

This is a repository copy of *Phytoestrogen intake and other dietary risk factors for low motile sperm count and poor sperm morphology*.

White Rose Research Online URL for this paper: https://eprints.whiterose.ac.uk/164085/

Version: Published Version

# Article:

Povey, A.C., Clyma, J., McNamee, R. et al. (5 more authors) (2020) Phytoestrogen intake and other dietary risk factors for low motile sperm count and poor sperm morphology. Andrology, 8 (6). pp. 1805-1814. ISSN 2047-2919

https://doi.org/10.1111/andr.12858

# Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here: https://creativecommons.org/licenses/

# Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



# ORIGINAL ARTICLE

# ANDROLOGY 😂 🔛 WILEY

# Phytoestrogen intake and other dietary risk factors for low motile sperm count and poor sperm morphology

Andy C. Povey<sup>1</sup> | Julie-Ann Clyma<sup>1</sup> | Roseanne McNamee<sup>2</sup> | Harry D. Moore<sup>3</sup> | Helen Baillie<sup>4</sup> | Allan A. Pacey<sup>3</sup> | Janet E. Cade<sup>5</sup> | Nicola M. Cherry<sup>6</sup> | Participating Centres of Chaps-UK

<sup>1</sup>Epidemiology and Public Health Group, Faculty of Biology, Medicine and Health, Division of Population Health, Health Services Research and Primary Care, School of Health Sciences, University of Manchester, Manchester Academic Health Science Centre, Manchester, UK

<sup>2</sup>Centre for Biostatistics, Faculty of Biology, Medicine and Health, Division of Population Health, Health Services Research and Primary Care, School of Health Sciences, University of Manchester, Manchester Academic Health Science Centre, Manchester, UK

<sup>3</sup>Department of Oncology and Metabolism, School of Medicine and Biomedical Sciences, University of Sheffield, Sheffield, UK

<sup>4</sup>Jessop Fertility, The Jessop Wing, Sheffield Teaching Hospitals, Tree Root Walk, Sheffield, UK

<sup>5</sup>Nutritional Epidemiology Group, School of Food Science and Nutrition, University of Leeds, Leeds, UK

<sup>6</sup>Division of Preventive Medicine, University of Alberta, Edmonton, AB, Canada

#### Correspondence

Andy C. Povey, Epidemiology and Public Health Group, Division of Population Health, Health Services Research and Primary Care, School of Health Sciences, University of Manchester, Manchester, UK. Email: apovey@manchester.ac.uk

#### **Funding information**

The study was funded by the UK Health and Safety Executive, the UK Department of Environment, Transport and the Regions, the UK Department of Health (grant code DoH 1216760), and the European Chemical Industry Council (grant code EMSG19).

#### Abstract

**Background:** Few potentially modifiable risk factors of male infertility have been identified, and while different diets and food groups have been associated with male infertility, evidence linking dietary factors including phytoestrogens and semen quality is limited and contradictory.

**Objectives:** To study the associations between phytoestrogen intake and other dietary factors and semen quality.

**Materials and Methods:** A case-referent study was undertaken of the male partners, of couples attempting conception with unprotected intercourse for 12 months or more without success, recruited from 14 UK assisted reproduction clinics. A total of 1907 participants completed occupational, lifestyle and dietary questionnaires before semen quality (concentration, motility and morphology) were assessed. Food intake was estimated by a 65-item food frequency questionnaire (FFQ) covering the 12 months prior to recruitment. Analyses of dietary risk factors for low motile sperm concentration (MSC: <4.8  $\times$  10<sup>6</sup>/mL) and poor sperm morphology (PM: <4% normal morphology) used unconditional logistic regression, accounting for clustering of subjects within the clinics, first without, and then with, adjustment for confounders associated with that outcome.

**Results:** High consumption of daidzein ( $\ge 13.74 \ \mu g/d$ ), a phytoestrogen found in soy products, was a protective factor for MSC with an odds ratio (95%CI) of 0.58 (0.42-0.82) after adjustment for clustering and potential confounding. Dietary risk factors for PM after similar adjustment showed that drinking whole milk (OR 0.67, 95%CI 0.47-0.96) and eating red meat were protective with an OR 0.67 (0.46-0.99) for eating red meat >3 times/wk.

**Discussion:** In this case-referent study of men attending an infertility clinic for fertility diagnosis, we have identified that low MSC is inversely associated with daidzein intake. In contrast, daidzein intake was not associated with PM but eating red milk and drinking whole milk were protective.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. Andrology published by Wiley Periodicals LLC on behalf of American Society of Andrology and European Academy of Andrology

**Conclusions:** Dietary factors associated with semen quality were identified, suggesting that male fertility might be improved by dietary changes.

KEYWORDS

semen quality, sperm motility, sperm morphology, daidzein, phytoestrogens, diet

# 1 | INTRODUCTION

Male subfertility is a significant factor in approximately 50% of all couples experiencing a period of infertility.<sup>1</sup> Despite widespread concern about reported but disputed declines in semen quality and more specifically total sperm count,<sup>2,3</sup> few modifiable factors of semen quality have so far been identified (eg 4-6). The characterization of these factors is of particular interest since they offer a means to improve male fertility and potentially reduce the need for assisted reproductive technologies.

