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Genotypic differences in shoot silicon content and the impact on grain arsenic accumulation in rice

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1 Title Page:
2 Genotypic differences in shoot silicon content and the impact on grain arsenic accumulation
3 in rice
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17 Figures:6 figures in the main text and 4 supplementary figures
18 Short running tile: Genotypic differences of shoot silicon in rice.

35 Genotypic differences in shoot silicon content and the impact on grain arsenic accumulation
36 in rice

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46 **Abstract:**

47 Silicon in rice has been demonstrated to be involved in resistance to lodging, tolerance to
48 both drought and salinity, and also enhances resistance to pests and diseases. The aim of this
49 study was to determine the range of silicon content in a set of rice (*Oryza sativa* L.)
50 accessions, and to determine if the natural variation of shoot silicon is linked to the
51 previously identified silicon transporters (*Lsi* genes). Silicon content was determined in 50
52 field-grown accessions, representing all sub-populations of rice, with all accessions being
53 genotyped with 700K SNPs. SNPs within 10 kb of the *Lsi* genes were examined to determine
54 if any were significantly linked with the phenotypic variation. An XRF method of silicon
55 determination compared favourably with digestion and colorimetric analysis. There were
56 significant genotypic differences in shoot silicon ranging from 16.5 – 42.4 mg g⁻¹ of plant dry
57 weight, there was no significant difference between the rice sub-populations. Plants with
58 different alleles for SNPs representing *Lsi2* and *Lsi3* were significantly different for shoot
59 silicon content. Shoot silicon correlated negatively with grain arsenic in the *tropical* and
60 *temperate japonica* sub-population, suggesting that accessions with high shoot silicon have

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61 reduced grain arsenic. This study indicates that alleles for *Lsi* genes are excellent candidate
62 genes for further study to explain the natural variation of shoot silicon in rice.

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64 **Key words:** arsenic, natural variation, rice, silicon, XRF

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1 Introduction

Global rice (*Oryza sativa* L.) production needs to increase continuously to ensure the world's food security (Hibberd et al., 2008). As a beneficial element, silicon alleviates biotic and abiotic stresses in rice which helps to maintain yield (Ma and Takahashi, 2002; Detmann et al., 2012; Meharg and Meharg, 2015). Silicon is mainly available as monosilicic acid ranging from 0.1 to 0.6 mM in the soil solution (Epstein, 1994; Ma and Takahashi, 2002). Previous studies have demonstrated that monosilicic acid is taken up by rice roots as an undissociated molecule and translocated into the shoots through the transpiration stream (Takahashi and Hino, 1978; Mitani-Ueno et al., 2005). It then polymerises on the surface of cells in the shoot in the form of a silica-cellulose double layer and silica-cuticle double layer. This silica-base layer improves resistance to lodging, salinity tolerance, drought tolerance, and enhances resistance to pests and diseases (Takahashi and Hino, 1978; Mitani-Ueno et al., 2005, Chen et al., 2011; Han et al., 2015).

Genetically rice can be classified into two major sub-species, *Japonica* and *Indica* (Chang, 2003) and these have been further classified into 5 sub-populations; *indica*, *aus*, (both *Indica* sub-species) *tropical japonica*, *temperate japonica*, and *aromatic* (all three *Japonica* sub-species) (Garris et al., 2005; Zhao et al., 2011). Several previous studies indicate that there are differences in shoot silicon content between the *Indica* and *Japonica* sub-species of rice. Deren et al., (1992) showed that *Japonica* sub-species usually have a higher silicon content than *Indica* rice varieties, based on screening 10 accessions in the greenhouse and 18 under field conditions. A study conducted by Winslow (1992) revealed that African upland *Japonica* rice accessions had 50 to 100% higher silicon content in mature flag leaves than Asian upland *Indica* accessions. In addition to the differences at the subspecies level several studies have looked at genotypic differences in silicon content, showing ranges of 41 to 60 mg g⁻¹ (Deren, 2001) and 28 to 61 mg g⁻¹ (Norton et al., 2010a). Ma et al., (2007a) also

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91 observed that silicon uptake by the root and the concentration silicon present in the shoot are
92 both higher in *Japonica* than *Indica* rice accessions, which they attributed to differences in
93 the expression of silicon transporter genes.

94 Two types of silicon transporters have been identified in rice to date. A gene
95 (LOC_Os02g51110) identified for silicic acid influx in rice is classified as an aquaporin
96 (Low silicon 1 or *Lsi1*) which is a member of the nodulin 26-like intrinsic protein (OsNIP2;
97 1) group of aquaporins (*Ma et al.*, 2006; *Ma et al.*, 2008). A homologue of *Lsi1*, known as
98 *Lsi6* (LOC_Os06g12310; OsNIP2; 2), responsible for shoot and husk silicon distribution in
99 rice is also classified as an aquaporin (*Yamaji et al.*, 2008). The efflux of silicic acid through
100 the plasma membrane protein known as low silicon 2 (*Lsi2*; LOC_Os03g01700) is an energy
101 dependent process in rice (*Ma et al.*, 2007b). A homologue of *Lsi2*, known as *Lsi3*
102 (LOC_Os10g39980), is also an energy dependent active transporter involved in regulating
103 shoot silicon accumulation in rice (*Yamaji et al.*, 2015).

104 It has been shown that arsenic, classified as a class one carcinogen, can be transported
105 through silicon transporters in rice (*Ma et al.*, 2008; *Zhao et al.*, 2010; *Mitani-Ueno et al.*,
106 2011). There are two different forms of arsenic present in rice: organic arsenic and inorganic
107 arsenic (*Williams et al.*, 2005). Organic arsenic is found in rice in two main types of
108 molecular species dimethylarsinic acid (DMA), and monomethylarsonic acid (MMA) as well
109 as tetramethylarsonium (*Williams et al.*, 2005; *Hansen et al.*, 2011). Inorganic arsenic is
110 found in rice as two molecular species; arsenate and arsenite (*Abedin et al.*, 2002; *Williams et*
111 *al.*, 2005). Arsenate is an analogue of phosphate and is taken up via phosphate transporters
112 while arsenite is taken up by silicic acid transporters in rice (*Abedin et al.*, 2002; *Ma et al.*,
113 2008). It has been shown that the silicon transporters *Lsi1*, *Lsi2* and *Lsi6* are also arsenic
114 transporters, using a combination of mutants and transgenic lines (*Ma et al.*, 2008; *Zhao et*
115 *al.*, 2010; *Mitani-Ueno et al.*, 2011). Several studies indicate that anaerobic rice cultivation

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3 116 leads to increased mobilisation of soil arsenic in the form of arsenite, which causes
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5 117 anaerobically-grown rice to accumulate more arsenic through silicon transporters (*Ma et al.*,
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7 118 2008; *Xu et al.*, 2008; *Carey et al.*, 2010). Silicon fertilisation has also been shown to
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9 119 decrease shoot and grain arsenic indicating that silicon could play an important role in
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11 120 decreasing total arsenic uptake in rice (*Li et al.*, 2009; *Seyfferth and Ferdorf*, 2012).
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14 121 This study was designed to address four questions all related to the process of silicon and
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16 122 arsenic accumulation in rice: How does the cultivation method affect silicon distribution in
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18 123 different organs of rice plants? Are there significant genotypic differences in shoot silicon
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20 124 concentration across a diverse panel of rice related to the 5 different sub-populations of rice?
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22 125 Is there a relationship between natural variation in shoot silicon and arsenic content in rice?
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24 126 Can natural variation in shoot silicon be linked to known silicon transporters in rice? The
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26 127 results provide a deeper understanding of the natural variation in silicon content across rice
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29 128 accessions and its relationship to arsenic accumulation in rice grains.
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2 Materials and Methods

2.1 Silicon content in different organs of rice (*Oryza sativa* L.) grown in flooded and non-flooded conditions

An experiment was conducted in a greenhouse at the University of Aberdeen, UK under both flooded and non-flooded conditions with four replicates for each treatment. One litre plastic pots were filled with soil (~530 g soil described in *Norton et al.*, 2013). For the flooded condition, a plastic liner was used to line the pots and hold the water within the pot whereas the non-flooded pots were kept without a liner to allow drainage of water through the pot. Five Italic Carolina (*temperate japonica*) seeds were sown in each pot, then thinned to one plant in each pot after two weeks. To maintain the flooded condition, tap water from the greenhouse was used to flood the pots to 2 cm above the soil surface when plants were 3 weeks old. Every two weeks during the first four weeks of growth 100 mL of half strength Yoshida’s nutrient solution was added (*Yoshida et al.*, 1976). The dose of Yoshida’s nutrient solution was increased up to 100 mL of full strength solution every week after four weeks and continued until the filled grains had matured when samples were harvested.

At harvest, samples were collected from different parts of the mature plants: root, 3rd sheath, 3rd node, 3rd leaf, 2nd sheath, 2nd node, 2nd leaf, flag sheath, 1st node, flag leaf, husk and unpolished grains. The sheath, node and leaves were taken from the main tiller, with the most recent leaf prior to the flag leaf designated 2nd leaf. Root samples were washed thoroughly with tap water followed by deionised water and confirmed to be free of soil particles by examining the roots under a microscope (Leica MZ8, 10445932, 16×/14B, PLAN 1.0X). Samples were dried at 80°C for 5 days until a constant weight was achieved. All samples were mixed and subsampled prior to being ball-milled. The silicon content was determined by Flow Injection Analyser (FIA) after alkali digestion.

2.2 Genotypic differences in shoot silicon content of rice

Seeds were obtained from the Rice Diversity Panel 1 (RDP1) (<http://ricediversity.org/>) (Ali et al., 2011; Eizenga et al., 2014). The classification of Zhao et al., (2011) was used for the sub-population classification of rice accessions. In 2009 a total of 312 accessions were cultivated at the experimental site in Bangladesh. Seedlings were transplanted by hand in a single 2m row of 10 hills, each hill (one seedling) 20 cm apart and each row 20 cm apart in a randomised complete block design with four replicates of each accession. The experimental site was kept flooded until the grains were filled. Plant material from the central six plants was harvested and used for chemical analysis. Detailed information about the experimental site and experimental methods are described in Norton et al., (2012). For shoot silicon analysis, fifty accessions (10 accessions from each rice sub-population) were randomly selected based on the initial sub-population assignment using single sequence repeat (SSR) markers (Ali et al., 2011) (Supplementary Table 1). Subsequently, after selection and silicon analysis, these accessions have been assigned to sub-populations based on the 700K SNP data (McCouch et al., 2016), these sub-population assignments are used for classification of the accessions in this study.

2.3 Analysis of rice shoot silicon by FIA

Plant material and certified reference material (CRM) were prepared for silicon analysis as described by Carneiro et al., (2007). A total of 1.5 g shoot material from each sample was sub-sampled at random and powderised using a ball mill (Retsch, MM200, Germany). From the powderised plant material, a sub-sample of 20 mg was weighed into a 50 mL polyethylene centrifuge tube (CORNING®, NY). To digest the sample, 0.6 mL of hydrogen peroxide (H₂O₂, > 30% W/V, Fisher Scientific) and 1.5 mL of sodium hydroxide (NaOH, solutions 50%, Fluka) were added and the samples were then vortexed (mixed using a vortex mixer). The samples were heated for 1 hour at 90°C in a water bath, then vortexed again and

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left overnight. The tubes were vortexed again after overnight extraction, then heated at 123°C under a pressure of 0.15 MPa for 1 h. Samples were kept at room temperature for 2 h then vortexed, followed by addition of 18.5 mL of ddH₂O. Prior to analysis, samples were diluted 1:5 with Milli-Q water. Silicon content was measured using an FIA spectrophotometer (Tecator FIAstar 5010) a wavelength of 410 nm (Carneiro et al., 2007; Norton et al., 2010a; Norton et al., 2010b).

2.4 Analysis of rice shoot silicon by P-XRF

A total of 1.5 g of dried shoot material for each rice accession was sub-sampled at random and powdered using a ball mill (Retsch, MM200, Germany). To perform the analysis of shoot silicon by P-XRF, 19 accessions were selected at random from the 50 accessions for which shoot silicon had been determined by FIA. For P-XRF analysis, 0.7 g of homogeneous powder sample was compressed into 13 mm diameter pellets using a manual hydraulic press with a 13mm die at a pressure of 10 tons (Specac, Orpington, United Kingdom). Shoot silicon content (% of silicon dry shoot weight) was measured using a commercial P-XRF instrument (Niton XL3t900 GOLDD analyzer: Thermo Scientific Winchester, UK), calibrated using Si-spiked synthetic methyl cellulose and validated using Certified Reference Materials of NCS DC73349 ‘Bush branches and leaves’ obtained from the China National Analysis Center for Iron and Steel, as described in Reidinger et al., (2012). The mean value of samples for each accession was used for correlation analysis between P-XRF and FIA measurements.

