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1 Reply to Comment Letter

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6 To the Editor

7 In a Letter to the Editor, Tenover *et al* suggest that on-label testing with nucleic acid amplification tests 8 (NAATs) for the diagnosis of *Clostridioides difficile* infection (CDI) is discussed, to put toxin testing in 9 perspective. The authors argue that guidelines from the Infectious Diseases Society of American-Society 10 for Healthcare Epidemiology of America (IDSA-SHEA)(1), the American Society for Microbiology (ASM) 11 (2), and the European Society for Clinical Microbiology and Infectious Diseases (ESCMID) (3), "all make it 12 clear that NAATs play an essential role in the laboratory diagnosis of CDI." This statement is potentially 13 misleading. Firstly, the ESCMID guidelines state that "using NAAT as a stand-alone test and relying on 14 clinical symptoms to discern patients with CDI from asymptomatic carriers is not an optimal approach: 15 patients colonized by a toxigenic C. difficile strain may very well develop diarrhea due to other causes 16 (3)." Secondly, the systematic review from ASM evaluated performance compared to detection of C. 17 difficile organism/toxin/toxin and not clinical diagnosis (2). However, this systematic review somewhat 18 disingenuously evaluated only testing methods and algorithms including NAAT, and excluded key studies 19 that have demonstrated the clinical value of toxin based testing (4, 5). Tenover et al claim that ASM 20 guidelines "endorses a role for stand-alone NAATs for CDI," while the ASM authors state that the use of 21 NAAT alone is recommended best practice "for the detection of the C. difficile toxin gene or organism 22 (2)." The two statements are certainly not synonymous. Thirdly, the IDSA-SHEA guidelines do

23 recommend that NAAT can be used alone, but only when there are pre-agreed institutional criteria for 24 patient stool submission (weak recommendation, low quality of evidence). These guidelines also clearly 25 recommend an algorithm approach to CDI diagnosis that includes toxin testing. The clinical criteria, unexplained and new-onset ≥3 unformed stools in 24 hours in patients not receiving laxatives, are 26 27 discussed in the guidelines: "some of these conditions and interventions associated with diarrhea in 28 their own right, [...], have been shown to have increased risk of CDI. So, in practice it is difficult to 29 exclude the possibility of CDI on clinical grounds alone in a patient with new-onset or worsened diarrhea 30 (1)."

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32 Tenover et al agree that using NAAT for the diagnosis of CDI leads to overdiagnosis, but only if the 33 clinical criteria for testing are not met. The authors further argue that any diagnostic C. difficile assay can 34 be positive in asymptomatic carriers. In a recent study, it was shown that the proportion of CDI 35 overdiagnosis was over three times higher in NAAT+/toxin- than in NAAT+/toxin+ patients when an 36 ultrasensitive toxin assays was used for CDI diagnosis, in an institution where rigorous stool-submission 37 criteria were recently successfully set in place (6, 7). CDI is a toxin-mediated disease and, although it has 38 been known for decades that toxin-producing strains can be present in asymptomatic carriers (8, 9), 39 presence of toxins better correlate with disease and outcome than presence of toxin genes (4–6). 40 Hospital-onset diarrhea is a common condition and importantly a recent large study showed that the 41 majority (85%) have multiple possible causes (median 3; IQR 2-5) (10). Thus, reliance on NAAT alone for 42 the diagnosis of CDI will still lead to overdiagnosis even if clinical criteria are used to guide who/when to 43 test.

A number of NAAT qualities are highlighted in the Letter: speed, sensitivity, high negative predictive
value, and cost-effectiveness when used appropriately (although the latter can be debated). The authors

have, rightfully so, left out high clinical specificity and high positive predictive value, both critical
components of any diagnostic test. Surprisingly, Tenover *et al* close their Letter with the statement "we
simply note that three recent guidelines support the value of NAATs for diagnosing CDI, while none
indicate a role of 'ultrasensitive' toxin tests." Notably, however, an ultrasensitive toxin test was not
commercially available at the time of publication of these guidelines, making such a recommendation
impossible.

