
Article:
Huang, Yong, Hussain, Muhammad Azhar, Luo, Dan et al. (10 more authors) (2020) A Brassica napus Reductase Gene Dissected by Associative Transcriptomics Enhances Plant Adaption to Freezing Stress. Frontiers in Plant Science. 971. ISSN 1664-462X

https://doi.org/10.3389/fpls.2020.00971

A Brassica napus reductase gene dissected by associative transcriptomics enhances plant adaption to freezing stress

Yong Huang[1], Muhammad A. Hussain[1], Dan Luo[1], Hongzhi Xu[2], Cuan Zeng[2], Lenka Havlickova[3], Ian Bancroft[3], Zhitao Tian[1], Xuekun Zhang[1], Yong Cheng[1], Xiling Zou[1], Guangyuan Lu[1*], Yan Lv[1*]

[1]Oil Crops Research Institute, Chinese Academy of Agricultural Sciences, China, [2]Chongqing Three Gorges Academy of Agricultural Sciences, China, [3]University of York, United Kingdom

Submitted to Journal:
Frontiers in Plant Science

Specialty Section:
Plant Abiotic Stress

Article type:
Original Research Article

Manuscript ID:
551620

Received on:
14 Apr 2020

Revised on:
12 Jun 2020

Frontiers website link:
www.frontiersin.org

Conflict of interest statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest

Author contribution statement

Figures, study design:Yong Huang, Muhammad Azhar Hussain;
Investigation and data collection: Yong Huang; Hongzhi Xu; Chuan Zeng;
Data analysis: Guangyuan Lu, Zhitao Tian, LenkaHavlickova, Ian Bancroft;
Validation: Dan Luo;
Resources: LenkaHavlickova, Ian Bancroft;
Writing-original draft: Yong Huang, Muhammad Azhar Hussain;
Writing-review&editing: Yan Lv, Guangyuan Lu;
Funding acquisition: Xuekun Zhang, Yong Cheng, Xiling Zou, Guangyuan Lu, Yan Lv;

Keywords

alkaloid 3, rapeseed, tropinone reductase, Photosynthetic gas exchange parameter, Associative Transcriptomics

Abstract

Word count: 216

Cold treatment (vernalization) is required for winter crops such as rapeseed (Brassica napus L.). However, excessive exposure to low temperature (LT) in winter is also a stress for the semi-winter, early-flowering rapeseed varieties widely cultivated in China. Photosynthetic efficiency is one of the key determinants and thus a good indicator for LT tolerance in plants. So far, the genetic basis underlying photosynthetic efficiency is poorly understood in rapeseed. Here the current study used Associative Transcriptomics to identify genetic loci controlling photosynthetic gas exchange parameters in a diversity panel comprising 123 accessions. A total of 201 significant Single Nucleotide Polymorphisms (SNPs) and 147 Gene Expression Markers (GEMs) were detected, leading to the identification of 22 candidate genes. Of these, Cab026133.1, an orthologue of the Arabidopsis gene AT2G29300.2 encoding a tropinone reductase (BnTR1), was further confirmed to be closely linked to transpiration rate. Ectopic expressing BnTR1 in Arabidopsis plants significantly increased the transpiration rate and enhanced LT tolerance under freezing conditions. Also, a much higher level of alkaloids content was observed in the transgenic Arabidopsis plants, which could help protect against LT stress. Together, the current study showed that AT is an effective approach for dissecting LT tolerance trait in rapeseed and that BnTR1 is a good target gene for the genetic improvement of LT tolerance in plant.

Contribution to the field

The genetic basis underlying the adaptation to unfavourable climate is limited in Brassica napus. In the present study, we performed Associative Transcriptomics (AT) analysis on photosynthetic gas exchange parameters in a panel of 123 late-sown Brassica napus accessions. Hundreds of candidate CDSs significantly associated with target traits were identified in this study. Then one of the candidate genes encoding tropinone reductase in Brassica napus (BnTR1), was functionally validated in Arabidopsis; ectopic expressing BnTR1 could enhance the transpiration rate/evapotranspiration value and confer freezing tolerance in Arabidopsis plants. To the best of our knowledge, this is the first study to use AT approach for the identification of genes responsible for photosynthetic related traits under chilling stress conditions. We believe that this manuscript would be of interest to the readers of Frontiers in Plant Science, because our findings indicate that AT approach provides an opportunity for genetic dissection of complex traits such as ecological adaptation in crops.

Funding statement

This research was financially supported by funds from the National Key Research and Development Program of China (2018YFD0100905), the Oil Crops Research Institution Basal Research fund of the Chinese Academy of Agricultural Sciences (CAAS), China (1610172018010), the major science and technology project, Ministry of science and technology, China (2018ZX08020001).

Ethics statements

Studies involving animal subjects
Generated Statement: No animal studies are presented in this manuscript.

Studies involving human subjects
Generated Statement: No human studies are presented in this manuscript.

Inclusion of identifiable human data
Generated Statement: No potentially identifiable human images or data is presented in this study.

Data availability statement

Generated Statement: The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/supplementary material.

A *Brassica napus* reductase gene dissected by associative transcriptomics enhances plant adaption to freezing stress

Yong Huang[1,2,4], Muhammad Azhar Hussain[1,4], Dan Luo[1], Hongzhi Xu[2], Cuan Zeng[2], Lenka Havlickova[3], Ian Bancroft[3], Zhitao Tian[1], Xuekun Zhang[1], Yong Cheng[1], Xiling Zou[1], Guangyuan Lu[1*], Yan Lv[1*]

[1]Key Laboratory of Biology and Genetic Improvement of Oil Crops, Ministry of Agriculture, Oil Crops Research Institute of the Chinese Academy of Agricultural Sciences, Wuhan 430062, China.

[2]The Chongqing Three Gorges Academy of Agricultural Sciences, Chongqing, China.

[3]Centre for Novel Agricultural Products (CNAP) M119, Department of Biology, University of York, York, United Kingdom.

[4]These authors contributed equally to this work.

*Correspondence: lvyan322@163.com; luguangyuan@caas.cn

Abstract： Cold treatment (vernalization) is required for winter crops such as rapeseed (*Brassica napus* L.). However, excessive exposure to low temperature (LT) in winter is also a stress for the semi-winter, early-flowering rapeseed varieties widely cultivated in China. Photosynthetic efficiency is one of the key determinants and thus a good indicator for LT tolerance in plants. So far, the genetic basis underlying photosynthetic efficiency is poorly understood in rapeseed. Here the current study used Associative Transcriptomics to identify genetic loci controlling photosynthetic gas exchange parameters in a diversity panel comprising 123 accessions. A total of 201 significant Single Nucleotide Polymorphisms (SNPs) and 147 Gene Expression Markers (GEMs) were detected, leading to the identification of 22 candidate genes. Of these, Cab026133.1, an orthologue of the *Arabidopsis* gene AT2G29300.2 encoding a tropinone reductase (*BnTR1*), was further confirmed to be closely linked to transpiration rate. Ectopic expressing *BnTR1* in *Arabidopsis* plants significantly increased the transpiration rate and enhanced LT tolerance under freezing conditions. Also, a much higher level of alkaloids content was observed in the transgenic *Arabidopsis* plants, which could help protect against LT stress. Together, the current study showed that AT is an effective approach for dissecting LT tolerance trait in rapeseed and that *BnTR1* is a good target gene for the genetic improvement of LT tolerance in plant.

Keywords: Rapeseed, Associative Transcriptomics, photosynthetic gas exchange parameter, tropinone reductase, alkaloid.

INTRODUCTION

Rapeseed (*Brassica napus* L.) is one of the major oil crops worldwide, with an average annual cropping area of 35.3 million hectares producing 72.8 million tons of seeds in the past five years (http://www.fao.org/faostat/). Meal cake, the byproduct of rapeseed is also an important source of protein-rich feed for livestock (Wanasundara et al., 2016). Due to the agronomic importance of this oil crop, there is a great interest to boost its yield via genetic improvement of major agronomic traits.

The winter type rapeseed is mainly grown in Europe, which requires strong vernalization and is cold tolerant (O'neill et al., 2019). However, the semi-winter type rapeseed grown in China only needs moderate or weak vernalization, and excessive exposure to low temperature (LT) stress in winter will lead to plant damage at vegetative stage and finally cause yield loss (Liao and Guan, 2001; O'neill et al., 2019; Zhang et al., 2015). Yangtze River basin is the major area for growing semi-winter rapeseed, which accounts for at least 80% of the nation's total production (Tian et al., 2018). The rapeseed is usually sown in early October shortly after the harvest of rice in this area (Cong et al., 2019). However, in recent years, the delay of rice harvest usually lead to the postpone of rapeseed sowing until late October or early November, which results in poor germination and seedling establishment due to LT (Luo et al., 2019). The biomass of rapeseed seedling is also significantly reduced at overwintering stage, and thus is more susceptible to LT stresses, i.e. chilling (0–15°C) or freezing (<0°C) (Sage and Kubien, 2007; Zhang et al., 2012; Zhang et al., 2016). Moreover, delay of floral initiation and floral bud differentiation processes (Luo et al., 2018) and decrease of effective pod number, pod length, and seed yield (Ozer, 2003) were observed in the late-sowing rapeseed. To cope with LT stresses, plants have evolved several elaborate regulatory mechanisms; among these, balancing or coordinating the photosynthetic processes could be a critical one (Leister, 2019).

It has been established that light-harvesting complex II (LHCII) proteins in higher plants can facilitate their adaption to external biotic or abiotic environmental stresses such as drought stress or blast fungus infection (Andersson et al., 2003; Ganeteg et al., 2004; Liu et al., 2019b; Xu et al., 2012), and the positive function of

LHCBs in abscisic acid (ABA) mediated signalling pathway is repressed by WARY40 (Liu et al., 2013; Xu et al., 2012). LT stress induces the accumulation of transcript encoding heat-shock proteins (HSPs), and the persistence of HSPs can enhance the chilling tolerance of tomato fruit (Ding et al., 2001; Sabehat et al., 1996). In *Arabidopsis*, HSP21 protects the photosynthetic electron transport chain against the deleterious effects imposed by heat stress (Bernfur et al., 2017; Zhong et al., 2013). However, it is unclear whether HSPs function similarly in rapeseed to alleviate injury from LT stress. The other observations have mechanistically described how the photosynthetic organisms maintain their PSII function under stressful conditions with continuing or fluctuating light (Liu et al., 2019a). For instance, the chloroplast protein HHL1 forms a complex with LQY1 to repair and re-assemble PSII, which in turn helps overcome excessive light stress (Jin et al., 2014). In rapeseed, CBF/DREB transcription factors appear to have important roles in maintaining stronger photosynthetic efficiency and higher Calvin circulating enzyme activity under LT conditions (Savitch et al., 2005).

To date, several genetic studies have been reported for quantitative trait loci (QTLs) mapping of photosynthesis (Basu et al., 2019; De Miguel et al., 2014; Li et al., 2014; Li et al., 2016; Liu et al., 2017; Oakley et al., 2018) and LT tolerance in different plant species (Jha et al., 2017). However, very few QTLs have been identified in *Brassica* species (Ge et al., 2012; Yan et al., 2015). The temperature causing 50% of the maximal damage (LT50) has been regarded as a good index for evaluating LT tolerance (Hincha et al., 1987; Steponkus et al., 1990), which is also significantly correlated with net photosynthesis rate (A_n) in rapeseed (Urban et al., 2013). Therefore, photosynthetic gas exchange parameters such as A_n are a suitable index for the evaluation of LT tolerance in rapeseed that can facilitate the follow-up genetic study.

Photosynthesis plays an indispensable role in ensuring adequate energy supply throughout the plant lifecycle. Therefore, enhancing photosynthetic efficiency has been a commonly adopted strategy for crop yield improvement (Evans, 2013; Lawson et al., 2012; Long et al., 2006). In rice, a new photorespiratory bypass was assembled

by over-expressing *OsGLO3*, *OsOXO3* and *OsCATC* genes in the chloroplast, which resulted in obvious increases in photosynthetic efficiency, biomass and grain yield (Shen et al., 2019). In rapeseed, photosynthetic efficiency has a notable effect on yield, oil content and fatty acid composition (Ju and Li, 2012; Wang et al., 2015). However, the utilization rate of light energy in rapeseed is only 0.615-1.056%, which is much lower than that in rice, wheat or soybean (Zhang et al., 2017a). Therefore, it is possible to further improve the yield by enhancing photosynthetic efficiency in rapeseed.

During the long history of evolution, plants appear to overcome abrupt or mild temperature stresses in winter through a series of changes at molecular, cellular, physiological and biochemical levels (Zhang et al., 2017b). Alkaloid is one of the major secondary metabolites that is inducible under unfavourable conditions, especially drought stress (Selmar and Kleinwachter, 2013). Heavy metals can also promote the accumulation of alkaloid in *Catharanthus roseus* L. (Srivastava and Srivastava, 2010). The short-chain dehydrogenase/reductase (SDR) protein, which belongs to the NAD(P)-binding Rossmann-fold superfamily, functions in the biosynthesis of benzylisiquinoline alkaloids (i.e. morphine, codeine) and tropane-derived alkaloids such as scopalamine, atropine and cocaine. Tropinone reductases (TRs) are a group of SDR proteins which play key roles as a branch point in the biosynthesis pathway of tropane alkaloids (Tonfack et al., 2011). Hence, a study on TRs could help understand how alkaloids function in response to different stresses.

