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Pellis, Alessandro, Comerford, James W. orcid.org/0000-0002-9977-5695, Maneffa, Andrew J. et al. (3 more authors) (2018) Elucidating enzymatic polymerisations:Chain-length selectivity of Candida antarctica lipase B towards various aliphatic diols and dicarboxylic acid diesters. European Polymer Journal. pp. 79-84. ISSN: 0014-3057

<https://doi.org/10.1016/j.eurpolymj.2018.07.009>

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Elucidating enzymatic polymerisations: chain-length selectivity of *Candida antarctica* lipase B towards various aliphatic diols and dicarboxylic acid diesters

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Keywords: enzymatic polymerisation, biobased polyesters, solventless synthesis, *Candida antarctica* lipase B, enzymatic selectivity

Abstract

The sustainable synthesis of polymers is a field with growing interest due to the need of modern society to preserve the environment whilst making used products and food sustainable for the future generations. In this work we investigate the possibility of synthesizing aliphatic polyesters derived from various dicarboxylic acid diesters and diols in a solvent-free reaction system. *Candida antarctica* lipase B was selected as biocatalyst and its selectivity towards the carbon and ester chain length were elucidated. The selected enzyme was able to synthesize various polyesters combining C₄-C₁₀ diesters and C₄-C₈ diols. All combinations led to monomer conversions above 90% in 24 h with the best number average molecular weights (M_n) being obtained through the combination of dimethyl adipate and 1,8-octanediol leading to a M_n of 7141 Da. Differential scanning

calorimetry analysis shows a clear trend with an increase in melting temperature of the polymers that correlates with both the increase of the M_n or of the polymer's constitutional repeat unit carbon chain length. Thermogravimetric analysis and rheology measurements performed on selected samples also confirm the trend showing a variation of the polymer's degradation temperatures and viscosity profiles.

Introduction

The application of biocatalysts in organic synthesis offers several advantages compared with traditional chemo-catalysts such as milder reaction conditions with regards to temperature (usually $T < 100\text{ }^{\circ}\text{C}$), pressure and pH (normally 3-8). Such conditions often lead to remarkable energy efficiency, high enantio-, regio- and chemo-selectivities as well as controlled stereochemistry. These features allow the development of new functional compounds for pharmaceuticals, agrochemicals and polymers using nontoxic natural catalysts with a significant "green" appeal having commercial benefits and satisfying ecological requirements [1].

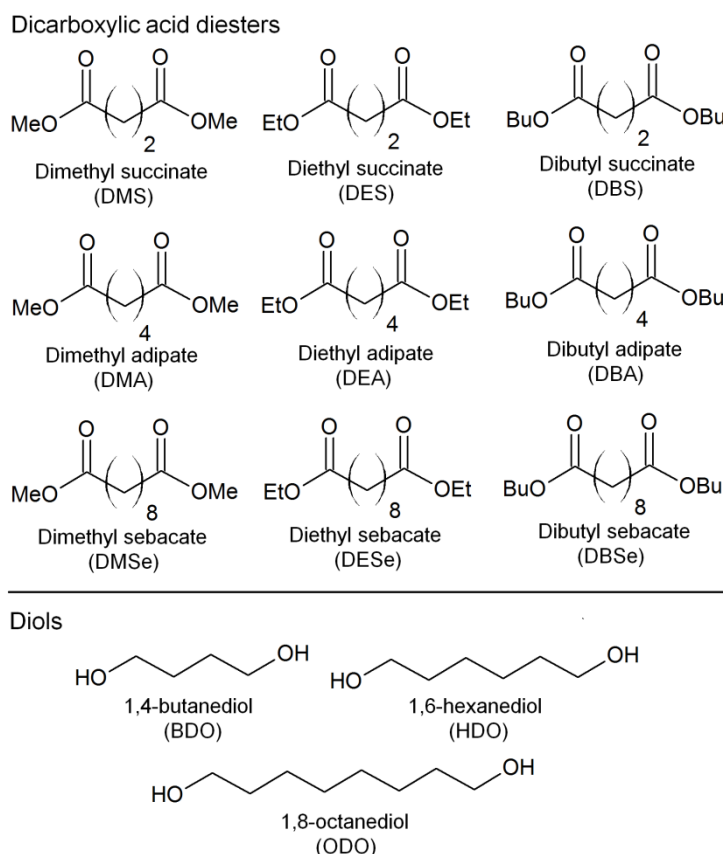
Despite studies focused on the use of glycosidases for the synthesis of natural and unnatural polysaccharides [2], as well as oxidoreductases for the polymerisation of phenol derivatives [3] and vinyl monomers [1] (mainly using laccases and peroxidases), the most investigated area of enzymatic synthesis is the production of polyesters via both polycondensation (transesterification) and ring opening polymerisations (ROPs) [4,5]. These areas have predominantly emerged thanks to the discovery and commercial availability of *Candida antarctica* lipase B (CaLB). Over recent years the extraordinary properties of this enzyme were brought to light from several research teams in the kinetic resolution of organofluorine rac-alcohols [6], the synthesis of glucoside esters [7] and the enantioselective synthesis of a β -amino acid ester via a solvent-free chemo-enzymatic reaction among others [8].

53 Further to this, CaLB has been shown to be an active catalyst for the synthesis of a wide
54 range of aliphatic [9], aliphatic functional [10, 11] (e.g. polyesters containing lateral
55 functionalities such as vinyl and hydroxy groups) and aliphatic-aromatic polyesters [12,13]
56 and polyamides [14]. In recent years these polyesters and polyamides have been derived
57 preferentially from renewable monomers such as 2,5-furandicarboxylic, adipic and succinic
58 acids and 1,4-butanediol among others [5].

