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Harnessing asymmetric N-heterocyclic carbene ligands to optimise SABRE hyperpolarisation†

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The catalytic signal amplification by reversible exchange process has become widely used for the hyperpolarisation of small molecules to improve their magnetic resonance detectability. It harnesses the latent polarisation of *para*hydrogen, and involves the formation of a labile metal complex that often contains an N-heterocyclic carbene (NHC) ligand (e.g. [Ir(H)₂(NHC)(pyridine)₃]Cl), which act as a polarisation transfer catalyst. Unfortunately, if the target molecule is too bulky, binding to the catalyst is poor and the hyperpolarisation yield is therefore low. We illustrate here the behaviour of a series of asymmetric NHC containing catalysts towards 3,4- and 3,5-lutidine in order to show how catalyst design can be used to dramatically improve the outcome of this catalytic process for sterically encumbered ligands.

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Introduction

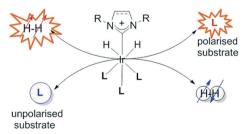
N-Heterocyclic carbenes (NHCs) are used widely as ligands in an array of chemical transformations such as hydrogenation, hydrogen transfer, hydroformylation, hydrosilylation, oxidation, isomerisation, telomerisation and various C-C bond forming reactions. ¹⁻⁴ By varying the N-bound substituents of classic imidazole based NHCs it is possible to diversify their electronic and steric properties to control reactivity. ⁵ For the free NHCs themselves, bulky substituents provide kinetic stabilisation whilst opening the carbene bond angle favours the triplet rather than the electronic singlet state formation. ^{6,7} When an NHC is bound to a metal complex the steric bulk of the NHC can be assessed *via* its percent buried volume ⁸⁻¹⁰ while the electronic effect is based on Tolman's electronic parameter. ¹¹ Both affect catalytic reactivity.

Here we target a novel form of catalysis, that is abbreviated to SABRE, signal amplification by reversible exchange. ¹² This process takes the latent magnetism of *para*hydrogen (*p*-H₂) and transfers it into a new material according to the process that is outlined in Scheme 1. Unlike a normal catalytic reaction, which chemically functionalises a substrate,

SABRE involves the catalytic transfer of polarisation into a

When this process happens in low magnetic field, the effects of chemical shift and coupling become important. Once a second order spin system is created, it becomes possible to transfer the *p*-H₂ derived hydride ligand polarisation into the NMR active nuclei and then to a reversibly bound ligand through the complex's scalar coupling network.¹⁵ The importance of this approach stems from the fact that the related hyperpolarisation method, dynamic nuclear polarisation (DNP),¹⁶ takes materials such as pyruvate and enhances their Magnetic Resonance Imaging (MRI) detectability to the point where they can be seen *in vivo*. In the case of pyruvate,¹⁷ the

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Scheme 1 Conceptual representation of the signal amplification by reversible exchange process which achieves the catalytic hyperpolarisation of a substrate (ι) via polarisation transfer within the iridium catalyst from a pair of protons that were previously located in a molecule of p-H₂.

hyperpolarisation target (1) from what is effectively p-H₂. The reversible addition of p-H₂ is therefore important, as it places the source of hyperpolarisation within the coordination sphere of the metal complex. Hence, the ligand exchange rates of the catalyst are important factors when considering how the catalyst works. $^{12-14}$

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 $[\]dagger$ Electronic supplementary information (ESI) available: NMR raw data can be found at https://doi.org/10.15124/42187464-6f06-4805-a487-9b469b7a15e4. CCDC 1848485. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/c8cy01214h

subsequent assessment of its metabolism has proven to be diagnostic of human cancer.17-20 Another approach using hyperpolarised ¹²⁹Xe has progressed to the point it is used for the diagnosis of diseases of the lung in humans.²¹ Hence, if SABRE catalysts can be optimised sufficiently, there is an expectation that a simple low cost route to hyperpolarised agents for disease diagnosis in humans would be possible. The benefits of SABRE for analytical science are however also likely to be substantial. 22-25

It is common to use precatalysts based on the [Ir(NHC)(COD)Cl] (1) motif in SABRE, and they react according to Scheme 2 to form active polarisation transfer catalysts such as [Ir(H)₂(NHC)(py)₃]Cl (2). During this process cyclooctadiene (COD) is first converted into cyclooctene (COE) and then cyclooctane (COA) and hence whilst the reaction steps shown are reversible, in practice 2 is stable.26 The ligand exchange rate of the substrate, in this case pyridine (py), also influences the efficiency of catalytic polarisation transfer because upon ligand loss the propagating spin-spin coupling framework within the catalyst is lost. Ideally, the lifetime of this complex needs to be commensurate with these small scalar couplings that are responsible for hyperpolarisation transfer. 26,27

It was found that iridium catalysts containing NHCs^{28,29} were more active than their mono phosphine counterparts because their lifetimes were closer to the optimum.30 Whilst other iridium complexes with bidentate,31 pincer ligands,32 and rhodium analogues have been tested33 NHCs have still proven to reflect the best ligand class and they have been modified to work in aqueous solution.³⁴ In fact SABRE transfers its hyperpolarisation successfully through multiple bonds to nitrogen containing compounds such as nicotinamide,³⁵ isoniazid,³⁶ pyrazole,³⁷ acetonitrile³⁸ and amines³⁹ to improve the detectability of appropriate 1H, 13C, 31P, 15N and 19F responses.40-44 It has also proven possible to use radio frequency excitation to mimic the low-field condition in order to allow high-field transfer. 15,45,46 A further exciting opportunity

Scheme 2 Route to the formation of the active SABRE catalyst 2 via the reaction of 1 with H2 and pyridine. The COD is converted first the COE and COA during this process.

revealed by SABRE is reflected in its ability to create hyperpolarised magnetic states with long lifetimes, 47 an array of research that is of growing importance as hyperpolarisation methods are beginning to feature in clinical diagnosis.