2.5 Relationship between silicon and arsenic content in rice

The plant material used in this study was previously examined for grain arsenic content (Norton et al., 2012) which provided an opportunity to examine the relationship between shoot silicon and grain arsenic in rice. The relationship between shoot silicon (log

transformed) and grain arsenic (log transformed) was investigated for the 50 rice accessions based on accession means.

2.6 Single Marker Analysis

The accessions used in this study have been genotyped using a high-density SNP chip (McCouch et al., 2016). SNPs for the accessions were extracted using PLINK (Purcell et al., 2007). SNPs were extracted from 10 kb upstream of the start codon to 10 kb downstream of the stop codon of the *Lsi1*, *Lsi2*, *Lsi3*, and *Lsi6* loci. SNPs were excluded from the analysis if they were invariant or if minor alleles were present in less than 5% of the accessions. The RDP1 population has a high degree of stratification by rice sub-population (Zhao et al., 2011; McCouch et al., 2016). To overcome this stratification, sub-population assignment was used (based on the 700 K SNP data; McCouch et al., 2016) as a factor in a two-way ANOVA, with SNP base call as the other factor. The two-way ANOVA was used to determine if the phenotype for the accession was significantly different for each SNP tested.

2.7 Sequence alignments

Based on the result achieved from the single-marker analysis the sequence diversity of *Lsi2* and *Lsi3* were investigated for 5 cultivars using BAM files produced after aligning sequence reads against Nipponbare version 7 reference genome. The genome sequences of the cultivars used in this study have been previously published (Kawahara et al., 2013; Cardoso et al., 2014; Schatz et al., 2014). The accessions were from the following sub-populations; 2 *indica* accessions (IR64 and Bala), 1 *aus* accession (DJ123) and 2 *tropical japonica* accessions (Azucena and Nipponbare). The genomic DNA sequence was visualised using the IGV (<https://www.broadinstitute.org/igv/>) to identify the difference of genomic DNA sequence within *Lsi2* and *Lsi3* in 5 cultivars (Thorvaldsdóttir et al., 2013; Robinson et al., 2011). Using

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224 Clustal Omega the DNA sequences of 5 cultivars were aligned for *Lsi2* and *Lsi3* separately
225 and showed in supplementary figure 3 and 4 respectively (Sievers *et al.*, 2011).

226 **2.8 Statistical analysis**

227 Statistical significance was set at $P < 0.05$ for all analyses, which were performed using
228 Minitab 16. The normality of distribution and homogeneity of variance of the data were
229 tested prior to one or two-way analysis of variance (ANOVA), as appropriate. Pearson
230 correlation analysis was used to investigate the relationship between measurements of shoot
231 silicon and grain arsenic.

232 **3 Results**

233 **3.1 Shoot silicon content in different organs of rice plants**

234 Flooding increased plant silicon content in the flag sheath, 1st node, flag leaf and husk
235 compared to plants grown under non-flooded conditions (Fig. 1). The content of silicon in
236 grain and root tissues were significantly lower than any in other organs of plants grown under
237 either condition. There was a significant difference ($P < 0.001$, $F = 27.40$, $R^2 = 78.20\%$) of
238 silicon content between different organs of the plant in non-flooded conditions: The highest
239 mean content was in the husks (46.8 mg g^{-1}), while the lowest was in the grain (3.5 mg g^{-1}).
240 For plants grown under flooded conditions: The highest silicon content was observed in the
241 flag leaf (67.3 mg g^{-1}) and the lowest was in the grains (4.4 mg g^{-1}).

242 **3.2 Genotypic difference in shoot silicon content of rice**

243 Fifty diverse rice accessions were examined by FIA to determine the difference in shoot
244 silicon content of rice. There was a significant genotypic difference in shoot silicon content
245 among the 50 accessions, where genotype explained 55% of the variation ($P < 0.001$; $F =$
246 5.80 ; $R^2 = 55.30\%$; $df = 49$). The mean shoot silicon content of the 50 accessions was 28.1
247 mg g^{-1} , and the lowest mean shoot silicon was observed in Dhala Shita (16.5 mg g^{-1}) The
248 highest mean shoot silicon was observed in Bala (42.4 mg g^{-1}) (Fig. 2). There was no
249 significant difference for shoot silicon content of the 5-major rice sub-populations (Fig. 3).

250 Nineteen rice accessions were selected at random from the 50 accessions analysed by FIA,
251 for measurement of shoot silicon content by P-XRF. The silicon content of four individual
252 field grown replicates of each accession were measured separately by P-XRF and FIA and the
253 mean value of each accession was used for correlation analysis. Using both methods,
254 genotypic differences were observed between the accessions ($P < 0.001$; $F = 9.90$; $df = 18$ for
255 P-XRF; $P < 0.001$; $F = 7.30$; $df = 18$ for FIA). Correlation analysis indicated that there was a

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3 256 significant and large positive correlation between the two methods ($r = 0.95$; $P < 0.001$; $df =$
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5 257 18) (Fig 4).

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8 258 **3.3 Correlation between shoot silicon and grain arsenic in rice**

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10 259 No significant correlation was observed between mean shoot silicon and mean shoot arsenic
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12 260 for all of the 50 accessions (supplementary Figure 1), or for within each of the 5 sub-
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14 261 populations. There was a weak negative correlation ($r = -0.31$; $P = 0.028$; $df = 49$)
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16 262 (supplementary Figure 2) between shoot silicon and grain arsenic content for all 50
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18 263 accessions. When correlation analysis was conducted separately for shoot silicon and grain
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20 264 arsenic on each of the sub-populations, significant negative correlations were found for the
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22 265 *temperate japonica* ($r = -0.78$; $P = 0.007$; $df = 9$) and *tropical japonica* ($r = -0.84$; $P = 0.002$;
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24 266 $df = 9$) accessions (Fig. 5). No significant correlations were observed for the other 3 major
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26 267 rice sub-populations (*indica*, *aus* and *aromatic*).

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30 268 **3.4 Testing accessions with different alleles of SNPs around and within *Lsi* genes for**
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32 269 **variation in shoot silicon concentration**

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35 270 A total of 10 SNPs from the SNP database are within 10 kb upstream and downstream of the
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37 271 *Lsi2* gene (selected SNPs for the rice accessions are presented in supplementary Table 2).
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39 272 Shoot silicon concentration for accessions with the different alleles for two of these SNPs
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41 273 was significantly different. SNP-3.434426 is located 2551 bp before the start codon, and
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43 274 revealed a significant difference between the C and T polymorphism ($P = 0.006$), where the
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45 275 mean silicon content of accessions with the C allele was 29.3 mg g^{-1} while the mean silicon
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47 276 content of the accessions with the T allele was 23.1 mg g^{-1} . SNP-3.438416 is located 6541 bp
48
49 277 before the start codon and revealed a significant difference between the A and C
50
51 278 polymorphism ($P = 0.008$), where the mean silicon content of the accessions with the A allele
52
53 279 was 29.6 mg g^{-1} while the silicon content of the accessions with the G allele had a mean of

23.1 mg g⁻¹. Both SNPs group the accessions in a similar way, the only difference was more missing SNP information for SNP-3.438416 (Fig. 6).

A total of 20 SNPs from the SNP database are within 10 kb upstream and downstream of the *Lsi3* gene (selected SNPs for the rice accessions are presented in supplementary Table 3). Shoot silicon concentration for accessions with the different alleles for one of these SNPs was significantly different. SNP- 10.21340470 is located 5299 bp prior to the start codon, and revealed a significant difference between the G and A polymorphism ($P = 0.016$), where the mean silicon content of accessions with the G allele was 28.4 mg g⁻¹ while the mean silicon content of the accessions with the A allele was 35.6 mg g⁻¹ (Fig. 6).

There were 20 SNPs and 19 SNPs observed within 10 kb of *Lsi1* and *Lsi6* respectively. However, at each of these SNPs the different alleles were not significantly different for shoot silicon concentration.

To explore further, the sequence alignments of *Lsi2* and *Lsi3* were performed using available high-quality genome sequences. The accessions used were Nipponbare, Azucena, IR64, Bala, and DJ123 which are from the *tropical japonica*, *tropical japonica*, *indica*, *indica* and *aus* rice subgroups respectively. From the sequence analyses of *Lsi2* and *Lsi3* a number of polymorphisms within the genes were identified. For *Lsi2*, there was a synonymous SNP substitution within the first exon, where DJ123 has “C” allele while the other 4 accessions have “T” allele (Supplementary Figure 3). For *Lsi3*, 4 SNPs were detected in exons and 6 SNPs in introns (Supplementary Figure 4). There was only one non-synonymous SNP observed in the first exon of *Lsi3* where DJ123 and Bala have “T” allele and other accessions have “A” allele. The available 3000 rice genome sequence data indicates that this polymorphism between “A” and “T” in *Lsi3* is associated with the *aus* sub-population in rice where 15 accessions have “A” allele and 184 accessions have “T” allele (Alexandrov et al.,

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304 2015). This non-synonymous polymorphism between “A” and “T” in *Lsi3* with the “T” allele
305 is very rarely observed in *indica* and *japonica* subpopulations of rice in 3000 rice genome
306 sequence data (Alexandrov et al., 2015).
307

308 4 Discussion

309 In this study, genotypic differences in shoot silicon content were identified from field grown
310 rice cultivars. However, no differences in shoot silicon were observed across the 5 different
311 sub-populations of rice. Additionally, SNPs detected in the accessions were significantly
312 linked to known silicon transporter genes in rice, which indicates that these genes are
313 potentially involved in natural variation of silicon accumulation in rice.

314 Flooded conditions increased silicon content in the upper part of the plant (flag sheath, 1st
315 node, flag leaf and husk) compared to the non-flooded conditions, which suggests that the
316 uptake or translocation of shoot silicon into these plant organs might be controlled by
317 different processes (compared to those determining silicon uptake in other tissues) which
318 differ between aerobic and anaerobic conditions. It has been shown that silicon dissolution
319 and bio-availability plays a significant role in the variation of silicon content in grasses
320 (Quigley et al., 2017). Therefore, the difference in dissolved silicon in flooded and non-
321 flooded conditions might affect the accumulation of silicon in the rice plants used in this
322 study. It was also notable that there was no significant difference in silicon content in
323 different tissues between the internodes (e.g. flag leaf, 2nd leaf) in non-flooded conditions but
324 there was a significant difference between the silicon content of internodes under flooded
325 conditions (Fig. 1). Previous studies have shown that transpiration is one of the most
326 important factors responsible for higher *silicification* in plants and that transpirational flow is
327 higher in anaerobic conditions than in aerobic ones (Mitani-Ueno et al., 2005; Kato and
328 Okami, 2011; Kumar et al., 2017; McLarnon et al., 2017). Therefore, one potential
329 explanation for increased silicon accumulation in the upper organs or developing organs of
330 rice plants (e.g. flag sheath, 1st node, flag leaf and husk) grown in flooded soils is a higher
331 transpirational flow in these plants. Importantly, the data presented in figure 1 shows that
332 tissue silicon content is reasonably evenly distributed across tissues with only that from

333 flooded plants in tissue associated with flowering and seed production significantly higher
334 than the rest. Since this reproductive tissue was removed from the field samples used in this
335 study we can be reassured that a mean value obtained from straw will be a good estimate of
336 the tissue concentration of the majority of rice plant.

337 Fifty accessions from 5 different sub-populations (10 accessions from each sub-population)
338 were selected at random to examine the difference of shoot silicon content in rice and this
339 revealed highly significant differences of shoot silicon content. A genotypic difference in
340 shoot silicon content across a wide group of accessions has been observed previously (*Deren*,
341 2001; *Norton et al.*, 2010a). The 2.6-fold difference of shoot silicon content in this study is
342 similar to the previous 2.2 fold range detected for genotypic differences of shoot silicon
343 content in rice (*Norton et al.*, 2010a). However, the maximum value observed in our study is
344 slightly lower than that detected previously (42.4 mg g⁻¹ in this study, 60 mg g⁻¹ (*Deren*,
345 2001), 61 mg g⁻¹ (*Norton et al.*, 2010a).

