



UNIVERSITY OF LEEDS

This is a repository copy of *Inhibition of histone deacetylase 1 or 2 reduces induced cytokine expression in microglia through a protein synthesis independent mechanism*.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/119679/>

Version: Accepted Version

Article:

Durham, BS, Grigg, R and Wood, IC orcid.org/0000-0003-1886-2533 (2017) Inhibition of histone deacetylase 1 or 2 reduces induced cytokine expression in microglia through a protein synthesis independent mechanism. *Journal of Neurochemistry*, 143 (2). pp. 214-224. ISSN 0022-3042

<https://doi.org/10.1111/jnc.14144>

© 2017 International Society for Neurochemistry. This is the peer reviewed version of the following article: Durham, B. S., Grigg, R. and Wood, I. C. (2017), Inhibition of histone deacetylase 1 or 2 reduces induced cytokine expression in microglia through a protein synthesis independent mechanism. *Journal of Neurochemistry*. doi:10.1111/jnc.14144, which has been published in final form at <https://doi.org/10.1111/jnc.14144>. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Inhibition of histone deacetylase 1 or 2 reduces induced cytokine expression in microglia through a protein synthesis independent mechanism.

Benjamin S. Durham^{1,3}, Ronald Grigg² and Ian C. Wood¹

¹School of Biomedical Sciences, and ² School of Chemistry, University of Leeds, Leeds LS2 9JT, UK. ³Current address: Department of Infection, Immunity and Cardiovascular Disease, University of Sheffield, Sheffield, S10 2RX

To whom correspondence should be addressed: Ian C Wood, Tel +44 (0)113 3437922. Fax +44 (0)113 3431407. E-mail: i.c.wood@leeds.ac.uk

Running title. HDAC1 and 2 regulate microglia activation

Number of pages: 27

Number of figures: 4

Conflict of Interest: Ian Wood is a handling editor of the Journal of Neurochemistry

Acknowledgements: The work was supported by a Biotechnology and Biological Sciences Research Council (BBSRC) studentship awarded to BSD. The authors declare no competing financial interests.

Abbreviations used

DMEM - Dulbecco's Modified Eagle's Medium

DMSO – Dimethyl sulphoxide

ECL – Enhanced chemiluminescence

EDTA – Ethylenediaminetetraacetic acid

ELISA – Enzyme-linked immunosorbent assay

FBS – Foetal bovine serum

HDAC – Histone deacetylase

IFN- γ – Interferon gamma

IL-6 – Interleukin 6

LPS – Lipopolysaccharide

MAPK – Microtubule associated protein kinase

PBS – Phosphate buffered saline

qPCR – Quantitative polymerase chain reaction

SDS-PAGE – Sodium dodecyl sulphate polyacrylamide gel electrophoresis

VPA – Valproic acid

Abstract

Histone deacetylase (HDAC) inhibitors prevent neural cell death in in vivo models of cerebral ischaemia, brain injury and neurodegenerative disease. One mechanism by which HDAC inhibitors may do this is by suppressing the excessive inflammatory response of chronically activated microglia. However, the molecular mechanisms underlying this anti-inflammatory effect and the specific HDAC responsible are not fully understood. Recent data from in vivo rodent studies has shown that inhibition of class I HDACs suppresses neuroinflammation and is neuroprotective. In our study we have identified that selective HDAC inhibition with inhibitors apicidin, MS-275 or MI-192, or specific knockdown of HDAC1 or 2 using siRNA, suppresses the expression of cytokines interleukin-6 (IL-6) and tumour necrosis factor-alpha (TNF- α) in BV2 murine microglia activated with lipopolysaccharide (LPS). Furthermore, we found that in the absence of HDAC1, HDAC2 is upregulated and these increased levels are compensatory, suggesting these two HDACs have redundancy in regulating the inflammatory response of microglia. Investigating the possible underlying anti-inflammatory mechanisms suggests an increase in protein expression is not important. Taken together, this study supports the idea that inhibitors selective towards HDAC1 or HDAC2, may be therapeutically useful for targeting neuroinflammation in brain injuries and neurodegenerative disease.

