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## 30        **Abstract**

31    *In vivo* estimates of tibiotalar and the subtalar joint kinematics can unveil unique information about gait  
32    biomechanics, especially in the presence of musculoskeletal disorders affecting the foot and ankle complex.  
33    Previous literature investigated the ankle kinematics on *ex vivo* data sets, but little has been reported for  
34    natural walking, and even less for pathological and juvenile populations. This paper proposes an MRI-based  
35    morphological fitting methodology for the personalised definition of the tibiotalar and the subtalar joint  
36    axes during gait, and investigated its application to characterise the ankle kinematics in twenty patients  
37    affected by Juvenile Idiopathic Arthritis (JIA). The estimated joint axes were in line with *in vivo* and *ex*  
38    *vivo* literature data and joint kinematics variation subsequent to inter-operator variability was in the order  
39    of 1°. The model allowed to investigate, for the first time in patients with JIA, the functional response to  
40    joint impairment. The joint kinematics highlighted changes over time that were consistent with changes in  
41    the patient's clinical pattern and notably varied from patient to patient. The heterogeneous and patient-  
42    specific nature of the effects of JIA was confirmed by the absence of a correlation between a semi-  
43    quantitative MRI-based impairment score and a variety of investigated joint kinematics indexes. In  
44    conclusion, this study showed the feasibility of using MRI and morphological fitting to identify the  
45    tibiotalar and subtalar joint axes in a non-invasive patient-specific manner. The proposed methodology  
46    represents an innovative and reliable approach to the analysis of the ankle joint kinematics in pathological  
47    juvenile populations.

48

49    **Key words:** Biomechanics, Ankle joint axis, Musculoskeletal modelling, Gait analysis, Patient-specific  
50    modelling

51

## 52        **Introduction**

53    Functional anatomy literature describes the ankle joint as a very complex structure allowing for multiple  
54    movements due to the combination of various mechanically coupled joints, including the tibiotalar (i.e.  
55    between tibia and talus) and subtalar (i.e. between talus and calcaneus) joints (Hicks et al., 1953; Siegler et  
56    al., 1988; Dettwyler et al., 2004). The biomechanical behaviour of the ankle during locomotion and its  
57    relationship with the anatomy have been investigated since the beginning of the last century (Fick, 1911;  
58    Manter, 1941; Barnett and Napier, 1952; Isman and Inman, 1969; Inman, 1976) and many authors have  
59    also estimated the kinematics of the tibiotalar and subtalar joints *ex vivo* (Hicks et al., 1953; Rasmussen and  
60    Tovborg-Jensen, 1982; van Langelaan, 1983; Siegler et al., 1988). The possibility of estimating the  
61    kinematics of the ankle's intrinsic joints from *in vivo* data is of interest when investigating musculoskeletal  
62    diseases. Nonetheless, a comprehensive understanding of the joint's intrinsic movement during walking is  
63    still lacking. This is because measuring the motion associated to foot inversion/eversion is not trivial and  
64    most literature has focused on the quantification of articular range of motion (ROM) for the various joint's  
65    degrees of freedom (DOFs) under controlled conditions (Lundberg et al., 1989; Mattingly et al., 2006;  
66    Lewis et al., 2009).

67    *In vivo* tracking of the relative movement of the talus relative to the calcaneus using skin markers and a  
68    standard gait analysis technique is complicated by the small size of these bones and the absence of visible  
69    superficial landmarks (Scott et al., 1991; Di Marco et al., 2016). Few studies have investigated the  
70    kinematics of the intrinsic joints of the ankle during walking and running (Arndt et al., 2004 and 2006)  
71    using intracortical bone pins, and compared the results to those from using superficial markers (Westblad  
72    et al., 2002). These studies clearly showed a description of plantar/dorsiflexion is possible with traditional  
73    gait analysis methods, however, estimates of inversion/eversion movement are still far from being accurate.  
74    Intracortical pin-based studies partially overcome this lack of accuracy but, due to the invasiveness of the  
75    technique, the number of participants is usually limited to few healthy volunteers, whose natural gait pattern  
76    can be altered by the possible pain and discomfort related to the implant. Both *in vivo* and *ex vivo* studies

77 reported high intra-subject and inter-subject variability in the subtalar joint kinematics with ROM up to 60°  
78 (Roaas and Anderson, 1982; Sepic et al., 1986; Lundberg, 1989).

79 The functional complexity of the subtalar joint led to a number of different modelling approaches, from the  
80 attempt to capture its mobility through multi-segmental foot models where the subtalar articulation was  
81 interpreted as a motion between hind-foot and fore-foot (Prinold et al., 2016; Saraswat et al., 2010), to a  
82 more anatomical representation as a universal or hinge joint (Delp et al., 1990; Malaquias et al., 2017). The  
83 hinge-like schematisation also applies to the tibiotalar joint and this approach is currently used within  
84 widely adopted musculoskeletal models (Delp et al., 1990). When simultaneously modelling both joints as  
85 hinges (Dul and Johnson, 1985), a reasonable simplification is made with respect to their real functional  
86 role (Siegler et al., 1988), according to which the tibiotalar and subtalar joints describe  
87 the plantar/dorsiflexion and inversion/eversion motions, respectively. This latter motion, despite its  
88 simplified appearance, is justified because the predominant motion occurs about a single axis of rotation  
89 (Scott and Winter, 1991). However, this DOF has been reported to be less accurately described with current  
90 musculoskeletal modelling approaches, mainly due to the difficulties in identifying the joint functional axis  
91 *in vivo* (Van den Bogert et al., 1994; Dettwyler et al., 2004; Parr et al., 2012). A high variability within-  
92 and between-subjects has been observed in the modelled joint axes, which is also related to the specific  
93 locomotion task (Leitch et al., 2010). In the presence of musculoskeletal disorders, the adoption of image-  
94 based patient-specific modelling approaches has been previously proposed (Prinold et al., 2016; Hannah et  
95 al., 2017) and proved to increase anatomical modelling accuracy (Correa and Pandy 2011; Durkin et al.,  
96 2006; Scheys et al., 2009). The use of this technique accounts for patients' anatomical features and  
97 peculiarities, crucial when impairments and gait limitations affect the subjects. In this study, we propose an  
98 image-based modelling procedure to define the tibiotalar and subtalar joints axes, avoiding operator-  
99 dependent steps and related variability issues (Prinold et al., 2016; Hannah et al., 2017). Once compared  
100 against literature, the procedure will be used as part of a patient-specific musculoskeletal modelling  
101 approach to investigate the gait ankle kinematics in children with Juvenile Idiopathic Arthritis (JIA), a

102 paediatric group of diseases of unknown aetiology characterised by joint inflammation potentially leading  
103 to cartilage damage. Altered gait patterns and physical disabilities (Ravelli and Martini, 2007) are possible  
104 outcomes in JIA. This longitudinal study will prove whether our modelling approach is capable of detecting  
105 clinical changes observed in the tibiotalar and the subtalar joint functions and quantify for the first time the  
106 relationship between these changes and the underlying joint impairments.