346 The plant material used for determination of shoot silicon content in the 50 rice accessions
347 was grown in flooded, irrigated conditions (*Norton et al.*, 2012). Previous studies estimated
348 that 27% - 44% of the silicon taken up by rice plants is supplied by irrigation, while the
349 remaining percentage must be supplied by soil constituents (*Desplanques et al.*, 2006;
350 *Klotzbücher et al.*, 2015). All the accessions tested in this study had a silicon content below
351 50 mg g⁻¹ which, according to *Dobermann and Fairhurst* (2000), is below the critical level of
352 mineral deficiency for rice production. The low shoot silicon content (16.5 mg g⁻¹ to 42.4 mg
353 g⁻¹) observed in this study may be due to removal of rice straw from the paddy field, which is
354 common practice in Bangladesh, and has been shown to contribute to lower shoot silicon in
355 the subsequent rice crop (*Seyfferth et al.*, 2013). Future work should focus on linking the
356 flooded and non-flooded pot based experiment and the removal of straw at the field scale to

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2
3 357 establish the importance of water management and field management on silicon accumulation
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5 358 in field grown rice.
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7 359 Several studies have demonstrated that the *Japonica* sub-species of rice have higher shoot
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9 360 silicon than *Indicas* (Winslow, 1992; Winslow et al., 1997; Ma et al., 2007a). These studies
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11 361 may have been limited by the number of accessions that were screened. For example, Ma et
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13 362 al., (2007b) only screened two rice accessions to examine the genotypic difference in silicon
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15 363 uptake of rice. To improve the current understanding of silicon biology in rice, we
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17 364 investigated field grown shoot samples of 50 rice accessions across 5 sub-populations. Within
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19 365 the material tested in this study the data suggests that the natural variation observed in shoot
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21 366 silicon is not governed by genetic differences between rice sub-populations, but rather is
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23 367 largely due to the genetic differences within individual sub-groups.
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26 368 Data on more than 50 accessions would have opened the opportunity to conduct genome-
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28 369 wide association (GWA) mapping where 200 accessions is considered a lower limit.
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31 370 However, the FIA colorimetric method for the determination of silicon in rice shoots proved
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33 371 not to be high throughput. However, in addition to the FIA method, a sub-set of samples
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35 372 were also analysed by P-XRF. The two different methods were strongly correlated, but not
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37 373 perfectly, and indicated that values for silicon content in samples measured by FIA were
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39 374 slightly higher than those measured by P-XRF. The observation that both methods provide
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41 375 comparable results highlights the conclusion that P-XRF can be used for silicon analysis to
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43 376 detect and measure genotypic differences across populations, instead of the more laborious
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45 377 and time-consuming alkali digestion method. Furthermore, a second advantage of P-XRF is
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47 378 that it is a non-destructive method. This would make it much more suitable for future GWA
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49 379 mapping studies.
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52 380 The plant material used in this study was previously used to examine the variation of shoot
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54 381 and grain arsenic (Norton et al., 2012). The comparison of shoot silicon and grain arsenic in
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3 382 this study is in agreement with previous studies where, in general, plants that had high shoot
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5 383 silicon also had lower grain arsenic (*Seyfferth and Ferdorf, 2012; Norton et al., 2012; Norton*
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7 384 *et al., 2013*). However, this study also adds more insight by taking into consideration the sub-
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9 385 population structure of rice cultivars. The correlation between shoot silicon and grain arsenic
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11 386 was sub-population specific. A strong relationship in between shoot silicon and grain arsenic
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13 387 was observed in *temperate japonica* and a weaker one in *tropical japonica*, but was not
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15 388 observed in *indica*, *aus* or *aromatic*. This important observation suggests that the genetic
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17 389 regulation of arsenic content in rice grain is different in *temperate* and *tropical japonicas*
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19 390 compared to the other rice sub-populations, implying that the silicon-transport-linked
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21 391 pathway implicated for arsenic accumulation (*Ma et al., 2007b; Norton et al., 2012*) may be
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23 392 less relevant in the *other* sub-populations.
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26

27 393 The accessions used in the study have been genotyped using a 700K SNP chip (*McCouch et*
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29 394 *al., 2016*). Single-marker analysis was used to test the candidacy of the known transporters of
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31 395 silica in rice. The study indicated that two SNPs within 10 kb of *Lsi2* and one within 10 kb
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33 396 of *Lsi3* were involved in contributing to the natural variation of shoot silicon accumulation in
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35 397 rice (Fig. 6). The *Lsi2* gene has been shown to be pivotal for transport of silicon and
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37 398 inorganic arsenic in studies conducted with mutants and transgenic plants (*Ma et al., 2006;*
38
39 399 *Ma et al., 2007b; Yamaji et al., 2008; Mitani-Ueno et al., 2011; Yamaji et al., 2015*). The
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41 400 identification of differences in shoot silicon and the link with three SNPs close to the genes
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43 401 further suggest that *Lsi2* and *Lsi3* are excellent candidate genes to explain the natural
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45 402 variation observed in shoot silicon content of rice. When looking at the sequencing variation
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47 403 of a number of diverse cultivars (which have been sequenced to a high depth) it is evident
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49 404 that there is only a small number of polymorphisms within the genes (Supplementary figures
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51 405 3 and 4). The highly conserved sequence for *Lsi2* may be due to its importance function for
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53 406 silicon accumulation in rice. However, the accessions screened in this study are likely to have
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greater sequence variation than the cultivars for which high quality sequence is available, and therefore there may be greater sequence variation for *Lsi2* (and the other *Lsi* genes) than that is represented in the 5 accessions reported here. A focus for future study will be to expand sequence information to more accessions to more fully explore sequence variation associated with the polymorphic SNPs presented in figure 6.

412

413 **5 Conclusion**

This study has demonstrated strong genotypic differences in shoot silicon in a diverse collection of rice cultivars, showing that there is potential to breed rice with increased silicon content that could improve resistance to both biotic and abiotic stresses in rice, which would help to maintain crop yields. The identification of significant SNPs linked with the shoot silicon phenotype within 10 kb of known silicon transporters warrants further study to investigate the impact of different alleles of these genes on silicon and arsenic accumulation in rice. Furthermore, the XRF method of silicon determination could be applied to GWA mapping studies that might reveal further candidate genes for silicon content in rice.

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2
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18 428 **References**

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Figure 1: Silicon in different organs of rice (bars are the mean of four replicates and error bars represent standard error of the mean). Letters above the columns (upper-case = anaerobic and lower case = aerobic) indicate statistically significant differences in silicon content of different plant organs using Tukey's test in two conditions. *denotes a significant difference between the two treatments for that plant organ.

Figure 2: Mean shoot silicon (mg g^{-1}) of 50 rice accessions determined by FIA. Different symbols refer to the accessions belonging to the different sub-populations; circle = *aus*, square = *indica*, cross = *aromatic*, triangle = *tropical japonica*, upside down triangle = *temperate japonica*. Error bars indicate the standard error of the mean ($n = 4$).

Figure 3: Shoot silicon (mg g^{-1}) content of 50 accessions in 5 different sub-populations of rice. ARO = *aromatic*, AUS = *aus*, IND = *indica*, TEJ = *temperate japonica* and TRJ = *tropical japonica*. The edges of each box show the upper and lower quantile and the bold line in the box shows the median value and the dotted line the mean value. The whiskers are the 10th and 90th percentiles.

Figure 4: Correlation of mean shoot silicon in 19 rice accessions determined by FIA and P-XRF. Error bars indicate the standard error of the mean ($n = 4$). Dotted line is the 1:1 line.

Figure 5: Correlation between shoot silicon (mg g^{-1}) and grain arsenic ($\mu\text{g kg}^{-1}$) in ARO= *aromatic*, AUS = *aus*, IND = *indica*, TEJ = *temperate japonica* and TRJ = *tropical japonica* subpopulations.

Figure 6: Variation in shoot silicon (mg g^{-1}) between different SNPs within 10 kb of *Lsi2* and *Lsi3*