Diet is possibly one such factor, and while the relationship between conventional semen quality parameters (concentration, motility and morphology) and diet patterns and specific dietary components including dietary phytoestrogens has been studied (eg 7-9), the results from these association studies are often inconsistent. Increasing consumption of "Western diets" has been associated with increased sperm concentration,<sup>10</sup> but not in all studies,<sup>11-13</sup> and furthermore has been both positively<sup>12</sup> and negatively<sup>10</sup> associated with sperm morphology. Similarly, increasing intake of diets considered healthy (eg Mediterranean) has been reported to improve sperm concentration or total sperm count in some<sup>12-14</sup> but not all<sup>10,15</sup> studies. In contrast, sperm motility has not been associated with diets considered healthy<sup>10,12,15</sup> or indeed a Western diet.<sup>10-12</sup> though a "prudent" dietary pattern has been positively associated with motility<sup>11</sup> but not consistently so.<sup>13,14</sup> In addition, despite the reported associations between endocrine disrupting chemicals and male fertility,<sup>16</sup> there are few studies that have examined associations between semen quality and oestrogenic phytoestrogens that are found in plant-based foods (especially soy). Soy food intake has been inversely associated with sperm concentration but not sperm motility nor morphology,<sup>17</sup> but a feeding trial of a phytoestrogen supplement did not find any evidence of changes in semen quality despite increases in plasma levels of the isoflavone phytoestrogens, genistein and daidzein.<sup>18</sup>

The inconsistency in the reported associations between these different food groups and semen quality may reflect methodological differences in study design (eg subfertile men vs healthy fertile men), study size or study populations (eg Asian vs Western). There are also intrinsic differences in the measurement of food intake and in how food groups were defined from one study to another. As a result, the evidence base linking diet to male subfertility is limited and often contradictory and there are no dietary recommendations currently reported in the UK National Institute for Health and Care Excellence guidelines on fertility problems: assessment and treatment.<sup>19</sup> To improve this evidence base, in this paper we examine associations

between dietary factors (with a particular focus on phytoestrogen intake) and low motile sperm concentration and sperm morphology in a large multi-centre case-referent study (CHAPS-UK).

# 2 | MATERIALS AND METHODS

## 2.1 | Design and recruitment

The CHAPS-UK study was a multi-site, case-referent study, and the design and methods have been described previously.<sup>20</sup> Cases and referents were the male partners of couples attempting conception with unprotected intercourse for 12 months or more without success and were recruited between 1 January 1999 and 31 January 2002. Men were excluded if they had prior knowledge of their own semen quality, were not able to understand English, had had a medical condition (eg cystic fibrosis) or medical treatment (eg chemotherapy) that could have caused their infertility and either they or their partner had been sterilized previously.<sup>20</sup> Prior to their first clinic visit following recruitment, participants completed a short questionnaire on work, lifestyle and health factors and further information on these factors was obtained by interview at the clinic. Men were asked to complete a dietary questionnaire at home with their partner in order to obtain the most complete and accurate information.

#### 2.2 | Semen analysis

Men were requested to abstain from ejaculation for a period of 3-5 days (depending on the clinic) prior to providing a semen sample for a diagnostic analysis. This sample was used not only for infertility investigations but also for this study, and it was analysed according to a protocol based upon the techniques outlined by the World Health Organization<sup>21</sup> as described previously.<sup>5,20</sup> Sperm concentration was estimated at an andrology laboratory associated with each centre using a haemocytometer. Motility was captured on videotape using a computer outstation commissioned for the study from Hobson Tracker Systems Limited, Sheffield, UK, and the tape was returned to the central laboratory for analysis of sperm motility by computer-assisted sperm analysis (CASA). The proportion of motile sperm was calculated as the % of sperm moving forward at 5  $\mu$ m/s or greater. Morphology slides were prepared and fixed at each site but analysed at the central laboratory by the Papanicolaou method and 200 spermatozoa assessed using a Computer Aided Sperm Morphometric Assessment system developed by Hobson Tracker

ANDROLOGY

Systems (Sheffield, UK). The machine was programmed to recognize as 'normal' stained sperm heads which fitted the dimensions given in WHO  $1999^{21}$ : a length of 4.0-5.0 mm and a width of 2.5-3.5 mm, with a length-to-width ratio between 1.50 and 1.75.

# 2.3 | Case definitions

Two case definitions were used, reflecting WHO guidelines.<sup>22</sup> The first was based on sperm motility, and cases were men with a motile sperm concentration (MSC) of  $<4.8 \times 10^6$ /mL: referents a MSC of  $\geq 4.8 \times 10^6$ /mL. The second definition was based on morphology, and cases with poor morphology (PM) were men whose sperm showed <4% normal morphology; referents had  $\geq 4\%$  normal forms.

## 2.4 | Dietary exposures

Information on dietary habits was collected using a questionnaire (Appendix S1) that included a 65-item food frequency questionnaire (FFQ) covering the 12 months prior to recruitment to assess phytoestrogen intake that was developed from then available information on phytoestrogen levels in food (eg 23-25). The following exposures were assessed:

- Type of diet (Q3.1; Appendix S1) coded into three categories: meat ± fish eater, fish eater, vegan or vegetarian.
- Frequency of red meat consumption (Q1.3; Appendix S1) recoded into three categories; ≤1/wk, >1 ≤ 3 times/wk, and >3 times/wk.
- Frequency of poultry consumption (Q1.2; Appendix S1) recoded into three categories ≤ 1/wk,> 1 ≤ 3 times/wk, and > 3 times/wk.
- Frequency of fish consumption (Q2.2; Appendix S1) recoded into three categories ≤ 1/wk,> 1 ≤ 3 times/wk, and > 3 times/wk.
- Number of portions of vegetables/d; (product of use and frequency summed over 28 questions (excluding potatoes) in the FFQ (Appendix S1) and grouped for those answering at least 60 of the FF questions into the following quartiles: low consumption (0-1.55); below average consumption (1.55-2.53); above average consumption (2.53-3.67); and high consumption (≥ 3.67).
- Number of portions of fruit/d; (product of use and frequency summed over 7 questions in the FFQ (Appendix S1) and grouped for those answering at least 60 of the FFQ into the following quartiles: low consumption (0-0.30); below average consumption (0.30-0.86); above average consumption (0.86-1.65); and high consumption (≥1.65).
- Type of milk used (Q4.1 Appendix S1) recoded into full fat, semiskim milk, skim milk, other (including soya milk), and none.
- Quantity of butter/margarine spread on bread (Q5.4 Appendix S1) categorized as thick spread, medium spread, scrape, and none at all.

