



UNIVERSITY OF LEEDS

This is a repository copy of *Fabrication of Multi-Layered Bone Scaffolds using Femtosecond Pulsed Lasers*.

White Rose Research Online URL for this paper:  
<http://eprints.whiterose.ac.uk/146035/>

Version: Accepted Version

---

**Conference or Workshop Item:**

Iqbal, N [orcid.org/0000-0002-2801-707X](https://orcid.org/0000-0002-2801-707X), Anastasiou, A, Maddi, C et al. (3 more authors) (2019) Fabrication of Multi-Layered Bone Scaffolds using Femtosecond Pulsed Lasers. In: Biophotonics Congress: Optics in the Life Sciences Congress 2019, Tucson, Arizona, USA.

---

This is an author produced version of presentation made at the Biophotonics Congress 2019 and published by the Optical Society of America. One print or electronic copy may be made for personal use only. Systematic reproduction and distribution, duplication of any material in this paper for a fee or for commercial purposes, or modifications of the content of this paper are prohibited.

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>

# Fabrication of Multi-layered Bone Scaffolds using Femtosecond Pulsed Lasers

Neelam Iqbal<sup>1</sup>, Antonios Anastasiou<sup>1</sup>, Chiranjeevi Maddi<sup>1</sup>, Mostafa El-Raif<sup>2</sup>, Peter V Giannoudis<sup>3</sup> and Animesh Jha<sup>1</sup>

<sup>1</sup> School of Chemical and Processing Engineering, Engineering Building, University of Leeds, LS2 9JT, UK

<sup>2</sup> Division of Oral Biology, Leeds Dental School, University of Leeds, UK

<sup>3</sup> Department of Trauma and Orthopaedic Surgery, Leeds General Infirmary, UK  
pml5ni@leeds.ac.uk



UNIVERSITY OF LEEDS

## Clinical Need

Healing of fractured bone is a complicated process therefore an ideal bone scaffold should have:

- exceptional osteogenic potential,
- load-bearing properties,
- promote angiogenesis for circulation of nutrients,
- antibacterial resistance to avoid infections that lead to failure of the surgery.

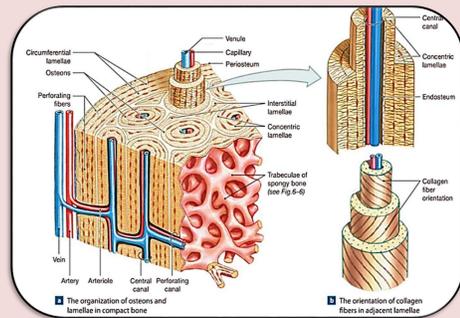


Fig. 1. Complex Concentric Structure of Bone

The structure of natural bone is complex and difficult to mimic (Fig.1). Current bone scaffolds do not possess the correct structure and environment to encourage mineralization and vascularization collectively.

## Aim

To fabricate **multi-layered** bone scaffold which has the potential to promote **bone mineralization, intrinsic vascularisation** and to provide **antibacterial properties**.

## Materials

We developed two types of chitosan mineral loaded samples.

The **Type-1** sample, aims to enhance osteogenic potential, it is loaded with calcium phosphate minerals (e.g.  $\beta$ -calcium pyrophosphate) and is fabricated through freeze drying in order to achieve a highly porous honeycomb structure (Fig.2.) that will promote osteoblast growth and proliferation.

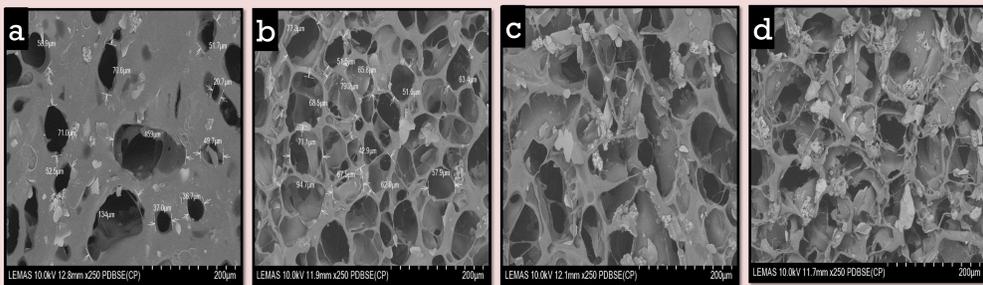


Fig.2. Comparison of Hitachi SU8230 SEM images of Type-1 freeze dried sample containing calcium phosphate minerals, (a) 20 wt%, (b) 30 wt%, (c) 40 wt%, and (d) 50 wt%.

