



Deposited via The University of Leeds.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/121884/>

Version: Accepted Version

Article:

Abuarab, N, Li, F and Sivaprasadarao, A (2017) Ionic signalling and mitochondrial dynamics. *Molecular and Cellular Oncology*, 4 (6). e1373889. ISSN: 2372-3556

<https://doi.org/10.1080/23723556.2017.1373889>

© 2017 Taylor & Francis. This is an Accepted Manuscript of an article published by Taylor & Francis in *Molecular & Cellular Oncology* on 18 September 2017, available online: <http://www.tandfonline.com/10.1080/23723556.2017.1373889>. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

Ionic Signalling and Mitochondrial Dynamics

Nada Abuarab, Fangfang Li & Asipu Sivaprasadarao

To cite this article: Nada Abuarab, Fangfang Li & Asipu Sivaprasadarao (2017): Ionic Signalling and Mitochondrial Dynamics, *Molecular & Cellular Oncology*, DOI: [10.1080/23723556.2017.1373889](https://doi.org/10.1080/23723556.2017.1373889)

To link to this article: <http://dx.doi.org/10.1080/23723556.2017.1373889>



Accepted author version posted online: 18 Sep 2017.



Submit your article to this journal [↗](#)



View related articles [↗](#)



View Crossmark data [↗](#)

Ionic Signalling and Mitochondrial Dynamics

Nada Abuarab¹, Fangfang Li¹, and Asipu Sivaprasadarao^{1,2,*}

¹School of Biomedical Sciences, Faculty of Biological Sciences, ²Multidisciplinary Cardiovascular Research Centre, University of Leeds, LS2 9JT, Leeds, U.K.

Corresponding author: Professor Asipu Sivaprasadarao, School of Biomedical Sciences, G6.44d, Garstang Building, University of Leeds, Leeds, LS2 9JT UK, Tel: +44-(0)-113-3434326, Fax: +44-(0)-113-3431407, E-mail: a.sivaprasadarao@leeds.ac.uk

Abstract

In age-related diseases, rise in intracellular reactive oxygen species (ROS) causes fragmentation of mitochondrial network. Our recent study demonstrated that ROS activation of TRPM2 (transient receptor potential melastatin-2) channels triggers lysosomal Zn²⁺ release that, in turn, triggers mitochondrial fragmentation. The findings provide new mechanistic insights that may have therapeutic implications.

Keywords

Mitochondrial dynamics, TRPM2, oxidative stress, reactive oxygen species, calcium, zinc, lysosomal membrane permeabilisation, cell proliferation, cell migration

In a normal cell, mitochondria exist as a tubular network that undergoes continuous fission and fusion¹. Fission helps to eliminate dysfunctional parts of the network via mitophagy, whilst fusion allows merger of the functionally intact parts with the healthy mitochondrial network. These processes, collectively known as 'mitochondrial dynamics' ensure maintenance of a healthy network required for efficient energy production and mitochondrial signalling.

The molecular machinery required for mitochondrial dynamics is mostly known¹. Mitochondrial fission is initiated by the ER (endoplasmic reticulum)-mediated constriction of the mitochondrial tubule and recruitment of Drp1 (dynamin-related protein) from the cytoplasm onto the mitochondria via its receptors, mitochondrial fission factor (MFF) and mitochondrial dynamics protein 51 (MID51). Drp-1 molecules form a spiral around the ER-constricted fission site, and together with dynamin 2, cause mitochondrial fission. Mitofusin-1/2 (MFN-1/2) and OPA1 (Optic atrophy type 1) catalyse the fusion of the outer and inner membranes of mitochondria, respectively.

Mitochondrial dynamics is finely regulated, but this regulation goes awry in a wide range of seemingly unrelated human diseases, including cardiovascular (e.g. ischemia), neuronal (Parkinson's, Alzheimer's and stroke) and infectious (some) diseases and certain cancers^{1, 2}. A unifying feature of these diseases, however, is that many of them are age-related and are associated with an increased production of reactive oxygen species (ROS). ROS is a powerful stimulant of mitochondrial fission, but how ROS signal mitochondrial fission has remained unclear.

In our recent publication³, we demonstrated that ROS use the oxidative stress-sensitive TRPM2 channel to signal mitochondrial fragmentation. We used high glucose (diabetic) stress to stimulate ROS production and mitochondrial fragmentation in endothelial cells⁴. We found that chemical inhibition, RNAi-silencing and knock-out of TRPM2 channels prevented high glucose-induced mitochondrial fragmentation³. Given that Drp1 recruitment to mitochondria is Ca²⁺-dependent, the results presented no surprise. However, as TRPM2 activation can also rise cytosolic Zn²⁺, we tested the effect of Zn²⁺ chelators. The result was rather unexpected: Zn²⁺ chelators completely inhibited high glucose-induced mitochondrial fission. Follow-up studies led to the discovery of a novel signalling pathway involving an intriguing interplay between Ca²⁺ and Zn²⁺ in inter-organelle communication that ultimately leads to mitochondrial fission (Figure 1)³.

