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## **Genotypic differences in shoot silicon content and the impact on grain arsenic accumulation in rice**

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2       1 Title Page:  
3       2 Genotypic differences in shoot silicon content and the impact on grain arsenic accumulation  
4       3 in rice  
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7       5 Partha Talukdar<sup>1</sup>, Sue E. Hartley<sup>2</sup>, Anthony J Travis<sup>1</sup>, Adam H. Price<sup>1</sup>, Gareth J. Norton<sup>1\*</sup>  
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2       35 Genotypic differences in shoot silicon content and the impact on grain arsenic accumulation  
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7       38 Partha Talukdar<sup>1</sup>, Sue E. Hartley<sup>2</sup>, Anthony J Travis<sup>1</sup>, Adam H. Price<sup>1</sup>, Gareth J. Norton<sup>1\*</sup>  
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33      46 **Abstract:**  
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Silicon in rice has been demonstrated to be involved in resistance to lodging, tolerance to both drought and salinity, and also enhances resistance to pests and diseases. The aim of this study was to determine the range of silicon content in a set of rice (*Oryza sativa* L.) accessions, and to determine if the natural variation of shoot silicon is linked to the previously identified silicon transporters (*Lsi* genes). Silicon content was determined in 50 field-grown accessions, representing all sub-populations of rice, with all accessions being genotyped with 700K SNPs. SNPs within 10 kb of the *Lsi* genes were examined to determine if any were significantly linked with the phenotypic variation. An XRF method of silicon determination compared favourably with digestion and colorimetric analysis. There were significant genotypic differences in shoot silicon ranging from 16.5 – 42.4 mg g<sup>-1</sup> of plant dry weight, there was no significant difference between the rice sub-populations. Plants with different alleles for SNPs representing *Lsi2* and *Lsi3* were significantly different for shoot silicon content. Shoot silicon correlated negatively with grain arsenic in the *tropical* and *temperate japonica* sub-population, suggesting that accessions with high shoot silicon have

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2       61 reduced grain arsenic. This study indicates that alleles for *Lsi* genes are excellent candidate  
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4       62 genes for further study to explain the natural variation of shoot silicon in rice.  
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11      64 **Key words:** arsenic, natural variation, rice, silicon, XRF  
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## 1 2      66    1 Introduction

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5      67    Global rice (*Oryza sativa* L.) production needs to increase continuously to ensure the world's  
6      68    food security (*Hibberd* et al., 2008). As a beneficial element, silicon alleviates biotic and  
7      69    abiotic stresses in rice which helps to maintain yield (*Ma and Takahashi*, 2002; *Detmann* et  
8      70    al., 2012; *Meharg and Meharg*, 2015). Silicon is mainly available as monosilicic acid ranging  
9      71    from 0.1 to 0.6 mM in the soil solution (*Epstein*, 1994; *Ma and Takahashi*, 2002). Previous  
10     72    studies have demonstrated that monosilicic acid is taken up by rice roots as an undissociated  
11     73    molecule and translocated into the shoots through the transpiration stream (*Takahashi and Hino*,  
12     74    1978; *Mitani-Ueno* et al., 2005). It then polymerises on the surface of cells in the shoot  
13     75    in the form of a silica-cellulose double layer and silica-cuticle double layer. This silica-base  
14     76    layer improves resistance to lodging, salinity tolerance, drought tolerance, and enhances  
15     77    resistance to pests and diseases (*Takahashi and Hino*, 1978; *Mitani-Ueno* et al., 2005, *Chen*  
16     78    et al., 2011; *Han* et al., 2015).

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19     79    Genetically rice can be classified into two major sub-species, *Japonica* and *Indica* (*Chang*,  
20     80    2003) and these have been further classified into 5 sub-populations; *indica*, *aus*, (both *Indica*  
21     81    sub-species) *tropical japonica*, *temperate japonica*, and *aromatic* (all three *Japonica* sub-  
22     82    species) (*Garris* et al., 2005; *Zhao* et al., 2011). Several previous studies indicate that there  
23     83    are differences in shoot silicon content between the *Indica* and *Japonica* sub-species of rice.  
24     84    *Deren* et al., (1992) showed that *Japonica* sub-species usually have a higher silicon content  
25     85    than *Indica* rice varieties, based on screening 10 accessions in the greenhouse and 18 under  
26     86    field conditions. A study conducted by *Winslow* (1992) revealed that African upland  
27     87    *Japonica* rice accessions had 50 to 100% higher silicon content in mature flag leaves than  
28     88    Asian upland *Indica* accessions. In addition to the differences at the subspecies level several  
29     89    studies have looked at genotypic differences in silicon content, showing ranges of 41 to 60  
30     90    mg g<sup>-1</sup> (*Deren*, 2001) and 28 to 61 mg g<sup>-1</sup> (*Norton* et al., 2010a). *Ma* et al., (2007a) also

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2       91 observed that silicon uptake by the root and the concentration silicon present in the shoot are  
3       92 both higher in *Japonica* than *Indica* rice accessions, which they attributed to differences in  
4       93 the expression of silicon transporter genes.  
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8       94 Two types of silicon transporters have been identified in rice to date. A gene  
9       95 (LOC\_Os02g51110) identified for silicic acid influx in rice is classified as an aquaporin  
10      96 (Low silicon 1 or *Lsi1*) which is a member of the nodulin 26-like intrinsic protein (OsNIP2;  
11      97 1) group of aquaporins (Ma et al., 2006; Ma et al., 2008). A homologue of *Lsi1*, known as  
12      98 *Lsi6* (LOC\_Os06g12310; OsNIP2; 2), responsible for shoot and husk silicon distribution in  
13      99 rice is also classified as an aquaporin (Yamaji et al., 2008). The efflux of silicic acid through  
14      100 the plasma membrane protein known as low silicon 2 (*Lsi2*; LOC\_Os03g01700) is an energy  
15      101 dependent process in rice (Ma et al., 2007b). A homologue of *Lsi2*, known as *Lsi3*  
16      102 (LOC\_Os10g39980), is also an energy dependent active transporter involved in regulating  
17      103 shoot silicon accumulation in rice (Yamaji et al., 2015).  
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21       104 It has been shown that arsenic, classified as a class one carcinogen, can be transported  
22       105 through silicon transporters in rice (Ma et al., 2008; Zhao et al., 2010; Mitani-Ueno et al.,  
23       106 2011). There are two different forms of arsenic present in rice: organic arsenic and inorganic  
24       107 arsenic (Williams et al., 2005). Organic arsenic is found in rice in two main types of  
25       108 molecular species dimethylarsinic acid (DMA), and monomethylarsonic acid (MMA) as well  
26       109 as tetramethylarsonium (Williams et al., 2005; Hansen et al., 2011). Inorganic arsenic is  
27       110 found in rice as two molecular species; arsenate and arsenite (Abedin et al., 2002; Williams et  
28       111 al., 2005). Arsenate is an analogue of phosphate and is taken up via phosphate transporters  
29       112 while arsenite is taken up by silicic acid transporters in rice (Abedin et al., 2002; Ma et al.,  
30       113 2008). It has been shown that the silicon transporters *Lsi1*, *Lsi2* and *Lsi6* are also arsenic  
31       114 transporters, using a combination of mutants and transgenic lines (Ma et al., 2008; Zhao et  
32       115 al., 2010; Mitani-Ueno et al., 2011). Several studies indicate that anaerobic rice cultivation  
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3     116 leads to increased mobilisation of soil arsenic in the form of arsenite, which causes  
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5     117 anaerobically-grown rice to accumulate more arsenic through silicon transporters (*Ma et al.*,  
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7     118 2008; *Xu et al.*, 2008; *Carey et al.*, 2010). Silicon fertilisation has also been shown to  
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9     119 decrease shoot and grain arsenic indicating that silicon could play an important role in  
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11     120 decreasing total arsenic uptake in rice (*Li et al.*, 2009; *Seyfferth and Feldorf*, 2012).

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14     121 This study was designed to address four questions all related to the process of silicon and  
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16     122 arsenic accumulation in rice: How does the cultivation method affect silicon distribution in  
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18     123 different organs of rice plants? Are there significant genotypic differences in shoot silicon  
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20     124 concentration across a diverse panel of rice related to the 5 different sub-populations of rice?  
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22     125 Is there a relationship between natural variation in shoot silicon and arsenic content in rice?  
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24     126 Can natural variation in shoot silicon be linked to known silicon transporters in rice? The  
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26     127 results provide a deeper understanding of the natural variation in silicon content across rice  
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28     128 accessions and its relationship to arsenic accumulation in rice grains.  
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2      129 **2 Materials and Methods**  
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6      130 **2.1 Silicon content in different organs of rice (*Oryza sativa L.*) grown in flooded and**  
7      131 **non-flooded conditions**  
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10     132 An experiment was conducted in a greenhouse at the University of Aberdeen, UK under both  
11     133 flooded and non-flooded conditions with four replicates for each treatment. One litre plastic  
12     134 pots were filled with soil (~530 g soil described in *Norton et al., 2013*). For the flooded  
13     135 condition, a plastic liner was used to line the pots and hold the water within the pot whereas  
14     136 the non-flooded pots were kept without a liner to allow drainage of water through the pot.  
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17     137 Five Italica Carolina (*temperate japonica*) seeds were sown in each pot, then thinned to one  
18     138 plant in each pot after two weeks. To maintain the flooded condition, tap water from the  
19     139 greenhouse was used to flood the pots to 2 cm above the soil surface when plants were 3  
20     140 weeks old. Every two weeks during the first four weeks of growth 100 mL of half strength  
21     141 Yoshida's nutrient solution was added (*Yoshida et al., 1976*). The dose of Yoshida's nutrient  
22     142 solution was increased up to 100 mL of full strength solution every week after four weeks  
23     143 and continued until the filled grains had matured when samples were harvested.  
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26     144 At harvest, samples were collected from different parts of the mature plants: root, 3<sup>rd</sup> sheath,  
27     145 3<sup>rd</sup> node, 3<sup>rd</sup> leaf, 2<sup>nd</sup> sheath, 2<sup>nd</sup> node, 2<sup>nd</sup> leaf, flag sheath, 1<sup>st</sup> node, flag leaf, husk and  
28     146 unpolished grains. The sheath, node and leaves were taken from the main tiller, with the most  
29     147 recent leaf prior to the flag leaf designated 2<sup>nd</sup> leaf. Root samples were washed thoroughly  
30     148 with tap water followed by deionised water and confirmed to be free of soil particles by  
31     149 examining the roots under a microscope (Leica MZ8, 10445932, 16×/14B, PLAN 1.0X).  
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33     150 Samples were dried at 80°C for 5 days until a constant weight was achieved. All samples  
34     151 were mixed and subsampled prior to being ball-milled. The silicon content was determined  
35     152 by Flow Injection Analyser (FIA) after alkali digestion.  
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2      153 **2.2 Genotypic differences in shoot silicon content of rice**

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4      154 Seeds were obtained from the Rice Diversity Panel 1 (RDP1) (<http://ricediversity.org/>) (Ali et  
5      155 al., 2011; Eizenga et al., 2014). The classification of Zhao et al., (2011) was used for the sub-  
6      156 population classification of rice accessions. In 2009 a total of 312 accessions were cultivated  
7      157 at the experimental site in Bangladesh. Seedlings were transplanted by hand in a single 2m  
8      158 row of 10 hills, each hill (one seedling) 20 cm apart and each row 20 cm apart in a  
9      159 randomised complete block design with four replicates of each accession. The experimental  
10     160 site was kept flooded until the grains were filled. Plant material from the central six plants  
11     161 was harvested and used for chemical analysis. Detailed information about the experimental  
12     162 site and experimental methods are described in Norton et al., (2012). For shoot silicon  
13     163 analysis, fifty accessions (10 accessions from each rice sub-population) were randomly  
14     164 selected based on the initial sub-population assignment using single sequence repeat (SSR)  
15     165 markers (Ali et al., 2011) (Supplementary Table 1). Subsequently, after selection and silicon  
16     166 analysis, these accessions have been assigned to sub-populations based on the 700K SNP data  
17     167 (McCouch et al., 2016), these sub-population assignments are used for classification of the  
18     168 accessions in this study.

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20      169 **2.3 Analysis of rice shoot silicon by FIA**

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22      170 Plant material and certified reference material (CRM) were prepared for silicon analysis as  
23      171 described by Carneiro et al., (2007). A total of 1.5 g shoot material from each sample was  
24      172 sub-sampled at random and powdered using a ball mill (Retsch, MM200, Germany). From  
25      173 the powdered plant material, a sub-sample of 20 mg was weighed into a 50 mL  
26      174 polyethylene centrifuge tube (CORNING®, NY). To digest the sample, 0.6 mL of hydrogen  
27      175 peroxide (H<sub>2</sub>O<sub>2</sub>, > 30% W/V, Fisher Scientific) and 1.5 mL of sodium hydroxide (NaOH,  
28      176 solutions 50%, Fluka) were added and the samples were then vortexed (mixed using a vortex  
29      177 mixer). The samples were heated for 1 hour at 90°C in a water bath, then vortexed again and

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2      178 left overnight. The tubes were vortexed again after overnight extraction, then heated at 123°C  
3      179 under a pressure of 0.15 MPa for 1 h. Samples were kept at room temperature for 2 h then  
4      180 vortexed, followed by addition of 18.5 mL of ddH<sub>2</sub>O. Prior to analysis, samples were diluted  
5      181 1:5 with Milli-Q water. Silicon content was measured using an FIA spectrophotometer  
6      182 (Tecator FI Astar 5010) a wavelength of 410 nm (*Carneiro et al., 2007; Norton et al., 2010a;*  
7      183 *Norton et al., 2010b*).