In addition, soy intake was estimated from 10 soy-based food questions (FFQ: Appendix S1). Soy intake (number of portions per day) was categorized into none, below median intake (<0.0506) and above median intake (>0.0506). The amount of daidzein and genistein consumed was estimated as described in Appendix S2 for those answering at least 60 FFQ questions (Appendix S1). For daidzein, the amount consumed was grouped into the following quartiles: low intake (0-1.47 µg/d); below average intake (1.47-3.72 µg/d); above average intake (3.72-13.74 µg/d); and high intake (≥13.74 µg/d). For genistein, the following quartiles were used: low intake (0-5.30 µg/d); below average intake (5.30-9.62 µg/d); above average intake (9.62-21.93 µg/d); and high intake (≥21.93 µg/d).

## 2.5 | Assessment of confounders

Confounders found to be significantly related to outcome in earlier analysis of these data<sup>4,5,20</sup> were re-examined here for those completing the dietary questionnaire (see Appendices S3 and S4) and all relating to outcome retained. Age was not related to motility or morphology. The factors retained for motility were ethnic group, testes surgery, manual work, wearing boxer shorts and abstinence (Appendix S3), and for morphology were BMI, cannabis use, season and abstinence (Appendix S4).

# 2.6 | Statistical methods

Analyses of the two outcomes (low motile sperm count and poor morphology) used unconditional logistic regression, accounting for clustering of subjects within the 14 fertility clinics, first without, and then with, adjustment for confounders associated with that outcome. Where more than one dietary factor was associated with the outcome (P < .10), the contribution of each factor was assessed using a Wald statistic. All analyses were carried out using the generalized linear latent and mixed models (gllamm) command within STATA 14.2 (Stata Corporation; Statcorp).

# 2.7 | Ethics

Ethical approval was given by the Multi-Centre Ethics Committee for the North West (ref. no. MREC 98/8/73) with subsequent approval given by the Local Research Ethics Committee at each site.

# 3 | RESULTS

# 3.1 | Study population

A total of 2249 men were recruited and provided a semen sample and 1907 of these (84.8%) returned the dietary questionnaire. Sperm motility was determined for all 1907 men, and 440 were cases (23.1%) with a low MSC (<4.8  $\times$  10<sup>6</sup>/mL). Sperm morphology was determined

# TABLE 1 Dietary factors for low motile sperm count

	Case	Referent		
	N (%)	N (%)	OR (95% CI) <sup>a</sup>	OR <sub>adj</sub> (95%Cl <sub>adj</sub> ) <sup>b</sup>
Type of diet				
Meat (± fish)	426 (97.0)	1415 (96.5)	1	1
Fish only	7 (1.6)	27 (1.8)	0.88 (0.38-2.05)	0.87 (0.36-2.06)
Vegetarian or vegan	6 (1.4)	24 (1.6)	0.84 (0.34-2.07)	0.99 (0.39-2.50)
Frequency of red meat consumption				
≤1/wk	98 (22.7)	326 (22.7)	1	1
>1 ≤ 3 times/wk	185 (42.9)	703 (49.0)	0.87 (0.66-1.16)	0.84 (0.63-1.12)
>3 times/wk	148 (34.3)	407 (28.3)	1.20 (0.89-1.62)	1.13 (0.83-1.53)
Frequency of poultry consumption				
≤1/wk	93 (21.6)	306 (21.3)	1	1
>1 ≤ 3 times/wk	232 (53.8)	810 (56.3)	0.93 (0.71-1.23)	0.92 (0.70-1.23)
>3 times/wk	106 (24.6)	320 (22.3)	1.08 (0.78-1.49)	1.06 (0.76-1.47)
Frequency of fish consumption				
≤1/wk	301 (70.0)	1031 (71.6)	1	1
>1 ≤ 3 times/wk	110 (25.6)	341 (23.7)	1.12 (0.87-1.45)	1.09 (0.84-1.41)
>3 times/wk	19 (4.4)	68 (4.7)	0.97 (0.57-1.64)	0.96 (0.55-1.65)
Portions of vegetables/d				
Low (0-1.55)	102 (26.0)	327 (24.3)	1	1
Below average (1.55-2.53)	118 (30.1)	321 (23.9)	1.18 (0.87-1.60)	1.24 (0.90-1.70)
Above average (2.53-3.67)	91 (23.2)	343 (25.5)	0.86 (0.62-1.19)	0.86 (0.61-1.20)
High (≥ 3.67)	81 (20.7)	354 (26.3)	0.75 (0.54-1.04)	0.78 (0.55-1.09)
Portions of fruit/d				
Low (0-0.30)	101 (25.8)	328 (24.4)	1	1
Below average (0.30-0.86)	95 (24.2)	329 (25.2)	0.90 (0.65-1.24)	0.90 (0.65-1.25)
Above average (0.86-1.65)	95 (24.2)	345 (25.7)	0.89 (0.65-1.23)	0.96 (0.69-1.33)
High (≥1.65)	101 (25.3)	333 (24.8)	0.97 (0.71-1.34)	0.99 (0.72-1.38)
Type of milk used				
Full fat	108 (24.7)	288 (19.7)	1	1
Semi-skim milk	256 (58.4)	945 (64.7)	0.72 (0.55-0.93)	0.77 (0.58-1.01)
Skim milk	53 (12.1)	179 (12.3)	0.81 (0.55-1.18)	0.89 (0.60-1.33)
Other	4 (0.9)	19 (1.3)	0.59 (0.19-1.78)	0.79 (0.26-2.44)
None	17 (3.9)	29 (2.0)	1.58 (0.83-3.00)	1.71 (0.88-3.31)
Quantity of butter/margarine used on b	oread			
Thick spread	25 (5.7)	91 (6.3)	1	1
Medium spread	236 (54.3)	836 (57.5)	1.01 (0.63-1.61)	1.02 (0.63-1.65)
Scrape	130 (29.9)	452 (31.1)	1.02 (0.63-1.67)	1.02 (0.62-1.68)
None	44 (10.1)	74 (5.1)	2.16 (1.21-3.87)	2.32 (1.28-4.23)

<sup>a</sup>Adjusted for clustering within centre.

<sup>b</sup>Adjusted for clustering and ethnicity, surgery, work, boxer shorts and abstinence

in 1673 men who had returned a dietary questionnaire, and of these, 274 men were cases (16.4%) who had poor sperm morphology (<4% normal forms). There was no relation between case status and likelihood of returning the questionnaire but those that did so were more likely to be older and of white ethnicity (Appendix S5).