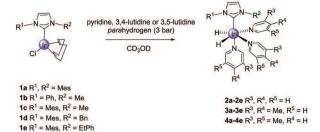
One of the limitations of SABRE has proven to be associated with its ability to hyperpolarise sterically encumbered agents due to the steric properties of the commonly used NHC IMes complex which prevents adequate binding. 48 However, without some steric interaction ligand loss will be slow.²⁶ Here we prepare a series of complexes using a range of asymmetrically substituted NHC's where we maintain some steric congestion and assess them for their SABRE activity. We achieve this by maintaining one mesityl arm in conjunction with the introduction of a smaller substituent to the second nitrogen centre. These substituents were selected carefully to prevent catalyst deactivation through well-known C-H bond activation. 49 The use of asymmetry in this way to drive exchange is somewhat analogous to the ansa-bridged metallocenes that are used to create isotactic polymers.⁵⁰ We start our studies with pyridine, which reflects the most widely used model ligand in SABRE51 before considering 3,4- and 3,5-lutidine so as to introduce additional steric interactions that we predict will lead to improved SABRE behaviour. It is noteworthy that previous studies with lutidine have identified it as a promising tool for pH imaging although steric effects acted to limit SABRE-hyperpolarization effects^{52,53} and quench it totally in ortho-substituted lutidines and picolines.⁵⁴

Results and discussion

1a and the asymmetric complexes 1b-1e were prepared according to Scheme 3. In the case of the vellow 1b, this necessitated the silver(1) oxide mediated transmetalation of [Ir(COD)Cl]₂. However, for 1c-1e a modified literature procedure⁵⁵ involving BPh₄ salts, [Ir(COD)Cl]₂ and K₂CO₃ was employed. In all cases the yields exceeded 60% and the products were characterised by 2D NMR spectroscopy, mass spectrometry and elemental analysis as detailed in the ESI.† X-ray data for 1e is included which map on to other reported structures.

$$\begin{array}{c} Ag_2O\\ [Ir(COD)Cl]_2\\ \hline CH_2Cl_2\\ \hline RT \end{array}$$

Scheme 3 Synthetic routes to 1 containing the specified C_2 asymmetric carbene.



Scheme 4 Formation of products 2, 3 and 4 from reactions with 1.

SABRE activity analysis of precatalysts 1a-1e

(a) Pyridine SABRE activity. The reactivity of precatalysts 1 towards pyridine and H2 was first investigated. In order to do this, each of the complexes 1 were dissolved in CD₃OD solutions that contained pyridine (py, 5-fold excess) prior to adding p-H2. In all cases, the solutions changed colour from yellow to colourless as 2 forms, with generic formula $[Ir(L_{NHC})(H)_2(py)_3]Cl$, where L_{NHC} is the corresponding monodentate carbene of Scheme 4. Hydride resonances were readily seen at around -22 ppm in these NMR spectra due to these species. We note that when 1b was re-examined with ¹⁵N labelled pyridine the corresponding hydride signal exhibits a ${}^{2}J_{H-(15)N}$ = 19.6 Hz splitting; two ${}^{15}N$ resonances can be seen at 236.1 and 253.4 ppm for the corresponding axial and equatorial ligands respectively.

The SABRE activities of these species were then compared at 298 K through a series of ¹H NMR studies using the 'shake and drop' method (see ESI†). The resulting per proton ortho phenyl 1H NMR resonance signal gains, and the total free pyridine substrates ¹H NMR signal gains, are reported in Table 1 for observations at 9.4 T. It can be readily seen that the order of SABRE efficiency is 2a > 2d > 2e > 2c > 2b for both data types. These values suggest that the performance of none of these new complexes compare favourably with that of 2a for pyridine signal enhancement. In order to identify the reason for this, the corresponding pyridine ligand loss rates were determined. These are detailed in Table 1 and follow the same trend. Hence we can conclude that the associated ligand loss rates are all too small, when compared to 2a, for optimal behaviour. This observation is consistent with reported literature behaviour. 56,57 When these experiments were repeated at 313 K, the corresponding signal gains increased although 2b proved unstable (see ESI†) and is not discussed further. Notably, the overall SABRE efficiency trend changes to 2a > 2d > 2c > 2e at 313 K.

(b) 3,4-Lutidine and 3,5-lutidine SABRE activity. In view of the fact that pyridine binds too tightly in 2b-2e we tested the bulkier 3,4- and 3,5-lutidine. These targets have pK_a values of 6.28 and 5.85 (ref. 58) respectively compared to 5.3 for pyridine.⁵⁹ Hence they should bind more strongly if the process is not sterically inhibited and their ligand loss rates should be lower. When CD₃OD solutions of 1 and 5-equivalents of these lutidines were examined under 3 bar of p-H2, the formation of [Ir(H)2(lut)3(LNHC)]Cl (3 and 4) was indicated as detailed in the ESI† in all cases. Tables 2 and 3 summarise the hydride chemical shifts of 3 and 4, the associated slower lutidine ligand loss rate and the ortho proton and total proton signal enhancement values that were obtained after SA-BRE transfer at 60 G and detection at 9.4 T (Fig. 1).