## 107 **Methods**

### 108 **2.1 Subjects and data acquisition**

109 Twenty participants (5 males, 15 females, age:  $11.6 \pm 3.1$  years, mass:  $47.6 \pm 18.2$  kg, height:  $148 \pm 17$  cm, 11  
110 new onsets) affected by Juvenile Idiopathic Arthritis (JIA) of various sub-types (oligoarticular onset JIA,  
111 polyarticular JIA, psoriatic arthritis, and undifferentiated arthritis) (Ravelli and Martini, 2007) were  
112 recruited among those referred to two different children's hospitals (Istituto Giannina Gaslini, Genoa (Lab  
113 1), and "Bambino Gesù" Children's Hospital, Rome (Lab 2)). The study was conducted following  
114 Helsinki's declaration on human rights and was approved by the ethical committee of both hospitals.  
115 Written informed consent was obtained by patients' parents.

116 Medical resonance images (MRI) and gait analysis data were collected at three time-points (6 months apart)  
117 to follow the disease progression. The imaging performed at month 0 (M0) and month 12 (M12) included  
118 a foot and ankle regional MRI (multi-slice multi-echo 3D Gradient Echo (mFFE) with water-only selection  
119 (WATS) with 0.5 mm in-plane resolution and 1 mm slice thickness). The month 6 (M6) imaging included  
120 a full lower limb MRI (3D T1-weighted fat-suppression sequence (e-THRIVE) with 1mm in-plane  
121 resolution and 1mm slice thickness). The core set of basic sequences and definitions suggested by the  
122 Outcome Measure in Rheumatology (OMERACT) MRI Working Group (Ostergaard et al., 2003; Nusman  
123 et al., 2016) was used to provide an MRI-based evaluation of the joints (Table I). A weighted, average index  
124 ( $I_{MRI}$ ) was used to quantify the overall level of impairment of the foot and ankle region.

Table I - MRI scoring.

| Index                   | MRI sequence                               | Scale  | Sites   |
|-------------------------|--|--|---|
| <b>Bone erosion</b>     | T1-weighted fat-saturated                  | Range 0-10<br>% of eroded articular surface ( <i>Ostergaard et al., 2003</i> )<br>0 = no erosion;<br>1 = 1–10%; 2 = 11–20%;<br>3 = 21–30%; 4 = 31–40%;<br>5 = 41–50%; 6 = 51–60%;<br>7 = 61–70%; 8 = 61–80%;<br>9 = 81–90%; 10 = 91–100% | Distal tibial epiphysis<br>Distal fibula epiphysis<br>Tarsal bones<br>Metatarsal bases  |
| <b>Cartilage damage</b> | WATS                                       | Range 0-3<br>% of damaged cartilage surface<br>0 = no damage;<br>1 = 1–33%;<br>2 = 34–66%;<br>3 = 67–100%;<br>4 = extensive damage causing ankyloses   | Tibiotalar<br>Between distal talus and calcaneus,<br>Talonavicular<br>Calcaneocuboid<br>Cuneonavicular<br>Between cuneiforms and I, II and III metatarsal bones<br>Between cuboid and IV and V metatarsal bones |
| <b>Synovitis</b>        | T1-weighted fat-saturated                  | Range 0-3<br>Degree of synovial enhancement and synovial thickness ( <i>Ostergaard et al., 2003; Malattia et al., 2011</i> )<br>0 = normal;<br>1 = mild;<br>2 = moderate;<br>3 = severe  | Tibio-peroneo-talar<br>Subtalar<br>Talonavicular<br>Calcaneocuboid<br>I-V tarsometatarsal<br>Cuneonavicular   |
| <b>Tenosynovitis</b>    | T1-weighted fat-saturated with enhancement | Range 0-3<br>Degree of peritendinous effusion or synovial proliferation<br>0 = normal;<br>1 = mild (< 2 mm);<br>2 = moderate (2 -5 mm);<br>3 = severe (> 5 mm)   | Anterior tibial<br>Extensor digitorum longus<br>Extensor hallucis longus<br>Posterior tibial<br>Flexor digitorum longus<br>Flexor hallucis longus<br>Peroneal tendons   |

126

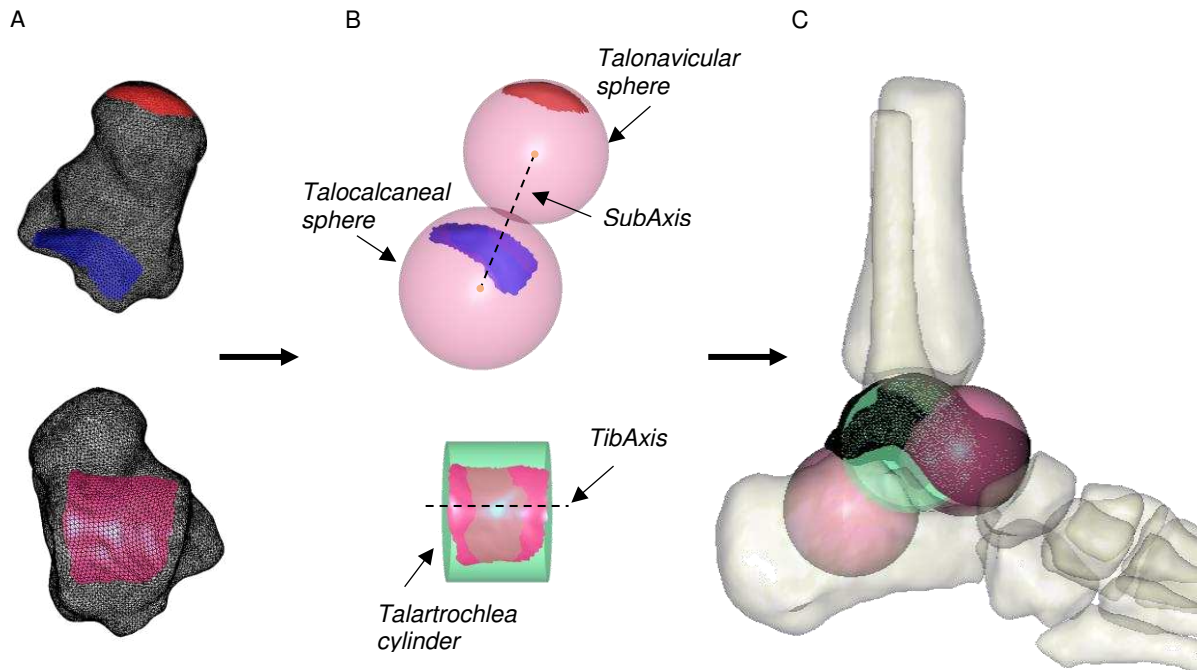
127 Gait analysis was based on stereophotogrammetry and data were collected using a 6-camera system (BTS,  
128 Smart DX, 100Hz) with two force plates (Kistler, 1kHz) in Lab 1, and an 8-camera system (Vicon, MX,  
129 200Hz) and two force plates (AMTI, OR6, 1kHz) in Lab 2. Five walking trials at self-selected speed were  
130 performed and a minimum of three trials were used for the analysis. The marker set included forty-four  
131 markers from the Vicon Plug in gait protocol (Vicon Motion System) and the modified Oxford Foot Model  
132 (mOFM) protocol (Stebbins et al., 2006). A subset of MRI-visible markers (twenty-eight in the lower limb  
133 MRI and six in the regional MRI scans) was retained during the imaging acquisition for data registration.