Associative Transcriptomics (AT) strategy, which combines association mapping and transcriptome, has greatly facilitated the genetic dissection of complex traits (Bazakos et al., 2017). Considerable progress has been achieved by AT in allopolyploidy crops such as oilseed rape and wheat, which provided a large number of causative genes or functional markers for molecular marker-assisted breeding. For instance, a transcription factor (HAG1) was identified in rapeseed by AT, which plays an indispensable role in the synthesis of aliphatic glucosinolates (Harper et al., 2012). With AT platform, the genetic studies of many other complex traits in rapeseed were

also reported, including homeostasis of nitrate, phosphate and sulfate anions (Koprivova et al., 2014), calcium and magnesium accumulation (Alcock et al., 2017), lodging resistance (Miller et al., 2018), clubroot resistance (Hejna et al., 2019), erucic acid and tocopherol (vitamin E) isoform accumulation in seeds (Havlickova et al., 2018), and leaf nutrition concentration (Alcock et al., 2018). Recently, the power of AT was further enhanced by using a much larger panel comprising 383 rapeseed accessions (Havlickova et al., 2018). In bread wheat, two causative genes underlying stem strength variation have also been detected by AT (Miller et al., 2016). Despite the above efforts, the AT approach has not yet been applied to photosynthetic related traits under LT conditions in rapeseed.

The present study aims to identify candidate genes associated with photosynthetic gas exchange parameters including A_n, stomatal conductance to water vapour (G_{sw}), internal CO_2 concentration (C_i) and transpiration rate/evapotranspiration (E) in rapeseed by AT. Twenty-two candidate genes were obtained, and one was functionally validated. Ectopic expressing tropinone reductase (*BnTR1*) in *Arabidopsis* can significantly enhance transpiration rate and LT tolerance, implying its great potential for the genetic improvement of LT tolerance in plant.

MATERIALS AND METHODS

Plant materials

A panel comprising 123 rapeseed accessions was used for association study, which is available from the John Innes Centre, Norwich, UK (**Supplementary Table S1**) (Havlickova et al., 2018). Within this panel, there are 37 winter type, 32 spring type, 47 semi-winter type and 7 unclassified rapeseed accessions. The panel was sown on 28th Oct in 2016 in Wuhan (114.30°E, 30.57°N), China. All accessions were planted using a completely randomized block design with three replications. The temperature was recorded during the field experiments (**Supplementary Figure S1)**. Compared with those sown on normal occasion (28th Sept in 2016), the late-sown (i.e. 28th Oct) rapeseed seedlings were subjected to LT stress during winter.

Determination of photosynthetic gas exchange parameters

The fourth true leaf of each of the 123 rapeseed accessions was chosen for the measurement of photosynthetic gas exchange parameters including A_n, G_{sw}, C_i and E in the open field at the 60-d-old seedling stage. Two independent plants of one accession from each block were measured by LI-6400 photosynthesis equipment (Li-Cor 6400, Li-CorInc, Lincoln, NE, USA) as described previously (Yan et al., 2015). The measurements were performed on a sunny day from 27th December to 30th December with a maximum temperature of 9°C in daytime and the lowest temperature of -1°C at night. The phenotypic data were collected from three blocks as biological replications (**Supplementary Table S2**). Broad-sense heritability was estimated according to a previous study (Kaler and Purcell, 2019).

The photosynthetic gas exchange parameters of *Arabidopsis* wild type (WT) and transgenic seedlings (ecotype Columbia) were measured on the second functional leaf with the light intensity of 800 μmol $m^{-2}s^{-1}$ and CO_2 concentration of 400 μL L^{-1}. All of the *Arabidopsis* plants were grown in the greenhouse (16-h-light/8-h-dark) with the light intensity of 120 μmol m^{-2} s^{-1} at 23°C. Each leaf was measured three times as technical replications, and five independent plants from each line were measured as biological replications (**Supplementary Table S2**).

Genome-wide association study

The association panel and the procedure of AT analysis have been reported in detail previously (Havlickova et al., 2018). In brief, RNA-Seq data were generated from young leaves of the association panel harvested 21 d after sowing under 16-h-light (20°C)/8-h-dark (14°C) glasshouse conditions. The transcriptome data were mapped onto the developed ordered *Brassica* A and C pan-transcriptomes (He et al., 2015) and resulted in a set of 355,536 SNPs and RPKM values for 116,098 CDS models. Following the removal of SNP markers with minor allele frequencies below 0.01, a total of 256,397 SNPs were retained (http://www.yorknowledgebase.info/) and used as marker input for the Associative Transcriptomics analysis as previously described (Harper et al., 2012; Lu et al., 2014). The current study adopted a compressed mixed linear model including both fixed and random effects according to a previous method (Lipka et al., 2012). The *P*-values ($-\log_{10}$ converted) for all SNPs

were plotted against their physical position in the 'pseudo-molecules' to produce a Manhattan plot. The Bonferroni significance threshold was set as $P=3.9\times10^{-6}$ (1/256397) (Duggal et al., 2008). Allelic effects of all candidate SNPs were calculated according to a previous study (Prado et al., 2017); a positive effect indicates that the allele increases the trait value, whereas a negative effect indicates that the alternative allele increases the trait value.

The transcript level was quantified as reads per kb per million aligned reads (RPKM) across the panel. After filtering (RPKM≤0.4), a total of 53,889 gene expression marker (GEM) was obtained. The GEM was regarded as the dependent variable and trait data as the independent variable (Wood et al., 2017). The fixed effect linear model was performed to assess the relationship between gene expression level and the traits (Alcock et al., 2017). The *P*-value for each GEM was converted ($-\log_{10}P$) and plotted against its physical position to generate a Manhattan plot. The Bonferroni significance threshold was set at $P=1.85\times10^{-5}$ (1/53889).

Growth conditions and stress treatments

To determine the gene function of *BnTR1* on *Arabidopsis* under freezing conditions, the seeds of WT and transgenic *Arabidopsis* plants were germinated on 1/2 MS medium and 1/2 MS medium plus 30 μg/mL hygromycin, respectively. After 1 week growth at 23°C (16-h-light/8-h-dark), healthy seedlings with uniform sizes were transplanted into 8×8 cm pots (four plants per pot). Then, the 21-d-old plants were transferred into a growth chamber at -4°C for 4 h after 24 h of cold acclimation and recover at 23°C (16-h-light/8-h-dark). Six pots from each transgenic line or WT plants were used to investigate the survival rate 3 d after recovery. The leaves were sampled for physiological and biochemical measurements.

To determine the effects of alkaloid on *Arabidopsis* and rapeseed seedlings, WT plants of *Arabidopsis* were firstly grown under the same conditions as above. Before the freezing treatment, a dosage of 0, 10, and 30 nmol alkaloid (atropine) per seedling was added to the soil for *Arabidopsis*, while a dosage of 0, 50 and 150 nmol per seedling was added to the soil for rapeseed accession Zhongshuang 11 (ZS11) according to the previous study (Hara and Kurita, 2014). After inoculation overnight,

the 21-d-old *Arabidopsis* plants were transferred into a growth chamber at -4°C for 4 h, the three-leaf-stage rapeseed plants were transferred into the growth chamber at -4°C for 4 h (Yan et al., 2019). The survival rate was investigated in six pots 3 d after recovery (**Supplementary Table S2**).

To investigate the expression of candidate genes under LT stress conditions, six rapeseed accessions showing extreme photosynthetic efficiency (i.e. Sv706118, Kajsa, Callypso, Libritta, Gefion and Jupiter; **Supplementary Table S1**) were used for expression analysis of candidate genes. The four-leaf seedlings were transferred into the plant growth chamber with -4°C for 4 h (Yan et al., 2019). The leaves were sampled before, and after freezing treatment, and then used for RNA extraction and molecular analysis. The detailed information was listed in **Supplementary Table S2.**

Physiological and Biochemical Measurements

To determine the effects of *BnTR1* on *Arabidopsis* at physiological and biochemical levels under LT stress conditions, the seedlings of transgenic and WT *Arabidopsis* plants were treated under freezing conditions (-4°C for 4 h) and, the leaves of 21-d-old seedlings were sampled at three time-point (i.e. before freezing treatment, after freezing treatment, and recovery for 3 d at 23°C) for measuring physiological and biochemical characteristics, including Fv/Fm, electrolyte leakage, 3,3'-diaminobenzidine (DAB) staining, proline content, soluble sugar content, reactive oxygen species (ROS) scavenging enzymes activity, H_2O_2 content, alkaloid content. All measurements were performed with at least three biological replications; detail information for experimental design and plant materials used was listed in **Supplementary Table S2**.

The Fv/Fm measurement was performed using the second functional leaves before and after the freezing treatment. The leaves were firstly immersed in 1% agarose overnight avoiding of the dark, then the chlorophyll measurement (Fv/Fm) was measured using the modulated chlorophyll fluorescence instrument (PAM-2500; Walz) as previously reported (Lv et al., 2017).

The electrolyte leakage measurement was performed according to the previous study (Lv et al., 2016). Briefly, six leaves from six were cut and immersed in 8 mL of

253 double-distilled H_2O in a 10-mL tube. After shaking overnight, the electrolyte leakage
254 was measured using a model DDS-IIA device (Leici Instrument) as R1; it was
255 measured again and recorded as R2 after boiling at 95°C in a water bath for 15 min
256 and cooling down. The relative electrolyte leakage was calculated as a ratio of R1/
257 R2.

258 The proline and soluble sugar contents were measured using the kits from Beijing
259 Solarbio Science & Technology as described before (Yan et al., 2019). In brief, 0.1 g
260 fresh tissue was powdered and incubated in 1 ml 3% sulfosalicylic acid (for proline)
261 or ddH_2O (for soluble sugar). After centrifuging, 400 μL supernatant was mix with
262 other reaction buffers and incubated at 95°C in a water bath for 15 min, then the
263 absorbance was measured using MULTISCAN FC (Thermo Scientific).

264 The ROS scavenging enzymes activity was measured by commercial kits
265 according to the manufacturer's instruction (Beijing Solarbio Science & Technology)
266 with minor modification (Yan et al., 2019). 0.1 g fresh tissue was powdered using 1
267 mL 0.05 mol/L PBS buffer (pH 7.8). The supernatant was obtained after centrifuging
268 at 8,000 g for 10 min at 4°C and used for superoxide dismutase (SOD) activity,
269 peroxidase (POD) activity, catalase (CAT) activity measurement using MULTISCAN
270 FC (Thermo Scientific).

271 DAB staining was performed as previously described (Ning et al., 2010; Zhang
272 et al., 2011). The fourth functional leaf of each plant was sampled and infiltrated in
273 0.1 mg/mL 3,3'-diaminobenzidine liquid (50 mM Tris-acetate buffer, pH 5.0). After
274 incubation overnight at 25°C in the dark, the stained leaves were photographed after
275 removing the chlorophyll by absolute ethanol. The H_2O_2 content was quantified
276 according to the instruction of the kit (Beijing Solarbio Science & Technology) (Yan
277 et al., 2019).

278 The total alkaloid was extracted as described previously (Chen et al., 2013;
279 Zhang et al., 2004). The freeze-dried leaves were powdered; 0.1 g powder was
280 homogenized overnight with 1.0 mL 70% aqueous methanol at 4°C. Following
281 centrifugation at 10,000 g for 10 min at 4°C, the extracts were absorbed (CNWBOND
282 Carbon-GCB SPE Cartridge, 250 mg, 3 mL; ANPEL) and filtrated (SCAA-104, 0.22

μm pore size; ANPEL). Next, the total alkaloid content was determined using the alkaloid ELISA kit (Hiton) according to the instructions.

RNA extraction and gene expression analyses

Total RNA was extracted from the *Arabidopsis* or rapeseed seedlings with TransZol reagent (Trans) and converted to the first-strand cDNA using the EasyScript®One-Step cDNA Synthesis SuperMix (Trans). The Quantitative Real-time PCR (qRT-PCR) was performed using Power SYBR®Green PCR Master Mix according to the manufacturer's instructions on a StepOnePlusReal-Time PCR System (Applied Biosystems). The primers used for expression analysis of candidate genes either detected by AT approach or involved in the known biosynthetic pathway of alkaloid were listed in **Supplementary Table S3**. The relative expression level was determined as previously described (Livak and Schmittgen, 2001).

Vector construction and gene transformation

To generate transgenic lines over-expressing candidate gene (*BnTR1*), the coding sequence (CDS) of *LOC106445422* was amplified from a rapeseed variety ZS11 and ligated into vector pCAMBIA1300 driven by a tobacco cauliflower mosaic virus 35S promoter (CaMV35S). The construct was introduced into *Arabidopsis* variety Columbia (Col) by *Agrobacterium*-mediated transformation (Zhang et al., 2006). Three T4 homozygous lines (L1, L3, L5) significantly over-expressing *BnTR1* were obtained by screening at 30 μg/mL hygromycin. The primers used for vector construction were listed in **Supplementary Table S3**.

Statistical analyses

Statistical analyses were conducted with Microsoft Excel (2003) and SPSS (version 22.0) software using one-way analysis of variance (ANOVA) with Tukey's multiple comparisons test or two-tailed Student's *t*-test. All data were presented as the means ± standard error (SE) based on three replicates. $P<0.05$ and $P<0.01$ were considered statistically significant and highly significant, respectively.

