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**Article:**
Firdaus, Mohd Anuar Mohd, Agatz, Annika orcid.org/0000-0003-3228-8822, Hodson, Mark E orcid.org/0000-0002-8166-1526 et al. (2 more authors) (2018) Fate, uptake, and distribution of nanoencapsulated pesticides in soil–earthworm systems and implications for environmental risk assessment. Environmental Toxicology and Chemistry. pp. 1420-1429. ISSN: 1552-8618

https://doi.org/10.1002/etc.4094

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# Fate, Uptake and Distribution of Nanoencapsulated Pesticides in Soil-Earthworm Systems and Implications for Environmental Risk Assessment

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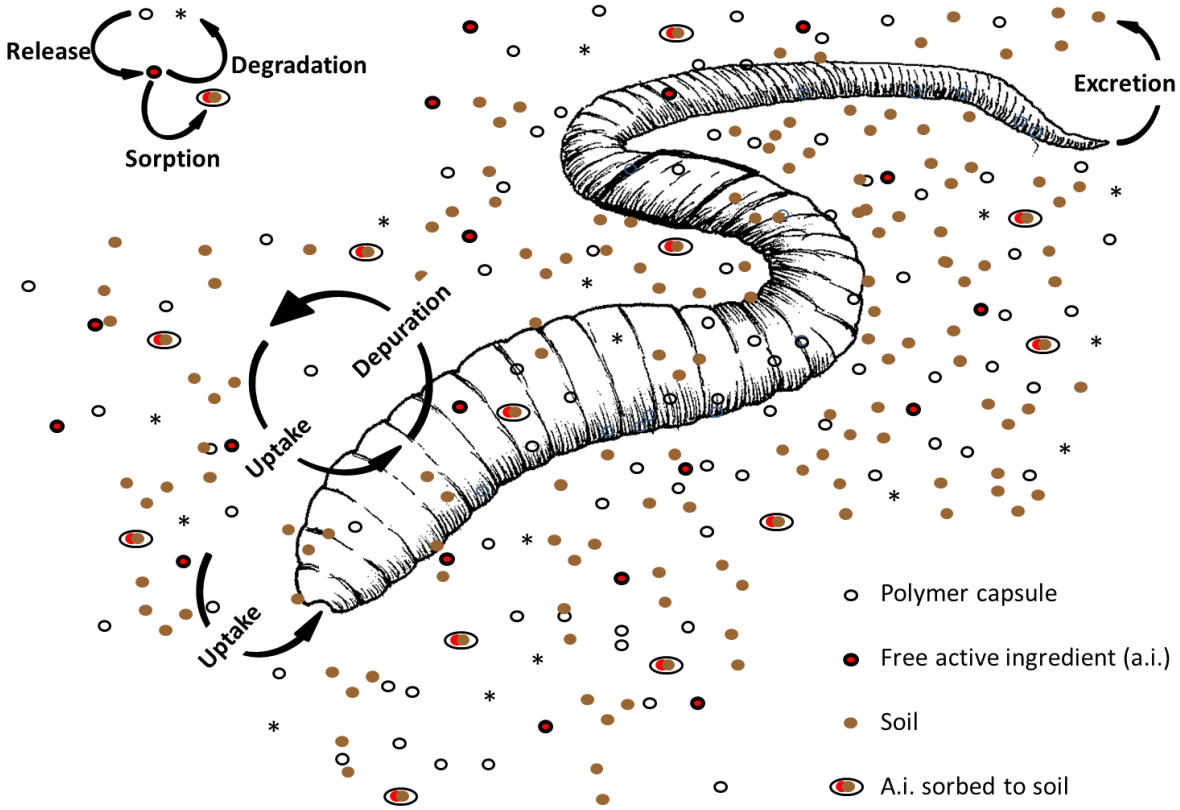
## Abstract

Nanopesticides are novel plant protection products offering numerous benefits. As nanoparticles behave differently from dissolved chemicals, environmental risks of these materials could differ from conventional pesticides. Here we used soil-earthworm systems to compare the fate and uptake of analytical grade bifenthrin to that of bifenthrin in traditional and nano-encapsulated formulations. Apparent sorption coefficients for bifenthrin in the nano-treatments were up to 3.8 times lower than in the non-nano treatments whereas dissipation half-lives of the nano-treatments were up to two time longer. Earthworms in the nano-treatments accumulated around 50% more bifenthrin than those in the non-nano treatments. In the non-nano treatments, most of the accumulated material was found in the earthworm tissue while in the nano-treatments, the majority resided in the gut. Evaluation of toxicokinetic modelling approaches showed that models incorporating the release rate of bifenthrin from the nanocapsule and distribution within the earthworm provided the best estimations of uptake from the nanoformulations. Overall, our findings indicate that the risks of nanopesticides may

be different from conventional formulations. The modelling presented here provides a starting point for assessing risks of these materials but needs to be further developed to better consider the behaviour of the nanoencapsulated pesticide within the gut system.

Keywords: Nanopesticides; Synthetic pyrethroids; Nanoencapsulation; Earthworms; Toxicokinetic modelling, *Eisenia fetida*, *Lumbricus terrestris*

32 Graphical abstract

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## Introduction

Recently, novel pesticide products have been developed that employ nanotechnology (Kah et al., 2013; Kah, 2015). These so called 'nanopesticides' comprise either nanoparticulate forms of a pesticide active ingredient or nanocapsules containing an active ingredient (a.i.). Nanopesticides offer a range of advantages over conventional pesticides in that they may increase efficacy of the a.i. and/or enhance the environmental and human health safety profiles of the products (Kah et al., 2013; Kookana et al., 2014). However, there is recognition that the application of nanotechnology could also have negative and unanticipated impacts on the environment so it is also possible that nanopesticides could pose a greater risk than equivalent conventional pesticide products. The applicability of existing environmental risk assessment approaches for pesticides to nanoformulations has also been questioned (Kah, 2015).

One group of organisms that will be exposed to nanopesticides are terrestrial invertebrates such as earthworms. Earthworms are known to bio-magnify inorganic and organic soil contaminants, including pesticides, polycyclic aromatic hydrocarbons, brominated flame retardants, and metals (Heikens et al., 2001; Matscheko et al., 2002; Langdon et al., 2005). Earthworms being at the base of a food chain hold an integral position. Uptake and accumulation of contaminants into earthworms not only poses a risk to the earthworm directly, but bioaccumulation and contaminant transfer through the food chain to top predators such as birds has the potential to result in secondary poisoning (Spurgeon and Hopkin, 1996).

Data for other non-pesticide nanoparticles shows that these materials can be taken up by earthworms (Kwak and Youn-Joo, 2005). Investigations determining distribution of nanoparticles show that highest concentrations of accumulated materials are associated with the earthworm gut (Unrine et al., 2010; Waissi-Leinonen et al. 2012). Adverse effects have also been reported in earthworms following exposure to carbon-based and metal and metal oxide nanoparticles (Kwak and Youn-Joo, 2005; Scott-Fordsmand et al. 2008).