59 Despite several other enzymes belonging to the hydrolases family, namely cutinases from
60 *Humicola insolens* [15], *Fusarium solani pisi* [16] and *Thermobifida cellulosilytica* [17],
61 being reported to be active for the synthesis of various polyesters and polyamides in their
62 lyophilized and immobilized forms (ranging from cross-linked enzyme aggregates to
63 covalent binding [18, 19]), the choice of the chemist is still often the readily available CaLB
64 adsorbed on a methacrylic resin known under the tradename of Novozym® 435. This
65 biocatalyst has been shown to be active and stable in several different conditions ranging
66 from water-based to anhydrous organic media and up to temperatures of ~100 °C.

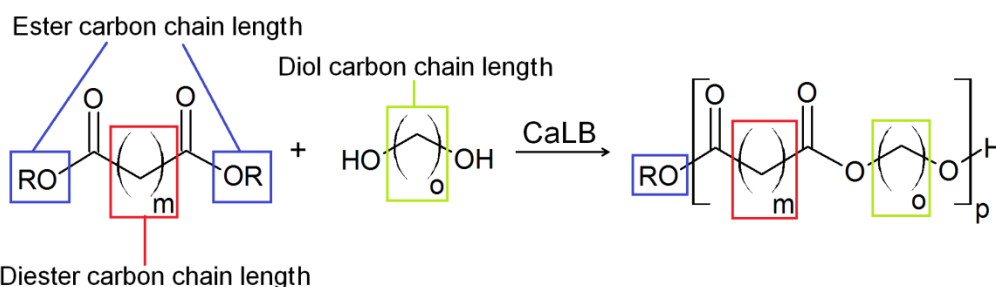
67 Among the many synthesis studies both on polycondensation and ROPs that were
68 performed over the years using this enzyme, we found there is a lack of comparative
69 studies on the range of monomers shown in Scheme 1 (most of which bio-based) [20] for
70 the synthesis of aliphatic polyesters.

71



Scheme 1. Dicarboxylic acid diesters (top) and linear diols (bottom) used in this study for the enzymatic synthesis of aliphatic polyesters in solvent-free systems using *Candida antarctica* lipase B as biocatalyst.

Despite the many studies on the topic, there remains an absence of any detailed systematic study into the implications of changing the diester and/or diol whilst applying identical methodologies for the polymerisation. In the present study we investigated the CaLB-catalyzed polycondensation of various methyl-, ethyl- and butyl- dicarboxylic acid esters with various linear diols having a carbon chain length of 4, 6 or 8, shedding light on how reactive different size diesters are when employing enzymatic catalysis in a solvent-free reaction system (Scheme 2). These results will/help(ed) us to understand the strengths and weaknesses of using this, until now, sporadically investigated enzyme for polyester synthesis.



Scheme 2. The three parameters that were investigated in the present study, namely 1) ester carbon chain length; 2) diester carbon chain length and 3) diol carbon chain length.

Materials and methods

Chemicals and enzymes

1,4-butanediol (BDO) was purchased from Alfa Aesar. Dimethyl adipate (DMA) and 1,8-octanediol (ODO) were purchased from Acros Organics. Dimethyl succinate (DMS) was purchased from Fluka. All other chemicals and solvents were purchased from Sigma-Aldrich and used as received if not otherwise specified. *Candida antarctica* lipase B (CaLB) immobilized onto methacrylic resin was purchased from Sigma-Aldrich (product code L4777) (N435). The enzyme was dried under vacuum for 96 h at 25 °C and stored in a desiccator prior to use.

Enzymatic polycondensation of aliphatic polyesters

0.006 mol of dicarboxylic acid ester (A) and 0.006 mol of linear diol (B) (diester:diol ratio= 1.0:1.0) were accurately weighted in a 25 mL round bottom flask. The mixture was then stirred at 85 °C until a homogeneous melt was obtained. 10% w w⁻¹ (calculated on the total amount of the monomers) of N435 was then added and the reaction was run for 6 h at 1 Atm. A vacuum of 20 mbar was subsequently applied for an additional 18 h maintaining the reaction temperature at 85 °C (total reaction time: 24 h). The reaction product was recovered by adding THF (or DCM in the case of the 24 h succinate-BDO polymers) in order to dissolve the solid reaction products. The biocatalyst was then removed via a filtration step and the solvent evaporated under vacuum. The polymers were then characterised without additional purification steps.

110 Nuclear Magnetic Resonance (NMR) Spectroscopy

111 ^1H -NMR analyses were performed on a JEOL JNM-ECS400A spectrometer at a frequency
112 of 400 MHz. CDCl_3 was used as NMR solvent for all synthesized polymers.

113 Gel Permeation Chromatography (GPC)

114 GPC was carried out using a PSS SDV High set composed of 3 analytical columns ($300 \times$
115 8 mm , particle diameter $5 \mu\text{m}$) of 1000, 1000×10^5 and 10^6 \AA pore sizes, plus guard column
116 (Polymer Standards Service GmbH, PSS) installed in a PSS SECcurity SEC system.
117 Elution was with THF at 1 mL min^{-1} with a column temperature of $23 \text{ }^\circ\text{C}$ and detection by
118 refractive index. $20 \mu\text{L}$ of a $\sim 2 \text{ mg mL}^{-1}$ sample in THF, adding a drop of toluene as
119 reference standard, was injected for each measurement and eluted for 50 min. Calibration
120 was carried out in the molecular weight range 370-2520000 Da using the ReadyCal
121 polystyrene standards supplied by Sigma Aldrich and referenced to the toluene peak.