RESULTS

Phenotypic variation of photosynthetic gas exchange parameters

A field-grown rapeseed association panel experienced long-term LT stress in winter (**Supplementary Figure S1**). The photosynthetic gas exchange parameters such as A_n, G_{sw}, C_i and E were measured since they can reflect photosynthetic efficiency for a plant. Substantial variations for the four traits were observed in 123 rapeseed accessions (**Figure 1**). A_n varied from 12.87 to 24.02 with a mean of 19.29 μmol (CO_2) $m^{-2}s^{-1}$. Similarly, G_{sw} ranged from 245.21 to 383.28 μmol (CO_2) mol^{-1} and E from 0.96 to 3.75 mmol (H_2O) $m^{-2}s^{-1}$. G_{sw} was the most variable trait since it has the largest coefficient of variation (0.69), with a minimum of 0.16 and a maximum of 2.53 mol (H_2O) $m^{-2}s^{-1}$; the range of broad-sense heritability varied from 49.49% to 68.91% (**Supplementary Table S4**). Moreover, the four traits in the association panel were positively correlated with each other ($P \leq 0.01$) (**Supplementary Table S5**). For instance, there was a strong correlation between A_n and E ($r = 0.785$, $P < 0.01$).

Associative Transcriptomics for photosynthetic gas exchange parameters

To identify genomic regions controlling photosynthetic related traits, AT was performed in rapeseed. Using the mixed linear model, a total of 201 significant SNPs were detected, which originated from 148 CDSs (**Figure 2**). Unexpectedly, most of the significant CDSs were associated with G_{sw} trait, while only one was related to E trait. The detail results including physical positions, *P*-values, allelic effects were summarized in **Supplementary Table S6**. For GEM analysis, a total of 145 CDSs above the corrected Bonferroni thresholds were identified (**Figure 2**). Of these, a respective of 5, 10, 20 and 110 CDSs were detected for A_n, G_{sw}, C_i and E. The detail information including physical positions and *P*-values for all significant GEMs was listed in **Supplementary Table S7**.

By annotating all the above significant CDS in public database TAIR (https://www.arabidopsis.org/), the present study shortlists the number of candidate genes to only 22 that putatively involved in photosynthesis or LT stress response (**Supplementary Table S8**). To verify their roles in rapeseed with or without LT stress, six accessions from the association panel were selected; Sv706118, Kajsa, and

Callypso (Accessions 1-3) exhibited higher photosynthesis efficiency and were tolerant to freezing stress, while Libritta, Gefion, and Jupiter (Accessions 4-6) presented lower photosynthesis efficiency and were sensitive to freezing stress (**Supplementary Table S1, Supplementary Figure S2**). The gene expression profiles were investigated by qRT-PCR in the six accessions under freezing conditions. Results showed that all of these genes exhibited a significantly different expression level in the six accessions, suggesting that these genes indeed involved in freezing stress response. *Cab026133.1* seemed to have a much higher expression level in the three tolerant accessions (Accessions 1-3) with or without freezing stress (**Figure 3A**); so did *Cab011968.1*, *Cab022014.2*, and *Cab007526.2*, an orthologue of inorganic carbon transport protein (AT1G70760.1), bZIP transcription factor (AT5G28770.3) and citrate synthase 2 (BnaA10g24440D), respectively. An opposite trend was observed for *Bo5g017460.1* and *Cab008128.1*, an orthologue of F-box family protein (AT2G32560.1) and dehydroascorbate reductase (AT5G16710.1), respectively (**Supplementary Figure S3**).

Selection and characterization of candidate gene

Cab026133.1 was selected for further analysis because it not only exhibited the highest *P*-value (6.33×10^{-9}) for E trait in GEM analysis (**Supplementary Table S8**) but also highly expressed in LT tolerant accessions (**Figure 3A**). Besides, the expression of *Cab026133.1* (presented as RPKM) across the rapeseed panel was positively correlated with E level (r=0.406, $P<10^{-3}$) and accounted for 16.5% of trait variation (**Figure 3B**). The ortholog of Cab026133.1 in *Arabidopsis* (AT2g29300) encodes an SDR protein involved in the oxidation-reduction process of secondary metabolites such as phenols, isoprene and alkaloid (Selmar and Kleinwachter, 2013). SDR proteins are classified into six subfamilies, and the tropinone reductase subfamily belongs to the major route of alkaloid biosynthesis (Tonfack et al., 2011). The alignment of amino acid sequence clearly illustrated that LOC106445422 shared 87.3% similarity with Cab026133.1 and 57% with CoTR, a known tropinone reductase in *Cochlearia officinalis* (Brock et al., 2008). LOC106445422 displayed

typical SDRs motifs (Gly-X_3-Gly-X-Gly) and four conserved residues that form the catalytic tetrad NSYK (N127, S155, Y168, K172) (**Supplementary Figure S4**). The current study name LOC106445422 as *BnTR1* and used as the candidate gene for the follow-up studies.

To assess the effect of allelic variation on *BnTR1* in the rapeseed association panel, the genomic region covering the whole gene as well as the 2-kb promoter region was amplified. One TAA/TAATAA insertion was detected in the fourth intron that formed two major haplotypes, i.e. Haplotype I (with TAA insertion) and Haplotype II (with TAATAA insertion) (**Figure 3C**). Haplotype I (n=31) displayed significantly higher E value than Haplotype II (n=60) (P=5.43×10^{-4}) (**Figure 3D**). Besides, the LT tolerant Accessions 1-3 were determined as Haplotype I while sensitive Accessions 4-6 as Haplotype II at *BnTR1* locus (**Supplementary Table S1**). The expression level of *BnTR1* in Accessions 1-3 was also significantly higher than that in accessions 4-6 with or without stress treatment (**Figure 3A**). Therefore, it was evident that E variation may be attributed to expression or allelic variation at *BnTR1*, which was expressed almost in all tissues of rapeseed at both vegetative and reproductive stages (**Supplementary Figure S5**).

Ecotopic expressing *BnTR1* enhances freezing tolerance in *Arabidopsis*

To analyze gene function, three independent *Arabidopsis* lines (L1, L3, L5) ectopic expressing *BnTR1* were generated. All transgenic lines showed an increased expression level of *BnTR1* in comparison to the WT plants (**Figure 4A**). At the seedling stage, E value of the transgenic lines was much higher than that of WT plants (**Figure 4B**), thus confirming that *BnTR1* controls transpiration rate. Since photosynthetic gas exchange parameters are significantly correlated with cold tolerance (Urban et al., 2013), it was speculated that BnTR1 also involves in LT stress. To test this hypothesis, seedlings were treated at -4°C for 4 h and then recover at 23°C. All of the transgenic lines were shown to be freezing-tolerant because there was no obvious syndrome, while the WT plants were wrinkled and hydrophanous (**Figure 4C**). After recovery for 3 d at 23°C, all transgenic plants survived, but 62% of the WT

plants died (**Figure 4D**). These results strongly suggested that BnTR1 enhanced the freezing tolerance of *Arabidopsis* plants.

To assess whether C-repeat-binding factors (CBFs) contributed to the enhanced freezing tolerance of the transgenic lines, the current study measured the relative expression level of *CBFs* in transgenic and WT plants after freezing stress (**Figure 4E-G**). In the transgenic lines grown under normal conditions, expressions of *CBFs* were much higher than that in WT plants, indicating that they were markedly induced by *BnTR1*. Moreover, the expression of *CBF1* and *CBF3* was significantly up-regulated and *CBF2* down-regulated by *BnTR1* during freezing treatment in comparison with those of normal conditions. The current study further examined the expression level of CBFs-targeted cold-responsive genes (COR genes) (**Figure 4H-I**). As expected, the transcript levels of COR genes (namely *COR15* and *RD29A*) in the transgenic lines were highly induced when compared with that in WT plants. These results indicated that *BnTR1* influences CBF regulon in the stress-signalling pathway to control freezing tolerance.

The chlorophyll fluorescence parameter Fv/Fm, an indicator for the potential maximum photosystem II (PSII) capacity of plants, has been widely used to determine the ability of tolerance to environmental stresses under laboratory conditions (Mishra et al., 2014; Thalhammer et al., 2014). Here, markedly higher Fv/Fm ratio was observed in leaves of the transgenic lines under freezing as well as normal conditions (**Figure 5A**). To further clarify whether the difference of photosynthetic capacity was caused by the excessive expression of *BnTR1*, the current study assessed the expression level of *BnTR1* and other genes involved in photosynthetic processes such as *RCA*, *SBPASE* (for CO_2 fixation or assimilation) and *CAB1-4* (for light-harvesting) (Basu et al., 2019; Sun et al., 2017) (**Figure 5B-H**). Under normal conditions, the expression level of *RCA*, *SPASE* and *CAB1* were slightly reduced in the transgenic lines compared to WT plants, whereas *CAB2*, *CAB3* and *CAB4* were induced. The freezing treatment led to a notable suppression of all genes; however, the expression of genes in the transgenic lines returned to a high level when freezing stress was removed.

BnTR1 contributes to cell membrane protection and antioxidants

Altering the osmotic balance to maintain the integrity and stability of cell membrane is proposed to be an efficient way for plants adapting to the changing environments (Morsy et al., 2005; Valerio et al., 2011). To test this hypothesis, physiological and biochemical assays were carried out. Results showed that freezing treatment led to only 40-200% increase of proline content in WT plants but as high as 100-300% in transgenic plants, indicating that the transgenic plants expressing *BnTR1* could accumulate more proline (**Figure 6A**). The soluble sugar content showed a similar pattern (**Figure 6B**). However, the electrolyte leakage increased more rapidly in WT plants than in transgenic lines (**Figure 6C**). These results implied that BnTR1 actively responded to freezing stress by maintaining cell membrane stability and osmotic balance.

Antioxidants, which function in scavenging the reactive oxygen species (ROS), are generally considered as another effective element in defending abiotic stresses (Choudhury et al., 2017). To determine whether *BnTR1* affects the antioxidant system, the accumulation of ROS was determined by DAB staining. The brown precipitate (H_2O_2) in WT was much larger than that in the transgenic lines (**Figure 6D**), indicating that WT plants had a higher level of H_2O_2 content than transgenic plants (**Figure 6E**). Oxidoreductases like POD, SOD and CAT also function in scavenging redundant ROS (Gupta et al., 2016). Here, we found that the transgenic lines exhibited stronger SOD activity than WT (**Figure 6F**), which help plants alleviate oxidation damage from freezing conditions. However, no significant difference was detected for POD and CAT activities. Together, the enhanced freezing adaption for transgenic plants could be attributed to the increased ROS scavenging ability.

BnTR1 positively affects alkaloid metabolism

BnTR1 (homolog of AT2g29300) is predicted to be a tropinone reductase involved in the biosynthesis of alkaloid (KO00960), which mainly produces atropine. Although all alkaloids (with more than 12,000 different structures) have been

well-documented in pharmacology, their roles in abiotic stress remain elusive (Schlager and Drager, 2016). To investigate the specific role of BnTR1 in alkaloid metabolism, the total alkaloids content was quantified. As expected, the transgenic lines led to one- to two-fold increase of alkaloids contents compared with WT plants after freezing stress (**Figure 7A**). To further confirm the effect of alkaloid on stressed plants, exogenous atropine was applied to WT plants, since atropine was considered to be the product of alkaloid metabolism (KO00960) (Hara and Kurita, 2014). Results demonstrated that application of 10 nmol atropine per plant significantly rescued the susceptibility of WT plants, but the protective effect was weakened when dosage increase to 30 nmol (**Figure 7B**). The survival rate increased by three- to four-fold compared with WT plants without atropine treatment (**Figure 7C**).

To further elucidate the protective role of alkaloid in rapeseed, the current study applied exogenous atropine to a widely cultivated rapeseed variety, ZS11, under freezing conditions. Phenotypic analysis showed that the wilting phenotype of ZS11 plants was partially rescued by exogenous atropine application (50 and 150 nmol per plant) (**Figure 7D**), while no significant difference was observed in the survival rate. It was concluded that alkaloids alleviated the damage on plants from extreme LT stress.