INTRODUCTION

Microglia are the innate immune cells of the brain that are responsible for the excessive and chronic neuroinflammatory response known to contribute to the pathogenesis of brain injury and disease (Block et al. 2007, Glass et al. 2010). Inhibitors of histone deacetylases (HDACs) reduce the inflammatory response of isolated microglia to stimulants such as lipopolysaccharide (LPS) (Kannan et al. 2013, Chen et al. 2007, Faraco et al. 2009, Peng et al. 2005, Suh et al. 2010, Suuronen et al. 2003). When delivered in vivo, HDAC inhibitors (HDACi) reduce neuroinflammation, promote neuroprotection and improve functional outcomes in models of cerebral ischaemia (Kim & Chuang 2014, Kim et al. 2007, Xuan et al. 2012, Sinn et al. 2007), traumatic brain injury (Shein & Shohami 2011, Zhang et al. 2008) and encephalomyelitis (Camelo et al. 2005, Zhang et al. 2010). Therapies for treating brain injury and disease are lacking, but these studies highlight a role for HDACs in neuroinflammation and suggest they are appropriate targets to inhibit.

The mechanism by which HDAC inhibition is anti-inflammatory is not understood, but we know HDACs remove acetyl groups from lysine residues on proteins including, histones (Strahl & Allis 2000), enzymes and transcription factors (Glozak et al. 2005, Yao & Yang 2011). As a consequence, deacetylation of histones promotes a compact chromatin structure and reduces gene expression, and deacetylation of specific lysine residues on transcription factors can modulate their activity (Boyes et al. 1998, Gu & Roeder 1997) .

The identity of which acetylated proteins are responsible for the anti-inflammatory responses observed when using HDAC inhibitors is unclear. There are 18 mammalian HDAC isoforms (class I HDACs (1, 2, 3 and 8), class II HDACs (4-7, 9 and 10), class III sirtuins (1-7) and the

class IV HDAC11) and the majority of studies to date have focused on using non-selective HDAC inhibitors such as suberoylanilide hydroxamic acid (SAHA), trichostatin-A (TSA) and valproic acid (VPA) which inhibit the majority of these isoforms. As a result, we do not know which HDACs modulate the microglial inflammatory response and how inhibition of these leads to the acetylation of specific proteins to reduce neuroinflammation. Recent studies have begun to address these questions and have shown that selective inhibition of Class I HDACs 1, 2 and 3 with the HDAC inhibitor MS-275 (Hu et al. 2003, Simonini et al. 2006, Beckers et al. 2007, Khan et al. 2008) can reduce neuroinflammation in a mouse model of Alzheimer's disease (Zhang & Schluesener 2013) and the inflammatory response of macrophages to the inflammatory stimulant LPS (Jeong et al. 2014).

In our study, we have used selective HDAC inhibitors and siRNA knockdown to identify HDAC1 and HDAC2 as the key HDACs involved in the neuroinflammatory response of microglia. We show that selective class I HDAC inhibitors and siRNA to specifically knock-down HDAC1 and 2, both suppressed the expression of cytokines in BV2 murine microglia. Knockdown of HDAC1 alone resulted in a compensatory increase in the levels of HDAC2 and did not suppress cytokine expression, showing these two enzymes have redundancy in the neuroinflammatory response. We show that the HDACi are effective in the absence of new protein synthesis suggesting that the mechanism of HDACi does not involve increased gene expression. This identification suggests that HDAC selective inhibitors may be therapeutically useful for targeting microglia and neuroinflammation, in brain injury and disease by modulating the acetylation levels and function of non-histone protein(s).

MATERIALS & METHODS

CELL CULTURE

BV-2 murine microglia (RRID:CVCL_0182, Interlab Cell Line Collection #ATL03001 (Blasi et al. 1990)) were cultured in Dulbecco's Modified Eagle's Medium (DMEM) high glucose AQmedia™ (Sigma-Aldrich) supplemented with 10% v/v foetal bovine serum (FBS, PAA Cell Culture Company) and 100U penicillin/100 µg streptomycin (Sigma-Aldrich). Cells were seeded into 6-well plates at either 350,000 or 500,000 cells/well and 24-well plates at 175,000 cells/well. The cells were cultured 24 hours before treatment. BV-2 cells were treated with vehicle control, or HDAC inhibitor with or without 500 ng/mL lipopolysaccharide (LPS, Sigma), 1 µM oligomeric Beta-amyloid 1-42 (rPeptide) or 100 ng/ml Interferon gamma (IFN- γ) and cells harvested after 6 or 24 hours. Inhibitors used were; Apicidin (Sigma-Aldrich), MI192 (University of Leeds), MS-275 (Cayman Chemicals), SAHA (Cayman Chemicals) all dissolved in DMSO and Valproic acid (VPA, Sigma-Aldrich) dissolved in PBS. For pre-treatment experiments, BV-2 cells were treated with vehicle control or appropriate drugs for 24 hours before addition of 500 ng/mL LPS for a further 6 hours.