122 Differential Scanning Calorimetry (DSC)

123 Traditional DSC analyses were performed on a TA Instruments Q2000 under nitrogen
124 atmosphere. The heating rate used was $5 \text{ }^\circ\text{C min}^{-1}$ over a T range of -60 - $200 \text{ }^\circ\text{C}$. Cooling
125 rate was set at $5 \text{ }^\circ\text{C min}^{-1}$ over the same T range. Sample mass was of 5-10 mg for all
126 measured samples. The polymer's melting ranges were calculated from the second
127 heating scan using the value at peak maximum option.

128 Thermogravimetric analysis (TGA)

129 TGA was performed on a PL Thermal Sciences STA 625 thermal analyzer. $\sim 10 \text{ mg}$ of
130 sample was weighted in an aluminium pan. The sample was then placed into the furnace
131 with a N_2 flow of 100 mL min^{-1} and heated from 21 to $625 \text{ }^\circ\text{C}$ at a heating rate of $10 \text{ }^\circ\text{C}$
132 min^{-1} . From the TGA profiles the temperatures at 5% and 50% mass loss (TD5 and TD50)
133 were subsequently determined.

134 Rheology measurements

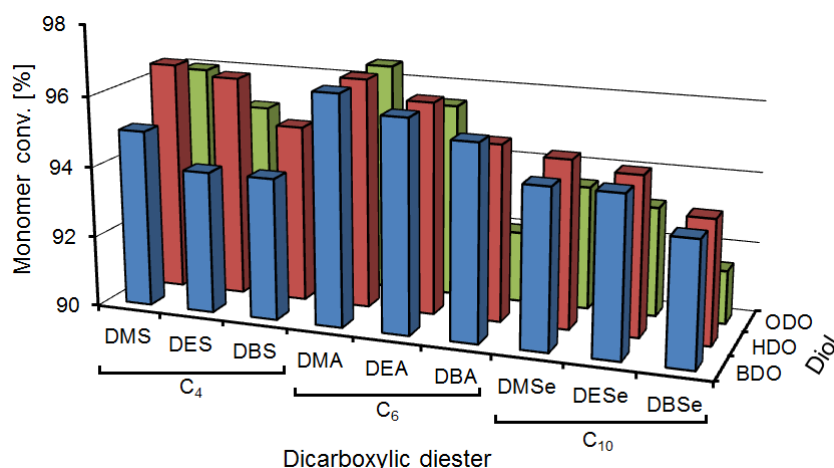
135 Rheological rotational measurement under controlled shear rate were recorded using a
 136 Brookfield R/S-CPS+ Rheometer fitted with a Peltier P-TS2 temperature controller and
 137 cone-plate geometry where the cone used had a 25 mm diameter, 2° angle and 0.045 mm
 138 gap height. The polymer samples were ground with a pestle and mortar prior to being
 139 loaded onto the pre-heated (80 °C) bottom plate. In all cases, a constant shear rate of 100
 140 s⁻¹ was maintained throughout the experiments with each polymer melt being initially held
 141 at 80 °C (under shear) for 120 s before the temperature was decreased by 0.01 °C s⁻¹.

142

143 Results and Discussion

144 The CaLB-catalysed polycondensation of methyl-, ethyl- and butyl- succinic (C₄), adipic
 145 (C₆) and sebacic (C₁₀) dicarboxylic acids esters with linear diols having a carbon chain
 146 length of 4, 6 or 8 atoms were investigated. In order to be environmentally friendly, all
 147 reactions were performed in bulk (i.e. solvent-less). All monomers were melted at 85 °C
 148 before adding the biocatalyst that allowed the initiation of the step-growth
 149 polycondensation. From the ¹H-NMR spectra analysis it is possible to observe that for all
 150 reactions excellent monomer conversion values (>90%) were obtained with the methyl and
 151 ethyl esters consistently giving slightly higher conversions than the equivalent butyl esters
 152 for all the dicarboxylic diesters investigated (Figure 1).

153

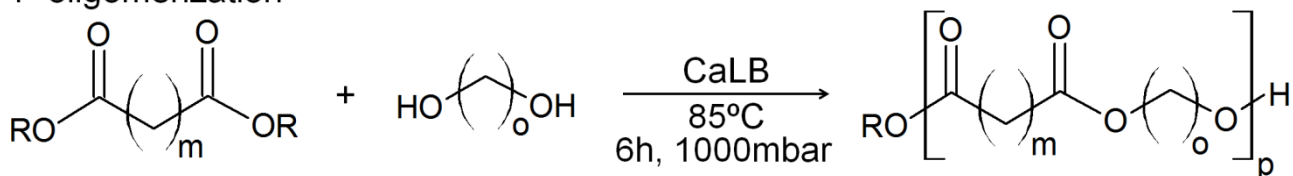


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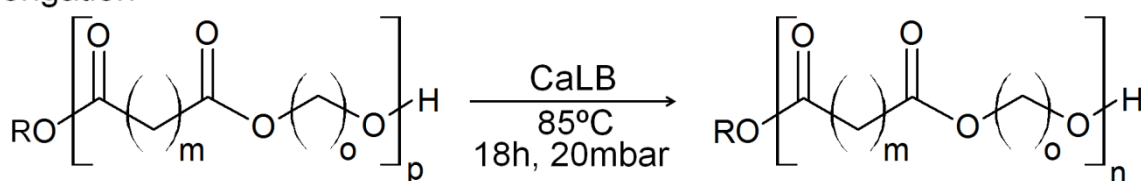
155 Figure 1. ¹H-NMR analysis of the crude polycondensation products after CaLB-catalyzed polycondensation
 156 reactions after 24 h. Reactions without catalyst led to no detectable transesterification products. All reactions
 157 were performed in duplicates. Statistical analysis reporting the mean value ± standard deviation for all
 158 reactions can be found in ESI, Figure S1 and the complete spectra assignment in Figure S5.
 159