# 3.2 | Risk factors for low MSC

In the fully adjusted model (adjusting for clustering and ethnicity, surgery, work, boxer shorts and abstinence: Appendix S3), low MSC was not positively associated with any dietary factor other than a report of no spread used on bread. Before adjustment for clustering and confounding, there was a trend (P = .017) towards lower risk with the consumption of more vegetables but this was no longer significant after adjustment (Table 1). After adjusting for confounders and clustering within centres, low MSC was inversely associated with above median intake of soy (OR = 0.75, 95%Cl 0.56-1.00), high daidzein intake (OR 0.58, 95%Cl (0.42-0.82) and high genistein intake (OR 0.60 95%Cl 0.43-0.83; Table 2). In a multivariable regression, multilevel consumption of soy (P = .85) and genistein (P = .73) did not add significantly to a model including daidzein. In a combined model of daidzein and spread, the OR (95%Cl) for high daidzein intake (>13.74) was 0.57 (0.41-0.81) and that of no spread 2.39 (1.56-3.67).

#### 3.3 | Risk factors for PM

After adjustment for clustering, body mass index, cannabis use, abstinence and season (see Appendix S4), poor sperm morphology was positively associated with use of semi-skimmed milk and inversely associated with consumption of red meat more than three times per week (Table 3). When all other milk types and no milk (with similar odds ratios in Table 3) were grouped together and contrasted with whole milk consumption, whole full fat milk was found to be protective (OR = 0.67 95%CI 0.47-0.96: Table 4) and associated with good morphology. When red meat and whole full fat milk were entered into the same model, both retained their protective effect but this was reduced slightly for each factor individually as the two were correlated: those who ate red meat 3 or more times a week were also more likely to drink whole milk. The interaction between them was not significant in a fully adjusted model (last column, Table 4).

**TABLE 2**Soy and phytoestrogenintake with low motile sperm count

-WILEY-

Consumption of soy, genistein and daidzein was not associated with poor morphology (Table 5).

# 4 | DISCUSSION

Dietary risk factors for male subfertility are potentially modifiable and hence of special interest. In this case-referent study of a male population attending an infertility clinic for fertility diagnosis, we have identified that being a case with a low MSC is inversely associated with daidzein intake. In contrast, daidzein intake was not associated with poor morphology but eating red milk and drinking whole milk were protective. These results suggest that dietary risk factors for low MSC and PM differ and that changes to a diet at an individual level, depending upon their type of subfertility, might improve semen quality.

The results of this study are indicative of a protective effect of daidzein (or genistein) on motile sperm count (but not morphology). This effect could potentially be attributed to their interactions with oestrogen receptors<sup>26</sup> but the potential mode of action is difficult to identify as isoflavones have a wider range of cellular effects that can be considered either protective, for example, antioxidant activity<sup>27</sup> or increased androgen levels<sup>28</sup> or detrimental, for example, increasing capacitation<sup>29</sup> and altering neonatal testicular development.<sup>30</sup> Our results initially appear to contradict previous smaller studies that have reported phytoestrogen intake reduces semen quality (eg 17). However, certain Asian populations consume high levels of soy products with little apparent effect on male fertility<sup>31</sup> and soy food intake has not been associated with IVF treatment outcomes.<sup>32</sup> Consistent with our results, daily consumption of a soy product reportedly increased semen quality in a man with oligozoospermia<sup>33</sup> but a larger trial, over 6 months, of 14

	Case	Referent		
	N (%)	N (%)	OR (95% CI) <sup>a</sup>	OR <sub>adj</sub> (95%Cl <sub>adj</sub> ) <sup>b</sup>
oy (Intake/d)				
None	201 (51.3)	617 (45.9)	1	1
Below median (<0.0506)	99 (25.3)	354 (26.3)	0.87 (0.66-1.15)	0.92 (0.69-1.22)
Above median (≥0.0506)	92 (23.5)	374 (27.8)	0.76 (0.58-1.01)	0.75 (0.56-1.00)
stimated daidzein Intake (μg/d	)			
Low (0-1.47)	119 (30.4)	315 (23.4)	1	1
Below average (1.47-3.72)	98 (25.0)	336 (25.0)	0.79 (0.58-1.07)	0.80 (0.58-1.11)
Above average (3.72-13.74)	97 (24.7)	338 (25.1)	0.78 (0.57-1.06)	0.81 (0.59-1.12)
High (≥ 13.74)	78 (19.9)	356 (26.5)	0.59 (0.43-0.82)	0.58 (0.42-0.82)
stimated genistein Intake (µg/	d)			
Low (0-5.30)	120 (30.6)	314 (23.3)	1	1
Below average (5.30-9.62)	98 (25.0)	336 (25.0)	0.76 (0.56-1.04)	0.82 (0.60-1.13)
Above average (9.62-21.93)	93 (23.7)	342 (25.4)	0.71 (0.52-0.97)	0.75 (0.50-1.03)
High (≥21.93)	81 (20.7)	353 (26.2)	0.60 (0.44-0.83)	0.60 (0.43-0.83)

<sup>a</sup>Adjusted for clustering within centre.

<sup>b</sup>Adjusted for clustering and ethnicity, surgery, work, boxer shorts and abstinence.