The signal enhancements seen for aromatic ¹H resonances of 3,4-lutidine with 3b-3e at 298 K proved to be larger than those seen for pyridine with 2b-2e. This suggests that the result of the chemical structure change and which focuses polarisation transfer into three rather than five proton sites, as for pyridine, is beneficial; this view is consistent with the comparatively poor methyl proton enhancements seen. Upon warming to 313 K the resulting signal gains increased such that 3a yields an average aromatic proton signal gain of 1700fold whilst 3d remains the best competitor with a value of 510-fold.

In contrast, the bulkier 3,5-lutidine shows much better signal enhancements at 298 K than 3,4-lutidine, which is now reflected by the total gain of 1615-fold with 4d. In both cases, the NHC 1-mesityl-3-benzylimidazole yields the best catalyst

Table 1 Rate of ligand loss per molecule in solution, level of pyridine ¹H NMR signal enhancement (fold) at 9.4 T and 298 K and 313 K for 2

Complex	2a	2b	2c	2d	2e
Pyridine ligand loss rate (298 K, s ⁻¹)	13.35	_	0.51	0.71	1.28
Ortho pyridine proton signal gain (per proton)	-1452 ± 81	-0.77 ± 0.04	-48 ± 2	-95 ± 5	-68 ± 3
and total proton gain, 298 K (fold)	5246 ± 262	2.1 ± 0.1	147 ± 7	304 ± 15	209 ± 10
Pyridine ligand loss rate (313 K, s ⁻¹)	87.82	_	3.66	6.82	7.36
Ortho pyridine proton signal gain (per proton)	-414 ± 21	_	-158 ± 8	-196 ± 10	-130 ± 6
and total proton gain, 313 K (fold)	3137 ± 156	_	544 ± 27	677 ± 34	450 ± 22

Table 2 Hydride peak positions, 3,4-lutidine loss rate, and indicated ¹H NMR signal enhancements yield for 3,4-lutidine after SABRE at 298 K

Complex	3a	3b	3 c	3d	3e
Ir–H chemical shift (ppm)	-22.80	-22.42	-22.09	-22.27	-22.32
Ligand loss rate (s ⁻¹)	9.72	_	0.39	0.76	1.9
Ortho proton signal gain (per proton)	-270 ± 13	_	-69 ± 3	-86 ± 4	-33 ± 2
and total proton gain (fold)	1709 ± 85	_	471 ± 23	506 ± 25	223 ± 11

Hydride peak positions, 3.5-lutidine loss rate, and ¹H NMR signal enhancements for 3.5-lutidine after SABRE at 298 K

Complex	4a	4b	4c	4d	4e
Ir–H chemical shift (ppm)	-22.79	-22.24	-22.35	-22.36	-22.34
Ligand loss rate (s ⁻¹)	2.10	0.34	1.01	2.14	1.9
Ortho proton signal gain (per proton) and total proton gain (fold)	-145 ± 7	-1.7 ± 0.1	-125 ± 6	-258 ± 13	-86.7 ± 4
	740 ± 37	10.5 ± 0.5	719 ± 36	1615 ± 81	504 ± 25

(3d and 4d) and we note there are two pairs of ortho phenyl protons in $[Ir(H)_2(lut)_3(L_{NHC})]Cl$ that provides a ${}^4J_{HH}$ coupling link to their hydride ligands which should be similar to those for pyridine. It is not surprising that both of these positions therefore receive strong polarisation under similar SABRE. Furthermore, it can be seen that, as predicted, the ability of the SABRE target to receive hyperpolarisation is controlled by the ligand scaffold.

Continuous flow experiments. The variation in the level of SABRE observed is therefore strongly dependent on the catalyst and substrate combination. According to the literature, it is also influenced by the magnetic field experienced by the sample at the point of polarisation transfer. This situation arises because polarisation transfer is relayed via the scalar coupling in the catalyst when under weak-field conditions. Hence the chemical shift difference that exists between the parahydrogen-derived hydride ligands and the interacting spin-½ nuclei of the ligated substrate influence the level of polarisation transfer observed.¹²

We used an automated polarizer that has been described previously^{60,61} to examine this effect by reference to pyridine and 3,4-lutidine. Fig. 2 shows how the ¹H NMR signal gain for 3,4-lutidine varies with the changing polarisation transfer field when using catalyst 3d. For ¹H transfer, the optimal value for pyridine was 70 G whilst for the aromatic resonances of 3,4-lutidine it is 60 G. It appears, however, that optimal transfer into the aliphatic groups is more efficient at 70 G. It is highly noteworthy that the methyl resonances of 3,4-lutidine are substantially enhanced when compared to related reports.³⁵

Fig. 3 shows the corresponding polarisation transfer field profile for 3,5-lutidine achieved with 4d. The optimal field

3.4-lutidine therma

Fig. 1 Typically thermal and hyperpolarised ¹H NMR spectra of 3,4-lutidine achieved by 3d illustrating the hyperpolarisation effect.

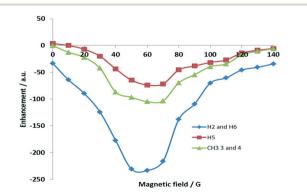
value for polarisation transfer now lies between 70 and 80 G with the methyl groups receiving a far greater share of the signal.

Effects of NMR relaxation. In order to further rationalise this behaviour, the relaxation times of the protons of these three substrates were measured at 9.4 T in the absence of catalysts in methanol-d4 whilst under H2 as detailed in Table 4. The corresponding values determined under SABRE conditions are also detailed. It is clear that, as expected, the presence of the catalyst consistently reduces these values. 35,62 We therefore determined the corresponding relaxation times of key protons in 3 and 4 when they were prepared from 1 with just a 3.05 fold excess based on iridium.