DISCUSSION

Power of AT approach

Brassica napus originated from the hybridization of *Brassica rapa* and *Brassica oleracea* which contribute the A and C genomes, respectively (Cheung et al., 2009). There is only 15% difference in nucleotide structure and 3% difference in transcriptional expression patterns between chromosomes A and C, which limits the development of SNP markers in genome-wide association analysis until the availability of the high throughput next-generation sequencing technology (Adams et al., 2003; Higgins et al., 2012). Over the past years, AT approach based on abundant SNP markers and GEMs, has successfully simplified the complexity of the whole genome (Harper et al., 2012), and has been widely applied in rapeseed, wheat and

493 other polyploidy crops (Alcock et al., 2017; Harper et al., 2016a; Harper et al., 2016b;
494 Harper et al., 2012; Havlickova et al., 2018; Koprivova et al., 2014; Lu et al., 2014;
495 Miller et al., 2018; Miller et al., 2016; Schuster et al., 2013). However, the genetic
496 basis of photosynthetic-related traits in oil crops remains elusive. Here, the genetic
497 architecture of photosynthetic gas exchange parameters was investigated by AT
498 approach, and a gene termed *BnTR1* was confirmed to be responsible for E trait
499 (Figure 4), which might be a promising candidate beneficial to rapeseed in coping
500 with climatic changes. Several other interesting candidates were also identified. For
501 instance, Bo5g155110.1 was found to be significantly associated with E traits
502 (P=1.1×10^{-5}), and down-regulated by freezing stress (**Supplementary Figure S3**).
503 The homolog in *Arabidopsis* is Cyclophilin38 (AtCYP38), which functions in the
504 assembly and maintenance of PSII super complex (**Supplementary Table S7**). The
505 loss-of-function mutant of *AtCYP38* shows reduced growth rate and photosynthetic
506 efficiency compared to its wild type. Additionally, the D1 and D2 proteins in PSII
507 reaction centre show a short half-life, resulting in susceptibility upon exposure to
508 excessive light (Fu et al., 2007; Sirpio et al., 2008). Bo3g153100.1 was hit by an SNP
509 marker, with -log10 (*P*-value) value as high as 9.06 (**Supplementary Table S6**), it
510 was also markedly up-regulated by freezing stress (**Supplementary Figure S3**).
511 Bo3g153100.1 was homologues to AT4G37930 in *Arabidopsis*, which has been
512 documented in the photorespiration process (Takahashi et al., 2007). In the knockout
513 mutant of AT4G37930, the photorespiration pathway is destroyed, and the
514 chlorophyll deficiency results in chlorosis (Voll et al., 2006). Therefore, it seems that
515 AT is a powerful tool to identify candidate genes for photosynthesis and LT stress in
516 rapeseed. It is worthy to further study the function of all 22 candidate genes identified
517 here.
518
519 **The positive role of BnTR1 under LT conditions**
520 It is generally accepted that photosynthesis is vulnerable to adverse
521 environmental stresses such as extreme temperature, salinity, drought or combined
522 stresses (Sainz et al., 2010; Strzepek et al., 2019). Abiotic stresses lead to

photoinhibition as well as excessive generation of ROS, which suppresses the photosynthetic progress and ultimately repress the growth and productivity in plants (Gabriel et al., 2010; Nishiyama et al., 2014). During the long-term evolution, plants have developed a variety of adaptive mechanisms to cope with the stressful conditions (Liu et al., 2019a; Strzepek et al., 2019). The CBF transcription factors in rapeseed are known to be responsible for the photosynthetic performance; CBF5 and CBF17 enhance the energy conversion efficiency under LT conditions (Dahal et al., 2012; Savitch et al., 2005). CBF1-CBF3, also termed dehydration-responsive element-binding factors, have been well-documented in plants. In *Arabidopsis*, CBF2 represents a negative regulator for LT response, while CBF1 and CBF3 are positive regulators (Novillo et al., 2004; Novillo et al., 2012). Interestingly, increased expression of *CBF1* and *CBF3* and repressed expression of *CBF2* were observed in the *BnTR1* transgenic lines (**Figure 4E-G**), indicating that BnTR1 represented a unique influence on CBF members. Both alleviated accumulation of ROS and activated SOD enzyme system was observed in the *BnTR1* transgenic lines (**Figure 6**), suggesting active impacts of BnTR1 on the ROS scavenging system. In addition, ectopic expressing *BnTR1* also promote the expression of the genes associated with plant photosynthesis (**Figure 5**). Specifically, the decrease of *RCA* transcripts leads to lower A_n value, which in turn slows down plant growth (Von Caemmerer et al., 2005; Yin et al., 2010). Moreover, RCA enhances growth and photosynthesis under moderate heat stress conditions (Kumar et al., 2009; Kurek et al., 2007). However, overexpression of *SBPASE* improves sugar accumulation and enhanced photosynthesis efficiency (De Porcellinis et al., 2018; Feng et al., 2007; Miyagawa et al., 2001). In the present study, PSII was severely repressed during freezing treatment, whereas the *BnTR1* transgenic *Arabidopsis* plants still exhibited higher Fv/Fm level compared to WT (**Figure 5A**). These observations suggest that BnTR1 triggered a series of responses including the ROS scavenging system, CBF pathway and photosynthetic processes. However, further work is required to confirm their roles in LT tolerance.

Protective role of alkaloids under LT conditions

Previous studies have been instrumental in revealing some metabolites underlying stress response mechanisms (Thalmann and Santelia, 2017). So far, definitions of alkaloids are generally focused on strong pharmacological effects, such as antimitotic, antidote, anticancer and antioxidants (Schlager and Drager, 2016). The concentration of alkaloid compounds is predominantly inducible when plants are subjected to multiple stresses (Cheng et al., 2018; Srivastava and Srivastava, 2010). However, few studies have recognized the positive correlations between alkaloids and stress resistance. Application of sanguinarine for *Arabidopsis* seedlings under heat stress condition could markedly enhance the tolerance, which presumably by promoting the expression of heat shock proteins like HSP70 and HSP90.1S (Hara and Kurita, 2014; Matsuoka et al., 2016). BnTR1 is predicted to encode a tropinone reductase, which is involved in the metabolic pathway of atropine alkaloids. The current study determined the total alkaloids content under stress conditions, which showed an increased level in *BnTR1* transgenic lines under normal and freezing stress conditions compared with WT plants (**Figure 7A**). Moreover, the application of exogenous atropine alleviated the damage caused by extreme temperature in both *Arabidopsis* and rapeseed seedlings (**Figure 7B, D**), which was in agreement with the observations in sanguinarine under heat stress conditions (Schlager and Drager, 2016). However, more studies are still required to confirm that alkaloids could function as a protectant for plants to confer stronger resistance to LT stresses. The current study has compared the expression level of stress-related genes in *Arabidopsis* plants treated with exogenous atropine under freezing conditions (**Supplementary Figure S6**). It was found that atropine could promote the expression of *CBF1*, *CBF3*, *CAB1*, *CAB3*, *CAB4*, *SPASE* before or after freezing treatment, but the extent is much lower than that induced by BnTR1 (**Figure 4**, **Figure 5**). Thus the results confirmed at least in part the protective role of atropine for a plant in adaption to LT stress. It is proposed that BnTR1 works as an effecter via metabolizing alkaloids accumulation, photosynthesis, CBF/DREB pathways and ROS scavenging system in stressed *Arabidopsis*, which in turn contributes to the adaptation under LT conditions.

CONCLUSIONS

During overwintering for the semi-winter type rapeseed grown in China, the extremely low temperature has a deleterious impact on plant productivity. Therefore, the identification of genes responsible for stress response is the prime interest of researchers. Despite of limited phenotypic data, our associative transcriptomics approach has been successfully used to dissect the genetics of photosynthetic-related traits under low temperature conditions. The first short-chain dehydrogenease/reductase, BnTR1 was identified in rapeseed, which improved the transpiration rate and freezing tolerance of *Arabidopsis* plants. Taken together, our findings illustrated the molecular mechanism of plant adaption to low temperature stress. Finally, this work sheds light on the way to increase low temperature tolerance in rapeseed by genetic engineering strategies.

Supplementary data

Supplementary Figure S1. Temperature record during the phenotypic investigation of the field-grown association panel.

Supplementary Figure S2. Phenotype investigation of six rapeseed accessions under LT stress conditions.

Supplementary Figure S3. Expression analysis of candidate genes in six rapeseed accessions under LT stress conditions.

Supplementary Figure S4. Sequence alignment of BnTR1 and SDR proteins.

Supplementary Figure S5. Spatio-temporal expression patterns of *BnTR1* in various tissues of rapeseed.

Supplementary Figure S6. Expression analysis of LT- or photosynthetic-related genes in *Arabidopsis* with atropine application.

Supplementary Table S1. Information of 123 accessions used in the association study.

Supplementary Table S2. Summary of experiment design for the present study

Supplementary Table S3. Primers used in the present study.

Supplementary Table S4. Phenotypic variations of photosynthesis-related traits in 123

rapeseed accessions.

Supplementary Table S5. Correlation analysis of photosynthesis-related traits in 123 rapeseed accessions.

Supplementary Table S6. SNP markers detected by association study on photosynthesis-related traits in rapeseed.

Supplementary Table S7. GEMs markers detected by association study on photosynthesis-related traits in rapeseed.

Supplementary Table S8. Candidate genes detected by association study on photosynthesis-related traits in rapeseed.

ACKNOWLEDGEMENTS

This research was financially supported by funds from the National Key Research and Development Program of China (2018YFD0100905), the Oil Crops Research Institution Basal Research Fund of the Chinese Academy of Agricultural Sciences (CAAS), China (1610172018010), the major science and technology project, Ministry of science and technology, China (2018ZX08020001).

REFERENCES

Adams, K. L., Cronn, R., Percifield, R., and Wendel, J. F. (2003). Genes duplicated by polyploidy show unequal contributions to the transcriptome and organ-specific reciprocal silencing. *Proc Natl Acad Sci U S A*. 100, 4649-4654. doi: 10.1073/pnas.0630618100.

Alcock, T. D., Havlickova, L., He, Z., Bancroft, I., White, P. J., Broadley, M. R., et al. (2017). Identification of candidate genes for calcium and magnesium accumulation in Brassica napus L. by association genetics. *Front Plant Sci*. 8, 1968. doi: 10.3389/fpls.2017.01968.

Alcock, T. D., Havlickova, L., He, Z., Wilson, L., Bancroft, I., White, P. J., et al. (2018). Species-wide variation in shoot nitrate concentration, and genetic loci controlling nitrate, phosphorus and potassium accumulation in Brassica napus L. *Front Plant Sci*. 9, 1487. doi: 10.3389/fpls.2018.01487.

Andersson, J., Wentworth, M., Walters, R. G., Howard, C. A., Ruban, A. V., Horton, P., et al. (2003). Absence of the Lhcb1 and Lhcb2 proteins of the light-harvesting complex of photosystem II - effects on photosynthesis, grana stacking and fitness. *Plant J.* 35, 350-361. doi: 10.1046/j.1365-313x.2003.01811.x.

Basu, U., Bajaj, D., Sharma, A., Malik, N., Daware, A., Narnoliya, L., et al. (2019). Genetic dissection of photosynthetic efficiency traits for enhancing seed yield in chickpea. *Plant Cell Environ*. 42, 158-173. doi: 10.1111/pce.13319.

Bazakos, C., Hanemian, M., Trontin, C., Jimenez-Gomez, J. M., and Loudet, O. (2017). New strategies and tools in quantitative genetics: how to go from the phenotype to the genotype. *Annu Rev Plant Biol*. 68, 435-455. doi: 10.1146/annurev-arplant-042916-040820.

Bernfur, K., Rutsdottir, G., and Emanuelsson, C. (2017). The chloroplast-localized small heat shock protein Hsp21 associates with the thylakoid membranes in heat-stressed plants. *Protein Sci*. 26, 1773-1784. doi: 10.1002/pro.3213.

Brock, A., Brandt, W., and Drager, B. (2008). The functional divergence of short-chain dehydrogenases involved in tropinone reduction. *Plant J*. 54, 388-401. doi: 10.1111/j.1365-313X.2008.03422.x.

Chen, W., Gong, L., Guo, Z., Wang, W., Zhang, H., Liu, X., et al. (2013). A novel integrated method for large-scale detection, identification, and quantification of widely targeted metabolites: application in the study of rice metabolomics. *Molecular Plant*. 6, 1769-1780. doi: 10.1093/mp/sst080.

Cheng, T., Hu, L., Wang, P., Yang, X., Peng, Y., Lu, Y., et al. (2018). Carbon monoxide potentiates high temperature-induced nicotine biosynthesis in tobacco. *Int J Mol Sci*. 19, 188. doi: 10.3390/ijms19010188.

Cheung, F., Trick, M., Drou, N., Lim, Y. P., Park, J. Y., Kwon, S. J., et al. (2009). Comparative analysis between homoeologous genome segments of Brassica napus and its progenitor species reveals extensive sequence-level divergence. *Plant Cell*. 21, 1912-1928. doi: 10.1105/tpc.108.060376.

Choudhury, F. K., Rivero, R. M., Blumwald, E., and Mittler, R. (2017). Reactive oxygen species, abiotic stress and stress combination. *Plant J*. 90, 856-867. doi: 10.1111/tpj.13299.

Cong, R. H., Zhang, Z., and Lu, J. W. (2019). Climate impacts on yield of winter oilseed rape in different growth regions of the Yangtze River Basin. *Chinese Journal of Oil Crop Sciences*. 41, 894-903. doi: 10.19802/j.issn.1007-9084.2019046

Dahal, K., Gadapati, W., Savitch, L. V., Singh, J., and Huner, N. P. (2012). Cold acclimation and BnCBF17-over-expression enhance photosynthetic performance and energy conversion efficiency during long-term growth of Brassica napus under elevated CO2 conditions. *Planta*. 236, 1639-1652. doi: 10.1007/s00425-012-1710-2.

De Miguel, M., Cabezas, J. A., De Maria, N., Sanchez-Gomez, D., Guevara, M. A., Velez, M. D., et al. (2014). Genetic control of functional traits related to photosynthesis and water use efficiency in Pinus pinaster Ait. drought response: integration of genome annotation, allele association and QTL detection for candidate gene identification. *BMC Genomics*. 15, 464. doi: 10.1186/1471-2164-15-464.

De Porcellinis, A. J., Norgaard, H., Brey, L. M. F., Erstad, S. M., Jones, P. R., Heazlewood, J. L., et al. (2018). Overexpression of bifunctional fructose-1,6-bisphosphatase/sedoheptulose-1,7-bisphosphatase leads to enhanced photosynthesis and global reprogramming of carbon metabolism in Synechococcus sp. PCC 7002. *Metab Eng*. 47, 170-183. doi: 10.1016/j.ymben.2018.03.001.

Ding, C.-K., Wang, C. Y., Gross, K. C., and Smith, D. L. (2001). Reduction of chilling injury and transcript accumulation of heat shock proteins in tomato fruit by methyl jasmonate and methyl salicylate. *Plant Science*. 161, 0-1159. doi: 10.1016/S0168-9452(01)00521-0

Duggal, P., Gillanders, E. M., Holmes, T. N., and Bailey-Wilson, J. E. (2008). Establishing an adjusted p-value threshold to control the family-wide type 1 error in genome wide association studies. *BMC Genomics*. 9, 516. doi: 10.1186/1471-2164-9-516.

