



UNIVERSITY OF LEEDS

This is a repository copy of *The effects of xylanase on grower pig performance, concentrations of volatile fatty acids and peptide YY in portal and peripheral blood.*

White Rose Research Online URL for this paper:  
<http://eprints.whiterose.ac.uk/132132/>

Version: Accepted Version

---

**Article:**

Taylor, AE, Bedford, MR and Miller, HM [orcid.org/0000-0003-1440-0454](https://orcid.org/0000-0003-1440-0454) (2018) The effects of xylanase on grower pig performance, concentrations of volatile fatty acids and peptide YY in portal and peripheral blood. *animal*, 12 (12). pp. 2499-2504. ISSN 1751-7311

<https://doi.org/10.1017/S1751731118000277>

---

(c) The Animal Consortium 2018, This article has been published in a revised form in *animal* <https://doi.org/10.1017/S1751731118000277>. This version is free to view and download for private research and study only. Not for re-distribution, re-sale or use in derivative works.

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>

1 **The effects of xylanase on grower pig performance, concentrations of volatile**  
2 **fatty acids and peptide YY in portal and peripheral blood.**

3

4 A. E. Taylor <sup>1</sup>, M. R. Bedford <sup>2</sup>, and H. M. Miller <sup>1</sup>

5

6 <sup>1</sup> *Faculty of Biological Sciences, University of Leeds, Leeds, LS2 9JT, UK*

7 <sup>2</sup> *AB Vista Feed Ingredients Ltd, 3 Woodstock Court, Marlborough, SM8 4AN, UK*

8

9

10 Corresponding author: Amy Taylor. Email: a.e.taylor@leeds.ac.uk

11

12

13 Short title: Optimum inclusion level of xylanase in pig diets

14

15

16

17

18

19

20

21

22

23 **Abstract**

24 Non-starch polysaccharides (NSP) present in wheat and barley can act as anti-  
25 nutrients leading to an increase in digesta viscosity and a reduction in nutrient  
26 digestibility. Xylanase, an NSP degrading enzyme, has been shown to increase  
27 nutrient digestibility in pigs. The objectives of this study were: 1) to identify the  
28 optimum inclusion level of xylanase in grower pig diets by measuring the effect of  
29 increasing enzyme levels on growth performance, the concentration of volatile fatty  
30 acids (VFA) and peptide YY concentration in portal and peripheral blood of grower  
31 pigs and 2) to increase our understanding of the interrelationships between xylanase  
32 inclusion, VFA production and peptide YY secretion. Five hundred and twelve grower  
33 pigs ((Large White x Landrace) x MAXGRO) were allocated to pens creating 32  
34 replicates of 4 pigs per pen per treatment. Pigs were allocated to trial weighing  $14.2$   
35  $\pm 0.31$  kg and remained on trial until approximately  $41.5 \pm 3.31$  kg. The experiment  
36 was a dose response design with 4 inclusion levels (0, 8 000, 16 000 or 32 000 BXU/  
37 kg) of xylanase (Econase XT). Diets were cereal based wheat, barley mix formulated  
38 to meet or exceed the nutrient requirements of grower pigs. Body weight and feed  
39 intake were recorded to calculate growth performance. Pen faecal samples were  
40 collected to estimate DM, organic matter (OM) and crude fibre (CF) apparent total  
41 tract digestibility. At the end of the trial 16 pigs per treatment were euthanised by  
42 schedule 1 procedures. Peripheral and portal blood samples were collected for  
43 peptide YY and VFA analysis. The addition of xylanase to the diet had no effect on  
44 growth performance, DM, OM or CF total tract digestibility, however xylanase tended  
45 to have a quadratic effect on ileum pH with higher pH values recorded for pigs fed a  
46 diet supplemented with 8 000 and 16 000 BXU/kg xylanase ( $P < 0.1$ ). Xylanase had  
47 no effect on peptide YY levels or VFA concentration. Total VFA concentration was

48 higher in portal compared to peripheral blood ( $P < 0.05$ ). In conclusion, the addition  
49 of xylanase had no effect on grower pig performance, nutrient digestibility, VFA  
50 concentration or peptide YY concentration when fed up to 32 000 BXU/ kg over a 35  
51 d period. Pig performance was good for all treatments throughout the trial suggesting  
52 that diet quality was sufficient thus there were no beneficial effects of adding  
53 xylanase.