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9      184 **2.4 Analysis of rice shoot silicon by P-XRF**

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11      185 A total of 1.5 g of dried shoot material for each rice accession was sub-sampled at random  
12      186 and powdered using a ball mill (Retsch, MM200, Germany). To perform the analysis of shoot  
13      187 silicon by P-XRF, 19 accessions were selected at random from the 50 accessions for which  
14      188 shoot silicon had been determined by FIA. For P-XRF analysis, 0.7 g of homogeneous  
15      189 powder sample was compressed into 13 mm diameter pellets using a manual hydraulic press  
16      190 with a 13mm die at a pressure of 10 tons (Specac, Orpington, United Kingdom). Shoot silicon  
17      191 content (% of silicon dry shoot weight) was measured using a commercial P-XRF instrument  
18      192 (Niton XL3t900 GOLDD analyzer: Thermo Scientific Winchester, UK), calibrated using Si-  
19      193 spiked synthetic methyl cellulose and validated using Certified Reference Materials of NCS  
20      194 DC73349 ‘Bush branches and leaves’ obtained from the China National Analysis Center for  
21      195 Iron and Steel, as described in *Reidinger et al., (2012)*. The mean value of samples for each  
22      196 accession was used for correlation analysis between P-XRF and FIA measurements.

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24      197 **2.5 Relationship between silicon and arsenic content in rice**

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26      198 The plant material used in this study was previously examined for grain arsenic content  
27      199 (*Norton et al., 2012*) which provided an opportunity to examine the relationship between  
28      200 shoot silicon and grain arsenic in rice. The relationship between shoot silicon (log

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2       201 transformed) and grain arsenic (log transformed) was investigated for the 50 rice accessions  
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4       202 based on accession means.  
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7       203 **2.6 Single Marker Analysis**  
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10      204 The accessions used in this study have been genotyped using a high-density SNP chip  
11      205 (*McCouch et al., 2016*). SNPs for the accessions were extracted using PLINK (*Purcell et al.,*  
12      206 2007). SNPs were extracted from 10 kb upstream of the start codon to 10 kb downstream of  
13      207 the stop codon of the *Lsi1*, *Lsi2*, *Lsi3*, and *Lsi6* loci. SNPs were excluded from the analysis if  
14      208 they were invariant or if minor alleles were present in less than 5% of the accessions. The  
15      209 RDP1 population has a high degree of stratification by rice sub-population (*Zhao et al., 2011*;  
16      210 *McCouch et al., 2016*). To overcome this stratification, sub-population assignment was used  
17      211 (based on the 700 K SNP data; *McCouch et al., 2016*) as a factor in a two-way ANOVA, with  
18      212 SNP base call as the other factor. The two-way ANOVA was used to determine if the  
19      213 phenotype for the accession was significantly different for each SNP tested.  
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22      214 **2.7 Sequence alignments**  
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25      215 Based on the result achieved from the single-marker analysis the sequence diversity of *Lsi2*  
26      216 and *Lsi3* were investigated for 5 cultivars using BAM files produced after aligning sequence  
27      217 reads against Nipponbare version 7 reference genome. The genome sequences of the cultivars  
28      218 used in this study have been previously published (*Kawahara et al., 2013; Cardoso et al.,*  
29      219 2014; *Schatz et al., 2014*). The accessions were from the following sub-populations; 2 *indica*  
30      220 accessions (IR64 and Bala), 1 *aus* accession (DJ123) and 2 *tropical japonica* accessions  
31      221 (Azucena and Nipponbare). The genomic DNA sequence was visualised using the IGV  
32      222 (<https://www.broadinstitute.org/igv/>) to identify the difference of genomic DNA sequence  
33      223 within *Lsi2* and *Lsi3* in 5 cultivars (*Thorvaldsdóttir et al., 2013; Robinson et al., 2011*). Using  
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2       224 Clustal Omega the DNA sequences of 5 cultivars were aligned for *Lsi2* and *Lsi3* separately  
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4       225 and showed in supplementary figure 3 and 4 respectively (*Sievers et al.*, 2011).  
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7       226 **2.8 Statistical analysis**  
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10     227 Statistical significance was set at  $P < 0.05$  for all analyses, which were performed using  
11     228 Minitab 16. The normality of distribution and homogeneity of variance of the data were  
12     229 tested prior to one or two-way analysis of variance (ANOVA), as appropriate. Pearson  
13     230 correlation analysis was used to investigate the relationship between measurements of shoot  
14     231 silicon and grain arsenic.  
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2      232 **3 Results**  
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6      233 **3.1 Shoot silicon content in different organs of rice plants**  
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9      234 Flooding increased plant silicon content in the flag sheath, 1<sup>st</sup> node, flag leaf and husk  
10     235 compared to plants grown under non-flooded conditions (Fig. 1). The content of silicon in  
11     236 grain and root tissues were significantly lower than any in other organs of plants grown under  
12     237 either condition. There was a significant difference ( $P < 0.001$ ,  $F = 27.40$ ,  $R^2 = 78.20\%$ ) of  
13     238 silicon content between different organs of the plant in non-flooded conditions: The highest  
14     239 mean content was in the husks ( $46.8 \text{ mg g}^{-1}$ ), while the lowest was in the grain ( $3.5 \text{ mg g}^{-1}$ ).  
15     240 For plants grown under flooded conditions: The highest silicon content was observed in the  
16     241 flag leaf ( $67.3 \text{ mg g}^{-1}$ ) and the lowest was in the grains ( $4.4 \text{ mg g}^{-1}$ ).  
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19     242 **3.2 Genotypic difference in shoot silicon content of rice**  
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22     243 Fifty diverse rice accessions were examined by FIA to determine the difference in shoot  
23     244 silicon content of rice. There was a significant genotypic difference in shoot silicon content  
24     245 among the 50 accessions, where genotype explained 55% of the variation ( $P < 0.001$ ;  $F =$   
25     246  $5.80$ ;  $R^2 = 55.30\%$ ;  $df = 49$ ). The mean shoot silicon content of the 50 accessions was  $28.1$   
26     247  $\text{mg g}^{-1}$ , and the lowest mean shoot silicon was observed in Dhala Shita ( $16.5 \text{ mg g}^{-1}$ ) The  
27     248 highest mean shoot silicon was observed in Bala ( $42.4 \text{ mg g}^{-1}$ ) (Fig. 2). There was no  
28     249 significant difference for shoot silicon content of the 5-major rice sub-populations (Fig. 3).  
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31     250 Nineteen rice accessions were selected at random from the 50 accessions analysed by FIA,  
32     251 for measurement of shoot silicon content by P-XRF. The silicon content of four individual  
33     252 field grown replicates of each accession were measured separately by P-XRF and FIA and the  
34     253 mean value of each accession was used for correlation analysis. Using both methods,  
35     254 genotypic differences were observed between the accessions ( $P < 0.001$ ;  $F = 9.90$ ;  $df = 18$  for  
36     255 P-XRF;  $P < 0.001$ ;  $F = 7.30$ ;  $df = 18$  for FIA). Correlation analysis indicated that there was a  
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3 256 significant and large positive correlation between the two methods ( $r = 0.95$ ;  $P < 0.001$ ; df =  
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5 257 18) (Fig 4).

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7 258 **3.3 Correlation between shoot silicon and grain arsenic in rice**

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9 259 No significant correlation was observed between mean shoot silicon and mean shoot arsenic  
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11 260 for all of the 50 accessions (supplementary Figure 1), or for within each of the 5 sub-  
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13 populations. There was a weak negative correlation ( $r = -0.31$ ;  $P = 0.028$ ; df = 49)  
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15 261 (supplementary Figure 2) between shoot silicon and grain arsenic content for all 50  
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17 262 accessions. When correlation analysis was conducted separately for shoot silicon and grain  
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19 263 arsenic on each of the sub-populations, significant negative correlations were found for the  
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21 264 *temperate japonica* ( $r = -0.78$ ;  $P = 0.007$ ; df = 9) and *tropical japonica* ( $r = -0.84$ ;  $P = 0.002$ ;  
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23 265 df = 9) accessions (Fig. 5). No significant correlations were observed for the other 3 major  
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25 266 rice sub-populations (*indica*, *aus* and *aromatic*).  
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29 268 **3.4 Testing accessions with different alleles of SNPs around and within *Lsi* genes for**  
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31 269 **variation in shoot silicon concentration**

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33 270 A total of 10 SNPs from the SNP database are within 10 kb upstream and downstream of the  
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35 271 *Lsi2* gene (selected SNPs for the rice accessions are presented in supplementary Table 2).  
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37 272 Shoot silicon concentration for accessions with the different alleles for two of these SNPs  
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39 273 was significantly different. SNP-3.434426 is located 2551 bp before the start codon, and  
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41 274 revealed a significant difference between the C and T polymorphism ( $P = 0.006$ ), where the  
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43 275 mean silicon content of accessions with the C allele was  $29.3 \text{ mg g}^{-1}$  while the mean silicon  
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45 276 content of the accessions with the T allele was  $23.1 \text{ mg g}^{-1}$ . SNP-3.438416 is located 6541 bp  
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47 277 before the start codon and revealed a significant difference between the A and C  
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49 278 polymorphism ( $P = 0.008$ ), where the mean silicon content of the accessions with the A allele  
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51 279 was  $29.6 \text{ mg g}^{-1}$  while the silicon content of the accessions with the G allele had a mean of  
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2 280 23.1 mg g<sup>-1</sup>. Both SNPs group the accessions in a similar way, the only difference was more  
3 missing SNP information for SNP-3.438416 (Fig. 6).  
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7 282 A total of 20 SNPs from the SNP database are within 10 kb upstream and downstream of the  
8 283 *Lsi3* gene (selected SNPs for the rice accessions are presented in supplementary Table 3).  
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10 284 Shoot silicon concentration for accessions with the different alleles for one of these SNPs was  
11 285 significantly different. SNP- 10.21340470 is located 5299 bp prior to the start codon, and  
12 286 revealed a significant difference between the G and A polymorphism ( $P = 0.016$ ), where the  
13 287 mean silicon content of accessions with the G allele was 28.4 mg g<sup>-1</sup> while the mean silicon  
14 288 content of the accessions with the A allele was 35.6 mg g<sup>-1</sup> (Fig. 6).  
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17 289 There were 20 SNPs and 19 SNPs observed within 10 kb of *Lsi1* and *Lsi6* respectively.  
18 290 However, at each of these SNPs the different alleles were not significantly different for shoot  
19 291 silicon concentration.  
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22 292 To explore further, the sequence alignments of *Lsi2* and *Lsi3* were performed using available  
23 293 high-quality genome sequences. The accessions used were Nipponbare, Azucena, IR64, Bala,  
24 294 and DJ123 which are from the *tropical japonica*, *tropical japonica*, *indica*, *indica* and *aus*  
25 295 rice subgroups respectively. From the sequence analyses of *Lsi2* and *Lsi3* a number of  
26 296 polymorphisms within the genes were identified. For *Lsi2*, there was a synonymous SNP  
27 297 substitution within the first exon, where DJ123 has “C” allele while the other 4 accessions  
28 298 have “T” allele (Supplementary Figure 3). For *Lsi3*, 4 SNPs were detected in exons and 6  
29 299 SNPs in introns (Supplementary Figure 4). There was only one non-synonymous SNP  
30 300 observed in the first exon of *Lsi3* where DJ123 and Bala have “T” allele and other accessions  
31 301 have “A” allele. The available 3000 rice genome sequence data indicates that this  
32 302 polymorphism between “A” and “T” in *Lsi3* is associated with the *aus* sub-population in rice  
33 303 where 15 accessions have “A” allele and 184 accessions have “T” allele (Alexandrov et al.,  
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3 304 2015). This non-synonymous polymorphism between “A” and “T” in *Lsi3* with the “T” allele  
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5 305 is very rarely observed in *indica* and *japonica* subpopulations of rice in 3000 rice genome  
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7 306 sequence data (Alexandrov et al., 2015).

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2     **308 4 Discussion**  
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6     309 In this study, genotypic differences in shoot silicon content were identified from field grown  
7     310 rice cultivars. However, no differences in shoot silicon were observed across the 5 different  
8     311 sub-populations of rice. Additionally, SNPs detected in the accessions were significantly  
9     312 linked to known silicon transporter genes in rice, which indicates that these genes are  
10    313 potentially involved in natural variation of silicon accumulation in rice.  
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17     314 Flooded conditions increased silicon content in the upper part of the plant (flag sheath, 1<sup>st</sup>  
18     315 node, flag leaf and husk) compared to the non-flooded conditions, which suggests that the  
19     316 uptake or translocation of shoot silicon into these plant organs might be controlled by  
20     317 different processes (compared to those determining silicon uptake in other tissues) which  
21     318 differ between aerobic and anaerobic conditions. It has been shown that silicon dissolution  
22     319 and bio-availability plays a significant role in the variation of silicon content in grasses  
23     320 (Quigley et al., 2017). Therefore, the difference in dissolved silicon in flooded and non-  
24     321 flooded conditions might affect the accumulation of silicon in the rice plants used in this  
25     322 study. It was also notable that there was no significant difference in silicon content in  
26     323 different tissues between the internodes (e.g. flag leaf, 2<sup>nd</sup> leaf) in non-flooded conditions but  
27     324 there was a significant difference between the silicon content of internodes under flooded  
28     325 conditions (Fig. 1). Previous studies have shown that transpiration is one of the most  
29     326 important factors responsible for higher *silicification* in plants and that transpirational flow is  
30     327 higher in anaerobic conditions than in aerobic ones (Mitani-Ueno et al., 2005; Kato and  
31     328 Okami, 2011; Kumar et al., 2017; McLarnon et al., 2017). Therefore, one potential  
32     329 explanation for increased silicon accumulation in the upper organs or developing organs of  
33     330 rice plants (e.g. flag sheath, 1<sup>st</sup> node, flag leaf and husk) grown in flooded soils is a higher  
34     331 transpirational flow in these plants. Importantly, the data presented in figure 1 shows that  
35     332 tissue silicon content is reasonably evenly distributed across tissues with only that from  
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2       333 flooded plants in tissue associated with flowering and seed production significantly higher  
3       334 than the rest. Since this reproductive tissue was removed from the field samples used in this  
4       335 study we can be reassured that a mean value obtained from straw will be a good estimate of  
5       336 the tissue concentration of the majority of rice plant.