To date, the focus of research into bioconcentration and impacts of nanoparticles on earthworms has been on metals and metal oxides (Kwak and Youn-Joo, 2005), carbon nanotubes (Petersen et al., 2008, Petersen et al., 2011, Scott-Fordsmand et al. 2008) and fullerenes (Li et al. 2010, Kelsey and White 2013). To the best of our knowledge, no-one has explored the uptake of nanopesticides, even though it is inevitable that earthworms will be exposed to these products during use. Therefore, here we investigate the effects of nanoencapsulation on the fate, uptake, depuration and distribution of a pesticide a.i. in soil-earthworm systems. The nanoencapsulated materials used in the study were developed by Vive Crop Protection Inc and comprise bifenthrin encapsulated in a polymer nanoparticle with the aim to better target the active ingredient to the pest species. We compare the fate, uptake and distribution of the analytical grade a.i. with that of conventional and nanoformulated products for the two earthworm species *Eisenia fetida* and *Lumbricus terrestris*. The findings are used to explore the suitability of existing and novel toxicokinetic models to better characterise the environmental risks of nanoencapsulated substances in the future.

## Materials and methods

### *Chemicals, soils and organisms*

Analytical PESTANAL® grade bifenthrin was purchased from Sigma-Aldrich (Dorset, UK), formulated bifenthrin (Capture LFR) was obtained from FMC Corporation (Philadelphia, USA). Two nanoencapsulated formulations of bifenthrin (Nano A and B) were obtained from Vive Crop Protection Inc. (Toronto, Canada). The precise make-up of these materials is proprietary but both formulations employ an acrylate copolymer to encapsulate the bifenthrin but contain different co-formulants. Acetonitrile (99.9%) was purchased from Fisher Scientific (Loughborough, UK). Details of the bifenthrin treatments are provided in the Supporting Information.

A sandy loam soil was obtained from Landlook (Midlands, UK). Prior to use, the soil was air dried, sieved to ≤ 2 mm to ensure homogeneity within the soil matrix and stored at room temperature. Characteristics of the study soil are provided in the Supporting Information.

89 *Eisenia fetida* and *Lumbricus terrestris* were obtained from Blades Biological Ltd. (Kent, UK).
90 The earthworms were cultured in a medium of horse manure and peat (50:50) for *E. fetida*,
91 and in moist soil for *L. terrestris*. They were kept moist with deionized water under laboratory
92 conditions (20 ± 3 °C). The horse manure used in this culture was collected from horses that
93 were not under medication to avoid any toxic effects on the earthworms. *E. fetida* were fed
94 twice weekly with homogenized mashed potato powder which was added to the surface of the
95 culture and *L. terrestris* were fed with dead birch leaves distributed on the surface of the moist
96 soil.

97 *Uptake and depuration studies*

98 Uptake and depuration experiments followed OECD Guideline 317 'Bioaccumulation in
99 Terrestrial Oligochaetes' and used only *E. fetida* (OECD, 2010). Experiments were performed
100 in glass jars at a concentration of 10 µg/g of active ingredient where each jar contained 50 ±
101 1 g of test soil and kept in an incubator at 20 ± 2 °C, using a 16:8 light/dark cycle. Assuming
102 a mixing depth of 20 cm, concentrations expected in the environment from the use of Capture
103 would be expected to range from 35 - 100 µg $g^{-1}$. The test concentration was one order of
104 magnitude lower than the concentration we used previously to assess the toxicity of the
105 different bifenthrin treatments to *E. fetida*. At 100 µg $g^{-1}$, no mortality, a slight increase in
106 growth and a small decrease in cocoon production were observed (Anuar, unpublished data).
107 Before the earthworms were exposed to the different treatments, they were acclimated to the
108 experimental conditions in the incubator for 48 h using non-treated soil. The different bifenthrin
109 treatments were then mixed with the soil using deionized water as solvent carrier to achieve
110 a moisture content between 60-70% of the maximum water holding capacity (MWHC). Treated
111 soil was left for 24 h before adding the earthworms.

112 For each bifenthrin treatment (analytical grade, conventional and two nanoformulations), 45
113 glass jars of treated soil were prepared. At the start of the uptake phase, one mature adult *E.*
114 *fetida* with a visible clitellum was added to each glass jar. Glass jars were then covered with
115 garden fleece (to prevent earthworms from escaping while allowing sufficient air supply to be

maintained) attached with an elastic band. The uptake phase of the experiment lasted for up to 21 d with triplicate samples being taken at 0 and 6 h and 1, 3, 7, 10, 14, 21 d. *E. fetida* in the remaining glass jars were then transferred to clean soil for up to another 21 d of depuration with samples being taken at 6 h and 1, 3, 7, 10, 14, 21 d after transfer. At each time point in both phases, the earthworm weight and mortality were recorded. Soil moisture content in each glass jars was monitored throughout both phases, and adjusted, where necessary, by adding deionized water so that it remained between 60-70% of the MWHC. The pH of the soils was measured at the beginning and end of the uptake phase and at the end of the depuration phase. Earthworms were fed weekly with mashed potato powder.

Once samples were collected, earthworms were removed, rinsed with deionized water, blotted dry, weighed and then placed for 48 h on moist filter papers to allow the earthworms to purge their gut contents (Dalby et al., 1996). The moist filter papers were changed twice a day (in the morning and evening). The earthworms were then frozen prior to analysis. Soil samples were taken for chemical analysis and to extract soil pore water.

*Distribution of bifenthrin in earthworms*

The distribution of bifenthrin following exposure to the different treatments was assessed using both *E. fetida* and *L. terrestris*. Experiments were performed at the same concentration and conditions as used in the uptake and depuration studies. *E. fetida*, were exposed to 50 ± 1 g of soil treated with each treatment or soil only while *L. terrestis* were exposed to 350 ± 5 g of treated soil or soil only. The duration of the uptake phase was 10 d while the depuration phase lasted for 7 d. There were six replicates per treatment and sampling point. Soil, faeces and earthworm samples were taken at the end of each phase for analysis.

The removed earthworms were placed on a dissecting tray with their dorsal side facing upwards. Using a pair of dissecting scissors, an opening cut was made below the clitellum. A straight line cut was made from the opening cut down to the posterior. The cut was made carefully and not too deeply to avoid damage to the internal organs. The skin was pulled apart using forceps and pinned back using dissecting pins. The earthworms were then separated

into skin, gut and other tissue (hereafter referred to as 'tissue') for *L. terrestris*. Separation of *E. fetida* tissues proved challenging so it was only possible to separate these samples into gut + tissue and skin. Prior to analysis, samples were washed with distilled water and centrifuged at 3000 rpm for 15 min. Samples and washing water were analysed separately for bifenthrin residues.

*Sample extraction and HPLC analysis*

Soil (5 ± 0.5 g) was extracted by adding 15 mL acetonitrile and then shaking the mixture on an orbital shaker (250 oscillations $min^{-1}$) at room temperature (20 ± 2 °C) for 2 h. Samples were then allowed to settle and 2 mL aliquots of supernatant were taken for analysis. Soil pore water was obtained by placing 10 ± 1 g of soil into a glass syringe with a layer of 3 cm of glass wool inserted into the bottom. The syringe was inserted into a glass centrifuge tube and centrifuged for 20 min at 2016 g to separate soil and soil pore water. The difference in density of the polymer capsules and water is small so centrifugation would not be expected to affect the recovery of the bifenthrin from the nano-treatments compared to the non nano-treatments (Kah et al., 2016).