54

55 **Keywords:** Digestibility, Enzyme, Fibre, Growth performance, Nutrition

56

## 57 **Implications**

58 The presence of non-starch polysaccharides (NSP) in wheat exerts anti nutritional  
59 effects in pigs, increasing digesta viscosity and reducing nutrient digestibility through  
60 encapsulation which results in poor feed efficiency and growth. The addition of NSP  
61 degrading enzymes such as xylanase to wheat based diets can help to reduce these  
62 effects. However the addition of xylanase as high as 32 000 BXU/ kg to wheat-barley  
63 cereal based diets formulated to meet or exceed the nutrient requirements of grower  
64 pigs had no effect on growth performance or nutrient utilisation. This suggests that  
65 the addition of xylanase to good quality wheat based grower pig diets has no  
66 beneficial effects.

67

## 68 **Introduction**

69 Exogenous enzymes are used in livestock diets in order to maximise utilisation of  
70 nutrients and improve feed efficiency. The presence of non-starch polysaccharides

71 (NSP) in wheat (predominately arabinoxylans) have been shown to have anti  
72 nutritional effects in pigs (Yin *et al.*, 2000, Northey *et al.*, 2007). Non-starch  
73 polysaccharides increase digesta viscosity and reduce nutrient digestibility through  
74 encapsulation (Kim *et al.*, 2005). The incorporation of NSP degrading enzymes can  
75 help to alleviate the negative effects of NSP. Xylanase, an NSP degrading enzyme,  
76 works by removing the nutrient encapsulating effects of the cell wall in feed by the  
77 breakdown of long-chain arabinoxylans, releasing nutrients for digestion by the  
78 animal and by reducing digesta viscosity (Masey O'Neill *et al.*, 2014b). An additional  
79 mechanism has been suggested as the addition of xylanase to maize–soybean  
80 based broiler diets consistently increased serum peptide YY (Singh *et al.*, 2012)  
81 suggesting a neuro-hormonal mechanism through the production of the satiety-  
82 related gut hormone peptide YY. Peptide YY slows down gastric emptying increasing  
83 gastric digestion of nutrients (Allen *et al.*, 1984) with a consequential increase in feed  
84 efficiency. Arabinoxylo-oligosaccharides, produced from enzymatic hydrolysis of  
85 arabinoxylans, are non-digestible oligosaccharides with prebiotic activity that can be  
86 fermented by intestinal microbiota (Grootaert *et al.*, 2009, Neyrinck *et al.*, 2012),  
87 resulting in an increase in volatile fatty acid (VFA) production, predominately acetate,  
88 propionate and butyrate (Goodlad *et al.*, 1987). Cucho *et al.* (2000) found that ileal  
89 VFA infusion increased peptide YY concentrations in pigs suggesting the production  
90 of VFAs stimulates the release of the gut hormone peptide YY from entero-endocrine  
91 cells.

92 The use of xylanase in pig diets has been extensively studied, however inconsistent  
93 results have been reported. Passos *et al.* (2015) found that xylanase  
94 supplementation from 0 to 1 400 LXU/kg enhanced ileal digestibility of neutral  
95 detergent fibre (NDF), dry matter (DM), organic matter (OM) and energy when fed to

96 grower pigs. Woyengo *et al.* (2008) found that xylanase supplementation had no  
97 effect on grower pig performance, however they did find that xylanase  
98 supplementation at 4 000 XU/kg increased amino acid ileal digestibility and Ca  
99 digestibility. Olukosi *et al.* (2007) found no effect of xylanase supplementation on  
100 performance or nutrient digestibility when added to wheat-barley-soybean meal-  
101 based diet for grower pigs. The inconsistent results reported may result from  
102 differences in the level of xylanase used. Inconsistent results along with limited  
103 information on the role of a xylanase enzyme on VFA production in grower pigs and  
104 the activity of peptide YY suggests there is a need for further investigation.

105 The current study was designed to test the hypotheses that hydrolysis of NSPs by  
106 xylanase creates shorter chain oligosaccharides altering caecal fermentation.

107 Changes in caecal fermentation will alter VFA production which in turn will affect  
108 peptide YY secretion. The objectives of this study were: 1) to identify the optimum  
109 inclusion level of xylanase (Econase XT) in grower pig diets by measuring the effect  
110 of increasing enzyme levels on growth performance, the concentration of VFA and  
111 peptide YY concentration in portal and peripheral blood of grower pigs and 2) to  
112 increase our understanding of the interrelationships between xylanase inclusion,  
113 VFA production and peptide YY secretion in grower pigs.

114

## 115 **Material and methods**

116

117 *Animals*

118 This experiment was carried out at the University of Leeds, Spen farm. This  
119 experiment received ethical approval from the University of Leeds Animal Welfare  
120 and Ethical Review Committee. Five hundred and twelve grower pigs ((Large White  
121 x Landrace) x MAXGRO) were allocated to pens on the basis of litter of origin, sex  
122 and liveweight (32 reps of 4 treatments with 4 pigs per pen). Pigs were allocated to  
123 trial at 7 weeks of age ( $14.2 \pm 0.31$  kg) and remained on trial until 12 weeks of age  
124 (approximately  $41.5 \pm 3.31$  kg). Pigs were housed in fully slatted grower  
125 accommodation throughout the trial with 16 pens per room. Each pen measured 135  
126 cm x 155 cm. Heating and ventilation were controlled to maintain temperature at  
127 approximately 22°C (Dicam system). Pigs had free access to feed and water.

