

This is a repository copy of *Temporal and spatial variation in pharmaceutical concentrations in an urban river system*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/128156/>

Version: Accepted Version

Article:

Burns, Emily E. orcid.org/0000-0003-4236-6409, Carter, Laura J., Kolpin, Dana W et al. (2 more authors) (2018) Temporal and spatial variation in pharmaceutical concentrations in an urban river system. Water research. pp. 72-85. ISSN: 0043-1354

<https://doi.org/10.1016/j.watres.2018.02.066>

Reuse

This article is distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs (CC BY-NC-ND) licence. This licence only allows you to download this work and share it with others as long as you credit the authors, but you can't change the article in any way or use it commercially. More information and the full terms of the licence here: <https://creativecommons.org/licenses/>

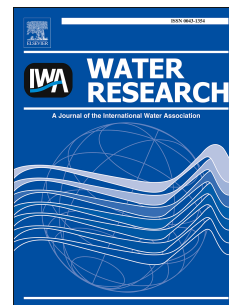
Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

Accepted Manuscript

Temporal and spatial variation in pharmaceutical concentrations in an urban river system

Emily E. Burns, Laura J. Carter, Dana W. Kolpin, Jane Thomas-Oates, Alistair B.A. Boxall



PII: S0043-1354(18)30179-9

DOI: [10.1016/j.watres.2018.02.066](https://doi.org/10.1016/j.watres.2018.02.066)

Reference: WR 13618

To appear in: *Water Research*

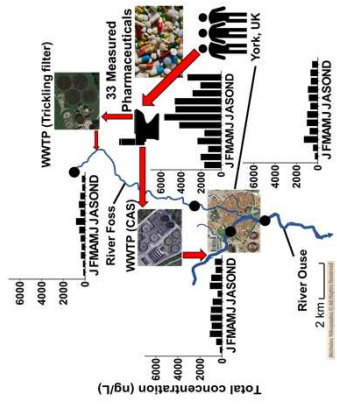
Received Date: 14 December 2017

Revised Date: 27 February 2018

Accepted Date: 28 February 2018

Please cite this article as: Burns, E.E., Carter, L.J., Kolpin, D.W., Thomas-Oates, J., Boxall, A.B.A., Temporal and spatial variation in pharmaceutical concentrations in an urban river system, *Water Research* (2018), doi: 10.1016/j.watres.2018.02.066.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Temporal and spatial variation in pharmaceutical concentrations in an urban river system

Emily E. Burns^a, Laura J. Carter^b, Dana W. Kolpin^c, Jane Thomas-Oates^a, Alistair B.A. Boxall^b

^aChemistry Department, University of York, York, YO10 5DD United Kingdom

^bEnvironment Department, University of York, York, YO10 5DD United Kingdom

^cU.S. Geological Survey, Iowa City, IA, 52242, United States

Corresponding author: Emily Burns (emily.burns@york.ac.uk)

Other Author emails: Laura Carter (laura.carter@york.ac.uk), Dana Kolpin (dw.kolpin@usgs.gov), Jane Thomas-Oates (jane.thomas-oates@york.ac.uk), Alistair Boxall (alistair.boxall@york.ac.uk)

Abstract

Many studies have quantified pharmaceuticals in the environment, few however, have incorporated detailed temporal and spatial variability due to associated costs in terms of time and materials. Here, we target 33 physico-chemically diverse pharmaceuticals in a spatiotemporal exposure study into the occurrence of pharmaceuticals in the wastewater system and the Rivers Ouse and Foss (two diverse river systems) in the city of York, UK. Removal rates in two of the WWTPs sampled (a carbon activated sludge (CAS) and trickling filter plant) ranged from not eliminated (carbamazepine) to >99% (paracetamol). Data comparisons indicate that pharmaceutical exposures in river systems are highly variable regionally, in part due to variability in prescribing practices, hydrology, wastewater management, and urbanisation and that select annual median pharmaceutical concentrations observed in this study were higher than those previously observed in the European Union and Asia thus far.

Significant spatial variability was found between all sites in both river systems, while seasonal variability was significant for 86% and 50% of compounds in the River Foss and Ouse, respectively. Seasonal variations in flow, in-stream attenuation, usage and septic effluent releases are suspected drivers behind some of the observed temporal exposure variability. When the data were used to evaluate a simple environmental exposure model for pharmaceuticals, mean ratios of predicted environmental concentrations (PECs), obtained using the model, to measured environmental concentrations (MECs) were 0.51 and 0.04 for the River Foss and River Ouse, respectively. Such PEC/MEC ratios indicate that the model underestimates actual concentrations in both river systems, but to a much greater extent in the larger River Ouse.

Keywords: LC-MS/MS; surface water; wastewater; seasonal; exposure; predicted environmental concentration

1.0 Introduction

Determining pharmaceutical exposures in environmental matrices has become a substantial area of research since the 1990s (Daughton, 2016). The presence of pharmaceuticals in freshwater systems has now been documented globally, with research especially focused in Europe and North America (aus der Beek et al., 2016). Pharmaceuticals primarily enter the environment through patient use when an unmetabolised fraction is excreted and subsequently passes through wastewater treatment plants (WWTPs), which are typically not designed to remove such organic contaminants (Luo et al., 2014). Consequently, WWTPs are significant sources of pharmaceuticals to the environment (Lindholm-Lehto et al., 2016). A recent study of United Kingdom (UK) WWTPs estimated that 13% of effluent discharges could pose risks to the receiving environment regarding pharmaceutical exposures (Comber et al.,

2018). Removal rates are highly variable between treatment types (Kasprzyk-Hordern et al., 2009; Luo et al., 2014), seasons (Golovko et al., 2014), and even within treatment plants themselves (Verlicchi et al., 2012). Moreover, removal rates have only been estimated for a small fraction of the total number of pharmaceuticals in use (Boxall et al., 2014) and only a few studies have reported WWTP removals in the UK specifically (Comber et al., 2018; Kasprzyk-Hordern et al., 2009, 2008). WWTP removal rates are valuable parameters, and their inclusion in occurrence modelling substantially improves the accuracy of pharmaceutical exposure predictions (Burns et al., 2017; Verlicchi et al., 2014).

The potential for, and extent of, effects posed by pharmaceutical exposure to non-target organisms, such as fish or invertebrates, is largely unknown (Vasquez et al., 2014). However, there is mounting evidence that select pharmaceuticals are having deleterious effects at environmentally relevant (i.e. real-world) concentrations. Examples of documented effects at environmentally relevant concentrations include antidepressants causing behavioural changes in fish (fluoxetine) (Mccallum et al., 2017), disruption during early development (venlafaxine) (Thompson et al., 2017), the equivalent of human side effects from exposure to the anti-diabetic drug metformin (Niemuth et al., 2015) or the feminization of wild fish populations downstream of a pharmaceutical manufacturing facility in France (Sanchez et al., 2011). It is therefore important to characterise the source and fate of pharmaceuticals in the aquatic environment to aid in risk assessment as approaches evaluating potential adverse effect concentrations emerge.

To adequately characterise the fate of pharmaceuticals in the environment, robust monitoring campaigns which include seasonal or year-long sampling covering a range of compounds at a reasonable spatial resolution are required. However, only a small number of spatiotemporal exposure studies have been performed that meet these

criteria (Baker and Kasprzyk-Hordern, 2013; Daneshvar et al., 2010; Kasprzyk-Hordern et al., 2008; Paíga et al., 2016). These exposure studies are extremely valuable as they provide detailed information which can be related back to the myriad of factors (many varying both seasonally and temporally) that influence environmental concentrations of pharmaceuticals including hydrology (Kasprzyk-Hordern et al., 2008), WWTP removal efficiency (Silva et al., 2014), pharmaceutical usage (Sun et al., 2014), and in-stream removal processes (e.g. biodegradation and sorption to sediment) (Daneshvar et al., 2010; Camacho-Munoz et al., 2010; Moreno-González et al., 2014). In combination, the impact of these processes on pharmaceutical exposure and fate is largely unknown but, if better defined, could improve exposure prediction approaches and offer greater confidence, in terms of exposure, when evaluating risks that pharmaceuticals may pose to the environment.

Recently, a handful of aqueous rapid pharmaceutical determination high-performance liquid chromatography tandem mass spectrometry (HPLC-MS/MS) methods have been developed that achieve comparable limits of detection (LODs) to those including sample pre-concentration or clean-up (Anumol et al., 2015; Boix et al., 2015; Campos-Mañas et al., 2017; Furlong et al., 2014; Oliveira et al., 2015). Such methods involve utilising larger than normal injection volumes (~100 μ L) to increase the likelihood of detection (Petrie et al., 2016). Removal of the extraction step reduces sample preparation time and can increase the number of samples that can be processed (highly beneficial to large spatiotemporal exposure campaigns). A significant analytical problem arising during pharmaceutical quantification is matrix effects (typically mass spectrometric ionisation enhancement or suppression). The presence of background interferences in “dirty” matrices (e.g. streams, WWTP effluent, etc.) can co-elute with target analytes and impair quantification past the point of suitability (Petrović et al., 2005). Several approaches have been attempted to reduce matrix effects including

sample pre-concentration and clean-up to help isolate target pharmaceuticals (Van De Steene et al., 2006). Such pre-concentration, however, is difficult to optimise, time consuming, costly, and may also concentrate interfering analytes, thus unintentionally increasing matrix effects (Yu et al., 2012). Matrix interferences have been reported to be comparatively lower for rapid determination methods than more costly and laborious sample pre-concentration/clean-up methods (Anumol et al., 2015).

In this study, which was performed in the frame of the Innovative Medicines Initiative iPiE project on intelligent assessment of pharmaceuticals in the environment, we validate and apply a rapid determination aqueous HPLC-MS/MS method for the quantification of 33 physico-chemically diverse pharmaceuticals to a year-long surface-water exposure campaign. Monitoring was conducted during 2016 at 11 sites along the urbanised and larger River Ouse and smaller, more rural River Foss which converge within the city of York, UK (Figure 1). The monthly sampling design provided good temporal resolution while unparalleled spatial resolution was achieved in the two contrasting river systems. In addition, influent and effluent samples from two of the WWTPs that serve the city were collected when possible and removal efficiencies estimated. Predicted exposure concentrations (PECs) were calculated for both rivers using a simple model and the model was then evaluated against annually averaged measured environmental concentrations (MECs) calculated from the monthly sampling data.

2.0 Methods

2.1 Study area and sample collection

2.1.1 Study Compounds

Study compounds were selected based on those previously detected in the York river system during an initial scoping study in which 95 pharmaceutical and degradation

products were surveyed (Burns et al., 2017). From these results, 32 pharmaceuticals were selected due to either their known or expected presence. An additional compound, gabapentin, was also included in the study due to its high usage, resistance to environmental degradation, and ecotoxic potential (Herrmann et al., 2015).

