



NEWS & VIEWS

Could Conservative Iron Chelation Lead to Neuroprotection in Amyotrophic Lateral Sclerosis?

Caroline Moreau,^{1,*} Véronique Danel,^{1,*} Jean Christophe Devedjian,² Guillaume Grolez,¹ Kelly Timmerman,² Charlotte Laloux,² Maud Petrault,² Flore Gouel,² Aurélie Jonneaux,² Mary Duthheil,² Cédric Lachaud,² Renaud Lopes,³ Grégory Kuchcinski,³ Florent Auger,⁴ Maeva Kyheng,⁵ Alain Duhamel,⁵ Thierry Pérez,⁶ Pierre François Pradat,^{7,8} Hélène Blasco,⁹ Charlotte Veyrat-Durebex,⁹ Philippe Corcia,⁹ Patrick Oeckl,¹⁰ Markus Otto,¹⁰ Luc Dupuis,¹¹ Guillaume Garçon,¹² Luc Defebvre,¹ Z. Ioav Cabantchik,¹³ James Duce,^{14,15} Régis Bordet,² and David Devos^{1,2}

Abstract

Iron accumulation has been observed in mouse models and in both sporadic and familial forms of amyotrophic lateral sclerosis (ALS). Iron chelation could reduce iron accumulation and the related excess of oxidative stress in the motor pathways. However, classical iron chelation would induce systemic iron depletion. We assess the safety and efficacy of conservative iron chelation (*i.e.*, chelation with low risk of iron depletion) in a murine preclinical model and pilot clinical trial. In *Sod1*^{G86R} mice, deferiprone increased the mean life span compared with placebo. The safety was good, without anemia after 12 months of deferiprone in the 23 ALS patients enrolled in the clinical trial. The decreases in the ALS Functional Rating Scale and the body mass index were significantly smaller for the first 3 months of deferiprone treatment (30 mg/kg/day) than for the first treatment-free period. Iron levels in the cervical spinal cord, medulla oblongata, and motor cortex (according to magnetic resonance imaging), as well as cerebrospinal fluid levels of oxidative stress and neurofilament light chains were lower after deferiprone treatment. Our observation leads to the hypothesis that moderate iron chelation regimen that avoids changes in systemic iron levels may constitute a novel therapeutic modality of neuroprotection for ALS. *Antioxid. Redox Signal.* 29, 742–748.

Keywords: amyotrophic lateral sclerosis, conservative iron chelator, oxidative stress, neuroprotection, treatment

¹Department of Neurology, ALS Center, Lille University, INSERM UMRS_1171, University Hospital Center, LICEND COEN Center, Lille, France.

Departments of ²Medical Pharmacology and ³Neuroradiology, Lille University, INSERM UMRS_1171, University Hospital Center, LICEND COEN Center, Lille, France.

⁴Department of Preclinical Radiology, Lille University, INSERM UMRS_1171, LICEND COEN Center, Lille, France.

⁵Department of Biostatistic, University of Lille, CHU Lille, EA 2694–Santé Publique: épidémiologie et qualité des soins, Lille, France.

⁶Department of Pneumology, Lille University, University Hospital Center, Lille, France.

⁷Laboratoire d'Imagerie Biomédicale, Sorbonne Universités, UPMC University Paris 06, CNRS, Inserm, Paris, France.

⁸Département de Neurologie, AP-HP, Hôpital Pitié-Salpêtrière, Paris, France.

⁹Laboratoire de Biochimie, Université François Rabelais, INSERM U930, CHRU, Tours, France.

¹⁰Department of Neurology, Center for Biomedical Research, Ulm University Hospital, Ulm, Germany.

¹¹INSERM UMR-S1118, Faculté de Médecine de, Strasbourg, France.

¹²EA4483 Department of Toxicology, CHU of Lille University, Lille, France.

¹³Della Pergola Chair, Alexander Silberman Institute of Life Sciences, Hebrew University, Jerusalem, Israel.

¹⁴Alzheimer's Research UK Cambridge Drug Discovery Institute, University of Cambridge, Cambridge Biomedical Campus, Cambridge, United Kingdom.

¹⁵The Florey Institute of Neuroscience and Mental Health, University of Melbourne, Parkville, Victoria, Australia.

*These authors contributed equally to this work.

Innovation

The present work provides the first clinical evidence about the neuroprotective potential of a therapeutically safe chelation treatment on early-stage amyotrophic lateral sclerosis (ALS) patients, which responded significantly to treatment in both brain iron deposits and indicators of disease progression. The novel treatment relied on oral administration of deferiprone that by chelation of labile iron it conferred on oxidation-stressed animals, improved motor functions, while essentially sparing systemic iron. The paradigmatic modality of chelation with deferiprone in ALS has prompted a multicenter study.