The Type-2 samples are membrane geometries, fabricated through casting of chitosan solutions loaded with  $CeO_2$  and  $Ce_2O_3$  nanoparticles (NPs) which are known for their excellent antibacterial and angiogenic properties (Fig.3) already observed in systems for in vitro and also in vivo models [2]. The membranes were laser micropatterned to enhance angiogenesis by allowing guided cell growth throughout the scaffold.

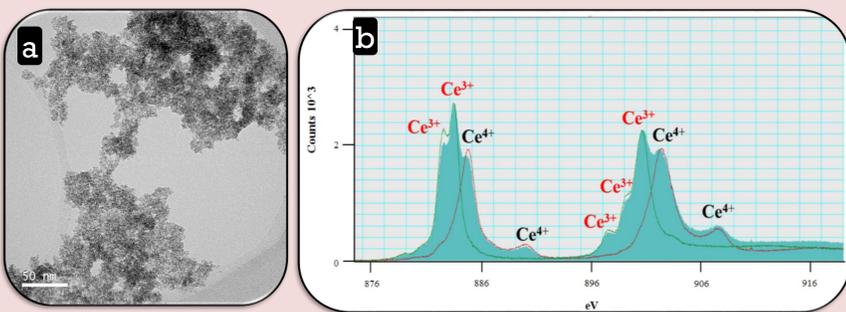


Fig.3. (a) TEM images of cerium oxide nanoparticle; (b) EELs analysis of Type-A nanoparticles depicting the presence of  $Ce^{3+}$  and  $Ce^{4+}$  oxidation states.

The scaffold can be fabricated through a **numbering up** approach where several layers of Type-1 and Type-2 samples are piled up together in order to mimic the natural structure of cancellous bone.

## Conclusions

○ Fabrication of microchannels on the membrane surfaces were successful, as can be seen in fig.4. Microchannels were created without damaging the surrounding material. The channel width sizes ranged between  $30 \mu m$  and  $140 \mu m$  for laser power between 40 mW and 220 mW.

- We fabricated 2 types of mineral loaded samples, when layered together the scaffold induces mineralization, enhances angiogenesis but also has antibacterial potential.
- Human osteoblast cells were successful grown on Type-1 samples indicating the potential for bone generation and mineralisation which is essential for bone remodelling.
- Type-2 samples containing cerium oxide NPs have the potential to reduce the prevalence of bacteria during and after bone scaffold implantation. This is linked to the samples ability to resorb allowing the NPs to be released at a steady rate, the NPs will aid in minimizing or preventing infection.

## Femtosecond Pulsed Laser

A Coherent Libra-S-1K femtosecond laser (1 kHz repetition rate, 100 fs pulse duration, wavelength of 800 nm) was utilized to fabricate microchannel networks and patterns on the membrane surfaces depicted (Fig.4.).

- Ablation threshold of Type-2 materials; this was determined to be  $5.4 J/cm^2$ .
- Initial laser power was set at 40 mW thus consecutive channels were formed by gradually increasing the power in steps of 20 mW per line (maximum power used was 220 mW).