2.1.2 Study Area

The River Ouse and River Foss were chosen for the study, as they flow through the city of York, UK, and converge downstream of the city centre (Figure 1). The two rivers represent differing levels of urbanisation and size. To minimise potential variability, grab water samples were collected from the network of 11 sampling sites in the same order and on approximately the same day and time each month from January to December 2016. Site locations were strategically chosen based on their ease of access and position in relation to WWTP outfalls. Both rivers were sampled with sufficient spatial resolution to build concentration profiles and increase the probability of detecting transient pharmaceuticals in the absence of composite sampling techniques. Three WWTPs serve the city within the sampling network (Figure 1). WWTP A is a trickling filter plant and serves a population of 18 600, WWTP B is a conventional activated sludge (CAS) facility serving a population of 27 900, while WWTP C is a surplus activated sludge (SAS) plant serving a population of 180 500. Sampling site and WWTP characteristics along with dates of sampling are detailed in Supplemental Material, Tables S1 and S2.

2.1.3 Sample Collection

All samples collected were subject to the same sampling protocol. At each site, three 1-L field replicates were collected from the centroid of flow (when possible); sampling sites had been previously determined to be well-mixed, therefore sampling in a single location was deemed appropriate (Supplemental Material, Figure S1). For each field replicate, a 10-mL aliquot was drawn into a 24-mL disposable syringe and filtered

through a primed 0.7- μ m glass-fibre filter (GF/F) (Whatman Inc.) into an amber glass vial and immediately frozen in the field using dry ice. To demonstrate that field filtration and collection did not contaminate samples, three field blanks per sampling visit were collected. HPLC-grade water was brought to the field, filtered and prepared identically to field samples. Samples were then returned to the laboratory and stored at -18°C until analysis which occurred within seven days. The concentration reported for each sample per site is the median of the three field replicates collected. The filtering of samples in the field is beneficial as it removes particulates which can extend HPLC column life, reduce instrument maintenance, as well as remove bacteria associated with particulates that could facilitate analyte degradation. There is a formal possibility that analytes could be retained on the filter; however pharmaceutical filtration studies including 26 compounds (acids, bases and amphoteres) ranging in hydrophobicity (logKow -2.3 to 6.3) suggest these losses will be insignificant (<5%) (Mompelat et al., 2013), thus an assessment of filter losses has not been repeated here.

2.2 High performance liquid chromatography-tandem mass spectrometry

A Thermo Scientific™ TSQ Endura MS operating in multiple reaction monitoring mode interfaced with an EASY-Max NG™ heated electrospray source operating in positive mode was used for pharmaceutical detection. Two transitions were monitored for each analyte and the m/z and collision energy optimised using the Thermo™ Tune 2.0 software, summarised in Supplemental Material, Table S3. Chromatographic separation was achieved with a Dionex Ultimate 3000 HPLC (Thermo Scientific™) equipped with a 100- μ L sample injection loop and autosampler maintained at 4°C. Mobile phase A consisted of HPLC-grade water amended with 12-mL of 1 M formic acid and 10-mL of 1 M ammonium hydroxide for a total volume of 1-L, and mobile phase B was 100% methanol (Furlong et al., 2014). The chromatographic conditions and program are reported in the Supplemental Material Table S4.

Internal standard (IS) calibration was used to quantify the pharmaceuticals in the method described. For reasons of expense and availability, not all pharmaceuticals had a corresponding isotopically labelled internal standard (ILIS) (Supplemental Material, Table S3). In these cases, atrazine- d_5 was used and has been previously determined suitable for this role (Furlong et al., 2014). Samples were fully thawed and a 995- μ L aliquot pipetted into a 1.5-mL LC vial and a 5- μ L spike of IS solution (80 ng/L) added. Samples were immediately analysed after preparation. Peak detection criteria were in accordance with Commission Decision (2002/657/EC). Due to analytical complications, fexofenadine could not be quantified in the April surface-water samples. Further details of peak qualification and quantitation are provided in the Supplementary Material.

The use of ILIS is a good strategy to compensate for matrix effects (Stüber and Reemtsma, 2004). This is not a perfect solution as matrix effects can still influence quantification, possibly due to a slight difference in retention time (t_R) between the ILIS and target analyte resulting in differing ionisation efficiencies (Wang et al., 2007). Therefore, sample matrix spikes were routinely prepared and analysed with all sample batches to provide an indication of the presence of interferences which cause signal suppression/enhancement and could impact quantification. In this study, acceptable matrix recovery was considered to be 70% to 120% in accordance with previously published methods (Boix et al., 2015; USEPA, 2016; Furlong et al., 2014). Matrix 'recovery' falling outside this range indicates signal suppression/enhancement could be occurring and samples should quantitatively be interpreted with caution. At least three matrix spike samples from different sampling sites were prepared per analytical batch to monitor for matrix effects throughout the sampling campaign as the sample matrices are heterogenous and likely to vary temporally. Surface-water matrix spikes were prepared by spiking 20 μ L of 80 ng/L or 200 ng/L calibration solution into a sample replicate with 5 μ L of IS solution. The much higher ambient concentration of pharmaceuticals in WWTP

influent and effluent required the matrix spike samples to be prepared at a higher concentration, 4000 ng/L. Matrix recovery was calculated by subtracting the ambient sample concentration and dividing by the concentration spiked.

With each sample batch at least three calibration check samples (CCSs) were prepared to monitor accuracy throughout the analytical batch (injected every 10 samples). These CCSs were prepared to a concentration of 80 ng/L by pipetting 20 μ L of the relevant calibration solution into 975 μ L of HPLC grade water and spiked with 5 μ L of IS solution. At the end of each batch a 4 ng/L calibration solution spike, prepared similarly, was also injected. The accuracy of these CCSs was required to be within 20% or affected samples were re-analysed (Furlong et al., 2014; USEPA, 2016).

This formed part of a rigorous quality control plan which was followed during environmental sample analysis using a series of sample matrix spikes, calibration solution spikes, field blanks, and laboratory blanks randomly dispersed throughout analytical batches. Further detail of quality control, how these samples were prepared and results are reported in the Supplementary Material.

2.3 Analytical method validation

Method validation included an assessment of precision (inter- and intra-day), limits of detection, limits of quantification, and recovery from all studied matrices. The methods and results with which each of these parameters were assessed are reported in the Supplemental Material.

2.4 WWTP removal efficiency

Due to access restrictions, 24 h composite samples for influent and effluent could only be collected once from WWTP A and B during summer 2016 (Supplementary Material, Table S2). Only grab samples unsuitable for estimating removals could be collected from WWTP C. WWTP removal efficiency was estimated, when appropriate,

for WWTP A and B based on mean influent and effluent concentrations according to Equation 1. In this context ‘removal’ is the change in concentration between influent and effluent which does not represent true removal, but rather partitioning to the solid phase and/or the formation of transformation products. Negative removals can occur, potentially due to sampling limitations (e.g. longer than 24 h hydraulic/sludge retention time) (Ort et al., 2010), from the conversion of conjugated metabolites back to the parent compound during treatment (Verlicchi et al. 2012), or desorption from sludge during secondary treatment (Blair et al., 2015).

$$\% \text{ Removal} = \left(1 - \frac{\text{Effluent}}{\text{Influent}}\right) \times 100 \quad [1]$$

2.5 Statistical Analysis

Data analysis was performed using Graphpad Prism (Graphpad Software, 2017). To use statistical tests when non-detects were present, data substitution according to Equation 2 was undertaken. This approach was suggested to be appropriate for left censoring of up to 40% of a dataset (Antweiler, 2015). If the non-detect frequency for a compound was greater than 40%, it was not included in statistical testing. To determine whether significant spatial differences existed between sites, pairwise t-tests were conducted based on the monthly concentrations (Furlong et al., 2017). To determine whether any analytes were seasonally variable in each river, concentrations from sites F3-F4 and O3-O4 were grouped by season and a Friedman’s Test followed by a Dunn’s multiple comparisons post hoc test was undertaken. These sites were used in the seasonality test due to their downstream location in relation to WWTP A and B, as well as their location in relation to Environment Agency flow gauges (Figure 1) as the flow recorded at these gauges was not representative of flow conditions at the remaining study sites (Center for Ecology & Hydrology, 2016).

$$\text{Substitution} = \frac{\sqrt{2}}{2} * \text{LOD} \quad [2]$$

2.6 Predicted environmental concentrations

Annual average MECs were compared to PECs to gauge the accuracy of simple exposure algorithms commonly used for the prioritisation of pharmaceuticals and risk assessment (Burns et al., 2017). Local annual pharmaceutical usage data were obtained from the National Health Service Business Authority (National Health Service, 2016), while wastewater generation was assumed to be 200 L/person·day (European Medicines Agency, 2006). Experimental WWTP removal rates (Eqn. 1) were used with river specific dilution factors based on the average flow from sampling days to generate a PEC for both rivers. PEC calculations were based on the approach suggested by the European Medicines Agency (2006). Parameters and equations used to predict the PECs are provided in the Supplemental Material Table S6.

3.0 Results & Discussion

3.1 Method performance and quality control

The method was determined to be sufficiently reproducible as assessed by the relative standard deviation of multiple injections ($n=8$) during (5.5 %RSD) and across (7.5 %RSD) analysis days according to USEPA (2016) guidelines and Boix et al. (2015) where an $RSD \leq 20\%$ above the LOQ (i.e. 80 ng/L) is desirable. The limits of detections (LOD) ranged from 0.9 ng/L (carbamazepine) to 12.4 (gabapentin) and an LOD <10 ng/L was achieved for 91% of analytes (Table S5). There were no quantifiable concentrations of any of the target pharmaceuticals in field blanks collected routinely throughout the monitoring campaign. Routine matrix spikes in surface water fell within the acceptable 70 – 120% recovery range for concentrations of 80 and 200 ng/L, indicating that throughout the sample analysis quantification was not unacceptably impaired due to matrix effects (Figure 2). Matrix effects were observed in WWTP effluent and influent, a phenomenon also reported by others, and suggested to be due to the presence of a greater proportion of chemical species that can affect consistent ionisation in

comparison to surface water (Boix et al., 2015; Oliveira et al., 2015). In effluent 13% and in influent 19% of analytes fell outside the acceptable matrix signal response, identified in Figure 2 and 3. Signal enhancement was most prominent for diphenhydramine in both influent and effluent (442% and 375%, respectively), while metformin (214%) and tramadol (156%) also exhibited significant signal enhancement in influent. In this study, a slight shift in relative t_R of the analyte with respect to its ILIS, was observed in WWTP influent and effluent in comparison to surface water, which, in addition to it containing a larger number of chemical constituents, could help explain why matrix effects were not well compensated for all analytes using isotopically labelled internal standards. WWTP influent and effluent matrix spikes indicate that caution is needed when interpreting quantitative results and removal efficiencies due to significant matrix effects, while matrix spikes in surface water indicate that matrix effects are sufficiently compensated for by the internal standards.

3.2 Pharmaceuticals in WWTPs

The highest summed pharmaceutical concentrations in influent were observed in samples from WWTP B, while highest summed concentrations in effluent were observed in samples taken at WWTP A. Paracetamol had the highest concentration in all WWTP influents, 282, 186 and 117 $\mu\text{g/L}$ at WWTP B, A and C, respectively. In effluent, gabapentin had the highest concentration (8541 ng/L) at WWTP C followed by metformin (6111 ng/L) at WWTP A and fexofenadine (2094 ng/L) in effluent at WWTP C. Seven pharmaceuticals (diphenhydramine, norethisterone, oseltamivir, raloxifene, sertraline, triamterene and verapamil) were not detected in any WWTP sample. Average concentration and standard deviation (SD) of WWTP influent and effluent samples are reported in the Supplemental Data Table S10.