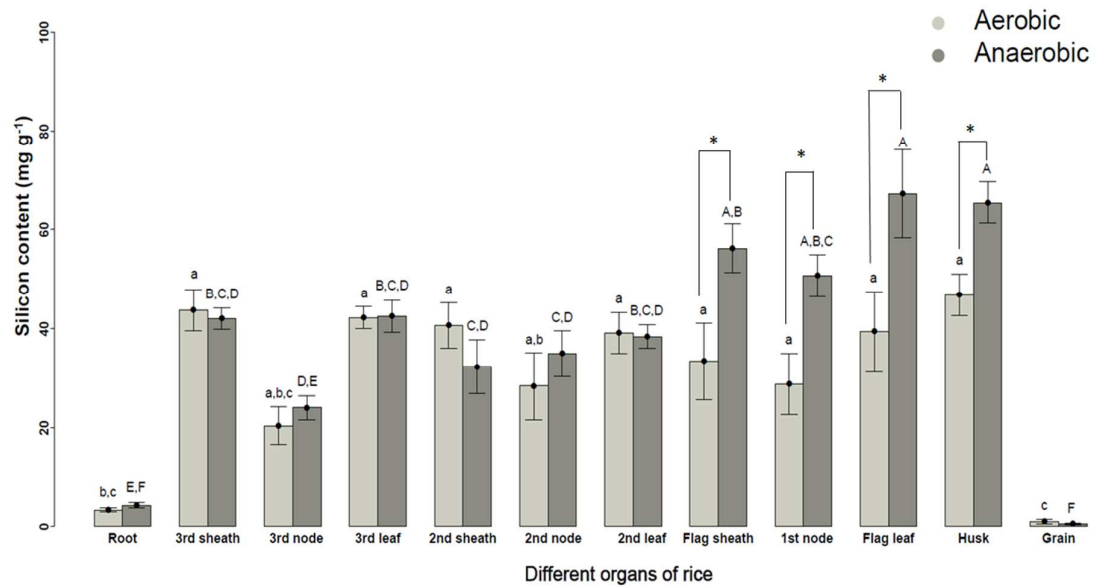


Figure 1

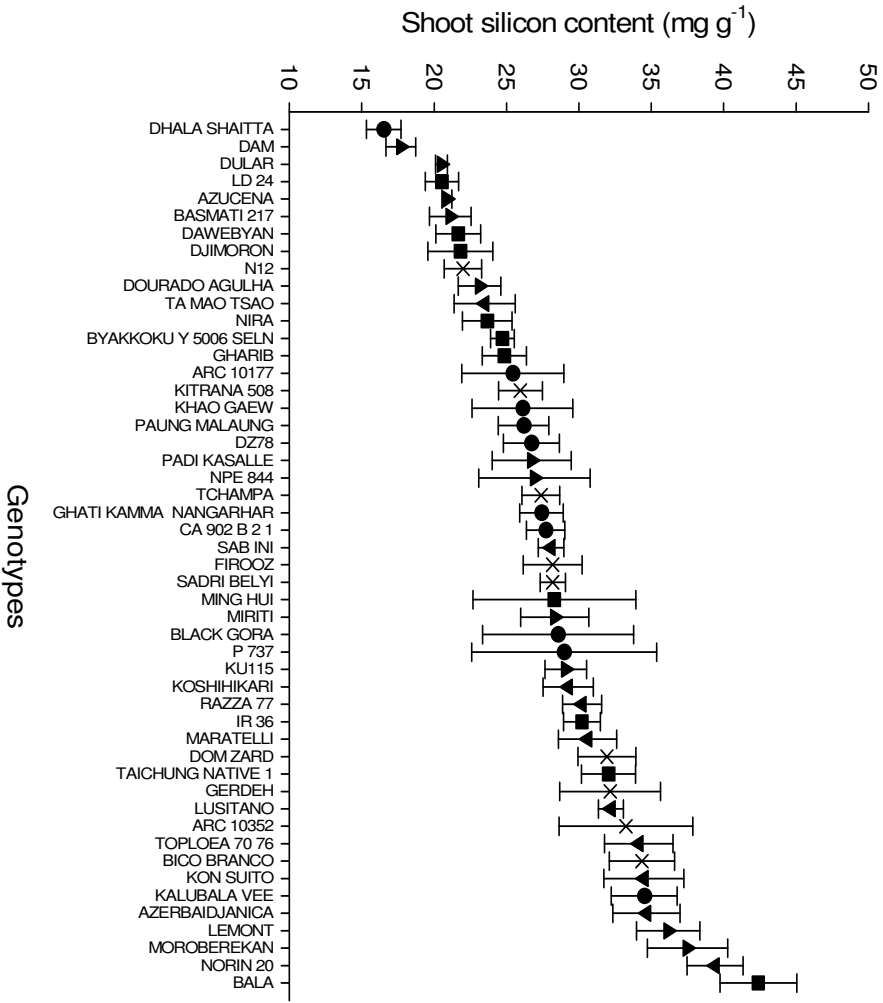


Figure 2

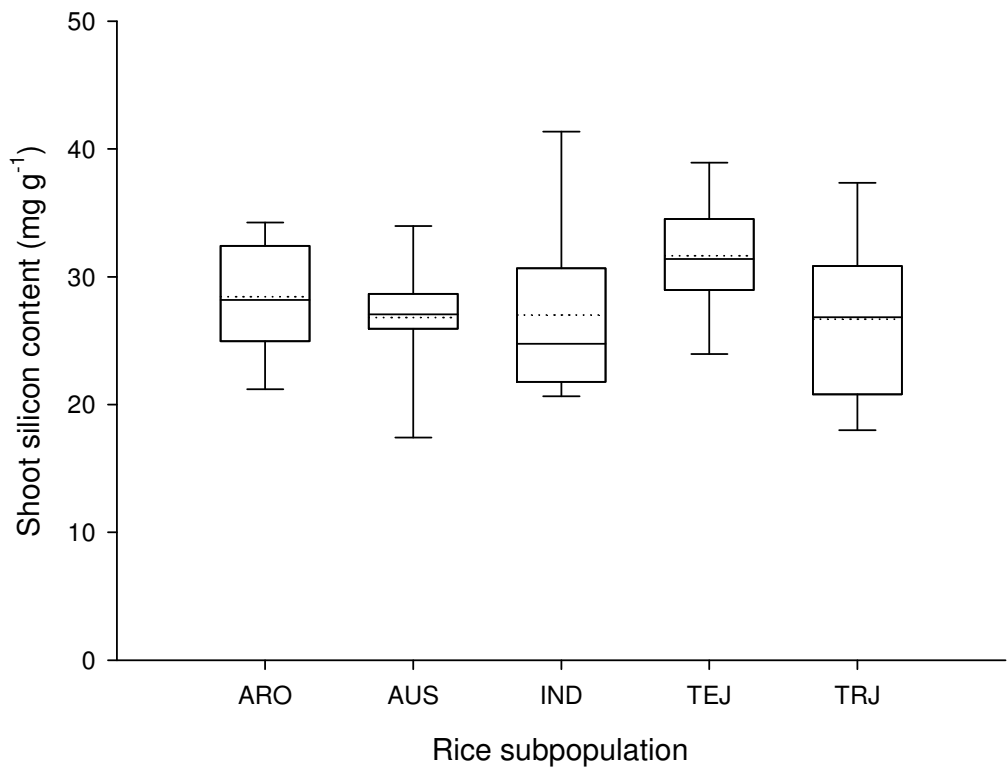
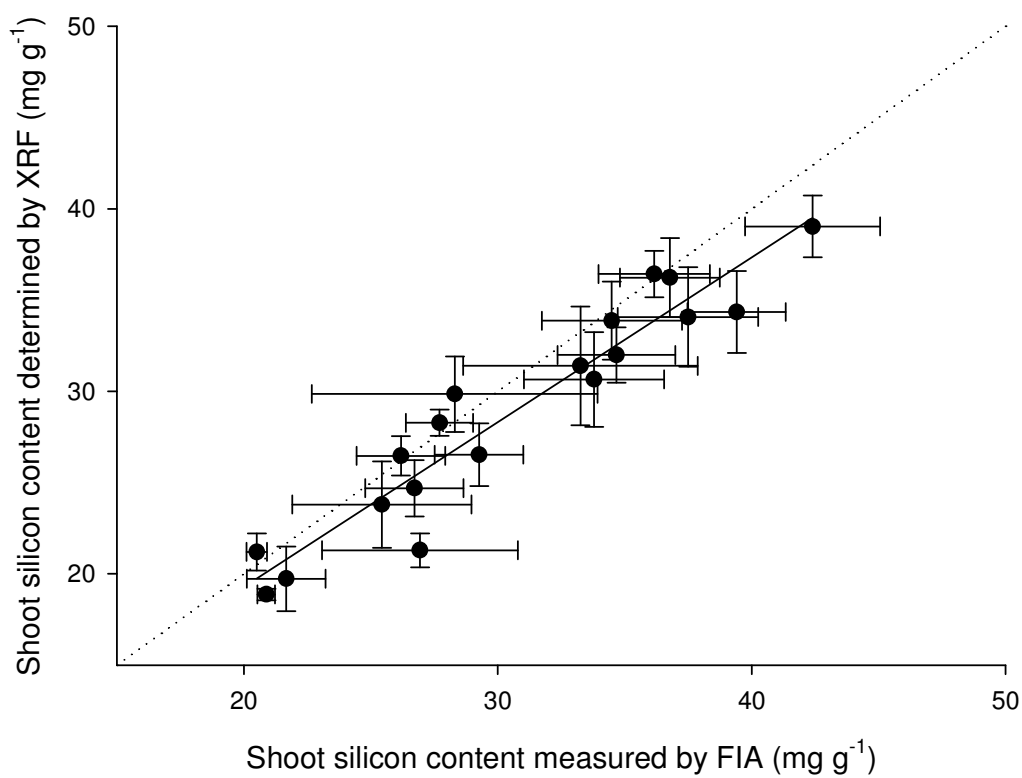


Figure 3



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685 **Figure 4**

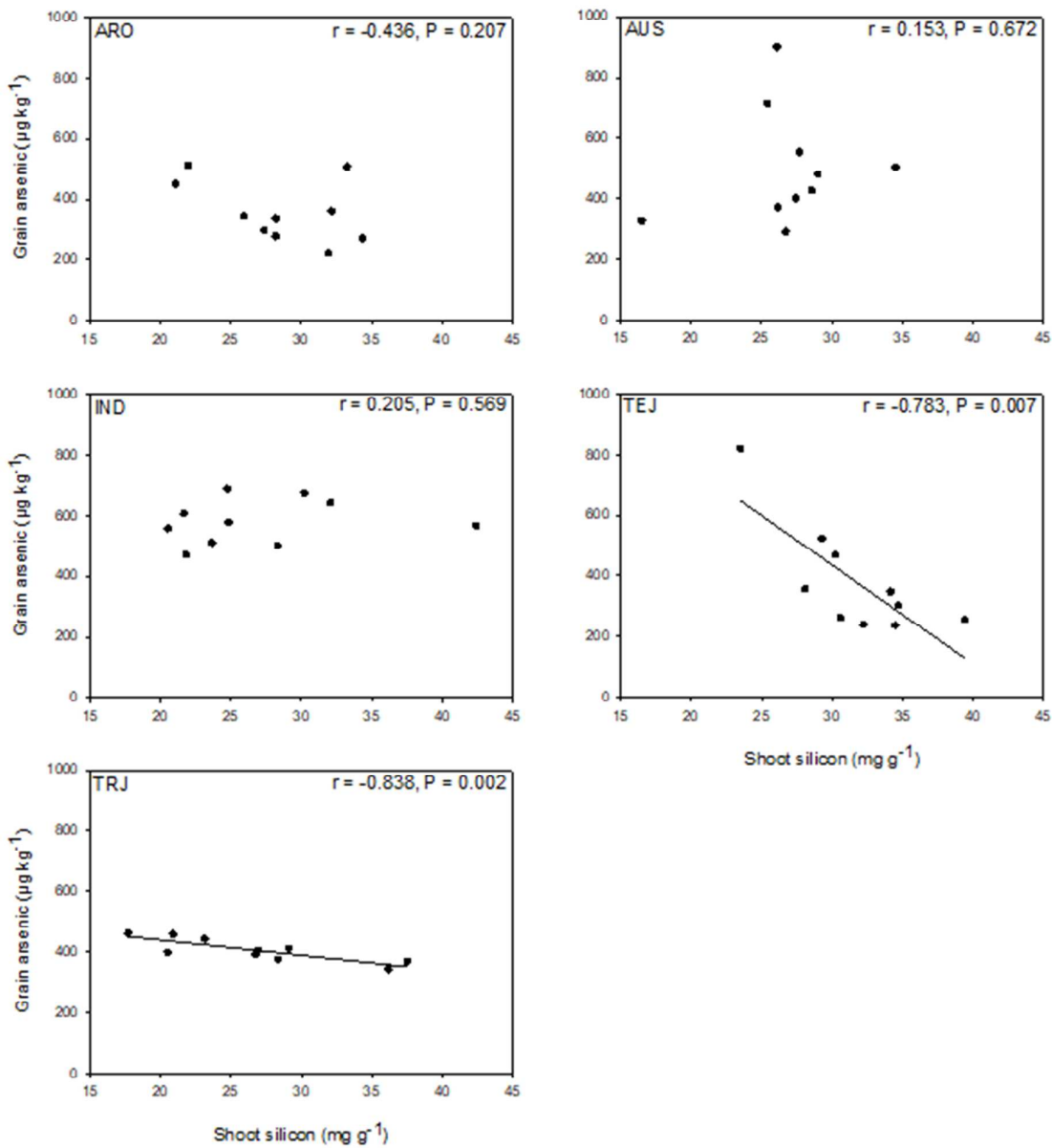
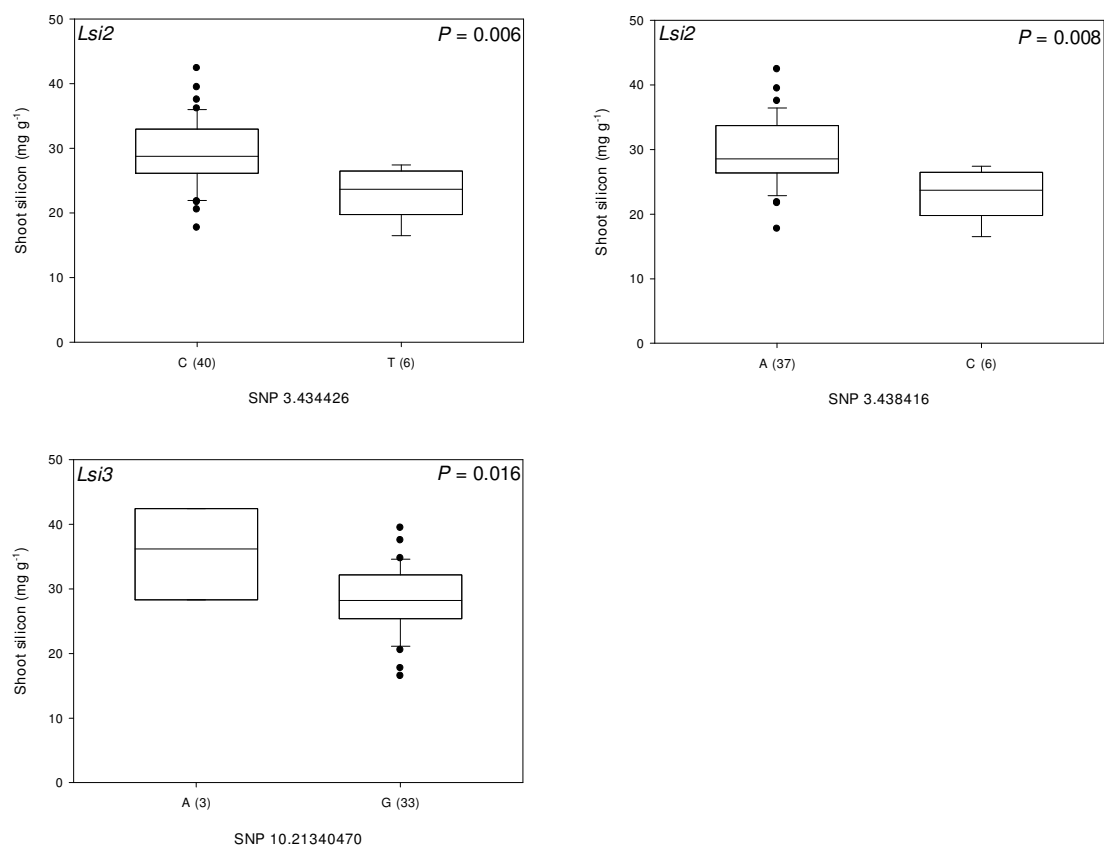