52 **References**

53	1.	McDonald LC, Gerding DN, Johnson S, Bakken JS, Carroll KC, Coffin SE, Dubberke ER, Garey KW,
54		Gould CV, Kelly C, Loo V, Shaklee Sammons J, Sandora TJ, Wilcox MH. 2018. Clinical Practice
55		Guidelines for Clostridium difficile Infection in Adults and Children: 2017 Update by the Infectious
56		Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA).
57		Clin Infect Dis Off Publ Infect Dis Soc Am.
58	2.	Kraft CS, Parrott JS, Cornish NE, Rubinstein ML, Weissfeld AS, McNult P, Nachamkin I, Humphries
59		RM, Kirn TJ, Dien Bard J, Lutgring JD, Gullett JC, Bittencourt CE, Benson S, Bobenchik AM, Sautter
60		RL, Baselski V, Atlas MC, Marlowe EM, Miller NS, Fischer M, Richter SS, Gilligan P, Snyder JW. 2019.
61		A Laboratory Medicine Best Practices Systematic Review and Meta-analysis of Nucleic Acid
62		Amplification Tests (NAATs) and Algorithms Including NAATs for the Diagnosis of Clostridioides
63		(Clostridium) difficile in Adults. Clin Microbiol Rev 32.
64	3.	Crobach MJT, Planche T, Eckert C, Barbut F, Terveer EM, Dekkers OM, Wilcox MH, Kuijper EJ. 2016.
65		European Society of Clinical Microbiology and Infectious Diseases: update of the diagnostic
66		guidance document for Clostridium difficile infection. Clin Microbiol Infect Off Publ Eur Soc Clin
67		Microbiol Infect Dis 22 Suppl 4:S63-81.
68	4.	Planche TD, Davies KA, Coen PG, Finney JM, Monahan IM, Morris KA, O'Connor L, Oakley SJ, Pope
69		CF, Wren MW, Shetty NP, Crook DW, Wilcox MH. 2013. Differences in outcome according to
70		Clostridium difficile testing method: a prospective multicentre diagnostic validation study of C
71		difficile infection. Lancet Infect Dis 13:936–945.
72	5.	Polage CR, Gyorke CE, Kennedy MA, Leslie JL, Chin DL, Wang S, Nguyen HH, Huang B, Tang Y-W,
73		Lee LW, Kim K, Taylor S, Romano PS, Panacek EA, Goodell PB, Solnick JV, Cohen SH. 2015.

74 Overdiagnosis of Clostridium difficile Infection in the Molecular Test Era. JAMA Intern Med
75 175:1792–1801.

- Sandlund J, Estis J, Katzenbach P. 2019. Evaluation of the Singulex Clarity C. diff Toxins A/B Assay
 for Diagnosis of Clostridioides difficile Infection. ASM Microbe. 2019.
- 78 7. Chow S-K, Naderpour A, Van Enk J. 2018. It Is Not about the Assay: Preanalytical Screening Is the
 79 Key to Reducing *Clostridioides difficile* Infection. J Clin Microbiol 57.
- 80 8. Kyne L, Warny M, Qamar A, Kelly CP. 2000. Asymptomatic carriage of Clostridium difficile and
- 81 serum levels of IgG antibody against toxin A. N Engl J Med 342:390–397.
- Kyne L, Warny M, Qamar A, Kelly CP. 2001. Association between antibody response to toxin A and
 protection against recurrent Clostridium difficile diarrhoea. Lancet Lond Engl 357:189–193.
- 10. Mawer D, Byrne F, Drake S, Brown C, Prescott A, Warne B, Bousfield R, Skittrall JP, Ramsay I,
- 85 Somasunderam D, Bevan M, Coslett J, Rao J, Stanley P, Kennedy A, Dobson R, Long S, Obisanya T,
- 86 Esmailji T, Petridou C, Saeed K, Brechany K, Davis-Blue K, O'Horan H, Wake B, Martin J,
- 87 Featherstone J, Hall C, Allen J, Johnson G, Hornigold C, Amir N, Henderson K, McClements C, Liew I,
- 88 Deshpande A, Vink E, Trigg D, Guilfoyle J, Scarborough M, Scarborough C, Wong THN, Walker T,
- 89 Fawcett N, Morris G, Tomlin K, Grix C, O'Cofaigh E, McCaffrey D, Cooper M, Corbett K, French K,
- 90 Harper S, Hayward C, Reid M, Whatley V, Winfield J, Hoque S, Kelly L, King I, Bradley A, McCullagh
- 91 B, Hibberd C, Merron M, McCabe C, Horridge S, Taylor J, Koo S, Elsanousi F, Saunders R, Lim F,
- 92 Bond A, Stone S, Milligan ID, Mack DJF, Nagar A, West RM, Wilcox MH, Kirby A, Sandoe J a. T. 2019.
- 93 Cross-sectional study of the prevalence, causes and management of hospital-onset diarrhoea. J
- 94 Hosp Infect.