



UNIVERSITY OF LEEDS

This is a repository copy of *Extending the applications of sediment profile imaging to geochemical interpretations using colour*.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/124802/>

Version: Accepted Version

Article:

Statham, PJ, Homoky, WB, Parker, ER et al. (6 more authors) (2019) Extending the applications of sediment profile imaging to geochemical interpretations using colour. *Continental Shelf Research*, 185. pp. 16-22. ISSN 0278-4343

<https://doi.org/10.1016/j.csr.2017.12.001>

© 2017 Elsevier Ltd. This manuscript version is made available under the CC-BY-NC-ND 4.0 license <http://creativecommons.org/licenses/by-nc-nd/4.0/>

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

1 **Extending the applications of sediment profile imaging to geochemical**
2 **interpretations using colour**

3
4 P.J. Statham^a, W.B. Homoky^b, E.R. Parker^c, J.K. Klar^{a,d}, B. Silburn^c, S.W. Poulton^e, S.
5 Kröger^c, R.B. Pearce^a, E.L. Harris^a
6

7 ^aOcean and Earth Science, University of Southampton, National Oceanography Centre,
8 Southampton, SO14 3ZH, United Kingdom

9 ^bUniversity of Oxford, Department of Earth Sciences, South Parks Road, Oxford, OX1 3AN,
10 United Kingdom

11 ^cCentre for Environment, Fisheries and Aquaculture Science, Pakefield Road, Lowestoft,
12 NR33 0HT, United Kingdom

13 ^dPresent address: LEGOS, Université de Toulouse, CNES, CNRS, IRD, UPS, 14 Avenue
14 Edouard Belin, 31400 Toulouse, France

15 ^eSchool of Earth and Environment, The University of Leeds, Leeds. LS2 9JT, United
16 Kingdom
17

18 Corresponding author: email pjs@soton.ac.uk

19 Keywords: iron, manganese, shelf sediments, SPI colour, geochemistry
20

21 **Abstract**

22 Whilst Sediment Profile Imaging (SPI) is a very widely used technique in the regulatory
23 assessment of seabed environmental health, and in the study of seafloor sediment-biology
24 interactions, the potential for SPI images to be used in a geochemical context has not been
25 rigorously assessed. Here we have examined relationships between colour and geochemistry
26 in a sediment core collected from the Celtic Sea, North West European Shelf, that was
27 digitally imaged and on which detailed geochemical analyses were also performed. Average
28 oxygen penetration depth was 4.08 ± 0.72 mm, (n=5), whilst the apparent redox potential
29 discontinuity (aRPD) as determined by sediment colour change was at 78 mm. As iron
30 (oxyhydr)oxides decreased with depth, black sulfide phases increased, and the aRPD most
31 closely correlated with this geochemical change rather than the oxygen penetration depth.
32 Colour analysis of the image showed a clear correlation of brightness with black FeS (acid

33 volatile sulfide). There was a general correlation of iron oxide phases with orange colour in
34 the upper part of the sediment profile, whilst in the lower part of the core the orange oxide
35 phases appeared to be obscured by the black FeS present. The sulfide-brightness relationship
36 indicates colour analysis can provide an estimate of FeS, and potentially the carrying capacity
37 for toxic metals such as cadmium, zinc and copper as sulfides in this type of sediment.
38 Additionally, detailed geochemical analyses of SPI cores may provide new insights into the
39 activity and impacts of infauna and the link with sediment biogeochemical cycles of carbon
40 and nutrients.

41

42 **1. Introduction**

43

44 The development of the in situ sediment profile imaging (SPI) technique has provided a
45 powerful tool for rapid spatial assessment of biological activity in surface sediments (Rhoads
46 and Cande, 1971). The SPI approach is now widely used for assessing environmental
47 quality parameters (Germano et al., 2011; Solan et al., 2003) in general and also in relation to
48 national and international standards. These applications are based on data from the images
49 being used with current conceptual models of sediment-organism relationships

50

51 An important parameter first derived from the SPI images in the 1970s was a redox potential
52 discontinuity (RPD) corresponding to colour changes and assumed variations in
53 geochemistry (Fenchel, 1969; Vismann, 1991). However, the RPD subsequently became
54 synonymous with biological mixing depth on the premise that the colour transition was
55 biologically mediated. The RPD was then used in gauging the ecological health within
56 successional models, and was subsequently used in the derived indices of Benthic Habitat
57 Quality (Rosenberg et al., 2004) and the Organism-Sediment Index (Rhoads and Germano,

58 1986). Whilst microbial communities associated with burrow structures do appear to be
59 linked to availability of oxidants (Bertics and Ziebis, 2009), little other data supports a strong
60 link between macro organism mixing depth and the RPD. The realisation of this limitation
61 led to the use of “apparent RPD” to describe the colour transitions seen. As measurements of
62 the redox potential, or *in situ* oxygen and other redox variables such as iron or nitrate, are not
63 possible with current SPI technology, it has been assumed that reddish-brown sediment
64 colour tones are indicative of sediments in an oxidative geochemical state, and that the
65 sediment porewaters are not intensely reducing. The rigorous interpretation of the aRPD in
66 relation to oxidising conditions thus remains difficult (Gerwing et al., 2015), and despite the
67 effort expended in obtaining these images only limited work has been done on exploring the
68 potential to use such images for geochemical investigations (Grizzle and Penniman, 1991;
69 Teal et al., 2009; Teal et al., 2010), and then largely on solute phases of sediment chemistry
70 rather than the particulate components which may contribute the bulk of the colour observed .

71

72 A direct approach to providing improved metal biogeochemical data used chemical sensors
73 incorporated into the SPI system (Teal et al., 2013; Teal et al., 2009). The diffusive gradients
74 in thin films (DGT) technique applied provides fluxes of dissolved Fe and Mn from sediment
75 to the collection gel. Whilst useful data on time integrated liberation of dissolved Fe and Mn
76 in the sediment matrix is provided, as the measurements are done *in situ* it is not possible to
77 relate fluxes to the oxygen penetration depth, or porewater and solid phase composition.

78

79 Here we have taken the novel approach of collecting a core that was digitally imaged and
80 also geochemically analysed by a wide range of techniques to provide data on porewater and
81 solid phase parameters. This approach allows a direct comparison of image and sediment
82 geochemistry at this shelf site, and provides a rigorous basis for assessing the potential of

83 using SPI images in a geochemical context. Additionally the geochemical signatures can
84 provide insights into linked biogeochemical cycles and biological processes occurring
85 (Bertics and Ziebis, 2009).

86

87 **2. Methods**

88

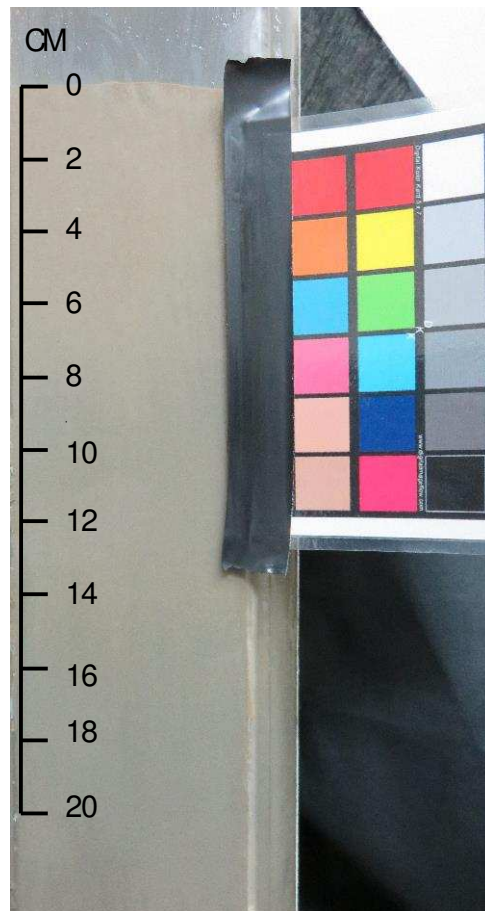
89 The work was carried out within the framework of the NERC funded Shelf Sea
90 Biogeochemistry Programme. A NIOZ box corer was used at cohesive Site A (sandy mud,
91 $\sim 51^{\circ} 12.6754' \text{ N}$, $6^{\circ} 8.0277' \text{ W}$) within the Celtic Sea, UK (Thompson et al., 2017) in August
92 2015. The NIOZ collected sediment was sub-sampled using a modified 10 cm diameter core
93 tube (Figure 1a) in which a flat sheet of transparent polymethyl methacrylate replaced part of
94 the tube wall. The flat face allowed a digital image to be taken under laboratory lighting
95 using a Canon G-15 camera (12.1 megapixel CMOS detector) of a cross section of the core
96 (Figure 1b; see supplementary material for a high-resolution [637×2577 pixels, 300dpi]
97 image file). A standard colour card (Digital Kolor Kard from digitalimageflow.com) was
98 photographed at the same time for scale and colour inter-comparison.