The first step in the signalling pathway was a rise in intracellular ROS by the diabetic stress. By stimulating the TRPM2 channel, ROS increased the cytosolic Ca²⁺. Rise in Ca²⁺ triggered lysosomal membrane permeabilisation (LMP), leading to the release of its contents, including free Zn²⁺. Inhibition of TRPM2 channels prevented LMP completely³. This result is interesting in itself because although it is known that Ca²⁺ plays a role in ROS-induced LMP, the signalling protein responsible for the Ca²⁺ rise was hitherto unknown. The finding could dispel the current notion that LMP is a nonspecific process and may have implications for lysosomal diseases⁵.

We found that the lysosomal Zn²⁺ release was accompanied an accumulation of free Zn²⁺ in mitochondria³. How Zn²⁺ escapes sequestration by the cytosolic buffers (e.g. metallothioneins) and enters mitochondria is unclear, but presence of lysosomes in the proximity of mitochondria and mitochondrial membrane transport mechanisms might be important. Rise in mitochondrial Zn²⁺ led to the recruitment of cytoplasmic Drp1 onto mitochondria and the consequent mitochondrial fragmentation. Drp1 recruitment to mitochondria requires depolarisation of mitochondrial membrane potential ($\Delta\psi_{mt}$) and is regulated by multiple posttranslational modifications¹. Zn²⁺ could cause a loss of $\Delta\psi_{mt}$ by virtue of its ability to inhibit the mitochondrial electron transport chain⁶. However, what effect Zn²⁺ might have on the posttranslational modifications to promote mitochondrial Drp1 recruitment remains to be established. Regardless of the mechanisms, our study demonstrated that Zn²⁺ is ultimately responsible for mitochondrial fragmentation.

Excessive mitochondrial fragmentation is generally associated with apoptotic cell death. Previous studies suggested that LMP leads to mitochondria-mediated intrinsic apoptosis. Proteolytic actions of cathepsins released during LMP are thought to activate the pro-apoptotic bcl-2 (B-cell lymphoma 2) pathway to induce intrinsic apoptosis⁵. Given our finding that Zn²⁺ induces mitochondrial fragmentation, and its previously known role in apoptosis⁷, it seems reasonable to suggest that Zn²⁺ might carry the apoptotic signal from lysosomes to mitochondria.

Mitochondrial fission is associated with cancer cell proliferation and migration^{1, 2}. Certain cancer cells display fragmented mitochondria due to increased expression or activation of Drp1, coupled with the downregulation of MFN-2. Importantly, reducing the mitochondrial fragmentation through Drp1 inhibition or MFN2 overexpression has been shown to inhibit cell proliferation^{1, 2}. Notably, growth factor stimulation of K-Ras (Kirsten ras oncogene) increases ERK (Extracellular Signal-regulated Kinase)-2-mediated Drp-1 phosphorylation, mitochondrial Drp1 recruitment and fragmentation leading to tumour growth¹. As cancer cells often contain increased levels of ROS, it is reasonable to speculate a role for ROS-activated, TRPM2-mediated Zn²⁺ signalling in cancer cell proliferation. Supporting this idea, TRPM2 is upregulated in several cancers and its inhibition prevented prostate cancer (PC-3) cell proliferation⁸. Mitochondrial fission is also associated with cancer cell migration where fragmented mitochondria move towards the leading edge (lamellipodia) of migrating cells, which again was prevented by Drp1 inhibition⁹. Whether TRPM2-mediated Zn²⁺ signalling plays a role in this process is not known, but we have demonstrated that TRPM2 inhibition as well as Zn²⁺ chelation inhibits PC-3 and HeLa cell migration¹⁰.