134 Despite being collected in different centres and with different equipment, the raw-data underwent the same  
135 pre-processing in terms of labelling, gap-filling (spline algorithm built in Vicon Nexus 1.8.5 (Woltring et  
136 al., 1986)), and smoothing (4th-order Butterworth filter, 6Hz cut-off (Barlett et al., 2007)).

## 137 **2.2 Anatomical model**

138 A statistical shape modelling approach (Steger et al., 2012) was used to segment the lower limb bones from  
139 the MRI and subject-specific anatomical models were produced using specialised software (NMSBuilder,  
140 Valente et al., 2017). For each patient, two bilateral three-segment anatomical models were built using the  
141 M0 and M12 datasets, resulting in 80 foot models. Twelve of these were excluded due to incompleteness  
142 of the experimental dataset, resulting in a final dataset of 68 feet. The joints' reference frames, namely  
143 tibiotalar joint (between tibia and talus) and subtalar (between talus and foot) were defined according to the  
144 ISB conventions (Baker et al., 2003) and the joint axes were identified through morphological fitting of  
145 articular surfaces (Figure 1A-C). The subtalar joint axis (*SubAxis*) was defined as the axis connecting the  
146 centres of the spheres fitted to the anterior (Talonavicular sphere) and to the posterior-inferior  
147 (*Talocalcaneal sphere*) facets of the talus respectively (Figure 1B). This was similar to that proposed by  
148 Parr et al., 2012, who, however, used the anterior-inferior portion of the talus surface to define the  
149 Talonavicular sphere. To define the tibiotalar joint axis (*TibAxis*), a cylinder was fitted to the entire trochlea  
150 (*Talar trochlea cylinder*) as a simplification of the approach proposed by Siegler et al., 2014 (Modenese et  
151 al., 2018). The fitting was implemented in Meshlab (Cignoni et al., 2008) by identifying the articular  
152 surfaces from the segmented geometries and minimising the least squares distance between the identified  
153 surface and the corresponding best fitting analytical shape (Least Squares Geometric Elements library,  
154 Matlab). The distal tibia (segmented from the M0/12 MRI) was afterwards registered to the entire tibia (M6  
155 dataset) using the Iterative Closest Point algorithm in Meshlab to obtain a full lower limb model. A  
156 comprehensive description of the modelling procedure is available as supplementary material in Modenese  
157 et al. (2018). The data and models presented in this paper are available on Figshare (doi:  
158 <https://doi.org/10.15131/shef.data.5863443.v1>).





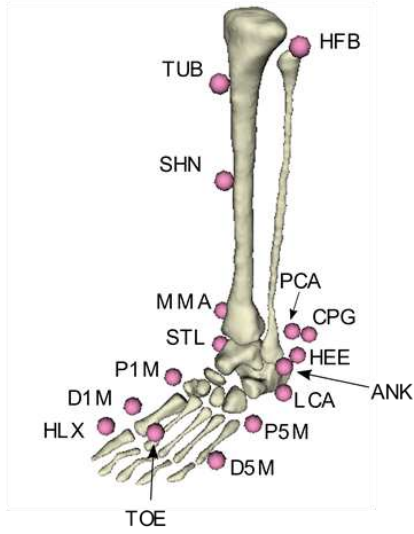
159

160 **Figure 1 - (A) Plantar (top) and dorsal (bottom) views of the right talus (black wireframe) with**  
 161 **highlighted articular regions: anterior facet (red), posterior-inferior facet (blue), trochlea (fuchsia). (B)**  
 162 **Fitting of analytical shapes to the selected articular regions: two spheres (light pink) identify the axis of**  
 163 **the subtalar joint (SubAxis) as the axis connecting the centres of the spheres and a cylinder (light green)**  
 164 **identifies the axis of the tibiotalar joint (TibAxis) as the cylinder axis. (C) Example of the fitted**  
 165 **geometries integrated within the ankle anatomical model.**

166

167 **2.3 Joint kinematics**

168 The OpenSim's (Delp et al., 2007) Inverse Kinematics (IK) tool was run to estimate the tibiotalar and  
 169 subtalar joint angles starting from a set of sixteen skin markers (five on the tibia, eleven on the foot, Figure  
 170 2), eight were also virtually palpated on the medical images. The difference between the virtual and  
 171 experimental markers estimated by the IK tool was less than 1cm on average over all the time-steps, as  
 172 suggested in the OpenSim best practice recommendations (Hicks et al., 2015).



| Label | Description  | Markers |        |
|-------|--|---------|--------|
|       |  | MRI     | Stereo |
| HFB   | Head of the fibula   | Yes     | Yes    |
| SHN   | Anterior aspect of shin  | Yes     | Yes    |
| TUB   | Tibial tuberosity  | -       | Yes    |
| MMA   | Medial malleolus   | Yes     | Yes    |
| ANK   | Lateral malleolus  | Yes     | Yes    |
| PCA   | Posterior medial aspect of heel  | -       | Yes    |
| STL   | Sustentaculum tali   | -       | Yes    |
| LCA   | Lateral calcaneus  | -       | Yes    |
| CPG   | Wand marker on posterior calcaneus aligned with transverse orientation | -       | Yes    |
| HEE   | Posterior distal aspect of heel  | -       | Yes    |
| P1M   | Lateral aspect of 1 <sup>st</sup> metatarsal base                      | -       | Yes    |
| P5M   | Lateral aspect of 5 <sup>th</sup> metatarsal base                      | -       | Yes    |
| TOE   | Between 2 <sup>nd</sup> and 3 <sup>rd</sup> metatarsal heads           | Yes     | Yes    |
| D1M   | Lateral aspect of 1 <sup>st</sup> metatarsal head                      | Yes     | Yes    |
| D5M   | Lateral aspect of 5 <sup>th</sup> metatarsal head                      | Yes     | Yes    |
| HLX   | Medial side of the proximal hallux                                     | Yes     | Yes    |

173

174 **Figure 2 - Experimental markers used in the imaging (MRI) and stereo-photogrammetric (Stereo)**  
 175 **measurements.**