99 Figure 1. a) Modified core tube. Original tube size 10 cm diameter by 60 cm long) b).

100 Colour image of core, showing depth scale and reference colour card used.



a)



b)

101

102 The aRPD was determined following a published procedure (Solan et al., 2004). Here the
103 image is analysed using the open access Image-J programme, in which the image is broken
104 down into RGB colours, the red channel chosen and the range of intensities selected to best
105 highlight the colour change associated with the aRPD.

106 A detailed description of most sampling and analytical techniques used is provided elsewhere
107 (Klar et al., 2017) and only an overview is given here, except for those techniques not
108 covered in the Klar et al. paper. The strategy was to choose analytes that indicate the redox
109 state of the core, and solid phases that would impact colour (thus for example iron
110 (oxyhydr)oxides would be expected to be an orange colour and initially formed under
111 oxidising conditions, whilst FeS is black and is indicative of typically reducing conditions).

112 Replicate dissolved oxygen profiles across the core surface were collected behind the flat
113 wall of the core tube before any subsampling was done. A Unisense Clark type electrode with
114 a 100µm tip was used, and data on oxygen was obtained at typically 200µm depth intervals
115 across the benthic interface and into the sediment. With core top water removed, porewaters
116 were extracted from the sediment core at typically 1 to 2 cm depth intervals using Rhizon
117 samplers (Seeberg-Elverfeldt et al., 2005) that prevent oxygen contamination of the collected
118 porewaters (*c.f.* Klar et al. 2017). After porewater extraction and filtration, the residual
119 sediment was sliced using a polycarbonate sheet at 0.5, 1 and 2 cm depth-intervals, and
120 stored at -20° C in zip-lock bags prior to further analyses.

121 The concentrations of Fe(II), and Fe(II) plus Fe(III) (i.e. after addition of a reducing agent),
122 were determined in the dissolved (<0.2 µm) size fractions of porewater samples using the
123 Fe(II)-complexing ferrozine ligand (Sigma-Aldrich) (Stookey, 1970; Viollier et al., 2000). In
124 order to examine associations of Fe and Mn with solid sediment phases, firstly an ascorbic
125 acid leach (Raiswell et al., 2010) was used to extract the easily reducible oxide phases, such
126 as amorphous ferrihydrite, but not the more crystalline oxide phases. A further citrate
127 dithionite reducing leach was then applied to each sample to remove more crystalline Fe
128 oxide phases including haematite and goethite (Poulton and Canfield, 2005; Raiswell et al.,
129 1994). The total dissolution of non-leached freeze dried and ground sediment samples used a
130 mixture of hydrofluoric, nitric and hydrochloric acids in PFA containers on a hotplate. Fe and
131 Mn in the leach solutions were determined using an inductively coupled plasma optical
132 emission spectrometer (ICP-OES, iCAP6000 Series, Thermo Scientific). Nutrient
133 concentrations in sediment porewaters were all analysed on board using a Bran and Luebbe
134 segmented flow colorimetric auto-analyser (Woodward and Rees, 2001). Particulate organic
135 carbon (POC) and nitrogen (PON) were determined using a Carlo-Erba CHNOS analyser
136 (Nieuwenhuize et al., 1994).

137 X-ray diffraction analysis of the sediment used a PANalytical X'Pert pro
138 diffractometer machine fitted with a Cu X-ray tube. The machine operating conditions
139 were 35kV, 40mA utilising automatic slits and a step size of $0.02^\circ 2\theta$ at 1 second/
140 step. The samples were prepared as randomly oriented powder samples with an
141 internal standard of 25% by weight of corundum and side-loaded to avoid preferred
142 orientation. Precision values for the samples are approximately $\pm 0.5-2\%$ for
143 crystalline materials and $\pm 10-20\%$ (of the amount present) for total clay. Scanning
144 electron microscope work used a Carl Zeiss LEO1450VP Scanning Electron
145 Microscope (SEM) fitted with an Oxford Instruments EDS system, and an X-Act
146 Silicon Drift Detector (10mm^2 area) using the AZtec Energy software system (v.3.1).

147
148 Acid volatile sulfide (AVS) and pyrite (PY) were determined in the sediment following the
149 approach of Canfield et al. (1986) using a sequential acid (6M HCl) reflux, and then acidified
150 chromous chloride reflux, to convert the solid phase sulfides to hydrogen sulfide that is
151 collected as silver sulfide (Ag_2S) in a silver nitrate solution. The two sulfide pools in the
152 sediment are calculated after gravimetric measurement of the Ag_2S formed from the known
153 mass of sediment added.

154
155 Analysis of the core colour was based on the iron (oxyhydr)oxides appearing in the red-
156 orange part of the spectrum (haematite and ferrihydrite are red-orange, goethite is orange)
157 and iron mono-sulfides (FeS ; AVS) being black and impacting image brightness. Colour
158 analysis was done using the open access ImageJ suite (Fiji implementation). In order to
159 compare brightness with measured AVS concentrations, the image was first converted to a
160 hue-saturation-brightness (HSB) stack, and the brightness layer examined. Brightness values
161 were read off at 5 points across the image at the sampled depth, and averaged to provide a

162 value to plot against AVS. For the iron (oxyhydr)oxide phases, the zone in the image
163 corresponding to the geochemical analysis was selected and the colour threshold selected to
164 correspond to orange values. Digital values of 20-37 bracketing the orange range (circa 590-
165 620 nm) were chosen and pixels in this range were picked out on the image. The image was
166 then analysed for the fraction of pixels that corresponded to the orange range. The fractions
167 were then compared to the corresponding iron (oxyhydr)oxide data.

168

169 3. Results and Discussion

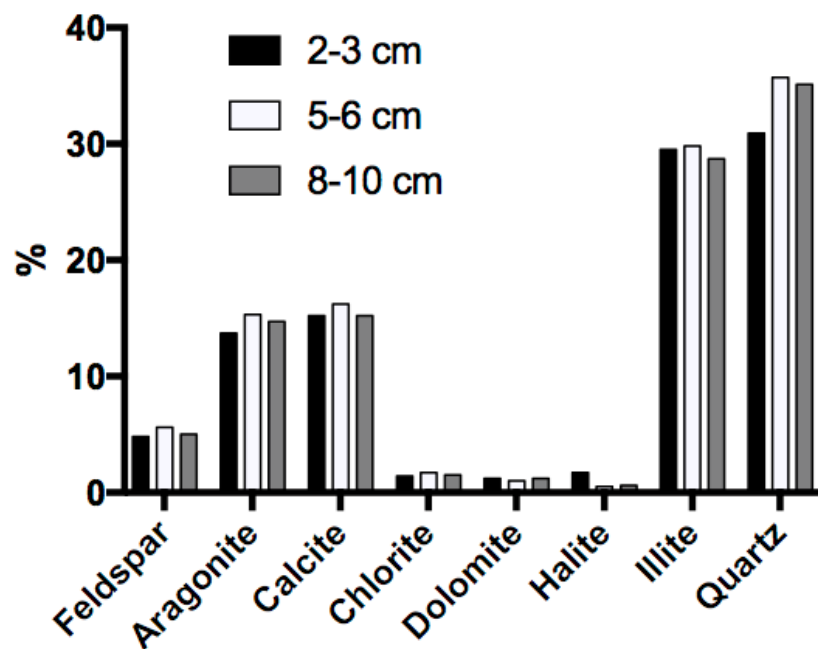
170

171 3.1 Bulk composition

172 Site A is a very poorly sorted, very fine skewed, mesokurtic, very coarse silt, classified
173 according to the Folk classification scheme as a sandy mud; see Thompson et al. (2017) for
174 further detail. Major components of the sediment matrix are shown in Figure 2.