	Case	Referent			
Dietary factor	N (%)	N (%)	OR (95% CI) <sup>a</sup>	OR <sub>adj</sub> (95% Cl <sub>adj</sub> ) <sup>b</sup>	
Type of diet					
Meat ( $\pm$ fish)	263 (6.0)	1350 (96.6)	1	1	
Fish only	6 (2.2)	25 (1.8)	1.25 (0.50-3.10)	0.96 (0.38-2.43)	
Vegetarian or vegan	5 (1.8)	23 (1.6)	1.07 (0.40-2.86)	1.09 (0.40-2.96)	
Frequency of red meat consu	Frequency of red meat consumption				
≤1/wk	70 (25.8)	300 (21.9)	1	1	
>1 ≤ 3 times/wk	133 (49.1)	652 (47.5)	0.89 (0.64-1.23)	0.87 (0.63-1.22)	
>3 times/wk	68 (25.1)	420 (30.6)	0.71 (0.49-1.03)	0.67 (0.46-0.99)	
Frequency of poultry consum	ption				
≤1/wk	73 (26.9)	282 (20.6)	1	1	
>1 ≤ 3 times/wk	142 (52.4)	771 (56.2)	0.71 (0.52-0.97)	0.75 (0.54-1.03)	
>3 times/wk	56 (20.7)	319 (23.3)	0.68 (0.46-0.99)	0.73 (0.49-1.08)	
Frequency of fish consumptio	'n				
≤1/wk	184 (68.4)	980 (71.4)	1	1	
>1 ≤ 3 times/wk	74 (27.5)	328 (23.9)	1.19 (0.89-1.61)	1.14 (0.84-1.54)	
>3 times/wk	11 (4.1)	65 (4.7)	0.90 (0.46-1.74)	0.85 (0.43-1.67)	
Portions of vegetables/d					
Low (0-1.55)	66 (24.9)	305 (24.0)	1	1	
Below average (1.55-2.53)	70 (27.2)	317 (25.0)	1.05 (0.72-1.52)	1.04 (0.71-1.52)	
Above average (2.53-3.67)	58 (22.6)	331 (26.1)	0.82 (0.55-1.21)	0.77 (0.52-1.15)	
High (≥3.67)	65 (25.3)	317 (25.0)	0.96 (0.65-1.41)	0.90 (0.61-1.33)	
Portions of fruit/d					
Low (0-0.30)	68 (26.5)	312 (24.6)	1	1	
Below average (0.30-0.86)	64 (24.9)	317 (25.0)	0.93 (0.64-1.35)	0.91 (0.62-1.33)	
Above average (0.86-1.65)	55 (21.4)	331 (26.1)	0.76 (0.52-1.12)	0.76 (0.51-1.13)	
High (≥1.65)	70 (27.2)	310 (24.4)	1.03 (0.71-1.49)	1.02 (0.70-1.49)	
Type of milk used					
Full fat	45 (16.4)	290 (20.8)	1	1	
Semi-skim milk	181 (66.1)	889 (63.9)	1.34 (0.94-1.90)	1.45 (1.01-2.09)	
Skim milk	37 (13.5)	165 (11.9)	1.48 (0.91-2.39)	1.61 (0.98-2.63)	
Other	3 (1.1)	15 (1.1)	1.28 (0.35-4.65)	1.40 (0.38-5.24)	
None	8 (2.9)	32 (2.3)	1.65 (0.71-3.82)	1.85 (0.78-4.38)	
Quantity of margarine/butter used on bread					
Thick spread	13 (4.8)	84 (6.1)	1	1	
Medium spread	153 (58.0)	795 (57.4)	1.20 (0.65-2.22)	1.18 (0.64-2.20)	
Scrape	91 (33.3)	418 (30.2)	1.36 (0.72-2.55)	1.37 (0.72-2.60)	
None	16 (5.9)	88 (6.4)	1.10 (0.49-2.44)	1.03 (0.46-2.32)	

**TABLE 3** Dietary factors for poormorphology

<sup>a</sup>Adjusted for clustering with centres.

<sup>b</sup>Adjusted for clustering and body mass index, cannabis use, abstinence and season

men with normal semen quality reported no alterations in semen quality although plasma daidzein and genistein levels increased.<sup>18</sup>

Dietary recommendations for red and processed meat consumption are controversial.<sup>34</sup> In this study, a protective effect of red meat consumption on sperm morphology was observed, a result consistent with reports that diets characterized by high meat intake are indeed associated with an improvement in certain aspects of semen quality, for example sperm concentration<sup>15</sup> and possibly sperm morphology.<sup>35</sup> However, previous hypotheses have suggested that meat consumption would increase risk of adverse semen quality due to increased consumption of, for example, saturated fat, xenobiotics including xenoestrogens and potentially hormone residues.<sup>36</sup> However, the evidence for such effects is limited with few studies reporting any negative associations between red meat consumption and semen quality.<sup>11,15,35,37-40</sup> **TABLE 4**Relation of red meat, fullfat milk and their interaction to poormorphology

# –<mark>andrology ⊜ </mark>–Wiley<sup>⊥</sup><sup>7</sup>

	OR (95% CI)				
	Unadjusted	Model 1 <sup>a</sup>	Model 2ª		
Red meat					
<3 times/wk	1	1	1		
≥3 times/wk	0.74 (0.54-1.00)	0.71 (0.49-1.01)	0.69 (0.49-0.95)		
Type of milk use					
All other	1	1	1		
Full fat	0.67 (0.45-0.95)	0.75 (0.55-1.02)	0.62 (0.40-0.96)		
Interaction between red meat and full fat milk					
None	-	-	1		
Both	_	_	1.50 (0.71-3.19)		

<sup>a</sup>Adjusted for confounders and clusters within centre

# **TABLE 5** Soy and phytoestrogen intake with poor morphology

	Case	Referent		
	N (%)	N (%)	OR (95% CI) <sup>a</sup>	OR <sub>adj</sub> (95%Cl <sub>adj</sub> ) <sup>b</sup>
Soy (Intake/d)				
None	121 (47.1)	596 (46.9)	1	1
Below medium (<0.0506)	68 (26.5)	336 (26.5)	1.01 (0.73-1.40)	0.98 (0.70-1.37)
Above medium (≥0.0506)	68 (26.5)	338 (26.6)	1.00 (0.72-1.39)	0.91 (0.65-1.28)
Estimated daidzein Intake (µg/	d)			
Low (0-1.47)	70 (27.2)	300 (23.6)	1	1
Below average (1.47-3.72)	60 (23.3)	327 (25.7)	0.77 (0.53-1.13)	0.77 (0.53-1.14)
Above average (3.72-13.74)	57 (22.2)	337 (26.5)	0.72 (0.49-1.06)	0.73 (0.49-1.08)
High (≥13.74)	70 (27.2)	306 (24.1)	0.98 (0.67-1.41)	0.90 (0.62-1.32)
Estimated genistein Intake (µg/d)				
Low (0-5.30)	65 (25.3)	308 (24.3)	1	1
Below average (5.30-9.62)	64 (24.9)	318 (25.0)	0.95 (0.65-1.40)	0.94 (0.64-1.38)
Above average (9.62-21.93)	59 (23.0)	338 (26.6)	0.83 (0.56-1.21)	0.81 (0.54-1.19)
High (≥21.93)	69 (26.9)	306 (24.1)	1.07 (0.73-1.56)	0.97 (0.66-1.43)

<sup>a</sup>Adjusted for clustering with centres.