176 **2.4 Model evaluation**

177 **Sensitivity to operator-dependent input**

178 The bone segmentations from three randomly chosen patients were used to investigate the effect of  
 179 operator-dependent variability in the definition of *TibAxis* and *SubAxis*. Three operators repeated the  
 180 morphological fitting three times and the coordinates of the *Talartrochlea cylinder*, *Talocalcaneal sphere*  
 181 and *Talonavicular sphere* centres were used for the comparison. A 3D quantification of their variability  
 182 ( $SD_{3d}$ ) was calculated from the standard deviation of the point coordinates ( $sd_x, sd_y, sd_z$ ) as:

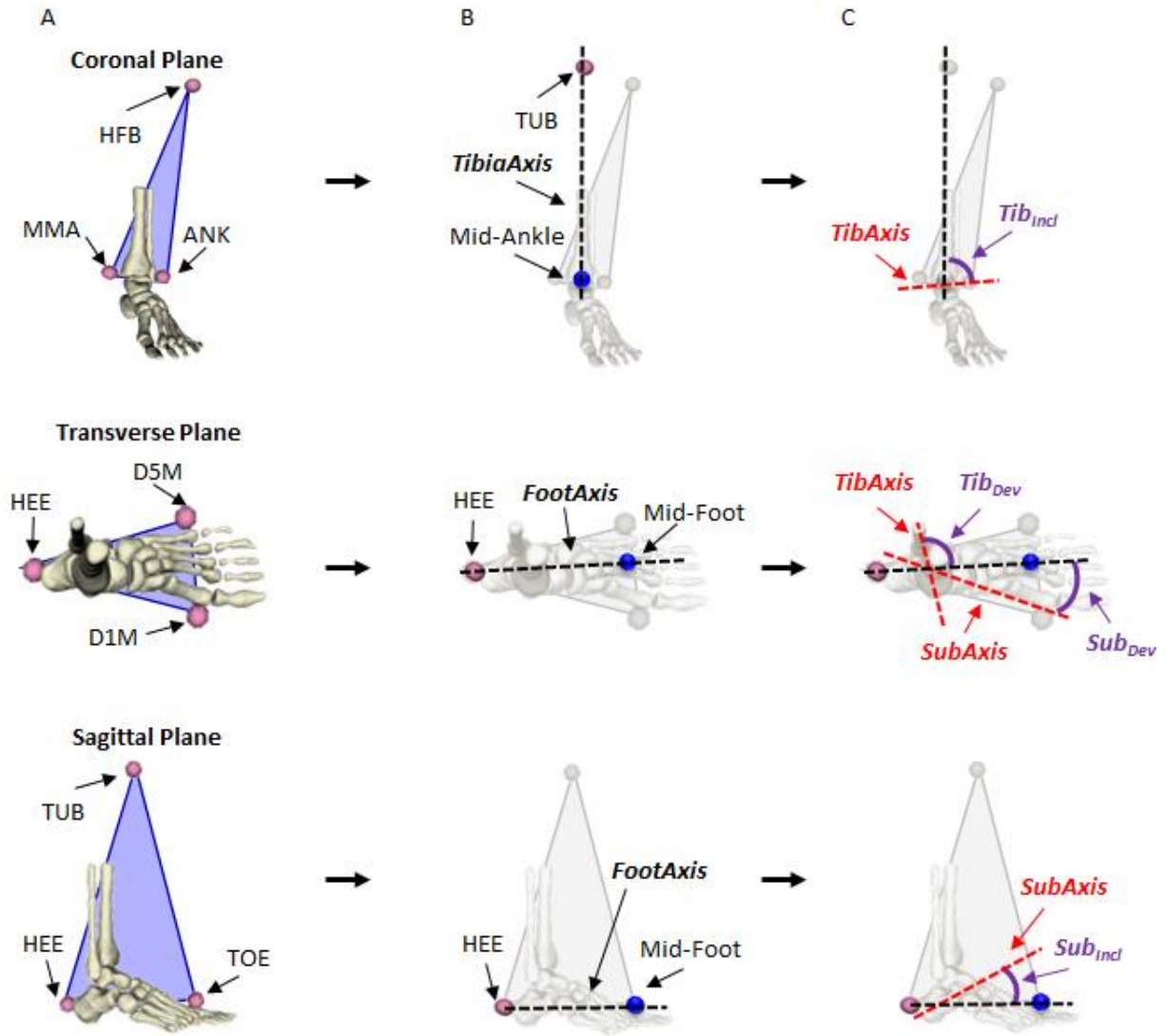
183 
$$SD_{3d} = \sqrt{sd_x^2 + sd_y^2 + sd_z^2}$$

184 For the foot that led to the worst-case scenario (higher inter-operator  $SD_{3d}$ ), a second level of analysis was  
 185 conducted to quantify the propagation of this error on the joint kinematics. The nine models built by the  
 186 three operators were then used to estimate the tibiotalar and subtalar joint kinematics using data from one

187 randomly selected gait trial from the same patient. The maximum value of the mean and standard deviation  
188 calculated over the nine repetitions for each point of the gait cycle was then used to quantify the maximum  
189 expected error.

## 190 **Consistency with literature data**

191 Among the 68 available models, 38 were selected (19 per side, preferentially from M12) to conduct the  
192 following analysis. A standing trial collected during the gait analysis session was used to identify the pose  
193 of each subject and the resulting neutral position of the foot. The transverse, sagittal, and coronal anatomical  
194 planes, the midline of the foot (*FootAxis*) and the long axis of tibia (*TibiaAxis*) were identified using the  
195 standing trial markers (Figure 3A-B). These allowed quantifying the tibiotalar inclination (*TibIncl*) and  
196 deviation (*TibDev*), and the subtalar inclination (*SubIncl*) and deviation (*SubDev*) as shown by the angles in  
197 Figure 3C. *TibIncl*, *TibDev*, *SubIncl* and *SubDev* were compared to literature data from *ex vivo* cadaveric  
198 specimens (Isman and Inman., 1969; Inman, 1976) and from healthy adults (Van den Bogert et al., 1994).  
199 The estimations of *TibAxis* and *SubAxis* at M0 and M12 were also compared. All 26 models for which the  
200 3D anatomy was available at both time-points (52 models) were used for a between-session comparison.  
201 For this analysis, the angle between the two joint axes (*InterAxis*) was preferred over the measures of *TibIncl*,  
202 *TibDev*, *SubIncl*, and *SubDev* to avoid the effect of experimental markers repositioning (between the two  
203 sessions) on these angles. Mean and maximum between-session variations were quantified, and a paired-  
204 two-sided Wilcoxon signed-rank test ( $\alpha=0.05$ ) was performed under the null hypothesis showed that no  
205 statistical difference existed between the two repeated measures. This was intended as a repeatability  
206 assessment of the proposed method, assuming in the investigated age range, and within 12 months, neither  
207 disease progression (Ravelli and Martini, 2007) nor growth (Evans, 2010) would cause changes in the joint  
208 morphology.