Earthworm samples were homogenized for 5 minutes using a LabGen Series 7 homogenizer with 5 mL of acetonitrile. The suspension was transferred, with rinsing using an additional 5 ml of acetonitrile, to a glass vial. The extracts were centrifuged for 20 min at 2016 g. The samples were then filtered using 0.45 µm nylon filters and a 2 mL aliquot of the supernatant was taken for further analysis.

Soil and earthworm extracts and pore water were analysed using High-performance Liquid Chromatography (HPLC; Perkin Elmer, Flexar) coupled with photodiode array detection. More detail on the methods used are provided in the Supporting Information. The limits of detection and quantification were 1.2 and 3.7 ng $mL^{-1}$ for the analytical grade, 1.5 and 4.7 ng $mL^{-1}$ for the conventional formulation, 1.9 and 5.9 ng $mL^{-1}$ for nano A and 2.1 and 6.5 ng $mL^{-1}$ for nano B. Recoveries for analytical method ranged from 90-107% for water, 88-103% for soil, 84-

169 102% for *E. fetida* and 90-107% for *L. terrestris* and recoveries of the filtration method ranged
170 from 87-100% (see supporting information).

171 *Data analysis*

172 Determination of sorption coefficient, $k_d$

173 Sorption coefficient, $k_d$ values were calculated at each time point (Equation 1) where: $C_{water}$
174 and $C_{soil}$ are the concentrations of bifenthrin in soil pore water (µg $mL^{-1}$) and soil (µg $g^{-1}$),
175 MWHC is the maximum water holding capacity of the soil (%), and %water is the moisture
176 content of the soil (%). Averages of $k_d$-values were then determined.

177 $$k_d = \frac{C_{soil}}{C_{water} * (\frac{\%water}{MWHC})} - 1 \quad (1)$$

178 Kinetic modelling

179 We wanted to evaluate whether data on the uptake and depuration characteristics could be
180 used to inform the uptake and depuration of bifenthrin resulting from exposure to Capture LFR
181 and the two nano formulations. Three models were explored with increasing complexity. Data
182 from the analytical grade bifenthrin treatment was always used to parameterise the models.

183 Model 1 was the first order one compartment toxicokinetic model outlined by Ashauer *et al.*
184 (2010) (Equation 2).

185 $$\frac{\mathrm{d}C\mathrm{organism}}{\mathrm{d}t} = k\mathrm{in} * C\mathrm{water}\,(t) - k\mathrm{out} * C\mathrm{organism}\,(t) \quad (2)$$

186 Where: $C_{organism}$ is the internal concentration (µg $g^{-1}$); $C_{water}$ is the concentration in the pore
187 water (µg $mL^{-1}$); and $k_{in}$ and $k_{out}$ are the uptake rate constant (mL $g^{-1}$ $h^{-1}$) and the depuration
188 rate constant ($h^{-1}$), respectively.

189 Model 2 was designed for estimating uptake of an active ingredient from a nonencapsulated
190 treatment. This model is an adaptation of Model 1 modified to account for the release of
191 bifenthrin from the polymer capsules into the soil pore water (Equation 3).

192 $$\frac{\mathrm{d}C\mathrm{organism}}{\mathrm{d}t} = k\mathrm{in} * C\mathrm{water}_2\,(t) - k\mathrm{out} * C\mathrm{organism}\,(t) \quad (3.1)$$

with

$$\frac{dC\text{water}_2}{dt} = (C\text{water (t)} - C\text{water}_2\ (t)) * k\text{r} \quad (3.2)$$

Where: $C\text{water}_2$ is the concentration of the compound in the pore water released from the nanoformulation (µg $g^{-1}$) and $k_r$ is the release rate of the nanoformulation ($h^{-1}$). The release rate can be calculated by comparing the degradation rate of the bifenthrin a.i. with the degradation rate of bifenthrin in the nanoencapsulated formulation (Kah et al., 2016). A full description of the approach for estimating release rate is provided in Kah et al. (2016).

Model 3 was used for the distribution studies. This model extends either Model 1 or 2 to account for the distribution of the compound in gut, skin and tissue (Equations 4).

$$C\text{skin} = C\text{organism}\ (t) * a \quad (4.1)$$

$$C\text{tissue} = C\text{organism}\ (t) * b \quad (4.2)$$

$$C\text{gut} = C\text{organism}\ (t) - C\text{skin} - C\text{tissue} \quad (4.3)$$

Where: *C*skin, *C*gut and Ctissue are the concentration of the compound in skin, gut and tissue (µg $g^{-1}$), and a and b are distribution coefficients between the total internal concentration and the skin, the total internal concentration and the tissue. The distribution coefficients are obtained using studies on analytical grade a.i..

Statistical analysis

Statistical analysis was performed using SigmaPlot (Version 13.0; Systat Software, San Jose, CA). Data were tested performing one-way - or two-way- Analysis of Variance (ANOVA) via the Holm-Sidak pairwise comparison method with the Shapiro-Wilk test for normality of data and the Brown-Forsythe test for equal variance of data. Modelling was conducted in OpenModel V 2.4.2. (http://openmodel.info/) using the Runge-Kulta (4th Order) ordinary differential equation method with Monte Carlo simulations to obtain the 95% confidence interval and the Nash–Sutcliffe Efficiency calculation for goodness of fit indication. Nash–

Sutcliffe values (hereafter called Nash index) between 0 and 1 represent an acceptable fit of the model to the data.

## Results and discussion

Fate of bifenthrin in soil

Generally, throughout the uptake phase, there was a decrease in concentration of bifenthrin in the soil and soil pore water which was associated with an increase in the concentration of bifenthrin in the earthworms (Data are summarised in the Supporting information). At the end of the uptake phase, 65-82% of bifenthrin was extractable and associated with the soil particles, 16-33% had dissipated/degraded, around 1% was present in the pore water and < 1% was taken up by the earthworms (Figure 1 in the Supporting Information). Apparent half-lives for bifenthrin in the different treatments increased in the order analytical grade bifenthrin – Capture LFR – Nano B – Nano A. The observed $DT_{50}$ for the analytical grade bifenthrin is at the lower end of the values reported in the field for bifenthrin and is lower than reported in laboratory studies (Pesticide Properties Database, 2017). Half-lives are also lower than those obtained by Kah *et al*. (2016) in similar investigations into the differences in persistence of analytical grade bifenthrin and the a.i. in Capture LFR and nanoencapsulated treatments, although the order of half-lives is the same (Kah et al., 2016). Release rates and associated release times ($RT_{50}$) for the nanoformulations (Table 1) are lower than those found by Kah *et al*. (2016). Overall, these results indicate that even traditional formulations can affect the persistence of an active ingredient but this impact is more enhanced in the nano-encapsulated treatments, possibly due to the nanocapsules 'shielding' the unreleased a.i. from the degrading microbes.

Sorption coefficients ($k_d$), based on the soil and soil-pore water concentrations ranged from 154 to 585 L $kg^{-1}$ and increased significantly in the order Nano A = Nano B < Capture LFR < analytical grade bifenthrin (Table 1; Two-Way ANOVA Holm-Sidak method $P < 0.001$). Sorption coefficients are lower than previously reported for the a.i. which range from 882 to 6000 mL $g^{-1}$ in different soil types (Pesticides Property Database, 2017). Sorption coefficients

244 are also lower than those observed by Kah et al. for bifenthrin a.i. and bifenthrin in traditional
245 and nanoencapsulated formulations (Kah et al., 2016). The mismatch is possibly explained by
246 the fact that we derived $k_d$ values based on pore water measurements, which is arguably more
247 realistic than the batch equilibrium approach employed in previous studies. The differences
248 might be explained by dissolved organic carbon in the pore water which may act as an
249 additional sink for the bifenthrin or due to differences in the nature of the organic carbon in the
250 soils used in the different studies.