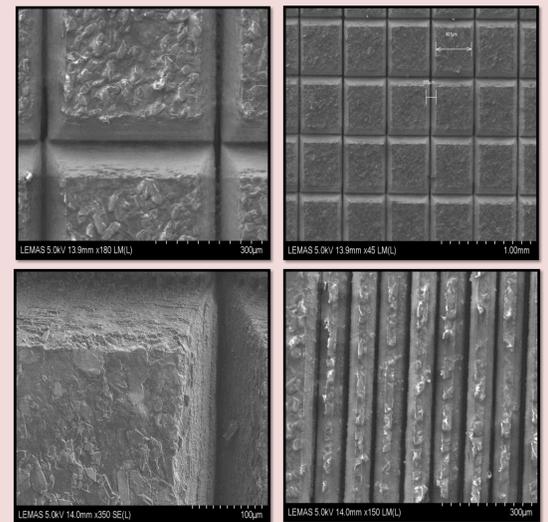


Fig.4. Hitachi SU8230 SEM images of consistent micro-networks created with the aid of a femtosecond laser (1kHz) upon chitosan membrane containing 1g of mineral.

## Results

During the synthesis of the nanoparticles it was

observed that the drying method (i.e. freeze drying or furnace drying) significantly affected the physicochemical properties, size, shape and the  $Ce^{4+}$  to  $Ce^{3+}$  ratio.

- Freeze dried NPs were successfully fabricated with particle sizes between 4nm and 8nm whereas the particle sizes of furnace dried NPs were between 15nm and 20 nm.

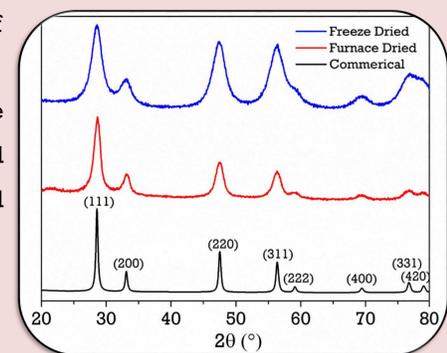


Fig.5. X-ray diffractometer spectrum of cerium oxide NPs, 2 $\theta$  scanning range was 20° to 80° at a scan speed of 5s and increment of 0.03.

- For **Type-2** samples the initial rate of resorption was significantly high for all the membranes, however; as time progressed the rate began to stabilize (Fig.6).

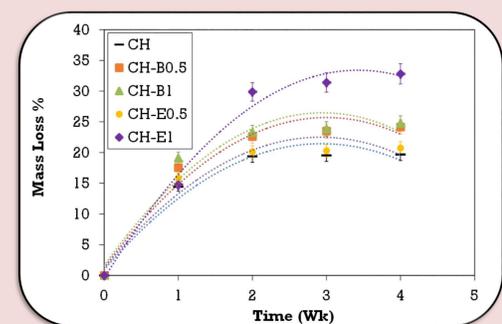


Fig.6. Mineral based membrane samples immersion in saline solution for a period of 4 weeks to test the rate of degradation.

- It is essential to optimize the rate of resorption of the scaffold with the rate of bone mineralization and tissue remodeling.

- Co-culturing is essential to ensure osteoblast cells (mineralisation) and endothelial cells (Fig.7) (vascularisation) are able to grow on all the membranes fabricated

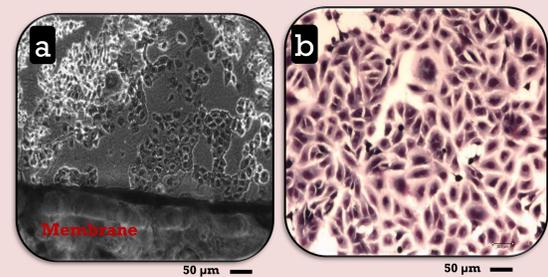


Fig.7. Microscope images of G292-osteoblast growth on membrane samples (a) chitosan film, (b) NaOH treated chitosan film.

- Cytotoxicity testing was successful thus; determining all membranes were not toxic to cells.

## Acknowledgments

This research is supported by The University of Leeds and is funded by the Engineering and Physical Sciences Research Council (EPSRC).

## References

- [1] Das, S., et al., *The induction of angiogenesis by cerium oxide nanoparticles through the modulation of oxygen in intracellular environments*. Biomaterials, 2012. 33(31): p. 7746-7755.
- [2] Boccaccini, A.R., et al., *Polymer/bioactive glass nanocomposites for biomedical applications: A review*. Composites Science and Technology, 2010. 70(13): p. 1764-76.