From all these data it can be seen that whilst 4d has a very similar ligand loss rate to 4a, the total signal enhancement it provides is improved by almost 50%. This change is clearly due to less efficient relaxation within the catalyst, which improves by 18% in 4d versus 4a. In support of this, 4c and 4a actually also deliver similar signal gains with similar relaxation times even though their ligand exchange rates differ by >50%. In view of these observations we can state that the effects of ligand exchange on SABRE activity must be less critical than those of relaxation within the catalyst.

¹³C SABRE transfer with 3 and 4

It has also been shown that SABRE can enhance the ¹³C responses of molecules. When the ¹³C hyperpolarisation of pyridine was evaluated for 2a, limited 13C signal gains are observed even though the IMes derivate works extremely well for ¹H. This suggests that the ligand exchange rate is too fast (13.35 s⁻¹) for substantial direct ¹³C transfer at this field. In contrast, the results with slower exchanging 2b-2e prove



Polarisation transfer field profile of 3,4-lutidine using precursor Fig. 2

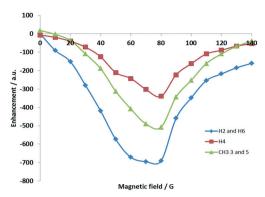


Fig. 3 Polarisation transfer field profile for 3,5-lutidine based on precursor 1d

Table 4 Observed T_1 values for the indicated protons of 3,4-lutidine and 3,5-lutidine in the free material at 9.4 T without, and in the presence of the indicated catalyst, alongside the corresponding values when bound as a ligand *trans* to hydride. The initial ratio of 1 to the indicated ligand was 1:3.05 to ensure free substrate was present

Sample/agent	T_1 (/s) value (sin	T_1 (/s) value (site)			
3,4-Lutidine	Ortho	Meta	CH ₃		
Without	5.75 & 5.58	5.14	3.31 & 3.83		
3a, free ligand	2.83 & 2.14	6.45	1.80 & 1.82		
3a, coordinated	3.20 & 2.49	5.09	1.51 & 1.20		
3,5-Lutidine					
Without	21.30	19.58	6.63		
4a, free ligand	3.23	6.26	2.10		
4a, coordinated	3.13	4.30	2.44		
4c, free ligand	3.86	5.47	2.45		
4c, coordinated	2.97	6.91	1.92		
4d, free ligand	3.74	6.63	2.49		
4d, coordinated	3.81	6.34	2.35		

better with 2d being the most efficient. The *ortho* and *meta* ¹³C positions exhibit the strongest signal gains on the molecule. When the polarisation transfer field (PTF) dependence was considered, optimum transfer was achieved at 50 G for 2d with the total carbon signal enhancement being 430-fold at this value.

When 3,4-lutidine was examined, 3d and 3e proved to be similarly effective. Fig. 4 shows a typical NMR spectrum that was obtained with 3d. For context, when a normal ¹³C{¹H} NMR spectrum was acquired, 1024 scans were needed to see the associated carbon signals (Fig. 4a).

The signals that are seen in these fully coupled NMR spectra contain anti-phase character that is associated with the detection of a heteronuclear longitudinal two spin order term (I_zS_z). In each case, the anti-phase splitting is ~8 Hz in size and reflects a long-range rather than direct one bond CH splitting. This is fully consistent with related reported observations. The SABRE ¹³C activity of 3d was also investigated more rigorously using the flow apparatus. The two inequivalent *meta*- and *para*-methyl ¹³C signals (positions 3 and 4) of 3,4-lutidine were found to show relatively large enhancements after transfer between 0–90 G, with no signal

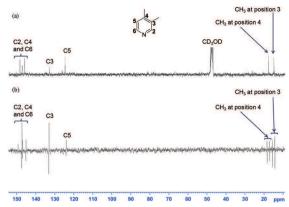


Fig. 4 (a) $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum recorded with 1024 scans for a mixture of 3d and 3,4-lutidine without $p\text{-H}_2$; (b) ^{13}C NMR spectra recorded in one scan on the same sample after SABRE showing the corresponding hyperpolarised carbon signals (the vertical expansion setting used to display the spectrum in Fig 4 (a) is four times that used in Fig. 4 (b)); the relaxation delay was 3 seconds alongside a 30° excitation pulse.

seen after transfer between 90–140 G. The *meta*-methyl ¹³C signals (position 3) generally exhibited lower polarisation levels to that of the *para*-methyl resonance (position 4). In the case of the aromatic region, the *meta* quaternary carbon at position 4 on 3,4-lutidine showed an individual enhancement value of 544-fold at 40 G whereas the *ortho*-CH value was 560-fold. The *meta*-CH resonance showed minimal polarisation enhancement at 40 G and this increase to 50-fold at 30 G. The trend in signal intensity with polarisation transfer field for these ¹³C resonances is very different to that seen in the corresponding ¹H NMR spectra.

In addition, the maximum signal gain seen in the ¹H NMR spectra is reflected in the *ortho*-CH resonances, followed by the methyl groups, and finally the *meta*-CH proton. Comparatively, 3a still yields significantly lower ¹³C polarisation than 3d, achieving only a 222-fold gain for the *meta*-quaternary carbon at 70 G. While the *ortho*-CH signal could be seen, the remaining carbon resonances exhibited signal enhancements of 20–300 fold depending on the magnetic field strength tested. Hence, the total ¹³C signal enhancements seen for 3d were 1300-fold after transfer at 40 G, which was significantly higher than the corresponding 780-fold at 0.5 G achieved with 3a as detailed in Fig. 5.