160 The 2-step polycondensation protocol (1st-oligomerization and 2nd-chain elongation)
 161 reported in Scheme 3 was found suitable for all the reactions (see Table 1). The
 162 polycondensation reactions occur via transesterification of dicarboxylic acids diesters with
 163 aliphatic diols and follow a step-growth polymerisation mechanism (the same as some
 164 other similar synthetic polymers such as polyamides and polyurethanes). In fact with low
 165 DP (~5-10) the conversion is already >80-90% and further increase in DP (two fold)
 166 influences only minor the monomer conversion. This allows the initial oligomerisation of
 167 most of the monomers without loss of any of the starting compounds. After oligomerisation
 168 is achieved, vacuum is applied to effectively remove the alcohol by-product and enables
 169 the elongation of the polymeric chain length. The biocatalyst is indeed able to catalyse
 170 synthetic reactions starting from long polymeric chains as recently reported by Vastano et
 171 al. who coupled dimethyl itaconate and poly(ethylene glycol) to poly(hydroxyalkanoates)
 172 having M_n of 50188 Da and M_w of 117720 Da produced using an engineered strain of
 173 Escherichia Coli [21].
 174

1- oligomerization



2- elongation



R= -methyl, -ethyl or -butyl

o= C₄, C₆ or C₈

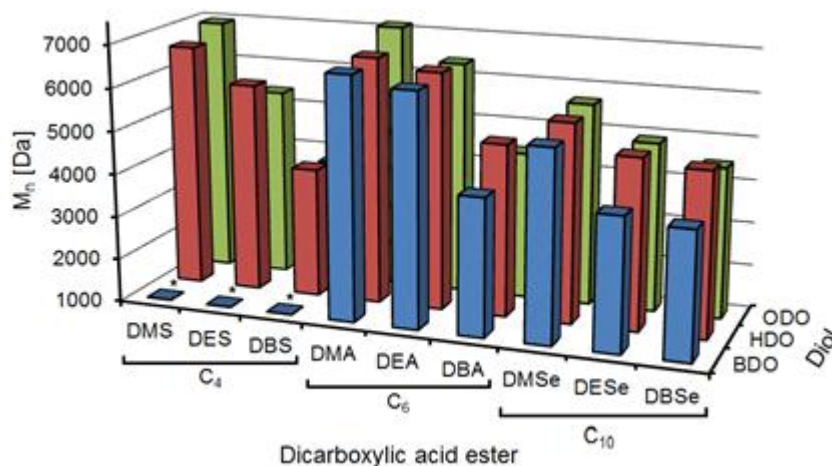
m= C₂, C₄ or C₈

175 Scheme 3. The 2-step polycondensation protocol used for the synthesis of various aliphatic-aliphatic
 176 polyesters via enzymatic catalysis in bulk (n>p).
 177

178

179 The number average molecular weight (M_n) (Figure 2) and the weight average molecular
 180 weight (M_w) (see Figure S2 in ESI) of the polymers were determined via GPC analysis.
 181 The M_n trend reported in Figure 2 agreed well with the $^1\text{H-NMR}$ spectroscopy monomer
 182 conversion analysis reported in Figure 1. The methyl and ethyl diesters lead to the
 183 synthesis of polymers having higher molecular weight relative to the butyl diesters. This
 184 effect of the size of the alkyl group of the diester (-methyl, -ethyl and -butyl) becomes less
 185 marked when increasing the diester internal chain length from C_6 to C_{10} . The lower
 186 conversions and molecular masses obtained using butyl diesters is most probably due to
 187 the relative boiling points of the alcohol by-products with methanol (BP= 64.7 °C) and
 188 ethanol (BP= 78.4 °C) being significantly less volatile than butanol (BP= 117.7 °C) and
 189 therefore easier to remove from the reaction system, especially when considering a
 190 reaction temperature of 85 °C was used. The polymers synthesized using the shortest
 191 dicarboxylic esters, dimethyl succinate (DMS), diethyl succinate (DES) and dibutyl
 192 succinate (DBS) together with 1,4-butanediol (BDO) were not soluble in the GPC mobile
 193 phase (THF) and therefore were not characterised using this technique.

194



195
 196 Figure 2. Number average molecular weights (M_n) calculated via GPC analysis of the crude
 197 polycondensation products using a 370-2520000 Da polystyrene calibration curve. Reactions without
 198 catalyst did not lead to detectable polyesterification products. All reactions were performed in duplicates.
 199 Statistical analysis reporting the mean value \pm standard deviation for all reactions can be found in ESI,
 200 Figure S2. *=reactions non soluble in the GPC mobile phase (THF).

201

202 The generally observed trend for the synthesised polymers based on HDO and ODO
203 having higher molecular masses than the BDO equivalents (with the exception of the
204 adipate-based polyesters that showed similar molecular masses) reflects very well the
205 data reported by Feder and Gross for the polycondensation of sebacic acid and various
206 diols using N435 at 70 °C. [15] In this previous work, a comparison between adipic and
207 sebacic acid was made and fully agrees with our observation that the adipic acid diesters
208 lead to higher molecular weight products than the sebacic diesters when methyl and ethyl
209 esters were used while no significant differences were observed when butyl esters were
210 used. It was noticed that CaLB, when operating in a solventless reaction system, led to
211 smaller differences between the different ester chains than when using cutinases and
212 serin-hydrolase enzymes belonging to the same family. [22] In fact, whereas CaLB leads
213 to similar conversions with BDO, HDO and ODO, cutinases have a preferred substrate.
214 Cutinase from *Humicola insolens* (70 °C, in bulk, diol diacid 1:1 mol/mol, 1% w/w of
215 enzyme-to monomer, application of vacuum after 2 h) produces polyesters with higher
216 molecular weight when ODO is used as a diol [15] while cutinase 1 from *Thermobifida*
217 *cellulosilytica* (Thc_cut1) (70 °C, in bulk, diol diacid 1:1 mol/mol, 10% w w⁻¹ CaLB
218 immobilized onto methacrylic resin-to monomer, application of vacuum for 24 h) prefers
219 BDO, leading to only short oligomers when HDO and ODO are used in combination with
220 dimethyl adipate. [23] For Thc_cut1, it was recently reported that the structure of the diol is
221 relevant for the chain length, but the conversion is not much affected. [17]