Introduction

AMYOTROPHIC LATERAL SCLEROSIS (ALS) is characterized by rapid progressive upper and lower motor neuron degeneration, leading to paralysis and death. Iron accumulation has been observed in mouse models and in both sporadic and familial forms of ALS (1, 3–6, 8, 9). Iron accumulation seems to occur at least in microglial cells within motor cortical regions (6) and has been observed in the motor cortex using magnetic resonance imaging (MRI) (1, 6). In patients with sporadic ALS, cerebrospinal fluid (CSF) levels of iron are elevated (4) and elevated serum ferritin levels correlate with shorter survival (9). Importantly, iron chelation has shown therapeutically relevant protective effects in animal models (3, 5). Deferiprone is a unique iron chelator; at low-dose levels, it can cross membranes, decrease regional iron accumulation, redeploy the captured iron to extracellular transferrin, and subsequently distribute iron throughout the body (thus avoiding anemia), defining the “conservative” iron chelation (2).

Dose and Sex Effect on Neuroprotection with Deferiprone in the Murine Model of ALS

A dose- and sex-dependent effect of deferiprone on survival was observed in the SOD, superoxide dismutase (SOD)1^{G86R} mouse model (Fig. 1). The female mice in the 50 mg/kg/day deferiprone groups survived for 13 days longer than those in the vehicle group (Fig. 1A). This corresponded to a 56% extension in survival from disease onset (defined as the peak in body weight) (Fig. 1B). The dose of 100 mg/kg/day was less effective and the dose of 200 mg/kg/day was not effective (not shown). A significant effect was observed in male mice only with the highest dose (200 mg/kg/day). Deferiprone improved the animals' physical examination, as shown by greater body weight (Fig. 1B), a lower peak in neurological impairment (*i.e.*, a lower Neuroscore, a quick phenotypic neurological scoring system) (Fig. 1C), and return of gene expression of *acetylcholine receptor subunit γ* (*Chrn3*) to wild-type levels (marker of denervation of muscles) (Fig. 1D). Importantly, treated mice had less iron accumulation in the spinal cord (shown by MRI T2* sequence) compared with vehicle control mice (Fig. 1E), demonstrating an ability of deferiprone to hit the biological target. As with treatment in other neurological models (2), deferiprone did not induce anemia, and

serum ferritin was only marginally below normal levels at the highest dose (Fig. 1F).

Safety Profile of Deferiprone in Early ALS Patients

Twenty-three consecutive sporadic patients were enrolled (22 limb onset and 1 bulbar onset) (Table 1). Four patients dropped out: one patient died after a fall, and 3 withdrew their consent. All were compliant to medication. The non-neurological physical examination remained unchanged. All patients displaying normal hematologic profiles, a slight elevation in urine iron levels, and a transient decrease in serum ferritin (in the normal ranges) was observed in the first 3 months of treatment.

Is There a Disease Modifying Effect of Deferiprone in Early ALS Patients?

The decrease in the ALS Functional Rating Scale (ALSFRS-R) score was significantly smaller for the first 3 months of deferiprone treatment than for the 3-month treatment-free period ($p=0.013$) (Table 1). Likewise, the decrease in the body mass index (BMI) was significantly different, with a decrease during the first 3 months and a small increase during the treatment period (no percutaneous endoscopic gastrostomy feeding) ($p=0.047$). Then, the BMI remained unchanged for 9 months. The reduction in manual muscle testing (MMT) scores was lower in patients on deferiprone than in matched patients from the Mitotarget study, although this difference did not reach statistical significance ($p=0.09$) (Table 2).

Iron Accumulation and Oxidative Stress Reduction Under Deferiprone in Early ALS Patients

A significant decrease in iron concentration (shown by a decrease in R2*) was observed in the cervical spinal cord, the medulla oblongata, and the motor cortex but not in areas outside the motor system (*i.e.*, the cerebellum and the occipital cortex) following treatment with deferiprone. Iron levels, oxidative stress markers, and neurofilament light chains were lower after deferiprone treatment in the CSF (Table 1).

Conservative Iron Chelation

The present study is a first to demonstrate the safety of conservative iron chelation in ALS. In both a murine model of familial ALS and sporadic ALS patients, low-dose deferiprone was associated with a decrease in pathologic iron accumulation in the central motor pathways but did not alter iron metabolism in other regions of the brain or in the periphery. This new therapeutic strategy appears to maintain the patient's overall aerobic metabolism and limit excess oxidative stress—as has been observed in Parkinson's disease (2).

Deferiprone significantly increased survival, as previously reported with other iron chelators (3, 5). This was observed despite treatment initiation at the symptomatic stage in the phenotypically aggressive SOD1^{G86R} model. An interaction between dose and sex was observed; disease severity, iron accumulation, and the required dose were higher in males than in females.

Deferiprone had a good safety profile in patients, with adverse events mostly relating to persistent ALS symptoms.