Precursor 1d also achieved the optimal ¹³C NMR signal enhancement in 3,5-lutidine and the results showed a lower level of enhancement compared to 3,4-lutidine, which is in agreement with the corresponding ¹H NMR changes. Now signals for all of the carbon sites, except the *para* position, are clearly visible after transfer in fields between 0 and 130 G. At 30 G, the optimum magnetic field, the total carbon NMR signal gain was 800-fold, with a 380-fold contribution coming from the quaternary *meta* carbon position, 96-fold for the methyl carbons, and the remainder being due to *ortho* ¹³C signal. The full PTF profile is presented in Fig. 6.

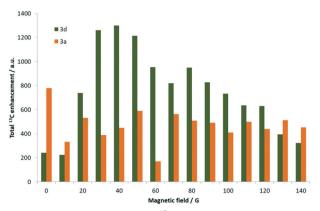


Fig. 5 Graph comparing the total ¹³C enhancement observed for the carbon resonances of 3,4-lutidine when using complex 3d (green) and 3a (orange) respectively as a function of the strength of the polarisation transfer field.

Early reaction stages

Paper

During such SABRE studies it has proven common in the early stages of reaction to observe complexes of the type [Ir(H)₂(COD)(NHC)(sub)]Cl. These materials form transiently as 1 converts into the final tris substituted product. They actually reflect H2 addition to the initial substitution product [Ir(sub)(NHC)(COD)]Cl which forms from the precursor [Ir(NHC)(COD)Cl]. Their observation is important because If the initial substitution is slow then inefficient H2 addition to [Ir(NHC)(COD)Cl] occurs. However, while the resulting product [Ir(H)₂(NHC)(COD)Cl] goes on to yield common [Ir(H)₂(sub)-(NHC)(COD)]Cl the reaction can take weeks at 298 K. For completeness, the COD ligand present in these complexes is converted to COE and finally COA during this process.

We therefore examined the early stages of these reactions with pyridine but observed [Ir(py)(NHC)(COD)]Cl to form only when the NHC was IMes. When the associated sample of 1b containing a 5-fold excess of pyridine was exposed to parahydrogen in methanol- d_4 at 263 K, and the solution warmed to 273 K, no reaction was initially evident in the corresponding 1-scan ¹H NMR spectrum. However, when 16-scans are av-

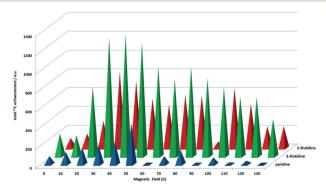


Fig. 6 Graph comparing the ¹³C enhancement observed for the carbon resonances of pyridine, 3,4-lutidine and 3,5-lutidine when using complex 3d as a function of the strength of the polarisation transfer field.

eraged, a pair of very weak antiphase hydride signals is detected at -12.68 and -17.67 ppm that share a common J_{H-H} coupling of -10 Hz. Subsequently the species giving rise to this material converts into 2b, although significant decomposition is evident.

When a similar sample of 1c was exposed to p-H₂ at 273 K in methanol- d_4 a pair of similar strongly enhanced antiphase hydride signals is detected. They now occur at -12.86 and -17.82 ppm and share a $J_{\rm HH}$ coupling of -5.5 Hz. Similar observations result from 1d and 1e and the associated NMR spectra are illustrated in Fig. 7. We note that the rate of formation of final products 2d and 2e exceeds that of 2b and 2c.

Experimental

The asymmetric monocarbene ligands of Scheme 1 were synthesised by modifying previously published procedures.⁶³ Phenylimidazole, or mesitylimidazole, were reacted with the respective iodomethane, benzyl bromide or phenethyl bromide in acetonitrile and stirred at 25 °C or at reflux. The solvent was evaporated and crude brown-oil was recrystallised from methanol: diethyl ether mixtures to give off-white solids [PhIMe]I, [MesIMe]I, [MesIBn]Br or [MesIEtPh]Br. These ligands were dissolved in dichloromethane and stirred for 1 hour with NaBPh4 to exchange the counter-ion. The mixture was filtered and the solvent evaporated in vacuo to yield [MesIMe]BPh4, [MesIBn]BPh4 or [MesIEtPh]BPh4 as white crystalline solids.

Synthesis of Ir(1) complexes containing asymmetric monocarbene ligands 1b-1e

1b was synthesised by the silver(1) transmetallation of [Ir(COD)Cl]₂. The reaction mixture was stirred overnight at 25

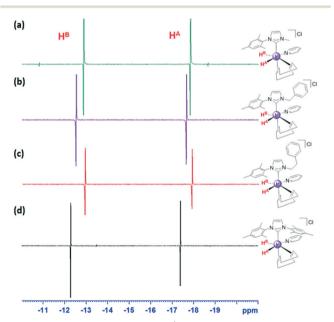


Fig. 7 Hydride region of a series of ¹H NMR spectra recorded using a $\pi/4$ pulse (400 MHz, CD₃OD, 273 K) showing signals for: (a) 5c; (b) 5d; (c) 5e and (d) 5a.

°C, filtered and the solvent evaporated. The crude product was then dissolved in dichloromethane, filtered, isolated and recrystallised from dichloromethane: hexane as a bright yellow solid. 1c-1e were synthesised by heating [Ir(COD)Cl]₂ and K₂CO₃ in acetone with the corresponding ligand at reflux for 4 hours. After cooling, the mixture was filtered, and the solvent evaporated. The crude product was redissolved in dichloromethane, and a white solid precipitates. [Ir(COD)(MesIMe)Cl] (1c), [Ir(COD)(MesIBn)Cl] (1d) and [Ir(COD)(MesIEtPh)Cl] (1e) resulted as yellow solids, and were used without further purification. All these complexes were air-stable as solids.

Single crystals of and 1e suitable for X-ray structure determination were obtained by slow diffusion of a concentrated dichloromethane solution of the complex in hexane at -20 °C (see ESI†).