CELL TRANSFECTION

BV2 microglia seeded into 6-well plates were washed with 1 mL PBS (Oxoid) and maintained in 1 mL Opti-MEM® (Gibco) throughout the transfection procedure. Cells were transfected with 50 pmoles of Silencer® Select Negative Control siRNA (Ambion) or Silencer® Select Pre-designed siRNA targeted against HDAC1 (id: s119557, Ambion) or HDAC2 (id: s67417, Ambion) as follows. For each well, 3 µL of Lipofectamine™ 2000 (Invitrogen) was dissolved

in 100 μ L of Opti-MEM® and 1 μ L of 50 μ M siRNA was dissolved in 100 μ L of Opti-Mem®, these were incubated for 5 minutes before combining and incubating for a further 20 minutes at room temperature. Afterwards, 200 μ L of this mix was added to the well, followed by incubation at 37°C in a humid atmosphere with 5% CO₂ for 4 hours. This was then removed and the cells were cultured in 3 mL DMEM high glucose AQmedia™ supplemented with 1% v/v FBS and 100U penicillin/100 μ g streptomycin for 24 hours followed by a medium change and culture for a further 24 hours before treating 500 ng/mL LPS for 6 hours.

WHOLE CELL PROTEIN EXTRACTION

BV2 cells seeded into 6-well plates were washed with 1 mL PBS and scraped into 250 μ L ice-cold RIPA Buffer [10 mM Tris-HCL pH 8.0, 1 mM EDTA, 1% Triton X-100, 0.1% sodium deoxycholate, 0.1% sodium dodecyl sulfate (SDS), 140 mM NaCl, 1 mM PMSF (all from Sigma-Aldrich)], then incubated for 30 minutes on ice. The lysate was clarified by centrifugation at 13400 \times g for 20 minutes at 4°C and the supernatant containing proteins was collected and the concentration determined using the Bicinchoninic Acid (BCA) protein assay (Sigma-Aldrich).

HISTONE PROTEIN EXTRACTION

Histone proteins were extracted from BV2 microglia cultured in 6-well plates. Cells were washed with 1 mL PBS then scraped into 1 mL ice-cold PBS and pelleted by centrifugation at 400 \times g for 5 minutes at room temperature. The cell pellet was resuspended in 400 μ L of Triton Lysis Buffer per 1 \times 10⁷ cells [0.5% v/v Triton X-100, 2 mM PMSF, 0.02% w/v NaN₃ (all from Sigma-Aldrich) and PBS] and incubated on ice for 10 minutes. Lysed cells were centrifuged at

6600 × g for 10 minutes at 4°C. The pellet was resuspended in half the volume of Triton Lysis Buffer used earlier then centrifuged at 6600 × g for 10 minutes at 4°C. The nuclei pellet was resuspended in 50 µL of 200 mM HCl (Acros Organics) and histone proteins were extracted overnight at 4°C. The samples were centrifuged at 6600 × g for 10 minutes at 4°C and the supernatant containing histone proteins was collected and the protein concentration determined using the Bradford protein assay (Sigma-Aldrich).

WESTERN BLOTTING

Protein samples (10 µg) were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and wet-transferred onto PVDF membrane at 30V for 30 minutes. The membranes were blocked overnight at 4°C with blocking solution [5% w/v nonfat dried milk powder, 0.1% v/v Tween® 20 (both from Sigma-Aldrich) and PBS] and then incubated with either; Anti-HDAC1 [1:2000, rabbit polyclonal, Abcam, [RRID:AB_305705](#)], Anti-HDAC2 [1:2000, rabbit polyclonal, Abcam, [RRID:AB_305706](#)], Anti-HDAC3 [1:2000, rabbit polyclonal, Abcam, [RRID:AB_305708](#)], Anti-β-actin [1:10,000, mouse monoclonal, Sigma, [RRID:AB_476744](#)], Anti-Histone H3 [1:1000, mouse monoclonal, Cell Signaling, [RRID:AB_1642229](#)], Anti-acetyl Histone H3 Lysine 9 [1:1000, rabbit polyclonal, Millipore, [RRID:AB_310544](#)] or Anti-acetyl Histone H4 Pan-lysine [1:10,000, rabbit polyclonal, Millipore, [RRID:AB_310270](#)] (all dissolved in blocking solution) for 1 hour at room temperature. Membranes were washed with PBS-0.1% v/v Tween® 20 and incubated with an appropriate secondary antibody; Goat-Anti-Rabbit IgG-horseradish peroxidase (HRP) linked [1:2000, Cell Signaling, [RRID:AB_2099233](#)] or Goat-Anti-Mouse IgG-HRP linked [1:2000, Cell Signaling, [RRID:AB_330924](#)] for 1 hour at room temperature followed by washing as before. Membranes were incubated with an enhanced chemiluminescence (ECL) substrate