222 In order to be able to analyze the differences between all reactions, including the formation
223 of aforementioned insoluble poly(butylene succinate), some selected reactions (reported in
224 Table 1) were run for only 6 h at 85 °C and atmospheric pressure in order to stop the
225 reaction after the initial oligomerisation phase.

226

Table 1. Comparison of the CaLB-catalyzed polycondensation reactions after 6 h at 85 °C and 1000 mbar and 24 h.

Diester	Diol	Time [h]	Conv. [%] [#]	M _n [Da] [*]	M _w [Da] [*]	Đ [*]	M ₀	DP _n
DES	BDO	6	89	851	1094	1.29	172.2	4.9
		24	94	ns	ns	ns		x
	HDO	6	92	2164	2398	1.26	200.2	9.8
		24	96	5853	10439	1.79		29.2
DMA	BDO	6	89	2227	2843	1.31		10.1
		24	97	6626	11549	1.71		33.1
DBA	HDO	6	83	2018	2247	1.07	228.3	9.5
		24	95	4978	8018	1.62		21.8

[#] Calculated via ¹H-NMR based on the conversion of the diol.

^{*} Calculated via GPC using a - Da polystyrene calibration curve and toluene as internal standard.

ns= non soluble in THF

227

228 From the collected data it is possible to see that the insolubility of poly(butylene succinate)
 229 in THF, even after stirring for several days, is not due to the high molecular weight of the
 230 polymers. It is well known that the dissolution of polymers depends not only on their
 231 physical properties, but also on their polarity, molecular weight, branching, crosslinking
 232 degree and crystallinity. [24] In this particular instance, given the crystalline nature of all
 233 produced polymers it is probable that the insolubility of the succinate-based aliphatic
 234 oligoesters after 24 h reaction was indeed a result of high crystallinity rather than the
 235 oligomers as evidenced by the molecular masses data reported in Table 1 (and the DSC
 236 data reported later). The calculation of the DP of the reactions after 6 and 24 h fully
 237 confirms the trend previously reported via ¹H-NMR spectroscopy and GPC for all of the
 238 other reactions carried out (Table 1).

239 In addition to monomer conversion and molecular weight analysis, differential scanning
 240 calorimetry analysis for the determination of the polymers' melting points and
 241 thermogravimetric analysis for the determination of the mass loss were performed. Figure
 242 3 reports the melting temperatures of all the synthesized polymers when the standard
 243 protocol (6 h at 1000 mbar, 18 h at 20 mbar, 85 °C, 10% w w⁻¹ CaLB, 24 h total reaction
 244 time) was applied. It is possible to observe a general trend where an increase in polymer
 245 melting point aligns with increasing carbon chain length of the diester, where C₄-based

polymers have lower melting points than the C₁₀-based polymers. This is due to the crystallinity of the C₄-based polymers as discussed above. A similar increase is also observed when increasing the diol carbon chain length from C₄ (BDO, blue) to C₈ (ODO, green). In this case, increasing the dicarboxylic ester length from C₄ to C₈ reduces the melting point differences among the produced polymers, analogous to number average molecular weight observations plotted in Figure 2.

Table 2. Calculated percentage of M_n reduction for any diethyl ester chain in relation to the alkyl group in relation to 1,8-octanediol.

		Diester end group		
		Me	Et	Bu
Diester carbon chain lenght	C4	6959 g mol ⁻¹ * 100%	5380 g mol ⁻¹ * 23%	3796g mol ⁻¹ * 45%
	C6	7141g mol ⁻¹ * 100%	6390 g mol ⁻¹ * 11%	4413 g mol ⁻¹ * 38%
	C10	5677 g mol ⁻¹ * 100%	4908 g mol ⁻¹ * 14%	4478 g mol ⁻¹ * 21%

* Calculated via GPC using a - Da polystyrene calibration curve and toluene as internal standard.

In Table 2 we reported the % of M_n reduction for any diethyl ester chain in relation to the alkyl group in relation to 1,8-octanediol. As we can see from the collected data, the methyl ester (Me) (taken as 100%) is always the best performing one, followed by the diethyl (Et) and the dibutyl (Bu) ones. Also to notice that the differences in the M_ns decrease for the Bu end group are lower with the increase of the carbon chain length of the diester for the considered reaction.

Further to this, the difference in polymer melting temperature between the different diol chain lengths becomes less pronounced when increasing the diester chain length (Figure 3).