Figure 5

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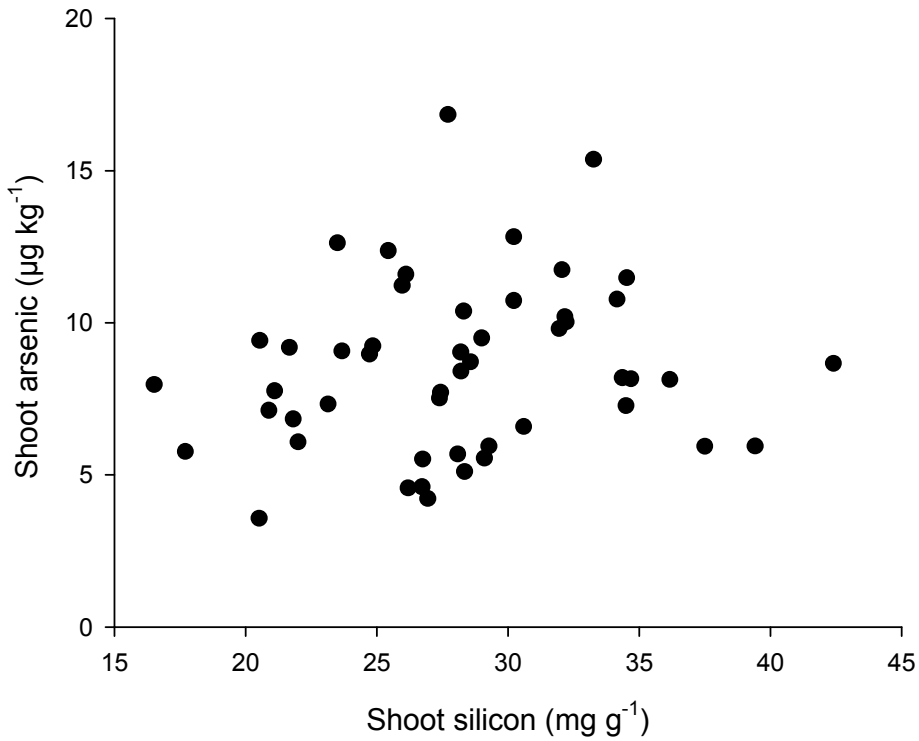
**Figure 6**

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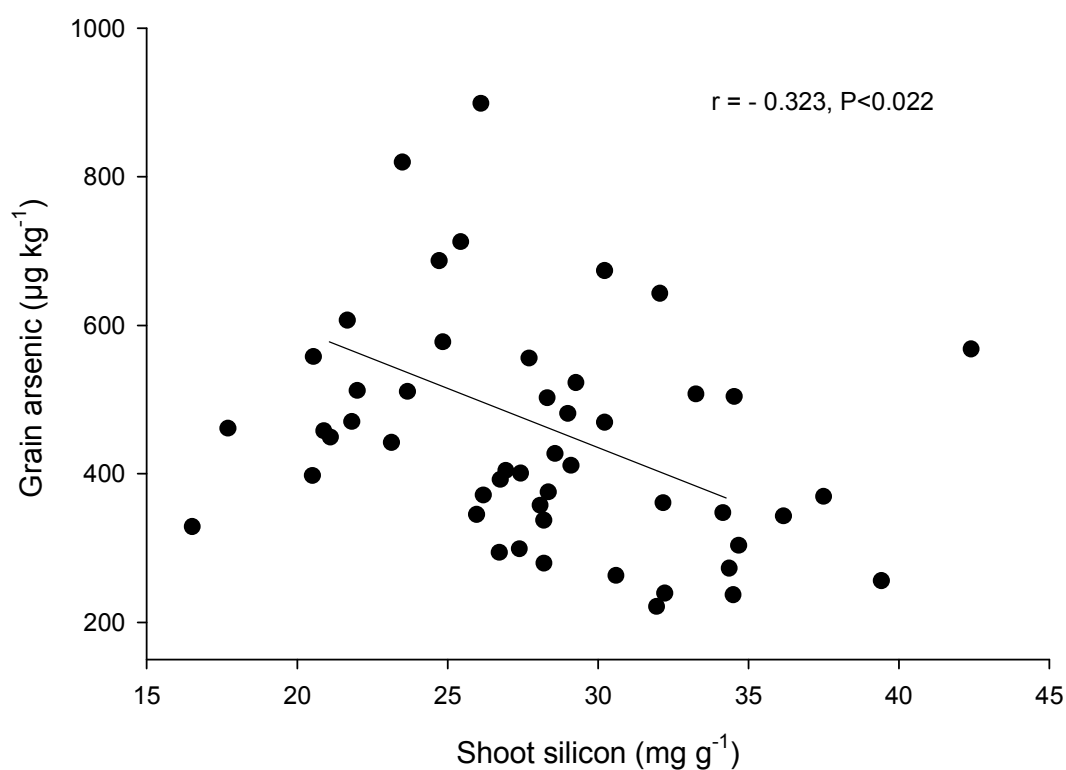
706 **Supplementary Table 1:** Selected genotype from RDP1 for shoot silicon analysis

707 **Supplementary Table 2:** SNPs with 10 kb (upstream and downstream) of *Lsi2*. SNPs data
708 taken from the High-density rice array (HDRA, 700k SNPs data) available at
709 <http://ricediversity.org/data/index.cfm>. After each SNP name (in brackets) is the location of
710 that SNP aligned to the Rice Genome Annotation Project – release 7 sequence available at
711 <http://rice.plantbiology.msu.edu/>

712 **Supplementary Table 3:** SNPs with 10 kb (upstream and downstream) of *Lsi3*. SNPs data
713 taken from the High-density rice array (HDRA, 700k SNPs data) available at
714 <http://ricediversity.org/data/index.cfm>. After each SNP name (in brackets) is the location of
715 that SNP aligned to the Rice Genome Annotation Project – release 7 sequence available at
716 <http://rice.plantbiology.msu.edu/>



Supplementary figure 1: Correlation between shoot silicon (mg g⁻¹) and shoot arsenic (µg kg⁻¹) within 50 accessions of RDP1.



Supplementary figure 2: Correlation between shoot silicon (mg g⁻¹) and grain arsenic (µg kg⁻¹) within 50 accessions of RDP1.

Lsi2 (LOC_Os03g01700)



Bala AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGCGTACCAGAGAGAGATCAT
 IR64 AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGCGTACCAGAGAGAGATCAT
 DJ123 AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGCGTACCAGAGAGAGATCAT
 Nipponbare AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGCGTACCAGAGAGAGATCAT
 Azucena AGCATCAGCAGCGCATGCACTAGCAGCTTAGCTACTGCGCGCGTACCAGAGAGAGATCAT

Bala CAGCTCAAGTAGCTAAGCTAGCTATAATCTGCTAGCTAGCTCGATCAGACACTTAATTAC
 IR64 CAGCTCAAGTAGCTAAGCTAGCTATAATCTGCTAGCTAGCTCGATCAGACACTTAATTAC
 DJ123 CAGCTCAAGTAGCTAAGCTAGCTATAATCTGCTAGCTAGCTCGATCAGACACTTAATTAC
 Nipponbare CAGCTCAAGTAGCTAAGCTAGCTATAATCTGCTAGCTAGCTCGATCAGACACTTAATTAC
 Azucena CAGCTCAAGTAGCTAAGCTAGCTATAATCTGCTAGCTAGCTCGATCAGACACTTAATTAC

Bala CTGCTAGGTGGTGGTCGATCGAAGAAGAA**AGATGAGTGAGCTTTCGTCGGCGCCCAAG**
 IR64 CTGCTAGGTGGTGGTGGTCGATCGAAGAAGAA**AGATGAGTGAGCTTTCGTCGGCGCCCAAG**
 DJ123 CTGCTAGGTGGTGGTCGATCGAAGAAGAA**AGATGAGTGAGCTTTCGTCGGCGCCCAAG**
 Nipponbare CTGCTAGGTGGTGGTGGTCGATCGAAGAAGAA**AGATGAGTGAGCTTTCGTCGGCGCCCAAG**
 Azucena CTGCTAGGTGGTGGTGGTCGATCGAAGAAGAA**AGATGAGTGAGCTTTCGTCGGCGCCCAAG**

Bala GTGGCGCTTGGATCGATCGCGTTTCGCGGTGTTCTGGATGATGGCGGTGTTCCCGTCGGTG
 IR64 GTGGCGCTTGGATCGATCGCGTTTCGCGGTGTTCTGGATGATGGCGGTGTTCCCGTCGGTG
 DJ123 GTGGCGCTTGGATCGATCGCGTTTCGCGGTGTTCTGGATGATGGCGGTGTTCCCGTCGGTG
 Nipponbare GTGGCGCTTGGATCGATCGCGTTTCGCGGTGTTCTGGATGATGGCGGTGTTCCCGTCGGTG
 Azucena GTGGCGCTTGGATCGATCGCGTTTCGCGGTGTTCTGGATGATGGCGGTGTTCCCGTCGGTG

Bala CCGTTCTCTGCCGATCGGGCGGACGGCGGGGTTCGTGCTGAGCGCGGTGCTGATGGTGATA
 IR64 CCGTTCTCTGCCGATCGGGCGGACGGCGGGGTTCGTGCTGAGCGCGGTGCTGATGGTGATA
 DJ123 CCGTTCTCTGCCGATCGGGCGGACGGCGGGGTTCGTGCTGAGCGCGGTGCTGATGGTGATA
 Nipponbare CCGTTCTCTGCCGATCGGGCGGACGGCGGGGTTCGTGCTGAGCGCGGTGCTGATGGTGATA
 Azucena CCGTTCTCTGCCGATCGGGCGGACGGCGGGGTTCGTGCTGAGCGCGGTGCTGATGGTGATA

Bala TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCCATCGACCTCCCAATCCTGGGCCCTC
 IR64 TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCCATCGACCTCCCAATCCTGGGCCCTC
 DJ123 TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCCATCGACCTCCCAATCCTGGGCCCTC
 Nipponbare TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCCATCGACCTCCCAATCCTGGGCCCTC
 Azucena TTCCACGTGATCAGCCCCGACGACGCGTACGCCCTCCATCGACCTCCCAATCCTGGGCCCTC

Bala CTCTTCGCCACCATGGTGGTGGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
 IR64 CTCTTCGCCACCATGGTGGTGGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
 D1J123 CTCTTCGCCACCATGGTGGTGGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
 Nipponbare CTCTTCGCCACCATGGTGGTGGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG
 Azucena CTCTTCGCCACCATGGTGGTGGGCAGCTACCTCCGGAACGCCGGGATGTTCAAGCACCTG

Bala GGGCGTCTGCTGGCGTGGAAAGAGCCAGGGCGGGCGCGACCTCATGTGCCGCGTCTGCGTC
 IR64 GGGCGTCTGCTGGCGTGGAAAGAGCCAGGGCGGGCGCGACCTCATGTGCCGCGTCTGCGTC
 DJ123 GGGCGTCTGCTGGCGTGGAAAGAGCCAGGGCGGGCGCGACCTCATGTGCCGCGTCTGCGTC
 Nipponbare GGGCGTCTGCTGGCGTGGAAAGAGCCAGGGCGGGCGCGACCTCATGTGCCGCGTCTGCGTC
 Azucena GGGCGTCTGCTGGCGTGGAAAGAGCCAGGGCGGGCGCGACCTCATGTGCCGCGTCTGCGTC