(Amersham) and exposed to either photographic film, a Fujifilm LAS-3000 imaging system or a cDigit® Scanner (LICOR). The intensity of each band was quantified and normalized to the β -actin loading control.

RNA EXTRACTION, REVERSE TRANSCRIPTION AND QUANTITATIVE PCR (qPCR)

Total RNA was extracted from BV2 microglia cells using TRI Reagent® (Sigma-Aldrich) as per manufacturer's instructions and resuspended in TE, pH 7.5 and concentration were determined using a NanoDrop 2000c (Thermo Scientific). RNA (2.5 μ g) was primed for reverse transcription at 65°C for 5 minutes with 1.25 μ L of Oligo(dT)15 primers (0.5 μ g/ μ L, Promega), 1.25 μ L of Random primers (0.5 μ g/ μ L, Promega) in a final volume of 32.5 μ L. cDNA was synthesised at 37°C for 60 min using M-MLV Reverse Transcriptase, RNase H Minus (200 U/ μ L and RNasin® Plus, Promega) with 2 mM dNTP (Bioline) in a final reaction volume of 50 μ L. Quantitative PCR (qPCR) reactions were carried out in duplicate using a Rotor Gene 6000 PCR Analyzer (Corbett) using SensiMix™ SYBR® & Fluorescein (Bioline). Each reaction comprised of 50 or 100 ng of sample cDNA, 300 nM of each gene primer, in a final volume of 20 μ L. Primers used were: IL-6 5'-CCCAACTTCCAATGCTCTCC and 5'-ACATGGGATTCCACAAAC, TNF- α 5'-TGA ACTTCGGGGTGATCG and 5'-GGGCTTGTC ACTCGAGTTTT, U6 5'- CCGCTTCGGCAGCACA and 5'-AACGCTTCACGAATTTGCGT, HDAC1 5'- GACCGCAAG-TGTGTGG and 5'-GAGCAACATTCCGGATGGTG, HDAC2 5'- CAACAGATCGCGTGAT-GACC and 5'-CCCTTTCCAGCACCAATATCC, HDAC3 5'-GACGTGCATCGTGCTCC-AGT and 5'-ACATTCCCCATGTCCTCGAAT. An RNA control (no reverse transcription) and a no template control were also run. PCR conditions were: 95°C for 10 min followed by 45 cycles

of 95°C for 10 s, 60°C for 15 s and 72°C for 30 s. Relative quantitation of transcript levels was performed using the $2^{-\Delta\Delta C_t}$ method (Livak & Schmittgen, 2001) and the house-keeping gene U6.

ENZYME LINKED IMMUNOSORBANT ASSAYS (ELISAs)

Cell culture supernatants were taken from BV2 microglia cells cultured in 24-well plates. The culture medium was removed and centrifuged for 30 seconds at 16000 × g to pellet any detached cells then 950 µL was removed for analysis. The concentration of mouse IL-6 protein was determined in triplicate by 96-well plate format ELISAs following Invitrogen's instructions.