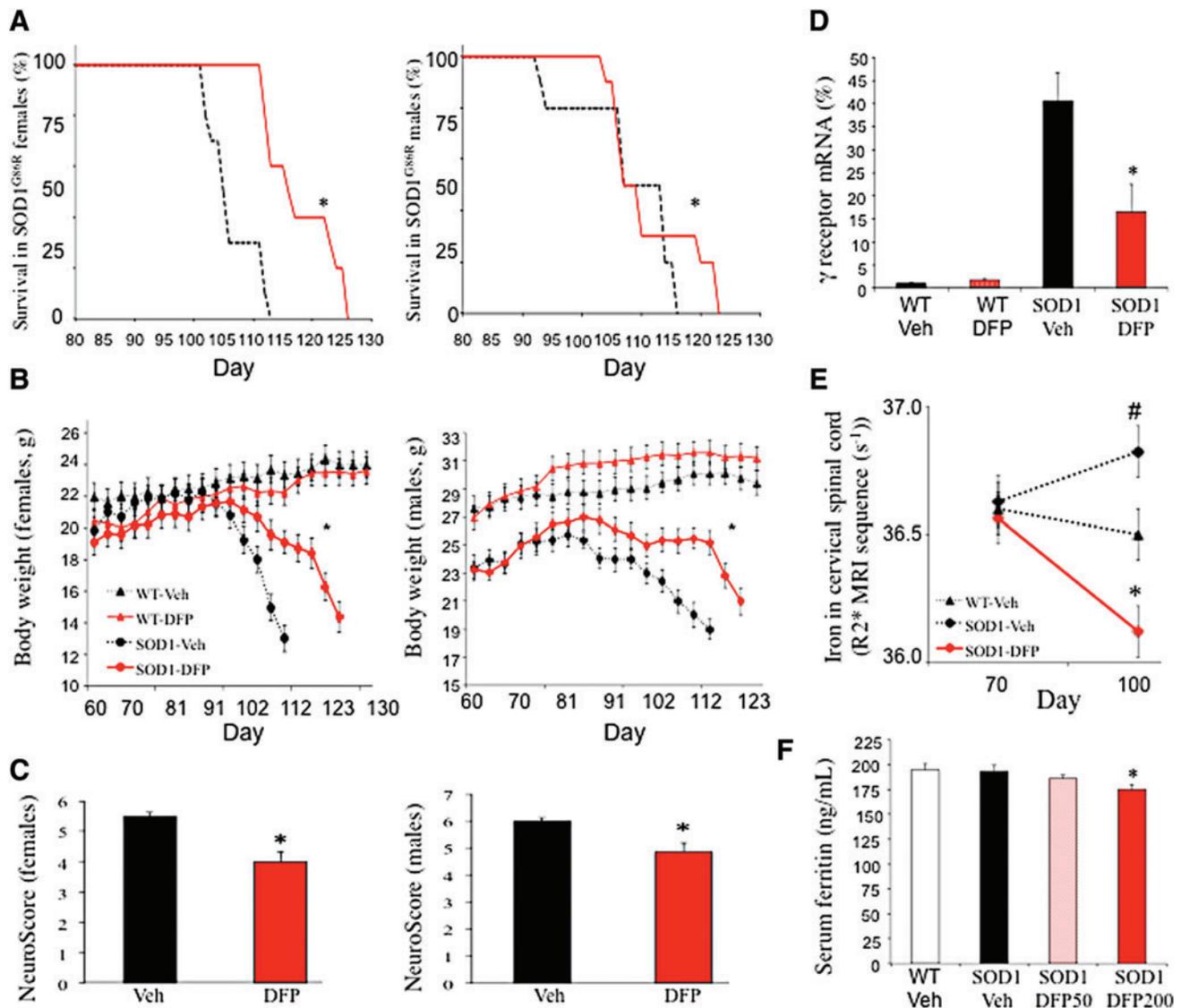


FIG. 1. Pharmacological effects of DFP in a murine model of ALS. Data are mean ± SEM in all experiments and doses, $n = 10$ per group for each sex and each dose. For the females, the best dose was 50 mg/kg/day, the dose of 100 mg/kg/day was less effective, and the dose of 200 mg/kg/day was not inefficient. For the males, the only efficient dose was 200 mg/kg/day. All the figures are presented with the dose of 50 mg/kg/day for the females and 200 mg/kg/day for the males. **(A)** The survival rates were significantly improved for female (in red) and male (in black) SOD1^{G86R} mice (from disease and treatment onset onward) in the DFP group (solid line) and the vehicle group (Veh, dotted line) (log-rank test; females: $p = 0.011$; males: $p = 0.03$). **(B)** Change in body weight in female and male SOD1^{G86R} mice from disease and treatment onset until death, in four groups: WT-Veh (black triangles with a dotted line), WT-DFP (red triangles with a solid line), SOD1^{G86R}-Veh (black circles with a dotted line), and SOD1^{G86R}-DFP (red circles with a solid line) (ANCOVA adjusted on baseline, $*p < 0.05$ vs. untreated SOD1^{G86R} mice). **(C)** The peak NeuroScore (a quick phenotypic neurological scoring system for evaluating disease progression in the mouse model of ALS: 0: no impairment; 6: greatest possible impairment) in the SOD1^{G86R}-Veh group (black bar) and the SOD1^{G86R}-DFP group (red bar) (Mann-Whitney test, $*p < 0.05$ vs. untreated SOD1^{G86R} mice). **(D)** As measured by qRT-PCR, acetylcholine receptor subunit γ receptor RNA expression as a percentage (%) of the control value in the left gastrocnemius muscle in SOD1^{G86R} female mice treated with 50 mg/kg/day DFP (Mann-Whitney test, $*p < 0.05$ vs. untreated SOD1^{G86R} mice). **(E)** Magnetic resonance imaging of the cervical spinal cord at day 70 (*i.e.*, just before the appearance of symptoms and treatment onset) and at day 100 (*i.e.*, with marked motor impairments before death) in untreated female SOD1^{G86R} mice (triangles with a black solid line), WT female mice (squares with a dotted line), and SOD1^{G86R} female mice treated with 50 mg/kg/day DFP (red line). $R2^*$ ($= 1/T2^*$) was obtained in a manually drawn region of interest using a monoexponential fitting of the signal decay with the echo time (Kruskal-Wallis test $*p < 0.05$ vs. untreated SOD1^{G86R} mice, $\#p < 0.05$ vs. WT untreated mice). **(F)** Serum ferritin levels (ng/mL) from serum obtained at end of treatment: not significantly reduced after 50 mg/kg/day and 100 mg/kg/day (data not shown), but significantly reduced after 200 mg/kg/day (Kruskal-Wallis test $*p < 0.05$ vs. untreated SOD1^{G86R} mice). ALS, amyotrophic lateral sclerosis; DFP, deferiprone; SOD, superoxide dismutase; WT, wild type. To see this illustration in color, the reader is referred to the web version of this article at www.liebertpub.com/ars