128

129 *Experimental design*

130 The experiment was a dose response design with four inclusion levels of xylanase  
131 (0, 8 000, 16 000 or 32 000 BXU/kg). The xylanase was a beta 1-4, endo-xylanase  
132 (Econase® XT, AB Vista, Marlborough, Wiltshire, UK) and its main function is to  
133 break down the fibre fraction from cereals, mainly arabinoxylan (European Food  
134 Safety Authority, 2008). Diets were a cereal based wheat, barley mix formulated to  
135 meet or exceed the nutrient requirements of grower pigs (BSAS, 2003) and  
136 contained 500 FTU Quantum Blue per kg diet (AB Vista, Marlborough, Wiltshire,  
137 UK). The diet composition is shown in Table 1. Titanium dioxide was added in all  
138 diets at 5 g/kg as an indigestible marker to determine apparent total tract digestibility  
139 of nutrients. Feed was analysed for crude protein, lysine, crude fibre, fat, ash,  
140 phosphorus and calcium (Sciantec Analytical Services Ltd. Stockbridge Technology

141 Centre, UK). Diets were also analysed for xylanase activity by ELISA method using  
142 Quantiplate Kits for Econase XT (Enzyme Services & Consultancy, Innovation &  
143 Technology Centre, Ystrad Mynach, UK).

144

#### 145 *Measurements*

146 Pigs were individually weighed at the start of the trial (7 weeks of age) and at 10 and  
147 12 weeks of age. Average daily intake (ADI), average daily gain (ADG) and feed  
148 conversion ratio (FCR) were recorded on a pen basis. Mean pen health scores were  
149 taken each morning throughout the experiment on a scale of 1-4 (1= no signs of poor  
150 health, 4= serious signs of poor health) as described by Taylor *et al.* (2013). Scores  
151 were assessed by the same trained individuals. A record was kept of any veterinary  
152 interventions and any mortality.

153 Pen faecal samples (250-300 g) were collected on the second last day of the trial.  
154 Samples from two pens of the same treatment were pooled to give 16 replicates per  
155 treatment and stored at -20 °C for subsequent analysis. All feed and pooled faecal  
156 samples underwent wet ash sample preparation (method 975.03 AOAC 2012) for  
157 analysis of TiO<sub>2</sub> (Short *et al.*, 1996). Digestibility values for DM, organic matter (OM)  
158 and crude fibre (CF) were calculated using TiO<sub>2</sub> as a marker. Briefly, each sample  
159 was oven dried at 102 °C for 24 hours. Samples were then ashed in a muffle furnace  
160 for 16 hours at 550 °C. Ashed samples were then digested in 7.4 M H<sub>2</sub>SO<sub>4</sub>. This  
161 solution was filtered and used for TiO<sub>2</sub> determination. Crude Fibre was determined  
162 using the Foss fibre Cap 2021 Fibre Analysis System (Foss Analytical, Hilleroed,  
163 Denmark).

164 At the end of the trial 16 pigs per treatment were euthanised by schedule 1  
165 procedures. Pigs were stunned using captive-bolt and then exsanguinated in



166 accordance with the Animals (Scientific Procedures) Act 1986. Pigs were selected by  
167 randomly choosing 16 of the 32 replicates. Within each replicate, one pig from each  
168 pen was selected based on performance. The performance of the sampled pig was  
169 as close to the mean for that treatment replicate as possible. Immediately following  
170 death, blood samples were collected from both portal and peripheral circulations into  
171 heparinized tubes. The blood was centrifuged at 2 000 x g, 4 °C for 15 minutes.  
172 Plasma was removed and stored at -20 °C prior to analysis. The gastrointestinal tract  
173 was removed and the pH of the distal ileum and caecum digesta were measured (HI-  
174 99163 Handheld Meat pH Meter, Hanna Instruments, UK).

175 Plasma samples were analysed for peptide YY concentration using a commercial  
176 ELISA kit (Pig Peptide YY (PYY) ELISA kit (CSB-EL019128PI)). The assay has high  
177 sensitivity for the detection of pig PYY with no significant cross-reactivity or  
178 interference between pig PYY and analogues. The intra-assay CV was <15% and  
179 the inter-assay CV was <15%. Plasma samples were also analysed for VFA  
180 concentrations (acetate, propionate, butyrate) using gas chromatography (Varian  
181 3400). Helium was used as carrier gas. Caproic acid was used as the internal  
182 standard. The sample was deproteinized using ultrafiltration (Amicon 0.5ml 10K;  
183 MerckMillipore, Darmstadt, Germany). Samples were spun at 14 000 x g for 20  
184 minutes. Phosphoric acid was then added to the ultrafiltrate. This ultrafiltrate was  
185 directly injected into the system.

