



This is a repository copy of *Salicylic acid, senescence, and heterosis*.

White Rose Research Online URL for this paper:  
<http://eprints.whiterose.ac.uk/145790/>

Version: Accepted Version

---

**Article:**

Smith, L.M. [orcid.org/0000-0003-2364-8187](https://orcid.org/0000-0003-2364-8187) (2019) Salicylic acid, senescence, and heterosis. *Plant Physiology*, 180 (1). pp. 3-4. ISSN 0032-0889

<https://doi.org/10.1104/pp.19.00260>

---

Copyright American Society of Plant Biologists. This is an author-produced version of a paper subsequently published in *Plant Physiology* [[www.plantphysiol.org](http://www.plantphysiol.org)]. Uploaded in accordance with the publisher's self-archiving policy.

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>

## Salicylic acid, senescence, and heterosis

For a number of crops agricultural yields have been boosted by the phenomenon of heterosis or hybrid vigour, where a hybrid outperforms either parent. Understanding the various mechanisms by which heterosis arises may facilitate the breeding of hybrids with increased vigour for further crop varieties or species. Vigour can be determined by measurement of a number of traits from seed yield and plant biomass through to biotic and abiotic stress resistance. Trade-offs may exist between some of these traits such that breeding for one may negatively affect another desirable phenotype (e.g. defence vs growth; [1]). A number of plant hormones act in regulating plant growth and defence with both synergistic and antagonistic effects. Maize (*Zea mays*) and rice (*Oryza sativa*) are two key crops for which heterosis has increased yields. In rice, altered regulation of gibberellic acid and abscisic acid biosynthesis genes can lead to hybrid vigour [4], while factors that contribute to heterosis in maize including the masking of recessive deleterious mutations [2] and changes in the epigenetic landscape associated with subsequent gene expression [3].

As for many biological processes, *Arabidopsis thaliana* has been used as a model in the study of hybrid vigour, with a number of pathways and genes that contribute to heterosis identified over the past decade. For *Ler* x C24 hybrids of *Arabidopsis*, auxin signalling and transport pathways, as well as epigenetic regulation of genes have been identified as mechanisms of heterosis [6, 7]. These studies confirm the potential for hormone signalling and epigenetic regulation of gene expression to contribute to hybrid vigour. For C24 x Col-0 hybrids, heterosis in rosette growth is linked to up-regulation of photosynthesis genes and down-regulation of stress response genes that are co-regulated by the circadian clock [8]. Although far from a comprehensive, these previous studies demonstrate the complexity of heterosis, which has different causes between hybrids and species and multiple factors contributing to heterosis within a single hybrid. In contrast to the varied mechanisms of heterosis, the opposite phenomenon of hybrid incompatibility or hybrid weakness is frequently based on up-regulation of defence response genes [5].

In this issue of *Plant Physiology*, Ian Greaves' group at CSIRO Agriculture and Food in Canberra further explored the basis of heterosis in C24 x *Ler* hybrids of *Arabidopsis* [9]. This study builds on their previous body of work showing that hormone pathway and biotic response genes are differently expressed in hybrids of C24 x *Ler*, C24 x Col-0 and Col-0 x *Ler* [10]. The group has previously shown that a reduction in the defence hormone salicylic acid leads to increased vigour in C24 plants, while decreased salicylic acid and auxin responses are present in C24 x *Ler* and C24 x Col-0 hybrids [10]. Based on these results, the authors hypothesized that hybrid vigour can in some instances be the reverse of hybrid incompatibility, i.e. that down-regulation of defence response genes in a hybrid may lead to heterosis.

It is important to note that C24 is physiologically an outlier compared to other *Arabidopsis* accessions [11]. With reduced stomatal conductance, possibly linked to the higher salicylic acid levels, C24 is drought, heat and ozone tolerant, but more sensitive to some other forms of abiotic stress [12-15]. C24 has been known for almost a decade to have constitutively high salicylic acid levels with small rosettes but normal levels of seed yield when not

stressed [12]. Thus, the use of rosette biomass as an indicator of overall fitness does not apply to C24.

In the current paper, the authors have explored further the role of salicylic acid in C24 and C24 x *Ler* hybrids. The authors confirmed their previous finding that decreasing salicylic acid levels in C24, through the introduction of a bacterial *NahG* gene that converts the hormone into inactive catechol, leads to an increase in plant size after 21 days of growth. Whether a further reduction in salicylic acid levels in the C24 x *Ler* hybrid would also lead to an increase in growth has not been tested, although the fresh weight of mature C24 *NahG* rosettes was equivalent to that of the C24 x *Ler* F1 plants. However, the hybrids were larger than either parent or the C24 *NahG* plants early in growth, indicating that there is an additional heterosis mechanism other than salicylic acid active during seedling establishment.

Through a detailed transcriptome comparison of C24 *NahG*, *Ler NahG*, and F1 C24 x *Ler* plants to the mid-parental value, Gonzalez-Bayon and colleagues found that the majority of differentially expressed genes were down-regulated, rather than up-regulated, and therefore focused on this subset of 771 repressed genes. Unsurprisingly, genes within the salicylic acid biosynthesis pathway were down-regulated in the C24 *NahG* and hybrid lines, consistent with salicylic acid levels being lower than the C24 value in these lines. In the *Ler NahG* line, where salicylic acid levels were not significantly reduced beyond the already low levels of the hormone in the *Ler* wild-type, various salicylic acid-responsive defence genes were also down-regulated, but to a lesser extent. In the C24 *NahG* and hybrid lines, lower gene expression was largely due to a reduction in expression of C24-derived genes rather than the *Ler* genes.