209  
210

211 **Figure 3 - (A) Identification of anatomical planes (blue triangles) as defined using the virtual markers**  
 212 **(pink) corresponding to the experimental markers listed in Figure 2. (B) Definition of the anatomical**  
 213 **axes (midline of the foot = FootAxis, long axis of the tibia = TibiaAxis, black dashed lines) by calculating**  
 214 **average points (blue markers) between virtual marker pairs (Mid-Foot = midpoint between D1M and**  
 215 **D5M; Mid-Ankle = midpoint between ANK and MMA). (C) Quantification of the inclination (TibIncl)**  
 216 **and deviation (TibDev) of tibiotalar joint and inclination (SubIncl) and deviation (SubDev) of subtalar**  
 217 **joint as the angles (purple arches) between the anatomical axes and the joint axes (red dashed lines) as**  
 218 **defined through morphological fitting (Figure 1).**

219 **Effect of clinical impairment on joint kinematics**

220 The models from 13 subjects (3 males, 10 females, age:  $11.0 \pm 3.1$  years, mass:  $44.5 \pm 16.9$  kg, height:  $143$   
 221  $\pm 13$  cm, 8 new onsets), for whom both clinical and biomechanical information was available, were used to

222 test the link between changes in the kinematics and impairment of the ankle as measured from the MRI.  
223 The  $I_{MRI}$  scores were used to classify the disability level of each ankle and identify better and worse time-  
224 points. They were then placed into “low-involvement” and “high-involvement” groups accordingly. The  
225 joint kinematics of the two groups were then compared using a non-parametric 1D two-tailed paired  $t$ -test  
226 ( $\alpha=0.05$ ) (Nichols and Holmes, 2002) based on Statistical Parametric Mapping (SPM) in MATLAB (v9.1,  
227 R2016b, Mathworks, USA), using the SPM1D package (Pataky et al., 2012). This was chosen since the  
228 data were not normally distributed. The following kinematic parameters were also calculated to investigate  
229 the correlation with the  $I_{MRI}$ : area under the curves of the tibiotalar and subtalar joint angles, maximum  
230 plantarflexion (PF) and dorsiflexion (DF) angles, maximum inversion (Inv) and eversion (Ev) angles, and  
231 joint ROM. Furthermore, the asymmetry between the left and right foot kinematics was quantified using  
232 the Root Mean Square Deviation (RMSD) and Mean Absolute Variability (MAV) (Di Marco et al., 2018),  
233 as well as the between-side difference of ROM and standard deviations (SD). RMSD, MAV, ROM and SD  
234 were measured at the two time-points and compared using a two-sided Wilcoxon signed-rank test ( $\alpha=0.05$ ).  
235 The absolute difference ( $\Delta I_{MRI}$ ) between left and right  $I_{MRI}$  was also calculated and a correlation analysis  
236 was used to assess whether an asymmetry in the clinical score, namely higher  $\Delta I_{MRI}$ , corresponded to higher  
237 values of the kinematic parameters.

## 238 **Results**

### 239 **Sensitivity to operator-dependent input**

240  $SD_{3d}$  of *Talonavicular sphere* and *Talocalcaneal sphere*'s centres are reported in Table II, as well as the  
241 resulting maximum angular variability of the *TibAxis* and *SubAxis*, whose maximum value ( $9.6^\circ$ ) was found  
242 for the inclination of *SubAxis* in patient P3. For this patient, the propagation of inter-operator variability on  
243 the articular kinematics introduced a maximum standard deviation of  $0.6^\circ$  and  $1.3^\circ$  for the tibiotalar and  
244 subtalar joints respectively, both occurring at 63% of the gait cycle.

245

**Table II – Inter-operator standard deviation (SD) of fitted surfaces centres and axes.**

|          | <i>Talartrochlea center</i> | <i>Talonavicular center</i> | <i>Talocalcaneal center</i> | <i>TibAxis</i> | <i>SubAxis</i> |
|----------|-----------------------------|-----------------------------|-----------------------------|----------------|----------------|
| Patients | $SD_{3d}$ [mm]              | $SD_{3d}$ [mm]              | $SD_{3d}$ [mm]              | SD [°]         | SD [°]         |
| P1       | 0.4                         | 0.4                         | 1.4                         | 0.6            | 1.7            |
| P2       | 0.5                         | 0.8                         | 1.5                         | 0.8            | 1.3            |
| P3       | 0.8                         | 2.1                         | 5.1                         | 2.0            | 5.6            |

246

247 **Consistency with literature data**

248 The residual error of the fitting algorithm (average ( $\pm$ SD) across the 52 models) was equal to 0.16 ( $\pm$ 0.05)  
 249 mm, 0.48 ( $\pm$ 0.21) mm, and 0.28 ( $\pm$ 0.11) mm for the *Talonavicular*, *Talocalcaneal*, and *Talartrochlea*  
 250 surfaces, respectively. The average ( $\pm$ SD) values of the measured foot angles (*Tib<sub>Incl</sub>*, *Tib<sub>Dev</sub>*, *Sub<sub>Incl</sub>*, and  
 251 *Sub<sub>Dev</sub>*) (Table III) were found to be in line with the corresponding *ex vivo* (Isman and Inman., 1969; Inman,  
 252 1976) and *in vivo* (Van den Bogert et al., 1994) measurements available in the literature. The average  
 253 absolute difference between the M0 and M12 measures of *InterAxes* was  $2.2^\circ \pm 2.1^\circ$ , which was not  
 254 statistically significant (Wilcoxon test  $p=0.648$ ).

255

**Table III - Inclination and deviation of tibiotalar and subtalar joint axes and comparison with published literature datasets (n = numebr of subjects).**