Evans, J. R. (2013). Improving photosynthesis. *Plant Physiol*. 162, 1780-1793. doi: 10.1104/pp.113.219006.

Feng, L., Wang, K., Li, Y., Tan, Y., Kong, J., Li, H., et al. (2007). Overexpression of SBPase enhances photosynthesis against high temperature stress in transgenic rice plants. *Plant Cell Rep*. 26, 1635-1646. doi: 10.1007/s00299-006-0299-y.

Fu, A., He, Z., Cho, H. S., Lima, A., Buchanan, B. B., and Luan, S. (2007). A chloroplast cyclophilin functions in the assembly and maintenance of photosystem II in Arabidopsis thaliana. *Proc Natl Acad Sci U S A*. 104, 15947-15952. doi: 10.1073/pnas.0707851104.

Gabriel, L. T., Michel, H., and Francois, O. (2010). The chloroplastic lipocalin AtCHL prevents lipid peroxidation and protects Arabidopsis against oxidative stress. *Plant Journal*. 60, 691-702. doi: 10.1111/j.1365-313X.2009.03991.x.

Ganeteg, U., Kulheim, C., Andersson, J., and Jansson, S. (2004). Is each light-harvesting complex protein important for plant fitness? *Plant Physiol*. 134, 502-509. doi: 10.1104/pp.103.033324.

Ge, Y., Wang, T., Wang, N., Wang, Z., Liang, C., Ramchiary, N., et al. (2012). Genetic mapping and localization of quantitative trait loci for chlorophyll content in Chinese cabbage (Brassica rapa ssp. pekinensis). *Scientia Horticulturae*. 147, 42–48. doi: 10.1016/j.scienta.2012.09.004.

Gupta, K., Sengupta, A., Chakraborty, M., and Gupta, B. (2016). Hydrogen peroxide and polyamines act as double edged swords in plant abiotic stress responses. *Front Plant Sci*. 7, 1343. doi: 10.3389/fpls.2016.01343.

Hara, M., and Kurita, I. (2014). The natural alkaloid sanguinarine promotes the expression of heat shock protein genes in Arabidopsis. *Acta Physiologiae Plantarum*. 36, 3337-3343. doi: 10.1007/s11738-014-1681-y.

Harper, A. L., Mckinney, L. V., Nielsen, L. R., Havlickova, L., Li, Y., Trick, M., et al. (2016a). Molecular markers for tolerance of European ash (Fraxinus excelsior) to dieback disease identified using Associative Transcriptomics. *Sci Rep*. 6, 19335. doi: 10.1038/srep19335.

Harper, A. L., Trick, M., He, Z., Clissold, L., Fellgett, A., Griffiths, S., et al. (2016b). Genome distribution of differential homoeologue contributions to leaf gene expression in bread wheat. *Plant Biotechnol J*. 14, 1207-1214. doi: 10.1111/pbi.12486.

Harper, A. L., Trick, M., Higgins, J., Fraser, F., Clissold, L., Wells, R., et al. (2012). Associative transcriptomics of traits in the polyploid crop species Brassica napus. *Nat Biotechnol*. 30, 798-802. doi: 10.1038/nbt.2302.

Havlickova, L., He, Z., Wang, L., Langer, S., Harper, A. L., Kaur, H., et al. (2018). Validation of an updated Associative Transcriptomics platform for the polyploid crop species Brassica napus by dissection of the genetic architecture of erucic acid and tocopherol isoform variation in seeds. *Plant J*. 93, 181-192. doi: 10.1111/tpj.13767.

He, Z., Cheng, F., Li, Y., Wang, X., Parkin, I. A., Chalhoub, B., et al. (2015). Construction of Brassica A and C genome-based ordered pan-transcriptomes for use in rapeseed genomic research. *Data Brief*. 4, 357-362. doi: 10.1016/j.dib.2015.06.016.

Hejna, O., Havlickova, L., He, Z., Bancroft, I., and Curn, V. (2019). Analysing the genetic architecture of clubroot resistance variation in Brassica napus by associative transcriptomics. *Mol Breed*. 39, 112. doi: 10.1007/s11032-019-1021-4.

Higgins, J., Magusin, A., Trick, M., Fraser, F., and Bancroft, I. (2012). Use of mRNA-seq to discriminate contributions to the transcriptome from the constituent genomes of the polyploid crop species Brassica napus. *BMC Genomics*. 13, 247. doi: 10.1186/1471-2164-13-247.

Hincha, D. K., Hofner, R., Schwab, K. B., Heber, U., and Schmitt, J. M. (1987). Membrane rupture is the common cause of damage to chloroplast membranes in leaves injured by freezing or excessive

735 wilting. *Plant Physiol*. 83, 251-253. doi: 10.1104/pp.83.2.251.
736 Jha, U. C., Bohra, A., and Jha, R. (2017). Breeding approaches and genomics technologies to increase
737 crop yield under low-temperature stress. *Plant Cell Rep*. 36, 1-35. doi:
738 10.1007/s00299-016-2073-0.
739 Jin, H., Liu, B., Luo, L., Feng, D., Wang, P., Liu, J., et al. (2014). HYPERSENSITIVE TO HIGH
740 LIGHT1 interacts with LOW QUANTUM YIELD OF PHOTOSYSTEM II1 and functions in
741 protection of photosystem II from photodamage in Arabidopsis. *Plant Cell*. 26, 1213-1229. doi:
742 10.1105/tpc.113.122424.
743 Ju, X., and Li, Z. (2012). Study of different types of rape (Brassica L.) cultivars (lines) relationship
744 between photosynthetic physiological indices and yield. *Journal of Anhui Agri.Sci*. 40,
745 11213-11215. doi: 10.13989/j.cnki.0517-6611.2012.22.106.
746 Kaler, A. S., and Purcell, L. C. (2019). Estimation of a significance threshold for genome-wide
747 association studies. *BMC Genomics*. 20, doi: 10.1186/s12864-019-5992-7
748 Koprivova, A., Harper, A. L., Trick, M., Bancroft, I., and Kopriva, S. (2014). Dissection of the control
749 of anion homeostasis by associative transcriptomics in Brassica napus. *Plant Physiol*. 166,
750 442-450. doi: 10.1104/pp.114.239947.
751 Kumar, A., Li, C., and Portis, A. R., Jr. (2009). Arabidopsis thaliana expressing a thermostable chimeric
752 Rubisco activase exhibits enhanced growth and higher rates of photosynthesis at moderately high
753 temperatures. *Photosynth Res*. 100, 143-153. doi: 10.1007/s11120-009-9438-y.
754 Kurek, I., Chang, T. K., Bertain, S. M., Madrigal, A., Liu, L., Lassner, M. W., et al. (2007). Enhanced
755 Thermostability of Arabidopsis Rubisco activase improves photosynthesis and growth rates under
756 moderate heat stress. *Plant Cell*. 19, 3230-3241. doi: 10.1105/tpc.107.054171.
757 Lawson, T., Kramer, D. M., and Raines, C. A. (2012). Improving yield by exploiting mechanisms
758 underlying natural variation of photosynthesis. *Curr Opin Biotechnol*. 23, 215-220. doi:
759 10.1016/j.copbio.2011.12.012.
760 Leister, D. (2019). Genetic engineering, synthetic biology and the light reactions of photosynthesis.
761 *Plant Physiol*. 179, 778-793. doi: 10.1104/pp.18.00360.
762 Li, H., Wang, G., Zheng, Q., Li, B., Jing, R., and Li, Z. (2014). Genetic analysis of biomass and
763 photosynthetic parameters in wheat grown in different light intensities. *J Integr Plant Biol*. 56,
764 594-604. doi: 10.1111/jipb.12174.
765 Li, H., Yang, Y., Zhang, H., Chu, S., Zhang, X., Yin, D., et al. (2016). A Genetic Relationship between
766 Phosphorus Efficiency and Photosynthetic Traits in Soybean As Revealed by QTL Analysis Using
767 a High-Density Genetic Map. *Front Plant Sci*. 7, 924. doi: 10.3389/fpls.2016.00924.
768 Liao, G., and Guan, C. (2001). Effect of seeding date on yield characteristics of different rapeseed
769 (Brassica napus) genotypes. *Chinese Journal of Applid Ecology*. doi: 853-858.
770 10.13287/j.1001-9332.2001.0203
771 Lipka, A. E., Tian, F., Wang, Q., Peiffer, J., Li, M., Bradbury, P. J., et al. (2012). GAPIT: genome
772 association and prediction integrated tool. *Bioinformatics*. 28, 2397-2399. doi:
773 10.1093/bioinformatics/bts444.
774 Liu, J., Lu, Y., Hua, W., and Last, R. L. (2019a). A new light on Photosystem II maintenance in
775 oxygenic photosynthesis. *Front Plant Sci*. 10, 975. doi: 10.3389/fpls.2019.00975.
776 Liu, M., Zhang, S., Hu, J., Sun, W., Padilla, J., He, Y., et al. (2019b). Phosphorylation-guarded
777 light-harvesting complex II contributes to broad-spectrum blast resistance in rice. *Proc Natl Acad*
778 *Sci U S A*. 116, 17572-17577. doi: 10.1073/pnas.1905123116.

Liu, R., Xu, Y. H., Jiang, S. C., Lu, K., Lu, Y. F., Feng, X. J., et al. (2013). Light-harvesting chlorophyll a/b-binding proteins, positively involved in abscisic acid signalling, require a transcription repressor, WRKY40, to balance their function. *J Exp Bot*. 64, 5443-5456. doi: 10.1093/jxb/ert307.

Liu, X., Fan, Y., Mak, M., Babla, M., Holford, P., Wang, F., et al. (2017). QTLs for stomatal and photosynthetic traits related to salinity tolerance in barley. *BMC Genomics*. 18, 9. doi: 10.1186/s12864-016-3380-0.

Livak, K. J., and Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods*. 25, 402-408. doi: 10.1006/meth.2001.1262.

Long, S. P., Zhu, X. G., Naidu, S. L., and Ort, D. R. (2006). Can improvement in photosynthesis increase crop yields? *Plant Cell Environ*. 29, 315-330. doi: 10.1111/j.1365-3040.2005.01493.x.

Lu, G., Harper, A. L., Trick, M., Morgan, C., Fraser, F., O'neill, C., et al. (2014). Associative transcriptomics study dissects the genetic architecture of seed glucosinolate content in Brassica napus. *DNA Res*. 21, 613-625. doi: 10.1093/dnares/dsu024.

Luo, T., Xian, M., Zhang, C., Zhang, C., Hu, L., and Xu, Z. (2019). Associating transcriptional regulation for rapid germination of rapeseed (Brassica napus L.) under low temperature stress through weighted gene co-expression network analysis. *Scientific Reports*. 9, doi: 10.1038/s41598-018-37099-0

Luo, T., Zhang, J., Khan, M. N., Liu, J., Xu, Z., and Hu, L. (2018). Temperature variation caused by sowing dates significantly affects floral initiation and floral bud differentiation processes in rapeseed (Brassica napus L.). *Plant Science*. 271, 40-51. doi: 10.1016/j.plantsci.2018.03.004.

Lv, Y., Guo, Z., Li, X., Ye, H., Li, X., and Xiong, L. (2016). New insights into the genetic basis of natural chilling and cold shock tolerance in rice by genome-wide association analysis. *Plant Cell Environ*. 39, 556-570. doi: 10.1111/pce.12635.

Lv, Y., Yang, M., Hu, D., Yang, Z., Ma, S., Li, X., et al. (2017). The OsMYB30 Transcription Factor Suppresses Cold Tolerance by Interacting with a JAZ Protein and Suppressing β-Amylase Expression. *Plant Physiology*. 173, 1475. doi: 10.1104/pp.16.01725.

Matsuoka, E., Matsubara, T., Takahashi, I., Murano, H., and Hara, M. (2016). The isoquinoline alkaloid sanguinarine which inhibits chaperone activity enhances the production of heat shock proteins in Arabidopsis. *Plant Biotechnol (Tokyo)*. 33, 409-413. doi: 10.5511/plantbiotechnology.16.1001a.

Miller, C. N., Harper, A. L., Trick, M., Wellner, N., Werner, P., Waldron, K. W., et al. (2018). Dissecting the complex regulation of lodging resistance in Brassica napus. *Mol Breed*. 38, 30. doi: 10.1007/s11032-018-0781-6.

Miller, C. N., Harper, A. L., Trick, M., Werner, P., Waldron, K., and Bancroft, I. (2016). Elucidation of the genetic basis of variation for stem strength characteristics in bread wheat by Associative Transcriptomics. *BMC Genomics*. 17, 500. doi: 10.1186/s12864-016-2775-2.

Mishra, A., Heyer, A. G., and Mishra, K. B. (2014). Chlorophyll fluorescence emission can screen cold tolerance of cold acclimated Arabidopsis thaliana accessions. *Plant Methods*. 10, 38. doi: 10.1186/1746-4811-10-38.

Miyagawa, Y., Tamoi, M., and Shigeoka, S. (2001). Overexpression of a cyanobacterial fructose-1,6-/sedoheptulose-1,7-bisphosphatase in tobacco enhances photosynthesis and growth. *Nat Biotechnol*. 19, 965-969. doi: 10.1038/nbt1001-965.