TABLE 1. NEUROLOGICAL AND SYSTEMIC PARAMETERS IN AMYOTROPHIC LATERAL SCLEROSIS PATIENTS AT INCLUSION AND WITH UP TO 12 MONTHS OF DEFERIPRONE TREATMENT

	Baseline	V3 3 months of no treatment	V6 3 months of DFP	p-value for V3 vs. V6	V9 6 months of DFP	V12 9 months of DFP	V15 12 months of DFPFP
← The SAFE-FAIR-ALS trial (months) →							
Population (<i>n</i> = 23)	Mean ± SD age: 56.2 ± 9.8 years; 4 female/13 male; mean ± SD disease duration (since first signs): 11.7 ± 5.7 months						
Drop out: 4 patients: 1 death related to brain hemorrhage after a fall and 3 withdrew their consent							
Adverse events during V3–V18 (<i>n</i> = 23)							
Falls with or without trauma (<i>n</i> = 9); respiratory failure with or without bronchitis (<i>n</i> = 4); arterial hypertension (<i>n</i> = 2); transient chest pain (<i>n</i> = 2); transient abdominal pain (<i>n</i> = 1); pulmonary embolism (<i>n</i> = 1); atrial fibrillation (<i>n</i> = 1); arterial thrombosis (<i>n</i> = 1); transient nausea (<i>n</i> = 1); transient postural tremor (<i>n</i> = 1); pruritus (<i>n</i> = 1); gastritis (<i>n</i> = 1)							
Handicap test (<i>n</i> = 19)							
ALSFRS-R	40.5 ± 4 41 [36–45]	35.6 ± 7 36 [31–42]	33.1 ± 8 33 [26–41]	0.013*	31.3 ± 9 31 [26–41]	29.6 ± 11 31 [20–40]	29 ± 11 35 [20–39]
BMI	26.3 ± 4 25 [24–28]	25.9 ± 4 26 [23–28]	26.0 ± 4 25 [24–28]	0.047**	26.4 ± 4 26 [24–28]	26.9 ± 3 27 [24–28]	25 ± 3 24 [24–27]
MMT	93.9 ± 7 92 [89–99]	88.0 ± 11 90 [81–95]	79.4 ± 13 88.0 ± 11	0.33**	77.0 ± 16 75 [68–88]	75.2 ± 17 77 [66–84]	72 ± 14 72 [66–79]
SVC	102.6 ± 18 102 [89–112]	95.3 ± 20 90 [79–110]	87.3 ± 25 90 [60–96]	0.1**	79.8 ± 23 83 [67–96]	78.8 ± 31 74 [57–96]	75 ± 35 76 [58–92]
Iron accumulation on MRI: R2* (s ⁻¹) (<i>n</i> = 19)							
Precentral-central cortex		14.9 ± 1.3	15.6 ± 1.4	14.3 ± 1.6	0.038** (V3–V6)	13.3 ± 2	
Medulla oblongata		16.2 ± 3	17.4 ± 2	15.5 ± 1	0.047** (V3–V6)	15.7 ± 2	
Cerebellum		16.9 ± 1	17.1 ± 1	16.9 ± 1	0.3** (V3–V6)	17.1 ± 1	
Cervical spinal cord		40.2 ± 5.6	42.2 ± 5.6	39.2 ± 4.8	0.002** (V3–V6)	39 ± 4.9	
Ferritin and iron in body fluids (<i>n</i> = 19)							
Ferritin		134 [100–175]	143 [89–193]	134 [72–170]	0.002 (V3–V6)	150 [99–206]	160 [101–220]
Ferritin (CSF)		—	12.3 [10–16]	—	0.027 (V3–V9)	9.7 [9–14]	
Iron (urine)		11.4 [4–19]	12.4 [8–12]	98.6 [32–99]	0.001 (V3–V6)	85 [42–122]	72 [21–99]
Oxidative stress marker in CSF (<i>n</i> = 19)							
8-OHdG		—	6 [5.4–6.7]	—	0.029 (V3–V9)	5.2 [5–6.5]	—
SOD-1 in CSF (<i>n</i> = 19)							
SOD		—	0.6 [0.5–0.7]	—	0.3 (V3–V9)	0.6 [0.5–0.7]	—
Neurofilaments in CSF (<i>n</i> = 19)							
NfL		—	5698 [3332–9599]	—	0.08 (V3–V9)	3492 [2807–6288]	—
pNfH		—	2578 [1338–3245]	—	0.3 (V3–V9)	2381 [1361–3255]	—