175

176 Figure 2. Bulk mineralogy of the core at discrete depths.



177

178 Dominant phases are carbonates (primarily aragonite and calcite), quartz, illite, and
179 plagioclase feldspar; changes in these components with depth are small. The carbonate
180 fractions are expected to come from two sources. Firstly, eroded Cretaceous carbonaceous
181 rocks would have been transported here during the last Ice Age when the major river system
182 running through what is now the English Channel deposited a fraction of its particle load.
183 Secondly, carbonate will have been generated by the ubiquitous shelled organisms amongst
184 the benthos living within the surface sediments. The independent particulate inorganic carbon
185 data is in reasonable agreement with the XRD data (averages of 36.5 and 30.6%
186 respectively). Particulate organic carbon (average 1.05%; see supplementary material) is a
187 small component of the total carbon in this matrix, and remains fairly constant with depth
188 except for a small increase towards the surface. The quartz, feldspar and illite are products of
189 lithogenic erosion and weathering.

190

191 **3.2 Redox status of the core**

192

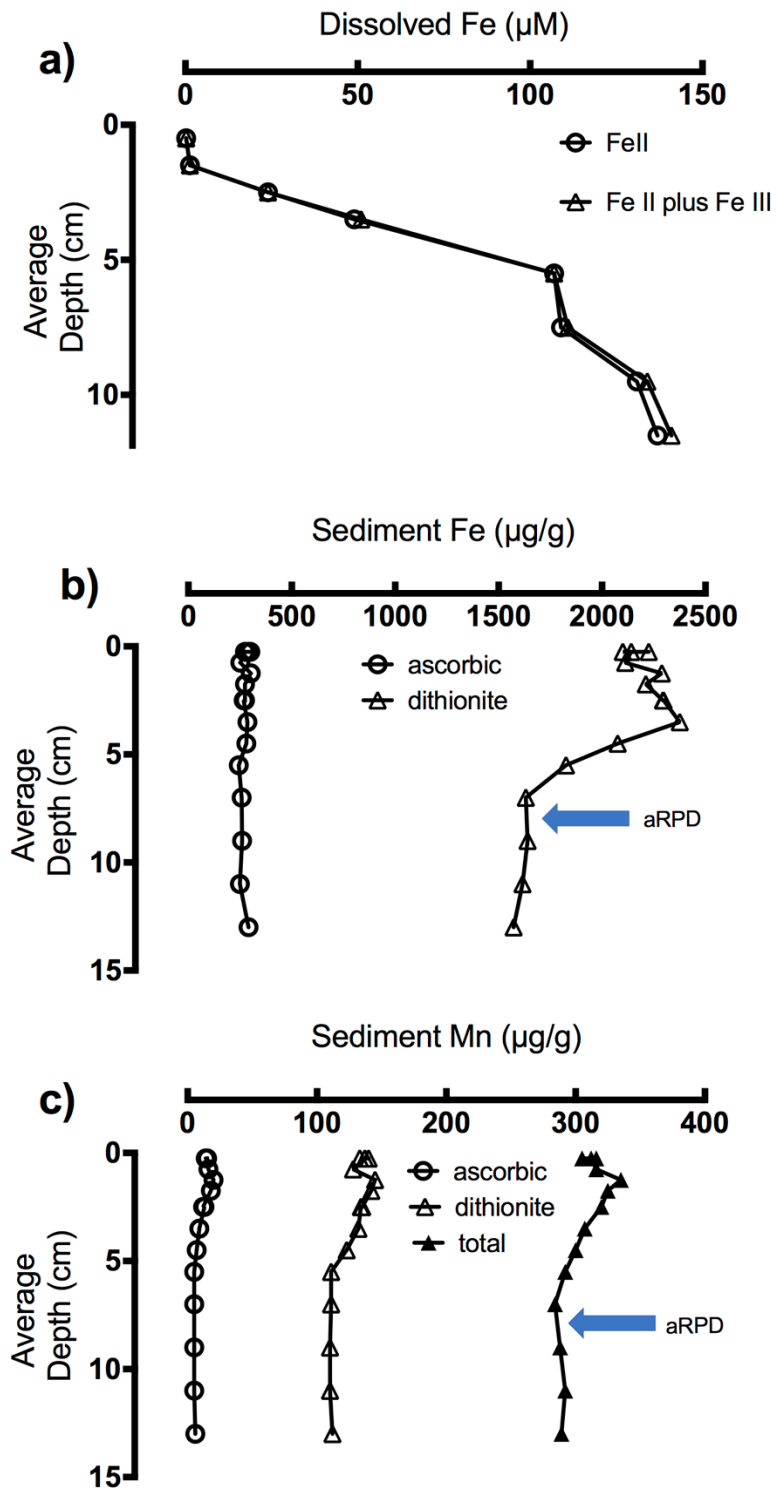
193 In accordance with the accepted general diagenetic sequence of terminal electron acceptors
194 (Burdige, 2006) oxygen and then nitrate rapidly disappear in the upper few mm of the core.
195 Measured oxygen penetration depths (4.08 ± 0.72 mm, $n=5$) are more than an order of
196 magnitude shallower than the aRPD (78 mm). Further degradation of organic carbon is
197 expected to sequentially use Mn and Fe oxides and then sulfate as electron acceptors. Mn
198 oxides can be important electron sinks when present at high enough concentrations
199 (Thamdrup et al., 1994). However, in this core average total Mn (297 $\mu\text{g/g}$) is only 2.1 % of
200 average total Fe (14110 $\mu\text{g/g}$) on a mass/mass basis, and the reducible Mn oxides (dithionite
201 released) are at low concentrations relative to the equivalent Fe oxide phases (127 c.f. 2027
202 $\mu\text{g/g}$). Thus iron (oxyhydr)oxides are expected to be the most important electron acceptors.

203

204 Porewater dissolved Fe(II) (dFe(II)) follows the trends expected with very low concentrations
205 near surface, where iron oxidation is possible, increasing down to about 6 cm, followed by
206 high and relatively constant dFe(II) concentrations deeper in the core (Figure 3a).

207

208 Figure 3 a) Dissolved (<0.2 μm) Fe, b) ascorbic acid and dithionite leachable Fe, c) Mn total,
209 ascorbic and dithionite leaches.



210

211

212 Isotope data demonstrates that this dFe(II) is predominantly derived from bacterial

213 dissimilatory iron reduction (DIR) (Klar et al., 2017). A further route for formation of FeII in

214 solution is the reaction between HS^- and iron oxides (see below). Porewater nutrients follow

215 anticipated patterns with nitrate rapidly disappearing in surface sediments, followed by an
216 increase in ammonium, phosphate and dissolved silicon with depth (see supplementary
217 material).

