneration. *J Am Coll Cardiol.* (2017) 70:2504–15. doi: 10.1016/j.jacc.2017.09.036
29. Liang J, Zhang H, Kong W, Deng W, Wang D, Feng X, et al. Safety analysis in patients with autoimmune disease receiving allogeneic mesenchymal stem cells infusion: a long-term retrospective study. *Stem Cell Res Ther.* (2018) 9:312. doi: 10.1186/s13287-018-1053-4
30. Magri C, Schramme M, Febre M, Cauvin E, Labadie F, Saulnier N, et al. Comparison of efficacy and safety of single versus repeated intra-articular injection of allogeneic neonatal mesenchymal stem cells for treatment of osteoarthritis of the metacarpophalangeal/metatarsophalangeal joint in horses: a clinical pilot study. *PLoS ONE.* (2019) 14:e0221317. doi: 10.1371/journal.pone.0221317

Conflict of Interest: EK-P was employed by company Stem CellX Europe Limited. PC and EK-T are shareholders of Stem CellX Europe Limited and PC is a shareholder of Assentra Limited.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Kriston-Pál, Haracska, Cooper, Kiss-Tóth, Szukacsov and Monostori. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.