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7       337 Fifty accessions from 5 different sub-populations (10 accessions from each sub-population)  
8       338 were selected at random to examine the difference of shoot silicon content in rice and this  
9       339 revealed highly significant differences of shoot silicon content. A genotypic difference in  
10      340 shoot silicon content across a wide group of accessions has been observed previously (*Deren*,  
11      341 2001; *Norton* et al., 2010a). The 2.6-fold difference of shoot silicon content in this study is  
12      342 similar to the previous 2.2 fold range detected for genotypic differences of shoot silicon  
13      343 content in rice (*Norton* et al., 2010a). However, the maximum value observed in our study is  
14      344 slightly lower than that detected previously (42.4 mg g<sup>-1</sup> in this study, 60 mg g<sup>-1</sup> (*Deren*,  
15      345 2001), 61 mg g<sup>-1</sup> (*Norton* et al., 2010a)).

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17      346 The plant material used for determination of shoot silicon content in the 50 rice accessions  
18      347 was grown in flooded, irrigated conditions (*Norton* et al., 2012). Previous studies estimated  
19      348 that 27% - 44% of the silicon taken up by rice plants is supplied by irrigation, while the  
20      349 remaining percentage must be supplied by soil constituents (*Desplanques* et al., 2006;  
21      350 *Klotzbücher* et al., 2015). All the accessions tested in this study had a silicon content below  
22      351 50 mg g<sup>-1</sup> which, according to *Dobermann and Fairhurst* (2000), is below the critical level of  
23      352 mineral deficiency for rice production. The low shoot silicon content (16.5 mg g<sup>-1</sup> to 42.4 mg  
24      353 g<sup>-1</sup>) observed in this study may be due to removal of rice straw from the paddy field, which is  
25      354 common practice in Bangladesh, and has been shown to contribute to lower shoot silicon in  
26      355 the subsequent rice crop (*Seyfferth* et al., 2013). Future work should focus on linking the  
27      356 flooded and non-flooded pot based experiment and the removal of straw at the field scale to

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2 357 establish the importance of water management and field management on silicon accumulation  
3 358 in field grown rice.  
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5 359 Several studies have demonstrated that the *Japonica* sub-species of rice have higher shoot  
6 360 silicon than *Indicas* (Winslow, 1992; Winslow et al., 1997; Ma et al., 2007a). These studies  
7 361 may have been limited by the number of accessions that were screened. For example, Ma et  
8 362 al., (2007b) only screened two rice accessions to examine the genotypic difference in silicon  
9 363 uptake of rice. To improve the current understanding of silicon biology in rice, we  
10 364 investigated field grown shoot samples of 50 rice accessions across 5 sub-populations. Within  
11 365 the material tested in this study the data suggests that the natural variation observed in shoot  
12 366 silicon is not governed by genetic differences between rice sub-populations, but rather is  
13 367 largely due to the genetic differences within individual sub-groups.

14 368 Data on more than 50 accessions would have opened the opportunity to conduct genome-  
15 369 wide association (GWA) mapping where 200 accessions is considered a lower limit.  
16 370 However, the FIA colorimetric method for the determination of silicon in rice shoots proved  
17 371 not to be high throughput. However, in addition to the FIA method, a sub-set of samples  
18 372 were also analysed by P-XRF. The two different methods were strongly correlated, but not  
19 373 perfectly, and indicated that values for silicon content in samples measured by FIA were  
20 374 slightly higher than those measured by P-XRF. The observation that both methods provide  
21 375 comparable results highlights the conclusion that P-XRF can be used for silicon analysis to  
22 376 detect and measure genotypic differences across populations, instead of the more laborious  
23 377 and time-consuming alkali digestion method. Furthermore, a second advantage of P-XRF is  
24 378 that it is a non-destructive method. This would make it much more suitable for future GWA  
25 379 mapping studies.

26 380 The plant material used in this study was previously used to examine the variation of shoot  
27 381 and grain arsenic (Norton et al., 2012). The comparison of shoot silicon and grain arsenic in  
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3 382 this study is in agreement with previous studies where, in general, plants that had high shoot  
4 silicon also had lower grain arsenic (*Seyfferth and Ferdorf*, 2012; *Norton et al.*, 2012; *Norton*  
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6 383 et al., 2013). However, this study also adds more insight by taking into consideration the sub-  
7 population structure of rice cultivars. The correlation between shoot silicon and grain arsenic  
8 was sub-population specific. A strong relationship in between shoot silicon and grain arsenic  
9 was observed in *temperate japonica* and a weaker one in *tropical japonica*, but was not  
10 observed in *indica*, *aus* or *aromatic*. This important observation suggests that the genetic  
11 regulation of arsenic content in rice grain is different in *temperate* and *tropical japonicas*  
12 compared to the other rice sub-populations, implying that the silicon-transport-linked  
13 pathway implicated for arsenic accumulation (*Ma et al.*, 2007b; *Norton et al.*, 2012) may be  
14 less relevant in the *other* sub-populations.  
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18 393 The accessions used in the study have been genotyped using a 700K SNP chip (*McCouch et*  
19 *al.*, 2016). Single-marker analysis was used to test the candidacy of the known transporters of  
20 silica in rice. The study indicated that two SNPs within 10 kb of *Lsi2* and one within 10 kb  
21 of *Lsi3* were involved in contributing to the natural variation of shoot silicon accumulation in  
22 rice (Fig. 6). The *Lsi2* gene has been shown to be pivotal for transport of silicon and  
23 inorganic arsenic in studies conducted with mutants and transgenic plants (*Ma et al.*, 2006;  
24 *Ma et al.*, 2007b; *Yamaji et al.*, 2008; *Mitani-Ueno et al.*, 2011; *Yamaji et al.*, 2015). The  
25 identification of differences in shoot silicon and the link with three SNPs close to the genes  
26 further suggest that *Lsi2* and *Lsi3* are excellent candidate genes to explain the natural  
27 variation observed in shoot silicon content of rice. When looking at the sequencing variation  
28 of a number of diverse cultivars (which have been sequenced to a high depth) it is evident  
29 that there is only a small number of polymorphisms within the genes (Supplementary figures  
30 3 and 4). The highly conserved sequence for *Lsi2* may be due to its importance function for  
31 silicon accumulation in rice. However, the accessions screened in this study are likely to have  
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3 407 greater sequence variation than the cultivars for which high quality sequence is available, and  
4 408 therefore there may be greater sequence variation for *Lsi2* (and the other *Lsi* genes) than that  
5 409 is represented in the 5 accessions reported here. A focus for future study will be to expand  
6 410 sequence information to more accessions to more fully explore sequence variation associated  
7 411 with the polymorphic SNPs presented in figure 6.  
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19 413 **5 Conclusion**  
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414 This study has demonstrated strong genotypic differences in shoot silicon in a diverse  
415 collection of rice cultivars, showing that there is potential to breed rice with increased silicon  
416 content that could improve resistance to both biotic and abiotic stresses in rice, which would  
417 help to maintain crop yields. The identification of significant SNPs linked with the shoot  
418 silicon phenotype within 10 kb of known silicon transporters warrants further study to  
419 investigate the impact of different alleles of these genes on silicon and arsenic accumulation  
420 in rice. Furthermore, the XRF method of silicon determination could be applied to GWA  
421 mapping studies that might reveal further candidate genes for silicon content in rice.

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18       428 **References**  
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3 647 **Figure 1:** Silicon in different organs of rice (bars are the mean of four replicates and error  
4 648 bars represent standard error of the mean). Letters above the columns (upper-case = anaerobic  
5 649 and lower case = aerobic) indicate statistically significant differences in silicon content of  
6 650 different plant organs using Tukey's test in two conditions. \*denotes a significant difference  
7 651 between the two treatments for that plant organ.  
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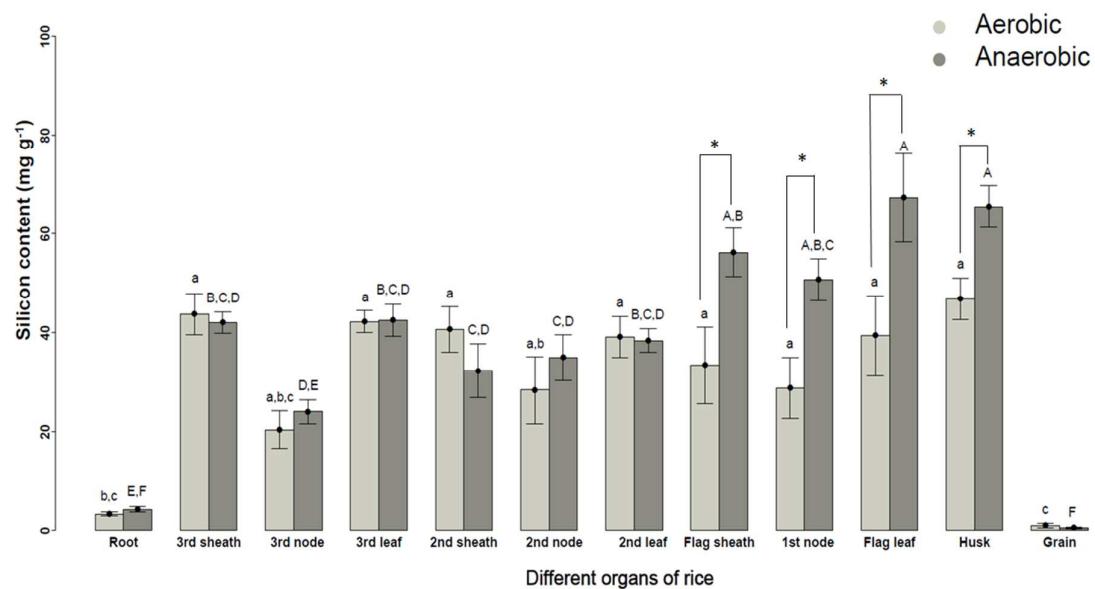
10 653 **Figure 2:** Mean shoot silicon ( $\text{mg g}^{-1}$ ) of 50 rice accessions determined by FIA. Different  
11 654 symbols refer to the accessions belonging to the different sub-populations; circle = *aus*,  
12 655 square = *indica*, cross = *aromatic*, triangle = *tropical japonica*, upside down triangle =  
13 656 *temperate japonica*. Error bars indicate the standard error of the mean ( $n = 4$ ).  
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15 657 **Figure 3:** Shoot silicon ( $\text{mg g}^{-1}$ ) content of 50 accessions in 5 different sub-populations of  
16 658 rice. ARO = *aromatic*, AUS = *aus*, IND = *indica*, TEJ = *temperate japonica* and TRJ =  
17 659 *tropical japonica*. The edges of each box show the upper and lower quantile and the bold line  
18 660 in the box shows the median value and the dotted line the mean value. The whiskers are the  
19 661 10<sup>th</sup> and 90<sup>th</sup> percentiles.  
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21 662 **Figure 4:** Correlation of mean shoot silicon in 19 rice accessions determined by FIA and P-  
22 663 XRF. Error bars indicate the standard error of the mean ( $n = 4$ ). Dotted line is the 1:1 line.  
23

24 664 **Figure 5:** Correlation between shoot silicon ( $\text{mg g}^{-1}$ ) and grain arsenic ( $\mu\text{g kg}^{-1}$ ) in ARO=  
25 665 *aromatic*, AUS = *aus*, IND = *indica*, TEJ = *temperate japonica* and TRJ = *tropical japonica*  
26 666 subpopulations.  
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28 667 **Figure 6:** Variation in shoot silicon ( $\text{mg g}^{-1}$ ) between different SNPs within 10 kb of *Lsi2* and  
29 668 *Lsi3*  
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675 **Figure 1**