251 The observations for Capture LFR demonstrate that even traditional co-formulants can affect
252 the distribution of the bifenthrin in soils although the effect is more enhanced in the nano-
253 encapsulated treatments. Other studies have explored the effects of formulation on pesticide
254 behaviour. In studies with chlorsulfuron, co-formulants reduced sorption (Foldenyi et al., 2013)
255 while studies with propyzamide (Khan and Brown, 2017) showed sorption to increase and
256 studies with triticonazole, cyprodinil, propetamphhos and fludioxinil showed sorption to
257 increase (Beigel et al., 1998; Beigel and Barriuso, 2000; Garcia-Ortega et al., 2006; Pose-
258 Juan et al., 2011). The impacts of co-formulants therefore likely depend on the active
259 ingredient and the nature of the co-formulants used in a product. The observed reduction in
260 the $k_d$ values for the nano-encapsulated materials is likely due to a combination of the co-
261 formulant effects and the fact that the polymer capsule shielded the bifenthrin from sorption
262 sites on the soil surface.

263 Uptake and depuration behaviour

264 *Uptake and depuration in* E. fetida

265 No mortality was recorded and the studies passed the validity criteria (based on earthworm
266 growth and mortality) according to the principles outlined in the OECD 317 (OECD, 2010).
267 Lower uptake and depuration was seen for bifenthrin in the analytical and Capture LFR
268 treatments compared to the two nanoformulation treatments (Figure 1). At the end of the
269 depuration phase, in the analytical grade and Capture LFR treatments, bifenthrin was still
270 detectable in the earthworms whereas for the two nanoformulation treatments, it was not

detectable. The pattern of uptake and depuration between the non-nano and nano treatments was also different. The non-nano exposures were characterised by a steady uptake and elimination of bifenthrin over time whereas in the nano treatments, an initial rapid period of uptake or elimination was observed and this then tailed off (Figure 1).

The first order one-compartment model (Model 1) was successfully fitted to the data from the analytical grade bifenthrin treatment (Nash index = 0.94) obtaining an uptake- and depuration - rate constant of 0.222 ± 0.009 mL $g^{-1}$ $h^{-1}$ and 0.0036 ± 0.0002 $h^{-1}$. Use of the uptake and depuration rates in the model to simulate the uptake and depuration bifenthrin from the Capture LFR formulation worked well (Nash index = 0.68) but failed to acceptably simulate the uptake of Nano A and Nano B (Nash index < -0.01) (Figure 1). The results for the Capture modelling do, however, suggest that it may be possible to extrapolate from studies into the uptake of analytical grade materials to estimate uptake of a.i.'s from traditional formulations.

Model 2, which incorporates the release rate of bifenthrin from the nanocapsule, underestimated uptake and depuration of the bifenthrin from the two nano treatments (Nash index < 0; Figure 1). Closer inspection of the simulation however revealed that this model more accurately simulated the internal concentration at the end of the depuration phase. The differences in kinetic patterns and model fits suggests that the nanoencapsulated bifenthrin was accumulated *via* a different mechanism than in the analytical grade material and Capture LFR treatments. As previous studies with earthworms have shown that other nanoparticles accumulate in the earthworm gut rather than the actual tissue (Unrine et al., 2010; Waissi-Leinonen 2012), we performed studies to explore whether there were any differences in the distribution of the bifenthrin in earthworms exposed to the different treatments. Here, we not only used *E. fetida* but also *L. terrestris* due to its larger size and consequent ease of handling.

*Distribution studies*

Concentrations of bifenthrin in *L. terrestis* were significantly greater than in *E. fetida* for all treatments ($P < 0.001$). Within each species, uptake was significantly different between the nano formulated and non-nano treatments, while within the non-nano and nano treatments no

298 significant difference in uptake was observed (P<0.001). These differences in uptake might
299 be explained by differences in the way the two earthworm species process soil organic matter,
300 their ecological strategy and/or lipid content (Kelsey et al., 2005). *Eisenia fetida* is smaller than
301 *L. terrestris* and is an epigeic species living primarily at or near the soil surface and consumes
302 coarse particulate organic matter and surface litter. *L. terrestris* is an anecic species that lives
303 in deep burrows and comes to surface to feed on surface litter [Bouche, 1983]. Interestingly,
304 the interspecies difference in uptake that we see is the opposite to that observed in a similar
305 study using pharmaceuticals covering a range of physico-chemical properties (Carter et al.,
306 2016).

307 Significant differences were also seen in the depuration of bifenthrin by the two species (P <
308 0.001). For the analytical grade and Capture LFR treatments, *L. terrestris* still contained 57 -
309 59% of the accumulated bifenthrin after the 7 d depuration phase while concentrations in
310 *E.fetida* were significantly lower (43-47%; P < 0.001). Depuration of bifenthrin from the two
311 nano treatments was faster with *L. terrestris* containing 20-22% of accumulated bifenthrin after
312 7 d depuration while *E. fetida* contained only 10-13% of the accumulated mass. Bifenthrin from
313 the nanoformulations was eliminated 2.8 and 4 times more quickly than from the non-nano
314 treatments (P < 0.001).

315 In *L. terrestris*, following the uptake phase, concentrations of bifenthrin from the analytical
316 grade and Capture LFR treatments was significantly higher (P < 0.01) in the tissue compared
317 to the gut and skin which had similar (P = 0.6) bifenthrin concentrations (Figure 2). In contrast,
318 for the nanoformulation treatments significantly (P < 0.001) higher concentrations were
319 observed in the gut of the earthworms compared to the skin and tissue. Concentrations in skin
320 and tissue were also significantly different (P < 0.001). The concentration in the gut of the
321 nano- exposed animals was significantly higher (P < 0.001) than the non-nano exposed
322 earthworms even though the concentration of bifenthrin in the soil was the same. A significant
323 difference was also observed between the two nanoformulations (P = 0.019). This might
324 indicate that the earthworms are selectively consuming the polymer capsules and/or that the

capsules are becoming 'trapped' in the gut of the animal and are eliminated more slowly than the bulk soil. This seems to be nanoformulation specific. While it is not known whether earthworms are able to select finer material from coarser particles, there is a body of evidence indicating that they do exhibit preferences for different food types (Curry and Schmidt, 2007).

At the end of the 7 d depuration phase, for the analytical grade and Capture LFR treatments, highest concentrations of bifenthrin in *L. terrestris* were seen in the tissue while for the nanoformulation treatments highest concentrations were seen in the gut (Figure 2). The observation that nanopesticide treatments result in highest bifenthrin concentrations in the gut are similar to findings from a previous study into the uptake and distribution of C60 and Au nanoparticles into earthworms (Unrine et al., 2010; Waissi-Leinonen 2012; Petersen et al., 2008, 2011). We found that gut associated bifenthrin was generally less eliminated *via* the gut for the non-nanoformulation treatments compared to the nanoformulated treatments (Figure 3). Furthermore, a temporal shift in elimination of the nanoformulated bifenthrin occurred. Gut associated elimination of bifenthrin was greatest for the non-nanoformulation treatments whilst the earthworms were still in soil during the elimination phase of the experiment whilst elimination for the nanoformulated treatments was greatest when the organisms were on the filter paper after the elimination phase of the experiment.