Data are mean ± standard deviation (first line) and median value [Quartile 1–Quartile 3] (second line). A revised ALSFRS-R score was measured as the primary outcome in the patient cohorts, whereas BMI, muscle strength (measured by MMT), and SVC were also evaluated as secondary outcomes (Student's *t* test (*) or Wilcoxon test (**), depending on the distribution assessed using histograms and the Shapiro–Wilk test).

Iron content was quantified by R2* transverse relaxation rates (= 1/T2*) measured using a 3-Tesla MRI system (analysis of variance for repeat measures; the statistically significant contrast analyses for V0 vs. V3, V3 vs. V6, and V6 vs. V9 are indicated).

Additional secondary biological outcomes were evaluated using previously characterized assays. Units observed in control patients were within the established ranges previously reported. Ferritin in CSF and serum in ng/mL. Iron in urines in µg/24 h. 8-oxdG: DNA adduct of 8-oxo-7,8-dihydro-2'-deoxyguanosine in ng/mL (Highly Sensitive 8-OHdG Check, Gentaur France SARL, Paris, France), SOD in U/mL (whole blood, as published elsewhere⁸); NfL: neurofilament light chain in pg/mL (Simoa/Digital ELISA platform) and pNfH: phosphorylated neurofilament heavy chain in pg/mL (BioVendor kit) (Student's *t*-test (*) or Wilcoxon test (**), depending on the distribution assessed using histograms and the Shapiro–Wilk test).

The bold values are statistically significant. ALS, amyotrophic lateral sclerosis; ALSFRS-R, ALS Functional Rating Scale; BMI, body mass index; CSF, cerebrospinal fluid; DFP, deferiprone; MMT, manual muscle testing; MRI, magnetic resonance imaging; SVC, slow vital capacity; SOD, superoxide dismutase.

TABLE 2. COMPARISONS OF 9 MONTHS OF DISEASE PROGRESSION IN PATIENTS ON DEFERIPRONE VERSUS A MATCHED POPULATION FROM THE MITOTARGET TRIAL