Conclusions

The results presented in this work demonstrate that the asymmetric carbene ligands of Scheme 2 can be used to prepare complexes of the type [Ir(NHC)(COD)Cl] (1). These complexes have been shown to react with hydrogen and substrates pyridine, 3,4-lutidine or 3,5-lutidine to form the corresponding Ir(III) products [Ir(H)₂(NHC)(sub)₃]Cl (2-4) of Scheme 3. These products were then assessed for activity in the SABRE process, which involves the catalytic transfer of polarisation from p-H2 into the substrate to enhance its NMR detectability. Key parameters in this process, namely the ligand exchange rates, and the relaxation times of the associated NMR resonances have been assessed and compared to those of the symmetric carbene bearing [Ir(H)₂(IMes)(sub)₃]Cl (2-4a), the established SABRE reference catalyst. For pyridine and 3,4-lutidine poor activity was observed as a consequence of slow ligand loss due to the reduced steric bulk associated with these NHC's relative to IMes and strong interactions with the associated nitrogen centres due to their high basicity. However, when 3,5-lutidine was examined, the enhanced steric encumbrance of this substrate out ways the pK_a benefit and increases the rates of ligand loss for the asymmetric NHC's such that they become competitive with their IMes derivatives. The result of this process is that 4d, achieved an ortho proton signal gain of 258-fold per proton in 3,5-lutidine in contrast to the 145-fold value achieved by 4a at 9.4 T.

One further benefit of these catalysts is reflected in their ability to sensitise the ¹³C response of these agents. Now the enhanced stability works favourably as evidenced by the spectra of Fig. 4. It is therefore likely that asymmetric carbenes offer a route to further improve the ability of SABRE to enhance the MR response of sterically encumbered substrates and hence the range of applications. For efficient polarization transfer to ¹³C, the utilization of a micro-Tesla field in conjunction with the presence of ¹⁵N to relay polarization transfer has proven highly efficient thereby suggesting a route to improve on these results. ^{64,65}

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 W. A. Herrmann, K. Öfele, D. v. Preysing and S. K. Schneider, J. Organomet. Chem., 2003, 687, 229–248.
- 2 G. Altenhoff, R. Goddard, C. W. Lehmann and F. Glorius, Angew. Chem., Int. Ed., 2003, 42, 3690–3693.
- 3 M. Eckhardt and G. C. Fu, J. Am. Chem. Soc., 2003, 125, 13642-13643.
- 4 H. Lebel, M. K. Janes, A. B. Charette and S. P. Nolan, J. Am. Chem. Soc., 2004, 126, 5046–5047.
- 5 H. Jacobsen, A. Correa, A. Poater, C. Costabile and L. Cavallo, *Coord. Chem. Rev.*, 2009, 253, 687–703.
- 6 B. C. Gilbert, D. Griller and A. S. Nazran, *J. Org. Chem.*, 1985, **50**, 4738–4742.
- 7 D. Bourissou, O. Guerret, F. P. Gabbaï and G. Bertrand, Chem. Rev., 2000, 100, 39–92.
- 8 A. C. Hillier, W. J. Sommer, B. S. Yong, J. L. Petersen, L. Cavallo and S. P. Nolan, *Organometallics*, 2003, 22, 4322–4326.
- 9 S. Díez-González and S. P. Nolan, Coord. Chem. Rev., 2007, 251, 874–883.
- 10 H. Clavier and S. P. Nolan, Chem. Commun., 2010, 46, 841–861.
- 11 C. A. Tolman, Chem. Rev., 1977, 77, 313-348.
- 12 R. W. Adams, J. A. Aguilar, K. D. Atkinson, M. J. Cowley, P. I. P. Elliott, S. B. Duckett, G. G. R. Green, I. G. Khazal, J. Lopez-Serrano and D. C. Williamson, *Science*, 2009, 323, 1708–1711.
- 13 R. W. Adams, S. B. Duckett, R. A. Green, D. C. Williamson and G. G. R. Green, *J. Chem. Phys.*, 2009, 131, 194505.
- 14 K. D. Atkinson, M. J. Cowley, P. I. P. Elliott, S. B. Duckett, G. G. R. Green, J. López-Serrano and A. C. Whitwood, *J. Am. Chem. Soc.*, 2009, 131, 13362–13368.
- 15 A. N. Pravdivtsev, A. V. Yurkovskaya, H. Zimmermann, H.-M. Vieth and K. L. Ivanov, *RSC Adv.*, 2015, 5, 63615–63623.
- 16 J.-H. Ardenkjaer-Larsen, G. S. Boebinger, A. Comment, S. Duckett, A. S. Edison, F. Engelke, C. Griesinger, R. G. Griffin, C. Hilty, H. Maeda, G. Parigi, T. Prisner, E. Ravera, J. van Bentum, S. Vega, A. Webb, C. Luchinat, H. Schwalbe and L. Frydman, *Angew. Chem., Int. Ed.*, 2015, 54, 9162–9185.
- 17 K. Golman, R. i. t. Zandt, M. Lerche, R. Pehrson and J. H. Ardenkjaer-Larsen, *Cancer Res.*, 2006, 66, 10855–10860.