Morsy, M. R., Almutairi, A. M., Gibbons, J., Yun, S. J., and De Los Reyes, B. G. (2005). The OsLti6 genes encoding low-molecular-weight membrane proteins are differentially expressed in rice

cultivars with contrasting sensitivity to low temperature. *Gene*. 344, 171-180. doi: 10.1016/j.gene.2004.09.033.

Ning, J., Li, X., Hicks, L. M., and Xiong, L. (2010). A Raf-like MAPKKK gene DSM1 mediates drought resistance through reactive oxygen species scavenging in rice. *Plant Physiol*. 152, 876-890. doi: 10.1104/pp.109.149856.

Nishiyama, Y., Yamamoto, H., Allakhverdiev, S., Inaba, M., Yokota, A., and Murata, N. (2014). Oxidative stress inhibits the repair of photodamage to the photosynthetic machinery. *Embo Journal*. 20, 5587-5594. doi: 10.1093/emboj/20.20.5587.

Novillo, F., Alonso, J. M., Ecker, J. R., and Salinas, J. (2004). CBF2/DREB1C is a negative regulator of CBF1/DREB1B and CBF3/DREB1A expression and plays a central role in stress tolerance in Arabidopsis. *Proc Natl Acad Sci U S A*. 101, 3985-3990. doi: 10.1073/pnas.0303029101.

Novillo, F., Medina, J., Rodriguez-Franco, M., Neuhaus, G., and Salinas, J. (2012). Genetic analysis reveals a complex regulatory network modulating CBF gene expression and Arabidopsis response to abiotic stress. *J Exp Bot*. 63, 293-304. doi: 10.1093/jxb/err279.

O'neill, C. M., Lu, X., Calderwood, A., Tudor, E. H., and Penfield, S. (2019). Vernalization and floral transition in autumn drive winter annual life history in oilseed rape. *Current Biology*. 29, doi: 10.1016/j.cub.2019.10.051

Oakley, C. G., Savage, L., Lotz, S., Larson, G. R., Thomashow, M. F., Kramer, D. M., et al. (2018). Genetic basis of photosynthetic responses to cold in two locally adapted populations of Arabidopsis thaliana. *J Exp Bot*. 69, 699-709. doi: 10.1093/jxb/erx437.

Ozer, H. (2003). Sowing date and nitrogen rate effects on growth, yield and yield components of two summer rapeseed cultivars. *European Journal of Agronomy*. 19, 453-463. doi: 10.1016/S1161-0301(02)00136-3.

Prado, S. A., Cabrera-Bosquet, L., Grau, A., Coupel-Ledru, A., Millet, E. J., Welcker, C., et al. (2017). Phenomics allows identification of genomic regions affecting maize stomatal conductance with conditional effects of water deficit and evaporative demand. *Plant Cell & Environment*. doi: 10.1111/pce.13083

Sabehat, A., Weiss, D., and Lurie, S. (1996). The correlation between heat-shock protein accumulation and persistence and chilling tolerance in tomato fruit. *Plant Physiology*. 110, 531-537. doi: 10.1104/pp.110.2.531.

Sage, R., and Kubien, D. (2007). The temperature response of C 3 and C 4 photosynthesis. *Plant, cell & environment*. 30, 1086-1106. doi: 10.1111/j.1365-3040.2007.01682.x.

Sainz, M., Diaz, P., Monza, J., and Borsani, O. (2010). Heat stress results in loss of chloroplast Cu/Zn superoxide dismutase and increased damage to Photosystem II in combined drought-heat stressed Lotus japonicus. *Physiol Plant*. 140, 46-56. doi: 10.1111/j.1399-3054.2010.01383.x.

Savitch, L. V., Allard, G., Seki, M., Robert, L. S., Tinker, N. A., Huner, N. P., et al. (2005). The effect of overexpression of two Brassica CBF/DREB1-like transcription factors on photosynthetic capacity and freezing tolerance in Brassica napus. *Plant Cell Physiol*. 46, 1525-1539. doi: 10.1093/pcp/pci165.

Schlager, S., and Drager, B. (2016). Exploiting plant alkaloids. *Curr Opin Biotechnol*. 37, 155-164. doi: 10.1016/j.copbio.2015.12.003.

Schuster, T. M., Setaro, S. D., and Kron, K. A. (2013). Age estimates for the buckwheat family Polygonaceae based on sequence data calibrated by fossils and with a focus on the amphi-Pacific Muehlenbeckia. *PLoS One*. 8, e61261. doi: 10.1371/journal.pone.0061261.

Selmar, D., and Kleinwachter, M. (2013). Stress enhances the synthesis of secondary plant products: the impact of stress-related over-reduction on the accumulation of natural products. *Plant Cell Physiol*. 54, 817-826. doi: 10.1093/pcp/pct054.

Shen, B. R., Wang, L. M., Lin, X. L., Yao, Z., Xu, H. W., Zhu, C. H., et al. (2019). Engineering a new chloroplastic photorespiratory bypass to increase photosynthetic efficiency and productivity in rice. *Mol Plant*. 12, 199-214. doi: 10.1016/j.molp.2018.11.013.

Sirpio, S., Khrouchtchova, A., Allahverdiyeva, Y., Hansson, M., Fristedt, R., Vener, A. V., et al. (2008). AtCYP38 ensures early biogenesis, correct assembly and sustenance of photosystem II. *Plant J*. 55, 639-651. doi: 10.1111/j.1365-313X.2008.03532.x.

Srivastava, N. K., and Srivastava, A. K. (2010). Influence of some heavy metals on growth, alkaloid content and composition in Catharanthus roseus L. *Indian J Pharm Sci*. 72, 775-778. doi: 10.4103/0250-474X.84592.

Steponkus, P. L., Lynch, D. V., Uemura, M., Heber, U., and Pearce, R. S. (1990). The Influence of Cold Acclimation on the Lipid Composition and Cryobehaviour of the Plasma Membrane of Isolated Rye Protoplasts [and Discussion]. *Phil. Trans. R. Soc. Lond. B*. 326, 571-583. doi: 10.1098/rstb.1990.0032.

Strzepek, R. F., Boyd, P. W., and Sunda, W. G. (2019). Photosynthetic adaptation to low iron, light, and temperature in Southern Ocean phytoplankton. *Proc Natl Acad Sci U S A*. 116, 4388-4393. doi: 10.1073/pnas.1810886116.

Sun, S. N., Wang, Q., Sun, C. C., Liu, F. J., Bi, H. G., and Ai, X. Z. (2017). Response and adaptation of photosynthesis of cucumber seedlings to high temperature stress. *Ying Yong Sheng Tai Xue Bao*. 28, 1603-1610. doi: 10.13287/j.1001-9332.201705.009.

Takahashi, S., Bauwe, H., and Badger, M. (2007). Impairment of the photorespiratory pathway accelerates photoinhibition of photosystem II by suppression of repair but not acceleration of damage processes in Arabidopsis. *Plant Physiol*. 144, 487-494. doi: 10.1104/pp.107.097253.

Thalhammer, A., Hincha, D. K., and Zuther, E. (2014). Measuring freezing tolerance: electrolyte leakage and chlorophyll fluorescence assays. *Methods Mol Biol*. 1166, 15-24. doi: 10.1007/978-1-4939-0844-8_3.

Thalmann, M., and Santelia, D. (2017). Starch as a determinant of plant fitness under abiotic stress. *New Phytol*. 214, 943-951. doi: 10.1111/nph.14491.

Tian, Z., Ji, Y. H., Sun, L. X., Xu, X. L., Fan, D. L., Zhong, H. L., et al. (2018). Changes in production potentials of rapeseed in the Yangtze River Basin of China under climate change: A multi-model ensemble approach. *Journal of Geographical Sciences*. 28, 1700-1714. doi: 10.1007/s11442-018-1538-1

Tonfack, L., Moummou, H., Latché, A., Youmbi, E., Benichou, M., Pech, J.-C., et al. (2011). The plant SDR superfamily: Involvement in primary and secondary metabolism. *Current Topics in Plant Biology*. 12, 41-53. doi: 10.1186/1471-2229-12-219.

Urban, M. O., Klima, M., Vitamvas, P., Vasek, J., Hilgert-Delgado, A. A., and Kucera, V. (2013). Significant relationships among frost tolerance and net photosynthetic rate, water use efficiency and dehydrin accumulation in cold-treated winter oilseed rapes. *J Plant Physiol*. 170, 1600-1608. doi: 10.1016/j.jplph.2013.07.012.

Valerio, C., Costa, A., Marri, L., Issakidis-Bourguet, E., Pupillo, P., Trost, P., et al. (2011). Thioredoxin-regulated beta-amylase (BAM1) triggers diurnal starch degradation in guard cells, and in mesophyll cells under osmotic stress. *J Exp Bot*. 62, 545-555. doi: 10.1093/jxb/erq288.

Voll, L. M., Jamai, A., Renne, P., Voll, H., Mcclung, C. R., and Weber, A. P. (2006). The photorespiratory Arabidopsis shm1 mutant is deficient in SHM1. *Plant Physiol*. 140, 59-66. doi: 10.1104/pp.105.071399.

Von Caemmerer, S., Hendrickson, L., Quinn, V., Vella, N., Millgate, A. G., and Furbank, R. T. (2005). Reductions of Rubisco activase by antisense RNA in the C4 plant Flaveria bidentis reduces Rubisco carbamylation and leaf photosynthesis. *Plant Physiol*. 137, 747-755. doi: 10.1104/pp.104.056077.

Wanasundara, P., Mcintosh, T., Perera, S., Withana-Gamage, T., and Mitra, P. (2016). Canola/rapeseed protein-functionality and nutrition. *OCL*. 23, D407. doi: 10.1051/ocl/2016028.

Wang, J., Jian, H., Wei, L., Qu, C., Xu, X., Lu, K., et al. (2015). Genome-wide analysis of seed acid detergent lignin (ADL) and hull content in rapeseed (Brassica napus L.). *PLoS One*. 10, e0145045. doi: 10.1371/journal.pone.0145045.

Wood, I. P., Pearson, B. M., Garcia-Gutierrez, E., Havlickova, L., He, Z., and Harper, A. L. (2017). Carbohydrate microarrays and their use for the identification of molecular markers for plant cell wall composition. *Proc Natl Acad Sci U S A*. 114, 6860-6865. doi: 10.1073/pnas.1619033114.

Xu, Y. H., Liu, R., Yan, L., Liu, Z. Q., Jiang, S. C., Shen, Y. Y., et al. (2012). Light-harvesting chlorophyll a/b-binding proteins are required for stomatal response to abscisic acid in Arabidopsis. *J Exp Bot*. 63, 1095-1106. doi: 10.1093/jxb/err315.

Yan, L., Tariq, S., Cheng, Y., Lü, Y., Zhang, X. K., and Zou, X. L. (2019). Physiological and molecular responses to cold stress in rapeseed (Brassica napus L.). *Journal of Integrative Agriculture*. 18, 2742-2752. doi: 10.1016/S2095-3119(18)62147-1

Yan, X., Qu, C., Li, J., Chen, L., and Liu, L. (2015). QTL analysis of leaf photosynthesis rate and related physiological traits in in brassica napus. *Sci. Agric. Sin*. 14, 2095-3119. doi: 10.1016/S2095-3119(14)60958-8.

Yin, Z., Meng, F., Song, H., Wang, X., Xu, X., and Yu, D. (2010). Expression quantitative trait loci analysis of two genes encoding rubisco activase in soybean. *Plant Physiol*. 152, 1625-1637. doi: 10.1104/pp.109.148312.

Zhang, L., Ding, R., Chai, Y., Bonfill, M., Moyano, E., Oksman-Caldentey, K.-M., et al. (2004). Engineering tropane biosynthetic pathway in Hyoscyamus niger hairy root cultures. *Proc Natl Acad Sci U S A*. 101, 6786. doi: 10.1073/pnas.0401391101.

Zhang, S. J., Li, L., and Zhang, C. L. (2012). Effects of sowing date and planting density on the seed yield and oil content of winter oilseed rape. *Ying Yong Sheng Tai Xue Bao*. 23, 1326-1332. doi: 10.13287/j.1001-9332.2012.0179.

Zhang, X., Feng, L., Yang, T., Xu, Z., and Hu, L. (2015). Effects of Chilling Stress on Physiological Characteristics of Rapeseed Seedlings in Winter. *Plant Physiology Journal*. 51, 737-746. doi: 10.13592/j.cnki.ppj.2014.0411.

Zhang, X., Wang, L., Meng, H., Wen, H., Fan, Y., and Zhao, J. (2011). Maize ABP9 enhances tolerance to multiple stresses in transgenic Arabidopsis by modulating ABA signaling and cellular levels of reactive oxygen species. *Plant Molecular Biology*. 75, 365-378. doi: 10.1007/s11103-011-9732-x.

Zhang, X. R., Henriques, R., Lin, S. S., Niu, Q. W., and Chua, N. H. (2006). Agrobacterium-mediated transformation of Arabidopsis thaliana using the floral dip method. *Nature Protocols*. 1, 641-646. doi: 10.1038/nprot.2006.97.

Zhang, Y., Zhang, D., Yu, H., Lin, B., Fu, Y., and Hua, S. (2016). Floral Initiation in Response to Planting Date Reveals the Key Role of Floral Meristem Differentiation Prior to Budding in Canola

(Brassica napus L.). *Front Plant Sci*. 7, 1369. doi: 10.3389/fpls.2016.01369.