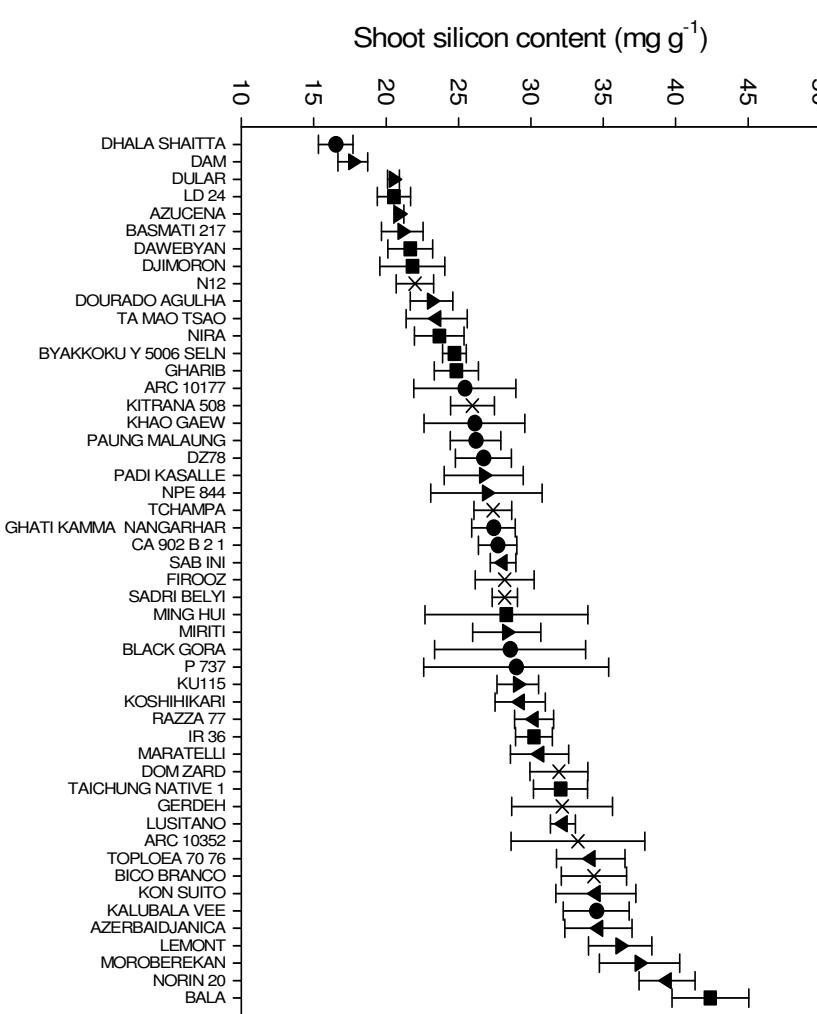
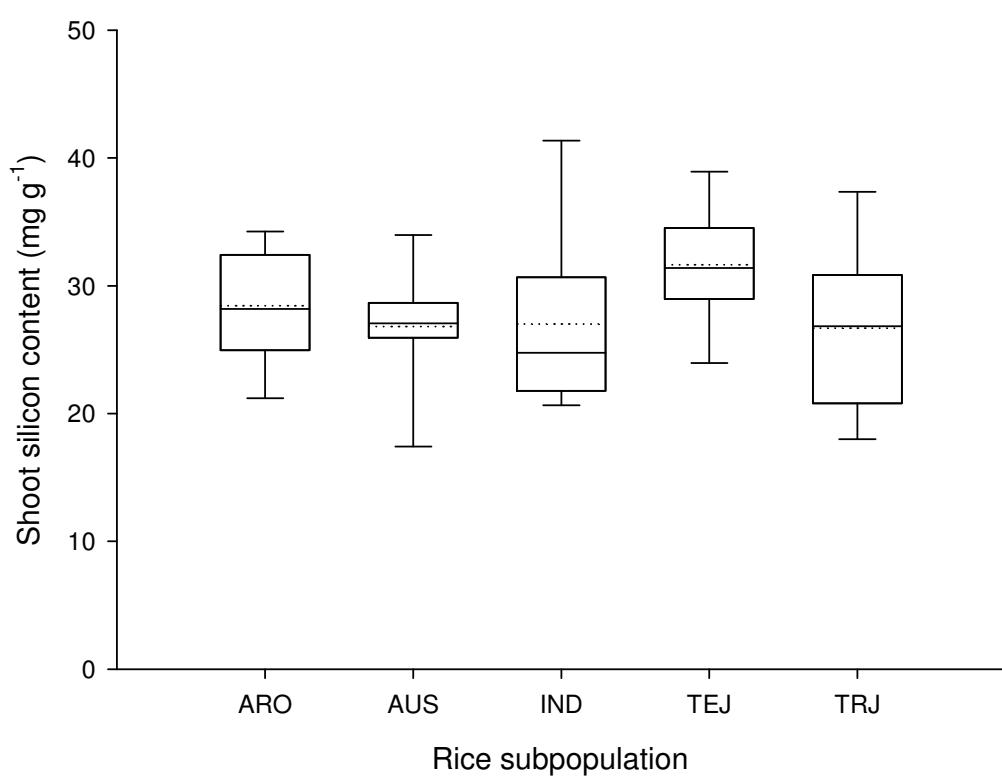


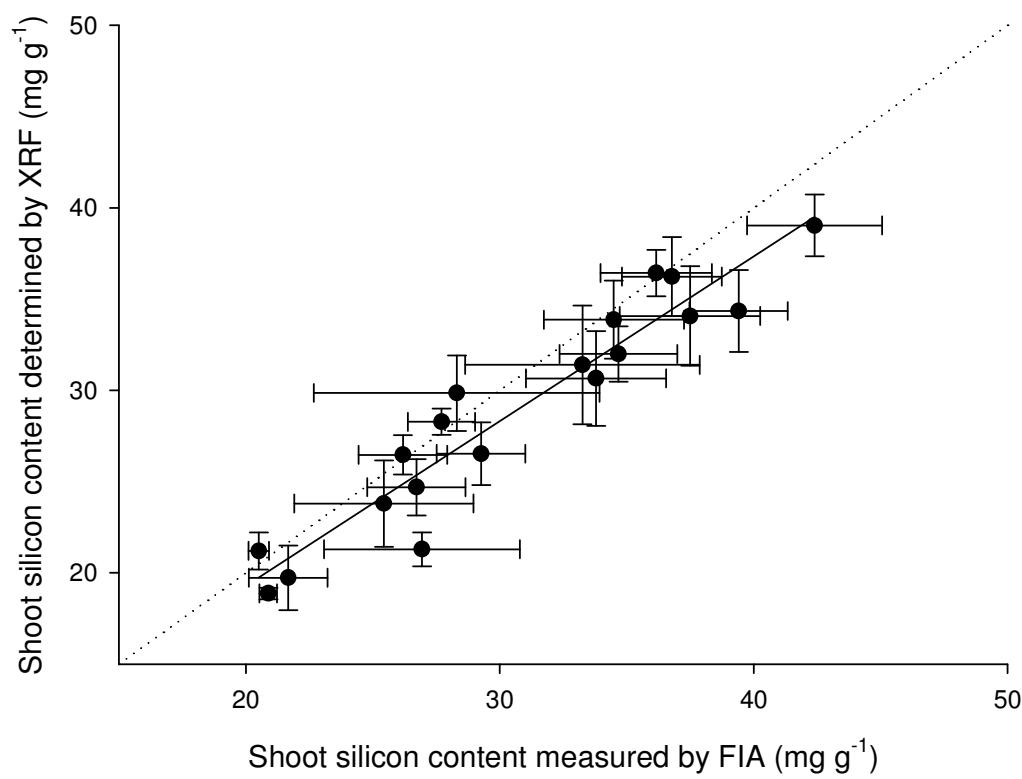
Figure 2

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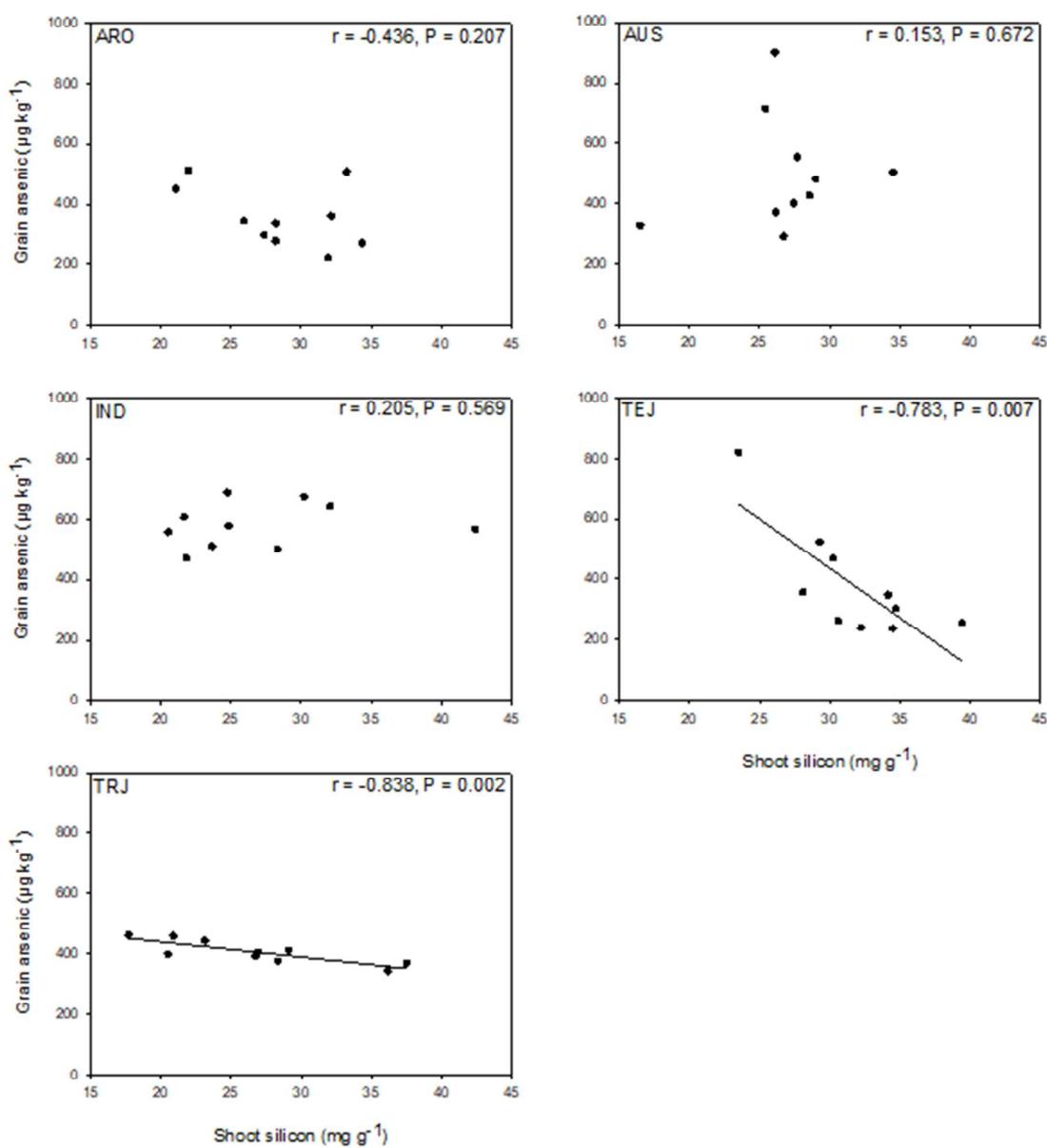
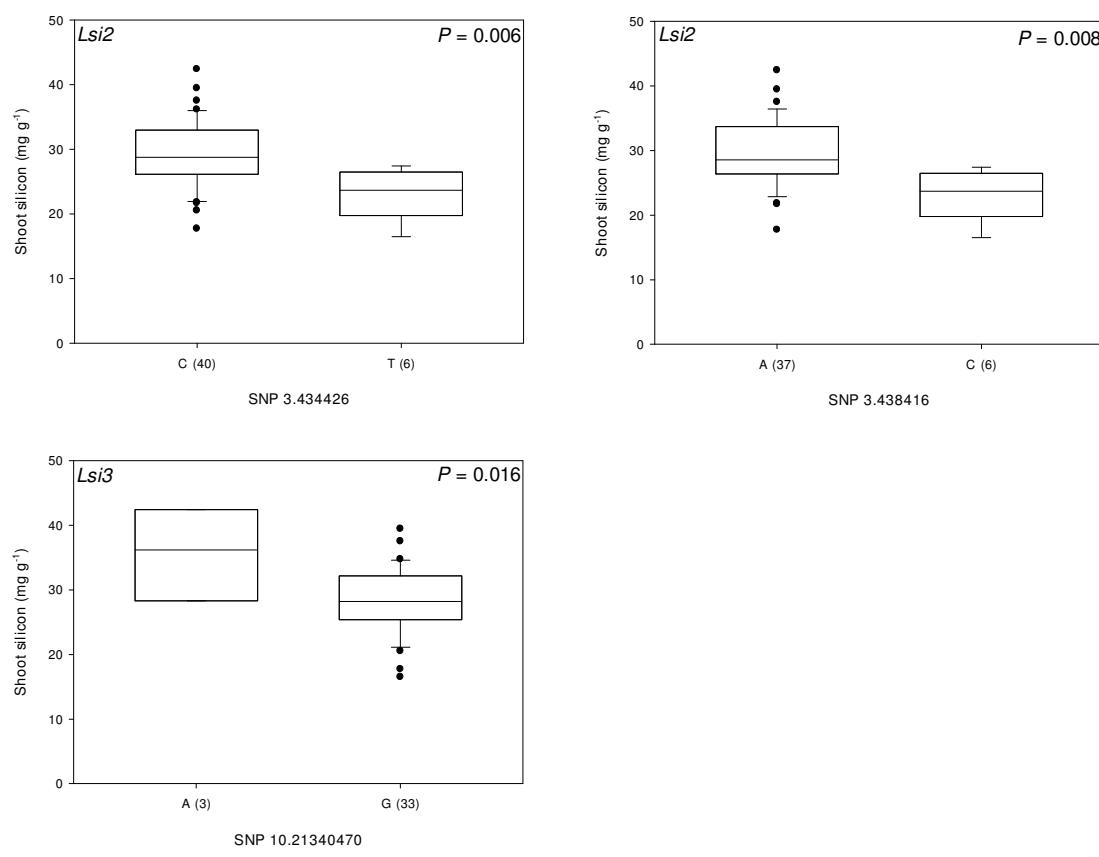
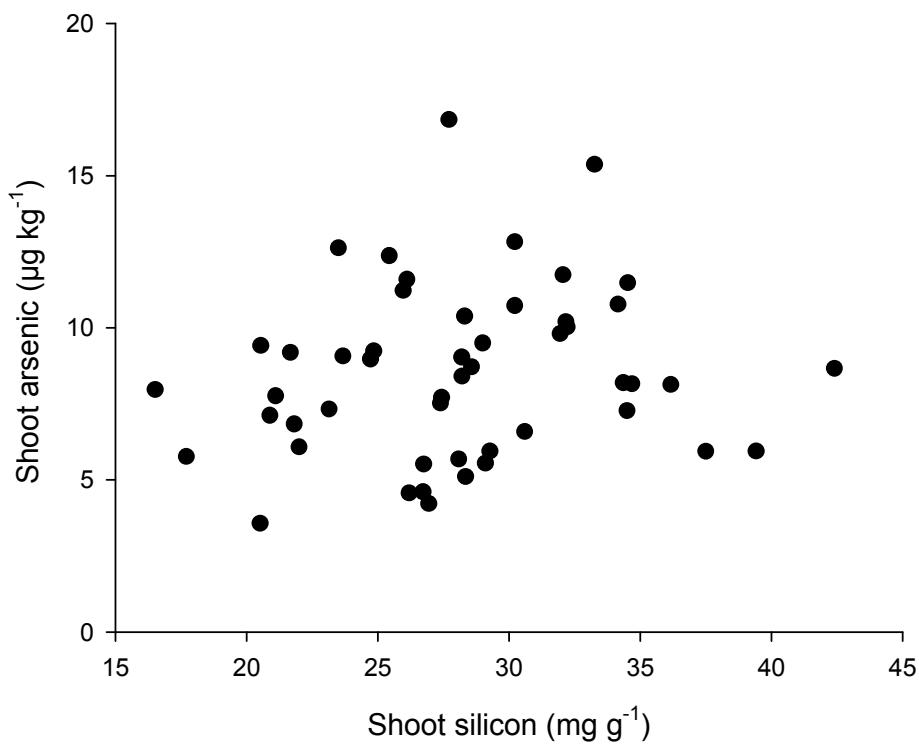