186

### 187 *Statistical analysis of results*

188 Mean pen data for liveweight, feed intake, daily gain, FCR and pH were analysed by  
189 ANOVA using the general linear model (GLM) procedure SPSS, version 22 (IBM

190 SPSS Statistics for Windows, Version 20.0. Armonk, NY: IBM Corp). All data were  
191 tested for normality of residuals and outliers. Orthogonal polynomial contrasts were  
192 applied to test for linear and quadratic responses to the inclusion of xylanase (SPSS,  
193 version 22). Responses were considered significant when  $P \leq 0.05$ , and trends were  
194 noted when  $P \leq 0.10$ . The number of pigs treated with antibiotics were analysed by  
195 the non-parametric Kruskal–Wallis one-way ANOVA. Any pigs removed from the trial  
196 due to ill health were weighed before removal. An ANOVA was run to compare  
197 treatment effects on portal and peripheral levels of peptide YY and total VFA,  
198 whereas a Repeated-Measures ANOVA was performed to compare portal and  
199 peripheral concentrations. Peptide YY data were not normally distributed and were  
200 log transformed in order to normalise the data for analysis. Total VFA data were not  
201 normally distributed and were transformed using square root transformation in order  
202 to normalise the data for analysis. Post analysis, any data which had been  
203 transformed was then reverse transformed prior to presentation in the results.  
204 Composition of VFA (%) in portal and peripheral blood were analysed by the non-  
205 parametric Kruskal–Wallis one-way ANOVA. The individual pig was the experimental  
206 unit for blood analysis.

## 207 **Results**

### 208 *General observations and health.*

209 The results of the dietary chemical analysis confirmed that analysed values were  
210 similar to the calculated values (Table 1). There was no difference between  
211 treatments for health score or in the numbers of pigs that were treated with  
212 antibiotics. Growth performance of pigs was considered normal throughout the trial  
213 with an ADG of  $781 \pm 50.7$  g/day. Two pigs were removed from the trial (8 000

214 BXU/kg) due to ill health. All remaining individual pig data was included for statistical  
215 analysis.

216

### 217 *Growth performance and digestibility*

218 At the start of the experiment there was no difference in liveweight between any of  
219 the dietary treatments. Performance data are shown in Table 2. The addition of  
220 xylanase to the diet had no effect on performance over the 35 days. The addition of  
221 xylanase had no effect on DM, OM or CF total tract digestibility

222

### 223 *pH of the gastrointestinal tract and blood analysis*

224 Supplementation of xylanase to the diet tended to have a quadratic effect on ileal pH  
225 with higher pH values recorded for pigs fed a diet supplemented with 8 000 and 16  
226 000 BXU/kg xylanase ( $P < 0.10$ ). pH levels returned to control levels when  
227 supplemented with 32 000 BXU/kg xylanase. Xylanase had no effect on caecal pH.

228 Xylanase had no effect on peptide YY levels in the portal ( $P = 0.967$ ) or the  
229 peripheral ( $P = 0.846$ ) blood (Figure 1). There was no significant difference in  
230 peptide YY concentrations between portal and peripheral blood ( $P = 0.355$ ). Total  
231 VFA concentration was higher in portal compared to peripheral blood ( $P < 0.05$ ;  
232 Table 3). In terms of the distribution of individual VFA in portal blood, acetate was  
233 present at the highest level ( $2.24 \pm 0.216$  mM/l), followed by propionate ( $0.494 \pm 0$   
234 .0891 mM/l) and then butyrate ( $0.175 \pm 0.0345$  mM/l). In contrast to this, the

235 composition of individual VFA in peripheral blood was predominately acetate.