INHIBITION OF PROTEIN SYNTHESIS

Cells were treated to 500 ng/mL LPS and either vehicle control, 1 µM SAHA or 500 nM Apicidin, in the presence or absence of 1 µg/mL cycloheximide (CHX) for 3 hours. Protein synthesis was assessed in the cells using a Click-iT® Plus O-propargyl-puromycin (OPP) Protein Synthesis Assay Kit (Molecular Probes, Life Technologies). Briefly, after 2.5 hours of drug treatments, Click-iT® OPP was added directly to each well to give a final concentration of 20 µM. The cells were incubated for a further 30 minutes at 37°C. The culture medium was removed, the cells washed with PBS, followed by fixation with 100 µL of 4% w/v paraformaldehyde (Sigma-Aldrich) for 15 minutes at room temperature. The cells were then permeabilised with 100 µL of 0.5% v/v Triton X-100 (Sigma-Aldrich) for 15 minutes at room temperature, washed twice with PBS and processed for imaging following the manufacturer's instructions. Imaging of labelled cells was carried out using the IncuCyte™ FLR with a 10× objective lens. Nine non-overlapping images were taken in phase-contrast and green-

fluorescence (excitation wavelength of 450-490 nm) per well. RNA was extracted from parallel treated cultures for qPCR analysis as described above.

STATISTICS

For qPCR experiments, data are presented either as mean percentage expression of the vehicle control \pm SEM or mean percentage expression of LPS +vehicle condition \pm SEM. Statistical analysis for each experimental condition vs. the vehicle condition was performed using a one-way repeated measures ANOVA followed by the Dunnet post hoc test (Lew 2007) at the 5% significance level. For ELISA experiments, data is presented as mean protein concentration \pm standard deviation (SD). Statistical analysis comparing the absolute absorbance values for each experimental condition vs. the vehicle control +LPS condition was performed using a Student's unpaired t-test assuming equal variances at the 5% significance level. For all quantitative data experiments were performed in triplicate and each experiment was performed at least three times, the "n" number represents number of experiments performed. No blinding was performed.

RESULTS

HDAC INHIBITION SUPPRESSES CYTOKINE EXPRESSION IN BV2 MICROGLIA

Previous studies using isolated murine microglia report that non-selective HDAC inhibitors (SAHA, TSA and VPA) suppress LPS induced inflammation as measured by a reduction in LPS induced cytokine expression (tumour necrosis factor-alpha, TNF- α and interleukin-6, IL-6) (Kannan et al. 2013, Suh et al. 2010). The specific HDAC(s) important in microglia and neuroinflammation are still largely unknown though identification of the specific HDAC is a requisite for the development of any targeted therapy. We first tested the response of LPS

activated BV2 microglia cells to the classical HDAC inhibitors SAHA and VPA. BV2 cells are a widely used model for primary microglia cells showing the same responses across a range of studies (Gresa-Arribas et al. 2012, Henn et al. 2009, Stansley et al. 2012, Horvath et al. 2008, Bocchini et al. 1992)). In response to LPS and IFN- γ BV2 cells show the same elevation of TNF- α , IL-6, COX2, iNOS and NO as primary microglia (Gresa-Arribas et al. 2012). A transcriptomic analysis of the LPS response of primary microglia and BV2 cells showed that the gene induction involved the same genes, though the magnitude of change was higher in primary microglia (Henn et al. 2009). Like primary microglia, BV2 cells are capable of triggering astrocyte activation when co-cultured with them (Henn et al. 2009, Horvath et al. 2008) phagocytose A β peptides (He et al. 2011) and pre-stimulation with IL-10 prevents LPS induced cytokine expression (Gresa-Arribas et al. 2012). Thus BV2 cells show the same responses as primary microglia, providing a suitable model system to study mechanisms underlying these responses with the additional advantage that, unlike primary microglia cultures which are mixed cultures, BV2 cultures are homogenous. Six hours after stimulation with LPS, oligomeric A β or IFN- γ , IL-6 mRNA expression was increased (Fig 1A). Incubation of BV2 cells with the HDAC inhibitor SAHA, inhibited the stimulation of IL-6 expression by each of the three stimulants used (Fig 1A). BV2 cells express the Class I HDACs, 1, 2 and 3 with highest levels of HDAC1 and lowest levels of HDAC3 (Fig 1B, C). Treatment with the HDAC inhibitors, SAHA and VPA produced an increase in the level of Histone H4 acetylation levels within 1hr which was stable over a period of 24 hr (Fig 1D) suggesting these inhibitors provide rapid and stable HDAC inhibition. Activation by LPS in the presence of either 1 μ M SAHA or 5 mM of VPA, produced a significantly reduced response in IL-6 mRNA expression by $84.1 \pm 2.8\%$ ($P = 0.004$) and $89.7 \pm 1.6\%$ respectively and TNF- α mRNA expression by $59.7 \pm 3.2\%$ and $77.9 \pm 2.5\%$ respectively (Fig 1E). Furthermore, SAHA significantly suppressed the LPS induced increase in IL-6 protein secretion by $85.6 \pm 2.5\%$

(Fig 1F). HDAC treatment did not have a significant effect on IL-6 levels in the absence of LPS stimulation (Fig 1A, E) and LPS treatment did not have a direct impact on HDAC mRNA levels (Fig 1G) or protein levels (Fig 1H) which remained unchanged.