Parameters and populations	Deferiprone treatment	Three months of DFP treatment	Nine months of DFP treatment	p ^a
ALSFERS				
SAFE-FAIR, mean (SD)	35.6 (6.7)	33.1 (8.2)	30.6 (11.2)	0.50
Median (IQR)	34.0 (30.0–43.0)	33.0 (26.0–41.0)	(35.0) (20.0–41.0)	
Mitotarget, mean (SD)	35.4 (8.4)	32.8 (9.9)	30.6 (10.5)	
Median (IQR)	38.0 (30.0–42.0)	35.5 (26.5–41.0)	35.5 (23.0–38.0)	
BMI				
SAFE-FAIR, mean (SD)	26.0 (4.5)	26.1 (4.9)	27.4 (2.9)	0.32
Median (IQR)	25.7 (23.6–28.4)	25.5 (24.0–29.0)	27.4 (25.2–28.9)	
Mitotarget, mean (SD)	23.8 (3.2)	23.2 (3.5)	23.0 (3.6)	
Median (IQR)	23.2 (21.4–25.7)	21.9 (20.7–25.3)	22.5 (20.5–25.5)	
MMT				
SAFE-FAIR, mean (SD)	88.1 (11.8)	79.4 (13.3)	76.0 (17.9)	0.090
Median (IQR)	90.5 (80.0–96.0)	78.5 (73.0–90.0)	78.5 (68.5–85.5)	
Mitotarget, mean (SD)	118.8 (26.4)	112.8 (29.7)	97.5 (39.0)	
Median (IQR)	127.0 (110.0–139.0)	121.5 (86.5–135.0)	111.0 (56.0–122.0)	
SVC				
SAFE-FAIR, mean (SD)	96.3 (20.2)	89.4 (24.1)	82.9 (28.3)	0.97
Median (IQR)	91.6 (80.4–113.0)	90.4 (66.7–102.8)	80.7 (61.8–105.4)	
Mitotarget, mean (SD)	92.1 (20.2)	84.7 (23.7)	76.4 (36.4)	
Median (IQR)	92.5 (78.0–106.0)	88.0 (75.0–98.0)	75.4 (49.5–107.5)	

The revised ALSFRS-R, BMI, muscle strength (measured by MMT), and SVC were evaluated.

Model for ALSFRS:

$ALSFRS_{ik} = \beta_0 + \beta_1 t_{6ik} + \beta_2 t_{12ik} + \beta_3 Group_{ik} + \beta_4 Group_{ik} * t_{6ik} + \beta_5 Group_{ik} * t_{12ik} + \beta_6 ALSFRS_{baseline_i} + \gamma_{0i} + \gamma_{1k} + \epsilon_{ik}$. ik = for any subject i, for any bloc k, $\gamma_{0i} + \gamma_{1k}$ are random effects.

The bold value is statistically significant.

^ap-Value for the overall interaction between group and time, adjusted for the outcome level before DFP treatment (baseline).

We did not detect any of the adverse events occasionally observed in patients, with systemic iron overload, treated with 100 mg/kg/day deferiprone.

Encouragingly, for a safety trial with a small number of patients, deferiprone treatment was associated with slower disability progression and weight loss. However, the occurrence of a nocebo effect and then a placebo effect cannot be ruled out, and so, a large, multicenter, double-blind, placebo-controlled, randomized clinical trial is underway.

Materials and Methods

SOD1^{G86R} transgenic mice

All animal experiments were carried out in accordance with the “Principles of Laboratory Animal Care,” the current French and European Union legislative and regulatory framework (APAFIS#4269-2015112317225759), and the European ALS group’s preclinical trial guidelines of 2010. A dose/response study was performed in FVB-Tg(Sod1*G86R) M1Jwg/J mice (JAX Laboratories) with 50, 100, or 200 mg/kg p.o. deferiprone or vehicle twice a day (10 in each group). Study treatment was initiated at 75 days, that is, an age at which these mice are devoid of motor symptoms but already present with weight loss. The investigators were blinded to the study treatment. MRI of the cervical spinal cord was done using a 7-T MR system (BioSpec Bruker, Ettlingen, Germany), using a multiecho T2*-weighted sequence (number of echoes: 12; first echo time: 4 ms; echo spacing: 7 ms; repetition time: 1500 ms; slice thickness: 1 mm; field of view: 200 × 250 mm; matrix: 256 × 256; number of signal averages: 2).

Clinical trial

A single-center, single-arm, 12-month pilot clinical trial was performed to evaluate the effect of deferiprone on patients with ALS. The patients were followed for a 3-month treatment-free period and then treated for 12 months with the liquid formulation of deferiprone at a dose level of 30 mg/kg/day (morning and evening dose). The patients were recruited between December 2013 and January 2015, and all provided written informed consent. All patients had been taking riluzole. Treatment compliance (>80%) was assessed by questioning the participants and inspection of the dispensed packs of medication. The primary outcome criterion was disease progression, measured using the revised ALSFRS-R. The 3-month treatment-free period was compared with the first 3 months of treatment. The secondary outcomes included MMT, BMI, slow vital capacity, and CSF levels of markers for oxidative stress and neurofilaments. Physical examination was assessed every 3 months together with adverse event reports and reviewed anonymously by an independent safety monitoring board. Weekly blood counts were used to monitor the risk of neutropenia. For exploratory purposes, 19 patients treated with deferiprone for 9 months were compared with 19 matched individuals from among the 512 patients in the Mitotarget trial (negative results with olesoxime; NCT:00868166) (7).