256

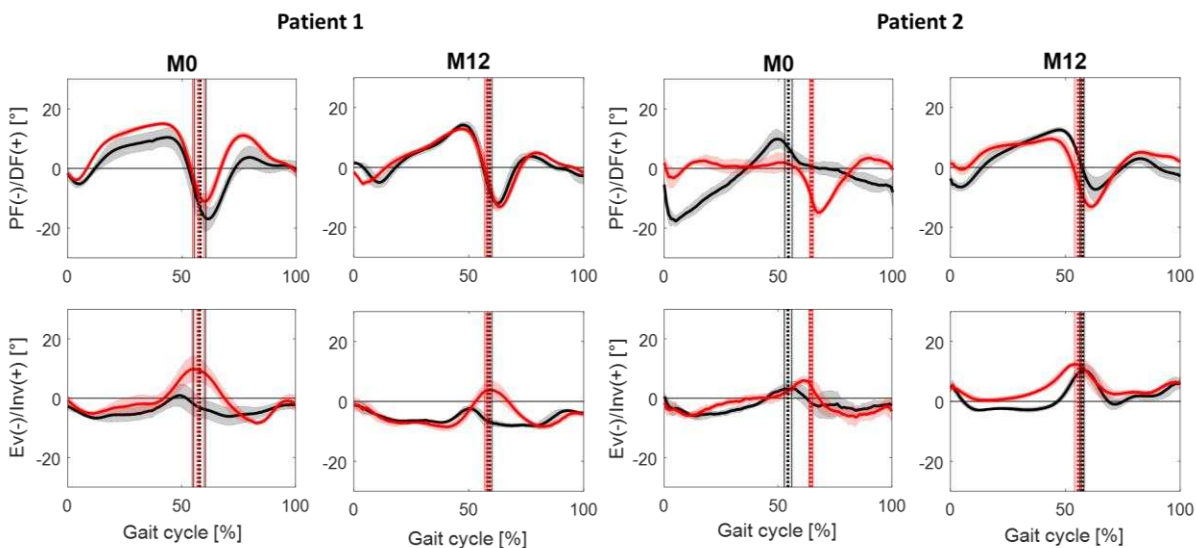
| Angle          | Isman and Inman, 1969<br>(n=46)<br>mean ( $\pm$ SD) [°] | Inman, 1976<br>(n=104)<br>mean ( $\pm$ SD) [°] | Van den Bogert, 1994<br>(n=14)<br>mean ( $\pm$ SD) [°] | This study<br>(n=38)<br>mean ( $\pm$ SD) [°] |
|----------------|---|--|--|--|
| Gender         | NA  | NA   | males  | 30 females/8 males                           |
| Age            | Adults (age not specified)                              | Adults (age not specified)                     | Adults (age not specified)                             | 11.2 $\pm$ 3.1 years                         |
| <i>TibIncl</i> | 80( $\pm$ 4)  | 82.7( $\pm$ 3.7) (n=107)                       | 85.4( $\pm$ 7.4)                                       | 90.7( $\pm$ 4.1)                             |
| <i>TibDev</i>  | 84( $\pm$ 7)  | -  | 89.0( $\pm$ 15.1)                                      | 82.7( $\pm$ 7.4)                             |
| <i>SubIncl</i> | 41( $\pm$ 9)  | 42( $\pm$ 9)                                   | 35.3( $\pm$ 4.8)                                       | 41.1( $\pm$ 14.1)                            |
| <i>SubDev</i>  | 23( $\pm$ 11)   | 23( $\pm$ 11)                                  | 18.0( $\pm$ 16.2)                                      | 27.0( $\pm$ 9.0)                             |

257

258 **Effect of clinical impairment on joint kinematics**

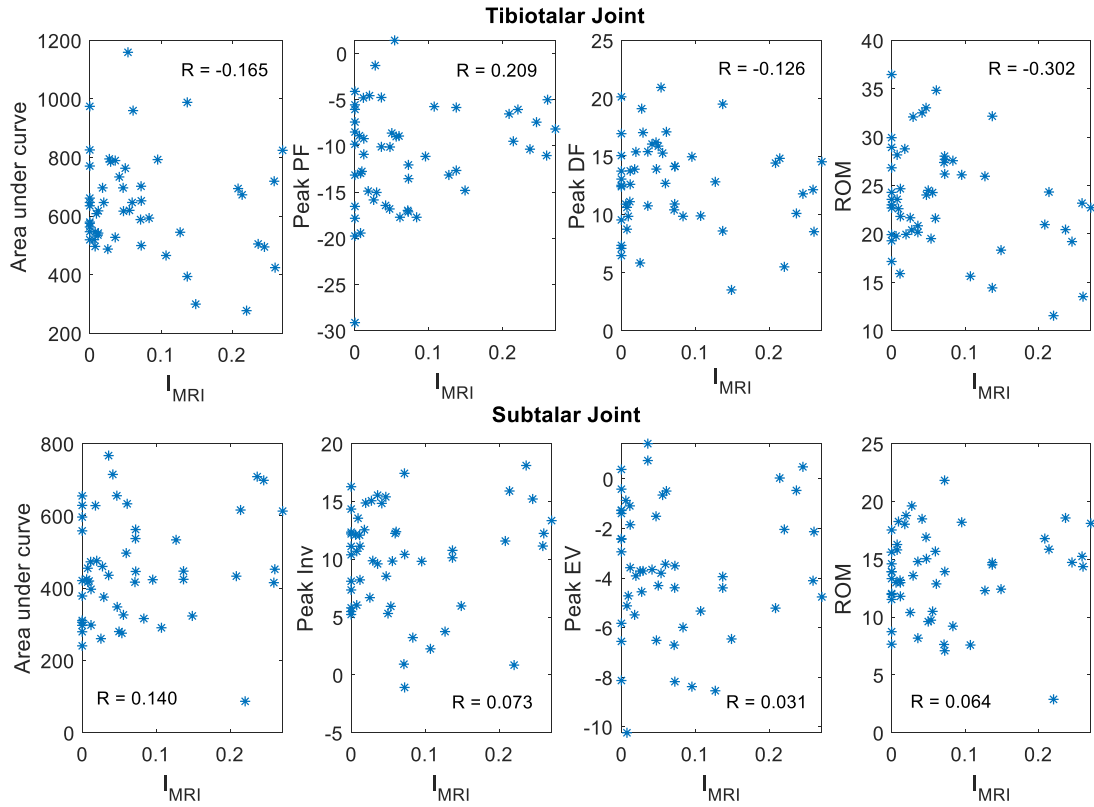
259 Figure 4 shows the estimated kinematics of two subjects with different clinical scoring: patient 1 was  
260 similarly affected by the pathology at the two observations, whereas at M12 patient 2 was in total remission,  
261 as defined by Ravelli and Martini (2007). This example highlights how the models clearly capture different  
262 kinematic patterns associated with different paths of disease progression. The observation of the joint angles  
263 also clearly indicates the ability of the model to describe changes in the gait patterns happening between  
264 the two time-points, which were also confirmed by consistent changes in the walking speed ( $1.51 \pm 0.05$  m/s  
265 at M0 and  $1.22 \pm 0.05$  m/s at M12 for subject 1;  $0.83 \pm 0.03$  m/s at M0 and  $1.20 \pm 0.04$  m/s at M12 for subject  
266 2). For the whole cohort, walking speed varied from  $1.01 \pm 0.24$  m/s at M0 to  $1.12 \pm 0.13$  m/s at M12, and was  
267  $1.14 \pm 0.17$  m/s and  $0.93 \pm 0.33$  m/s at the “low-involvement” and “high-involvement” time-points  
268 respectively, with no significant difference. Walking speed values did not correlate with the joint  
269 impairment level, as measured with the  $I_{MRI}$  ( $R = -0.21$  and  $R = 0.16$  at M0 and M12, respectively). Similarly,  
270 no correlation was observed between  $I_{MRI}$  and the kinematic parameters (Figure 5). This was confirmed by  
271 the absence of a group-wise statistically significant difference between the joint kinematics of the ankles at  
272 the “low-involvement” and “high-involvement” time-points throughout the gait cycle (Figure 6). Figure 7  
273 clearly shows the absence of a significant correspondence between the asymmetry of impairment ( $\Delta I_{MRI}$ )  
274 and the RMSD, MAV,  $\Delta ROM$  and  $\Delta SD$  observed at M0 and M12. However, a smaller  $\Delta I_{MRI}$  at M12 was

275 generally associated to a smaller value of the kinematics indices at that time-point, except for the  $\Delta$ SD of  
276 the tibiotalar joint and the  $\Delta$ ROM of the subtalar joint.