218

219 **3.3 Solid metal phases within the sediment core**

220

221 Total Fe and Mn in sediments (averages 1.41 % and 306 $\mu\text{g/g}$ respectively) are low relative
222 to continental crustal values of 5.13% for Fe and 852 $\mu\text{g/g}$ for Mn (Albarede, 2003). This
223 reflects dilution with illite (variable Fe content but nominally 1.43%), carbonate (normally
224 regarded as a relatively pure diluent in analysis of geological matrices), and quartz. Quartz
225 typically has low concentrations of trace metals and most Fe associated with the mineral is as
226 oxide coatings. Any coatings would be released through the dithionite-citrate leaching
227 techniques applied here (see below).

228

229 Leachable Fe falls into two groups: 1) The ascorbic fraction, that is reported to correspond
230 primarily with amorphous ferrihydrite phases (Raiswell et al., 2010), remains relatively
231 constant throughout the sampled core (average 274 $\mu\text{g/g}$, Figure 3b). This constancy is at first
232 sight surprising as one would expect the geochemically reactive oxide phases to be rapidly
233 reduced in the oxygen deficient zone beneath the surface few mm. In another core from this
234 site (Klar et al., 2017) an increase in ascorbic Fe was noted just below the surface in the
235 oxidised layer, reflecting precipitation of dissolved Fe diffusing upward, whilst average
236 deeper concentrations were similar to those found here, demonstrating consistency at depth
237 between cores. This low but consistent concentration of ascorbic leachable Fe throughout the
238 core most probably reflects a combination of primary ferrihydrite and iron that has been
239 reduced at the surface of Fe oxides, but not yet released to solution (Poulton, 2003). 2) The

240 dithionite-citrate reagent applied after the ascorbic leach will remove the more crystalline
241 oxide phases including goethite and haematite (Poulton and Canfield, 2005; Raiswell et al.,
242 2010). These phases are present at concentrations about an order of magnitude higher than
243 that of the ferrihydrite (Figure 3b). The amorphous phases are expected to be the most
244 geochemically reactive, and unless reduced they are expected to gradually reorder their
245 structures to more crystalline forms. The other important Fe bearing non-lithogenic mineral
246 phases in sediments are sulfides.

247

248 Bacterial sulphate reduction is reported for a range of similar temperate shelf and coastal
249 environments when oxygen and other more energetically favourable terminal electron
250 receptors are not available (Teal et al., 2009; Thamdrup et al., 1994), but the expected
251 resulting dissolved sulfide was not detected at any of the sampled depths of a core taken at
252 the same site (Klar et al., 2017). In this system any dissolved sulphide will rapidly react with
253 both reduced FeII and reactive iron oxides to generate FeS that gives the black colour in
254 reducing sediments (Bull and Williamson, 2001a), and so dissolved sulfide will be consumed
255 immediately after formation. Additional fates for HS^- include release of oxidised S species
256 (Burdige, 2006), and mineralisation with any chalcophilic trace metals (e.g. Cd, Cu, Hg, Pb,
257 Zn). The FeS transforms further to the more stable pyrite (FeS_2), which is the major long
258 term sink for S in sediments (Burdige, 2006). In this core AVS (principally FeS) and pyrite
259 are present at low concentrations (Figure 4).

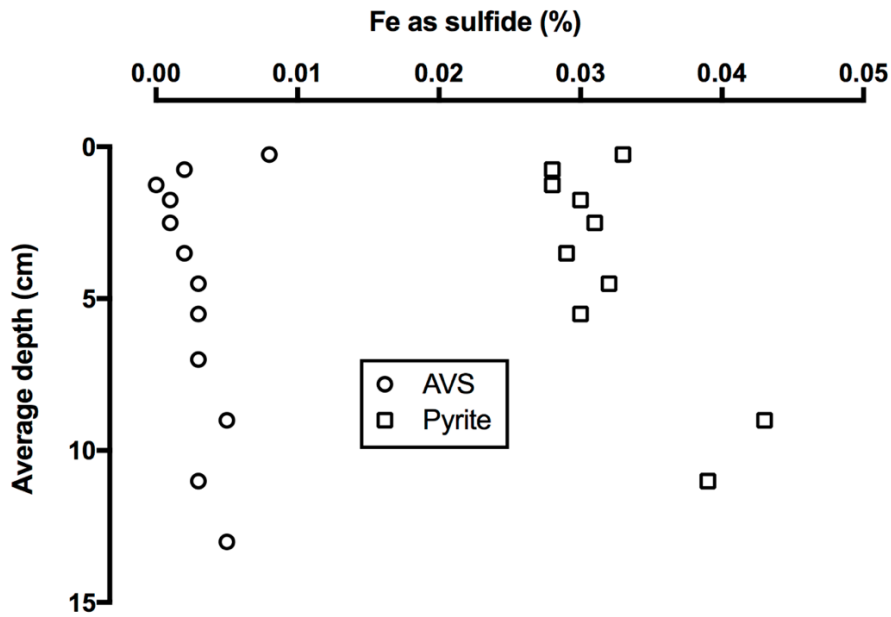
260

261

262

263

Figure 4. Iron sulfide phases in the core.



264

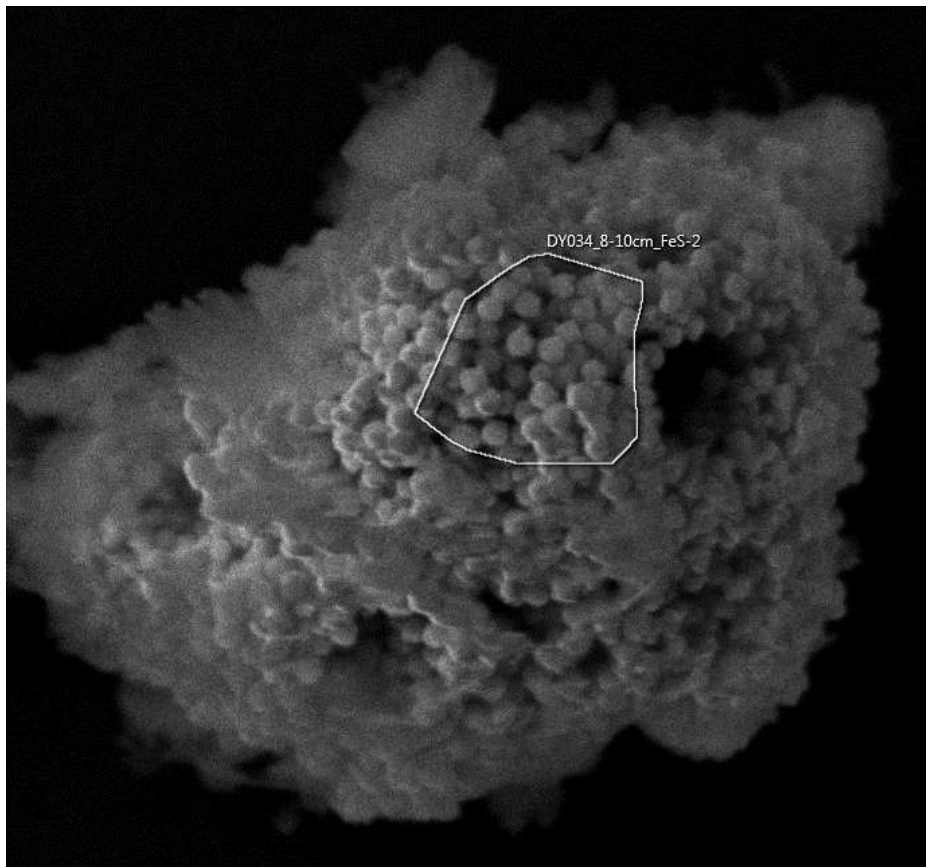
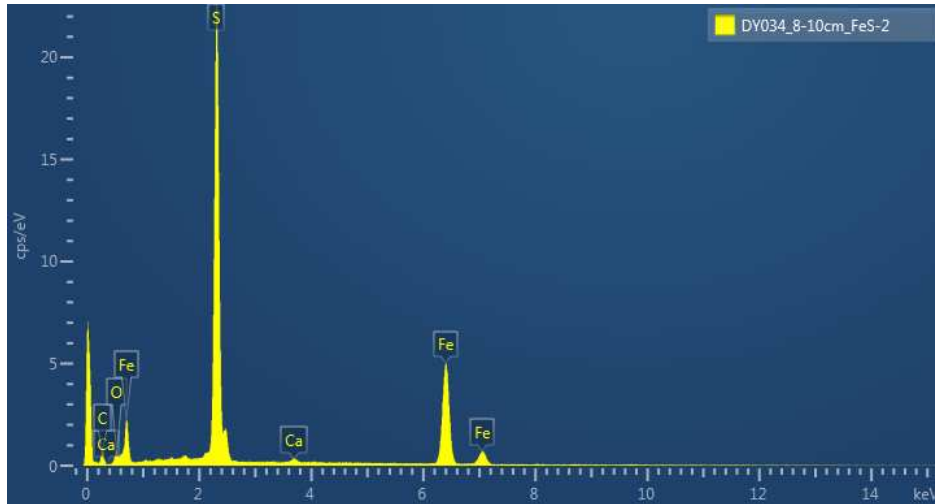
265