Unfortunately, it was not practically possible to separate out the internal organs of *E. fetida* from the tissue so we could only distinguish between bifenthrin in the skin and in tissue combined with the gut (Figure 2). For the analytical grade and Capture LFR treatments, concentrations in the gut + tissue and in the skin were significantly different after the uptake phase (Figure 2; $P < 0.001$) with higher concentrations being seen in the skin. For the nanoformulation treatments, concentrations of bifenthrin accumulated in the gut combined with the internal organs were significantly higher than in the skin ($P < 0.001$). Following the depuration phase, concentrations of bifenthrin in the gut + tissue and skin in all treatments were similar (Figure 2) and not significantly different within all treatment ($P > 0.1$) except for

351 the analytical grade treatment (P = 0.003). As we were unable to fully characterise the
352 distribution of the a.i. in *E. fetida* our modelling efforts focused on the *L. terrestris* studies.

353 Model 1, and combinations of models 1 and 3 and models 2 and 3 were used to simulate
354 uptake and depuration in the different *L. terrestris* treatments (Figure 4). Model 1 performed
355 very well for estimating the concentrations of the a.i. in the whole organism for the analytical
356 grade and Capture LFR treatments (Nash Index > 0.90). Predictions for the nano treatments
357 were also good (Nash Index > 0.48) although the model overestimated whole organism
358 concentrations at the end of the depuration phase (Figure 4). When model 1 was combined
359 with model 3 to simulate distribution of the a.i. between gut- skin- and remaining-tissues, good
360 predictions were obtained for the analytical grade and Capture LFR treatments for all tissues
361 (Nash Index > 0.76) and for the skin in the two nano treatments (Nash Index > 0.39).
362 Underestimates of concentrations in the gut and overestimates of concentrations in the tissue
363 by a factor of 8-11 were obtained using a combination of models 1 and 3 for both nano
364 treatments (Nash Index < 0.05). The fact that the two models worked well for estimating
365 behaviour in the Capture treatment is encouraging and suggests that estimates of uptake and
366 distribution based on analytical grade material can be used to extrapolate to behaviours in
367 traditional formulations. The approach worked less well for the nanoformulations so we then
368 extended the modelling to factor in the effect of the release rate from the capsule.

369 Incorporation of the a.i. release from the nanocapsule into the modelling of the
370 nanoformulations (i.e. model 2 and model 3 were used) resulted in predictions that fitted the
371 whole organism data and the skin data well (Nash Index > 0.58). This approach
372 underestimated concentrations in the gut at the end of the uptake phase while predictions of
373 concentrations at the end of the depuration phase were close to the measured data. This is a
374 direct result of the model assumption that compound distribution between different tissues
375 (skin, gut and remaining tissue) is instantaneous and fixed by distribution factors and a
376 temporal change in gut clearance between nano and non-nano formulations (Figure 3).
377 Nonetheless, inclusion of the release rate resulted in better predictions, compared to the

approach not considering release, of internal tissue concentrations of the a.i. with concentrations being a factor of 3.8 (nano A) and 5.1 (nano B) of measured data at the end of the uptake phase and within a factor of 5.7 (nano A) to 7.5. (nano B) at the end of the depuration phase. While the predictions were not perfect, these results indicate that to model internal tissue exposure, which will likely represent the toxicologically important fraction of the accumulate a.i., it is necessary to factor in the release rate of the a.i. from the nanocapsule into the toxicokinetic modelling.

Implications for risk and a potential modelling approach

We have demonstrated that nanoencapsulation will affect the behaviour and uptake of pesticides in soil. For bifenthrin, it will decrease the sorption of the active ingredient to soils, increase the apparent persistence of the compound and alter the uptake behaviour of the active ingredient into earthworms and the subsequent distribution. Consequently, the risk of nanoencapsulated bifenthrin to earthworms will be different from a conventional product. Whether nanoencapsulation increases, decreases or has no effect on risk is difficult to establish at this stage. While nanoencapsulated bifenthrin is taken up more quickly by the earthworms, from the *L. terristris* studies, it appears that the majority of the bifenthrin taken up is contained in the gut so the internalised concentration is lower than in earthworms exposed to the analytical grade substance and a conventional formulation. If less active ingredient is internalised, one would assume that less of the active ingredient will reach the site of toxic action so the effects of the nanoformulation will be lower. However, nanoencapsulation also increases the apparent persistence of the active ingredient which will lengthen the exposure duration of the earthworms to the active ingredient in the nanoformulation compared to a conventional ingredient. The increased efficacy of the nanoformulation compared to conventional formulations could mean that application rates to field are decreased which will also affect risk. The risks to birds and mammals feeding of earthworms could also be altered. If a nanoformulation is applied at the same rate as a conventional product then the oral exposure of these organisms will increase but differences in the bioaccessibility of

405 nanoencapsulated bifenthrin compared to free bifenthrin could be lower meaning less is
406 internalised. Again the duration of exposure will increase.

407 To answer some of these questions around the implications of changes in fate and uptake or
408 effects, a toxicokinetic toxicodynamic modelling approach is probably required (Ashauer and
409 Escher, 2010). In Figure 5, we present a conceptual model, based on our experimental
410 findings and investigations into the performance of the different toxicokinetic modelling
411 approaches, that could be used to model the toxicokinetics of a nanoencapsulated active
412 ingredient. The model assumes sorption to soil is instantaneous following release of pesticide
413 from the capsule and that it is the free (i.e. dissolved pore water) pesticide that is taken up into
414 the earthworm tissue – this assumption is supported by our distribution studies in *L. terrestris*
415 and the testing of Models 2 and 3. The internal concentration in the earthworm tissue over
416 time, needed for toxicokinetic toxicodynamic modelling of the effects, are then calculated
417 based on the release rate from the capsule, the soil-water distribution coefficient and the
418 uptake and depuration rates of the free active ingredient into/out of the earthworm. To estimate
419 oral exposure of birds and mammals, the mass concentration of the active ingredient in the
420 gut also needs to be considered and this is estimated based on the feeding rate of the
421 earthworm on whole soil and on the nanoparticles.

422 We believe that this conceptual model is a useful first step towards developing improved
423 environmental risk assessment approaches for estimating the uptake and effects of
424 nanoencapsulated pesticides in earthworms. The approach might also be applicable to other
425 materials (e.g. nanoencapsulated pharmaceuticals) and other organisms. In the future, we
426 recommend that the model be further parameterised for bifenthrin. We also recommend that
427 studies of the type reported here are done on a wider range of organisms using other pesticide
428 active ingredients with different persistence and physico-chemical properties contained in a
429 wider range of nanocarrier materials in order to evaluate the broader applicability of the model.

430 **Acknowledgements**

We would like to thank Dr. Darren Anderson from Vive Crop Protection Inc. who provided the nanoencapsulated test materials. MFMA was funded by the Ministry of Higher Education of Malaysia. AAs contribution to the study received support from the Innovate UK-funded VFETL project.

## Data Availability

The experimental data on which this manuscript is based can be obtained, on request, from the corresponding author.