In order to understand the role of specific HDACs in microglia and neuroinflammation, we tested HDAC inhibitors that show some selectivity towards specific HDAC isoforms *in vitro*. We treated LPS induced cells with MS-275 (which has reported selectivity for HDAC1 (Bradner, 2008)), 500 nM apicidin, or 1 μ M MI-192, (both of which have reported selectivity for HDAC2 and 3 (Khan, et al 2008; Boissinot et al., 2012)). Treatment of BV2 cells with these inhibitors showed a rapid and stable increase in acetylated Histone H4 for apicidin but a gradual increase for MS-275 and MI-192 over a 24 hour period (Fig 2A). Quantification of histone acetylation levels showed that apicidin produced a similar rate of increase to SAHA while MS-275 and MI-192 required longer incubation periods to induce high levels of histone acetylation (Fig 2B). MS-275 and MI-192 are members of the benzamide class of HDAC inhibitors which have previously been shown to bind HDACs with a slower association rate compared to hydroxamic acid inhibitors such as SAHA (Lauffer et al. 2013). Consistent with these data, co-treatment of LPS with apicidin was sufficient to reduce the induction of IL-6 and TNF- α expression by $82.7 \pm 2.3\%$ and $50.3 \pm 4.5\%$ but co-treatment of MS-275 or MI-192 was not, (not shown). Pre-treatment of BV2 cells, to mitigate the slow kinetics of inhibition, with either apicidin, MI-912 or MS-275 for 24 hr prior to LPS stimulation significantly reduced the LPS stimulated expression of IL-6 and TNF- α (Fig 2C). Together these data suggest that HDAC1 and HDAC2/3 contribute to the inflammatory response in microglia.

KNOCKDOWN OF HDAC1 OR HDAC2 SUPPRESSES CYTOKINE EXPRESSION IN BV2 MICROGLIA

Although the HDAC inhibitors show selectivity, they are not isoform specific, therefore we used an siRNA approach to specifically knockdown HDAC1 and HDAC2 expression to determine their involvement in the inflammatory response of microglia. We were able to significantly knock-down HDAC1 protein expression by $62.6 \pm 4.5\%$ (Fig 3A) and HDAC2 protein expression by $68.8 \pm 7.7\%$ (Fig 3A). Knockdown of HDAC1 resulted in an increased expression of HDAC2 with no change in HDAC3 expression (Fig 3A) while knockdown of HDAC2 did not result in any change of expression in HDACs 1 or 3 (Fig 3A). Following knock-down, we treated cells to 500 ng/mL LPS for 6 hours and assessed the expression of IL-6 and TNF- α mRNA. We found, that cells in which HDAC1 was knocked down, we did not observe any change in the response to LPS compared to control cells (not shown) but cells in which HDAC2 was knocked down showed a reduced induction of IL-6 (by $48.2 \pm 13\%$) and TNF- α (by $22.0 \pm 3.6\%$) expression in response to LPS (Fig 3B). To determine if the increase in HDAC2 expression, as a result of HDAC1 knockdown, was acting as a compensatory mechanism we used siRNA to HDAC1 to knockdown HDAC1 in combination with a titrated amount of HDAC2 siRNA to reduce HDAC2 to levels seen in control cells (Fig. 3, HDAC1 + 2). Using this titrated level of HDAC2 siRNA we were able to reduce HDAC1 levels by $63.5 \pm 2.4\%$, while maintaining the HDAC2 levels seen in control cells ($89.7 \pm 6.2\%$, Fig 3A HDAC1 + 2). Expression of HDAC3 was not significantly altered ($116 \pm 5\%$ of Scr siRNA). Cells in which HDAC1 levels are reduced and HDAC2 levels are unchanged resulted in a reduced response of IL-6 and TNF- α mRNA to LPS of $34.8 \pm 3.0\%$ and $35.7 \pm 4.8\%$ respectively (Fig 3B).