266 Both sulfide phases gradually increase with depth, with the biggest increase occurring below
 267 the aRPD, and throughout the pyrite concentrations are an order of magnitude higher than the
 268 corresponding AVS values (Figure 4). Other intermediary Fe-S compounds exist that are only
 269 partially dissolved during our AVS treatment (greigite, Fe_3S_4), but these generally occur at
 270 much lower concentrations than FeS and pyrite. Pyrite is typically found in framboidal or
 271 euhedral forms as shown in the SEM image in Figure 5.

272

273 Figure 5. SEM-image of sediment at 8-10 cm in the core showing pyrite framboidal crystals
274 (bottom) and EDS analysis of polygonal zone (top) shown in the SEM, confirming
275 composition as FeS₂ (maximum width of polygon is 5µm).

276



277

278 The amorphous Fe(oxyhydr)oxides formed by upwards diffusion of FeII and precipitation on
279 reaching oxygen containing porewaters, will be a highly reactive phase with which the HS⁻
280 can react rapidly (Canfield et al., 1992; Poulton et al., 2004). Additionally organic rich

281 microniches can be sites of intensive redox reactions (Lehto et al. 2017). Therefore, the
282 surface few mm of sediment can be an important formation site for FeS and FeS₂, consistent
283 with the higher AVS and PY concentrations at the surface of this core (Figure 4). In addition
284 to in situ formation, bioturbation may also transfer pyrite from deeper in the sediment to
285 surface layers. The crystalline Fe oxides found deeper in the sediments will react more
286 slowly with HS⁻ (Poulton et al., 2004) and HS⁻ production is typically slower here than in
287 surface layers (Teal et al., 2009), which may lead to accumulation of HS⁻, but at depths
288 greater than those studied here. The relatively low concentrations of solid sulfide phases and
289 the presence of iron (oxyhydr)oxides at depth in the core indicate this sediment is reactive
290 iron, rather than sulfur, dominated. In sediments where organic carbon supply is greater, and
291 sulphate reduction is enhanced, greater concentrations of sulfides would be expected
292 (Devereux et al., 2015).

293
294 Leachable Mn phases increase gradually above the aRPD (Figure 3c). The reagents with
295 strongest reducing action (dithionite) released most Mn, and constituted on average 41.3% of
296 the total values. These Mn reducible forms are therefore an important fraction of the total Mn
297 present. The increase in these fractions above the aRPD is consistent with trapping of
298 dissolved Mn as the more oxic surface conditions are approached, and biological mixing
299 down of surface formed Mn oxides (Thamdrup et al., 1994).

300

301 **3.4 Linking solid phases and processes within the core to colour**

302

303 The pure forms of the major constituents in the core (carbonate, illite, quartz) have no
304 significant colour in the visible spectrum, and therefore act as a white “canvas” against which
305 the colour from minor mineral components can be more readily seen. Sands that are

306 predominantly quartz may have a reddish hue due to surface Fe oxides that will be removed
307 by the analytical procedures used here, and will thus be observed in the oxide fraction.

308

309 The main coloured minerals expected in this sediment matrix are iron (oxyhydr)oxides, and
310 iron sulfides. The primary black coloured sulfide phase is expected to be an amorphous
311 mono-sulfide that gives the colour to hydrothermal “black smokers”, and this mineral will be
312 measured by the AVS technique. Although pyrite is more abundant than the AVS liberated
313 sulfides, its colour is typically a muted gold, and can be found in framboidal crystalline forms
314 (Figure 5). The main concentration changes for these minerals are around the aRPD depth at
315 7.8 cm (Figure 4). Above this depth, the dithionite Fe phases (expected to be mainly goethite
316 and haematite) increase by ~27% to the surface whilst below, pyrite and AVS increase by
317 ~25%.

318

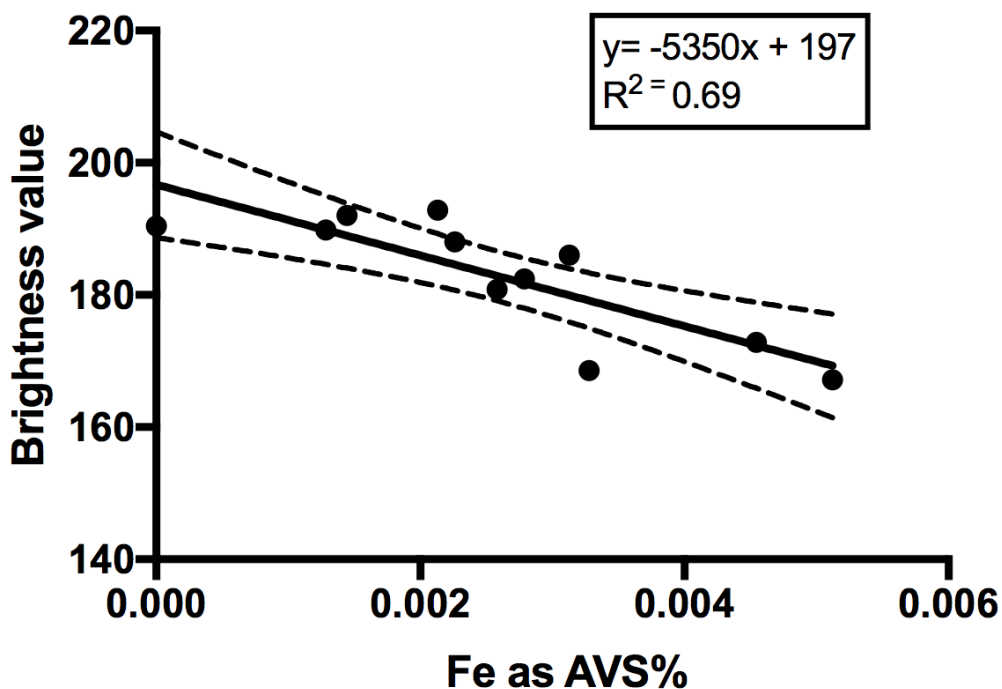
319 In order to link these geochemical observations with core colour requires an appropriate
320 methodological approach. The most useful starting point was the work of Bull and
321 Williamson (2001a) who investigated correlations between “amorphous” iron oxides and acid
322 volatile sulfur and colour properties on core samples collected from an estuarine system;
323 image analysis was used to generate colour intensity and colour saturation to compare,
324 respectively, to AVS and iron (oxyhydr)oxide concentrations. The underlying rationale was
325 to provide an image based estimate of the binding capacity of the sediment for toxic metals
326 such as Cu and Zn. Indeed, the need for such innovations that may improve the quantification
327 of metal source-sink relationships in marine sediments have been identified (Homoky et al.,
328 2016). A related image analysis approach was taken here.