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| 1 | | |
| 2 | | |
| 3 | Bala | ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC |
| 4 | IR64 | ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC |
| 5 | DJ123 | ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC |
| 6 | Nipponbare | ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC |
| 7 | Azucena | ATGGTGGGGCTCAACATGTCGTGGACGGCCATCACCACGGCGCTGGCGCTGGTGGTGGTC |
| 8 | | ***** |
| 9 | Bala | GACTTCCGCGACGCCGAGCCGTGCCTGGACACCGTGTCTACTCGCTGCTCGTCTTCTTC |
| 10 | IR64 | GACTTCCGCGACGCCGAGCCGTGCCTGGACACCGTGTCTACTCGCTGCTCGTCTTCTTC |
| 11 | DJ123 | GACTTCCGCGACGCCGAGCCGTGCCTGGACACCGTGTCTACTCGCTGCTCGTCTTCTTC |
| 12 | Nipponbare | GACTTCCGCGACGCCGAGCCGTGCCTGGACACCGTGTCTACTCGCTGCTCGTCTTCTTC |
| 13 | Azucena | GACTTCCGCGACGCCGAGCCGTGCCTGGACACCGTGTCTACTCGCTGCTCGTCTTCTTC |
| 14 | | ***** |
| 15 | Bala | TCCGGGATGTTTCATCACCGTCAGCGGCTTCAACAAGACGGGCCTCCCGGGAGCCATCTGG |
| 16 | IR64 | TCCGGGATGTTTCATCACCGTCAGCGGCTTCAACAAGACGGGCCTCCCGGGAGCCATCTGG |
| 17 | DJ123 | TCCGGGATGTTTCATCACCGTCAGCGGCTTCAACAAGACGGGCCTCCCGGGAGCCATCTGG |
| 18 | Nipponbare | TCCGGGATGTTTCATCACCGTCAGCGGCTTCAACAAGACGGGCCTCCCGGGAGCCATCTGG |
| 19 | Azucena | TCCGGGATGTTTCATCACCGTCAGCGGCTTCAACAAGACGGGCCTCCCGGGAGCCATCTGG |
| 20 | | ***** |
| 21 | Bala | GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGCGGCATCTCCGTCTCTCCGTC |
| 22 | IR64 | GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGCGGCATCTCCGTCTCTCCGTC |
| 23 | DJ123 | GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGCGGCATCTCCGTCTCTCCGTC |
| 24 | Nipponbare | GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGCGGCATCTCCGTCTCTCCGTC |
| 25 | Azucena | GACTTCATGGCCCCCTACTCCAAGGTCAACAGCGTCGGCGGCATCTCCGTCTCTCCGTC |
| 26 | | ***** |
| 27 | Bala | ATCATCTCTCCTCTCCAACCTCGCATCAAACGTACCAACGGGTAATTATACATTCTAT |
| 28 | IR64 | ATCATCTCTCCTCTCCAACCTCGCATCAAACGTACCAACGGGTAATTATACATTCTAT |
| 29 | DJ123 | ATCATCTCTCCTCTCCAACCTCGCATCAAACGTACCAACGGGTAATTATACATTCTAT |
| 30 | Nipponbare | ATCATCTCTCCTCTCCAACCTCGCATCAAACGTACCAACGGGTAATTATACATTCTAT |
| 31 | Azucena | ATCATCTCTCCTCTCCAACCTCGCATCAAACGTACCAACGGGTAATTATACATTCTAT |
| 32 | | ***** |
| 33 | Bala | TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATACAAAAATAT |
| 34 | IR64 | TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATACAAAAATAT |
| 35 | DJ123 | TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATACAAAAATAT |
| 36 | Nipponbare | TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATACAAAAATAT |
| 37 | Azucena | TAATCCAGAATTAATGTATACTACTCCATATAAGTATAACTAATTAACATACAAAAATAT |
| 38 | | ***** |
| 39 | Bala | ATATGCGCATGCATGCAGTGCTTCTTATGGGTGATGAGGTGGCGAAGGCGGCGCGCTGA |
| 40 | IR64 | ATATGCGCATGCATGCAGTGCTTCTTATGGGTGATGAGGTGGCGAAGGCGGCGCGCTGA |
| 41 | DJ123 | ATATGCGCATGCATGCAGTGCTTCTTATGGGTGATGAGGTGGCGAAGGCGGCGCGCTGA |
| 42 | Nipponbare | ATATGCGCATGCATGCAGTGCTTCTTATGGGTGATGAGGTGGCGAAGGCGGCGCGCTGA |
| 43 | Azucena | ATATGCGCATGCATGCAGTGCTTCTTATGGGTGATGAGGTGGCGAAGGCGGCGCGCTGA |
| 44 | | ***** |
| 45 | Bala | TATCGCCGGCGGCGGTGACGACGTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGG |
| 46 | IR64 | TATCGCCGGCGGCGGTGACGACGTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGG |
| 47 | DJ123 | TATCGCCGGCGGCGGTGACGACGTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGG |
| 48 | Nipponbare | TATCGCCGGCGGCGGTGACGACGTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGG |
| 49 | Azucena | TATCGCCGGCGGCGGTGACGACGTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGG |
| 50 | | ***** |
| 51 | Bala | GGAACCTGTCGCTGCTGGGGTCGGCGGCGAACCTGATAGTGTGCGAGCAGGCGAGGAGGG |
| 52 | IR64 | GGAACCTGTCGCTGCTGGGGTCGGCGGCGAACCTGATAGTGTGCGAGCAGGCGAGGAGGG |
| 53 | DJ123 | GGAACCTGTCGCTGCTGGGGTCGGCGGCGAACCTGATAGTGTGCGAGCAGGCGAGGAGGG |
| 54 | Nipponbare | GGAACCTGTCGCTGCTGGGGTCGGCGGCGAACCTGATAGTGTGCGAGCAGGCGAGGAGGG |
| 55 | Azucena | GGAACCTGTCGCTGCTGGGGTCGGCGGCGAACCTGATAGTGTGCGAGCAGGCGAGGAGGG |
| 56 | | ***** |

Bala CGCCCAGGAACGCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
 IR64 CGCCCAGGAACGCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
 DJ123 CGCCCAGGAACGCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
 Nipponbare CGCCCAGGAACGCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA
 Azucena CGCCCAGGAACGCCTACGACCTCACCTTCTGGCAGCACATCGTCTTCGGCGTCCCATCCA

 Bala CCCTCATCGTCACCGCCGTCGGCATACCCCTCATCGGCAAGATCTGATCTCATCTCATCG
 IR64 CCCTCATCGTCACCGCCGTCGGCATACCCCTCATCGGCAAGATCTGATCTCATCTCATCG
 DJ123 CCCTCATCGTCACCGCCGTCGGCATACCCCTCATCGGCAAGATCTGATCTCATCTCATCG
 Nipponbare CCCTCATCGTCACCGCCGTCGGCATACCCCTCATCGGCAAGATCTGATCTCATCTCATCG
 Azucena CCCTCATCGTCACCGCCGTCGGCATACCCCTCATCGGCAAGATCTGATCTCATCTCATCG

 Bala ACCCATCCAAATTAATTAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTCGTCGT
 IR64 ACCCATCCAAATTAATTAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTCGTCGT
 DJ123 ACCCATCCAAATTAATTAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTCGTCGT
 Nipponbare ACCCATCCAAATTAATTAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTCGTCGT
 Azucena ACCCATCCAAATTAATTAATTATGAGATCGACAAACATCCAAGCTTGCTAGGCTCGTCGT

 Bala CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGCACGTATATATATGCT
 IR64 CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGCACGTATATATATGCT
 DJ123 CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGCACGTATATATATGCT
 Nipponbare CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGCACGTATATATATGCT
 Azucena CGTCGTCGTCGACCACCGTACCATATATATGCATGCATGCCACGCACGTATATATATGCT

 Bala CCTCAACCTCAAGTCAATTAATTAAGAGAATGGATGAATGAATCAATGTTGTGTATGATT
 IR64 CCTCAACCTCAAGTCAATTAATTAAGAGAATGGATGAATGAATCAATGTTGTGTATGATT
 DJ123 CCTCAACCTCAAGTCAATTAATTAAGAGAATGGATGAATGAATCAATGTTGTGTATGATT
 Nipponbare CCTCAACCTCAAGTCAATTAATTAAGAGAATGGATGAATGAATCAATGTTGTGTATGATT
 Azucena CCTCAACCTCAAGTCAATTAATTAAGAGAATGGATGAATGAATCAATGTTGTGTATGATT

 Bala TCCTTTTTGTTTTGTTTTTACCACCGTATATGTGCTGTGTGGTGTATGGCCGGCGAGCT
 IR64 TCCTTTTTGTTTTGTTTTTACCACCGTATATGTGCTGTGTGGTGTATGGCCGGCGAGCT
 DJ123 TCCTTTTTGTTTTGTTTTTACCACCGTATATGTGCTGTGTGGTGTATGGCCGGCGAGCT
 Nipponbare TCCTTTTTGTTTTGTTTTTACCACCGTATATGTGCTGTGTGGTGTATGGCCGGCGAGCT
 Azucena TCCTTTTTGTTTTGTTTTTACCACCGTATATGTGCTGTGTGGTGTATGGCCGGCGAGCT

 Bala TGAAATTGAATTGGCCTTGCTTGCATGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
 IR64 TGAAATTGAATTGGCCTTGCTTGCATGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
 DJ123 TGAAATTGAATTGGCCTTGCTTGCATGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
 Nipponbare TGAAATTGAATTGGCCTTGCTTGCATGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA
 Azucena TGAAATTGAATTGGCCTTGCTTGCATGCATGCATGGTGATCAGATCAGCTCAGCTAGCTA

 Bala GCTAAATCTTGCTTAATTTTATCAAGTGTCAATCATGATGAGAAGAGAGGGAACCTAGATG
 IR64 GCTAAATCTTGCTTAATTTTATCAAGTGTCAATCATGATGAGAAGAGAGGGAACCTAGATG
 DJ123 GCTAAATCTTGCTTAATTTTATCAAGTGTCAATCATGATGAGAAGAGAGGGAACCTAGATG
 Nipponbare GCTAAATCTTGCTTAATTTTATCAAGTGTCAATCATGATGAGAAGAGAGGGAACCTAGATG
 Azucena GCTAAATCTTGCTTAATTTTATCAAGTGTCAATCATGATGAGAAGAGAGGGAACCTAGATG

 Bala GAGCTAGCTAGCTAGCAACACATGCAGTGTGTGCTAAGCAGTGCACCTGTCATGCTTAAA
 IR64 GAGCTAGCTAGCTAGCAACACATGCAGTGTGTGCTAAGCAGTGCACCTGTCATGCTTAAA
 DJ123 GAGCTAGCTAGCTAGCAACACATGCAGTGTGTGCTAAGCAGTGCACCTGTCATGCTTAAA
 Nipponbare GAGCTAGCTAGCTAGCAACACATGCAGTGTGTGCTAAGCAGTGCACCTGTCATGCTTAAA
 Azucena GAGCTAGCTAGCTAGCAACACATGCAGTGTGTGCTAAGCAGTGCACCTGTCATGCTTAAA

| | | |
|----|------------|---|
| 1 | | |
| 2 | | |
| 3 | Bala | TTTGCTTGCTTAATTCAACATCGTATTACATGTATGTTGATATGTGCATGTTCCCTCAAAA |
| 4 | IR64 | TTTGCTTGCTTAATTCAACATCGTATTACATGTATGTTGATATGTGCATGTTCCCTCAAAA |
| 5 | DJ123 | TTTGCTTGCTTAATTCAACATCGTATTACATGTATGTTGATATGTGCATGTTCCCTCAAAA |
| 6 | Nipponbare | TTTGCTTGCTTAATTCAACATCGTATTACATGTATGTTGATATGTGCATGTTCCCTCAAAA |
| 7 | Azucena | TTTGCTTGCTTAATTCAACATCGTATTACATGTATGTTGATATGTGCATGTTCCCTCAAAA |
| 8 | | ***** |
| 9 | Bala | TACATGGAAAAAAATGTTTCTACACATCTCAATCGTATAGAGTATGTAGCATGTCTTGT |
| 10 | IR64 | TACATGGAAAAAAATGTTTCTACACATCTCAATCGTATAGAGTATGTAGCATGTCTTGT |
| 11 | DJ123 | TACATGGAAAAAAATGTTTCTACACATCTCAATCGTATAGAGTATGTAGCATGTCTTGT |
| 12 | Nipponbare | TACATGGAAAAAAATGTTTCTACACATCTCAATCGTATAGAGTATGTAGCATGTCTTGT |
| 13 | Azucena | TACATGGAAAAAAATGTTTCTACACATCTCAATCGTATAGAGTATGTAGCATGTCTTGT |
| 14 | | ***** |
| 15 | Bala | TCAAACAAATATTTGTAGCCCTCTCTCTCTCTCTCTCTTTAGGTTTATTAATTAATTAGT |
| 16 | IR64 | TCAAACAAATATTTGTAGCCCTCTCTCTCTCTCTCTCTTTAGGTTTATTAATTAATTAGT |
| 17 | DJ123 | TCAAACAAATATTTGTAGCCCTCTCTCTCTCTCTCTCTTTAGGTTTATTAATTAATTAGT |
| 18 | Nipponbare | TCAAACAAATATTTGTAGCCCTCTCTCTCTCTCTCTCTTTAGGTTTATTAATTAATTAGT |
| 19 | Azucena | TCAAACAAATATTTGTAGCCCTCTCTCTCTCTCTCTCTTTAGGTTTATTAATTAATTAGT |
| 20 | | ***** |
| 21 | Bala | CTGTTCAATGTCACCTCAGTTCATAGAGATGCTATACTTAATTAATTGCTCCATGTATGC |
| 22 | IR64 | CTGTTCAATGTCACCTCAGTTCATAGAGATGCTATACTTAATTAATTGCTCCATGTATGC |
| 23 | DJ123 | CTGTTCAATGTCACCTCAGTTCATAGAGATGCTATACTTAATTAATTGCTCCATGTATGC |
| 24 | Nipponbare | CTGTTCAATGTCACCTCAGTTCATAGAGATGCTATACTTAATTAATTGCTCCATGTATGC |
| 25 | Azucena | CTGTTCAATGTCACCTCAGTTCATAGAGATGCTATACTTAATTAATTGCTCCATGTATGC |
| 26 | | ***** |
| 27 | | |
| 28 | Bala | AGAGTTGTACTACCTTAAAATATCAGTATGCATGCTCTTGTTGCCATTCCAAAAGAACAA |
| 29 | IR64 | AGAGTTGTACTACCTTAAAATATCAGTATGCATGCTCTTGTTGCCATTCCAAAAGAACAA |
| 30 | DJ123 | AGAGTTGTACTACCTTAAAATATCAGTATGCATGCTCTTGTTGCCATTCCAAAAGAACAA |
| 31 | Nipponbare | AGAGTTGTACTACCTTAAAATATCAGTATGCATGCTCTTGTTGCCATTCCAAAAGAACAA |
| 32 | Azucena | AGAGTTGTACTACCTTAAAATATCAGTATGCATGCTCTTGTTGCCATTCCAAAAGAACAA |
| 33 | | ***** |
| 34 | Bala | A |
| 35 | IR64 | A |
| 36 | DJ123 | A |
| 37 | Nipponbare | A |
| 38 | Azucena | A |
| 39 | | * |

Supplementary figure 3. Genomic sequence alignment of *Lsi2*. Regions highlighted in bold are predicted exon regions based on the Rice Genome Annotation Project – release 7 sequence available at <http://rice.plantbiology.msu.edu/>.