Figure 5

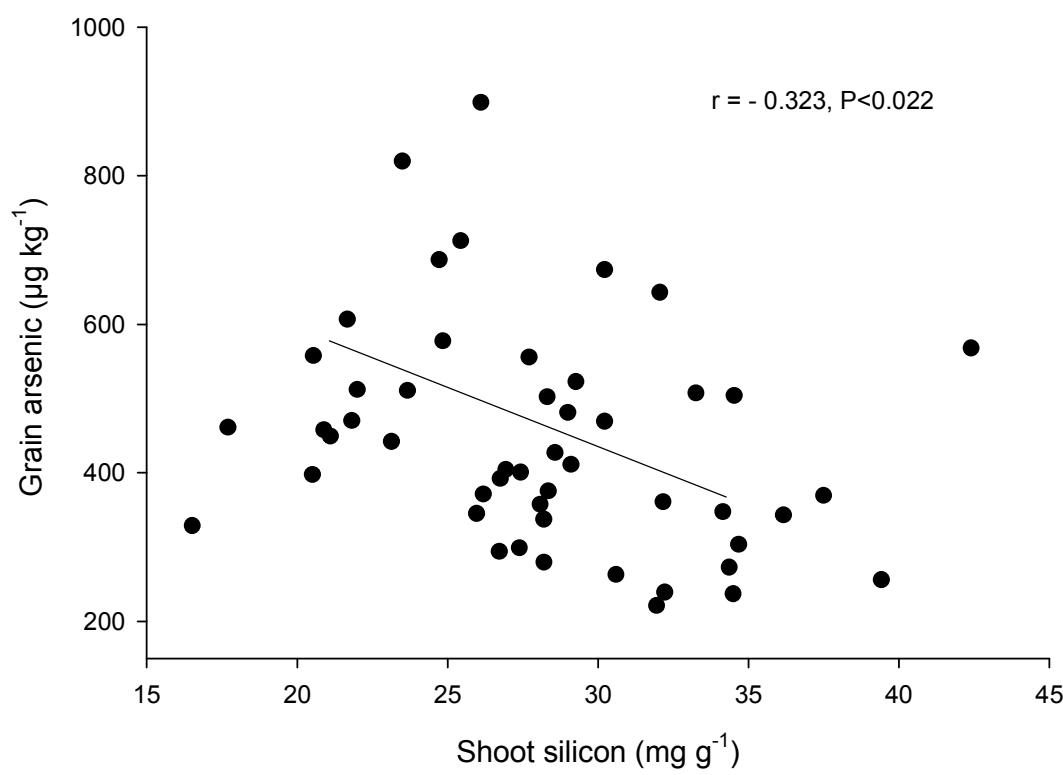
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2      706 **Supplementary Table 1:** Selected genotype from RDP1 for shoot silicon analysis  
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4      707 **Supplementary Table 2:** SNPs with 10 kb (upstream and downstream) of *Lsi2*. SNPs data  
5 taken from the High-density rice array (HDRA, 700k SNPs data) available at  
6 <http://ricediversity.org/data/index.cfm>. After each SNP name (in brackets) is the location of  
7 that SNP aligned to the Rice Genome Annotation Project – release 7 sequence available at  
8 <http://rice.plantbiology.msu.edu/>  
9  
10     712 **Supplementary Table 3:** SNPs with 10 kb (upstream and downstream) of *Lsi3*. SNPs data  
11 taken from the High-density rice array (HDRA, 700k SNPs data) available at  
12 <http://ricediversity.org/data/index.cfm>. After each SNP name (in brackets) is the location of  
13 that SNP aligned to the Rice Genome Annotation Project – release 7 sequence available at  
14 <http://rice.plantbiology.msu.edu/>  
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**Supplementary figure 1:** Correlation between shoot silicon (mg g<sup>-1</sup>) and shoot arsenic (µg kg<sup>-1</sup>) within 50 accessions of RDP1.



**Supplementary figure 2:** Correlation between shoot silicon (mg g<sup>-1</sup>) and grain arsenic (µg kg<sup>-1</sup>) within 50 accessions of RDP1.

## Lsi2 (LOC\_Os03g01700)



10	Bala	AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGTACAGAGAGAGATCAT
11	IR64	AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGTACAGAGAGAGATCAT
12	DJ123	AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGTACAGAGAGAGATCAT
13	Nipponbare	AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGTACAGAGAGAGATCAT
14	Azucena	AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGTACAGAGAGAGATCAT
15		*****
16	Bala	CAGCTCAAGTAGCTAAAGCTAGCTATAATCTGCTAGCTAGTCGATCAGACACTTAATTAC
17	IR64	CAGCTCAAGTAGCTAAAGCTAGCTATAATCTGCTAGCTAGTCGATCAGACACTTAATTAC
18	DJ123	CAGCTCAAGTAGCTAAAGCTAGCTATAATCTGCTAGCTAGTCGATCAGACACTTAATTAC
19	Nipponbare	CAGCTCAAGTAGCTAAAGCTAGCTATAATCTGCTAGCTAGTCGATCAGACACTTAATTAC
20	Azucena	CAGCTCAAGTAGCTAAAGCTAGCTATAATCTGCTAGCTAGTCGATCAGACACTTAATTAC
21		*****
22	Bala	CTGCTAGGTGGTGGTCATCGAAGAAGAAGAAGATGAGTGAGCTGGCTCGGCCAAG
23	IR64	CTGCTAGGTGGTGGTCATCGAAGAAGAAGAAGATGAGTGAGCTGGCTCGGCCAAG
24	DJ123	CTGCTAGGTGGTGGTCATCGAAGAAGAAGAAGATGAGTGAGCTGGCTCGGCCAAG
25	Nipponbare	CTGCTAGGTGGTGGTCATCGAAGAAGAAGAAGATGAGTGAGCTGGCTCGGCCAAG
26	Azucena	CTGCTAGGTGGTGGTCATCGAAGAAGAAGAAGATGAGTGAGCTGGCTCGGCCAAG
27		*****
28	Bala	GTGGCGCTTGGATCGATCGCGTTCGCGGTGTTCTGGATGATGGCGGTGTTCCGTCGGTG
29	IR64	GTGGCGCTTGGATCGATCGCGTTCGCGGTGTTCTGGATGATGGCGGTGTTCCGTCGGTG
30	DJ123	GTGGCGCTTGGATCGATCGCGTTCGCGGTGTTCTGGATGATGGCGGTGTTCCGTCGGTG
31	Nipponbare	GTGGCGCTTGGATCGATCGCGTTCGCGGTGTTCTGGATGATGGCGGTGTTCCGTCGGTG
32	Azucena	GTGGCGCTTGGATCGATCGCGTTCGCGGTGTTCTGGATGATGGCGGTGTTCCGTCGGTG
33		*****
34	Bala	CCGTTCTCGCGATCGGGCGACGGCGGGGTCGCTGCTGAGCGCGTGTGATGGTGATA
35	IR64	CCGTTCTCGCGATCGGGCGACGGCGGGGTCGCTGCTGAGCGCGTGTGATGGTGATA
36	DJ123	CCGTTCTCGCGATCGGGCGACGGCGGGGTCGCTGCTGAGCGCGTGTGATGGTGATA
37	Nipponbare	CCGTTCTCGCGATCGGGCGACGGCGGGGTCGCTGCTGAGCGCGTGTGATGGTGATA
38	Azucena	CCGTTCTCGCGATCGGGCGACGGCGGGGTCGCTGCTGAGCGCGTGTGATGGTGATA
39		*****
40	Bala	TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCATCGACCTCCAACTCTGGGCCTC
41	IR64	TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCATCGACCTCCAACTCTGGGCCTC
42	DJ123	TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCATCGACCTCCAACTCTGGGCCTC
43	Nipponbare	TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCATCGACCTCCAACTCTGGGCCTC
44	Azucena	TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCATCGACCTCCAACTCTGGGCCTC
45		*****
46	Bala	CTCTTCGCCACCATGGTGGTGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
47	IR64	CTCTTCGCCACCATGGTGGTGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
48	DJ123	CTCTTCGCCACCATGGTGGTGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
49	Nipponbare	CTCTTCGCCACCATGGTGGTGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
50	Azucena	CTCTTCGCCACCATGGTGGTGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
51		*****
52	Bala	GGGCGTCTGCTGGCGTGGAAAGAGGCCAGGGCGGGCGCACCTCATGTGCCCGTCTCGC
53	IR64	GGGCGTCTGCTGGCGTGGAAAGAGGCCAGGGCGGGCGCACCTCATGTGCCCGTCTCGC
54	DJ123	GGGCGTCTGCTGGCGTGGAAAGAGGCCAGGGCGGGCGCACCTCATGTGCCCGTCTCGC
55	Nipponbare	GGGCGTCTGCTGGCGTGGAAAGAGGCCAGGGCGGGCGCACCTCATGTGCCCGTCTCGC
56	Azucena	GGGCGTCTGCTGGCGTGGAAAGAGGCCAGGGCGGGCGCACCTCATGTGCCCGTCTCGC
57		*****

Bala	GTCACCGCCCTGCCAGGCCCTTCACCAAGCACACCTGCTGCGTGTCTCACCAGG
IR64	GTCACCGCCCTGCCAGGCCCTTCACCAAGCACACCTGCTGCGTGTCTCACCAGG
DJ123	GTCACCGCCCTGCCAGGCCCTTCACCAAGCACACCTGCTGCGTGTCTCACCAGG
Nipponbare	GTCACCGCCCTGCCAGGCCCTTCACCAAGCACACCTGCTGCGTGTCTCACCAGG
Azucena	GTCACCGCCCTGCCAGGCCCTTCACCAAGCACACCTGCTGCGTGTCTCACCAGG
*****	
Bala	TTCGTCTCGAGCTGCCGCCAGGCACACCTCCCCGCAAGCCCTTCCTCGCCCTC
IR64	TTCGTCTCGAGCTGCCGCCAGGCACACCTCCCCGCAAGCCCTTCCTCGCCCTC
DJ123	TTCGTCTCGAGCTGCCGCCAGGCACACCTCCCCGCAAGCCCTTCCTCGCCCTC
Nipponbare	TTCGTCTCGAGCTGCCGCCAGGCACACCTCCCCGCAAGCCCTTCCTCGCCCTC
Azucena	TTCGTCTCGAGCTGCCGCCAGGCACACCTCCCCGCAAGCCCTTCCTCGCCCTC
*****	
Bala	GCCTCCAGGCCAACATGGCTCCGCCACCCCCATGGCAACCCCCAGAACCTGGTC
IR64	GCCTCCAGGCCAACATGGCTCCGCCACCCCCATGGCAACCCCCAGAACCTGGTC
DJ123	GCCTCCAGGCCAACATGGCTCCGCCACCCCCATGGCAACCCCCAGAACCTGGTC
Nipponbare	GCCTCCAGGCCAACATGGCTCCGCCACCCCCATGGCAACCCCCAGAACCTGGTC
Azucena	GCCTCCAGGCCAACATGGCTCCGCCACCCCCATGGCAACCCCCAGAACCTGGTC
*****	
Bala	ATCGCCTCAATTAGCAAGATCACCTTCCCCAAGTTCTCATGGGAATCTCCCGGCCATG
IR64	ATCGCCTCAATTAGCAAGATCACCTTCCCCAAGTTCTCATGGGAATCTCCCGGCCATG
DJ123	ATCGCCTCAATTAGCAAGATCACCTTCCCCAAGTTCTCATGGGAATCTCCCGGCCATG
Nipponbare	ATCGCCTCAATTAGCAAGATCACCTTCCCCAAGTTCTCATGGGAATCTCCCGGCCATG
Azucena	ATCGCCTCAATTAGCAAGATCACCTTCCCCAAGTTCTCATGGGAATCTCCCGGCCATG
*****	
Bala	CTCGTGGGATGGCCGTCAACATGGTCATGCTCTGCATGTACTGGAGGGAGCTGGGC
IR64	CTCGTGGGATGGCCGTCAACATGGTCATGCTCTGCATGTACTGGAGGGAGCTGGGC
DJ123	CTCGTGGGATGGCCGTCAACATGGTCATGCTCTGCATGTACTGGAGGGAGCTGGGC
Nipponbare	CTCGTGGGATGGCCGTCAACATGGTCATGCTCTGCATGTACTGGAGGGAGCTGGGC
Azucena	CTCGTGGGATGGCCGTCAACATGGTCATGCTCTGCATGTACTGGAGGGAGCTGGGC
*****	
Bala	GGAGGGGGCGAGCTCAGCGTCAGCGCAAGCAGATGGAGGGGGTGAGGCAAGGTG
IR64	GGAGGGGGCGAGCTCAGCGTCAGCGCAAGCAGATGGAGGGGGTGAGGCAAGGTG
DJ123	GGAGGGGGCGAGCTCAGCGTCAGCGCAAGCAGATGGAGGGGGTGAGGCAAGGTG
Nipponbare	GGAGGGGGCGAGCTCAGCGTCAGCGCAAGCAGATGGAGGGGGTGAGGCAAGGTG
Azucena	GGAGGGGGCGAGCTCAGCGTCAGCGCAAGCAGATGGAGGGGGTGAGGCAAGGTG
*****	
Bala	CCGGCGTGGCCAAGAGCACGCCAGCTGAACGGCAACGGCACAGCATGATGTCGCTG
IR64	CCGGCGTGGCCAAGAGCACGCCAGCTGAACGGCAACGGCACAGCATGATGTCGCTG
DJ123	CCGGCGTGGCCAAGAGCACGCCAGCTGAACGGCAACGGCACAGCATGATGTCGCTG
Nipponbare	CCGGCGTGGCCAAGAGCACGCCAGCTGAACGGCAACGGCACAGCATGATGTCGCTG
Azucena	CCGGCGTGGCCAAGAGCACGCCAGCTGAACGGCAACGGCACAGCATGATGTCGCTG
*****	
Bala	GAGATGTCGGAGAACATAACGACCAAGCACCCATGGTCATGCACTGACGGAGGGCGGG
IR64	GAGATGTCGGAGAACATAACGACCAAGCACCCATGGTCATGCACTGACGGAGGGCGGG
DJ123	GAGATGTCGGAGAACATAACGACCAAGCACCCATGGTCATGCACTGACGGAGGGCGGG
Nipponbare	GAGATGTCGGAGAACATAACGACCAAGCACCCATGGTCATGCACTGACGGAGGGCGGG
Azucena	GAGATGTCGGAGAACATAACGACCAAGCACCCATGGTCATGCACTGACGGAGGGCGGG
*****	
Bala	CGGAAGCTGTTCTCAAGAGCTTCGCGTACGTGGTACGGTGGGGATGGTGGTGGCTAC
IR64	CGGAAGCTGTTCTCAAGAGCTTCGCGTACGTGGTACGGTGGGGATGGTGGTGGCTAC
DJ123	CGGAAGCTGTTCTCAAGAGCTTCGCGTACGTGGTACGGTGGGGATGGTGGTGGCTAC
Nipponbare	CGGAAGCTGTTCTCAAGAGCTTCGCGTACGTGGTACGGTGGGGATGGTGGTGGCTAC
Azucena	CGGAAGCTGTTCTCAAGAGCTTCGCGTACGTGGTACGGTGGGGATGGTGGTGGCTAC