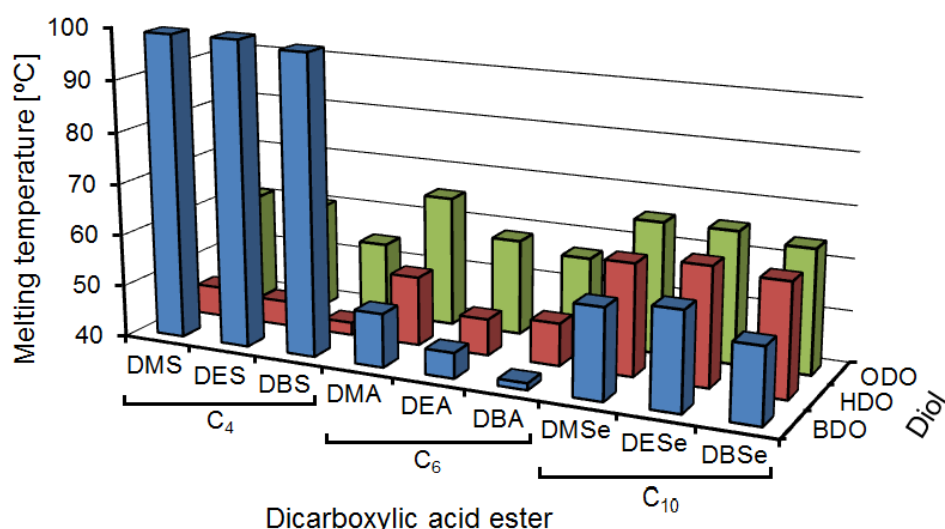


Figure 3. DSC determination of the polymer's melting points (see Table S1 for details).

The inert atmosphere (N_2) thermogravimetric profile of the polymers presented in Figure 4 is also consistent with the previously discussed molecular mass data. In fact, with the increase of the polymer's M_n and the increase of the diol's carbon chain length (see Figure 4a insert) the degradation temperature increases accordingly (Figure 4a).

A comparison of the polymerisations conducted using the same diol (1,8-octanediol, ODO, C_8) with three different diesters having methanol, ethanol or butanol alkyl groups on the monomers also show a remarkable difference in the obtained M_n (see discussion above and Figure 4b insert), with the dimethyl adipate-derived polyester proving more thermally stable than the diethyl equivalent, with this being more stable than the dibutyl. This indicates that the selection of the diester remains highly important for these transesterification reactions since the volatility of the leaving group proved to be a key point in determining the extent to which the polyester chain grows. From the TGA profiles the temperatures at 5% and 50% mass loss (TD5 and TD50) were subsequently determined and are reported in Table S2 in ESI.

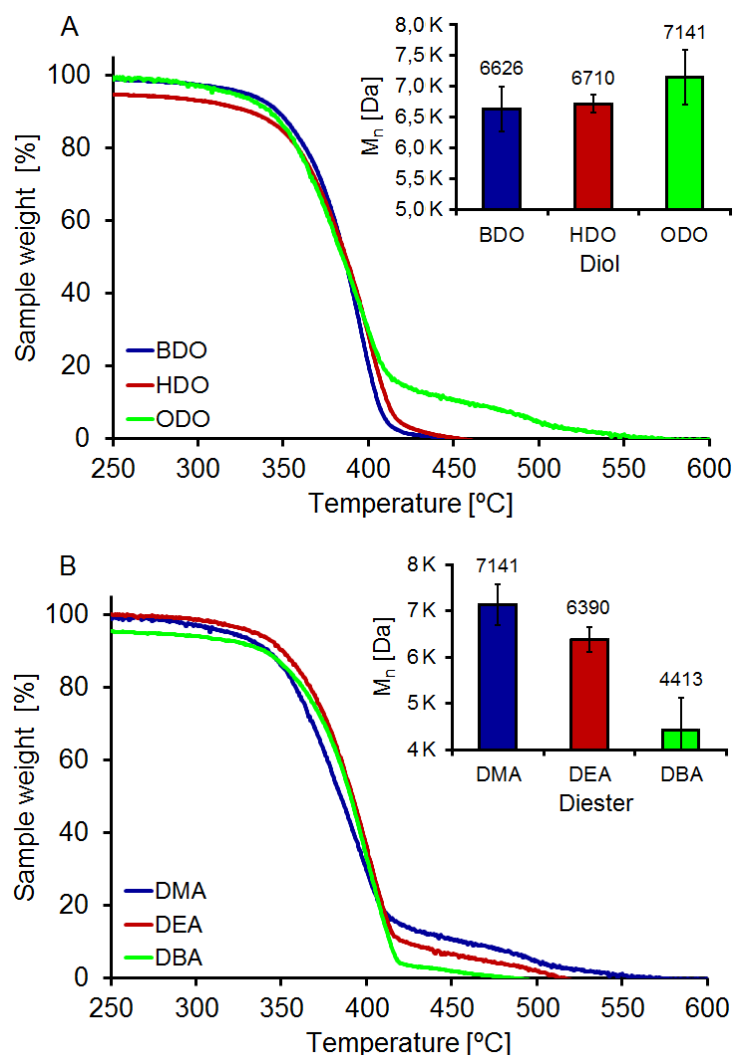


Figure 4. TGA analysis of A) polymers obtained from the polycondensation of dimethyl adipate and the three different aliphatic diols 1,4-butanediol (C4, BDO), 1,6-hexanediol (C6, HDO) and 1,8-octanediol (C8, ODO) and B) polymers obtained from the polycondensation of ODO with three different diesters dimethyl adipate (DMA), diethyl adipate (DEA) and dibutyl adipate (DBA).

Rheology was used to further characterise the polymers and identify differences in their viscosity. Figure 5 shows the viscosity change of the polymer melts based on a decrease of temperature for poly(1,6-hexylene adipate) synthesized from dimethyl adipate (blue), diethyl adipate (red) and dibutyl adipate (green). In all cases, viscosity steadily rose with decreasing temperature until the onset of freezing was reached, at which point viscosity increased rapidly. This further exemplifies how the differing molecular weights (see Fig.5 insert) lead to different viscosity profiles. The shorter chain polymer behaves remarkably different compared with the other two polymers with similar M_n values. Comparative studies of the presented reactions in organic media are needed in order to better elucidate

the effect of viscosity on the reaction progression. Moreover, we noted that the viscosity of any initial mixture appeared to be below the lower detection limit (ca. <0.2 Pa.s) of the apparatus when used under the same conditions.