277 **Figure 4 - Tibiotalar (PF/DF) and subtalar (Ev/Inv) joints kinematics for two JIA patients at M0 and**  
278 **M12. Average right (left) kinematics is shown with black (red) solid line with shadow representing  $\pm 1$**   
279 **standard deviation. Toe off is shown with dotted vertical lines  $\pm 1$  standard deviation (solid vertical lines).**  
280 **Walking speed changed from  $1.51 \pm 0.05$  m/s at M0 to  $1.22 \pm 0.05$  m/s at M12 for patient 1 and from  $0.83$**   
281  **$\pm 0.03$  m/s at M0 and  $1.20 \pm 0.04$  m/s at M12 for patient 2.**





282

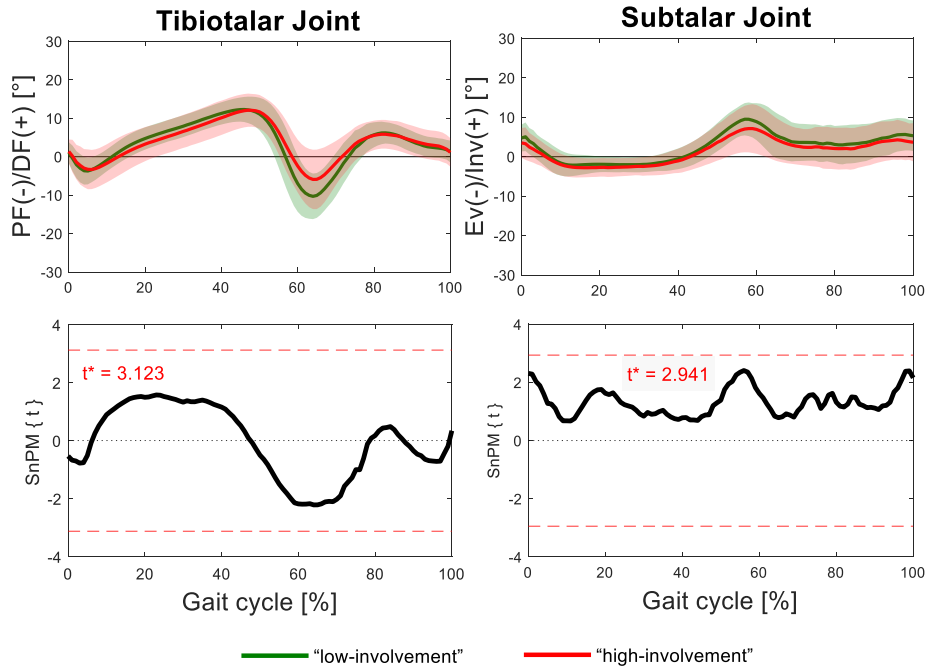
283 **Figure 5 - Correlation between joint impairment level ( $I_{MRI}$ ) and joint kinematics parameters (area**  
 284 **under the curve, peak of plantarflexion (Peak PF) and dorsiflexion (Peak DF), peak of Inversion (Peak**  
 285 **Inv) and eversion (Peak Ev), ROM) for all feet and observations.**

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287

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289

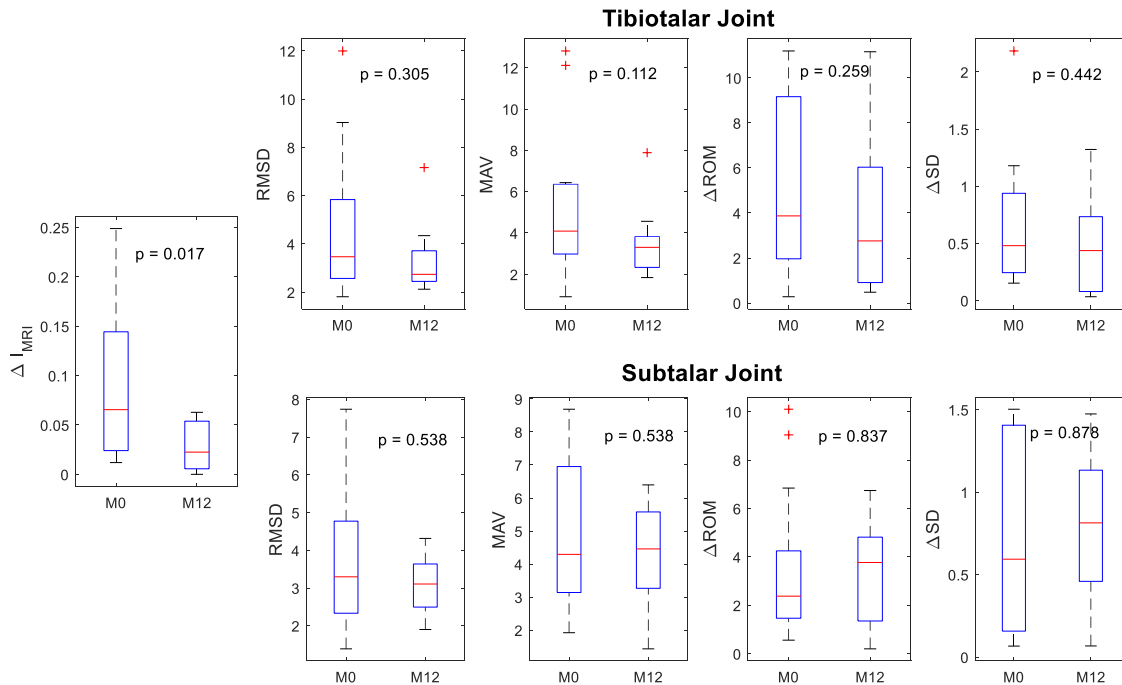


290

291 **Figure 6 - Tibiotalar (PF/DF) and subtalar (Ev/Inv) joint kinematics of the 13 subjects as calculated at**  
 292 **the “low-involvement” (green) and “high-involvement” (red) time-point. Solid lines in the left graphs**  
 293 **represent mean values and bands represent  $\pm 1$  standard deviation. The right figures show the**  
 294 **corresponding distribution of  $t$ -values (SnPM{t}) throughout the gait cycle as obtained from the non-**  
 295 **parametric 1D paired  $t$ -test (Nichols and Holmes, 2002), calculated using the SPM1D package (Pataky**  
 296 **et al., 2012). Each group includes 24 mono-lateral models (2 models were excluded from the analysis).**