1  
 2                   Bala  
 3                   IR64  
 4                   DJ123  
 5                   Nipponbare  
 6                   Azucena  
 7                   \*\*\*\*\*  
 8                   Bala  
 9                   IR64  
 10                  DJ123  
 11                  Nipponbare  
 12                  Azucena  
 13                  \*\*\*\*\*  
 14                  Bala  
 15                  IR64  
 16                  DJ123  
 17                  Nipponbare  
 18                  Azucena  
 19                  \*\*\*\*\*  
 20                  Bala  
 21                  IR64  
 22                  DJ123  
 23                  Nipponbare  
 24                  Azucena  
 25                  \*\*\*\*\*  
 26                  Bala  
 27                  IR64  
 28                  DJ123  
 29                  Nipponbare  
 30                  Azucena  
 31                  \*\*\*\*\*  
 32                  Bala  
 33                  IR64  
 34                  DJ123  
 35                  Nipponbare  
 36                  Azucena  
 37                  \*\*\*\*\*  
 38                  Bala  
 39                  IR64  
 40                  DJ123  
 41                  Nipponbare  
 42                  Azucena  
 43                  \*\*\*\*\*  
 44                  Bala  
 45                  IR64  
 46                  DJ123  
 47                  Nipponbare  
 48                  Azucena  
 49                  \*\*\*\*\*  
 50                  Bala  
 51                  IR64  
 52                  DJ123  
 53                  Nipponbare  
 54                  Azucena  
 55                  \*\*\*\*\*  
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ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC
ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC
ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC
ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC
*****  

GACTTC CGCGACGCCAGCCGTGCCTGGACACCGTGTCCCTACTCGCTGCTCGTCTTCTTC
GACTTC CGCGACGCCAGCCGTGCCTGGACACCGTGTCCCTACTCGCTGCTCGTCTTCTTC
GACTTC CGCGACGCCAGCCGTGCCTGGACACCGTGTCCCTACTCGCTGCTCGTCTTCTTC
GACTTC CGCGACGCCAGCCGTGCCTGGACACCGTGTCCCTACTCGCTGCTCGTCTTCTTC
GACTTC CGCGACGCCAGCCGTGCCTGGACACCGTGTCCCTACTCGCTGCTCGTCTTCTTC
*****  

TCCGGGATGTTCATACCGTCAGCGGCTTCAACAAGACGGGCTCCGGGAGCCATCTGG
TCCGGGATGTTCATACCGTCAGCGGCTTCAACAAGACGGGCTCCGGGAGCCATCTGG
TCCGGGATGTTCATACCGTCAGCGGCTTCAACAAGACGGGCTCCGGGAGCCATCTGG
TCCGGGATGTTCATACCGTCAGCGGCTTCAACAAGACGGGCTCCGGGAGCCATCTGG
TCCGGGATGTTCATACCGTCAGCGGCTTCAACAAGACGGGCTCCGGGAGCCATCTGG
*****  

GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGGGCATCTCCGTCTCTCCGTC
GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGGGCATCTCCGTCTCTCCGTC
GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGGGCATCTCCGTCTCTCCGTC
GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGGGCATCTCCGTCTCTCCGTC
GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGGGCATCTCCGTCTCTCCGTC
*****  

ATCATCCTCCCTCTCCAACCTCGCATCAAACGTACCAACGGTAATTATAACATTCTAT
ATCATCCTCCCTCTCCAACCTCGCATCAAACGTACCAACGGTAATTATAACATTCTAT
ATCATCCTCCCTCTCCAACCTCGCATCAAACGTACCAACGGTAATTATAACATTCTAT
ATCATCCTCCCTCTCCAACCTCGCATCAAACGTACCAACGGTAATTATAACATTCTAT
ATCATCCTCCCTCTCCAACCTCGCATCAAACGTACCAACGGTAATTATAACATTCTAT
*****  

TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATAACAAATAT
TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATAACAAATAT
TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATAACAAATAT
TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATAACAAATAT
TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATAACAAATAT
*****  

ATATGCGCATGCATGCA GTGCTTCTTATGGGTGATGAGGTGGCGAAGGGCGCGCGCTGA
ATATGCGCATGCATGCA GTGCTTCTTATGGGTGATGAGGTGGCGAAGGGCGCGCGCTGA
ATATGCGCATGCATGCA GTGCTTCTTATGGGTGATGAGGTGGCGAAGGGCGCGCGCTGA
ATATGCGCATGCATGCA GTGCTTCTTATGGGTGATGAGGTGGCGAAGGGCGCGCGCTGA
ATATGCGCATGCATGCA GTGCTTCTTATGGGTGATGAGGTGGCGAAGGGCGCGCGCTGA
*****  

TATCGCCGGCGCGCGTGACGACGTGCTGGCTGCTGGCGTGGTGAGCACGGTGGCGG
TATCGCCGGCGCGCGTGACGACGTGCTGGCTGCTGGCGTGGTGAGCACGGTGGCGG
TATCGCCGGCGCGCGTGACGACGTGCTGGCTGCTGGCGTGGTGAGCACGGTGGCGG
TATCGCCGGCGCGCGTGACGACGTGCTGGCTGCTGGCGTGGTGAGCACGGTGGCGG
TATCGCCGGCGCGCGTGACGACGTGCTGGCTGCTGGCGTGGTGAGCACGGTGGCGG
*****  

GGAACCTGTCGCTGCTGGGTGGCGGCGAACCTGATAGTGTGCGAGCAGGCAGGGAGGG
GGAACCTGTCGCTGCTGGGTGGCGGCGAACCTGATAGTGTGCGAGCAGGCAGGGAGGG
GGAACCTGTCGCTGCTGGGTGGCGGCGAACCTGATAGTGTGCGAGCAGGCAGGGAGGG
GGAACCTGTCGCTGCTGGGTGGCGGCGAACCTGATAGTGTGCGAGCAGGCAGGGAGGG
GGAACCTGTCGCTGCTGGGTGGCGGCGAACCTGATAGTGTGCGAGCAGGCAGGGAGGG
*****  


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Bala	CCCCCAGGAACGCCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
IR64	CCCCCAGGAACGCCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
DJ123	CCCCCAGGAACGCCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
Nipponbare	CCCCCAGGAACGCCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
Azucena	*****
Bala	CCCTCATCGTACCGCCGTCGGCATACCCCTATCGCAAGATCTGA TCTCATCTCATCG
IR64	CCCTCATCGTACCGCCGTCGGCATACCCCTATCGCAAGATCTGA TCTCATCTCATCG
DJ123	CCCTCATCGTACCGCCGTCGGCATACCCCTATCGCAAGATCTGA TCTCATCTCATCG
Nipponbare	CCCTCATCGTACCGCCGTCGGCATACCCCTATCGCAAGATCTGA TCTCATCTCATCG
Azucena	CCCTCATCGTACCGCCGTCGGCATACCCCTATCGCAAGATCTGA TCTCATCTCATCG
Bala	*****
IR64	ACCCATCAAATTAAATTAAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTGTCGT
DJ123	ACCCATCAAATTAAATTAAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTGTCGT
Nipponbare	ACCCATCAAATTAAATTAAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTGTCGT
Azucena	ACCCATCAAATTAAATTAAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTGTCGT
Bala	*****
IR64	CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGACGTATATATATGCT
DJ123	CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGACGTATATATATGCT
Nipponbare	CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGACGTATATATATGCT
Azucena	CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGACGTATATATATGCT
Bala	*****
IR64	CCTCAACCTCAAGTCATTAATTAAAGAGAATGGATGAATGAATCAATGTTGTATGATT
DJ123	CCTCAACCTCAAGTCATTAATTAAAGAGAATGGATGAATGAATCAATGTTGTATGATT
Nipponbare	CCTCAACCTCAAGTCATTAATTAAAGAGAATGGATGAATGAATCAATGTTGTATGATT
Azucena	CCTCAACCTCAAGTCATTAATTAAAGAGAATGGATGAATGAATCAATGTTGTATGATT
Bala	*****
IR64	TCCCTTTGTTTTGTTTACCCACCGTATAATGTGCTGTGGTGTATGGCGGGCGAGCT
DJ123	TCCCTTTGTTTTGTTTACCCACCGTATAATGTGCTGTGGTGTATGGCGGGCGAGCT
Nipponbare	TCCCTTTGTTTTGTTTACCCACCGTATAATGTGCTGTGGTGTATGGCGGGCGAGCT
Azucena	TCCCTTTGTTTTGTTTACCCACCGTATAATGTGCTGTGGTGTATGGCGGGCGAGCT
Bala	*****
IR64	TGAAATTGAATTGGCCTGCTGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
DJ123	TGAAATTGAATTGGCCTGCTGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
Nipponbare	TGAAATTGAATTGGCCTGCTGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
Azucena	TGAAATTGAATTGGCCTGCTGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
Bala	*****
IR64	GCTAAATCTGCTTAATTTCATCAAGTGTCAATCATGATGAGAAGAGAGGGAACTAGATG
DJ123	GCTAAATCTGCTTAATTTCATCAAGTGTCAATCATGATGAGAAGAGAGGGAACTAGATG
Nipponbare	GCTAAATCTGCTTAATTTCATCAAGTGTCAATCATGATGAGAAGAGAGGGAACTAGATG
Azucena	GCTAAATCTGCTTAATTTCATCAAGTGTCAATCATGATGAGAAGAGAGGGAACTAGATG
Bala	*****
IR64	GAGCTAGCTAGCAACACATGCAGTGTGCTAACGAGCTGCAGTCAGTGCACCTGCATGCTAAA
DJ123	GAGCTAGCTAGCAACACATGCAGTGTGCTAACGAGCTGCAGTCAGTGCACCTGCATGCTAAA
Nipponbare	GAGCTAGCTAGCAACACATGCAGTGTGCTAACGAGCTGCAGTCAGTGCACCTGCATGCTAAA
Azucena	GAGCTAGCTAGCAACACATGCAGTGTGCTAACGAGCTGCAGTCAGTGCACCTGCATGCTAAA
Bala	*****

Bala	TTTGCTTGCCTAACATCGTATTACATGTATGTTGATATGTCATGTTCCCTCAAAA
IR64	TTTGCTTGCCTAACATCGTATTACATGTATGTTGATATGTCATGTTCCCTCAAAA
DJ123	TTTGCTTGCCTAACATCGTATTACATGTATGTTGATATGTCATGTTCCCTCAAAA
Nipponbare	TTTGCTTGCCTAACATCGTATTACATGTATGTTGATATGTCATGTTCCCTCAAAA
Azucena	TTTGCTTGCCTAACATCGTATTACATGTATGTTGATATGTCATGTTCCCTCAAAA
*****	
Bala	TACATGGAAAAAAATGTTCTACACATCTCAATCGTATAGAGTATGTCATGCTTGT
IR64	TACATGGAAAAAAATGTTCTACACATCTCAATCGTATAGAGTATGTCATGCTTGT
DJ123	TACATGGAAAAAAATGTTCTACACATCTCAATCGTATAGAGTATGTCATGCTTGT
Nipponbare	TACATGGAAAAAAATGTTCTACACATCTCAATCGTATAGAGTATGTCATGCTTGT
Azucena	TACATGGAAAAAAATGTTCTACACATCTCAATCGTATAGAGTATGTCATGCTTGT
*****	
Bala	TCAAACAAATATTGTAGCCCTCTCTCTCTCTCTCTTTAGGTTATTAATTAAATTAGT
IR64	TCAAACAAATATTGTAGCCCTCTCTCTCTCTCTCTCTTTAGGTTATTAATTAAATTAGT
DJ123	TCAAACAAATATTGTAGCCCTCTCTCTCTCTCTCTCTCTTTAGGTTATTAATTAAATTAGT
Nipponbare	TCAAACAAATATTGTAGCCCTCTCTCTCTCTCTCTCTCTCTTTAGGTTATTAATTAAATTAGT
Azucena	TCAAACAAATATTGTAGCCCTCTCTCTCTCTCTCTCTCTCTTTAGGTTATTAATTAAATTAGT
*****	
Bala	CTGTTCAATGTCACCTCAGTTCATAGAGATGCTACTTAATTAAATTGCTCCATGTATGC
IR64	CTGTTCAATGTCACCTCAGTTCATAGAGATGCTACTTAATTAAATTGCTCCATGTATGC
DJ123	CTGTTCAATGTCACCTCAGTTCATAGAGATGCTACTTAATTAAATTGCTCCATGTATGC
Nipponbare	CTGTTCAATGTCACCTCAGTTCATAGAGATGCTACTTAATTAAATTGCTCCATGTATGC
Azucena	CTGTTCAATGTCACCTCAGTTCATAGAGATGCTACTTAATTAAATTGCTCCATGTATGC
*****	
Bala	AGAGTTGTACTACCTTAAATATCAGTATGCATGCTCTTGTGCCATTCCAAAAGAACAA
IR64	AGAGTTGTACTACCTTAAATATCAGTATGCATGCTCTTGTGCCATTCCAAAAGAACAA
DJ123	AGAGTTGTACTACCTTAAATATCAGTATGCATGCTCTTGTGCCATTCCAAAAGAACAA
Nipponbare	AGAGTTGTACTACCTTAAATATCAGTATGCATGCTCTTGTGCCATTCCAAAAGAACAA
Azucena	AGAGTTGTACTACCTTAAATATCAGTATGCATGCTCTTGTGCCATTCCAAAAGAACAA
*****	
Bala	A
IR64	A
DJ123	A
Nipponbare	A
Azucena	A
	*

**Supplementary figure 3.** Genomic sequence alignment of *Lsi2*. Regions highlighted in bold are predicted exon regions based on the Rice Genome Annotation Project – release 7 sequence available at <http://rice.plantbiology.msu.edu/>.