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521 Table 1: Sorption coefficients ($k_d$), dissipation half-lives (DT50), release half times (RT50),
522 bioconcentration factors (BCFs) and rates for release, uptake and depuration (k) for the
523 different bifenthrin treatments studied in the *Eisenia fetida* and *L. terrestris* studies.

| Endpoint | Study | Unit | Bifenthrin | Capture LFR | Nano A | Nano B |
|---|---|---|---|---|---|---|
| $k_d$ | Uptake and depuration | L $Kg^{-1}$ | 550 ± 21 | 394 ± 15 | 186 ± 4 | 233 ± 17 |
| $k_d$ | Distribution study *E.fetida* | L $Kg^{-1}$ | 494 ± 71 | 371 ± 47 | 154 ± 7 | 163 ± 9 |
| $k_d$ | Distribution study *L. terrestris* | L $Kg^{-1}$ | 585 ± 92 | 488 ± 32 | 274 ± 42 | 251 ± 31 |
| $DT_{50}$ | Uptake and depuration | d | 25 - 27 | 33-35 | 49 - 50 | 38 - 40 |
| Release rate ($k_r$) | Uptake and depuration | $h^{-1}$ | NA | NA | 0.104 ± 0.008 | 0.182 ± 0.016 |
| $RT_{50}$ | Uptake and depuration | d | NA | NA | 6 - 7 | 3 - 4 |
| $k_{in}$ *E.fetida* | Uptake and depuration | $LKg^{-1}h^{-1}$ | 0.222 ± 0.009 | NA | NA | NA |
| $k_{out}$ *E.fetida* | Uptake and depuration | $h^{-1}$ | 0.0036 ± 0.0002 | NA | NA | NA |
| BCF *E.fetida* | Uptake and depuration | - | 61.7 | NA | NA | NA |
| $k_{in}$ *L. terrestris* | Uptake and depuration | $LKg^{-1}h^{-1}$ | 0.7021 ± 0.0336 | NA | NA | NA |
| $k_{out}$ *L. terrestris* | Uptake and depuration | $h^{-1}$ | 0.0033 ± 0.0003 | NA | NA | NA |
| BCF *L. terrestris* | Uptake and depuration | - | 212 | NA | NA | NA |

524 [1] Determined from data presented here with additional data on soil concentrations over time in four other soils
525 (unpublished).

526

Figure Legends

**Figure 1:** Measured (dots) and predicted (lines) total internal concentration of different formulations of bifenthrin in *E. fetida* over time using Model 1 (black) and Model 2 (grey). Dotted lines indicate the 95% confidence interval for the predictions. Model parameterisation was conducted with data from bifenthrin a.i..

**Figure 2:** Proportion of total internal concentration of different formulations of bifenthrin in earthworms after 10 d uptake and 7 d depuration as average ± SD in relation to the internal concentration at the end of the uptake phase.

**Figure 3:** Proportion of the gut concentration at the end of the uptake phase of different formulations of bifenthrin recovered from *L. terrestris* faeces samples as average ± SD.

**Figure 4**: Measurements (dots) and predictions from different model combinations (lines) of internal concentration of different formulations of bifenthrin in whole organism and different compartments of *L. terrestris* over time. Grey backgrounds indicate that predictions account for the released fraction of bifenthrin from the nanoformulation. Dotted lines indicate the 95% confidence intervals for the predictions.

**Figure 5:** Conceptual model for estimating residues of active ingredients contained in nanoencapsulated formulations in terrestrial invertebrates over time.

Figure 1

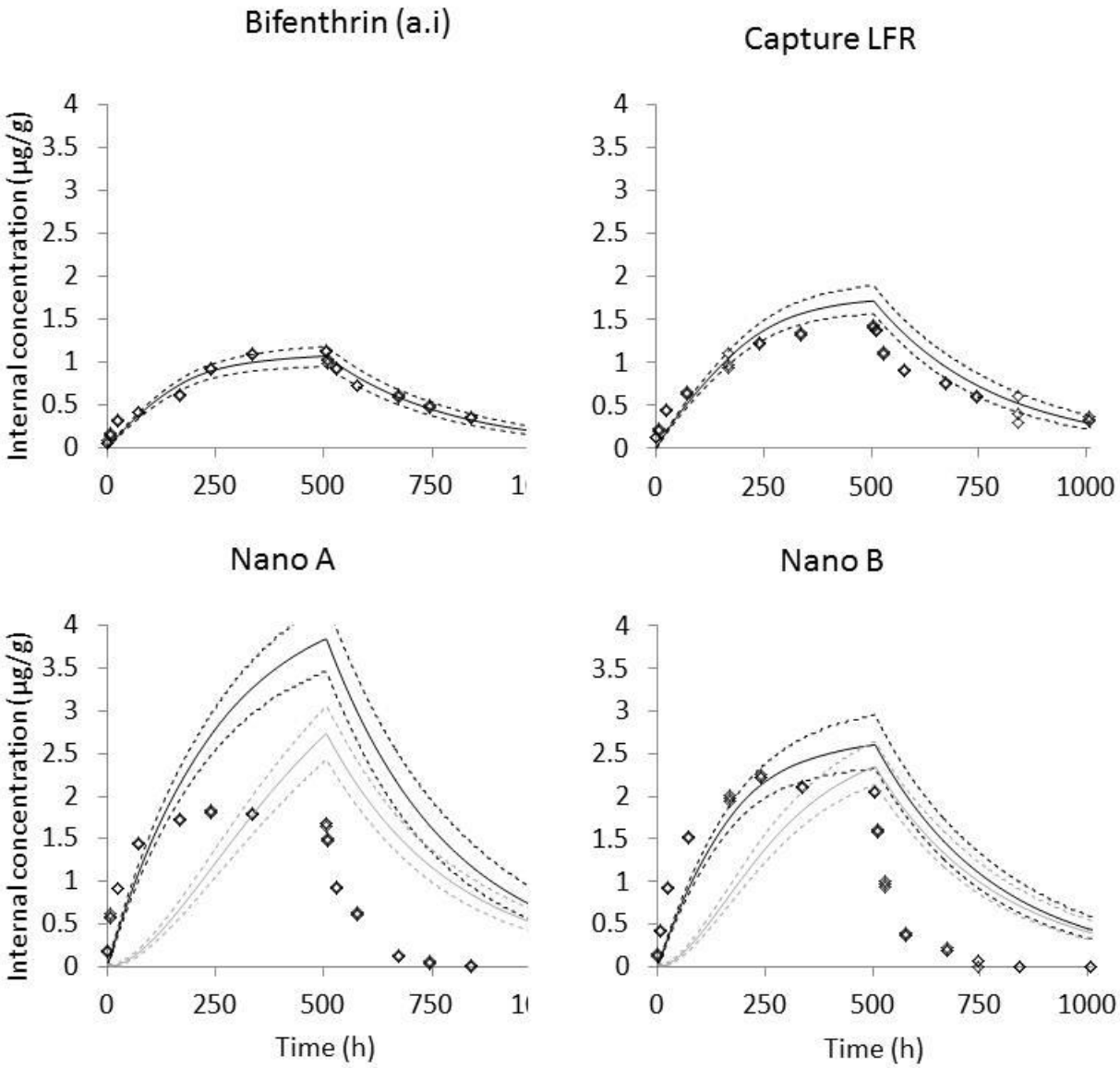

Bifenthrin (a.i)
Capture LFR
Nano A
Nano B
Internal concentration (µg/g)
Time (h)
0
0.5
1
1.5
2
2.5
3
3.5
4
0
250
500
750
1000