In a global review of pharmaceuticals in WWTPs, Verlicchi et al. (2012) reported influent concentrations for many compounds also observed in the WWTP samples in this

study. Codeine, paracetamol, gabapentin, hydrocodone, tramadol, erythromycin, trimethoprim, diltiazem, atenolol, propranolol, carbamazepine, gabapentin, cimetidine, and ranitidine influent concentrations all fell within the ranges reported by Verlicchi et al. (2012), while concentrations of amitriptyline were an order of magnitude lower. A study of effluents in the European Union (EU) reported average concentrations an order of magnitude lower than those determined here for tramadol, codeine, citalopram, fexofenadine, diltiazem, ranitidine, and amitriptyline, while effluent concentrations were similar for venlafaxine, trimethoprim, carbamazepine, and sulfamethoxazole in the York samples (Loos et al., 2013).

The estimated removal efficiency in each WWTP is presented for all detected analytes in Figure 3. The median removal efficiency was estimated to be 75% in WWTP A and 38% in WWTP B. Paracetamol was the analyte most efficiently removed at both treatment plants (>99%), while removals greater than 75% were reported for gabapentin, ranitidine, atenolol, sulfamethoxazole, metformin, and codeine. Despite being a trickling filter plant which might be expected to have poorer pharmaceutical removal than CAS systems (Kasprzyk-Hordern et al., 2009), WWTP A had similar and even greater removals for select compounds (i.e. carbamazepine, diltiazem, citalopram, erythromycin, cimetidine, and ranitidine). In the UK specifically, similar removals were reported previously (Kasprzyk-Hordern et al., 2009) for trimethoprim, amitriptyline, diltiazem, cimetidine, gabapentin, and paracetamol, while sulfamethoxazole, erythromycin, codeine, tramadol, carbamazepine, propranolol and ranitidine were, in general, more efficiently removed for this study. WWTPs with similar treatment capabilities were also studied previously in the UK (Kasprzyk-Hordern et al., 2009). In comparison with results reported here, WWTP removal rates were highly variable despite operating in the same region and employing similar treatments, a conclusion also observed in other regions (Verlicchi et al., 2012). The single sampling event in the WWTPs is limited, however

these estimates are still useful for comparative purposes. For example, sitagliptin removal efficiency (25 - 40%) has not been previously reported to the authors' knowledge. Therefore, while WWTPs are significant sources of pharmaceuticals entering the environment, analysis of WWTP removal efficiencies (i.e. reduction in parent pharmaceutical concentration from influent to effluent) as documented in this and previously published studies, demonstrate that WWTPs are generally decreasing the aquatic environmental burden by significantly reducing certain parent pharmaceutical concentrations (not considering degradates or transformation products) for many of the compounds studied.

3.3 Pharmaceuticals in Surface Water

Of the 33 pharmaceuticals monitored, 21 were detected in all 12 months in samples from the River Foss. Three compounds, oxazepam, verapamil, and triamterene, were not detected in any Foss sample. The remaining nine study compounds, diazepam, diphenhydramine, loratadine, norethisterone, oseltamivir, raloxifene, sulfamethoxazole, sertraline, and temazepam, were sporadically detected from month to month in this river. In comparison, ten compounds (carbamazepine, codeine, fexofenadine, gabapentin, hydrocodone, lidocaine, metformin, paracetamol, tramadol, and trimethoprim) were detected in all 12 months in the River Ouse samples. Eight compounds were not detected in any Ouse sample: diazepam, loratadine, oseltamivir, oxazepam, raloxifene, sulfamethoxazole, triamterene, and verapamil. The highest five annual median concentrations followed the same trend in both rivers: metformin>gabapentin>paracetamol>fexofenadine>tramadol, indicating that usage patterns, WWTP removal and environmental fate for the most prevalent pharmaceuticals are similar in these two systems. The range, detection frequency and annual median for each pharmaceutical in both river systems is reported in Tables 1 and 2.

Monthly total pharmaceutical concentrations at each sampling site are presented in Figures 4 and 5. These concentration figures provide a spatiotemporal overview of the relationship between sampling sites, rivers, and WWTPs serving the city. Monthly summed concentrations are higher in the River Foss (e.g. above 2000 ng/L) at sites downstream of the WWTP in comparison to the River Ouse, where most monthly summed concentrations are below 1000 ng/L despite the WWTPs on the River Ouse serving a larger population. This is due to greater dilution of discharged effluent in the Ouse; for example, flow ranged from 9.2 to 233 m³/s in the Ouse, compared with 0.0096 to 1.68 m³/s in the Foss on sampling days (Figure 1). For the sites immediately downstream of the WWTPs (O3, O6, and F2), the months with the lowest flows, July and June, yielded both the most analytes and the highest concentrations. Thus, concentrations appear to be inversely proportional to flow at site F2, similarly to observations reported previously (Kolpin et al., 2004). The trend is not continued moving downstream in the River Foss (sites F3-F5), potentially due to pharmaceutical losses stemming from dilution or in-stream removal processes such as biodegradation or sorption to sediment (Moreno-González et al., 2014), or due to pharmaceutical contributions from domestic septic systems (Carmona et al., 2014), and/or inputs from combined sewer overflows (CSO) (Phillips et al., 2012). In the Foss, a substantial spike downstream of F2 in paracetamol (9822 ng/L) was detected in the March sampling along with less intense spikes from other pharmaceuticals, such as metformin (2592 ng/L). These observations may be explained by local septic tank effluent entering the river downstream of the F2 site, captured during the March sampling period. Paracetamol can be >99% removed and metformin >93%, in conventional water treatment (Figure 3), therefore the spike in March concentrations might be explained by releases of septic effluent (James et al., 2016). James et al. (2012) reported paracetamol concentrations of 5000 ng/L at a septic effluent impacted site and identified it as a possible tracer of septic

system contamination. Combined sewer overflow (CSO) releases could provide an alternative explanation for the concentration spike (Philips et al. 2012), as a CSO is located just upstream of the F3 site. Low rainfall (University of York, 2018) prior to sampling suggest CSOs would not likely be in operation, therefore septic effluent releases provide a plausible explanation. Concentrations in the River Ouse varied less month to month than in the Foss, and a relationship with flow was less clear, with March and May in general having slightly greater total concentrations. March has also been reported to have the highest monthly concentration in recent temporal studies (Padhye et al., 2014; Sun et al., 2014). Sun et al. (2014) suggested March coincided with a spike in pharmaceutical usage and reduced WWTP removal capacity. This may explain the slightly higher concentrations observed in the River Ouse at sites upstream of the Foss confluence (O1-O4), while the spike in May (River Ouse) coincides with decreased river flow (Figure 1).

Metformin, a type II diabetes drug, had the highest annual median concentration (1117 and 237 ng/L in the Foss and Ouse, respectively), followed by gabapentin (anti-convulsant) (843 and 230 ng/L, Foss and Ouse, respectively) and paracetamol (analgesic) (209 and 77.6 ng/L, Foss and Ouse, respectively). This trend is different from those observed in previous temporal exposure campaigns studying similar compounds throughout the world. For example in China, Zhang et al. (2015) studied urbanized rivers and found antibiotics the most frequently detected pharmaceuticals. They did, however, report atenolol as having one of the highest annual median concentrations (53 ng/L), which is similar to the median concentration for this compound reported at site F2 (55.4 ng/L) in the current study. In Spain, Camacho-Munoz et al. (2010) reported propranolol most frequently detected in surface water, with a higher average concentration (80 ng/L) than observed in this study (20.1 ng/L). In Portugal, Paíga et al. (2016) reported carbamazepine the most frequently detected pharmaceutical with an annual median of

31.7 ng/L, while other similarly studied compounds, citalopram and venlafaxine had annual median concentrations of 0.86 and 40.1 ng/L, respectively and trimethoprim was not detected. In the River Foss, the highest annual median concentrations for carbamazepine, citalopram and venlafaxine was 66 ng/L, 15.4 and 21 ng/L, respectively while trimethoprim was detected in 100% of samples with an annual median of 30 ng/L. In Sweden, carbamazepine was also most frequently detected and at a higher annual mean than observed in York, 204 ng/L versus 66 ng/L in the River Foss, while atenolol concentration was similar to that reported here (60.2 ng/L, compared to 55.4 ng/L) (Daneshvar et al., 2010). In a similar temporal study in Wales, tramadol and gabapentin had the highest annual median concentrations (968 ng/L and 227 ng/L, respectively) (Kasprzyk-Hordern et al., 2008). Median concentrations of: gabapentin, tramadol, trimethoprim, paracetamol, carbamazepine, cimetidine and atenolol, in Wales were higher than we saw in York, while concentrations of diltiazem, atenolol, sulfamethoxazole, and erythromycin concentrations in the River Foss were lower than observed in Wales (Kasprzyk-Hordern et al., 2008). These comparisons suggest that annual pharmaceutical exposures in river systems are highly variable regionally, in part due to variability in prescribing practices, hydrology, wastewater management, and the degree of urbanisation. In addition, certain annual median concentrations of pharmaceuticals observed in this study are higher than those previously observed in the European Union and Asia.

3.3.1 Spatial Trends

The spatial trends for both rivers are presented in Figure 6; significant differences between a site and the adjacent downstream site are also noted. Spatial trends are apparent in both rivers, the greatest number of significant differences ($p < 0.05$) were found between the sites upstream and downstream of the WWTPs (i.e. F1-F2, O3-O4 and O5-O6) (Figure 6). In addition, significance increases were found when comparing

to sites further downstream. WWTPs make a significant contribution to pharmaceutical concentrations in both river systems, however upstream sources of certain pharmaceuticals exist in both rivers as significance was not achieved for cimetidine in the Foss and paracetamol, codeine, trimethoprim, and atenolol in the Ouse. There are WWTPs along the River Nidd (Figure 5) and upstream of sites O1 and F1 (>10 km) demonstrating that pharmaceuticals from upstream sources are transported into the city. Concentrations are generally highest immediately downstream of the WWTPs and decrease moving to downstream sites, evidenced by difference in height (i.e. concentration) between the bars from each site (Figure 6), similarly to observations in previous studies (Kasprzyk-Hordern et al., 2008). The decrease in concentrations moving downstream is variable between compounds indicating that in-stream attenuation is compound specific. For example, carbamazepine concentrations are similar between sites downstream of the WWTP in the River Foss (i.e. F2-F5), while over the same stretch of river concentrations of hydrocodone and citalopram decreases by 51% and 38%, respectively (Figure 6). In the Ouse, all concentrations decrease slightly from O3 to O4, however there is a slight increase occurring at O5, likely due to the confluence with the River Foss and again at O6, which is downstream of WWTP C.

