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Fabrication of Multi-Layered Bone Scaffolds using Femtosecond Pulsed Lasers

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Abstract: An IR femtosecond pulsed laser was used for micropatterning of biomineral containing chitosan membranes, aiming to enhance bone mineralization and angiogenesis. Pre and post irradiation materials have been characterized with XRD, SEM and spectroscopic techniques.

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1. Introduction

Bone is a complex living tissue with significant metabolic and regenerative activities, which are disrupted when the tissue is damaged. Healing of fractured bone is a complicated process therefore; an ideal bone scaffold should have: i) exceptional osteogenic potential in order to promote new bone formation; ii) load-bearing properties; iii) appropriate microstructure for promoting angiogenesis for circulation of nutrients and iv) antibacterial resistance to avoid infections that lead to failure of the surgery. Research efforts have been focused on development of the “perfect” biomaterial, targeting to achieve all the aforementioned properties with a single material system; e.g. blocks of calcium phosphates, coated Ti-implants [1], polymers [2] etc. The aim of our research is to establish a novel approach, for the development, fabrication and implantation of bone scaffolds, based on the fundamentals of advanced manufacturing using femtosecond lasers.

In the present work we discuss the fabrication of a multi-layered scaffold which has the potential to promote both bone mineralization and the formation of intrinsic vasculature. To achieve this we developed two types of chitosan mineral loaded samples. The **Type-1** sample, aims to enhance osteogenic potential, it is loaded with calcium phosphate minerals (e.g. β -calcium pyrophosphate) and is fabricated through freeze drying in order to achieve a highly porous honeycomb structure. The **Type-2** samples are based on membrane geometries, fabricated through casting of chitosan solutions loaded with CeO_2 and Ce_2O_3 nanoparticles which are known for their excellent antibacterial and angiogenetic properties already observed in systems for *in vitro* and also *in vivo* models [3]. A Coherent Libra-S-1K (100fs) femtosecond laser (1 kHz repetition rate), at a wavelength of 800 nm, was utilised to fabricate microchannel networks and patterns on the Type-2 membrane surfaces. Micropatterning and microchannels are expected to promote guided cellular growth leading eventually to the formation of vasculature. In

addition, the increased capillary forces in the channels will allow for better circulation of the nutritional components which are necessary for the formation of new blood vessels. The scaffold can be constructed through a **numbering up** approach where several layers of Type-1 and Type-2 membranes are piled up together in order to mimic the natural structure of cancellous bone.

2. Materials & Characterizations

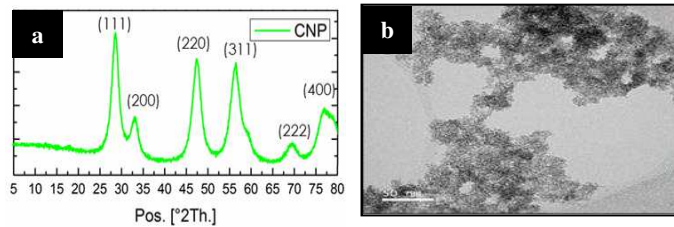


Fig. 1. (a) XRD analysis of cerium oxide nanoparticles; (b) TEM image of cerium oxide nanoparticles with particle size between 10-20nm

Cerium oxide nanoparticles were formed using a hydroxide mediated approach. A 0.3 M aqueous solution of sodium hydroxide was added dropwise to 1 M aqueous cerium nitrate hexahydrate solution at 25°C. The mixture was left under continuous stirring for 24 hrs. The nanoparticles were collected by filtration and washed several times with distilled water. They were subsequently frozen at -80°C for 24 hrs and then placed into a freeze drier for 24 hrs. X-Ray powder diffraction and TEM (fig.1) were used to verify the formation of the cerium oxide nanoparticles.

For the synthesis of both Type-1 and Type-2 materials, 6g of chitosan flakes was dissolved in a 2% v/v aqueous acetic acid solution under continuous mixing for 24 hrs. Type-1 samples were synthesised by adding to the chitosan solution different quantities of calcium phosphate minerals (i.e. 20, 30 and 40% w/w). After freeze drying, porous membranes have been obtained with pore sizes in the range of 42 μm and 94 μm (fig.2a) which are appropriate for bone regeneration (the apparent porosities are shown in fig.2b). For Type-2 membranes, CeO₂ nanoparticles have been added to chitosan solution and the mixture left to dry until the formation of a compact membrane.

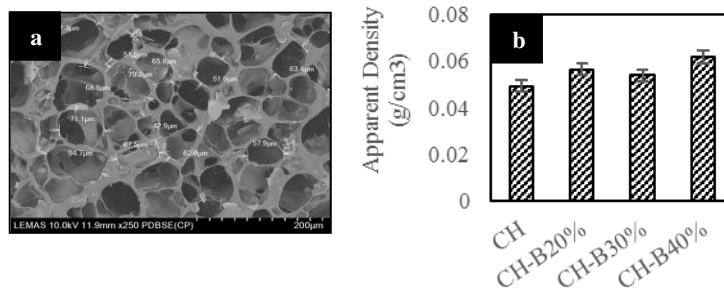


Fig. 2. (a) Hitachi SU8230 SEM image of **Type-1** freeze dried sample containing 30 wt% calcium phosphate minerals; (b) calculated apparent density of freeze dried calcium phosphate-chitosan samples.

3. Femtosecond Pulse Laser & Cell Study

Experiments were carried out to identify the ablation threshold of Type-2 materials; this was determined to be 5.4 J/cm². For the fabrication of channels and micropatterns the initial average power of the laser was set at 40 mW and

then consecutive channels were formed by gradually increasing the power in steps of 20 mW per line (maximum power used was 220 mW). As it was expected, the channel width was increased by increasing the power of the laser in a range between 30 μm and 140 μm . Fabrication of microchannels on the membrane surfaces were successful, as can be seen in fig.3 (a).

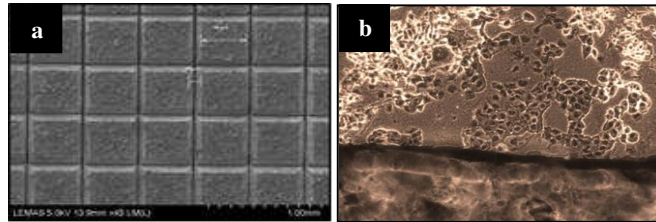


Fig. 3. (a) Hitachi SU8230 SEM image of regular and periodic microchannels fabricated with the aid of a femtosecond laser operating at 800 nm; (b) Optical microscope image of initial osteoblast contact test on chitosan-calcium phosphate membrane

Initial tests verified that both Type-1 and Type-2 samples do not present any toxicological risks. Osteoblast cells (cell line G292) were grown on and close to our samples as shown in fig.3b. Further experiments are still under process in order to confirm that cell growth and proliferation occurs within the micropatterns/channels.

4. References

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- [3] Boccaccini, A.R., et al., *Polymer/bioactive glass nanocomposites for biomedical applications: A review*. *Composites Science and Technology*, 2010. **70**(13): p. 1764-76.