Figure 2

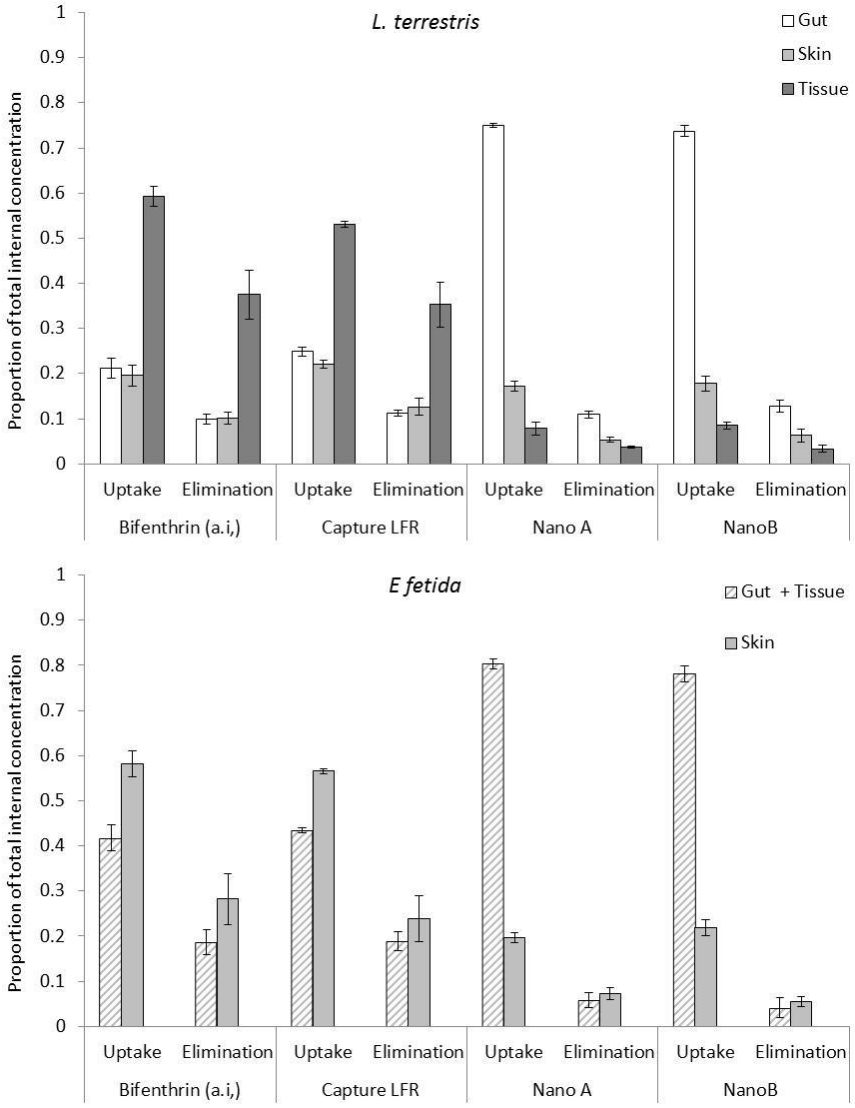

L. terrestris
Gut
Skin
Tissue
Proportion of total internal concentration
1
0.9
0.8
0.7
0.6
0.5
0.4
0.3
0.2
0.1
0
Uptake
Elimination
Bifenthrin (a.i,)
Capture LFR
Nano A
NanoB
E fetida
Gut + Tissue
Skin

Figure 3

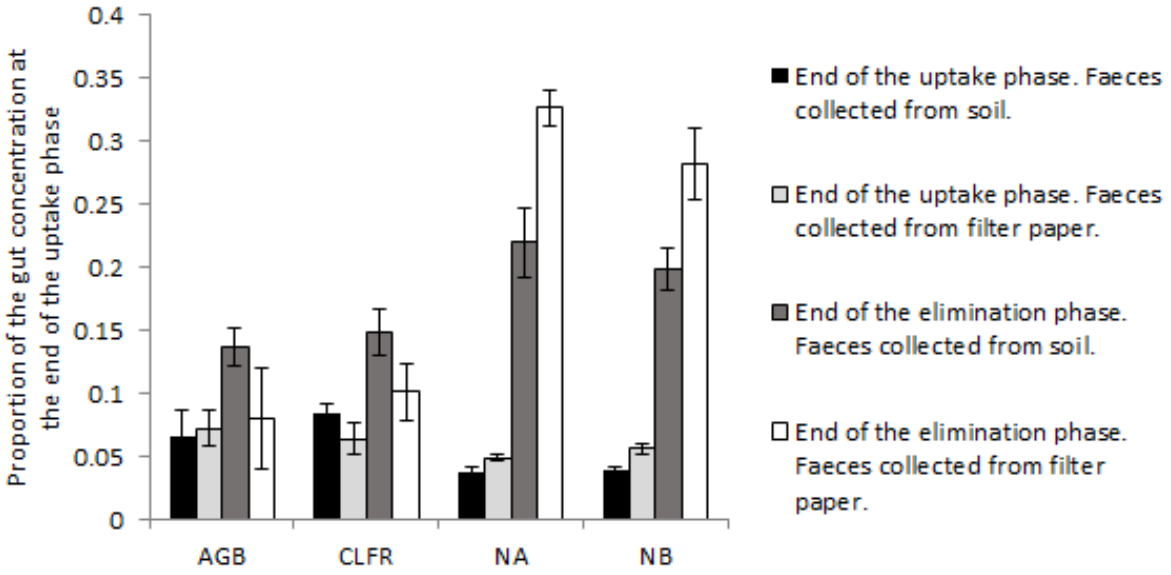

Proportion of the gut concentration at the end of the uptake phase
0.4
0.35
0.3
0.25
0.2
0.15
0.1
0.05
0
AGB
CLFR
NA
NB
End of the uptake phase. Faeces collected from soil.
End of the uptake phase. Faeces collected from filter paper.
End of the elimination phase. Faeces collected from soil.
End of the elimination phase. Faeces collected from filter paper.