The produced polyesters can find applications that range from the biomedical field (as carriers in protein- and peptide-delivery systems) [25] and, if further coupled with poly(ethylene glycol), for the production of eco-friendly water-soluble polymers and coatings [26].

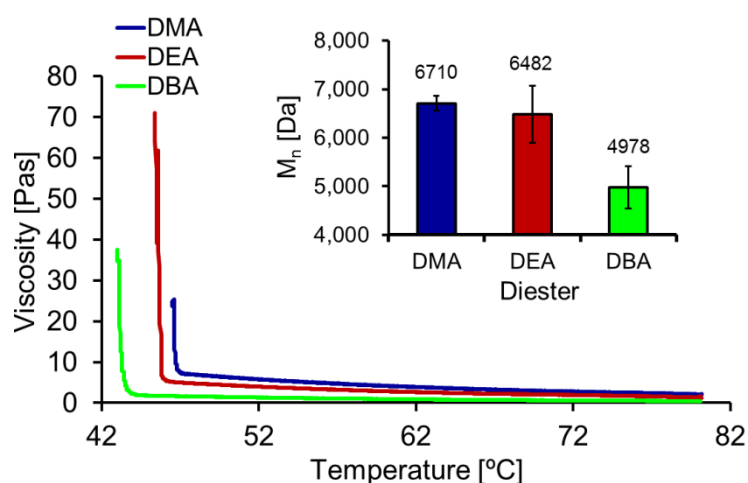


Figure 5. Change of the polymer's viscosity relative to the temperature decrease over time for different poly(hexamethylene adipate) polyesters whose chain length alters due to the chose dialkyl adipate monomer (dimethyl adipate – DMA, diethyl adipate – DEA and dibutyl adipate – DBA).

Conclusions

From the collected data we can conclude that CaLB is an effective catalyst for the synthesis of a range of aliphatic polyesters using a solventless reaction system. Polyesters based on diesters with an internal carbon chain length between 4 (succinate) and 10 (sebacate) and diols with a carbon chain length between 4 (1,4-butanediol) and 8 (1,8-octanediol) were successfully synthesized via transesterification. A strong effect of the selected alkyl group of the diester (dimethyl, diethyl and dibutyl) was observed for all polyesters. Lower molecular weights and monomer conversions were obtained using dibutyl esters since it proved more difficult to remove the butanol by-product during the

319 reaction due to its higher boiling point (relative to methanol and ethanol). DSC, TGA and
320 rheology measurements confirm the trend showing a large difference in degradation
321 temperatures and viscosity of the melts due to the polymers' molecular weight. This work
322 has sets the basis for efficient biocatalyzed syntheses of aliphatic polyesters having M_n of
323 ~ 6 and M_w of ~ 13 KDa and with monomers conversions $>94\%$. This also represents the
324 first systematic study of different dialkyl aliphatic diesters with various diols under identical
325 reaction conditions.

326

327 Conflicts of interest

328 The authors declare no conflicts of interest.

329

330 Acknowledgements

331 A. P. thanks the FWF Erwin Schrödinger fellowship (grant agreement J 4014-N34) for
332 financial support. M. H. S. thanks Academy of Finland for funding (grant #296547). T. J. F.,
333 J. H. C. and J. W. C. would like to thank the UK Engineering and Physical Sciences
334 Research Council (EPSRC, grant EP/L017393/1) and the Biotechnology and Biological
335 Sciences Research Council (BBSRC, grant BB/N023595/1) for funding their involvement in
336 this research.

337

338 Notes and references

339 Electronic Supplementary Information (ESI) available: Details of: ^1H -NMR spectroscopy
340 conversions (Fig. S1), M_n (Fig.S3) and M_w (Fig. S2 and S4) GPC data, DSC-determined
341 polymer's melting temperatures (Table S1) and TD5 and 50 (Table S2) are reported in
342 ESI. All data gathered via funding from grants EP/L017393/1 and BB/N023595/1 is
343 contained within the manuscript or the ESI.

- 344 1. S. Kobayashi, H. Uyama and S. Kimura, Enzymatic Polymerization, Chem. Rev.
345 101 (2001) 3793-3818.
- 346 2. S. Kobayashi, New developments of polysaccharide synthesis via enzymatic
347 polymerization, Proc. Jpn. Acad. Ser. B Phys. Biol. Sci. 83 (2007) 215-247.
- 348 3. D. Huber, A. Pellis, A. Daxbacher, G. S. Nyanhongo and G. M. Guebitz,
349 Polymerization of Various Lignins via Immobilized Myceliophthora thermophila Laccase
350 (MtL), Polymers 8 (2016) 280.
- 351 4. A. Pellis, E. Herrero Acero, V. Ferrario, D. Ribitsch, G. M. Guebitz and L. Gardossi.,
352 The Closure of the Cycle: Enzymatic Synthesis and Functionalization of Bio-Based
353 Polyesters, Trends Biotechnol. 34 (2016) 316-328.
- 354 5. A. Pellis, E. Herrero Acero, L. Gardossi, V. Ferrario and G. M. Guebitz, Renewable
355 building blocks for sustainable polyesters: new biotechnological routes for greener plastics,
356 Polym. Int. 65 (2016) 861-871.
- 357 6. S.S. Ribeiro, C. Raminelli and A. L. M. Porto, Enzymatic resolution by CALB of
358 organofluorine compounds under conventional condition and microwave irradiation. J.
359 Fluor. Chem.154 (2013) 53-59.
- 360 7. O. Kirk, F. Björkling, S. E. Godtfredsen and T. O. Larsen. Fatty Acid Specificity in
361 Lipase-Catalyzed Synthesis of Glucoside Esters, Biocatalysis 6 (1992) 127-134.
- 362 8. S. Strompen, M. Weiß, H. Gröger, L. Hilterhaus and A. Liese, Development of a
363 Continuously Operating Process for the Enantioselective Synthesis of a β -Amino Acid
364 Ester via a Solvent-Free Chemoenzymatic Reaction Sequence, Adv. Synth. Catal. 355
365 (2013) 2391-2399.
- 366 9. A. Pellis, G. M. Guebitz and T. J. Farmer, On the Effect of Microwave Energy on
367 Lipase-Catalyzed Polycondensation Reactions, Molecules 21 (2016) 1245.