1  
2  
3 *Lsi3* (LOC\_Os10g39980)  
45'  3'

6  
7  
8     Bala                         ACTCTCATGCCACCACCCACCTCCCACCTCCAAGACGCTAGTCATTGTAGTGAGAGAGA  
9     IR64                         ACTCTCATGCCACCACCCACCTCCCACCTCCAAGACGCTAGTCATTGTAGTGAGAGAGA  
10    DJ123                         ACTCTCATGCCACCACCCACCTCCCACCTCCAAGACGCTAGTCATTGTAGTGAGAGAGA  
11    Nipponbare                     ACTCTCATGCCACCACCCACCTCCCACCTCCAAGACGCTAGTCATTGTAGTGAGAGAGA  
12    Azucena                         ACTCTCATGCCACCACCCACCTCCCACCTCCAAGACGCTAGTCATTGTAGTGAGAGAGA  
13                                 \*\*\*\*\*  
14    Bala                         GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGTATGGGTCGGTGGCGTTCGGT  
15    IR64                         GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGTATGGGTCGGTGGCGTTCGGT  
16    DJ123                         GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGTATGGGTCGGTGGCGTTCGGT  
17    Nipponbare                     GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGTATGGGTCGGTGGCGTTCGGT  
18    Azucena                         GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGTATGGGTCGGTGGCGTTCGGT  
19                                 \*\*\*\*\*  
20    Bala                         GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCCCTGCCGATCGGGCGGACGGCG  
21    IR64                         GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCCCTGCCGATCGGGCGGACGGCG  
22    DJ123                         GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCCCTGCCGATCGGGCGGACGGCG  
23    Nipponbare                     GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCCCTGCCGATCGGGCGGACGGCG  
24                                 \*\*\*\*\*  
25    Bala                         GGGGCCTGCTGGCGCGTGCTGATGATCGTGTTCACCGTATCAGCGCCGACGACGCC  
26    IR64                         GGGGCCTGCTGGCGCGTGCTGATGATCGTGTTCACCGTATCAGCGCCGACGACGCC  
27    DJ123                         GGGGCCTGCTGGCGCGTGCTGATGATCGTGTTCACCGTATCAGCGCCGACGACGCC  
28    Nipponbare                     GGGGCCTGCTGGCGCGTGCTGATGATCGTGTTCACCGTATCAGCGCCGACGACGCC  
29    Azucena                         GGGGCCTGCTGGCGCGTGCTGATGATCGTGTTCACCGTATCAGCGCCGACGACGCC  
30                                 \*\*\*\*\*  
31    Bala                         TACGCTCCATCGACCTCCCCATCCTGGGCCCTCTTCGCCACCATGGTCGTGGCGG  
32    IR64                         TACGCTCCATCGACCTCCCCATCCTGGGCCCTCTTCGCCACCATGGTCGTGGCGG  
33    DJ123                         TACGCTCCATCGACCTCCCCATCCTGGGCCCTCTTCGCCACCATGGTCGTGGCGG  
34    Nipponbare                     TACGCTCCATCGACCTCCCCATCCTGGGCCCTCTTCGCCACCATGGTCGTGGCGG  
35                                 \*\*\*\*\*  
36    Bala                         TACCTCAAGAACGCCGGCATGTTCAAGGCACCTGGGCCGGTGTGGCGTGGCGAGCCAG  
37    IR64                         TACCTCAAGAACGCCGGCATGTTCAAGGCACCTGGGCCGGTGTGGCGTGGCGAGCCAG  
38    DJ123                         TACCTCAAGAACGCCGGCATGTTCAAGGCACCTGGGCCGGTGTGGCGTGGCGAGCCAG  
39    Nipponbare                     TACCTCAAGAACGCCGGCATGTTCAAGGCACCTGGGCCGGTGTGGCGTGGCGAGCCAG  
40    Azucena                         TACCTCAAGAACGCCGGCATGTTCAAGGCACCTGGGCCGGTGTGGCGTGGCGAGCCAG  
41                                 \*\*\*\*\*  
42    Bala                         GGCGGGCGCGACCTCATGTGCCCGTCTGCGTCACCGCGCTGCCAGCGCGCTTTC  
43    IR64                         GGCGGGCGCGACCTCATGTGCCCGTCTGCGTCACCGCGCTGCCAGCGCGCTTTC  
44    DJ123                         GGCGGGCGCGACCTCATGTGCCCGTCTGCGTCACCGCGCTGCCAGCGCGCTTTC  
45    Nipponbare                     GGCGGGCGCGACCTCATGTGCCCGTCTGCGTCACCGCGCTGCCAGCGCGCTTTC  
46    Azucena                         GGCGGGCGCGACCTCATGTGCCCGTCTGCGTCACCGCGCTGCCAGCGCGCTTTC  
47                                 \*\*\*\*\*  
48    Bala                         ACCAACGACACCTGCTGCCGTCTGCCACCGAGTTCTGCTGAGCTCGCCGCCAGCGC  
49    IR64                         ACCAACGACACCTGCTGCCGTCTGCCACCGAGTTCTGCTGAGCTCGCCGCCAGCGC  
50    DJ123                         ACCAACGACACCTGCTGCCGTCTGCCACCGAGTTCTGCTGAGCTCGCCGCCAGCGC  
51    Nipponbare                     ACCAACGACACCTGCTGCCGTCTGCCACCGAGTTCTGCTGAGCTCGCCGCCAGCGC  
52                                 \*\*\*\*\*  
53  
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60

Bala	AACCTCCGGCGAAGCCGTTCTGCTCGCCTGCCACCAGGCCAACATCGGCTCCAGC
IR64	AACCTCCGGCGAAGCCGTTCTGCTCGCCTGCCACCAGGCCAACATCGGCTCCAGC
DJ123	AACCTCCGGCGAAGCCGTTCTGCTCGCCTGCCACCAGGCCAACATCGGCTCCAGC
Nipponbare	AACCTCCGGCGAAGCCGTTCTGCTCGCCTGCCACCAGGCCAACATCGGCTCCAGC
Azucena	*****
Bala	GCGACGCCATCGCAACCGCAGAACCTGGTATCGCCTTAACAGCAAGATCTCCTT
IR64	GCGACGCCATCGCAACCGCAGAACCTGGTATCGCCTTAACAGCAAGATCTCCTT
DJ123	GCGACGCCATCGCAACCGCAGAACCTGGTATCGCCTTAACAGCAAGATCTCCTT
Nipponbare	GCGACGCCATCGCAACCGCAGAACCTGGTATCGCCTTAACAGCAAGATCTCCTT
Azucena	GCGACGCCATCGCAACCGCAGAACCTGGTATCGCCTTAACAGCAAGATCTCCTT
Bala	*****
IR64	ATTTCCCTCCTCGGCATCCTCCCCGATGCTGCCGGCATGGGATCAAACATGCTC
DJ123	ATTTCCCTCCTCGGCATCCTCCCCGATGCTGCCGGCATGGGATCAAACATGCTC
Nipponbare	ATTTCCCTCCTCGGCATCCTCCCCGATGCTGCCGGCATGGGATCAAACATGCTC
Azucena	ATTTCCCTCCTCGGCATCCTCCCCGATGCTGCCGGCATGGGATCAAACATGCTC
Bala	*****
IR64	ATGCTGCTCTGCATGTACTGGAAGGAGCTGACGGCGGCCCTGCAGCCCCGACGAGGTC
DJ123	ATGCTGCTCTGCATGTACTGGAAGGAGCTGACGGCGGCCCTGCAGCCCCGACGAGGTC
Nipponbare	ATGCTGCTCTGCATGTACTGGAAGGAGCTGACGGCGGCCCTGCAGCCCCGACGAGGTC
Azucena	ATGCTGCTCTGCATGTACTGGAAGGAGCTGACGGCGGCCCTGCAGCCCCGACGAGGTC
Bala	*****
IR64	GCCGCCGCAAGCAGATGGAGGCCATGAGGAGGGCCGCCACGGCGCTCAAACAACAAAC
DJ123	GCCGCCGCAAGCAGATGGAGGCCATGAGGAGGGCCGCCACGGCGCTCAAACAACAAAC
Nipponbare	GCCGCCGCAAGCAGATGGAGGCCATGAGGAGGGCCGCCACGGCGCTCAAACAACAAAC
Azucena	GCCGCCGCAAGCAGATGGAGGCCATGAGGAGGGCCGCCACGGCGCTCAAACAACAAAC
Bala	*****
IR64	AAGAAAGACGGCGACGCCACCCCGATCGCCGGAGGACGACGACGGCGGCGAC
DJ123	AAGAAAGACGGCGACGCCACCCCGATCGCCGGAGGACGACGACGGCGGCGAC
Nipponbare	AAGAAAGACGGCGACGCCACCCCGATCGCCGGAGGACGACGACGGCGGCGAC
Azucena	AAGAAAGACGGCGACGCCACCCCGATCGCCGGAGGACGACGACGGCGGCGAC
Bala	*****
IR64	GCCGAGTCATGATGTCGGAGAACATCTGACGAAGCACCGGTGGTCATGCACTGCTCG
DJ123	GCCGAGTCATGATGTCGGAGAACATCTGACGAAGCACCGGTGGTCATGCACTGCTCG
Nipponbare	GCCGAGTCATGATGTCGGAGAACATCTGACGAAGCACCGGTGGTCATGCACTGCTCG
Azucena	GCCGAGTCATGATGTCGGAGAACATCTGACGAAGCACCGGTGGTCATGCACTGCTCG
Bala	*****
IR64	GAGCACGGCGGAAGCTATTCTCTCAAGTGCTTCGCGTACGTTGGTCACCGCGCATGCTC
DJ123	GAGCACGGCGGAAGCTATTCTCTCAAGTGCTTCGCGTACGTTGGTCACCGCGCATGCTC
Nipponbare	GAGCACGGCGGAAGCTATTCTCTCAAGTGCTTCGCGTACGTTGGTCACCGCGCATGCTC
Azucena	GAGCACGGCGGAAGCTATTCTCTCAAGTGCTTCGCGTACGTTGGTCACCGCGCATGCTC
Bala	*****
IR64	GTCGCCTACATGCTCGGCCTAACATGTCGTGGACCGCCATCACCAACCGCCATCGCCCTC
DJ123	GTCGCCTACATGCTCGGCCTAACATGTCGTGGACCGCCATCACCAACCGCCATCGCCCTC
Nipponbare	GTCGCCTACATGCTCGGCCTAACATGTCGTGGACCGCCATCACCAACCGCCATCGCCCTC
Azucena	GTCGCCTACATGCTCGGCCTAACATGTCGTGGACCGCCATCACCAACCGCCATCGCCCTC