<sup>b</sup>Adjusted for clustering and body mass index, cannabis use, abstinence and season.

In this study, poor sperm morphology was associated with the lack of milk consumption and also the intake of skimmed or semiskimmed milk, suggesting that whole milk is protective of semen quality. Previously, a high intake of omega-3 polyunsaturated fats has been reported to be positively associated with sperm morphology<sup>41</sup> and fish oil supplements with semen volume and total sperm count.<sup>42</sup> Such results are consistent with the presence, in whole milk, of numerous fat-soluble vitamins and other components that might affect semen quality, for example, by providing protection against oxidative stress.<sup>43</sup> These would be lost upon processing to semi skimmed and skimmed milk.<sup>44,45</sup> Other studies, of smaller populations, have reported positive associations between consumption of low-fat milk and semen quality.<sup>38,39</sup>

There are a number of strengths associated with this study. Firstly, the number of men providing dietary information via a FFQ is much larger than many other previously published studies resulting in a study of sufficient power (80%) to detect an OR of 2 assuming 1 in 40 controls would be exposed, and a 2:1 ratio of referents to cases. The second strength of the study is that those men who knew the results of the assessment of their semen quality were excluded from the study. Such prior knowledge of their results could have biased their answers to specific questions or even modified behaviours prior to recruitment. Furthermore, semen analysis was carried out according to WHO protocols, with CASA being undertaken at each recruitment site but analysed centrally to ensure consistency in semen analysis. Case definitions were also as used previously<sup>5,20</sup> and defined a priori according to WHO definitions whereas other studies have used a wide variety of different measures of semen quality of unknown clinical relevance.

However, there are certain limitations to this work. Firstly, the study population was men attending infertility clinics as part of a couple and so potentially they might not be representative of all -WILFY-ANDROLOGY 📾 🕮

men. In addition, not all eligible men were recruited to the study and the reasons why they did not want to participate are not known. It is possible that they did want to be asked about lifestyle factors (including diet). In addition to the recognized limitations of using FFQs to assess food consumption, which in this study was over a 12month period because of the sustained period of infertility required to be eligible for the study, there are potential issues regarding the questionnaire coverage of phytoestrogen containing foods and also the accuracy of the levels of phytoestrogens in the covered dietary items.<sup>46</sup> The food items and phytoestrogen values used here are not those of more recent studies (eg 47-49). These issues are likely to be independent of case status and so result in non-differential bias, which is normally associated with a reduced estimate of effect size ('bias towards the null'). Assessment of phytoestrogen intake may be improved by urinary measurement of specific phytoestrogen metabolites: however, inconsistent results linking urinary daidzein and semen quality have also been reported.<sup>50,51</sup> Participants were also not asked about the intake of dietary supplements, and while there is little evidence that soy-based [18] or protein-based<sup>52</sup> supplements may alter male semen quality, other dietary supplements may improve male semen guality<sup>53</sup> and potentially interact with soy-based products to alter semen quality. Semen quality measures do not fully correlate with male fertility, and these can explain why dietary food groups associated with semen quality are not necessarily associated with time to pregnancy or other measures of reproductive success. Furthermore, urinary daidzein levels in men were not associated with time to pregnancy.<sup>54</sup> It has also been suggested that pre-pubertal exposures may be important but we were not able to study them.

In conclusion, this large multi-centre case-control study identified different risk factors for low motile sperm count and poor sperm morphology. Further work is required to confirm these associations but suggests that semen quality may be improved by targeted interventions.

# ACKNOWLEDGEMENTS

The study was designed and initiated with the support of Professor lan Cooke of the University of Sheffield. Participating centres were as follows: Department of Obstetrics and Gynaecology, Queens University, Belfast; Assisted Conception Unit, Birmingham Women's Hospital; Division of Obstetrics and Gynaecology, St Michael's Hospital, Bristol; Directorate of Women's Health, Southmead Hospital, Bristol; Cardiff Assisted Reproduction Unit, University of Wales; MRC Reproductive Biology Unit, Edinburgh; Reproductive Medicine Unit, Liverpool Women's Hospital; St Bartholomew's Hospital, London; Department of Obstetrics and Gynaecology, Royal Free and University College, London; Department of Reproductive Medicine, St Mary's Hospital, Manchester; IVF/Immunology Laboratory, Hope Hospital, Salford; Department of Histopathology, Wythenshawe Hospital, Manchester; International Centre for Life, Newcastle; Department of Obstetrics and Gynaecology, Jessop Hospital for Women, Sheffield; and Shropshire and Mid-Wales Fertility Centre, Royal Shrewsbury NHS Trust.

We are greatly indebted to the teams at each centre, which included clinicians, nursing and technical staff, for their contributions to the study. The research was made possible by the input of clinicians and scientists at the participating centres including Sheena Lewis, Neil McClure (Belfast), Masoud Afnan (Birmingham), Chris Ford (Bristol), Lucas Klentzeris, Linda Gregory (Cardiff), Stewart Irvine (Edinburgh), Iwan Lewis-Jones (Liverpool), Brian Leiberman, David Poulson and Paul Bishop (Manchester), Adrian Lower and Melanie Davies (London), Alison Murdoch (Newcastle), Jason Kasraie (Shrewsbury) and William Ledger (Sheffield). We also wish to thank the research coordinators, technical and administrative staff including Yvonne King, Priscilla Appelbe, Chris Pappas, Deborah Saxton, Ana Roby, Jeanette Tenggren, Mark Carus, Kelly Morris, Louise Reeve, Linda Street, Andrew Thomas and Hayley Willis. The authors also wish to thank Jennie Pollard for initial support in the development of the FFQ and for data entry and Sinead Boylan for other work on phytoestrogens.