236 Xylanase had no effect on total VFA concentration.

237

## 238 **Discussion**

239 Supplementing wheat-barley based diets with xylanase had no effect on growth  
240 performance or nutrient digestibility of grower pigs over a 35 d trial period, however  
241 pig performance across all treatments was good which may suggest that diet quality  
242 was sufficient or that there were no antinutritional effects from NSPs and thus there  
243 were no beneficial effects from adding xylanase. Yin *et al.* (2000) found that  
244 xylanase addition (5 000 U/kg) to grower pigs fed wheat based or wheat byproduct  
245 based diets improved nutrient digestibility. However they observed the greatest  
246 improvement in digestibility in pigs fed diets containing wheat plus bran which had a  
247 higher fibre and NSP content compared to wheat alone. Nortey *et al.* (2007) also  
248 found that the addition of xylanase (4 375 U/ kg) to grower pigs improved nutrient  
249 digestibility, however pigs were fed diets containing wheat millrun which again had a  
250 higher concentration of NSP in comparison to wheat. Dry matter digestibility of the  
251 control pigs in the current study was 86%. In the study by Yin *et al.* (2000), DM  
252 digestibility for pigs fed the wheat based diet was 89% whereas DM digestibility for  
253 pigs fed the wheat plus bran diet was 75.8%. Nortey *et al.* (2007) found that millrun  
254 inclusion reduced total tract DM digestibility in pigs from 86.7% (wheat control diet)  
255 to 74.2% (millrun diet). The addition of xylanase improved DM digestibility in the  
256 millrun diets (80.1%) but digestibility remained lower than that of the wheat control  
257 diet. Studies have shown an inverse relationship between control performance and  
258 exogenous enzyme response and thus the better the performance of the control

259 animals, the poorer the response to exogenous enzymes (Rosen, 2006, Rosen,  
260 2010). The age of the pig may also influence the pigs' response to xylanase. Olukosi  
261 *et al.* (2007) found that NSP-degrading enzymes increased nutrient digestibility when  
262 fed to weaner pigs, however NSP-degrading enzymes did not affect nutrient  
263 digestibility when fed to grower pigs. They suggested that this was due to an  
264 increase in the gut microbial population of grower pigs resulting in microbes capable  
265 of breaking down NSP.

266 In the current study xylanase tended to increase ileal pH values for pigs fed a diet  
267 supplemented with 8 000 and 16 000 BXU/kg xylanase. The higher ileal pH levels  
268 observed at 8 000 and 16 000 xylanase BXU/kg are difficult to explain. In contrast  
269 Sheng *et al.* (2013) found that xylanase increased microflora proliferation and  
270 fermentation when fed to broiler chicks resulting in a reduction in ileal digesta pH.  
271 An increase in caecal fermentation has been shown to stimulate the release of  
272 peptide YY from enterocyte cells (Keenan *et al.*, 2012). As peptide YY slows both  
273 gastric emptying and transit time it has been proposed that the release of peptide YY  
274 increases gastric digestion of nutrients. Masey O'Neill *et al.* (2014a) investigated the  
275 effects of xylanase supplementation (16 000 and 32 000 BXU/ kg) to wheat and corn  
276 based broiler diets on broiler performance, nutrient digestibility and caecal VFA  
277 content. They found that xylanase supplementation improved FCR irrespective of  
278 cereal or enzyme dose and improved nutrient digestibility. They also found that  
279 caecal VFA content was higher when broiler diets were supplemented with xylanase.  
280 Yin *et al.* (2000) found a numerical increase in apparent ileal VFA production with the  
281 addition of xylanase to grower pig diets, increasing VFA production by 38% when  
282 added to wheat plus bran based diets which had a high fibre and NSP content. This  
283 is in contrast to the current study where the addition of xylanase to wheat based

284 diets did not affect VFA concentrations in plasma. Volatile fatty acids were measured  
285 in portal blood in order to obtain information on the end products of fermentation in  
286 the gastrointestinal tract. The fact that xylanase had no effect on VFA concentrations  
287 suggests that the fermentation of fibre was already optimised in the control diet and  
288 therefore the addition of xylanase could not enhance this process. Alternatively it  
289 may be that the cereals in this experiment were resistant to xylanase attack. The  
290 higher levels of VFAs observed in the portal blood compared to peripheral blood was  
291 expected as the portal blood reflects the production of VFAs in the gastrointestinal  
292 tract and not consumed by colonocytes before uptake by the liver and peripheral  
293 tissues (Hooda *et al.*, 2010, den Besten *et al.*, 2013)

294 The current study also found no effect of xylanase on plasma peptide YY levels  
295 which is not surprising as VFA concentrations were not affected. Singh *et al.* (2012)  
296 observed an increase in serum levels of peptide YY when broilers were fed a maize  
297 soybean meal based diet under *ad libitum* conditions when diets were supplemented  
298 with xylanase (16 000 u/ kg Econase XT). However in the same experiment xylanase  
299 addition reduced total caecal VFA concentration. The increase in peptide YY was  
300 more pronounced for the broilers fed an energy deficient diet, thus the production of  
301 peptide YY may have been the result of the energy limiting diets in order to increase  
302 nutrient digestion.

303 In conclusion, the addition of xylanase had no effect on grower pig performance,  
304 nutrient digestibility, VFA concentration or peptide YY concentration when fed up to  
305 32 000 BXU/ kg over a 35 d period. The lack of response in the current study may  
306 suggest that NSP levels were low and diet quality was sufficient and as a result the  
307 addition of xylanase had no beneficial effects.