Lsi3 (LOC_Os10g39980)

Bala ACTCTCATCGCCACCACCACCTCCCACTCCAAGACGCTAGCTGCATTGTAGTGAGAGAGA
 IR64 ACTCTCATCGCCACCACCACCTCCCACTCCAAGACGCTAGCTGCATTGTAGTGAGAGAGA
 DJ123 ACTCTCATCGCCACCACCACCTCCCACTCCAAGACGCTAGCTGCATTGTAGTGAGAGAGA
 Nipponbare ACTCTCATCGCCACCACCACCTCCCACTCCAAGACGCTAGCTGCATTGTAGTGAGAGAGA
 Azucena ACTCTCATCGCCACCACCACCTCCCACTCCAAGACGCTAGCTGCATTGTAGTGAGAGAGA

Bala GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGGTGGGGTCGGTGGCGTTCGGT
 IR64 GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGGTGGGGTCGGTGGCGTTCGGT
 DJ123 GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGGTGGGGTCGGTGGCGTTCGGT
 Nipponbare GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGGTGGGGTCGGTGGCGTTCGGT
 Azucena GAGGCGGCCATGGCGCTGGCGTCGCTGCCGAAGGTGGTGGTGGGGTCGGTGGCGTTCGGT

Bala GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCTGCCGATCGGGCGGACGGCG
 IR64 GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCTGCCGATCGGGCGGACGGCG
 DJ123 GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCTGCCGATCGGGCGGACGGCG
 Nipponbare GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCTGCCGATCGGGCGGACGGCG
 Azucena GTGTTCTGGATGCTGGCGGTGTTCCCGTCGGTGCCGTTCTGCCGATCGGGCGGACGGCG

Bala GGGGCGCTGCTGGGCGCGGTGCTGATGATCGTGTTCACGTGATCAGCGCCGACGACGCC
 IR64 GGGGCGCTGCTGGGCGCGGTGCTGATGATCGTGTTCACGTGATCAGCGCCGACGACGCC
 DJ123 GGGGCGCTGCTGGGCGCGGTGCTGATGATCGTGTTCACGTGATCAGCGCCGACGACGCC
 Nipponbare GGGGCGCTGCTGGGCGCGGTGCTGATGATCGTGTTCACGTGATCAGCGCCGACGACGCC
 Azucena GGGGCGCTGCTGGGCGCGGTGCTGATGATCGTGTTCACGTGATCAGCGCCGACGACGCC

Bala TACGCCTCCATCGACCTCCCCATCCTGGGCCTCCTCTTCGCCACCATGGTCGTGGCGGC
 IR64 TACGCCTCCATCGACCTCCCCATCCTGGGCCTCCTCTTCGCCACCATGGTCGTGGCGGC
 DJ123 TACGCCTCCATCGACCTCCCCATCCTGGGCCTCCTCTTCGCCACCATGGTCGTGGCGGC
 Nipponbare TACGCCTCCATCGACCTCCCCATCCTGGGCCTCCTCTTCGCCACCATGGTCGTGGCGGC
 Azucena TACGCCTCCATCGACCTCCCCATCCTGGGCCTCCTCTTCGCCACCATGGTCGTGGCGGC

Bala TACCTCAAGAACGCCGGCATGTTACGGCACCTGGGCGGCTGCTGGCGTGGCGGAGCCAG
 IR64 TACCTCAAGAACGCCGGCATGTTACGGCACCTGGGCGGCTGCTGGCGTGGCGGAGCCAG
 DJ123 TACCTCAAGAACGCCGGCATGTTACGGCACCTGGGCGGCTGCTGGCGTGGCGGAGCCAG
 Nipponbare TACCTCAAGAACGCCGGCATGTTACGGCACCTGGGCGGCTGCTGGCGTGGCGGAGCCAG
 Azucena TACCTCAAGAACGCCGGCATGTTACGGCACCTGGGCGGCTGCTGGCGTGGCGGAGCCAG

Bala GGC GGCGCGACCTCATGTGCCGCGTCTGCGTCGTACCGCGCTCGCCAGCGCGCTCTTC
 IR64 GGC GGCGCGACCTCATGTGCCGCGTCTGCGTCGTACCGCGCTCGCCAGCGCGCTCTTC
 DJ123 GGC GGCGCGACCTCATGTGCCGCGTCTGCGTCGTACCGCGCTCGCCAGCGCGCTCTTC
 Nipponbare GGC GGCGCGACCTCATGTGCCGCGTCTGCGTCGTACCGCGCTCGCCAGCGCGCTCTTC
 Azucena GGC GGCGCGACCTCATGTGCCGCGTCTGCGTCGTACCGCGCTCGCCAGCGCGCTCTTC

Bala ACCAACGACACCTGCTGCGTCGTCCTACCGAGTTCGTGCTCGAGCTCGCCGCCGAGCGC
 IR64 ACCAACGACACCTGCTGCGTCGTCCTACCGAGTTCGTGCTCGAGCTCGCCGCCGAGCGC
 DJ123 ACCAACGACACCTGCTGCGTCGTCCTACCGAGTTCGTGCTCGAGCTCGCCGCCGAGCGC
 Nipponbare ACCAACGACACCTGCTGCGTCGTCCTACCGAGTTCGTGCTCGAGCTCGCCGCCGAGCGC
 Azucena ACCAACGACACCTGCTGCGTCGTCCTACCGAGTTCGTGCTCGAGCTCGCCGCCGAGCGC

| | | |
|----|------------|--|
| 1 | | |
| 2 | | |
| 3 | Bala | AACCTCCCGGCGAAGCCGTTCCCTGCTCGCGCTCGCCACCAGCGCCAACATCGGCTCCAGC |
| 4 | IR64 | AACCTCCCGGCGAAGCCGTTCCCTGCTCGCGCTCGCCACCAGCGCCAACATCGGCTCCAGC |
| 5 | DJ123 | AACCTCCCGGCGAAGCCGTTCCCTGCTCGCGCTCGCCACCAGCGCCAACATCGGCTCCAGC |
| 6 | Nipponbare | AACCTCCCGGCGAAGCCGTTCCCTGCTCGCGCTCGCCACCAGCGCCAACATCGGCTCCAGC |
| 7 | Azucena | AACCTCCCGGCGAAGCCGTTCCCTGCTCGCGCTCGCCACCAGCGCCAACATCGGCTCCAGC |
| 8 | | ***** |
| 9 | Bala | GCGACGCCCATCGGCAACCCGCGAGAACCCTGGTCATCGCCTTCAACAGCAAGATCTCCTTT |
| 10 | IR64 | GCGACGCCCATCGGCAACCCGCGAGAACCCTGGTCATCGCCTTCAACAGCAAGATCTCCTTT |
| 11 | DJ123 | GCGACGCCCATCGGCAACCCGCGAGAACCCTGGTCATCGCCTTCAACAGCAAGATCTCCTTT |
| 12 | Nipponbare | GCGACGCCCATCGGCAACCCGCGAGAACCCTGGTCATCGCCTTCAACAGCAAGATCTCCTTT |
| 13 | Azucena | GCGACGCCCATCGGCAACCCGCGAGAACCCTGGTCATCGCCTTCAACAGCAAGATCTCCTTT |
| 14 | | ***** |
| 15 | Bala | ATTTCCCTTCCTCCTCGGCATCCTCCCCGCCATGCTCGCCGGCATGGGGATCAACATGCTC |
| 16 | IR64 | ATTTCCCTTCCTCCTCGGCATCCTCCCCGCCATGCTCGCCGGCATGGGGATCAACATGCTC |
| 17 | DJ123 | ATTTCCCTTCCTCCTCGGCATCCTCCCCGCCATGCTCGCCGGCATGGGGATCAACATGCTC |
| 18 | Nipponbare | ATTTCCCTTCCTCCTCGGCATCCTCCCCGCCATGCTCGCCGGCATGGGGATCAACATGCTC |
| 19 | Azucena | ATTTCCCTTCCTCCTCGGCATCCTCCCCGCCATGCTCGCCGGCATGGGGATCAACATGCTC |
| 20 | | ***** |
| 21 | Bala | ATGCTGCTCTGCATGTACTGGAAGGAGCTCGACGGCGGCGCCTGCAGCCCCGACGAGGTC |
| 22 | IR64 | ATGCTGCTCTGCATGTACTGGAAGGAGCTCGACGGCGGCGCCTGCAGCCCCGACGAGGTC |
| 23 | DJ123 | ATGCTGCTCTGCATGTACTGGAAGGAGCTCGACGGCGGCGCCTGCAGCCCCGACGAGGTC |
| 24 | Nipponbare | ATGCTGCTCTGCATGTACTGGAAGGAGCTCGACGGCGGCGCCTGCAGCCCCGACGAGGTC |
| 25 | Azucena | ATGCTGCTCTGCATGTACTGGAAGGAGCTCGACGGCGGCGCCTGCAGCCCCGACGAGGTC |
| 26 | | ***** |
| 27 | Bala | GCCGCCGGCAAGCAGATGGAGGCCATCGAGGAGGGCCGCCGACGGCGCTCAACAACAAC |
| 28 | IR64 | GCCGCCGGCAAGCAGATGGAGGCCATCGAGGAGGGCCGCCGACGGCGCTCAACAACAAC |
| 29 | DJ123 | GCCGCCGGCAAGCAGATGGAGGCCATCGAGGAGGGCCGCCGACGGCGCTCAACAACAAC |
| 30 | Nipponbare | GCCGCCGGCAAGCAGATGGAGGCCATCGAGGAGGGCCGCCGACGGCGCTCAACAACAAC |
| 31 | Azucena | GCCGCCGGCAAGCAGATGGAGGCCATCGAGGAGGGCCGCCGACGGCGCTCAACAACAAC |
| 32 | | ***** |
| 33 | Bala | AAGAAAGACGACGGCGACGCCGCCACCCCGCATCGCCGGAGGACGACGACGGCGGCGAC |
| 34 | IR64 | AAGAAAGACGACGGCGACGCCGCCACCCCGCATCGCCGGAGGACGACGACGGCGGCGAC |
| 35 | DJ123 | AAGAAAGACGACGGCGACGCCGCCACCCCGCATCGCCGGAGGACGACGACGGCGGCGAC |
| 36 | Nipponbare | AAGAAAGACGACGGCGACGCCGCCACCCCGCATCGCCGGAGGACGACGACGGCGGCGAC |
| 37 | Azucena | AAGAAAGACGACGGCGACGCCGCCACCCCGCATCGCCGGAGGACGACGACGGCGGCGAC |
| 38 | | ***** |
| 39 | Bala | GCCGAGTCGATGATGTCGGAGAACATCTCGACGAAGCACCGGTGGTTTCATGCAGTGCTCG |
| 40 | IR64 | GCCGAGTCGATGATGTCGGAGAACATCTCGACGAAGCACCGGTGGTTTCATGCAGTGCTCG |
| 41 | DJ123 | GCCGAGTCGATGATGTCGGAGAACATCTCGACGAAGCACCGGTGGTTTCATGCAGTGCTCG |
| 42 | Nipponbare | GCCGAGTCGATGATGTCGGAGAACATCTCGACGAAGCACCGGTGGTTTCATGCAGTGCTCG |
| 43 | Azucena | GCCGAGTCGATGATGTCGGAGAACATCTCGACGAAGCACCGGTGGTTTCATGCAGTGCTCG |
| 44 | | ***** |
| 45 | Bala | GAGCACCGGCGGAAGCTATTCCCTCAAGTCTTCGCGTACGTGGTCACCGTCGGCATGCTC |
| 46 | IR64 | GAGCACCGGCGGAAGCTATTCCCTCAAGTCTTCGCGTACGTGGTCACCGTCGGCATGCTC |
| 47 | DJ123 | GAGCACCGGCGGAAGCTATTCCCTCAAGTCTTCGCGTACGTGGTCACCGTCGGCATGCTC |
| 48 | Nipponbare | GAGCACCGGCGGAAGCTATTCCCTCAAGTCTTCGCGTACGTGGTCACCGTCGGCATGCTC |
| 49 | Azucena | GAGCACCGGCGGAAGCTATTCCCTCAAGTCTTCGCGTACGTGGTCACCGTCGGCATGCTC |
| 50 | | ***** |
| 51 | Bala | GTCGCCTACATGCTCGGCCTCAACATGTCGTGGACCGCCATCACGACCGCCATCGCCCTC |
| 52 | IR64 | GTCGCCTACATGCTCGGCCTCAACATGTCGTGGACCGCCATCACGACCGCCATCGCCCTC |
| 53 | DJ123 | GTCGCCTACATGCTCGGCCTCAACATGTCGTGGACCGCCATCACGACCGCCATCGCCCTC |
| 54 | Nipponbare | GTCGCCTACATGCTCGGCCTCAACATGTCGTGGACCGCCATCACGACCGCCATCGCCCTC |
| 55 | Azucena | GTCGCCTACATGCTCGGCCTCAACATGTCGTGGACCGCCATCACGACCGCCATCGCCCTC |
| 56 | | ***** |