329

330 When brightness is plotted against AVS (Figure 6), a reasonable negative correlation is seen,
331 despite the shelf core having a smaller range of intensity values than the estuarine core of
332 Bull and Williamson (Bull and Williamson, 2001a). The surface-most sample that has
333 relatively high values for both AVS and pyrite appears an outlier (perhaps because of highly
334 reactive forms of iron (oxyhydr)oxides present in this surface layer, and the higher
335 concentration of organic carbon fuelling sulfate reduction) and is not included in the main
336 correlation shown. However, the data does indicate that in this sediment type an estimate of
337 AVS should be possible from SPI images.

338

339 Figure 6. Acid volatile sulfur relative to core HSB brightness. Dotted lines are 95%
340 confidence limits. Surface point omitted as appears an outlier.



341

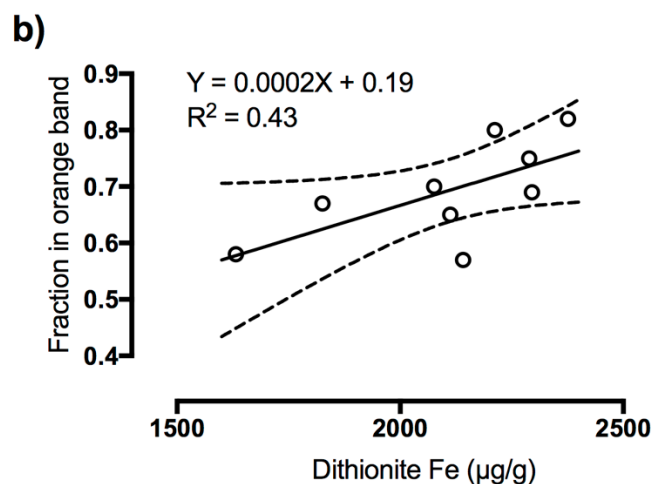
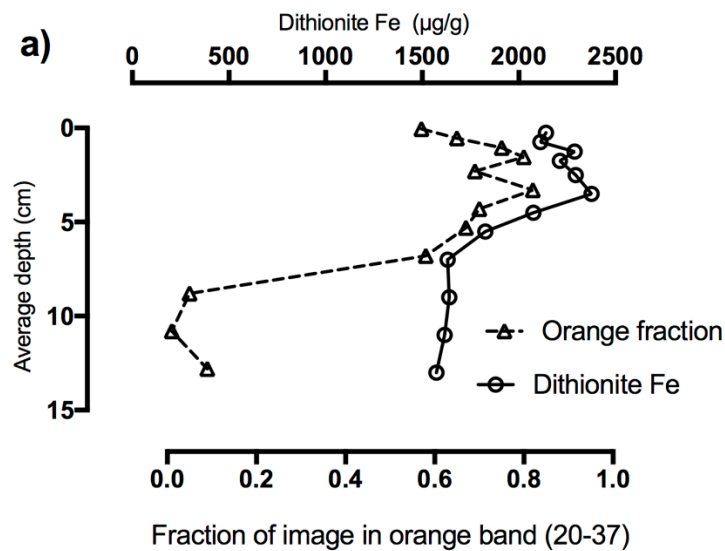
342

343 For the iron (oxyhydr)oxides (dithionite-citrate leach) there is a modest correlation with the
344 fraction of an image identified as orange in the zone above the aRPD (Figure 7). Below the
345 aRPD there is no observable orange colour, presumably reflecting the formation of FeS at the

346 surface of oxides that obscures their colour. The relationship of leachable Fe with colour will
 347 depend on: 1) mineralogy of oxide phases with intermediate and mixed forms showing
 348 different orange colours, and 2) the colour chosen for the image analysis (there was a shift to
 349 a more yellow colour with depth). Given these limitations it is unsurprising that only a weak
 350 correlation was observed. However, the general trends in colour relative to the Fe phases
 351 measured in the upper part of the core, are clear (Figure 7).

352

353 Figure 7. a) Fraction of image in orange band and Fe dithionite leach data, with depth; b)
 354 correlation of orange fraction with dithionite leach Fe in zone above aRPD.



355

356 The colour of complex mixtures of mineral phases is difficult to anticipate, but here where it
357 is assumed only two main coloured phases are present some success is achieved. Many
358 sediments will be more complex, e.g. increased concentrations of sulfide phases caused by
359 higher organic carbon content, and Mn rich systems where Mn oxides will impact colour.
360 The illite here is relatively colourless but, reduced Fe incorporated into montmorillonite is
361 reported to impart a green colour in reducing zones of open ocean sediments (Lyle, 1983),
362 and if montmorillonite is present it may complicate interpretation.

363

364 A variety of factors in addition to those directly impacting colour may complicate
365 interpretation in other shelf and coastal systems. Frequently, more complex structure than in
366 the core discussed here may be seen, with, for example, localised zones of organic matter
367 decomposition leading to intense carbon turnover that generates a halo effect of compressed
368 redox zones. This zone is often visually manifested as a black colouration resulting from
369 sulfate reduction and production of iron sulfide phases. Burrow structures that penetrate into
370 reducing sediments and transport oxygen can also produce compressed redox features in their
371 walls (Forster and Graf, 1992). Additionally, physical disturbances, including tidal mixing
372 and trawling can perturb sediment structure and redox zones. However, the general concept
373 of colour reflecting changes in the relative importance of the main coloured mineral phases
374 within the sediment should still apply, even under these conditions where heterogeneity of
375 the colour zones is increased.

376

377 **4. Conclusions**

378

379 The aRPD in the SPI image represents a significant visible change in the concentrations of
380 the iron (oxyhydr)oxides and iron sulfide phases in the core, as has been shown by detailed

381 analyses of the core solid phases and pore waters. Whilst both oxides and sulfur phases are
382 present throughout the measured core depths the decrease in one and increase in the other
383 leads to the net change in colour around the aRPD. It is important to note that in the shelf
384 sediments studied here there is no obvious correlation between oxygen penetration and
385 aRPD, and caution is needed in attempting to correlate colour changes with the depth to
386 which oxygen penetrates (Gerwing et al., 2015).

387

388 The aRPD is a key component of the metrics that are increasingly used in, or under
389 consideration for, management frameworks to assess seabed environmental health. These
390 multi-metric indices include the OSI or Organism Sediment Index (Rhoads and Germano,
391 1986) and BHQ or Benthic Habitat Quality index (Nilsson and Rosenberg, 1997) and they or
392 derivatives are finding use in, for example, the EU Water Framework Directive and Marine
393 Strategy Framework Directive (Borja et al., 2008). The work reported here will help
394 understand the aRPD so that its variability within these metrics or used alone (Teal et al.,
395 2010), and its relevance to biological communities, biogeochemical cycles and benthic
396 ecosystem health will be better understood. Indeed, whilst the uncertainty around the nature
397 of the aRPD remains, caution should be applied in using indices derived from it across
398 regions where variability in factors impacting the aRPD are not fully understood. As
399 Germano et al. recommended (2011) combining aRPD with other parameters in multivariate
400 analyses might be more appropriate than aRPD derived indices. Further biogeochemical
401 ground-truthing of the aRPD in other types of sedimentary systems with different sediment
402 fabric, particle sizes and biological communities will inform and potentially extend its use
403 further. It seems that the aRPD alone is something of a blunt tool for assessing ecosystem
404 health and combination with other parameters is an important way ahead.

405

406 The correlation between AVS and SPI image colour may prove useful if estimates of
407 sediment carrying capacity for toxic metals, e.g. Cu and Zn as in Bull and Williamson (2001),
408 and potentially other chalcophile elements, can be obtained from SPI spatial mapping within
409 disposal sites. More work on the relative stability constants of these heavy metal sulfides and
410 their formation in such sedimentary systems will be needed in this application. For more
411 detailed geochemical interpretation of routine SPI images, development and application of
412 new sensors and devices attached to SPI during deployment (e.g. oxygen penetration depth,
413 pH, Fe/Mn, S), are needed so that the range of parameters used in assessment of seabed
414 health can be extended.