Figure 4

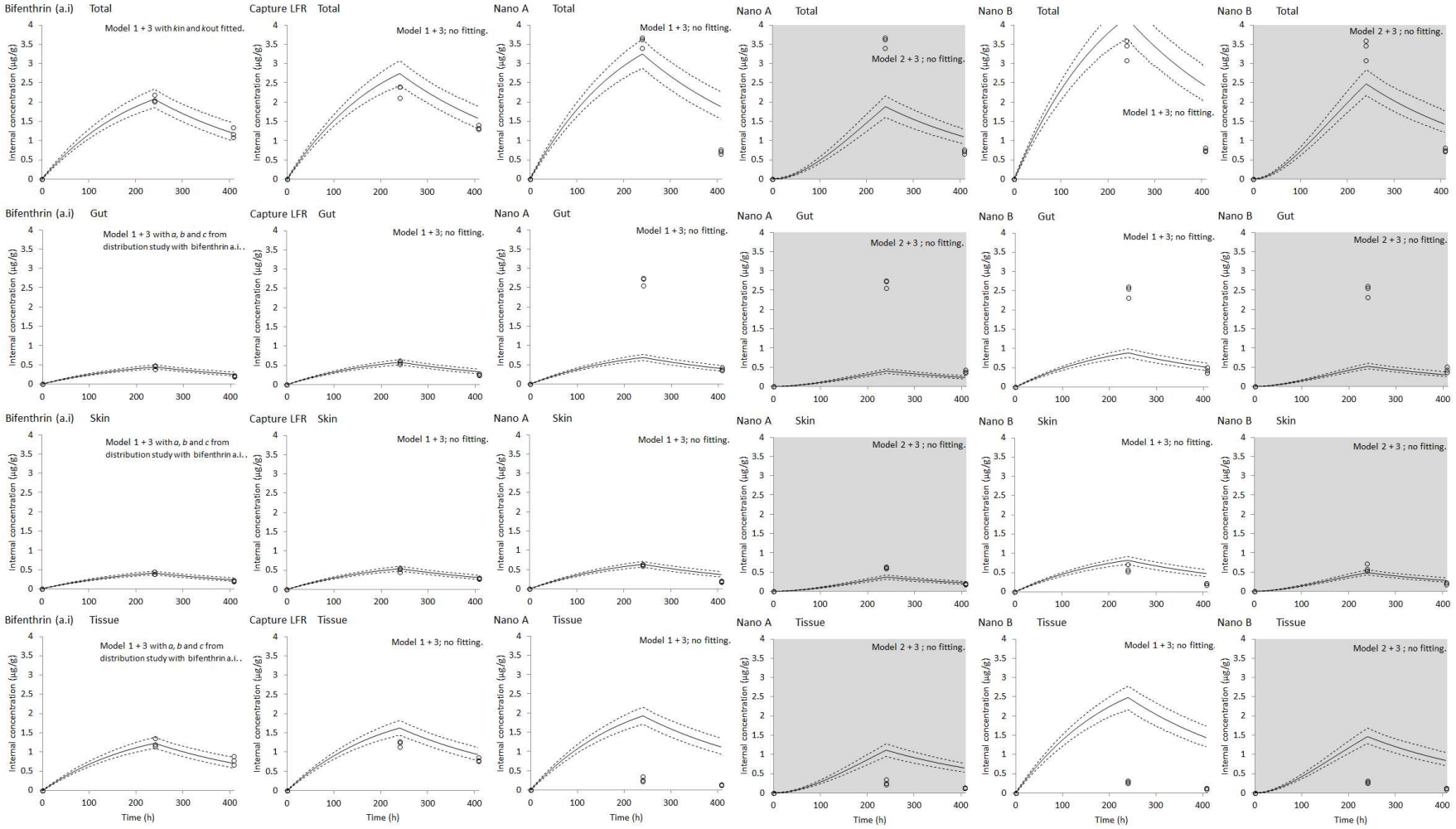
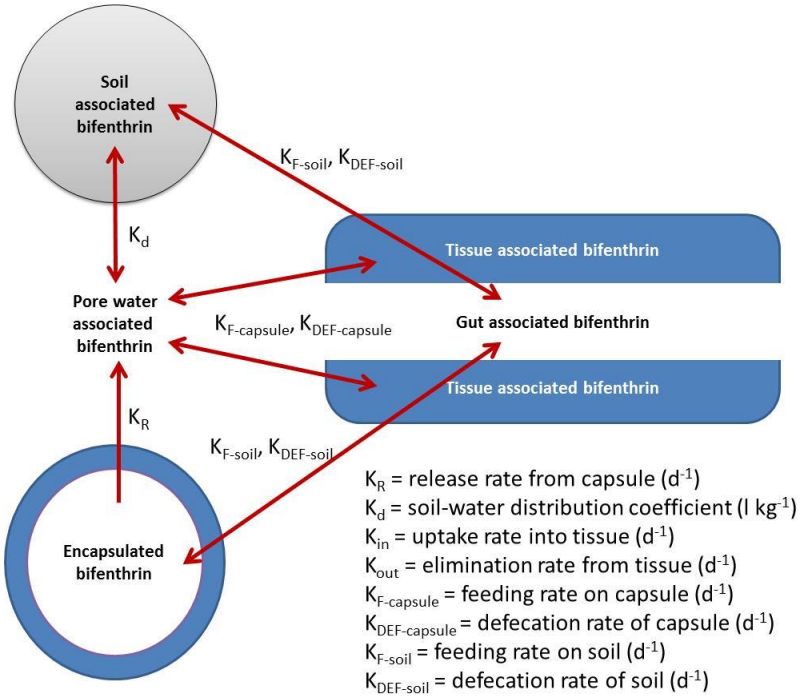

Bifenthrin (a.i) Total
Model 1 + 3 with kin and kout fitted.
Capture LFR Total
Model 1 + 3; no fitting.
Nano A Total
Model 1 + 3; no fitting.
Nano A Total
Model 2 + 3 ; no fitting.
Nano B Total
Model 1 + 3; no fitting.
Nano B Total
Model 2 + 3 ; no fitting.
Bifenthrin (a.i) Gut
Model 1 + 3 with a, b and c from distribution study with bifenthrin a.i. .
Capture LFR Gut
Model 1 + 3; no fitting.
Nano A Gut
Model 1 + 3; no fitting.
Nano A Gut
Model 2 + 3 ; no fitting.
Nano B Gut
Model 1 + 3; no fitting.
Nano B Gut
Model 2 + 3 ; no fitting.
Bifenthrin (a.i) Skin
Model 1 + 3 with a, b and c from distribution study with bifenthrin a.i. .
Capture LFR Skin
Model 1 + 3; no fitting.
Nano A Skin
Model 1 + 3; no fitting.
Nano A Skin
Model 2 + 3 ; no fitting.
Nano B Skin
Model 1 + 3; no fitting.
Nano B Skin
Model 2 + 3 ; no fitting.
Bifenthrin (a.i) Tissue
Model 1 + 3 with a, b and c from distribution study with bifenthrin a.i. .
Capture LFR Tissue
Model 1 + 3; no fitting.
Nano A Tissue
Model 1 + 3; no fitting.
Nano A Tissue
Model 2 + 3 ; no fitting.
Nano B Tissue
Model 1 + 3; no fitting.
Nano B Tissue
Model 2 + 3 ; no fitting.
Internal concentration (µg/g)
Time (h)

Figure 5.


Soil associated bifenthrin
$K_{F\text{-}soil}$, $K_{DEF\text{-}soil}$
$K_d$
Tissue associated bifenthrin
Pore water associated bifenthrin
$K_{F\text{-}capsule}$, $K_{DEF\text{-}capsule}$
Gut associated bifenthrin
Tissue associated bifenthrin
$K_R$
$K_{F\text{-}soil}$, $K_{DEF\text{-}soil}$
Encapsulated bifenthrin
$K_R$ = release rate from capsule ($d^{-1}$)
$K_d$ = soil-water distribution coefficient (l $kg^{-1}$)
$K_{in}$ = uptake rate into tissue ($d^{-1}$)
$K_{out}$ = elimination rate from tissue ($d^{-1}$)
$K_{F\text{-}capsule}$ = feeding rate on capsule ($d^{-1}$)
$K_{DEF\text{-}capsule}$ = defecation rate of capsule ($d^{-1}$)
$K_{F\text{-}soil}$ = feeding rate on soil ($d^{-1}$)
$K_{DEF\text{-}soil}$ = defecation rate of soil ($d^{-1}$)