308

309 **Acknowledgements**

310 None.

311

312 **Declaration of interest**

313 There are no conflict of interests.

314

315 **Ethics statement**

316 This experiment received ethical approval from the University of Leeds Animal Welfare and  
317 Ethical Review Committee

318

319 **Software and data repository resources**

320 Data is not deposited in an official repository.

321

322

323

324

325

326

327

328

329

330

331

332

333

334 **References**

335 Allen JM, Fitzpatrick ML, Yeats JC, Darcy K, Adrian TE and Bloom SR 1984. Effects of Peptide YY and  
336 Neuropeptide Y on Gastric Emptying in Man. *Digestion* 30, 255-262.

337

338 Animals (Scientific Procedures) Act 1986. Appropriate methods of humane killing. Retrieved on 12  
339 December 2017, from <https://www.legislation.gov.uk/ukpga/1986/14/schedule/1>

340

341 Authority EFS 2008. Scientific Opinion of the Panel on Additives and Products or Substances used in  
342 Animal Feed and of the Panel on Genetically Modified Organisms on the safety and efficacy of  
343 Econase XT P/L as feed additive for chickens for fattening, chickens reared for laying, turkeys for  
344 fattening, turkeys reared for breeding and piglets (weaned). *The EFSA Journal* 712, 1-19.

345

346 BSAS 2003. Nutrient Requirement Standards for Pigs. British Society of Animal Science, Midlothian,  
347 UK.

348

349 Cuche G, Cuber JC and Malbert CH 2000. Ileal short-chain fatty acids inhibit gastric motility by a  
350 humoral pathway. *American Journal of Physiology-Gastrointestinal and Liver Physiology* 279, G925-  
351 G930.

352

353 den Besten G, van Eunen K, Groen AK, Venema K, Reijngoud D-J and Bakker BM 2013. The role of  
354 short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism.  
355 *Journal of Lipid Research* 54, 2325-2340.

356

357 Goodlad RA, Lenton W, Ghatei MA, Adrian TE, Bloom SR and Wright NA 1987. Effects of an  
358 elemental diet, inert bulk and different types of dietary fibre on the response of the intestinal  
359 epithelium to refeeding in the rat and relationship to plasma gastrin, enteroglucagon, and PYY  
360 concentrations. *Gut* 28, 171-180.

361

362 Grootaert C, Van den Abbeele P, Marzorati M, Broekaert WF, Courtin CM, Delcour JA, Verstraete W  
363 and Van de Wiele T 2009. Comparison of prebiotic effects of arabinoxylan oligosaccharides and  
364 inulin in a simulator of the human intestinal microbial ecosystem. *Fems Microbiology Ecology* 69,  
365 231-242.

366

367 Hooda S, Matte JJ, Vasanthan T and Zijlstra RT 2010. Dietary Oat beta-Glucan Reduces Peak Net  
368 Glucose Flux and Insulin Production and Modulates Plasma Incretin in Portal-Vein Catheterized  
369 Grower Pigs. *Journal of Nutrition* 140, 1564-1569.

370

371 Keenan MJ, Martin RJ, Raggio AM, McCutcheon KL, Brown IL, Birkett A, Newman SS, Skaf J, Hegsted  
372 M, Tulley RT, Blair E and Zhou J 2012. A microarray study indicates high-amylose resistant starch  
373 increases hormones and improves structure and function of the GI tract. *Journal of nutrigenetics and  
374 nutrigenomics* 5, 26-44.