297

298



299

300 **Figure 7 – Boxplot distribution of  $\Delta I_{MRI}$  and kinematics indices (RMSD, MAV,  $\Delta ROM$  and  $\Delta SD$ ) for**  
 301 **both tibiotalar and subtalar joints (n=13) at M0 and M12.  $p$ -values from two-sided Wilcoxon signed-**  
 302 **rank test are reported in each plot. Data outliers are marked with a +.**

## 303 Discussion

304 The aim of the study was to propose a kinematic model of the tibiotalar and subtalar joints, and to use this  
 305 model to investigate the ankle joint kinematics in a group of children with JIA. The anatomical model was  
 306 based on a morphological fitting approach and underwent repeatability analysis.

307 The procedure proved to be robust to the operator-dependent input. Even in the worst-case scenario, where  
 308 the definition of the subtalar axis was associated with high inter-operator error ( $9.6^\circ$ ), the joint kinematics  
 309 varied less than  $1.3^\circ$ . The inter-operator variability was mainly associated with the quality of the segmented  
 310 images, i.e. low resolution, bias field or noise in the MRI, and to the complexity of segmenting bone tissue  
 311 in young subjects, where cortical bone is not completely ossified (Evans, 2010). Nonetheless, this error was  
 312 still acceptable when compared to other possible sources of variability coming from the experimental errors,  
 313 such as instrumental error and marker placement error (up to  $6^\circ \pm 2^\circ$  at the toe off (Di Marco et al., 2016)),

314 or soft tissue artefact (up to 20% of variability in the ankle kinematics (Lamberto et al., 2016)), confirming  
315 the chosen morphological fitting approach is suitable in the presence of low quality images and/or poor  
316 bone reconstructions.

317 An *in vivo* validation of the proposed technique was not possible within the framework of this project due  
318 to ethics constraint in the use of approaches like dual-fluoroscopy in a paediatric population. However, the  
319 comparison with *ex vivo* (Isman and Inman, 1969; Inman, 1976), and *in vivo* (Van den Bogert et al., 1994)  
320 data certainly support the validity of the technique. Previous studies (Leitch et al., 2010; Van den Bogert et  
321 al., 1994) reported the highest between-subject variabilities in the deviation angle (up to 15°); conversely,  
322 we found the biggest differences in the inclination of the subtalar axis (14 °). This could be ascribed to the  
323 subtalar axis' definition relying on the identification of the anterior facet of the talus. In the youngest  
324 children, in fact, this surface can present a layer of unossified cartilage (Evans et al., 2010), which can  
325 complicate the identification of the bone contour in the MRI, consequently affecting the results of  
326 segmentation and morphological fitting.

327 The second goal of the study involved the application of the modelling approach as part of the clinical gait  
328 assessment of patients with JIA. The between-session repeatability showed no statistically significant  
329 difference between the measures of *InterAxis* at M0 and M12, confirming our hypothesis.

330 The observed joint kinematics reflected the heterogeneous and patient-specific nature of the pathology,  
331 which presents several sub-types, each with a specific progression (Ravelli and Martini, 2007). In fact, the  
332 individual differences (Figure 4) were not representative of a group behaviour (Figure 6) as a consequence  
333 of different possible evolutions of the disease. The absence of a recognisable group pattern was  
334 demonstrated by the lack of a direct relationship between a joint's clinical impairment and its kinematics.  
335 The inter-subject variability was probably exacerbated by the heterogeneity of the cohort in terms of age,  
336 anthropometry, disease subtype and activity level. This explains the lack of correlation between joint  
337 kinematics (and their changes between time points) and the patient's  $I_{MRI}$  scores. This also held true for  
338 the walking speed, which was not correlated with the MRI scores, but was found in line with the

339 1.17±0.02m/s reported by Esbjörnsson et al., 2015 for a group of JIA children with similar ankle  
340 involvement. If group stratification needs to be pursued, then further investigation should aim at involving  
341 larger subgroups for every sub-type of JIA and matching them by age and size.

342 The analysis of the between-limb asymmetry at the two time-points showed similar trends in the distribution  
343 of  $\Delta I_{MRI}$  and in the observed kinematics indices, despite none of the latter was significantly different  
344 between the two time-points. In the tibiotalar articulation, lower  $\Delta I_{MRI}$  at M12 corresponded to smaller  
345 RMSD and MAV, confirming the asymmetry in the clinical involvement of the ankles is reflected by an  
346 asymmetry in the biomechanics of gait. The subtalar kinematics was in general less informative and this is  
347 probably associated to a smaller ROM of this joint when compared to the tibiotalar joint, potentially  
348 resulting in smaller sensitivity to kinematics changes. Furthermore, disease-related alterations in the  
349 movement are likely to be compensated by the tibiotalar joint being dominant in the ankle kinematics  
350 (Lundberg et al., 1989) and therefore limiting the role of the subtalar joint. The lack of an independent  
351 clinical assessment of the two joints must be considered as a limitation in the study. In fact, the present  
352 work is based on the assumption that the  $I_{MRI}$  score, evaluating the overall condition of the ankle joint, is  
353 representative of both tibiotalar and subtalar impairment level. Nonetheless, a different level of involvement  
354 of the two joints could justify their different biomechanical response. Lastly, the assumption made in  
355 schematising the joints as hinge-like mechanisms represents a substantial simplification of the true  
356 articulating surfaces, potentially limiting the representation of their true 3D motion. However, the tibiotalar  
357 kinematics was only marginally affected by this modelling choice, as this movement mainly occurs in the  
358 sagittal plane (Roach et al., 2016). On the contrary, the subtalar joint might benefit from a more detailed  
359 representation and further studies are needed to investigate this aspect.

360 In conclusion, this study showed the feasibility of using morphological fitting of MRI-based bone  
361 segmentation to identify the tibiotalar and subtalar joint axes in a non-invasive patient-specific manner.  
362 Including these joints in a musculoskeletal model of the lower limb, coupled with an appropriate marker  
363 set, can give a better understanding of their individual contribution to the ankle biomechanics. This supports

364 the adoption of the proposed modelling procedure into the practice of lower limb musculoskeletal modelling  
365 for the quantification of ankle biomechanics. The application to a pathological population, children with  
366 JIA, unveiled for the first time the absence of correlation between ankle impairment and biomechanical  
367 function, confirming the heterogeneous and systemic nature of this disease.

## 368 **Conflict of interest**

369 The authors declare they do not have any financial or personal relationships with other people or  
370 organizations that could have inappropriately influenced this study.

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