## CONFLICT OF INTEREST

Professor JE Cade is a Director of the University of Leeds company, Dietary Assessment Limited.

#### AUTHOR CONTRIBUTIONS

All members of the co-ordinating group contributed to the collection of data for the study and discussions on the design, conduct and interpretation of the results. Data management was done by J-AC, HB, NC and HM. ACP, NC, JC and AAP drafted the manuscript, and NC and ACP discussed and performed the statistical analysis. JC oversaw the design and analysis of the dietary questionnaire. All authors critically revised the paper and approved the submitted version. J-AC and HB co-ordinated the study and took care of communication and distribution of study materials to group members.

# ORCID

Andy C. Povey (D) https://orcid.org/0000-0002-9127-8328

#### REFERENCES

- Pan MM, Hockenberry MS, Kirby EW, Lipshultz LI. Male infertility diagnosis and treatment in the era of in vitro fertilization and intracytoplasmic sperm injection. *Med Clin North Am.* 2018;102:337-347.
- 2. Levine H, Jørgensen N, Martino-Andrade A, et al. Temporal trends in sperm count: a systematic review and meta-regression analysis. *Hum Reprod Update*. 2017;23:646-659.
- Ravanos K, Petousis S, Margioula-Siarkou C, et al. Declining sperm counts... or rather not? A mini review. Obstet Gynecol Surv. 2018;73:595-605.
- Povey AC, Clyma JA, McNamee R, et al. Modifiable and non-modifiable risk factors for poor semen quality: a case-referent study. *Hum Reprod.* 2012;27:2799-2806.
- Pacey AA, Povey AC, Clyma JA, et al. Modifiable and non-modifiable risk factors for poor sperm morphology. *Hum Reprod.* 2014;29:1629-1636.

- Database Syst Rev. 2019;3:CD007411.
  7. Nassan FL, Chavarro JE, Tanrikut C. Diet and men's fertility: does diet affect sperm quality? *Fertil Steril*. 2018;110:570-577.
- Salas-Huetos A, James ER, Aston KI, Jenkins TG, Carrell DT. Diet and sperm quality: nutrients, foods and dietary patterns. *Reprod Biol.* 2019;19:219-224.
- Salas-Huetos A, Bulló M, Salas-Salvadó J. Dietary patterns, foods and nutrients in male fertility parameters and fecundability: a systematic review of observational studies. *Hum Reprod Update*. 2017;23:371-389.
- 10. Liu CY, Chou YC, Chao JC, Hsu CY, Cha TL, Tsao CW. The Association between dietary patterns and semen quality in a general Asian population of 7282 males. *PLoS One.* 2015;10:e0134224.
- Gaskins AJ, Colaci DS, Mendiola J, Swan SH, Chavarro JE. Dietary patterns and semen quality in young men. *Hum Reprod*. 2012;27:2899-2907.
- Cutillas-Tolín A, Mínguez-Alarcón L, Mendiola J, et al. Mediterranean and western dietary patterns are related to markers of testicular function among healthy men. *Hum Reprod.* 2015;30:2945-2955.
- Nassan FL, Jensen TK, Priskorn L, Halldorsson TI, Chavarro JE, Jørgensen N. Association of dietary patterns with testicular function in young Danish men. JAMA Netw Open. 2020;3(2):e1921610.
- Jurewicz J, Radwan M, Sobala W, Radwan P, Bochenek M, Hanke W. Dietary patterns and their relationship with semen quality. *Am J Mens Health.* 2018;12:575-583.
- Vujkovic M, de Vries JH, Dohle GR, et al. Associations between dietary patterns and semen quality in men undergoing IVF/ICSI treatment. *Hum Reprod.* 2009;24:1304-1312.
- Cederroth CR, Auger J, Zimmermann C, Eustache F, Nef S. Soy, phyto-oestrogens and male reproductive function: a review. Int J Androl. 2010;33:304-316.
- 17. Chavarro JE, Toth TL, Sadio SM, Hauser R. Soy food and isoflavone intake in relation to semen quality parameters among men from an infertility clinic. *Hum Reprod*. 2008;23:2584-2590.
- Mitchell JH, Cawood E, Kinniburgh D, Provan A, Collins AR, Irvine DS. Effect of a phytoestrogen food supplement on reproductive health in normal males. *Clin Sci (Lond)*. 2001;100:613-618.
- National Institute for Health and Care Excellence. Fertility problems: assessment and treatment. Clinical guideline. 2013. https:// www.nice.org.uk/guidance/cg156. Accessed March 04, 2020
- Cherry N, Moore H, McNamee R, et al. Occupation and male infertility: glycol ethers and other exposures. *Occup Environ Med.* 2008;65:708-714.
- World Health Organization. WHO Laboratory Manual for the Examination of Human Semen and Sperm-Cervical Mucus Interaction, 4th edn. Cambridge, UK: Cambridge University Press; 1999.
- World Health Organization. WHO Laboratory Manual for the Examination and Processing of Human Semen, 5th edn. Geneva: World Health Organization; 2010.
- 23. Reinli K, Block G. Phytoestrogen content of foods-a compendium of literature values. *Nutr Cancer*. 1996;26:123-148.
- 24. Thompson LU, Robb P, Serraino M, Cheung F. Mammalian lignan production from various foods. *Nutr Cancer*. 1991;16:43-52.
- Liggins J, Bluck LJ, Runswick S, Atkinson C, Coward WA, Bingham SA. Daidzein and Genistein contents of vegetables. Br J Nutr. 2000;84:717-725.
- Rietjens IM, Sotoca AM, Vervoort J, Louisse J. Mechanisms underlying the dualistic mode of action of major soy isoflavones in relation to cell proliferation and cancer risks. *Mol Nutr Food Res.* 2013;57:100-113.
- Chadha R, Bhalla Y, Jain A, Chadha K, Karan M. Dietary soy isoflavone: a mechanistic insight. Nat Prod Commun. 2017;12:627-634.