In the River Foss, carbamazepine had only a single significant spatial difference between the site upstream of WWTP A discharge (site F1) and the sites downstream of the discharge. Carbamazepine has been reported to be resistant to biodegradation and stable in the environment (Moreno-González et al., 2014). In the River Ouse, all pharmaceuticals exhibited spatially significant trends. Carbamazepine was significantly different between each site downstream of WWTP B tested (i.e. O3 to O6). Since this did not occur in the River Foss over a similar distance, 13.3 km between sites F2 and F5 versus 11 km between sites O3 and O6, and the literature agrees that carbamazepine is resistant to biotransformation, a combination of dilution (e.g. urban drainage/runoff) and

other pharmaceutical sources (i.e. River Foss) moving downstream could be a plausible explanation.

Overall, these results indicate that a wide variety of environmental processes such as dilution and in-stream degradation are operating to differing extents in neighbouring rivers leading to different spatial patterns in pharmaceutical concentrations between sampling sites. For example, the reduction in concentrations moving downstream in the River Foss may be symptomatic of in-stream removal processes such as photolysis or microbial degradation (Daneshvar et al., 2010), while fluctuating concentrations in the River Ouse could be due to a complex dynamic between dilution and other pharmaceutical sources (i.e. tributaries, urban drainage) while natural removal processes potentially operating in the Foss may be masked or occur to a lesser extent in the larger Ouse system.

3.3.2 Seasonal Variability

Temporal variability between the seasons (Figure 7) is presented similarly to the approach for displaying spatial variability between sampling sites (Figure 6). Seasonal differences in pharmaceutical concentrations exist in the two river systems, especially in the River Foss. In both rivers, the lowest concentrations correspond with winter, the season which had the highest average flow (2.7 times higher than the next highest season, autumn). Conversely, the highest mass loads occur in winter, 1.4 times higher than the next highest season, spring. Lower concentrations in winter have also been reported previously (Baker and Kasprzyk-Hordern, 2013; Kasprzyk-Hordern et al., 2008), however several studies report higher concentrations in winter (Kot-Wasik et al., 2016; Lindholm-Lehto et al., 2016; Zhang et al., 2015). In addition, the extent of concentration variability between seasons differs between compounds, which could be due to seasonal patterns in usage (Sun et al., 2014) or seasonal variability in photodegradation or biodegradation, of which both processes can peak in summer, thus having a greater

impact on more readily biodegradable compounds (Lindholm-Lehto et al., 2016). In general, autumn was the season with the second highest median concentrations, except for paracetamol, where highest median values were observed during spring in both rivers. This could be due to increased usage coinciding with symptomatic treatment of illnesses more common in spring such as colds (Vatovec et al., 2016) in conjunction with lower flows than winter. To determine whether concentrations between seasons were significant, Friedman's test was used for pharmaceuticals with sufficient detections. Concentrations of 17 compounds (86%) were found to vary significantly by season in the River Foss, while amitriptyline, codeine, cimetidine, metformin, and ranitidine did not vary seasonally. Nine compounds (50%) had significant seasonal differences in the River Ouse, atenolol, carbamazepine, codeine, desvenlafaxine, gabapentin, lidocaine, ranitidine, sitagliptin, and trimethoprim.

The reasons for temporal variations in pharmaceutical concentrations have varied between studies with several reporting flow as the major driver, observing higher concentrations during times of low flow (Kasprzyk-Hordern et al., 2008; Kolpin et al., 2004). Others suggest higher pharmaceutical concentrations in winter months coincide with higher winter usage patterns (Sun et al., 2014) or decreased biodegradation in winter (Moreno-González et al., 2014), while others found no significant differences between sampled seasons (Camacho-Munoz et al., 2010). Due to higher concentrations coinciding with low-flow months in this study, we also suggest that flow appears to be a major driver behind the observed seasonal variability in pharmaceutical concentrations in the current study. The lack of significant seasonal differences found in the River Ouse could be explained by a lower annual variability in flow on sampling days than the River Foss (i.e. two orders of magnitude versus three). Further detailed investigation into the drivers behind the pharmaceutical concentrations observed both temporally and spatially is required to differentiate between the possible explanations, and could include

comparing results with prescription data and flow and estimating the impact of in-stream losses seasonally in different climates, and in river hydrological properties (e.g. depth and flow). Such analyses will be facilitated by the detailed pharmaceutical monitoring data reported in this study.

3.4 Comparisons of PECs and MECs

The PEC/MEC ratios for each compound for which it was possible to calculate an annual average MEC are reported in Figure 8. A ratio greater than 1 indicates PECs were higher than MECs and lower when less than 1. The PECs are severely underestimated in the Ouse; this may be due to pharmaceuticals being transported from upstream or problems with sewer connectivity within the sampling network not being accounted for in the simplistic PEC calculation. Several studies have attempted to gauge the accuracy of PECs by calculating a ratio with MECs, however the criterion for what constitutes accurate is variable across studies (Burns et al., 2017). This assessment has been previously limited to a small number of compounds and based on a limited number of sampling events not representative of the annual average MEC which the PEC was designed to predict. In this way, we present novel findings that indicate when an annual average MECs is calculated, less hydrologically complex river systems where pharmaceutical sources are limited, PECs characterise annual exposure within a factor 2 for 41% of compounds in this study (average factor 2.8), with no factor greater than 11. However paracetamol is an exception (underestimated by a factor of 73); the usage estimate did not incorporate over-the-counter contributions therefore underestimates were not unexpected (Burns et al., 2017). Conversely, the results from the River Ouse indicate that major limitations are associated with this predictive approach. All ratios

were off by a factor of at least 7 (average 27) and up to 139, which according to studies characterising the PEC/MEC, is outside an acceptable range (Verlicchi et al., 2014).

As the simple exposure model is routinely used for regulatory environmental risk assessment (ERA) of new pharmaceuticals, our findings have important regulatory implications. The predictions of exposure, currently being used to assess new compounds, are likely under- or over-estimating concentrations, depending on the type of compound. The use of a spatially referenced 'down the drain' hydrological model such as LF2000-WQX (Williams et al., 2012) or GREAT-ER (Feijtel et al., 1997) would likely result in improved predictions, as these models have the capacity to incorporate inputs from upstream sources; this is appropriate, as many rivers in the region pass through multiple urbanised areas and thus are subject to multiple WWTP inputs. In addition, the hydrological aspect can incorporate contributions or dilutions from the confluence with other river systems. Work currently being performed in the iPiE project involves the development of a spatially resolved model for European surface waters. The high-quality monitoring data presented in this study will be used to help evaluate this model. Our work also shows that inputs from other sources, potentially septic effluent, can be very important for some compounds at certain time of year. The consideration of these direct inputs in the risk assessment process may therefore be warranted.

4.0 Conclusion

A rapid determination HPLC-MS/MS method for 33 pharmaceuticals was validated and applied in a 12-month spatiotemporal pharmaceutical exposure campaign. WWTP removal efficiency was found to be similar between CAS and trickling filter technology for the target pharmaceuticals. Pharmaceutical concentrations in two contrasting river systems that run through the city of York, UK were found to vary significantly spatially and temporally, with the greatest variation observed for paracetamol in the River Foss, ranging from not detected to over 9822 ng/L. Temporal variations in concentration were

less frequently observed in the larger River Ouse, potentially due to the lower variability in flow which could be an important driver behind pharmaceutical concentrations in the study system. PEC/MEC ratios indicated that compounds in both rivers were generally underestimated by commonly used simple predictive exposure algorithms. In total, 41% of PEC/MEC ratios for the River Foss data were within a factor of 2, while for the River Ouse average ratios indicated predictions were off by a factor of 27. This analytical method and extensive monitoring results will be instrumental in improving the understanding of temporal pharmaceutical fate and occurrence in river systems.

Acknowledgement

The present work is funded by the European Union's Seventh Framework Programme for research, technological development and demonstration under grant agreement no. 608014 (CAPACITIE) and partly supported by the EU/EFPIA Innovative Medicines Initiative Joint Undertaking (iPiE Grant 115635). The York Centre of Excellence in Mass Spectrometry was created thanks to a major capital investment through Science City York, supported by Yorkshire Forward, with funds from the Northern Way Initiative, and subsequent support from EPSRC (EP/K039660/1; EP/M028127/1). The authors would also like to thank the U.S. Geological Survey (USGS) Toxic Substances Hydrology Program for its support of this research and P. Phillips for valuable comments and suggestions to improve the quality of the paper. Any use of trade, firm, or product names is for descriptive purposes only and does not imply endorsement by the U.S. Government.

References

- Antweiler, R.C., 2015. Evaluation of statistical treatments of left-censored environmental data using coincident uncensored data sets. II. Group comparisons. *Environ. Sci. Technol.* 49, 13439–13446.
- Anumol, T., Wu, S., Santos, M., D., Daniels, K.D., Snyder, S.A., 2015. Rapid direct

- 596 injection LC-MS/MS method for analysis of prioritized indicator compounds in
597 wastewater effluent. *Environ. Sci. Water Res. Technol* 1, 632-643.
- 598 aus der Beek, T., Weber, F.-A., Bergmann, A., Hickmann, S., Ebert, I., Hein, A., Küster,
599 A., 2016. Pharmaceuticals in the environment-Global occurrences and
600 perspectives. *Environ. Toxicol. Chem.* 35, 823–835.
- 601 Baker, D.R., Kasprzyk-Hordern, B., 2013. Spatial and temporal occurrence of
602 pharmaceuticals and illicit drugs in the aqueous environment and during wastewater
603 treatment: New developments. *Sci. Total Environ.* 454–455, 442–456.
- 604 Blair, B., Nikolaus, A., Hedman, C., Klaper, R., Grundl, T., 2015. Evaluating the
605 degradation, sorption, and negative mass balances of pharmaceuticals and
606 personal care products during wastewater treatment. *Chemosphere* 134, 395–401.
- 607 Boix, C., Ibáñez, M., Sancho, J. V, Rambla, J., Aranda, J.L., Ballester, S., Hernández,
608 F., 2015. Fast determination of 40 drugs in water using large volume direct injection
609 liquid chromatography–tandem mass spectrometry. *Talanta* 131, 719-727.
- 610 Boxall, A.B.A., Keller, V.D.J., Straub, J.O., Monteiro, S.C., Fussell, R., Williams,
611 R.J., 2014. Exploiting monitoring data in environmental exposure modelling and risk
612 assessment of pharmaceuticals. *Environ. Int.* 73, 176–185.
- 613 Burns, E.E., Thomas-Oates, J., Kolpin, D.W., Furlong, E.T., Boxall, A.B., 2017. Are
614 exposure predictions, used for the prioritization of pharmaceuticals in the
615 environment, fit for purpose? *Environ. Toxicol. Chem.* 36, 2823-2832.
- 616 Camacho-Muñoz, D., Martín, J., Santos, J.L., Aparicio, I., Alonso, E., 2010. Occurrence,
617 temporal evolution and risk assessment of pharmaceutically active compounds in
618 Dónana Park (Spain). *J. Hazard. Mater.* 183, 602–608.
- 619 Campos-Mañas, M.C., Plaza-Bolanos, P., Sánchez-Pérez, J.A., Malato, S., Agüera, A.,
620 2017. Fast determination of pesticides and other contaminants of emerging concern
621 in treated wastewater using direct injection coupled to highly sensitive ultra-high
622 performance liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A*
623 1507, 84–94.
- 624 Carmona, E., Andreu, V., Picó, Y., 2014. Occurrence of acidic pharmaceuticals and
625 personal care products in Turia River Basin: From waste to drinking water. *Sci.*
626 *Total Environ.* 484, 53–63.