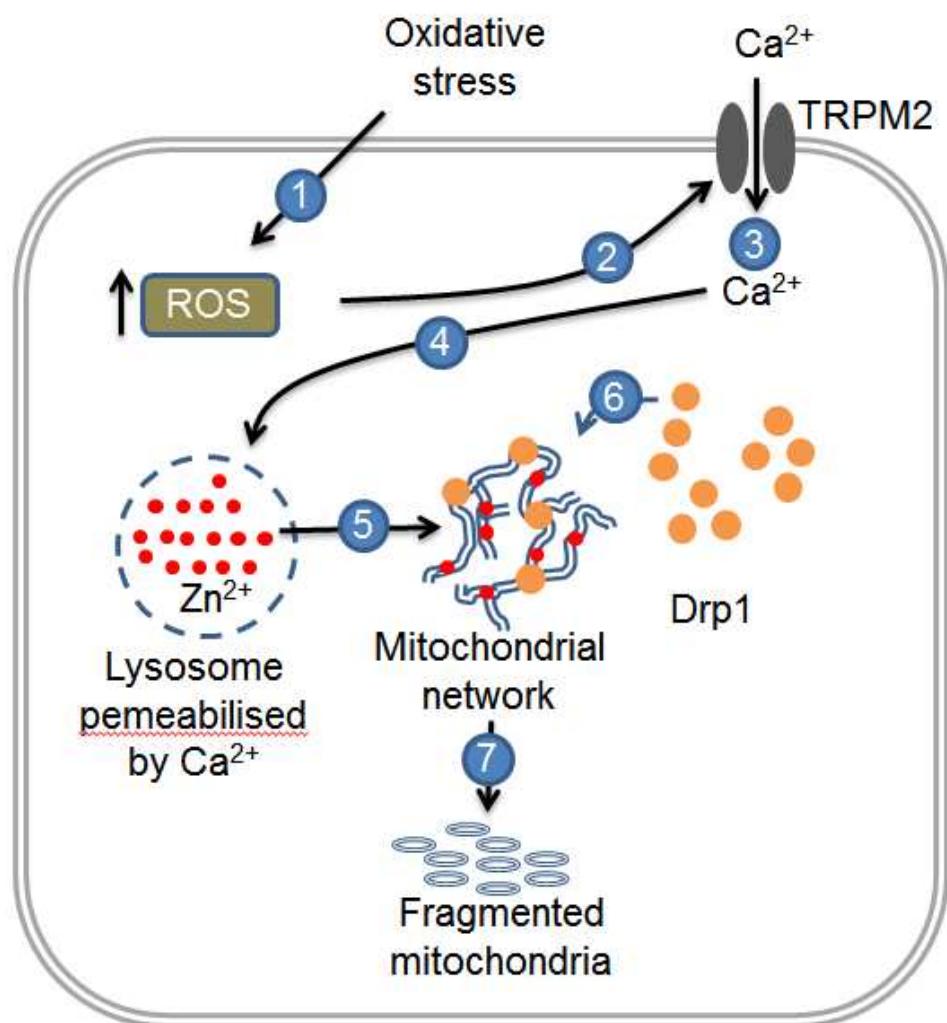
In conclusion, the stress signalling pathway identified in our study has provided new fundamental knowledge on how ionic signalling facilitates inter-organelle communication to drive mitochondrial fragmentation. The results have implications for a number of human diseases including cancer, and may have therapeutic potential.

Acknowledgements

This work was supported by the British Heart Foundation (PG/10/68/28528) and Ministry of Higher Education, Saudi Arabia.

References

1. Chen H, Chan DC. Mitochondrial Dynamics in Regulating the Unique Phenotypes of Cancer and Stem Cells. *Cell Metab* 2017; 26: 39-48.
2. Archer SL. Mitochondrial dynamics--mitochondrial fission and fusion in human diseases. *N Engl J Med* 2013; 369: 2236-2251.
3. Abuarab N, Munsey TS, Jiang LH, Li J, Sivaprasadarao A. High glucose-induced ROS activates TRPM2 to trigger lysosomal membrane permeabilization and Zn²⁺-mediated mitochondrial fission. *Sci Signal* 2017; 10(490).
4. Shenouda SM, Widlansky ME, Chen K, Xu G, Holbrook M, Tabit CE, et al. Altered mitochondrial dynamics contributes to endothelial dysfunction in diabetes mellitus. *Circulation* 2011; 124: 444-453.
5. Serrano-Puebla A, Boya P. Lysosomal membrane permeabilization in cell death: new evidence and implications for health and disease. *Ann N Y Acad Sci* 2016; 1371: 30-44.
6. Sensi SL, Paoletti P, Bush AI, Sekler I. Zinc in the physiology and pathology of the CNS. *Nat Rev Neurosci* 2009; 10: 780-791.
7. Manna PT, Munsey TS, Abuarab N, Li F, Asipu A, Howell G, et al. TRPM2 mediated intracellular Zn²⁺ release triggers pancreatic beta cell death. *The Biochemical Journal* 2015; 466: 537-546.
8. Zeng X, Sikka SC, Huang L, Sun C, Xu C, Jia D, et al. Novel role for the transient receptor potential channel TRPM2 in prostate cancer cell proliferation. *Prostate Cancer Prostatic Dis* 2010; 13: 195-201.
9. Zhao J, Zhang J, Yu M, Xie Y, Huang Y, Wolff DW, et al. Mitochondrial dynamics regulates migration and invasion of breast cancer cells. *Oncogene* 2013; 32: 4814-4824.
10. Li F, Abuarab N, Sivaprasadarao A. Reciprocal regulation of actin cytoskeleton remodelling and cell migration by Ca²⁺ and Zn²⁺: role of TRPM2 channels. *Journal of Cell Science* 2016; 129: 2016-2029.



Schematic of how diabetic stress induces mitochondrial fragmentation in endothelial cells. High glucose induces ROS (reactive oxygen species) production (1) leading to TRPM2 (transient receptor potential melastatin-2) activation (2), extracellular Ca^{2+} entry (3), Ca^{2+} -induced lysosomal membrane permeabilisation (4), Zn^{2+} (red dots) transfer from lysosomes to mitochondria (5), Zn^{2+} -induced Drp1 (dynamamin-related protein-1, orange filled circles) recruitment to mitochondria (6) and finally, mitochondrial fragmentation (7).