Iron content was quantified by R2* transverse relaxation rates (=1/T2*) measured using a 3-Tesla MRI system (Achieva, Philips Medical Systems, Best, The Netherlands), using a 2D fast-field echo multiecho sequence (number of echoes: 15; first echo time: 3.6 ms; echo spacing: 3.3 ms;

repetition time: 1803 ms). Two stacks were subsequently acquired in the axial plane; 17 slices for each (slice thickness: 2 mm; isotropic, no gap, field of view: 230 × 190 mm; matrix: 116 × 95; number of signal averages: 2) to cover a volume between the floor of the fourth ventricle and the corpus callosum convexity. Images were processed using a T2* tool on an IDL virtual machine (V2, www.rsinc.com/IDL). A monoexponential signal decay with the echo time was obtained by voxel-by-voxel nonlinear least-squares fitting of the multiecho data [$S(t) = S_0 \cdot e^{-t \cdot R2^*}$; where t = echo time, S = measured signal, $R2^*$ = transverse relaxation rate]. Regions of interest were manually drawn on $R2^*$ maps by the same operator, who was blinded to the clinical data.

Statistical analysis

Differences in main outcomes between the treatment-free period (months 0–3) and the first three months of deferiprone treatment (months 3–6) were assessed with a paired T test (for normally distributed variables) or Wilcoxon signed-rank test.

Taking into account the differences in baseline characteristics between the patients on deferiprone and the Mitotarget population, we performed 1:1 matching on three prespecified factors: age (± 5 years), disease duration (± 2 months), and sex. Changes in the main outcomes between month 3 and 15 (9 months of treatment) in the paired groups were compared using linear mixed models with random coefficients. *Group*, *time*, the *group* × *time* interaction, and the baseline value were considered as fixed effects, with the participant and block matching considered as random effects. All statistical tests were two sided ($p < 0.05$), and all data were analyzed using SAS software (version 9.4, SAS Institute, Inc., Cary, NC).

Study approval

All clinical investigations were performed in accordance with the tenets of the Declaration of Helsinki. All patients provided written informed consent to participation. A local institutional review board approved the aims and procedures of the main study (national reference number: 2013-001228-21; ClinicalTrials.gov reference: NCT02164253) and a compassionate 12-month extension. The study and the article followed the consort statement.

Acknowledgments

The authors thank the Lille University Medical Center, the ARSLA charity (*Association pour la Recherche sur la Sclérose Latérale Amyotrophique et autres maladies du motoneurones*), and ApoPharma, Inc. for support. They also thank Michael Spino, Fernando Tricta, John Connelly, and C.F. for valuable advice on deferiprone and the Fédération de la Recherche Clinique du CHU de Lille (Lille, France, with Professor Dominique Deplanque, Pauline Guyon, Edouard Millois, Francine Niset, Marie Pleuvret, P.G., and Bertrand Accard) for support. The authors also thank Andreas Jeromin for the kits of neurofilament assaying, Delphine Taillieu and Yann Lepage from the EOPS1 rodent behavioral exploration facility (Federation of Neurosciences, University of Lille, Lille, France), Dr. P.M., Jean Francois Wiart, and Valerie Cuvier for technical support. They thank Dr. David Fraser (Biotech Communication, Damery, France) for editing the article.

Authors' Contributions

A: study concept and design; B: acquisition of data; C: analysis and interpretation; D: critical revision of the article for important intellectual content; E: study supervision. Caroline Moreau: A, B, C, D; Véronique Danel: A, B, C, D; G.G., K.T., M.P., C.L., F.G., A.J., M.D., C.L., R.L., C.V.-D., G.K., T.P., P.J., F.A.: B, C, D; M.K. and A.D.: C, D, and independent and blind statistical analyses; P.F.P., H.B., P.C., L.D., Guillaume Garçon, L.D., J.D., R.B.: A, C, D; M.O., P.Ö.: B, C, D; D.D.: A, C, D, E; the indicated authors take responsibility for data collection and analysis and the principal investigator (DD), who had full access to all the study data, takes full responsibility for submitting the final work for publication.

Author Disclosure Statement

C.M. has received grants from the France Parkinson charity. She has received various honoraria from pharmaceutical companies for consultancy and lectures on Parkinson's disease at symposia, including Aguetant, AbbVie, Medtronic, and Novartis. V.D., J.C.D., G.G., K.T., M.P., C.L., F.G., A.J., M.D., C.L., R.L., C.V.-D., G.K., F.A., A.D., M.K., H.B., T.P., P.J.-T., L.D., G.G., P.O., R.B., and J.D. have nothing to declare. P.C. served on the advisory board and received honoraria from Roche for consultancy and grants from the ARSLA charity. P.F.P. has received grants from the ARSLA charity, the *Association Française contre les Myopathies-Téléthon* (AFM-Téléthon), the *Institut pour la Recherche sur la Moelle épinière et l'Encéphale* (IRME), the T.L. Foundation, and the Target ALS Foundation. L.D. served on the Scientific Advisory Board for Novartis and Aguetant, and has received honoraria from pharmaceutical companies for consultancy and lectures. M.O. received grants from the European Union (FAIRPARK-II), German Ministry of Science and Technology (KNDD-FTLDC), T.L. Foundation, ALS Foundation, Foundation of the state Baden-Wuerttemberg, and the German Science Foundation. He has served as advisor for Axon, Biogen, and given lectures for Lilly, Fujirebio, and Teva. D.D. has received PHRC grants from the French Ministry of Health and research funding from the ARSLA charity, the France Parkinson charity, and the Credit Agricole Foundation. He has led two investigator-driven pilot studies involving deferiprone (FAIRPARK-I and SAFE-FAIR ALS-I), provided free of charge by ApoPharma. He has served on advisory boards, served as a consultant, and given lectures for pharmaceutical companies such as Orkyn, Aguetant, AbbVie, Medtronic, Novartis, Teva, UCB, Lundbeck, and ApoPharma.