Down-regulation of defence responses may facilitate increased plant growth as a trade-off between growth and defence generally is accepted in plant biology [16] (although this connection is not absolute, see [17]). One important salicylic acid-inducible regulatory gene that has been linked to the balance between growth to defence is *TL1 BINDING TRANSCRIPTION FACTOR 1 (TBF1)*; [18]), a gene whose transcript levels were significantly reduced in the F1 hybrids and C24 *NahG* plants. Likewise, genes connected to salicylic acid-induced senescence were down-regulated in these lines, which Gonzalez-Bayon and colleagues linked to a brief delay in senescence providing leaves with a longer period of maximum photosynthetic potential.

Previously, the Greaves group has reported that increased plant size in C24 x *Ler* F1 hybrids and C24 *NahG* plants is partially due to increase in cell size [10]. In the current paper, they expand on this interesting result, showing that a handful of genes associated with increased cell size (encoding expansins and xyloglucan modifying proteins) were up-regulated in these lines, while extensins that limit cell size were down-regulated. Whether these genes act downstream of salicylic acid remains to be explored.

In summary, down-regulation of salicylic acid levels can increase growth in an accession with unusually high salicylic acid levels. In comparison, a hybrid from this accession that exhibits heterosis also showed down-regulation of salicylic acid-regulated defence genes. Both genotypes exhibit delayed senescence, allowing leaves to contribute more to growth through a prolonged period of maximum photosynthetic capacity. However, as evidenced

by i) the small size of the Ler parental line despite its low levels of salicylic acid, and ii) previous literature, hormone levels are only one part of the jigsaw puzzle in plant growth and heterosis.

The wider impact of the research presented by Gonzalez-Bayon and colleagues is that selection for down-regulation of the salicylic acid pathway in inbred accessions may prove to be a viable strategy to increase plant size and hence crop yields. Whether this approach is also useful for hybrids will require further investigation and may only be effective in cases where one parent has uncommonly high salicylic acid levels to begin with.

#### Cited papers:

1. Albrecht, T. and C.T. Argueso, *Should I fight or should I grow now? The role of cytokinins in plant growth and immunity and in the growth–defence trade-off*. Annals of Botany, 2017. **119**(5): p. 725.
2. Yang, J., et al., *Incomplete dominance of deleterious alleles contributes substantially to trait variation and heterosis in maize*. PLOS Genetics, 2017. **13**(9): p. e1007019.
3. Seifert, F., et al., *Parental Expression Variation of Small RNAs Is Negatively Correlated with Grain Yield Heterosis in a Maize Breeding Population*. Frontiers in Plant Science, 2018. **9**(13).
4. Chen, L., et al., *Genetic analysis for the grain number heterosis of a super-hybrid rice WFYT025 combination using RNA-Seq*. Rice, 2018. **11**(1): p. 37.
5. Chae, E., et al., *Species-wide genetic incompatibility analysis identifies immune genes as hot spots of deleterious epistasis*. Cell, 2014. **159**(6): p. 1341-51.
6. Groszmann, M., et al., *Changes in 24-nt siRNA levels in Arabidopsis hybrids suggest an epigenetic contribution to hybrid vigor*. Proc Natl Acad Sci USA, 2011. **108**(6): p. 2617-22.
7. Shen, H., et al., *Genome-Wide Analysis of DNA Methylation and Gene Expression Changes in Two Arabidopsis Ecotypes and Their Reciprocal Hybrids*. The Plant Cell, 2012. **24**(3): p. 875.
8. Miller, M., et al., *Natural variation in timing of stress-responsive gene expression predicts heterosis in intraspecific hybrids of Arabidopsis*. Nature communications, 2015. **6**: p. 7453.
9. Gonzalez-Bayon, R., et al., *Senescence and defence pathways contribute to heterosis*. Plant Physiology, 2019.
10. Groszmann, M., et al., *Hormone-regulated defense and stress response networks contribute to heterosis in Arabidopsis F1 hybrids*. Proc Natl Acad Sci U S A, 2015. **112**(46): p. E6397-406.
11. Ferguson, J.N., et al., *Natural variation of life-history traits, water use, and drought responses in Arabidopsis*. Plant Direct, 2018. **2**(1): p. e00035.
12. Bechtold, U., et al., *Constitutive salicylic acid defences do not compromise seed yield, drought tolerance and water productivity in the Arabidopsis accession C24*. Plant, cell & environment, 2010. **33**(11): p. 1959-1973.
13. Xu, E., et al., *Quantitative trait loci mapping and transcriptome analysis reveal candidate genes regulating the response to ozone in Arabidopsis thaliana*. Plant, cell & environment, 2015. **38**(7): p. 1418-1433.

14. Brosché, M., et al., *Natural variation in ozone sensitivity among Arabidopsis thaliana accessions and its relation to stomatal conductance*. Plant, cell & environment, 2010. **33**(6): p. 914-925.
15. Bechtold, U., J.N. Ferguson, and P.M. Mullineaux, *To defend or to grow: lessons from Arabidopsis C24*. Journal of Experimental Botany, 2018. **69**(11): p. 2809-2821.
16. Karasov, T.L., et al., *Mechanisms to Mitigate the Trade-Off between Growth and Defense*. The Plant Cell, 2017. **29**(4): p. 666.
17. Campos, M.L., et al., *Rewiring of jasmonate and phytochrome B signalling uncouples plant growth-defense tradeoffs*. Nature Communications, 2016. **7**: p. 12570.
18. Pajerowska-Mukhtar, K.M., et al., *The HSF-like transcription factor TBF1 is a major molecular switch for plant growth-to-defense transition*. Current Biology, 2012. **22**(2): p. 103-112.