- 18 I. Marco-Rius, M. C. D. Tayler, M. I. Kettunen, T. J. Larkin, K. N. Timm, E. M. Serrao, T. B. Rodrigues, G. Pileio, J. H. Ardenkjaer-Larsen, M. H. Levitt and K. M. Brindle, NMR Biomed., 2013, 26, 1696-1704.
- 19 S. R. Y. Xu, C. Ø. Mariager, L. B. Bertelsen, M. Schroeder, H. Oi, C. Laustsen and H. Stødkilde-Jørgensen, Tomography, 2017, 3, 67-73.
- 20 C. H. Cunningham, J. Y. C. Lau, A. P. Chen, B. J. Geraghty, W. J. Perks, I. Roifman, G. A. Wright and K. A. Connelly, Circ. Res., 2016, 119, 1177-1182.
- 21 J. P. Mugler and T. A. Altes, Magn. Reson. Imaging, 2013, 37, 313-331.
- 22 N. K. J. Hermkens, R. Aspers, M. C. Feiters, F. Rutjes and M. Tessari, Magn. Reson. Chem., 2018, 56, 633-640.
- 23 I. Reile, N. Eshuis, N. K. J. Hermkens, B. J. A. van Weerdenburg, M. C. Feiters, F. Rutjes and M. Tessari, Analyst, 2016, 141, 4001-4005.
- 24 N. Eshuis, R. Aspers, B. J. A. van Weerdenburg, M. C. Feiters, F. Rutjes, S. S. Wijmenga and M. Tessari, Angew. Chem., Int. Ed., 2015, 54, 14527-14530.
- 25 L. S. Lloyd, R. W. Adams, M. Bernstein, S. Coombes, S. B. Duckett, G. G. R. Green, R. J. Lewis, R. E. Mewis and C. J. Sleigh, J. Am. Chem. Soc., 2012, 134, 12904-12907.
- 26 L. S. Lloyd, A. Asghar, M. J. Burns, A. Charlton, S. Coombes, M. J. Cowley, G. J. Dear, S. B. Duckett, G. R. Genov, G. G. R. Green, L. A. R. Highton, A. J. J. Hooper, M. Khan, I. G. Khazal, R. J. Lewis, R. E. Mewis, A. D. Roberts and A. J. Ruddlesden, Catal. Sci. Technol., 2014, 4, 3544-3554.
- 27 N. Eshuis, R. Aspers, B. J. A. van Weerdenburg, M. C. Feiters, F. Rutjes, S. S. Wijmenga and M. Tessari, J. Magn. Reson., 2016, 265, 59-66.
- 28 L. S. Lloyd, A. Asghar, M. J. Burns, A. Charlton, S. Coombes, M. J. Cowley, G. J. Dear, S. B. Duckett, G. R. Genov, G. G. R. Green, L. A. R. Highton, A. J. J. Hooper, M. Khan, I. G. Khazal, R. J. Lewis, R. E. Mewis, A. D. Roberts and A. J. Ruddlesden, Catal. Sci. Technol., 2014, 4, 3544-3554.
- B. J. A. van Weerdenburg, S. Gloeggler, N. Eshuis, A. H. J. Engwerda, J. M. M. Smits, R. de Gelder, S. Appelt, S. S. Wymenga, M. Tessari, M. C. Feiters, B. Bluemich and F. P. J. T. Rutjes, Chem. Commun., 2013, 49, 7388-7390.
- 30 R. W. Adams, S. B. Duckett, R. A. Green, D. C. Williamson and G. G. R. Green, J. Chem. Phys., 2009, 131, 194505-194505.
- 31 A. J. Ruddlesden, R. E. Mewis, G. G. R. Green, A. C. Whitwood and S. B. Duckett, Organometallics, 2015, 34, 2997-3006.
- 32 A. J. Holmes, P. J. Rayner, M. J. Cowley, G. G. R. Green, A. C. Whitwood and S. B. Duckett, Dalton Trans., 2015, 44, 1077-1083.
- 33 V. V. Zhivonitko, I. V. Skovpin and I. V. Koptyug, Chem. Commun., 2015, 51, 2506-2509.
- 34 P. Spannring, I. Reile, M. Emondts, P. P. M. Schleker, N. K. J. Hermkens, N. G. J. v. d. Zwaluw, B. J. A. v. Weerdenburg, P. Tinnemans, M. Tessari, B. Blümich, F. P. J. T. Rutjes and M. C. Feiters, Chem. - Eur. J., 2016, 22, 9277-9282.