375  
376 Kim JC, Simmins PH, Mullan BP and Pluske JR 2005. The digestible energy value of wheat for pigs,  
377 with special reference to the post-weaned animal [Review]. *Animal Feed Science and Technology*  
378 122, 257-287.  
379  
380 Masey O'Neill HV, Singh M and Cowieson AJ 2014a. Effects of exogenous xylanase on performance,  
381 nutrient digestibility, volatile fatty acid production and digestive tract thermal profiles of broilers fed  
382 on wheat- or maize-based diet. *British Poultry Science* 55, 351-359.  
383  
384 Masey O'Neill HV, Smith JA and Bedford MR 2014b. Multicarbohydrase Enzymes for Non-ruminants.  
385 *Asian-Australasian Journal of Animal Sciences* 27, 290-301.  
386  
387 Neyrinck AM, Van Hee VF, Piront N, De Backer F, Toussaint O, Cani PD and Delzenne NM 2012.  
388 Wheat-derived arabinoxylan oligosaccharides with prebiotic effect increase satietogenic gut  
389 peptides and reduce metabolic endotoxemia in diet-induced obese mice. *Nutrition & Diabetes* 2, 1-  
390 9.  
391  
392 Nortey TN, Patience JF, Simmins PH, Trottier NL and Zijlstra RT 2007. Effects of individual or  
393 combined xylanase and phytase supplementation on energy, amino acid, and phosphorus  
394 digestibility and growth performance of grower pigs fed wheat-based diets containing wheat millrun.  
395 *Journal of Animal Science* 85, 1432-1443.  
396  
397 Olukosi OA, Sands JS and Adeola O 2007. Supplementation of carbohydrases or phytase individually  
398 or in combination to diets for weanling and growing-finishing pigs. *Journal of Animal Science* 85,  
399 1702-1711.  
400  
401 Passos AA, Park I, Ferket P, von Heimendahl E and Kim SW 2015. Effect of dietary supplementation  
402 of xylanase on apparent ileal digestibility of nutrients, viscosity of digesta, and intestinal morphology  
403 of growing pigs fed corn and soybean meal based diet. *Animal Nutrition* 1, 19-23.  
404  
405 Rosen GD 2006. Holo-analysis. *Poultry Science* 85, 957-959.  
406  
407 Rosen GD 2010. Holo-analysis of the efficacy of exogenous enzyme performance in farm animal  
408 nutrition. In *Enzymes in farm animal nutrition* (eds. MR Bedford and G Partridge), pp. 273–303, CAB  
409 International, Wallingford, UK.  
410  
411 Sheng QK, Yang LQ, Zhao HB, Wang XL and Wang K 2013. Effects of Low Level Water-soluble  
412 Pentosans, Alkaline-extractable Pentosans, and Xylanase on the Growth and Development of Broiler  
413 Chicks. *Asian-Australasian Journal of Animal Sciences* 26, 1313-1319.  
414  
415 Short FJ, Gorton P, Wiseman J and Boorman KN 1996. Determination of titanium dioxide added as an  
416 inert marker in chicken digestibility studies. *Animal Feed Science and Technology* 59, 215-221.  
417  
418 Singh A, O'Neill HVM, Ghosh TK, Bedford MR and Haldar S 2012. Effects of xylanase supplementation  
419 on performance, total volatile fatty acids and selected bacterial population in caeca, metabolic  
420 indices and peptide YY concentrations in serum of broiler chickens fed energy restricted maize–  
421 soybean based diets. *Animal Feed Science and Technology* 177, 194-203.  
422  
423 Taylor AE, Jagger S, Toplis P, Wellock IJ and Miller HM 2013. Are compensatory live weight gains  
424 observed in pigs following lysine restriction during the weaner phase? *Livestock Science* 157, 200-  
425 209.

426  
 427 Woyengo TA, Sands JS, Guenter W and Nyachoti CM 2008. Nutrient digestibility and performance  
 428 responses of growing pigs fed phytase- and xylanase-supplemented wheat-based diets. Journal of  
 429 Animal Science 86, 848-857.  
 430  
 431 Yin YL, McEvoy JDG, Schulze H, Hennig U, Souffrant WB and McCracken KJ 2000. Apparent  
 432 digestibility (ileal and overall) of nutrients and endogenous nitrogen losses in growing pigs fed wheat  
 433 (var. Soissons) or its by-products without or with xylanase supplementation. Livestock Production  
 434 Science 62, 119-132.

435

436 **Table 1** *Ingredient composition and nutrient content of experimental diets fed throughout the*  
 437 *trial.*

Xylanase, BXU/kg	0	8 000	16 000	32 000
<b>Raw Material (%)</b>				
Raw Wheat	41.229	41.224	41.219	41.209
Barley 62 kg/hl	20	20	20	20
Wheatfeed	5	5	5	5
Hypro soya	26.32	26.32	26.32	26.32
Herring meal	1.25	1.25	1.25	1.25
Soya Oil	1.71	1.71	1.71	1.71
Dicalcium Phosphate	1.41	1.41	1.41	1.41
Salt	0.39	0.39	0.39	0.39
L-Lysine-HCL	0.402	0.402	0.402	0.402
DL-Methionine	0.171	0.171	0.171	0.171
L-Threonine	0.166	0.166	0.166	0.166
L-Tryptophan	0.016	0.016	0.016	0.016
Vitamin/mineral supp	0.501	0.501	0.501	0.501
Lignobond	0.625	0.625	0.625	0.625
Quantum Blue 5G	0.01	0.01	0.01	0.01
Econase XT	0	0.005	0.01	0.02
Benzoic acid	0.5	0.5	0.5	0.5
Limestone flour	0.3	0.3	0.3	0.3
<b>Calculated nutrient content</b>				
Net energy (MJ/kg)	9.97	9.97	9.97	9.97
Digestible energy (MJ/kg)	14.17	14.17	14.17	14.17
Protein (%)	21.32	21.32	21.32	21.32
Fibre (%)	3.14	3.14	3.14	3.14
Salt (%)	0.65	0.65	0.65	0.65
Calcium (%)	0.71	0.71	0.71	0.71
Phosphorous (%)	0.68	0.68	0.68	0.68
Sodium (%)	0.18	0.18	0.18	0.18
SID <sup>1</sup> lysine	1.25	1.25	1.25	1.25
SID methionine	0.28	0.28	0.28	0.28
SID met + cys	0.58	0.58	0.58	0.58
SID theronine	0.8	0.8	0.8	0.8
SID tryptophan	0.24	0.24	0.24	0.24