Bala	GTCTGTCGACTTCCCGGACGCCATCCCTGCACAAGGTAAACCATATCCGCCA
IR64	GTCTGTCGACTTCCCGGACGCCATCCCTGCACAAGGTAAACCATATCCGCCA
DJ123	GTCTGTCGACTTCCCGGACGCCATCCCTGCACAAGGTAAACCATATCCGCCA
Nipponbare	GTCTGTCGACTTCCCGGACGCCATCCCTGCACAAGGTAAACCATATCCGCCA
Azucena	GTCTGTCGACTTCCCGGACGCCATCCCTGCACAAGGTAAACCATATCCGCCA
*****	
Bala	AATCACTATAACTCTAGGCCACACGAGCAAGTACACGTGGCGGCCATGCGGACAAGAAC
IR64	AATCACTATAACTCTAGGCCACACGAGCAAGTACACGTGGCGGCCATGCGGACAAGAAC
DJ123	AATCACTATAACTCTAGGCCACACGAGCAAGTACACGTGGCGGCCATGCGGACAAGAAC
Nipponbare	AATCACTATAACTCTAGGCCACACGAGCAAGTACACGTGGCGGCCATGCGGACAAGAAC
Azucena	AATCACTATAACTCTAGGCCACACGAGCAAGTACACGTGGCGGCCATGCGGACAAGAAC
*****	
Bala	AATATGTGAACGTGGGTGCCACGTGTGCGTGGATGTGAATGGATGATGCAGGTGTGTACT
IR64	AATATGTGAACGTGGGTGCCACGTGTGCGTGGATGTGAATGGATGATGCAGGTGTGTACT
DJ123	AATATGTGAACGTGGGTGCCACGTGTGCGTGGATGTGAATGGATGATGCAGGTGTGTACT
Nipponbare	AATATGTGAACGTGGGTGCCACGTGTGCGTGGATGTGAATGGATGATGCAGGTGTGTACT
Azucena	AATATGTGAACGTGGGTGCCACGTGTGCGTGGATGTGAATGGATGATGCAGGTGTGTACT
*****	
Bala	CGCTGCTGGTGTCTCTCGGGGATGTTCTGACGGTGAGCGGATTCAACAAGACGGGGC
IR64	CGCTGCTGGTGTCTCTCGGGGATGTTCTGACGGTGAGCGGATTCAACAAGACGGGGC
DJ123	CGCTGCTGGTGTCTCTCGGGGATGTTCTGACGGTGAGCGGGTTCAACAAGACGGGGC
Nipponbare	CGCTGCTGGTGTCTCTCGGGGATGTTCTGACGGTGAGCGGGTTCAACAAGACGGGGC
Azucena	CGCTGCTGGTGTCTCTCGGGGATGTTCTGACGGTGAGCGGGTTCAACAAGACGGGGC
*****	
Bala	TACCCGGGGCCATCTGGAACGTGATGGGCCCTACTCAAAGATCAACCACGTCACGGGC
IR64	TACCCGGGGCCATCTGGAACGTGATGGGCCCTACTCAAAGATCAACCACGTCACGGGC
DJ123	TACCCGGGGCCATCTGGAACGTGATGGGCCCTACTCAAAGATCAACCACGTCACGGGC
Nipponbare	TACCCGGGGCCATCTGGAACGTGATGGGCCCTACTCAAAGATCAACCACGTCACGGGC
Azucena	TACCCGGGGCCATCTGGAACGTGATGGGCCCTACTCAAAGATCAACCACGTCACGGGC
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Bala	TCACCGTCTCTCCGTACATCATCCTCCTCCAAACCTCGCTCCAACGTCCCCACCG
IR64	TCACCGTCTCTCCGTACATCATCCTCCTCCAAACCTCGCTCCAACGTCCCCACCG
DJ123	TCACCGTCTCTCCGTACATCATCCTCCTCCAAACCTCGCTCCAACGTCCCCACCG
Nipponbare	TCACCGTCTCTCCGTACATCATCCTCCTCCAAACCTCGCTCCAACGTCCCCACCG
Azucena	TCACCGTCTCTCCGTACATCATCCTCCTCCAAACCTCGCTCCAACGTCCCCACCG
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Bala	GTACGTACTTACTCACAACCTTCTCGCACCATGCTATTGCTAGTACCCAATGTGTCCC
IR64	GTACGTACTTACTCACAACCTTCTCGCACCATGCTATTGCTAGTACCCAATGTGTCCC
DJ123	GTACGTACTTACTCACAACCTTCTCGCACCATGCTATTGCTAGTACCCAATGTGTCCC
Nipponbare	GTACGTACTTACTCACAACCTTCTCGCACCATGCTATTGCTAGTACCCAATGTGTCCC
Azucena	GTACGTACTTACTCACAACCTTCTCGCACCATGCTATTGCTAGTACCCAATGTGTCCC
*****	
Bala	ACTCACCCAGCACTAACATTGTACTTACAGTACATCAGTTCTTTAATACATGATAT
IR64	ACTCACCCAGCACTAACATTGTACTTACAGTACATCAGTTCTTTAATACATGATAT
DJ123	ACTCACCCAGCACTAACATTGTACTTACAGTACATCAGTTCTTTAATACATGATAT
Nipponbare	ACTCACCCAGCACTAACATTGTACTTACAGTACATCAGTTCTTTAATACATGATAT
Azucena	ACTCACCCAGCACTAACATTGTACTTACAGTACATCAGTTCTTTAATACATGATAT
*****	
Bala	CTGACTTTTTTATCGATGAATGGATCTTATTCAATTCTTTGTAATATAAAAAGAT
IR64	CTGACTTTTTTATCGATGAATGGATCTTATTCAATTCTTTGTAATATAAAAAGAT
DJ123	CTGACTTTTTTATCGATGAATGGATCTTATTCAATTCTTTGTAATATAAAAAGAT
Nipponbare	CTGACTTTTTTATCGATGAATGGATCTTATTCAATTCTTTGTAATATAAAAAGAT
Azucena	CTGACTTTTTTATCGATGAATGGATCTTATTCAATTCTTTGTAATATAAAAAGAT
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 3 Bala TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATTTTTAATAAGAT  
 4 IR64 TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATTTTTAATAAGAT  
 5 DJ123 TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATTTTTAATAAGAT  
 6 Nipponbare TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATTTTTAATAAGAT  
 7 Azucena TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATTTTTAATAAGAT  
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 9 Bala GAATGATTATTAAGAAAAAAATTTTTGTCTCTCATATTAAAAAGAACAACTCA  
 10 IR64 GAATGATTATTAAGAAAAAAATTTTTGTCTCTCATATTAAAAAGAACAACTCA  
 11 DJ123 GAATGATTATTAAGAAAAAAATTTTTGTCTCTCATATTAAAAAGAACAACTCA  
 12 Nipponbare GAATGATTATTAAGAAAAAAATTTTTGTCTCTCATATTAAAAAGAACAACTCA  
 13 Azucena GAATGATTATTAAGAAAAAAATTTTTGTCTCTCATATTAAAAAGAACAACTCA  
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 15 Bala TTTTTACATGTTATTTGAGTGGATTGTAATTGTAATTATATGTTGGGTGAGTGTT  
 16 IR64 TTTTTACATGTTATTTGAGTGGATTGTAATTGTAATTATATGTTGGGTGAGTGTT  
 17 DJ123 TTTTTACATGTTATTTGAGTGGATTGTAATTGTAATTATATGTTGGGTGAGTGTT  
 18 Nipponbare TTTTTACATGTTATTTGAGTGGATTGTAATTGTAATTATATGTTGGGTGAGTGTT  
 19 Azucena TTTTTACATGTTATTTGAGTGGATTGTAATTGTAATTATATGTTGGGTGAGTGTT  
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 21 Bala GTTGATGGGGATGAGGTGGCGCGCGCGCGACGATATCACGGCGCGGTGACGCG  
 22 IR64 GTTGATGGGGATGAGGTGGCGCGCGCGCGACGATATCACGGCGCGGTGACGCG  
 23 DJ123 GTTGATGGGGATGAGGTGGCGCGCGCGCGACGATATCACGGCGCGGTGACGCG  
 24 Nipponbare GTTGATGGGGATGAGGTGGCGCGCGCGCGACGATATCACGGCGCGGTGACGCG  
 25 Azucena GTTGATGGGGATGAGGTGGCGCGCGCGCGACGATATCACGGCGCGGTGACGCG  
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 27 Bala GTCGTGGCTGCTGGCGTGGGTGAGCACGGTGGCGGGAACCTGTCGCTGGGTG  
 28 IR64 GTCGTGGCTGCTGGCGTGGGTGAGCACGGTGGCGGGAACCTGTCGCTGGGTG  
 29 DJ123 GTCGTGGCTGCTGGCGTGGGTGAGCACGGTGGCGGGAACCTGTCGCTGGGTG  
 30 Nipponbare GTCGTGGCTGCTGGCGTGGGTGAGCACGGTGGCGGGAACCTGTCGCTGGGTG  
 31 Azucena GTCGTGGCTGCTGGCGTGGGTGAGCACGGTGGCGGGAACCTGTCGCTGGGTG  
 32 \*  
 33 Bala GGCGGCGAACCTGATCGTGTGCGAGCGAGGCGAGGGCGACGCCAACGCC  
 34 IR64 GGCGGCGAACCTGATCGTGTGCGAGCGAGGCGAGGGCGACGCCAACGCC  
 35 DJ123 GGCGGCGAACCTGATCGTGTGCGAGCGAGGCGAGGGCGACGCCAACGCC  
 36 Nipponbare GGCGGCGAACCTGATCGTGTGCGAGCGAGGCGAGGGCGACGCCAACGCC  
 37 Azucena GGCGGCGAACCTGATCGTGTGCGAGCGAGGCGAGGGCGACGCCAACGCC  
 38 \*  
 39 Bala CACCTCTGGAACCACTGATCTCGGACTCCCATTCCACCCCTCGTCGTCACCGCC  
 40 IR64 CACCTCTGGAACCACTGATCTCGGACTCCCATTCCACCCCTCGTCGTCACCGCC  
 41 DJ123 CACCTCTGGAACCACTGATCTCGGACTCCCATTCCACCCCTCGTCGTCACCGCC  
 42 Nipponbare CACCTCTGGAACCACTGATCTCGGACTCCCATTCCACCCCTCGTCGTCACCGCC  
 43 Azucena CACCTCTGGAACCACTGATCTCGGACTCCCATTCCACCCCTCGTCGTCACCGCC  
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 45 Bala CATCCCTCTCATCGCAAGATCAACATCTAGCTGTCATTAAATTAAATTAAATATAAT  
 46 IR64 CATCCCTCTCATCGCAAGATCAACATCTAGCTGTCATTAAATTAAATTAAATATAAT  
 47 DJ123 CATCCCTCTCATCGCAAGATCAACATCTAGCTGTCATTAAATTAAATTAAATATAAT  
 48 Nipponbare CATCCCTCTCATCGCAAGATCAACATCTAGCTGTCATTAAATTAAATTAAATATAAT  
 49 Azucena CATCCCTCTCATCGCAAGATCAACATCTAGCTGTCATTAAATTAAATTAAATATAAT  
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 51 Bala CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT  
 52 IR64 CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT  
 53 DJ123 CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT  
 54 Nipponbare CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT  
 55 Azucena CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT  
 56 \*  
 57 Bala CATGCAAATTACAAGGGATGTGATATGATCATATTAAATCCAAGGAAGCTAGTATGATGAT  
 58 IR64 CATGCAAATTACAAGGGATGTGATATGATCATATTAAATCCAAGGAAGCTAGTATGATGAT  
 59 DJ123 CATGCAAATTACAAGGGATGTGATATGATCATATTAAATCCAAGGAAGCTAGTATGATGAT  
 60 Nipponbare CATGCAAATTACAAGGGATGTGATATGATCATATTAAATCCAAGGAAGCTAGTATGATGAT  
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4   Bala          TAAGTTGTGATCAAGCCGTACTGTGTGCATGCCAATGCATGCATGTGTTCAAGTGTACA
5   IR64          TAAGTTGTGATCAAGCCGTACTATGTGCATGCCAATGCATGCATGTGTTCAAGTGTACA
6   DJ123         TAAGTTGTGATCAAGCCGTACTATGTGCATGCCAATGCATGCATGTGTTCAAGTGTACA
7   Nipponbare    TAAGTTGTGATCAAGCCGTACTATGTGCATGCCAATGCATGCATGTGTTCAAGTGTACA
8   Azucena        TAAGTTGTGATCAAGCCGTACTATGTGCATGCCAATGCATGCATGTGTTCAAGTGTACA
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Bala          CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCATGCTCTGGGATAATTAGTTAA
IR64          CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCATGCTCTGGGATAATTAGTTAA
DJ123         CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCATGCTCTGGGATAATTAGTTAA
Nipponbare    CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCATGCTCTGGGATAATTAGTTAA
Azucena        CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCATGCTCTGGGATAATTAGTTAA
Bala          TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAGGCA
IR64          TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAGGCA
DJ123         TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAGGCA
Nipponbare    TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAGGCA
Azucena        TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAGGCA
Bala          AATGAACAGCAGAACATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT
IR64          AATGAACAGCAGAACATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT
DJ123         AATGAACAGCAGAACATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT
Nipponbare    AATGAACAGCAGAACATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT
Azucena        AATGAACAGCAGAACATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT
Bala          TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC
IR64          TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC
DJ123         TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC
Nipponbare    TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC
Azucena        TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC
Bala          ATGCCGATTACCTATTGCTCCAAACTTGCATTCAGTTGAACCTTTAATAAAAGTTAA
IR64          ATGCCGATTACCTATTGCTCCAAACTTGCATTCAGTTGAACCTTTAATAAAAGTTAA
DJ123         ATGCCGATTACCTATTGCTCCAAACTTGCATTCAGTTGAACCTTTAATAAAAGTTAA
Nipponbare    ATGCCGATTACCTATTGCTCCAAACTTGCATTCAGTTGAACCTTTAATAAAAGTTAA
Azucena        ATGCCGATTACCTATTGCTCCAAACTTGCATTCAGTTGAACCTTTAATAAAAGTTAA
Bala          TTTGGCTTGTGCAATTGCTCAATTGCTCCCTCTTGTGTTAACCGATCTTGCCTAGTGCG
IR64          TTTGGCTTGTGCAATTGCTCAATTGCTCCCTCTTGTGTTAACCGATCTTGCCTAGTGCG
DJ123         TTTGGCTTGTGCAATTGCTCAATTGCTCCCTCTTGTGTTAACCGATCTTGCCTAGTGCG
Nipponbare    TTTGGCTTGTGCAATTGCTCCCTCTTGTGTTAACCGATCTTGCCTAGTGCG
Azucena        TTTGGCTTGTGCAATTGCTCCCTCTTGTGTTAACCGATCTTGCCTAGTGCG
Bala          TACGTAGCTAGATTGCACATTAAAG
IR64          TACGTAGCTAGATTGCACATTAAAG
DJ123         TACGTAGCTAGATTGCACATTAAAG
Nipponbare    TACGTAGCTAGATTGCACATTAAAG
Azucena        TACGTAGCTAGATTGCACATTAAAG

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**Supplementary figure 4.** Genomic sequence alignment of *Lsi3*. Regions highlighted in bold are predicted exon regions based on the Rice Genome Annotation Project – release 7 sequence available at <http://rice.plantbiology.msu.edu/>.