- McVey MJ, Cooke GM, Curran IHA. Increased serum and testicular androgen levels in F1 rats with lifetime exposure to soy isoflavones. *Reprod Toxicol.* 2004;18:677-685.
- Fraser LR, Beyret E, Milligan SR, Adeoya-Osiguwa SA. Effects of estrogenic xenobiotics on human and mouse spermatozoa. *Hum Reprod.* 2006;21:1184-1193.
- Napier ID, Simon L, Perry D, et al. Testicular development in male rats is sensitive to a soy-based diet in the neonatal period. *Biol Reprod.* 2014;90:40.
- Messina M. Soybean isoflavone exposure does not have feminizing effects on men: a critical examination of the evidence. *Fertil Steril.* 2010;93:2095-2104.
- Mínguez-Alarcón L, Afeiche MC, Chiu YH, et al. Male soy food intake was not associated with in vitro fertilization outcomes among couples attending a fertility center. *Andrology*. 2015;3:702-708.
- Casini ML, Gerli S, Unfer V. An infertile couple suffering from oligospermia by partial sperm maturation arrest: can phytoestrogens play a therapeutic role? A case report study. *Gynecol Endocrinol*. 2006;22:399-401.
- Johnston BC, Zeraatkar D, Han MA, et al. Unprocessed Red meat and processed meat consumption: dietary guideline recommendations from the nutritional recommendations (NutriRECS) Consortium. Ann Intern Med. 2019;171:756-764.
- Afeiche MC, Williams PL, Gaskins AJ, et al. Meat intake and reproductive parameters among young men. *Epidemiology*. 2014;25:323-330.
- Andersson AM, Skakkebaek NE. Exposure to exogenous estrogens in food: possible impact on human development and health. Eur J Endocrinol. 1999;140:477-485.
- Braga DP, Halpern G, Figueira Rde C, Setti AS, laconelli A Jr, Borges E Jr. Food intake and social habits in male patients and its relationship to intracytoplasmic sperm injection outcomes. *Fertil Steril.* 2012;97:53-59.
- Eslamian G, Amirjannati N, Rashidkhani B, Sadeghi M-R, Hekmatdoost A. Intake pf food groups and idiopathic asthenozoospermia: a case-control study. *Hum Reprod*. 2012;11:3328-3336.
- Afeiche MC, Gaskins AJ, Williams PL, et al. Processed meat intake is unfavorably and fish intake favorably associated with semen quality indicators among men attending a fertility clinic. J Nutr. 2014;144:1091-1098.
- Maldonado-Cárceles AB, Mínguez-Alarcón L, Mendiola J, et al. Meat intake in relation to semen quality and reproductive hormone levels among young men in Spain. Br J Nutr. 2019;121:451-460.
- Attaman JA, Toth TL, Furtado J, Campos H, Hauser R, Chavarro JE. Dietary fat and semen quality among men attending a fertility clinic. *Hum Reprod.* 2012;27:1466-1474.
- 42. Jensen TK, Priskorn L, Holmboe SA, et al. Associations of fish oil supplement use with testicular function in young men. JAMA Netw Open. 2020;3:e1919462.
- 43. Wong WY, Thomas CMG, Merkus JMWM, Zielhuis GA, Steegers-Theunissen RPM. Male factor subfertility: possible causes and the impact of nutritional factors. *Fertil Steril*. 2000;73:4335-4342.
- 44. Pereira PC. Milk nutritional composition and its role in human health. *Nutrition*. 2014;30:619-627.
- 45. Vargaas-Bell-Perez E, Toro-Mujica P, Enriquez-Hidalgo D, Fellenberg MA, Gomez-Cortes P. Discrimination between retail bovine milks with different fat contents using chemometrics and fatty acid profiling. J Dairy Sci. 2017;100:4253-4257.
- Schwartz H, Sontag G, Plumb J. Inventory of phytoestrogen databases. Food Chem. 2009;113:736-747.
- Carmichael SL, Gonzalez-Feliciano AG, Ma C, Shaw GM, Cogswell ME. Estimated dietary phytoestrogen intake and major food sources among women during the year before pregnancy. *Nutr J*. 2011;10:105.

ILEY- ANDROLOGY 📾 😫

- Clarke DB, Lloyd AS, Lawrence JM, et al. Development of a food compositional database for the estimation of dietary intake of phyto-oestrogens in a group of postmenopausal women previously treated for breast cancer and validation with urinary excretion. *Brit J Nutr.* 2013;109:2261-2268.
- Lee A, Beaubernard L, Lamothe V, Bennetau-Pelissero C. New evaluation of isoflavone exposure in the French population. *Nutrients*. 2019;11:2308.
- Xia Y, Chen M, Zhu P, et al. Urinary phytoestrogen levels related to idiopathic male infertility in Chinese men. *Environ Int.* 2013;59:161-167.
- Qin Y, Du G, Chen M, et al. Combined effects of urinary phytoestrogens metabolites and polymorphisms in metabolic enzyme gene on idiopathic male infertility. *Arch Toxicol*. 2014;88:1527-1536.
- Tøttenborg SS, Glazer CH, Hærvig KK, et al. Semen quality among young healthy men taking protein supplements. *Fertil Steril.* 2020;114(1):89-96.
- Salas-Huetos A, Rosique-Esteban N, Becerra-Tomás N, Vizmanos B, Bulló M, Salas-Salvadó J. The effect of nutrients and dietary supplements on sperm quality parameters: a systematic

review and meta-analysis of randomized clinical trials. Adv Nutr. 2018;9:833-848.

54. Mumford SL, Sundaram R, Schisterman EF, et al. Higher urinary lignan concentrations in women but not men are positively associated with shorter time to pregnancy. *J Nutr.* 2014;144:352-358.

# SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Povey AC, Clyma J-A, McNamee R, et al; Participating Centres of Chaps-UK. Phytoestrogen intake and other dietary risk factors for low motile sperm count and poor sperm morphology. *Andrology*. 2020;00:1–10. https://doi.org/10.1111/andr.12858