Zhang, Y., Zhao, X., Guan, Z., Wang, X., Hou, J., and Tian, J. (2017a). A review on photosynthetic physiological research and high photosynthetic efficiency breeding of Brassica napus. *Chin. Agri. Sci. Bull*. 33, 44-54.

Zhang, Z., Li, J., Pan, Y., Li, J., Zhou, L., Shi, H., et al. (2017b). Natural variation in CTB4a enhances rice adaptation to cold habitats. *Nat Commun*. 8, 14788. doi: 10.1038/ncomms14788.

Zhong, L., Zhou, W., Wang, H., Ding, S., Lu, Q., Wen, X., et al. (2013). Chloroplast small heat shock protein HSP21 interacts with plastid nucleoid protein pTAC5 and is essential for chloroplast development in Arabidopsis under heat stress. *Plant Cell*. 25, 2925-2943. doi: 10.1105/tpc.113.111229.

966 **FIGURE LEGENDS**

967 **Figure 1.** Phenotypic variation of photosynthetic gas exchange parameters in 123
968 rapeseed accessions. Trait definition: Net photosynthesis rate (A_n), Stomatal
969 conductance to water vapour (G_{sw}), Internal CO_2 concentration (C_i), Transpiration rate
970 (E).

971

972 **Figure 2.** Manhattan plots for AT analysis in 123 rapeseed accessions. Manhattan
973 plots from left to right, represented for A_n, G_{sw}, C_i and E using SNPs (upper section)
974 and GEMs (bottom section), respectively. The -log10 (*P*-values) were plotted against
975 the position of the SNPs or GEMs on 19 chromosomes of *Brassica napus*. The black
976 line represents the -log10 (*P*-values) converted Bonferroni significance threshold for
977 SNP (5.41) and GEM (4.73), respectively.

978

979 **Figure 3.** Expression and allelic variation of *BnTR1* in rapeseed. (A) Expression
980 analysis of *BnTR1* (homolog of *Cab026133.1*) in six accessions corresponding to Hap
981 1 and Hap 2 under freezing conditions. The name of accessions 1-6 was Sv706118,
982 Kajsa, Callypso, Libritta, Gefion and Jupiter, respectively. *ACTIN* gene was used as
983 an internal control. Bars indicate the SE of three biological replicates. Different letters
984 indicate significant differences at $P< 0.05$ (one-way ANOVA with Tukey's multiple
985 comparisons test). (B) Correlation analysis between Transpiration rate (E) value and
986 expression level of *Cab026133.1* in the association panel (n=123). R^2 indicates the
987 coefficient of determination in linear regression. (C-D) Allelic variations at *BnTR1*
988 formed two main haplotypes and their effects on E value.

989

990 **Figure 4.** BnTR1 confers freezing tolerance in *Arabidopsis*. (A) Expression analysis
991 of the *BnTR1* transgenic plants (L1, L3, L5) and WT plants under normal condition
992 (i.e. 23°C). (B) Investigation of transpiration rate (E) value in the *BnTR1* transgenic
993 lines and WT plants under normal conditions. (C) Performance of the transgenic lines
994 and WT plants before and after freezing treatment (-4°C for 4 h). Scale=2 cm. (D)
995 Survival rates of the transgenic lines and WT plants after freezing treatment. (E-I)

Relative expression levels of *CBF1* (E), *CBF2* (F), *CBF3* (G), *COR15* (H), *RD29A* (I) in the transgenic lines and WT plants before and after freezing stress with the *Arabidopsis ACTIN* gene used as an internal control. Normal represents 23°C, freezing treatment represents 4 h at -4°C, recovery represents 3 d of recovery at 23°C. Bars indicate the SE of three biological replicates. Significant differences are determined by Student's *t*-test (**P*< 0.05, or ***P*< 0.01).

Figure 5. Variation of photosynthetic related traits and genes expression pattern in *BnTR1* transgenic plants. (A) Fv/Fm ratio in the transgenic lines and WT plants under freezing stress conditions. (B-H) Relative expression levels of *BnTR1* (B), *RCA* (C), *SBPASE* (D), *CAB1* (E), *CAB2* (F), *CAB3* (G), *CAB4* (H) in the transgenic lines and WT plants before and after freezing stress treatment with *Arabidopsis ACTIN* gene used as an internal control. L1, L3, L5 represent three independent homozygous lines of *BnTR1* transgenic plants. Bars indicate the SE of three biological replicates. Significant differences are determined by Student's *t*-test (**P*< 0.05, or ***P* < 0.01).

Figure 6. Physiological characterization of *BnTR1* transgenic plants under freezing stress conditions. (A-F) Investigation of the proline content (A), soluble sugar content (B), relative leakage (C), DAB staining analysis (D), H_2O_2 content (E), SOD activity (F) in the transgenic lines and WT plants under freezing stress conditions. L1, L3, L5 represent three independent homozygous lines of *BnTR1* transgenic *Arabidopsis* plants. Bars indicate the SE of three biological replicates. Significant differences are determined by the Student's *t*-test (**P*< 0.05, or ***P*< 0.01).

Figure 7. BnTR1 mediates alkaloid accumulation and exogenous atropine application enhances freezing tolerance. (A) Total alkaloids accumulation in *BnTR1* transgenic lines and WT plants under freezing stress conditions. L1, L3, L5 represent three independent homozygous lines of *BnTR1* transgenic *Arabidopsis* plants. (B) Phenotypes of *Arabidopsis* WT plants with exogenous atropine application (0 nmol per plant, 10 nmol per plant, 30 nmol per plant) under freezing stress conditions.

Scale=2 cm. (C) Survival rates of *Arabidopsis* WT plants with exogenous atropine application after the freezing treatment. (D) Phenotypes of rapeseed WT plants with exogenous atropine application (0 nmol per plant, 50 nmol per plant, 150 nmol per plant) under LT conditions. Scale=5 cm. Bars indicate the SE of three biological replicates. Significant differences are determined by Student's *t*-test (**P*< 0.05, or ***P*< 0.01).