415

416 **5. Acknowledgements**

417 This project was funded through Work Package 3 of the UK Shelf Sea Biogeochemistry
418 Programme (NE/K001973/1 and NE/K001787/1), jointly funded by the Natural
419 Environmental Research Council (NERC) and the Department for Environment, Food and
420 Rural Affairs (Defra). The views expressed are those of the authors and do not necessarily
421 represent those of NERC or Defra. W.B.H. was further supported by a NERC Fellowship
422 (NE/K009532/1). The samples for this project were collected with the excellent support of
423 captains, crew and NMF staff on the RRS *Discovery*. We are particularly grateful to fellow
424 researchers within the SSB programme for providing assistance at sea. We express special
425 thanks to Carolyn Harris for analysing nutrient samples during cruise DY034 and Malcolm
426 Woodward for collating these data, Ross Williams for XRD work, David Chatelet (Biological
427 imaging unit, University of Southampton) for help with ImageJ, Shir Akbari for POC/PIC
428 analysis, and Matt Cooper for ICP-OES analyses. We are grateful for the useful comments
429 provided by 2 anonymous reviewers that improved the manuscript.

430

431 **6. References**

432 Albarede, F., 2003. Geochemistry- an introduction. Cambridge University Press.

433 Bertics, V.J., Ziebis, W., 2009. Biodiversity of benthic microbial communities in bioturbated
434 coastal sediments is controlled by geochemical microniches. *Isme Journal* 3, 1269-1285.

- 435 Borja, A., Dauer, D.M., Diaz, R., Llanso, R.J., Muxika, I., Rodriguez, J.G., Schaffner, L.,
436 2008. Assessing estuarine benthic quality conditions in Chesapeake Bay: A comparison of
437 three indices. *Ecological Indicators* 8, 331-337.
- 438 Bull, D.C., Williamson, R.B., 2001a. Prediction of principal metal-binding solid phases in
439 estuarine sediments from color image analysis. *Environ. Sci. Technol* 35, 1658–1662.
- 440 Bull, D.C., Williamson, R.B., 2001b. Prediction of principal metal-binding solid phases in
441 estuarine sediments from color image analysis. *Environ. Sci. Technol* 35, 1658–1662.
- 442 Burdige, D.J., 2006. *Geochemistry of marine sediments*. Princeton University Press,
443 Princeton, NJ.
- 444 Canfield, D.E., Raiswell, R., Bottrell, S., 1992. The reactivity of sedimentary iron minerals
445 toward sulfide. *American Journal of Science* 292, 659-683.
- 446 Canfield, D.E., Raiswell, R., Westrich, J.T., Reaves, C.M., Berner, R.A., 1986. The use of
447 chromium reduction in the analysis of reduced inorganic sulfur in sediments and shales.
448 *Chemical Geology* 54, 149-155.
- 449 Devereux, R., Lehrter, J.C., Beddick, D.L., Yates, D.F., Jarvis, B.M., 2015. Manganese, iron,
450 and sulfur cycling in Louisiana continental shelf sediments. *Continental Shelf Research* 99,
451 46-56.
- 452 Fenchel, T., 1969. The ecology of marine microbenthos. IV. Structure and function of the
453 benthic ecosystem, its chemical and physical factors and microfauna communities with
454 special reference to the ciliated Protozoa. *Ophelia* 6, 1-182.
- 455 Forster, S., Graf, G., 1992. Continuously measured changes in redox potential influenced by
456 oxygen penetrating from burrows of callianassa-subterranea. *Hydrobiologia* 235, 527-532.
- 457 Germano, J.D., Rhoads, D.C., Valente, R.M., Carey, D.A., Solan, M., 2011. The use of
458 sediment profile imaging (spi) for environmental impact assessments and monitoring studies:
459 lessons learned from the past four decades, in: Gibson, R.N., Atkinson, R.J.A., Gordon,
460 J.D.M. (Eds.), *Oceanography and Marine Biology: An Annual Review*, Vol 49, pp. 235-297.
- 461 Gerwing, T.G., Gerwing, A.M.A., Hamilton, D.J., Barbeau, M.A., 2015. Apparent redox
462 potential discontinuity (aRPD) depth as a relative measure of sediment oxygen content and
463 habitat quality. *International Journal of Sediment Research* 30, 74-80.
- 464 Grizzle, R.E., Penniman, C.A., 1991. Effects of organic enrichment on estuarine macrofaunal
465 benthos - a comparison of sediment profile imaging and traditional methods. *Marine Ecology*
466 *Progress Series* 74, 249-262.
- 467 Klar, J.K., Homoky, W.B., Statham, P.J., Birchill, A.J., Harris, E.L., Woodward, E.M.S.,
468 Silburn, B., Cooper, M.J., James, R.H., Connelly, D.P., Chever, F., Lichtschlag, A., Graves,
469 C., 2017. Stability of dissolved and soluble Fe(II) in shelf sediment pore waters and release to
470 an oxic water column. *Biogeochemistry*.
- 471 Lyle, M., 1983. The brown green color transition in marine-sediments - a marker of the
472 Fe(iii)-Fe(ii) redox boundary. *Limnology and Oceanography* 28, 1026-1033.

- 473 Nieuwenhuize, J., Maas, Y.E.M., Middelburg, J.J., 1994. Rapid analysis of organic-carbon
474 and nitrogen in particulate materials. *Marine Chemistry* 45, 217-224.
- 475 Nilsson, H.C., Rosenberg, R., 1997. Benthic habitat quality assessment of an oxygen stressed
476 fjord by surface and sediment profile images. *Journal of Marine Systems* 11, 249-264.
- 477 Poulton, S.W., 2003. Sulfide oxidation and iron dissolution kinetics during the reaction of
478 dissolved sulfide with ferrihydrite. *Chemical Geology* 202, 79-94.
- 479 Poulton, S.W., Canfield, D.E., 2005. Development of a sequential extraction procedure for
480 iron: implications for iron partitioning in continentally derived particulates. *Chemical*
481 *Geology* 214, 209-221.
- 482 Poulton, S.W., Krom, M.D., Raiswell, R., 2004. A revised scheme for the reactivity of iron
483 (oxyhydr)oxide minerals towards dissolved sulfide. *Geochimica Et Cosmochimica Acta* 68,
484 3703-3715.
- 485 Raiswell, R., Canfield, D.E., Berner, R.A., 1994. A comparison of iron extraction methods
486 for the determination of degree of pyritisation and the recognition of iron-limited pyrite
487 formation. *Chemical Geology* 111, 101-110.
- 488 Raiswell, R., Vu, H.P., Brinza, L., Benning, L.G., 2010. The determination of labile Fe in
489 ferrihydrite by ascorbic acid extraction: Methodology, dissolution kinetics and loss of
490 solubility with age and de-watering. *Chemical Geology* 278, 70-79.
- 491 Rhoads, D.C., Cande, S., 1971. Sediment profile camera for in-situ study of organism-
492 sediment relations. *Limnology and Oceanography* 16, 110-&.
- 493 Rhoads, D.C., Germano, J.D., 1986. Interpreting long-term changes in benthic community
494 structure: a new protocol. *Hydrobiologia* 142, 291-308.
- 495 Rosenberg, R., Blomqvist, M., Nilsson, H.C., Cederwall, H., Dimming, A., 2004. Marine
496 quality assessment by use of benthic species-abundance distributions: a proposed new
497 protocol within the European Union Water Framework Directive. *Marine Pollution Bulletin*
498 49, 728-739.
- 499 Seeberg-Elverfeldt, J., Schlüter, M., Feseker, T., Kölling, M., 2005. Rhizon sampling of
500 porewaters near the sediment-water interface of aquatic systems. *Limnol Oceanogr.: Methods*
501 3, 361-371.
- 502 Solan, M., Germano, J.D., Rhoads, D.C., Smith, C., Michaud, E., Parry, D., Wenzhofer, F.,
503 Kennedy, B., Henriques, C., Battle, E., Carey, D., Iocco, L., Valente, R., Watson, J.,
504 Rosenberg, R., 2003. Towards a greater understanding of pattern, scale and process in marine
505 benthic systems: a picture is worth a thousand worms. *Journal of Experimental Marine*
506 *Biology and Ecology* 285, 313-338.
- 507 Solan, M., Wigham, B.D., Hudson, I.R., Kennedy, R., Coulon, C.H., Norling, K., Nilsson,
508 H.C., Rosenberg, R., 2004. In situ quantification of bioturbation using time-lapse fluorescent
509 sediment profile imaging (f-SPI), luminophore tracers and model simulation. *Marine Ecology*
510 *Progress Series* 271, 1-12.