In summary our data identify HDAC1 and 2 activities as important contributors to the neuroinflammatory response of microglia. HDAC1 and 2 may have the same or independent roles in activation though the observation that they show redundancy in this function with

increased HDAC2 levels being compensatory for reduced HDAC1 suggests there must be at least some overlap.

HDAC INHIBITION IS EFFECTIVE IN REDUCING THE INFLAMMATORY RESPONSE IN THE ABSENCE OF NEW PROTEIN SYNTHESIS

The mechanism by which HDAC inhibitors exert their effects is often assumed to involve increase in gene expression brought about as a result of increased histone acetylation. Incubation of BV2 cells with these HDAC inhibitors do result in increased histone acetylation (eg Fig 1D) and the association of increased acetylation with increased gene expression was first identified nearly 30 years ago (Hebbes et al. 1988). However recent proteomic data has identified in excess of 4000 proteins that are modified by acetylation (Choudhary et al. 2009, Liu et al. 2014, Lundby et al. 2012), a number comparable to targets of phosphorylation and suggests that acetylation is involved in many more processes than gene regulation alone. To test a requirement for changes in gene expression for the anti-inflammatory action of HDAC inhibition we blocked new protein synthesis using cycloheximide and tested the effectiveness of HDAC inhibitors to block IL-6 and TNF- α stimulation by LPS. Incubation of BV2 cells with cycloheximide for 1 or 3 hours completely blocked new protein synthesis as measured by O-propargyl-puromycin incorporation and protein synthesis was blocked under all conditions used to quantify gene expression levels (Fig 4A). Continued exposure to cycloheximide for 6 hr led to cell death (not shown) though 3 hr is sufficient time to observe the induction of IL-6 mRNA by LPS stimulation (Fig 4B, left bar) and cells were still healthy at this time point. The presence of cycloheximide did not affect the induction of IL-6 mRNA expression by LPS (Fig 4B, compare left two bars) and did not prevent either SAHA or Apicidin inhibiting this response (Fig 4B, right two bars). Thus these data indicate that the mechanism by which HDAC inhibition reduces the inflammatory response in microglia is manifest within 3 hours and does

not require new protein synthesis. Together this suggests that increased gene expression resulting from enhanced histone acetylation is not important for ability of HDAC inhibitors to reduce microglia activation and future work should aim to identify the important molecular targets.

DISCUSSION

It has been previously shown that inhibitors of HDACs can reduce the inflammatory response in activated microglia (Chen & Greene 2004, Faraco et al. 2009, Kannan et al. 2013, Suh et al. 2010) and by suppressing microglia activation show neuroprotective effects following transient ischaemia in vivo (Kim & Chuang 2014, Kim et al. 2007, Sinn et al. 2007, Xuan et al. 2012). However, the identity of the important HDACs involved has not been uncovered and the mechanism by which HDAC inhibition is beneficial is yet to be elucidated. Here we have shown that the function of both HDAC1 and HDAC2 contribute to the inflammatory response in microglia and that in the absence of HDAC1, increased HDAC2 levels compensate suggesting that these 2 HDACs show redundancy in this function. Furthermore the effectiveness of HDAC inhibition in the absence of new protein synthesis suggests that the HDACs are promoting the inflammatory response by regulating the acetylation levels of a non-histone protein rather than increasing levels of gene expression as a result of increased histone acetylation.

Microglia are often referred to as the immune cells of the brain and recently, selective inhibition and genetic knock-down of class I HDACs, was shown to reduce the production of cytokines in the inflammatory response of macrophages (Jeong et al. 2014). In macrophages, knockdown of either HDAC1 or 2 resulted in increased expression of the other and only a combined knockdown of HDACs1, 2 and 3 resulted in reduced inflammatory response to LPS (Jeong et al. 2014). In T lymphocytes deletion of HDAC1 resulted in an increase in HDAC2 protein levels but deletion of HDAC2 had no effect on HDAC1 (Dovey et al. 2013). Here we show that in microglial cells, that HDAC1 is the most highly expressed class I HDAC and knockdown of HDAC1 resulted in a compensatory increase in the levels of HDAC2 (Fig 3A).