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| Bala | TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATATTTTTTTAATAAGAT |
| IR64 | TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATATTTTTTTAATAAGAT |
| DJ123 | TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATATTTTTTTAATAAGAT |
| Nipponbare | TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATATTTTTTTAATAAGAT |
| Azucena | TAAATCAGACAATAAATCAAGTCAAAATAAAATAAATGTAATTATATTTTTTTAATAAGAT |
| ***** | |
| Bala | GAATGATTATTAAGAAAAAATTTTTTTGTCTCTCATAATTAAGAAAAAGAACACTCA |
| IR64 | GAATGATTATTAAGAAAAAATTTTTTTGTCTCTCATAATTAAGAAAAAGAACACTCA |
| DJ123 | GAATGATTATTAAGAAAAAATTTTTTTGTCTCTCATAATTAAGAAAAAGAACACTCA |
| Nipponbare | GAATGATTATTAAGAAAAAATTTTTTTGTCTCTCATAATTAAGAAAAAGAACACTCA |
| Azucena | GAATGATTATTAAGAAAAAATTTTTTTGTCTCTCATAATTAAGAAAAAGAACACTCA |
| ***** | |
| Bala | TTTTTTACATGTTATTTGAGTGGAATTGTAAATTAATTTATATATGTGGGTGTAGT GT |
| IR64 | TTTTTTACATGTTATTTGAGTGGAATTGTAAATTAATTTATATATGTGGGTGTAGT GT |
| DJ123 | TTTTTTACATGTTATTTGAGTGGAATTGTAAATTAATTTATATATGTGGGTGTAGT GT |
| Nipponbare | TTTTTTACATGTTATTTGAGTGGAATTGTAAATTAATTTATATATGTGGGTGTAGT GT |
| Azucena | TTTTTTACATGTTATTTGAGTGGAATTGTAAATTAATTTATATATGTGGGTGTAGT GT |
| ***** | |
| Bala | GTTGATGGGGGATGAGGTGGCGCGCGCGCGCGCGACGATATCACCGCGCGCGGTGACGCG |
| IR64 | GTTGATGGGGGATGAGGTGGCGCGCGCGCGCGCGACGATATCACCGCGCGCGGTGACGCG |
| DJ123 | GTTGATGGGGGATGAGGTGGCGCGCGCGCGCGCGACGATATCACCGCGCGCGGTGACGCG |
| Nipponbare | GTTGATGGGGGATGAGGTGGCGCGCGCGCGCGCGACGATATCACCGCGCGCGGTGACGCG |
| Azucena | GTTGATGGGGGATGAGGTGGCGCGCGCGCGCGCGACGATATCACCGCGCGCGGTGACGCG |
| ***** | |
| Bala | GTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGGGGAACCTGTCGCTGCTGGGGTC |
| IR64 | GTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGGGGAACCTGTCGCTGCTGGGGTC |
| DJ123 | GTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGGGGAACCTGTCGCTGCTGGGGTC |
| Nipponbare | GTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGGGGAACCTGTCGCTGCTGGGGTC |
| Azucena | GTCGTGGCTGCTGCTGGCGTGGGTGAGCACGGTGGCGGGGAACCTGTCGCTGCTGGGGTC |
| ***** | |
| Bala | GGCGGCGAACCTGATCGTGTGCGAGCAGGCGAGGCGGGCGACGCGCAACGCCTACGACCT |
| IR64 | GGCGGCGAACCTGATCGTGTGCGAGCAGGCGAGGCGGGCGACGCGCAACGCCTACGACCT |
| DJ123 | GGCGGCGAACCTGATCGTGTGCGAGCAGGCGAGGCGGGCGACGCGCAACGCCTACGACCT |
| Nipponbare | GGCGGCGAACCTGATCGTGTGCGAGCAGGCGAGGCGGGCGACGCGCAACGCCTACGACCT |
| Azucena | GGCGGCGAACCTGATCGTGTGCGAGCAGGCGAGGCGGGCGACGCGCAACGCCTACGACCT |
| ***** | |
| Bala | CACCTTCTGGAACACGTATCTTTCGGACTCCCATCCACCCTCGTCGTACCGCCATCGG |
| IR64 | CACCTTCTGGAACACGTATCTTTCGGACTCCCATCCACCCTCGTCGTACCGCCATCGG |
| DJ123 | CACCTTCTGGAACACGTATCTTTCGGACTCCCATCCACCCTCGTCGTACCGCCATCGG |
| Nipponbare | CACCTTCTGGAACACGTATCTTTCGGACTCCCATCCACCCTCGTCGTACCGCCATCGG |
| Azucena | CACCTTCTGGAACACGTATCTTTCGGACTCCCATCCACCCTCGTCGTACCGCCATCGG |
| ***** | |
| Bala | CATCCCTCTCATCGGCAAGATCAACATCTAGT CG TCATTAAATTAATTAATTAATATAAT |
| IR64 | CATCCCTCTCATCGGCAAGATCAACATCTAGT CG TCATTAAATTAATTAATTAATATAAT |
| DJ123 | CATCCCTCTCATCGGCAAGATCAACATCTAGT CG TCATTAAATTAATTAATTAATATAAT |
| Nipponbare | CATCCCTCTCATCGGCAAGATCAACATCTAGT CG TCATTAAATTAATTAATTAATATAAT |
| Azucena | CATCCCTCTCATCGGCAAGATCAACATCTAGT CG TCATTAAATTAATTAATTAATATAAT |
| ***** | |
| Bala | CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT |
| IR64 | CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT |
| DJ123 | CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT |
| Nipponbare | CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT |
| Azucena | CCGGCTAGCTACTATCTACAACGATCAGAAGAAGGCAGAATGATCGAGCATGCAAGAATT |
| ***** | |
| Bala | CATGCAAATTACAAGGGATGTGATATGATCATATTAATCCAAGGAAGCTAGTATGATGAT |
| IR64 | CATGCAAATTACAAGGGATGTGATATGATCATATTAATCCAAGGAAGCTAGTATGATGAT |
| DJ123 | CATGCAAATTACAAGGGATGTGATATGATCATATTAATCCAAGGAAGCTAGTATGATGAT |
| Nipponbare | CATGCAAATTACAAGGGATGTGATATGATCATATTAATCCAAGGAAGCTAGTATGATGAT |
| Azucena | CATGCAAATTACAAGGGATGTGATATGATCATATTAATCCAAGGAAGCTAGTATGATGAT |
| ***** | |

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|------------|---|
| Bala | TAAGTTTGTGATCAAGCCGTACT GT TGTGCATGCCAATGCATGCATGTGTTCAAGTGATACA |
| IR64 | TAAGTTTGTGATCAAGCCGTACT GT TGTGCATGCCAATGCATGCATGTGTTCAAGTGATACA |
| DJ123 | TAAGTTTGTGATCAAGCCGTACT GT TGTGCATGCCAATGCATGCATGTGTTCAAGTGATACA |
| Nipponbare | TAAGTTTGTGATCAAGCCGTACT GT TGTGCATGCCAATGCATGCATGTGTTCAAGTGATACA |
| Azucena | TAAGTTTGTGATCAAGCCGTACT GT TGTGCATGCCAATGCATGCATGTGTTCAAGTGATACA |
| ***** | |
| Bala | CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCCATGCTCTGGGATAAATTAGTTAA |
| IR64 | CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCCATGCTCTGGGATAAATTAGTTAA |
| DJ123 | CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCCATGCTCTGGGATAAATTAGTTAA |
| Nipponbare | CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCCATGCTCTGGGATAAATTAGTTAA |
| Azucena | CTATGCTTGGTCTCAAGTGGATCAATTATATTGGTGCCATGCTCTGGGATAAATTAGTTAA |
| ***** | |
| Bala | TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAAGGCA |
| IR64 | TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAAGGCA |
| DJ123 | TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAAGGCA |
| Nipponbare | TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAAGGCA |
| Azucena | TTAGTACTTATGATAAGCTAGAGAGGAGGATGCATTATATTGGTGCATGGTAATAAAGGCA |
| ***** | |
| Bala | AATGAACAGCAGAATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT |
| IR64 | AATGAACAGCAGAATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT |
| DJ123 | AATGAACAGCAGAATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT |
| Nipponbare | AATGAACAGCAGAATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT |
| Azucena | AATGAACAGCAGAATGCATGTATATATGTACTGTATCCCTGTTGCCAGCTGGTCTCTGTT |
| ***** | |
| Bala | TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC |
| IR64 | TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC |
| DJ123 | TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC |
| Nipponbare | TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC |
| Azucena | TGAATGAAGTTGATAGCAACAAAGCTAGCACTAGCGAATGAGCTAGTGACTCAAGTACGC |
| ***** | |
| Bala | ATGCCGGATTACCTCATTGCTCCAAACTTGCAATCAGTTTTGAACTTTAAATAAAGTTAA |
| IR64 | ATGCCGGATTACCTCATTGCTCCAAACTTGCAATCAGTTTTGAACTTTAAATAAAGTTAA |
| DJ123 | ATGCCGGATTACCTCATTGCTCCAAACTTGCAATCAGTTTTGAACTTTAAATAAAGTTAA |
| Nipponbare | ATGCCGGATTACCTCATTGCTCCAAACTTGCAATCAGTTTTGAACTTTAAATAAAGTTAA |
| Azucena | ATGCCGGATTACCTCATTGCTCCAAACTTGCAATCAGTTTTGAACTTTAAATAAAGTTAA |
| ***** | |
| Bala | TTTGGCTTTGTGCACCTTGTCATTTGCTCCCTCTTTGTTTAAACCGATCTTGCCTAGTGCG |
| IR64 | TTTGGCTTTGTGCACCTTGTCATTTGCTCCCTCTTTGTTTAAACCGATCTTGCCTAGTGCG |
| DJ123 | TTTGGCTTTGTGCACCTTGTCATTTGCTCCCTCTTTGTTTAAACCGATCTTGCCTAGTGCG |
| Nipponbare | TTTGGCTTTGTGCACCTTGTCATTTGCTCCCTCTTTGTTTAAACCGATCTTGCCTAGTGCG |
| Azucena | TTTGGCTTTGTGCACCTTGTCATTTGCTCCCTCTTTGTTTAAACCGATCTTGCCTAGTGCG |
| ***** | |
| Bala | TACGTAGCTAGATTGCACATTTAAG |
| IR64 | TACGTAGCTAGATTGCACATTTAAG |
| DJ123 | TACGTAGCTAGATTGCACATTTAAG |
| Nipponbare | TACGTAGCTAGATTGCACATTTAAG |
| Azucena | TACGTAGCTAGATTGCACATTTAAG |
| ***** | |

Supplementary figure 4. Genomic sequence alignment of *Lsi3*. Regions highlighted in bold are predicted exon regions based on the Rice Genome Annotation Project – release 7 sequence available at <http://rice.plantbiology.msu.edu/>.