References

- Adachi YI, Sato N, Saito Y, Kimura Y, Nakata Y, Ito K, Kamiya K, Matsuda H, Tsukamoto T, and Ogawa M. Usefulness of SWI for the detection of iron in the motor cortex in amyotrophic lateral sclerosis. *J Neuroimaging* 25: 443–451, 2015.
- Devos D, Moreau C, Devedjian JC, Kluza J, Petrault M, Laloux C, Jonneau X, Ryckewaert G, Garçon G, Rouaix N, Duhamel A, Jissendi P, Dujardin K, Auger F, Ravasi L, Hopes L, Grolez G, Firdaus W, Sablonnière B, Strubi-Vuillaume I, Zahr N, Destée A, Corvol JC, Pörtl D, Leist M, Rose C, Defebvre L, Marchetti P, Cabantchik ZI, and Bordet R.

- Targeting chelatable iron as a therapeutic modality in Parkinson's disease. *Antioxid Redox Signal* 21: 195–210, 2014.
3. Golko-Perez S, Amit T, Bar-Am O, Youdim MB, and Weinreb O. A novel iron chelator-radical scavenger ameliorates motor dysfunction and improves life span and mitochondrial biogenesis in SOD1^{G93A} ALS Mice. *Neurotox Res* 31: 230–244, 2017.
 4. Ignjatović A1, Stević Z, Lavrić D, Nikolić-Kokić A, Blagojević D, Spasić M, and Spasojević I. Inappropriately chelated iron in the cerebrospinal fluid of amyotrophic lateral sclerosis patients. *Amyotroph Lateral Scler* 13: 357–362, 2012.
 5. Jeong SY, Rathore KI, Schulz K, Ponka P, Arosio P, and David S. Dysregulation of iron homeostasis in the CNS contributes to disease progression in a mouse model of amyotrophic lateral sclerosis. *J Neurosci* 29: 610–619, 2009.
 6. Kwan JY, Jeong SY, Van Gelderen P, Deng HX, Quezado MM, Danielian LE, Butman JA, Chen L, Bayat E, Russell J, Siddique T, Duyn JH, Rouault TA, and Floeter MK. Iron accumulation in deep cortical layers accounts for MRI signal abnormalities in ALS: correlating 7 tesla MRI and pathology. *PLoS One* 7: e35241, 2012.
 7. Lenglet T, Lacomblez L, Abitbol JL, Ludolph A, Mora JS, Robberecht W, Shaw PJ, Pruss RM, Cuvier V, and Meininger V; Mitotarget Study Group. A phase II-III trial of olesoxime in subjects with amyotrophic lateral sclerosis. *Eur J Neurol* 21: 529–536, 2014.
 8. Lu A, Rajanala M, Mikkilineni S, Cahill S, Brown C, Berry RD, and Rogers J. The 5'-untranslated region of the C9orf72 mRNA exhibits a phylogenetic alignment to the cis-aconitase iron-responsive element; novel therapies for amyotrophic lateral sclerosis. *Neurosci Med* 7: 15–26, 2016.
 9. Veyrat-Durebex C, Corcia P, Mucha A, Benzimra S, Mallet C, Gendrot C, Moreau C, Devos D, Piver E, Pagès JC, Maillot F, Andres CR, Vourc'h P, and Blasco H. Iron metabolism disturbance in a French cohort of ALS patients. *Biomed Res Int* 485723, 2014.

Address correspondence to:

Dr. David Devos
 Département de Pharmacologie Médicale
 Université Lille INSERM 1171
 CHU de Lille
 Lille F-59037
 France

E-mail: david.devos@chru-lille.fr

Date of first submission to ARS Central, December 24, 2017;
 date of acceptance, December 24, 2017.

Abbreviations Used

ALS = amyotrophic lateral sclerosis
 ALSFRS-R = ALS Functional Rating Scale
 BMI = body mass index
 CSF = cerebrospinal fluid
 DFP = deferiprone
 MMT = manual muscle testing
 MRI = magnetic resonance imaging
 SCV = slow vital capacity
 SOD = superoxide dismutase