



Deposited via The University of Sheffield.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/123061/>

Version: Accepted Version

Proceedings Paper:

Hadley, L., Colley, H.E. and Murdoch, C. (2016) Development of macrophage-containing in vitro 3D models of oral cancer. In: Oral Diseases. 13th Biennial Congress of the European Association of Oral Medicine, 15-17 Sep 2016, Torino, Italy. Wiley, pp. 10-11. ISSN: 1354-523X. EISSN: 1601-0825.

<https://doi.org/10.1111/odi.12558>

This is the peer reviewed version of the following article: Hadley, L. et al, Development of macrophage-containing in vitro 3D models of oral cancer, Oral Diseases, 22 (S2), 10-11, which has been published in final form at <https://doi.org/10.1111/odi.12558>. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

Development of a macrophage-containing in-vitro 3D model of oral cancer

L. Hadley, H.E. Colley, C. Murdoch

School of Clinical Dentistry, The University of Sheffield, Sheffield, S10 2TA, UK

Objectives: Tumour-associated macrophages (TAM) represent a prominent component of the leukocytic infiltrate of human tumours and their accumulation in oral squamous cell carcinoma (OSCC) has been shown to be a predictor of poor prognosis. Evidence suggests that the tumour microenvironment drives TAM into a pro-tumour phenotype that exacerbates tumour growth but evidence for this in OSCC is lacking. In this study, we further develop a 3D *in vitro* model of OSCC to incorporate human macrophages to be used as a tool to examine the role of TAM in OSCC.

Methods: Human monocyte-derived macrophages were cultured with human oral fibroblasts within a type 1 collagen hydrogel with either human normal oral keratinocytes or OSCC cell lines seeded on top and cultured at an air-to-liquid interface. After 14 days, key features of the tumour microenvironment were measured using ELISA, immunohistochemistry and rheology. Macrophages were retrieved from the connective tissue by collagenase digestion and analysed using qPCR and 8-colour flow cytometry.

Results: Macrophages were viable and functional after 14-day 3D culture, with IL-6 and CXCL8 release observed upon LPS stimulation. Comparison of a malignancy-free environment to an induced tumour microenvironment showed a positive correlation between OSCC cell invasion and matrix stiffness that was enhanced by the presence of macrophages. In addition, flow cytometric analysis showed a marked increase in the number of macrophages expressing key markers, such as CD163 in an induced cancer compared to normal environment.

Conclusions: Macrophages remain viable and responsive to exogenous stimuli even when cultured within a 3D *in vitro* model for prolonged periods. These immuno-responsive 3D models provide a new, reproducible and adaptable tool that are able to mimic the tumour microenvironment during OSCC and therefore will be of great use in gaining a deeper understanding of the role of TAM and the tumour stroma in OSCC progression.