- Center for Ecology & Hydrology, 2016. National River Flow Archive. 27009- Ouse Skelton; 27083 – Foss Huntington. URL <http://nrfa.ceh.ac.uk/data/search> (accessed 3.9.17).
- Comber, S., Gardner, M., Sörme, P., Leverett, D., Ellor, B., 2018. Active pharmaceutical ingredients entering the aquatic environment from wastewater treatment works: A cause for concern? *Sci. Total. Environ.* 613-614, 538-547.
- European Commission, 2002. Commission Decision 2002 implementing Council Directive 96/23/EC concerning the performance of analytical methods and the interpretation of results (2002/657/EC). *Off. J. Eur. Communities*. L221/8-L221/36.
- Daneshvar, A., Svanfelt, J., Kronberg, L., Prévost, M., Weyhenmeyer, G.A., 2010. Seasonal variations in the occurrence and fate of basic and neutral pharmaceuticals in a Swedish river–lake system. *Chemosphere* 80, 301–309.
- Daughton, C.G., 2016. Pharmaceuticals and the Environment (PiE): Evolution and impact of the published literature revealed by bibliometric analysis. *Sci. Total Environ.* 562, 391–426.
- European Medicines Agency, 2006. Guideline on the Environmental Risk Assessment of Medicinal Products for Human Use. EMEA/CHMP/SWP/4447/00. Committee for Medicinal Products for Human Use, London, UK.
- Feijtel, T., Boeije, G., Matthies, M., Young, A., Morris, G., Gandolfi, C., Hansen, B., Fox, K., Holt, M., Koch, V., Schroder, R., Casani, G., Schowanek, D., Rosenblom, J., Niessen, H., 1997. Development of a geography-referenced regional exposure assessment tool for European rivers – GREAT-ER contribution to GREAT-ER #1. *Chemosphere* 34, 2351–2373.
- Furlong, E.T., Batt, A.L., Glassmeyer, S.T., Noriega, M.C., Kolpin, D.W., Mash, H., Schenck, K.M., 2017. Nationwide reconnaissance of contaminants of emerging concern in source and treated drinking waters of the United States: Pharmaceuticals. *Sci. Total Environ.* 579, 1629–1642.
- Furlong, E.T., Kanagy, C.J., Kanagy, L.K., Coffey, L.J., Burkhardt, M.R., 2014. Determination of human-use pharmaceuticals in filtered water by direct aqueous injection-high-performance liquid chromatography/tandem mass spectrometry: U.S. Geological Survey Techniques and Methods, Book 5, Laboratory Analysis. p. 49.

- 658 Golovko, O., Kumar, V., Fedorova, G., Randak, T., Grabic, R., 2014. Removal and
659 seasonal variability of selected analgesics/anti-inflammatory, anti-
660 hypertensive/cardiovascular pharmaceuticals and UV filters in wastewater treatment
661 plant. *Environ. Sci. Pollut. Res.* 21, 7578–7585.
- 662 Graphpad Software, 2017. Graphpad Prism version 7.00 for Windows, Graphpad
663 Software, La Jolla California USA, URL www.graphpad.com.
- 664 Herrmann, M., Menz, J., Olsson, O., Kümmerer, K., 2015. Identification of
665 phototransformation products of the antiepileptic drug gabapentin: Biodegradability
666 and initial assessment of toxicity. *Water Res.* 85, 11–21.
- 667 James, C.A.; Miller-Schulze, J.P.; Ultican, S.; Gipe, A.D. 2016. Evaluation of
668 contaminants of emerging concern from septic systems. *Water Res.* 101, 241–251.
- 669 Kasprzyk-Hordern, B., Dinsdale, R.M., Guwy, A.J., 2009. The removal of
670 pharmaceuticals, personal care products, endocrine disruptors and illicit drugs
671 during wastewater treatment and its impact on the quality of receiving waters. *Water*
672 *Res.* 43, 363–380.
- 673 Kasprzyk-Hordern, B., Dinsdale, R.M., Guwy, A.J., 2008. The occurrence of
674 pharmaceuticals, personal care products, endocrine disruptors and illicit drugs in
675 surface water in South Wales, UK. *Water Res.* 42, 3498–3518.
- 676 Kolpin, D.W., Skopec, M., Meyer, M.T., Furlong, E.T., Zaugg, S.D., 2004. Urban
677 contribution of pharmaceuticals and other organic wastewater contaminants to
678 streams during differing flow conditions. *Sci. Total Environ.* 328, 119–130.
- 679 Kot-Wasik, A., Jakimska, A., Śliwka-Kaszyńska, M., 2016. Occurrence and seasonal
680 variations of 25 pharmaceutical residues in wastewater and drinking water
681 treatment plants. *Environ. Monit. Assess.* 188, 661.
- 682 Lindholm-Lehto, P.C., Ahkola, H.S.J., Knuutinen, J.S., Herve, S.H., 2016. Widespread
683 occurrence and seasonal variation of pharmaceuticals in surface waters and
684 municipal wastewater treatment plants in central Finland. *Environ. Sci. Pollut. Res.*
685 23, 7985–7997.
- 686 Loos, R., Carvalho, R., António, D.C., Comero, S., Locoro, G., Tavazzi, S., Paracchini,
687 B., Ghiani, M., Lettieri, T., Blaha, L., Jarosova, B., Voorspoels, S., Servaes, K.,
688 Haglund, P., Fick, J., Lindberg, R.H., Schwesig, D., Gawlik, B.M., 2013. EU-wide

- 689 monitoring survey on emerging polar organic contaminants in wastewater treatment
690 plant effluents. *Water Res.* 47, 6475–6487.
- 691 Luo, Y., Guo, W., Ngo, H.H., Nghiem, L.D., Hai, F.I., Zhang, J., Liang, S., Wang, X.C.,
692 2014. A review on the occurrence of micropollutants in the aquatic environment and
693 their fate and removal during wastewater treatment. *Sci. Total Environ.* 473–474,
694 619–641.
- 695 McCallum, E.S., Bose, A.P.H., Warriner, T.R., Balshine, S., 2017. An evaluation of
696 behavioural endpoints: The pharmaceutical pollutant fluoxetine decreases
697 aggression across multiple contexts in round goby (*Neogobius melanostomus*).
698 *Chemosphere* 175, 401–410.
- 699 Mompelat, S., Jaffrezic, A., Jardé, E., Le Bot, B., 2013. Storage of natural water samples
700 and preservation techniques for pharmaceutical quantification. *Talanta* 109, 31–45.
- 701 Moreno-González, R., Rodríguez-Mozaz, S., Gros, M., Pérez-Cánovas, E., Barceló, D.,
702 León, V.M., 2014. Input of pharmaceuticals through coastal surface watercourses
703 into a Mediterranean lagoon (Mar Menor, SE Spain): Sources and seasonal
704 variations. *Sci. Total Environ.* 490, 59–72.
- 705 National Health Service, 2016. GP practice prescribing presentation-level data: January
706 to December 2016. URL <https://digital.nhs.uk/article/4214/Prescribing> (accessed
707 24.2.18).
- 708 Niemuth, N.J., Jordan, R., Crago, J., Blanksma, C., Johnson, R., Klaper, R.D., 2015.
709 Metformin exposure at environmentally relevant concentrations causes potential
710 endocrine disruption in adult male fish. *Environ. Toxicol. Chem.* 34, 291–296.
- 711 Oliveira, T.S., Murphy, M., Mendola, N., Wong, V., Carlson, D., Waring, L., 2015.
712 Characterization of Pharmaceuticals and Personal Care products in hospital effluent
713 and waste water influent/effluent by direct-injection LC-MS-MS. *Sci. Total Environ.*
714 518–519, 459–478.
- 715 Ort, C., Lawrence, M.G., Reungoat, J., Mueller, J.F., 2010. Sampling for PPCPs in
716 wastewater systems: Comparison of different sampling modes and optimization
717 strategies. *Environ. Sci. Technol.* 44, 6289–6296.
- 718 Padhye, L., Yao, H., Kung'u, F., Huang, C., 2014. Year-long evaluation on the
719 occurrence and fate of pharmaceuticals, personal care products, and endocrine

- 720 disrupting chemicals in an urban drinking water treatment plant. *Water Res.* 51,
721 266–276.
- 722 Paíga, P., Santos, L.H.M.L.M., Ramos, S., Jorge, S., Silva, J.G., Delerue-Matos, C.,
723 2016. Presence of pharmaceuticals in the Lis river (Portugal): Sources, fate and
724 seasonal variation. *Sci. Total Environ.* 573, 164–177.
- 725 Petrie, B., Youdan, J., Barden, R., Kasprzyk-Hordern, B., 2016. Multi-residue analysis of
726 90 emerging contaminants in liquid and solid environmental matrices by ultra-high-
727 performance liquid chromatography tandem mass spectrometry. *J. Chromatogr. A*
728 1431, 64–78.
- 729 Petrović, M., Hernando, M.D., Díaz-Cruz, M.S., Barceló, D., 2005. Liquid
730 chromatography-tandem mass spectrometry for the analysis of pharmaceutical
731 residues in environmental samples: A review. *J. Chromatogr. A* 1067, 1–14.
- 732 Phillips, P.J., Chalmers, A.T., Gray, J.L., Kolpin, D.W., Foreman, W.T., Wall, G.R., 2012.
733 Combined Sewer Overflows: An Environmental Source of Hormones and
734 Wastewater Micropollutants. *Environ. Sci. Technol.* 46, 5336–5343.
- 735 Sanchez, W., Sremski, W., Piccini, B., Palluel, O., Maillot-Maréchal, E., Betoulle, S.,
736 Jaffal, A., Aït-Aïssa, S., Brion, F., Thybaud, E., Hinfrey, N., Porcher, J.-M., 2011.
737 Adverse effects in wild fish living downstream from pharmaceutical manufacture
738 discharges. *Environ. Int.* 37, 1342–1348.
- 739 Silva, L.J.G., Pereira, A.M.P.T., Meisel, L.M., Lino, C.M., Pena, A., 2014. A one-year
740 follow-up analysis of antidepressants in Portuguese wastewaters: Occurrence and
741 fate, seasonal influence, and risk assessment. *Sci. Total Environ.* 490, 279–287.
- 742 Stüber, M. Reemtsma, T., 2004. Evaluation of three calibration methods to compensate
743 matrix effect in environmental analysis with LC-ESI-MS. *Anal. Bioanal. Chem.* 378,
744 910–196
- 745 Sun, Q., Lv, M., Hu, A., Yang, X., Yu, C.-P., 2014. Seasonal variation in the occurrence
746 and removal of pharmaceuticals and personal care products in a wastewater
747 treatment plant in Xiamen, China. *J. Hazard. Mater.* 277, 69–75.
- 748 Thompson, W.A., Arnold, V., Vijayan, M.M., 2017. Venlafaxine in embryos stimulates
749 neurogenesis and disrupts larval behavior in zebrafish. *Environ. Sci. Technol.* 51,
750 12889–12897.