Figure 1.JPEG

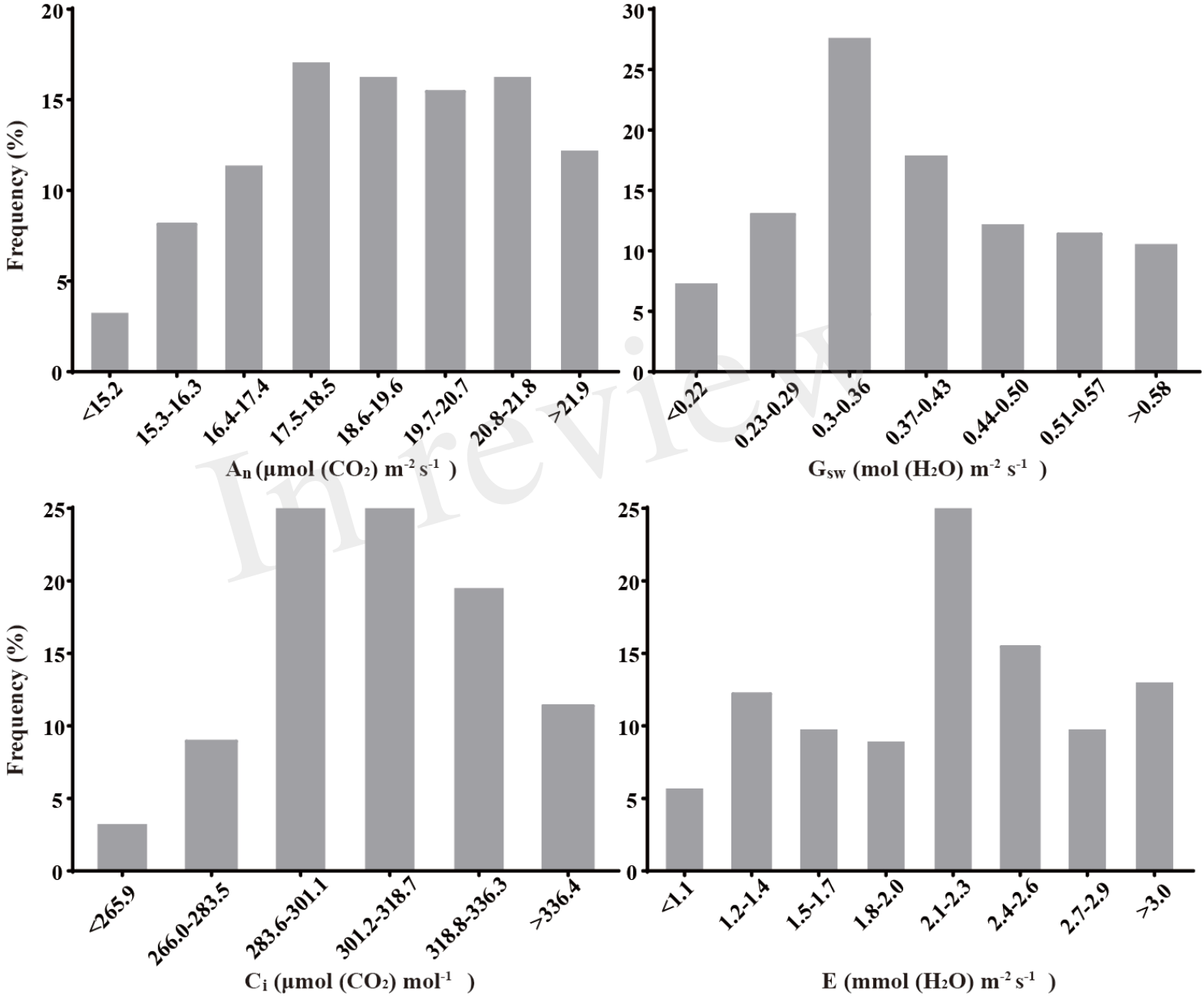

Figure 2.JPEG

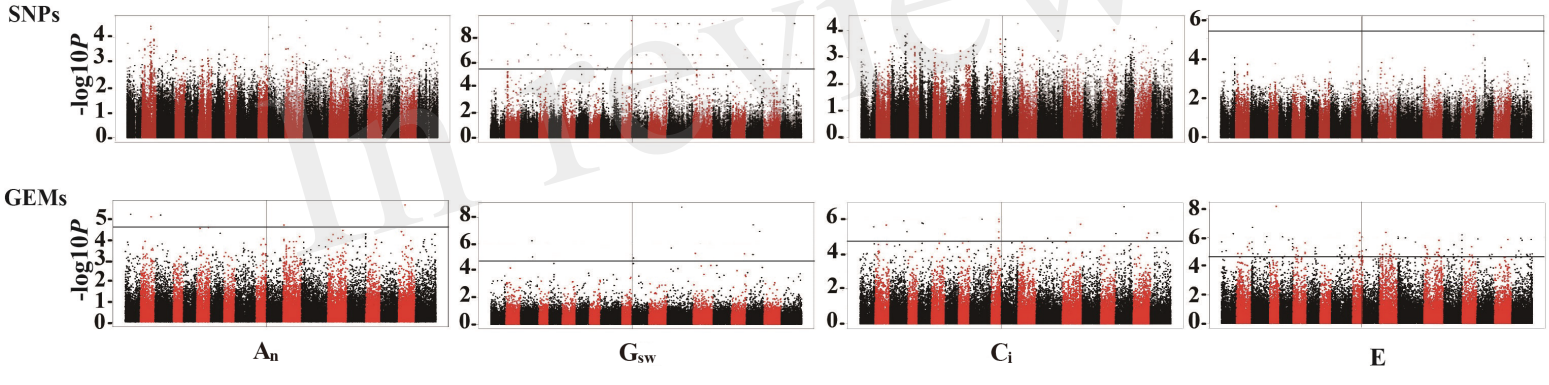

Figure 3.JPEG

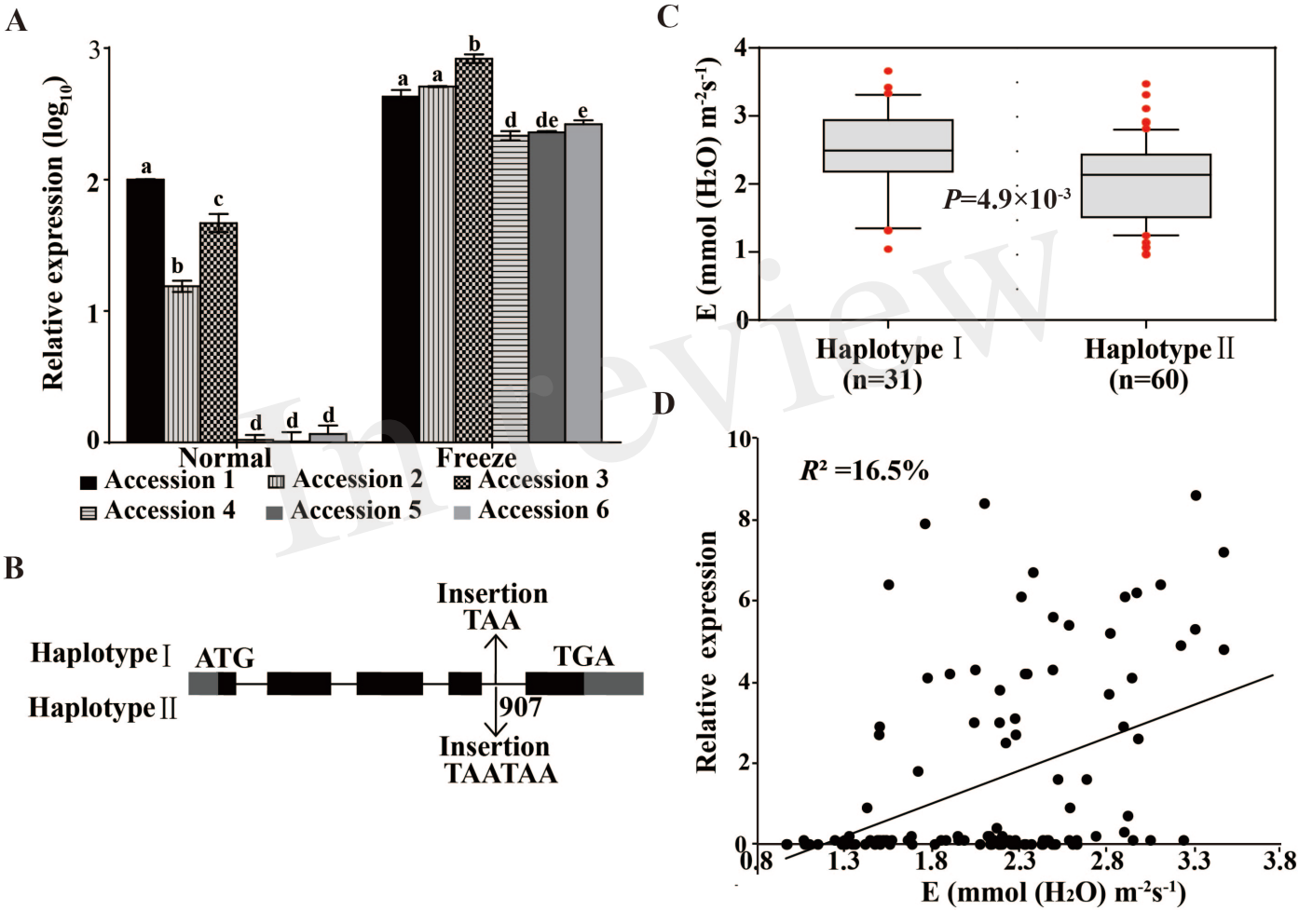

Figure 4.JPEG

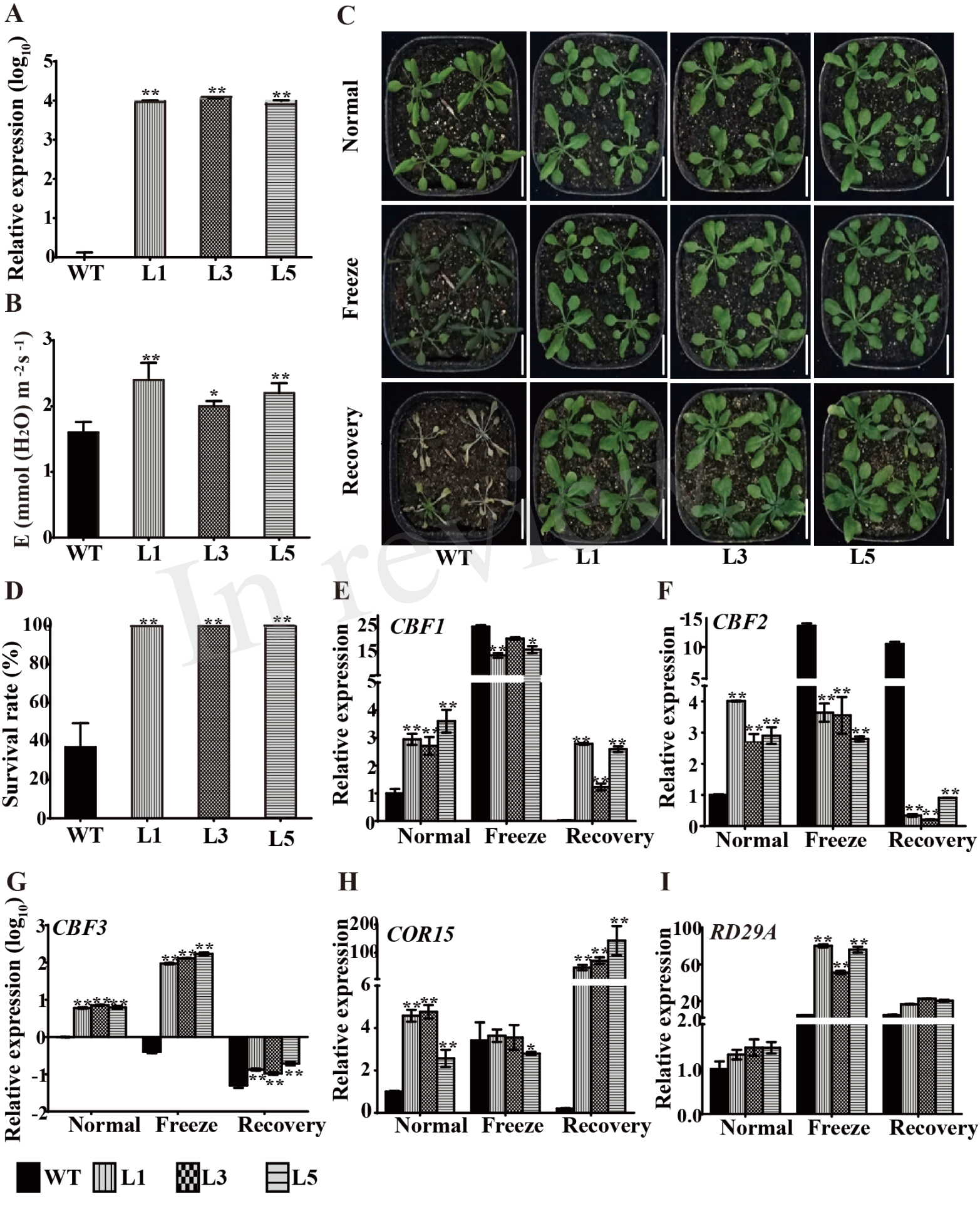

Figure 5.JPEG

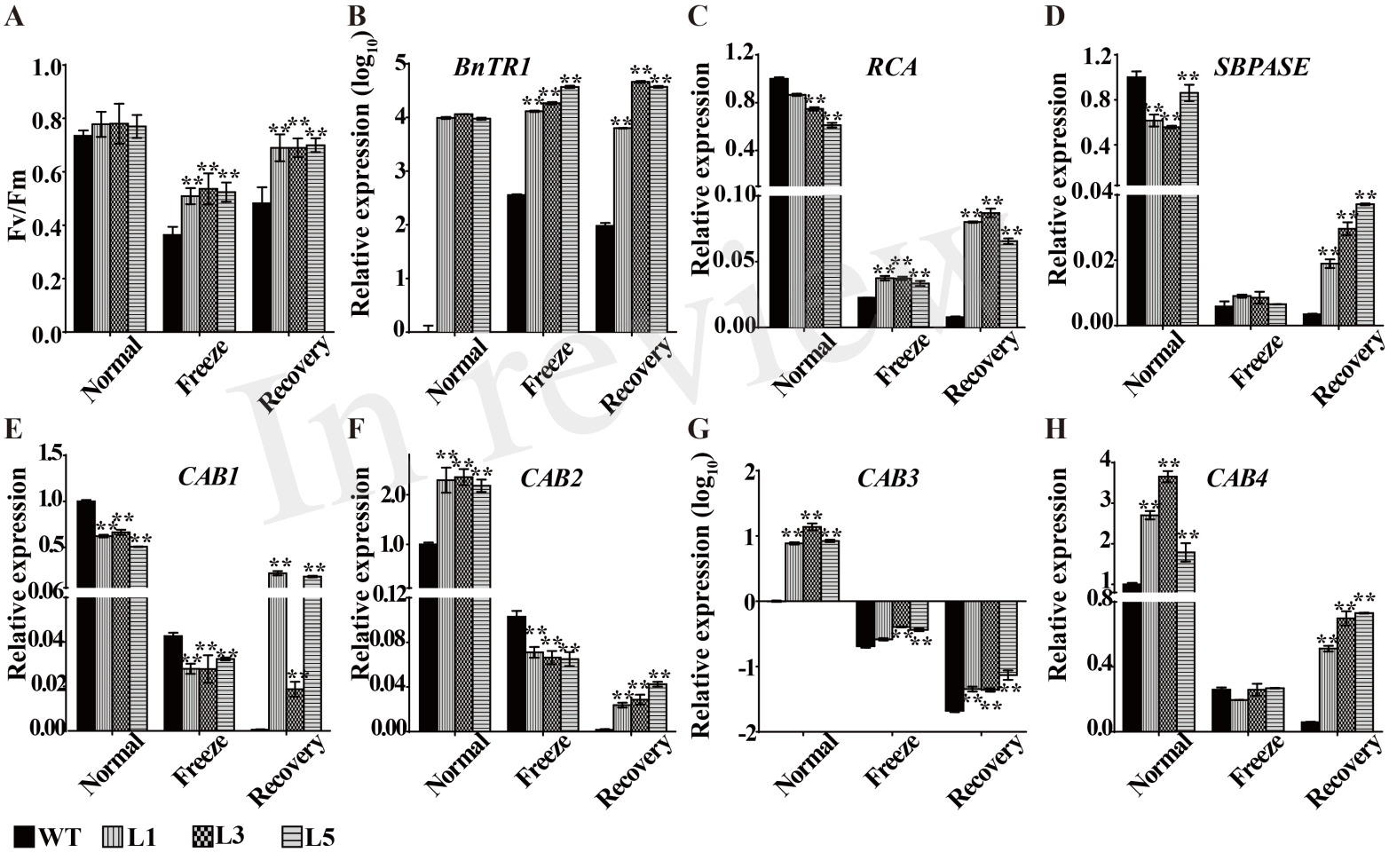

Figure 6.JPEG

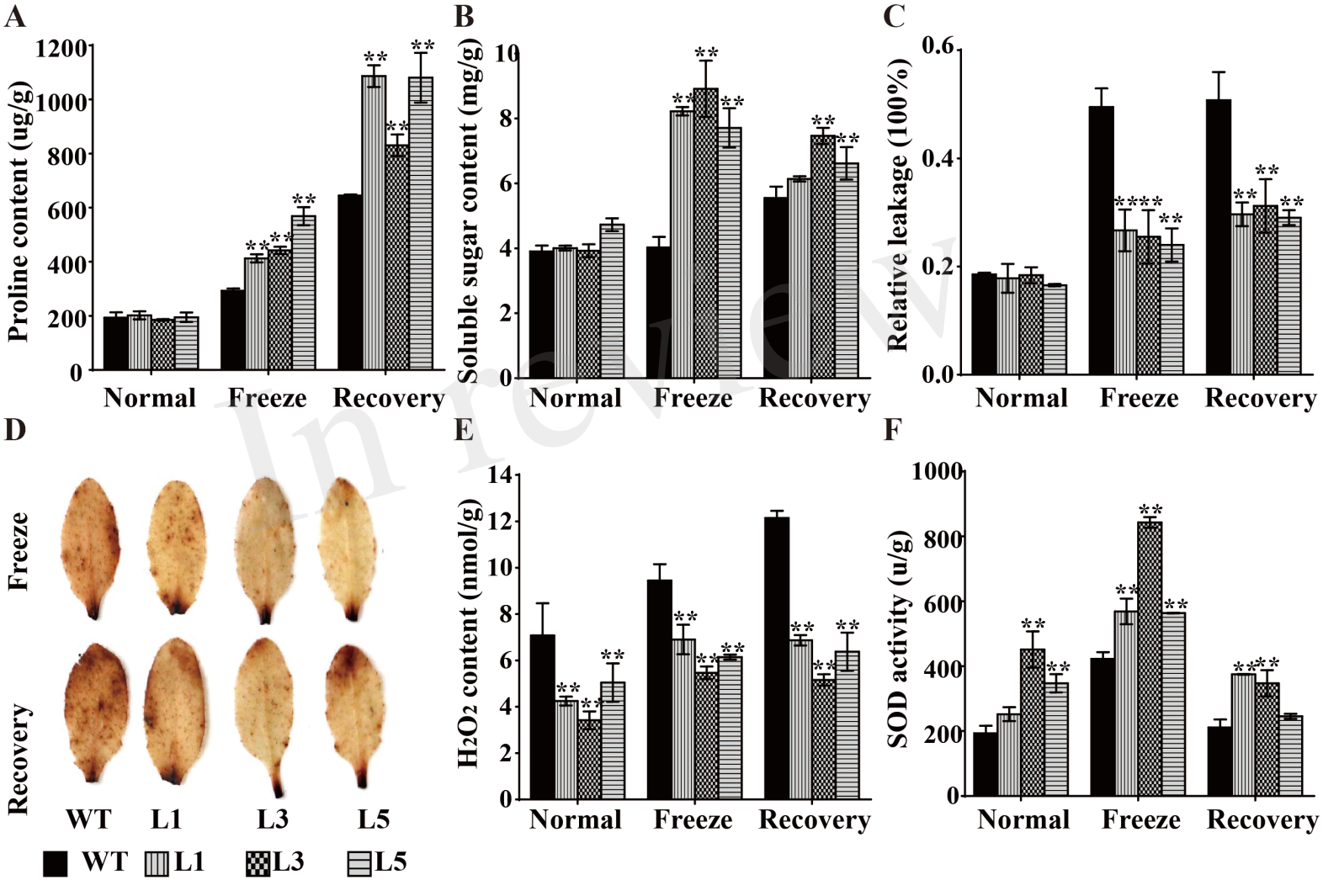

Figure 7.JPEG