**Analysed<sup>2</sup>**

Crude protein (%)	21.5	21.1	21.2	20.8
Lysine, total (%)	1.42	1.51	1.46	1.37
Calcium (%)	0.61	0.6	0.63	0.61
Phosphorous (%)	0.68	0.69	0.74	0.67
Xylanase activity, BXU/kg <sup>3</sup>	<2 000	10 900	22 800	42 500

438

439

440 **Table 2** *Effects of increasing concentrations of xylanase on liveweight, average daily gain,*441 *average daily intake, feed conversion ratio and pH of the ileum and caecum.*

Xylanase BXU/kg <sup>1</sup>	0	8 000	16 000	32 000	SE	Linear	Quadratic
Wk 7 wt, kg	14.2	14.2	14.2	14.2	0.31	0.955	0.956
Wk 10 wt, kg	29.2	28.9	29.1	28.8	0.54	0.432	0.754
Wk 12 wt, kg	41.4	41.9	41.3	41.4	0.62	0.626	0.628
7-10 weeks							
ADI 7-10, kg	1.034	1.029	1.031	1.024	0.0141	0.869	0.965
ADG 7-10, kg	0.716	0.699	0.708	0.693	0.0153	0.400	0.899
FCR 7-10	1.45	1.49	1.47	1.50	0.027	0.351	0.794
10-12 weeks							
ADI 10-12, kg	1.507	1.559	1.518	1.51	0.0272	0.739	0.690
ADG 10-12, kg	0.870	0.923	0.876	0.898	0.0181	0.639	0.855
FCR 10-12	1.74	1.7	1.76	1.70	0.044	0.618	0.668
7-12 weeks							
ADI 7-12, kg	1.223	1.24	1.226	1.218	0.0164	0.783	0.563
ADG 7-12, kg	0.777	0.788	0.774	0.774	0.0082	0.688	0.984
FCR 7-12	1.57	1.58	1.58	1.58	0.018	0.815	0.722
Health score	1.411	1.426	1.411	1.400	0.0292	0.677	0.741
Ileum pH	6.85	6.99	6.98	6.87	0.069	0.837	0.057
Caecum pH	5.67	5.73	5.69	5.62	0.069	0.371	0.333
Total-tract digestibilities (%)							
DM	86.90	86.22	84.01	84.75	1.011	0.105	0.238
OM	88.43	87.74	86.01	86.45	0.936	0.077	0.552
CF	50.67	45.74	39.07	39.85	4.829	0.105	0.313

442 <sup>1</sup> BXU = xylanase units, *n* = 32.

443 Wk = week; Wt = weight; ADFI = average daily feed intake; ADG = average daily gain; FCR  
 444 = feed conversion ratio; DM = dry matter; OM = organic matter; CF = crude fibre

445

446

447

448

449

450 **Table 3** *Volatile fatty acid (VFA) concentrations in portal and peripheral plasma of weaner-*  
 451 *grower pigs fed four levels of xylanase (d 35)*

452

Xylanase BXU/kg <sup>1</sup>	0	8 000	16 000	32 000	SE	P-value
Total VFA Portal, mM/l	2.52	2.180	2.55	2.55	0.627	0.993
Total VFA Peripheral mM/l	1.00	0.72	0.86	0.52	0.890	0.501
<b>Molar VFA proportion (%) Portal</b>						
Acetate	82.8	88.8	84.1	79.9	5.48	0.474
Propionate	10.3	8.2	12.5	15.0	3.15	0.378
Butyrate	7.0	3.0	3.4	5.1	2.30	0.368
<b>Molar VFA proportion (%) Peripheral</b>						
Acetate	95.3	100	100	100	3.2	0.121

453 <sup>1</sup>BXU = xylanase units, *n* = 16.

454

455

456

457

458

459

460

461

462

463

464

465

466 **Figure 1** Peptide YY concentrations in portal and peripheral plasma of grower pigs  
467 fed either 0, 8 000, 16 000 or 32 000 BXU/kg xylanase at d 35 (*n* = 16).