- University of York, 2018. Department of Electronics: Data Archive: *March 2016*. URL <https://weather.elec.york.ac.uk/data/vaisala/archives/0316.txt> (accessed 24/02/2018).
- USEPA, 2016. Method 542: Determination of pharmaceuticals and personal care products in drinking water by solid phase extraction and liquid chromatography electrospray ionisation tandem mass spectrometry (LC/ECS-MS/MS). EPA 815-R-15-012. 35 p.
- Van De Steene, J.C., Mortier, K.A., Lambert, W.E., 2006. Tackling matrix effects during development of a liquid chromatographic–electrospray ionisation tandem mass spectrometric analysis of nine basic pharmaceuticals in aqueous environmental samples. *J. Chromatogr. A* 1123, 71–81.
- Vasquez, M.I., Lambrianides, A., Schneider, M., Kümmerer, K., Fatta-Kassinos, D., 2014. Environmental side effects of pharmaceutical cocktails: What we know and what we should know. *J. Hazard. Mater.* 279, 169–189.
- Vatovec, C., Phillips, P., Van Wagoner, E., Scott, T.-M., Furlong, E., 2016. Investigating dynamic sources of pharmaceuticals: Demographic and seasonal use are more important than down-the-drain disposal in wastewater effluent in a University City setting. *Sci. Total Environ.* 572, 906–914.
- Verlicchi, P., Al Aukidy, M., Jelic, A., Petrović, M., Barceló, D., 2014. Comparison of measured and predicted concentrations of selected pharmaceuticals in wastewater and surface water: A case study of a catchment area in the Po Valley (Italy). *Sci. Total Environ.* 470–471, 844–854.
- Verlicchi, P., Al Aukidy, M., Zambello, E., 2012. Occurrence of pharmaceutical compounds in urban wastewater: Removal, mass load and environmental risk after a secondary treatment-A review. *Sci. Total Environ.* 429, 123–155.
- Wang, S., Cyronak, M., Yang, E., 2007. Does a stable isotopically labelled internal standard always correct analyte response? A matrix effect study on a LC/MS/MS method for the determination of carvedilol enantiomers in human plasma. *J. Pharm. Biomed. Anal.* 43, 701–707.
- Williams, R.J., Churchley, J.H., Kanda, R., Johnson, A.C., 2012. Comparing predicted against measured steroid estrogen concentrations and the associated risk in two United Kingdom river catchments. *Environ. Toxicol. Chem.* 31, 892–898.

- Yu, K., Li, B., Zhang, T., 2012. Direct rapid analysis of multiple PPCPs in municipal wastewater using ultrahigh performance liquid chromatography–tandem mass spectrometry without SPE pre-concentration. *Anal. Chim. Acta* 738, 59–68.
- Zhang, H., Du, M., Jiang, H., Zhang, D., Lin, L., Ye, H., Zhang, X., 2015. Occurrence, seasonal variation and removal efficiency of antibiotics and their metabolites in wastewater treatment plants, Jiulongjiang River Basin, South China. *Environ. Sci. Process. Impacts* 17, 225–234.

Table 1. Summary results (ng/L) for the River Foss from the January to December 2016 monitoring campaign. The annual median, concentration range and frequency of detection for each sampling site are reported.

Compound	F1		F2		F3		F4		F5	
	Range (Med)	%	Range (Med)	%	Range (Med)	%	Range (Med)	%	Range (Med)	%
Amitriptyline	n.d.	0	n.d. – 25.7 (10.3)	92	1.2* – 12.2 (5.7)	100	n.d. – 11.2 (2.6)	83	n.d. – 6.4 (2.0*)	75
Atenolol	n.d.	0	18.9 – 100 (55.4)	100	12.3* – 98.2 (43.6)	100	13.7* – 97.8 (34.8)	100	10.1* – 67.0 (21.8)	100
Carbamazepine	n.d. – 11.8 (4.5)	67	19.0 – 195 (45.2)	100	8.7 – 194 (66.0)	100	12.5 – 175 (61.6)	100	11.4 – 193 (36.8)	100
Cimetidine	n.d. – 49.6 (19.8)	83	n.d. – 44.0 (19.9)	92	3.0* – 40.5 (10.6)	100	2.1* – 16.9 (7.3*)	100	n.d. – 11.8 (7.2*)	67
Citalopram	n.d.	0	5.0 – 71.4 (15.4)	100	3.8* – 31.0 (15.3)	100	3.1* – 13.5 (7.8)	100	n.d. – 11.4 (5.9)	83
Codeine	n.d. – 10.8 (5.9*)	83	8.0 – 101 (59.2)	100	11.5 – 84.2 (57.3)	100	12.9 – 97.7 (44.0)	100	12.0 – 64.7 (29.1)	100
Desvenlafaxine	n.d. – 55.8 (16.8)	83	25.8 – 268 (70.0)	100	4.6* – 195 (86.2)	100	11.7 – 170 (77.3)	100	8.5* – 96.4 (44.5)	100
Diazepam	n.d.	0	n.d. – 1.6* (n.d.)	8.3	n.d. – 1.6* (n.d.)	8.3	n.d. – 1.8* (n.d.)	8.3	n.d. – 2.3* (n.d.)	8.3
Diltiazem	n.d. – 4.1 (1.2*)	75	4.7 – 48.7 (16.4)	100	4.7 – 36.0 (14.5)	100	4.4 – 25.0 (10.6)	100	n.d. – 12.7 (5.8)	92
Diphenhydramine	n.d.	0	n.d. – 12.7 (9.5)	67	n.d. – 3.8 (n.d.)	25	n.d. – 1.6* (n.d.)	17	n.d. – 3.4 (n.d.)	8.3
Erythromycin	n.d. – 34.5 (20.2*)	58	26.8 – 242 (90.0)	100	15.0* – 263 (88.8)	100	18.8* – 142 (80.5)	100	14.4 – 116 (45.9)	100
Fexofenadine ¹	n.d. – 104 (24.9)	83	43.8 – 1144 (177)	100	17.2 – 956 (253)	100	27.5 – 638 (166)	100	26.4 – 268 (92.5)	100
Gabapentin	17.4* – 229 (82.7)	100	476 – 1429 (789)	100	260 – 1445 (843)	100	404 – 1183 (768)	100	223 – 1341 (544)	100
Hydrocodone	n.d. – 5.7 (n.d.)	43	11.2 – 91.8 (21.6)	100	6.4 – 60.3 (25.0)	100	6.8 – 43.5 (20.6)	100	5.2 – 22.2 (11.1)	100
Lidocaine	n.d. – 3.9 (2.6*)	58	4.6 – 40.4 (8.2)	100	1.7* – 39.7 (11.8)	100	3.1 – 36.9 (10.4)	100	n.d. – 16.0 (6.1)	92
Loratadine	n.d.	0	n.d.	0	n.d. – 6.46 (n.d.)	8.3	n.d.	0	n.d.	0
Metformin	45.2 – 291 (121)	100	246 – 1783 (856)	100	266 – 2339 (1117)	100	340 – 2595 (888)	100	263 – 1750 (664)	100

Table 1. Summary results for the River Foss from the January to December 2016 monitoring campaign. The annual median, concentration range and frequency of detection for each sampling site are reported.

Compound	F1		F2		F3		F4		F5	
	Range (Med)	%	Range (Med)	%	Range (Med)	%	Range (Med)	%	Range (Med)	%
Norethisterone	n.d.	0	n.d. – 7.4* (n.d.)	8.3	n.d.	0	n.d.	0	n.d.	0
Oseltamivir	n.d.	0	n.d. – 8.8* (n.d.)	8.3	n.d.	0	n.d.	0	n.d.	0
Oxazepam	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Paracetamol	n.d. – 119 (60.0)	67	14.3* – 749 (74.4)	100	n.d. – 9822 (97.2)	92	32.0 – 9676 (209)	100	25.0 – 5445 (180)	100
Propranolol	n.d.	0	n.d. – 64.9 (17.8)	92	n.d. – 29.9 (20.1)	92	n.d. – 20.6 (10.0*)	92	n.d. – 18.3 (10.4*)	50
Raloxifene	n.d.	0	n.d.	0	n.d. – 7.2*	8.3	n.d. – 7.2*	8.3	n.d.	0
Ranitidine	n.d. – 10.8* (n.d.)	17	n.d. – 69.6 (53.4)	83	6.6* – 74.0 (27.9)	100	n.d. – 60.6 (22.2)	92	n.d. – 30.0 (13.6*)	92
Sertraline	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d. – 21.2 (n.d.)	8.3
Sitagliptin	n.d.	0	16.5 – 121 (35.2)	100	9.3* – 103 (46.5)	100	15.2 – 85.7 (36.9)	100	12.2* – 33.9 (19.5)	100
Sulfamethoxazole	n.d.	0	n.d. – 10.2* (n.d.)	33	n.d. – 33.0 (n.d.)	50	n.d. – 27.5 (n.d.)	42	n.d. – 18.1* (n.d.)	17
Temazepam	n.d.	0	n.d. – 38.2 (12.1)	67	n.d. – 25.0 (16.7)	75	n.d. – 27.8 (15.9)	67	n.d. – 12.6 (7.1*)	58
Tramadol	n.d. – 48.1 (31.2)	75	54.4 – 650 (117)	100	21.0 – 456 (177)	100	34.0 – 368 (169)	100	29.2 – 201 (84.7)	100
Triamterene	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Trimethoprim	n.d. – 9.8 (2.5*)	75	13.2 – 76.0 (30.3)	100	10.1 – 60.3 (26.4)	100	15.2 – 49.4 (19.8)	100	5.3 – 38.0 (13.8)	100
Venlafaxine	n.d. – 4.3 (2.2*)	42	9.2 – 102 (16.2)	100	2.4* – 82.6 (20.6)	100	5.9 – 37.9 (17.6)	100	2.3* – 17.8 (9.2)	100
Verapamil	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0

*Below LOQ

¹ data for 11 months only available (April 2016 missing).

n.d. No detect

(Med) Median

% Detection frequency (100% = 12 months)

Table 2. Summary results (ng/L) for the River Ouse from the January to December 2016 monitoring campaign. The annual median, concentration range and frequency detection for each sampling site are reported.