- 511 Stookey, L.L., 1970. Ferrozine- a new spectrophotometric reagent for iron. *Analytical*
512 *Chemistry* 42, 779-781.
- 513 Teal, L.R., Parker, E.R., Solan, M., 2010. Sediment mixed layer as a proxy for benthic
514 ecosystem process and function. *Marine Ecology Progress Series* 414, 27-40.
- 515 Teal, L.R., Parker, E.R., Solan, M., 2013. Coupling bioturbation activity to metal (Fe and
516 Mn) profiles in situ. *Biogeosciences* 10, 2365-2378.
- 517 Teal, L.R., Parker, R., Fones, G., Solan, M., 2009. Simultaneous determination of in situ
518 vertical transitions of color, pore-water metals, and visualization of infaunal activity in
519 marine sediments. *Limnology and Oceanography* 54, 1801-1810.
- 520 Thamdrup, B., Fossing, H., Jorgensen, B.B., 1994. Manganese, iron, and sulfur cycling in a
521 coastal marine sediment, Aarhus Bay, Denmark. *Geochimica Et Cosmochimica Acta* 58,
522 5115-5129.
- 523 Thompson, C.E.L., Silburn, B., Williams, M.E., Hull, T., Sivyer, D., Amoudry, L.O.,
524 Widdicombe, S., Ingels, J., Carnovale, G., McNeill, C.L., Hale, R., Marchais, C.L., Hicks,
525 N., Smith, H.E.K., Klar, J.K., Hiddink, J.G., Kowalik, J., Kitidis, V., Reynolds, S.,
526 Woodward, E.M.S., Tait, K., Homoky, W.B., Kröger, S., Bolam, S., Godbold, J.A., Aldridge,
527 J., Mayor, D.J., Benoist, N.M.A., Bett, B.J., Morris, K.J., Parker, E.R., Ruhl, H.A., Statham,
528 P.J., Solan, M., 2017. An approach for the identification of exemplar sites for scaling up
529 targeted field observations of benthic biogeochemistry in heterogeneous environments.
530 *Biogeochemistry*.
- 531 Viollier, E., Inglett, P.W., Hunter, K., Roychoudhury, A.N., Van Capellen, P., 2000. The
532 ferrozine method revisited: Fe(II)/Fe(III) determination in natural waters. *Applied*
533 *Geochemistry* 15, 785-790.
- 534 Vismann, B., 1991. Sulfide tolerance - physiological-mechanisms and ecological
535 implications. *Ophelia* 34, 1-27.
- 536 Woodward, E.M.S., Rees, A.P., 2001. Nutrient distributions in an anticyclonic eddy in the
537 northeast Atlantic Ocean, with reference to nanomolar ammonium concentrations. *Deep-Sea*
538 *Research Part II-Topical Studies in Oceanography* 48, 775-793.
539
- 540 Supplementary Material
- 541 1. TIFF version of core image (not included here because of file size).
- 542 2. Data table (this will be an Excel spreadsheet).

CSR Geochemical investigation of a digitally imaged shelf core, and implications for interpretation of sediment colour; Statham et al.

SUPPLEMENTARY MATERIAL, Data Table

Cruise DY034, August 2015. Site A sample 467

See main text and Klar et al. (2017) for details of analytical methods used, and precision and detection limit estimates

ND= No data, problem during analysis

Space= no analysis done

SOLID PHASE Unless otherwise shown units are µg/g dry weight

<i>Average Depth (cm)</i>	<i>% water</i>	<i>POC (%)</i>	<i>Carbonate (%)</i>	<i>Total Fe</i>	<i>Ascorbic Fe</i>	<i>Dithi o Fe</i>	<i>Fraction of image in orange band</i>	<i>Total Mn</i>	<i>Ascorbic Mn</i>	<i>dithio Mn</i>	<i>Fe AVS %</i>	<i>Brightness value</i>	<i>Fe PY %</i>
0.25	40.8			13970	300	2226		305	14	140			
0.25	40.7			14250	290	2100		316	15	133			
0.25	40.8	1.20	36.00	14090	274	2141	0.57	312	15	137	0.008	189.4	0.033
0.75	40.1	1.08	36.72	14180	252	2112	0.65	316	16	128	0.002	192.8	0.028
1.25	40.4	1.07	36.86	14820	302	2289	0.75	335	20	145	0.000	190.4	0.028
1.75	39.9	0.94	38.42	14500	274	2212	0.80	325	18	142	0.001	192	0.030
2.5	40.7	ND	ND	14890	274	2295	0.69	320	12	135	0.001	189.8	0.031
2.5	41.4				268	2298			13	134			
3.5	38.7	1.25	35.21	14660	286	2376	0.82	307	9	132	0.002	188	0.029
4.5	37.7	1.08	36.50	14500	280	2076	0.70	300	7	123	0.003	186	0.032
5.5	36.8	1.04	35.87	13750	244	1826	0.67	292	5	111	0.003	182.4	0.030
7	37.4	1.01	35.91	13350	258	1632	0.58	284	5	111	0.003	180.8	ND
9	36.3	0.97	36.12	13390	260	1640	0.05	288	5	110	0.005	172.8	0.043
11	38.2	0.92	37.08	13810	249	1616	0.01	292	5	110	0.003	168.6	0.039
13	38.2	1.00	37.35	13440	291	1573	0.09	289	6	112	0.005	167.2	ND

Core Interval	Bulk Mineralogy (%)								
	Plag. Feldspar	Aragonite	Calcite	Chlorite	Dolomite	Halite	Illite	Quartz	TOTAL
2-3cm	4.8	13.7	15.2	1.4	1.2	1.7	29.5	30.9	98.4
5-6cm	5.6	15.3	16.2	1.7	1	0.5	29.8	35.7	105.8
8-10cm	5	14.7	15.2	1.5	1.2	0.6	28.7	35.1	102

POREWATER								OXYGEN and aRPD depth		
Av Depth (cm)	Nitrite (μM)	Nitrate (μM)	Ammonium (μM)	Silicon (μM)	Phosphate (μM)	dFe II (μM)	dFe (μM)	Oxygen penetration depth (OPD) in mm across face of core		
0.5	0.01	6.27	6.5	85.1	1.7	0.20	0.21	Profile #	OPD	
1.5	0.04	1.61	20.2	130	2.5	1.30	1.32	1	3.9	
2.5	0.07	1.18	35.4	174	6.9	24	24	2	4.6	
3.5	0.15	1.02	42.3	195	10.3	49	51	3	4.6	
5.5	0.32	0.46	55.4	250	25.4	107	107	4	4.4	
7.5	0.41	0.63	64.6	294	32.7	109	111	5	2.9	
9.5	0.44	0.82	80.4	334	38.5	131	134			
11.5	0.57	0.09	92.2	347	44.2	137	141	aRPD	78	mm

Note that nitrate plus nitrite data are close to detection level below surface