- 368 10. A. Pellis, L. Corici, L. Sinigoi, N. D'Amelio, D. Fattor, V. Ferrario, C. Ebert and L.
369 Gardossi, Towards feasible and scalable solvent-free enzymatic polycondensations:
370 integrating robust biocatalysts with thin film reactions, *Green Chem.* 17 (2015) 1756-1766.
- 371 11. H. Uyama, M. Kuwabara, T. Tsujimoto and S. Kobayashi, Enzymatic synthesis and
372 curing of biodegradable epoxide-containing polyesters from renewable resources,
373 *Biomacromolecules* 4 (2003) 211-215.
- 374 12. Y. Jiang, A. J. J. Woortman, G. O. R. Alberda van Ekenstein and K. Loos, A
375 biocatalytic approach towards sustainable furanic–aliphatic polyesters. *Polym. Chem.* 6
376 (2015) 5198-5211.
- 377 13. Y. Jiang, A. J. J. Woortman, G. O. R. Alberda van Ekenstein, D. M. Petrović and K.
378 Loos, Enzymatic Synthesis of Biobased Polyesters Using 2,5-Bis(hydroxymethyl)furan as
379 the Building Block. *Biomacromolecules* 15 (2014) 2482-2493.
- 380 14. Y. Jiang, D. Maniar, A. J. J. Woortman and K. Loos, Enzymatic synthesis of 2,5-
381 furandicarboxylic acid-based semi-aromatic polyamides: enzymatic polymerization
382 kinetics, effect of diamine chain length and thermal properties. *RSC Adv.* 6 (2016) 67941-
383 67953.
- 384 15. D. Feder and R. A. Gross, Exploring Chain Length Selectivity in HIC-Catalyzed
385 Polycondensation Reactions. *Biomacromolecules* 11 (2010) 690-697.
- 386 16. E. Stavila, R. Z. Arsyi, D. M. Petrovic and K. Loos, *Fusarium solani* pisi cutinase-
387 catalyzed synthesis of polyamides. *Eur. Polym. J.* 49 (2013) 834-842.
- 388 17. A. Pellis, V. Ferrario, M. Cespugli, L. Corici, A. Guarneri, B. Zartl, E. Herrero Acero,
389 C. Ebert, G. M Guebitz and L. Gardossi, Fully renewable polyesters via polycondensation
390 catalyzed by *Thermobifida cellulosilytica* cutinase 1: an integrated approach, *Green Chem.*
391 19 (2017) 490-502.

- 392 18. M. Hunsen, A. Abul, W. Xie and R. A. Gross, Humicola insolens Cutinase-Catalyzed
393 Lactone Ring-Opening Polymerizations: Kinetic and Mechanistic Studies,
394 Biomacromolecules 9 (2008) 518-522.
- 395 19. A. Pellis, M. Vastano, F. Quartinello, E. Herrero Acero and G. M. Guebitz, His-Tag
396 Immobilization of Cutinase 1 From *Thermobifida cellulosilytica* for Solvent-Free Synthesis
397 of Polyesters, Biotechnol. J. 12 (2017) 1700322
- 398 20. T. J. Farmer and M. Mascal, Platform Molecules. in Introduction to Chemicals from
399 Biomass (eds. J. H. Clark and F. Deswarte) 89-155 (John Wiley & Sons, Ltd) 2015.
- 400 21. M. Vastano, A. Pellis, B. Immirzi, G. Dal Poggetto, M. Malinconico, G. Sannia, G. M.
401 Guebitz and C. Pezzella, Enzymatic production of clickable and PEGylated recombinant
402 polyhydroxyalkanoates, Green Chem. 19 (2017) 5494-5504.
- 403 22. V. Ferrario, A. Pellis, M. Cespugli, G. M. Guebitz and L. Gardossi, Nature Inspired
404 Solutions for Polymers: Will Cutinase Enzymes Make Polyesters and Polyamides
405 Greener?, Catalysts 6 (2016) 205.
- 406 23. A. Pellis, V. Ferrario, B. Zartl, M. Brandauer, C. Gamerith, E. Herrero Acero, C.
407 Ebert, L. Gardossi and G. M. Guebitz, Enlarging the tools for efficient enzymatic
408 polycondensation: structural and catalytic features of cutinase 1 from *Thermobifida*
409 *cellulosilytica*. Catal. Sci. Technol. 6 (2016) 3430-3442.
- 410 24. C. S. Brazel and S. L. Rosen, Fundamental Principles of Polymeric Materials, 3rd
411 Edition, Wiley, 2012.
- 412 25. S. Zhou, X. Deng, X. Li, W. Jia and L. Liu, Synthesis and Characterization of
413 Biodegradable Low Molecular Weight Aliphatic Polyesters and Their Use in Protein-
414 Delivery Systems, J. App. Polym. Sci. 91 (2004) 1848-1856.
- 415 26. Y. Ikada and H. Tsuji, Biodegradable polyesters for medical and ecological
416 applications, Macromol. Rapid Commun. 21 (2000) 117-132.