Compound	O1		O2		O3		O4		O5		O6	
	Range (med)	%	Range (med)	%	Range (med)	%	Range (med)	%	Range (med)	%	Range (med)	%
Amitriptyline	n.d.	0	n.d.	0	n.d. – 2.7 (n.d.)	17	n.d. -1.2* (n.d.)	17	n.d. – 1.5* (n.d.)	8	n.d. -2.5 (n.d.)	17
Atenolol	n.d.	0	n.d. – 22.0 (11.1*)	58	n.d. – 19.5 (10.7*)	67	n.d. – 16.9* (10.2*)	75	n.d. – 20.4 (10.4*)	67	n.d. – 18.8 (13.6*)	92
Carbamazepine	1.0* – 14.0 (5.8)	100	1.1* - 34.8 (9.2)	100	1.4* - 54.4 (19.2)	100	1.1* - 31.4 (12.1)	100	1.7* - 33.9 (15.0)	100	7.9 – 48.0 (23.4)	100
Cimetidine	n.d. – 2.3* (n.d.)	8	n.d. – 2.4* (n.d.)	8	n.d. - 5.7* (n.d.)	33	n.d. – 2.9* (n.d.)	17	n.d.	0	n.d. – 3.7 (n.d.)	42
Citalopram	n.d. - 3.3* (n.d.)	8	n.d. – 3.7* (n.d.)	33	n.d. – 7.0 (4.0*)	75	n.d. – 3.2* (n.d.)	50	n.d. – 4.0* (2.2*)	67	n.d. – 7.2 (4.8)	83
Codeine	n.d. – 13.5 (10.5*)	92	3.3 – 17.1 (10.7)	100	3.0* – 20.5 (14.3)	100	3.5* – 17.5 (13.8)	100	4.5* – 17.4 (14.9)	100	6.4* - 17.8 (8.8)	100
Desvenlafaxine	n.d. – 14.8 (n.d.)	50	n.d. – 27.5 (11.3)	75	n.d. – 46.8 (21.5)	83	n.d. -31.0 (14.2)	83	n.d. – 28.8 (15.2)	75	12.3 – 40.1 (26.8)	100
Diazepam	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Diltiazem	n.d. – 1.6* (n.d.)	25	n.d. – 2.5 (n.d.)	50	n.d. – 8.0 (3.6)	92	n.d. – 6.4 (1.8*)	67	n.d. – 3.7 (1.8*)	75	n.d. – 4.3 (3.7)	92
Diphenhydramine	n.d.	0	n.d. – 1.7* (n.d.)	8	n.d.- 2.9 (n.d.)	25	n.d.	0	n.d.- 4.8 (n.d.)	8	n.d. - 2.2* (n.d.)	8
Erythromycin	n.d.	0	n.d. – 17.3* (n.d.)	33	n.d. – 31.1 (21.3*)	92	n.d. – 20.3* (15.3*)	67	n.d. – 21.7* (n.d.)	50	n.d. – 33.9 (21.3*)	83
Fexofenadine ¹	n.d. – 41.7 (17.9)	83	n.d. – 48.7 (24.1)	83	n.d. – 77.8 (46.1)	92	n.d. – 68.2 (25.8)	83	n.d. – 44.0 (29.2)	92	7.4* – 98.5 (33.4)	100
Gabapentin	28.1* -242 (130)	100	39.4 – 351 (191)	100	24.5* - 429 (230)	100	30.0* - 369 (202)	100	33.8* - 364 (192)	100	39.5 – 450 (208)	100
Hydrocodone	n.d. – 2.9 (n.d.)	50	n.d. – 5.7 (3.6)	83	n.d. – 14.9 (7.8)	92	n.d. – 8.0 (4.0)	92	n.d. – 6.9 (4.0)	92	2.2 – 10.7 (6.0)	100
Lidocaine	n.d. – 4.1 (n.d.)	50	n.d. – 5.0 (2.7*)	83	n.d. – 6.5 (3.7)	92	n.d. – 5.4 (2.8)	83	n.d. – 5.6 (3.1)	83	1.6* – 8.8 (4.1)	100
Loratadine	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Metformin	52.5 – 323 (180)	100	63.4 – 431 (223)	100	60.6 – 422 (237)	100	60.2 – 422 (237)	100	73.6 – 445 (233)	100	142 – 483 (276)	100
Norethisterone	n.d.	0	n.d. -7.7 (n.d.)	8	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Oseltamivir	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0

Table 2. Summary results (ng/L) for the River Ouse from the January to December 2016 monitoring campaign. The annual median, concentration range and frequency detection for each sampling site are reported.

Compound	O1		O2		O3		O4		O5		O6	
	Range (med)	%	Range (med)	%	Range (med)	%	Range (med)	%	Range (med)	%	Range (med)	%
Oxazepam	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Paracetamol	22.3* – 191 (46.4)	100	15.4* – 202 (51.7)	100	16.8* – 186 (54.5)	100	20.1* – 186 (54.3)	100	22.7 – 369 (77.6)	100	21.2 – 226 (66.9)	100
Propranolol	n.d.	0	n.d.	0	n.d. – 8.3* (n.d.)	33	n.d.	0	n.d.	0	n.d. – 7.6* (n.d.)	8
Raloxifene	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Ranitidine	n.d. – 10.3* (n.d.)	25	n.d. – 10.5* (n.d.)	25	n.d. – 30.6 (15.1*)	75	n.d. – 13.3* (n.d.)	42	n.d. – 12.0* (n.d.)	25	n.d. – 15.5* (9.2*)	75
Sertraline	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Sitagliptin	n.d. – 10.7 (n.d.)	33	n.d. – 16.2 (9.3*)	75	n.d. – 32.5 (15.0)	92	n.d. – 16.9 (12.0*)	83	n.d. – 15.8 (10.4*)	83	n.d. – 26.5 (18.2)	92
Sulfamethoxazole	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Temazepam	n.d.	0	n.d.	0	n.d. – 7.2* (n.d.)	8	n.d.	0	n.d. – 4.4* (n.d.)	8	n.d. – 4.7* (n.d.)	8
Tramadol	n.d. – 27.0 (19.6)	83	3.9* – 39.9 (19.8)	100	n.d. – 57.2 (34.6)	92	n.d. – 44.8 (28.9)	92	n.d. – 47.4 (27.4)	92	20.7 – 52.4 (40.5)	100
Triamterene	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Tramadol	n.d. – 27.0 (19.6)	83	3.9* – 39.9 (19.8)	100	n.d. – 57.2 (34.6)	92	n.d. – 44.8 (28.9)	92	n.d. – 47.4 (27.4)	92	20.7 – 52.4 (40.5)	100
Triamterene	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0
Trimethoprim	n.d. – 19.0 (2.7)	92	2.0* – 8.9 (5.3)	100	2.8* – 19.3 (12.4)	100	n.d. – 11.1 (5.4)	92	2.3* – 12.1 (5.5)	100	7.3 – 22.9 (14.2)	100
Venlafaxine	n.d. – 2.6* (n.d.)	42	n.d. – 5.2 (2.6*)	75	n.d. – 8.5* (4.9)	83	n.d. – 4.3 (2.9*)	75	n.d. – 5.0 (3.1)	75	n.d. – 8.2 (4.5)	83
Verapamil	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0	n.d.	0

* Below LOQ

¹ data for 11 months only available (April 2016 missing).

n.d. No detect

(Med) Median

% Detection frequency (100% = 12 months)

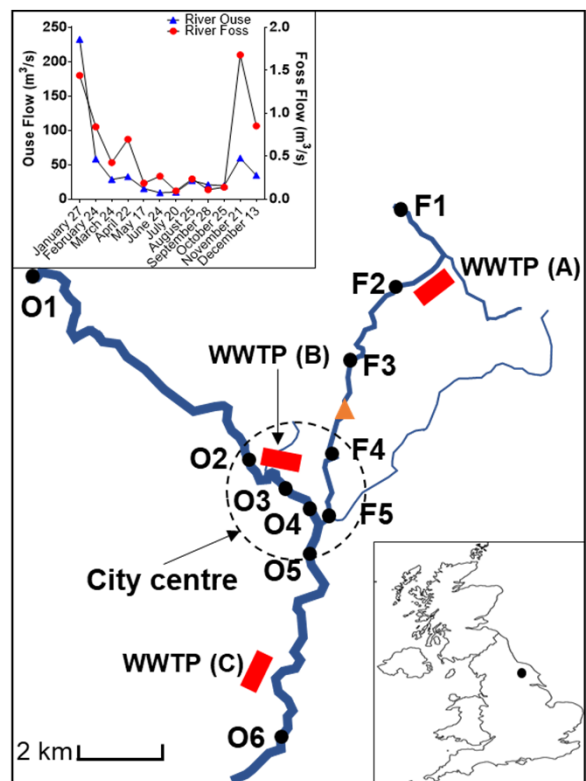


Figure 1. Map of 11 sampling sites within the sampling network. River flows recorded from a gauge in each river (orange triangle) from each sampling day (m^3/s) are pictured top left. WWTPs that serve the city (3) are represented by the red rectangles. Sites F1-F5 are along the smaller River Foss, while sites O1-O6 are along the larger River Ouse.

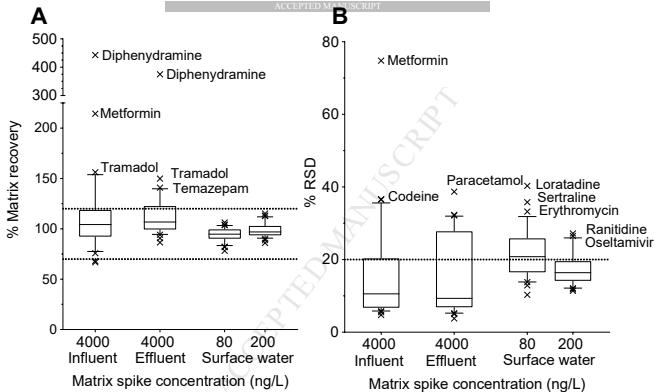


Figure 2. A) Routine matrix spikes run alongside environmental samples during the 12-month monitoring campaign in WWTP influent, effluent, surface water and reagent water. The dotted lines represent the 70 – 120% acceptable recovery range. B) %RSD of matrix spike replicates. An RSD below 20% is desirable (depicted with dotted line). The median, 25th and 75th quartiles are presented while the whiskers represent the 10th to 90th percentile, compounds outside this range are depicted with an X.

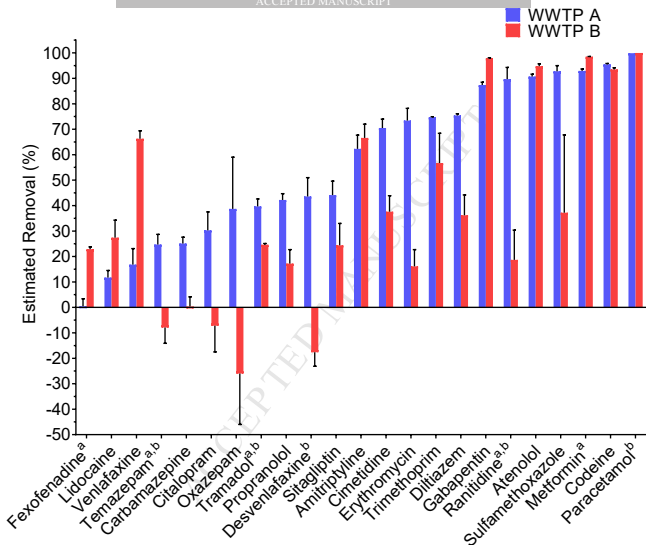


Figure 3. Estimated % removal in WWTP A (trickling filter), WWTP B (carbon activated sludge). Hydrocodone not shown, estimated removal in WWTP A -307% and in WWTP B -597%. Matrix recovery was outside the 70 -120% desired range is identified for (a) influent and (b) effluent.