- 35 P. J. Ravner, M. J. Burns, A. M. Olaru, P. Norcott, M. Fekete, G. G. R. Green, L. A. R. Highton, R. E. Mewis and S. B. Duckett, Proc. Natl. Acad. Sci. U. S. A., 2017, 114, E3188-E3194.
- 36 H. Zeng, J. Xu, J. Gillen, M. T. McMahon, D. Artemov, J.-M. Tyburn, J. A. B. Lohman, R. E. Mewis, K. D. Atkinson, G. G. R. Green, S. B. Duckett and P. C. M. van Zijl, J. Magn. Reson., 2013, 237, 73-78.
- 37 E. B. Dücker, L. T. Kuhn, K. Münnemann and C. Griesinger, J. Magn. Reson., 2012, 214, 159-165.
- 38 R. E. Mewis, R. A. Green, M. C. R. Cockett, M. J. Cowley, S. B. Duckett, G. G. R. Green, R. O. John, P. J. Rayner and D. C. Williamson, J. Phys. Chem. B, 2015, 119, 1416-1424.
- 39 W. Iali, P. J. Rayner and S. B. Duckett, Sci. Adv., 2018, 4, eaa06250.
- 40 P. J. Rayner, M. J. Burns, A. M. Olaru, P. Norcott, M. Fekete, G. G. R. Green, L. A. R. Highton, R. E. Mewis and S. B. Duckett, Proc. Natl. Acad. Sci. U. S. A., 2017, 114, E3188-E3194.
- 41 J. F. P. Colell, A. W. J. Logan, Z. Zhou, R. V. Shchepin, D. A. Barskiy, G. X. Ortiz, Q. Wang, S. J. Malcolmson, E. Y. Chekmenev, W. S. Warren and T. Theis, J. Phys. Chem. C, 2017, 121, 6626-6634.
- 42 R. V. Shchepin, B. M. Goodson, T. Theis, W. S. Warren and E. Y. Chekmenev, ChemPhysChem, 2017, 18, 1961-1965.
- 43 M. J. Burns, P. J. Rayner, G. G. R. Green, L. A. R. Highton, R. E. Mewis and S. B. Duckett, J. Phys. Chem. B, 2015, 119, 5020-5027.
- 44 D. A. Barskiy, R. V. Shchepin, C. P. N. Tanner, J. F. P. Colell, B. M. Goodson, T. Theis, W. S. Warren and E. Y. Chekmenev, ChemPhysChem, 2017, 18, 1493-1498.
- 45 T. Theis, M. Truong, A. M. Coffey, E. Y. Chekmenev and W. S. Warren, J. Magn. Reson., 2014, 248, 23-26.
- 46 S. S. Roy, G. Stevanato, P. J. Rayner and S. B. Duckett, J. Magn. Reson., 2017, 285, 55-60.
- 47 S. S. Roy, P. J. Rayner, P. Norcott, G. G. R. Green and S. B. Duckett, Phys. Chem. Chem. Phys., 2016, 18, 24905-24911.
- 48 M. J. Cowley, R. W. Adams, K. D. Atkinson, M. C. R. Cockett, S. B. Duckett, G. G. R. Green, J. A. B. Lohman, R. Kerssebaum, D. Kilgour and R. E. Mewis, J. Am. Chem. Soc., 2011, 133, 6134-6137.
- 49 N. Phillips, J. Rowles, M. J. Kelly, I. Riddlestone, N. H. Rees, A. Dervisi, I. A. Fallis and S. Aldridge, Organometallics, 2012, 31, 8075-8078.
- 50 H. H. Brintzinger and D. Fischer, in Polyolefins: 50 years after Ziegler and Natta II: Polyolefins by Metallocenes and Other Single-Site Catalysts, ed. W. Kaminsky, Springer Berlin Heidelberg, Berlin, Heidelberg, 2013, pp. 29-42.
- 51 W. Jiang, L. Lumata, W. Chen, S. Zhang, Z. Kovacs, C. Khemtong and A. D. Sherry, Sci. Rep., 2015, 5, 9104.
- 52 W. Jiang, L. Lumata, W. Chen, S. Zhang, Z. Kovacs, A. D. Sherry and C. Khemtong, Sci. Rep., 2015, 5, 9104.
- 53 R. V. Shchepin, D. A. Barskiy, A. M. Coffey, T. Theis, F. Shi, W. S. Warren, B. M. Goodson and E. Y. Chekmenev, ACS Sens., 2016, 1, 640-644.
- 54 R. V. Shchepin, M. L. Truong, T. Theis, A. M. Coffey, F. Shi, K. W. Waddell, W. S. Warren, B. M. Goodson and E. Y. Chekmenev, J. Phys. Chem. Lett., 2015, 6, 1961–1967.

- 55 R. Savka and H. Plenio, *Dalton Trans.*, 2015, 44, 891–893.
- 56 J.-B. Hovener, N. Schwaderlapp, T. Lickert, D. Leibfritz, J. Hennig, D. von Elverfeldt, B. Duckett Simon, E. Mewis Ryan, A. R. Highton Louise, M. Kenny Stephen, G. R. Green Gary and G. Korvink Jan, *Nat. Commun.*, 2013, 4, 2946.
- 57 G. W. Vuister, M. Tessari, Y. Karimi-Nejad and B. Whitehead, in *Modern Techniques in Protein NMR*, ed. N. R. Krishna and L. J. Berliner, Springer US, Boston, MA, 2002, pp. 195–257.
- 58 C. S. Handloser, M. R. Chakrabarty and M. W. Mosher, J. Chem. Educ., 1973, 50, 510–511.
- 59 C. S. Handloser, M. R. Chakrabarty and M. W. Mosher, J. Chem. Educ., 1973, 50, 510.
- 60 M. J. Cowley, R. W. Adams, K. D. Atkinson, M. C. R. Cockett, S. B. Duckett, G. G. R. Green, J. A. B. Lohman, R.

- Kerssebaum, D. Kilgour and R. E. Mewis, *J. Am. Chem. Soc.*, 2011, 133, 6134–6137.
- 61 R. E. Mewis, K. D. Atkinson, M. J. Cowley, S. B. Duckett, G. G. R. Green, R. A. Green, L. A. R. Highton, D. Kilgour, L. S. Lloyd, J. A. B. Lohman and D. C. Williamson, *Magn. Reson. Chem.*, 2014, 52, 358–369.
- 62 R. E. Mewis, M. Fekete, G. G. R. Green, A. C. Whitwood and S. B. Duckett, *Chem. Commun.*, 2015, 51, 9857–9859.
- 63 C. Nieto-Oberhuber, S. Lopez and A. M. Echavarren, *J. Am. Chem. Soc.*, 2005, 127, 6178–6179.
- 64 R. V. Shchepin, L. Jaigirdar, T. Theis, W. S. Warren, B. M. Goodson and E. Y. Chekmenev, *J. Phys. Chem. C*, 2017, 121, 28425–28434.
- 65 T. Theis, M. L. Truong, A. M. Coffey, R. V. Shchepin, K. W. Waddell, F. Shi, B. M. Goodson, W. S. Warren and E. Y. Chekmenev, J. Am. Chem. Soc., 2015, 137, 1404–1407.