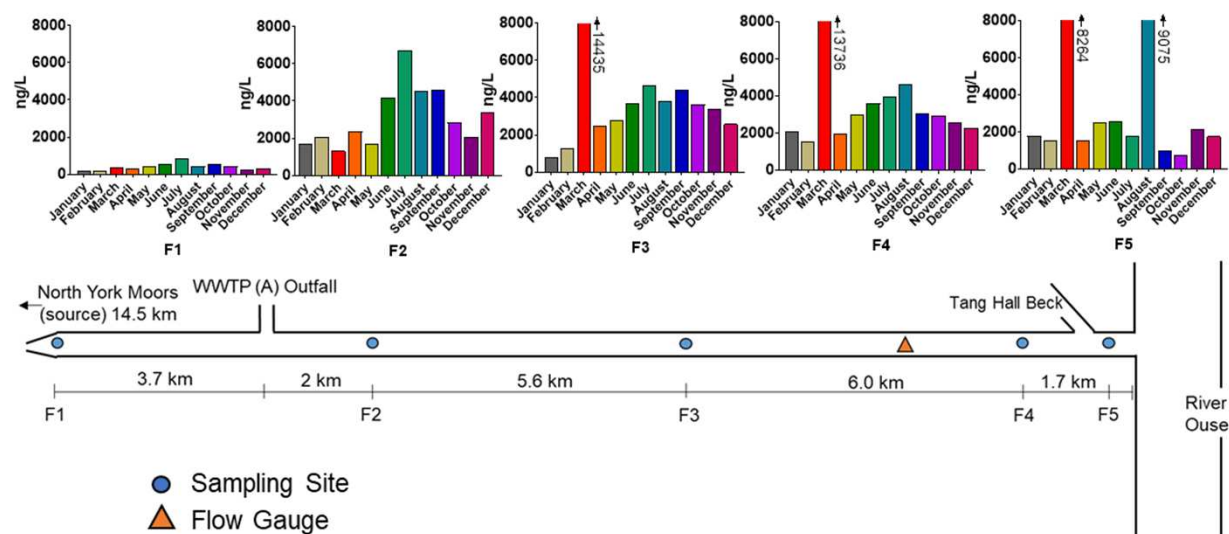


Figure 4. Total pharmaceutical concentration (summed) of all detected analytes at each sampling site from each month during 2016 in the River Foss. Sampling locations (blue circles) in relation to Environment Agency Flow gauges (orange triangles) are depicted along the river.

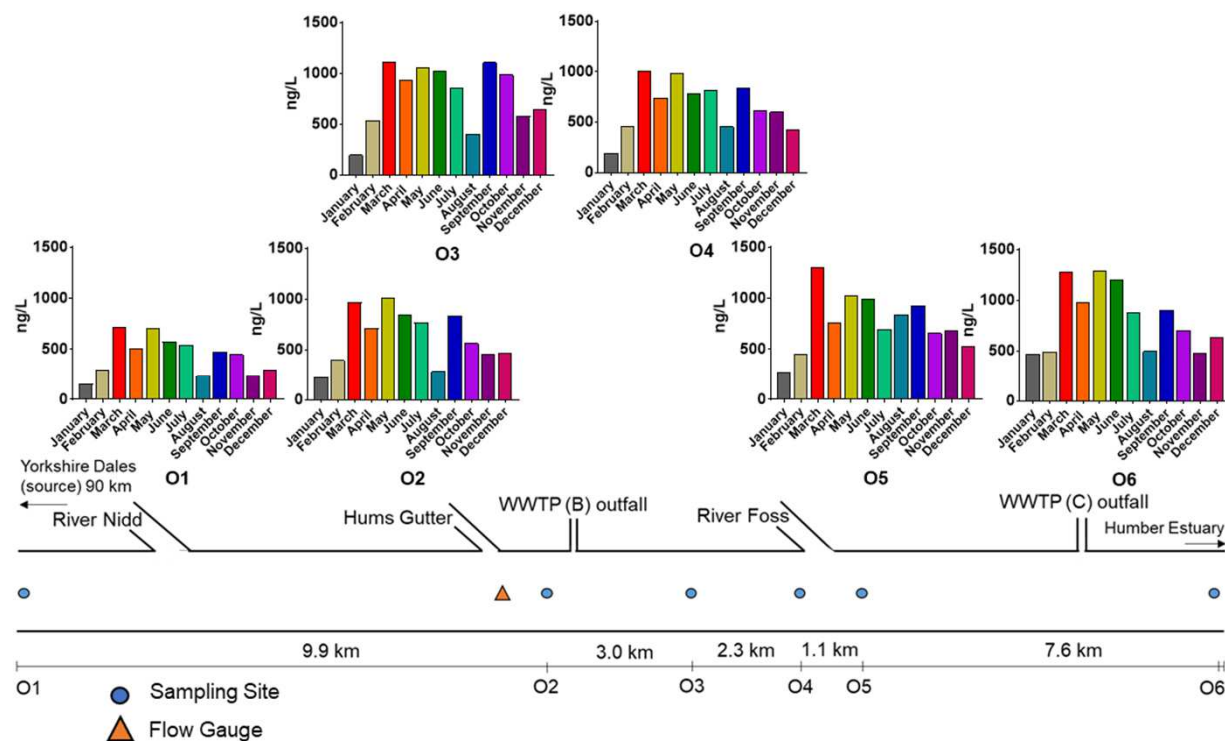


Figure 5. Total pharmaceutical concentration (summed) of all detected analytes at each sampling site from each month during 2016 along the River Ouse. Sampling locations (blue circles) in relation to Environment Agency Flow gauges (orange triangles) are depicted along the river.

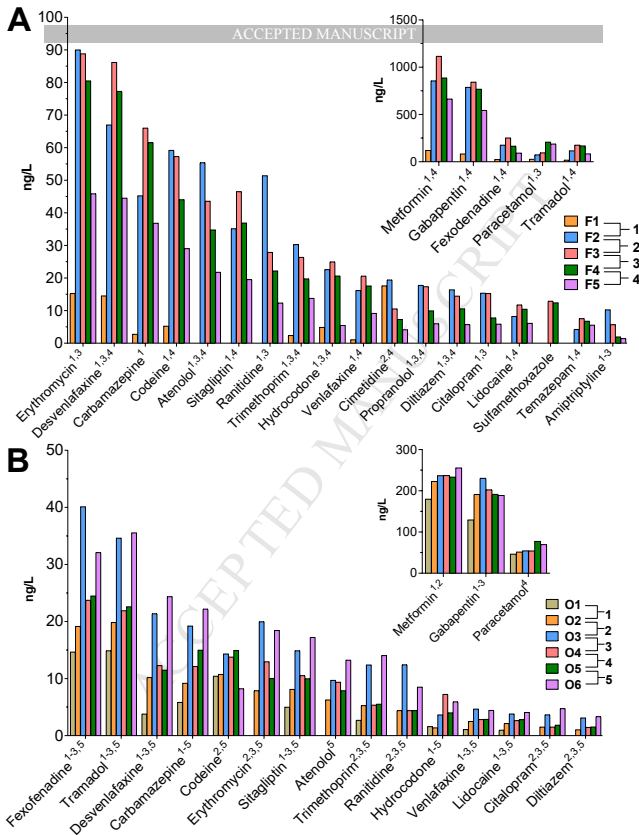


Figure 6. Annual median concentration from all sampled sites in (A) the River Foss and (B) River Ouse. Pairwise t-tests were conducted between neighbouring sites and significant differences are denoted by the corresponding number. Sites F1-F2, O1-O2 =1; F2-F3, O2-O3 =2; F3-F4, O3-O4 =3; F4-F5, O4-O5 =4; O5-O6 =5.

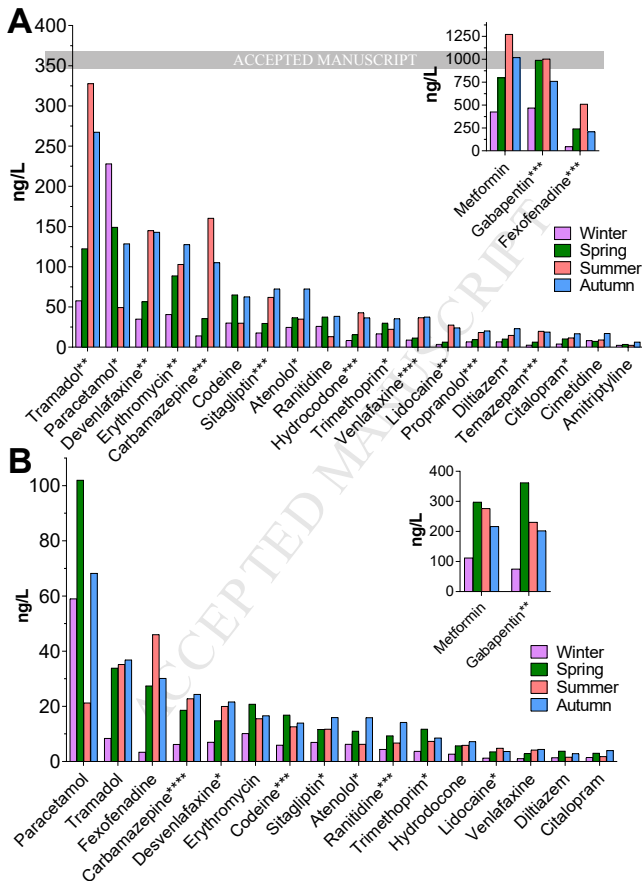


Figure 7. Median seasonal concentration from sites F3-F4 in the River Foss (A) and O3-O4 in the River Ouse (B) for select pharmaceuticals. Temporal variations were tested using Friedman's Test and results are reported for each compound where a significant result was found, $p < 0.05$ (*), $p < 0.01$ (**), $p < 0.0005$ (***), $p < 0.0001$ (****).

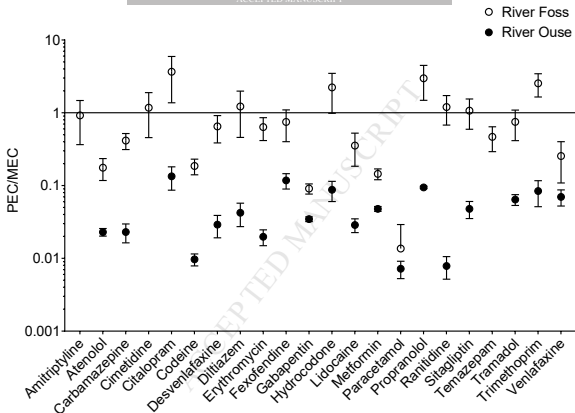


Figure 8. The annual average PEC/MEC ratios are plotted for the River Foss (open circles) and the River Ouse (closed circles). PECs were calculated for each river based on experimental WWTP removals and the average flow from sampling days. PEC/MEC ratios were calculated for site F2-F5 and O3-O6 and averaged, error bars represent the standard deviation.

Highlights

- 11 sites from two nested river systems sampled monthly for one year.
- Seasonal and spatial variation due to flow, usage and compound stability.
- Removal efficiency estimated for 24 pharmaceuticals in two WWTPs.
- Disagreement between measured concentrations and exposure model predictions.