likewise, we did not observe any compensatory increase in the levels of HDAC1 protein upon knockdown of HDAC2 (Fig 3A) however this contrasts to observations made using macrophages (Jeong et al. 2014). The mechanisms resulting in a compensatory increase in one HDAC upon loss of another are not known though HDAC1 does regulate its own promoter (Schuettengruber et al. 2003) and may also repress expression of other HDACs. One prediction of such a model would be that chemical inhibition of HDAC activity would also result in such compensatory increase. However, we did not observe any compensatory changes in HDAC expression in cells treated with HDAC inhibitors (not shown), suggesting it is not brought about by loss of HDAC enzyme activity but is potentially a mechanism involving the absence of the protein itself. In the absence of HDAC1 in T-lymphocytes the levels of SIN3 and MTA2 are reduced which may indicate that incomplete co-repressor complexes are turned over quickly (Dovey et al. 2013). This structural, rather than enzymatic, requirement for HDAC1 may underlie the reason that knockdown of HDAC1, but not inhibition results in a compensatory increase in HDAC2. Additionally, compensatory changes in HDAC protein levels have been observed in the absence of changes in mRNA levels, suggesting the mechanism involves enhanced translation or protein stability (Jurkin et al. 2011).

HDAC1 and 2 do not exist in the cell as isolated enzymes but are components of 3 independent co-repressor complexes; Sin3, NuRD and CoREST (for a review see (Kelly & Cowley 2013)). Each co-repressor complex contains 2 molecules of HDAC which may consist of two molecules of HDAC1, two molecules of HDAC2 or one of each. Others have observed that upon a loss of HDAC1, HDAC2 can become incorporated into the Sin3, NuRD and CoREST multi-protein complexes in its place (Dovey et al. 2013). The compensatory effect of HDAC2 in the inflammatory response may be explained by such a mechanism. Following a loss of HDAC1, HDAC2 is upregulated and this HDAC is incorporated into a specific complex in

place of HDAC1. This complex, specifically targets a protein, (which regulates the inflammatory response) for deacetylation. Regardless of the HDAC composition, be it two molecules of HDAC1, HDAC2 or one of each, the specificity for the substrate to be deacetylated comes from the complex itself rather than the HDACs. This hypothesis would suggest it doesn't matter which of the two HDAC isoforms is inhibited, an anti-inflammatory effect depends on a reduction in the number and activity of this specific functional multi-protein complex. Similarly, the compensatory effect of HDAC3 when HDAC1 and 2 are both lost in macrophages (Jeong et al. 2014) may be explained by HDAC3 being in a specific complex that targets the same substrate as the complex with either HDAC1 or 2. Further research is now needed to investigate these hypotheses and identify the complexes (and composition of them) that when inhibited is responsible for the suppression of pro-inflammatory mediator expression in BV2 microglia.

What is the important target of HDAC1 and 2 that promotes the inflammatory response? HDAC enzymes were originally characterised by their ability to deacetylate histone proteins, however these are not their only target and the acetylome may contain on the order of 4000 proteins (Choudhary et al. 2009, Liu et al. 2014). Additionally, the original idea, that HDAC inhibition leads to increased histone acetylation and increased gene expression is likely too simplistic because as many genes are repressed as are activated upon HDAC inhibition by SAHA (Peart et al. 2005). The specific HDAC target(s) important for the microglial response has not been unequivocally identified though a number of potential target proteins can be implicated based on a correlation of their acetylation with microglial activation. Perhaps the most studied non-histone protein involved in the inflammatory response and regulated by acetylation is the transcription factor NF- κ B (Greene & Chen 2004). Quiescent NF- κ B is restricted to the cytoplasm via its inhibitory binding partner I κ B but upon cell stimulation

becomes dissociated from I κ B and moves into the nucleus where it activates target gene expression. Initially, it was proposed that deacetylation of NF- κ B enhanced its interaction with I κ B and removal from the nucleus however NF- κ B can be acetylated at multiple sites and more recent data suggests that deacetylation at specific residues can result in activation of a subset of NF- κ B targets (Rothgiesser et al. 2010) thus inhibition of HDACs may enhance the level of acetylated NF- κ B, reducing its activity. In support of this idea, Furumai et al., 2011 showed that inhibition of HDACs in HeLa cells, with TSA, caused a reduction in the recruitment of NF- κ B, and RNA polymerase II to the promoter of IL-8 and a reduction in its expression (Furumai et al. 2011). In macrophages HDAC1, 2 and 3 can deacetylate MKP-1 (which is a member of the MAPK signalling pathway) regulating its phosphorylation and reducing its activity (Jeong et al. 2014). MKP-1 is a negative regulator of the inflammatory response not just in macrophages but also in microglia (Eljaschewitsch et al. 2006) making this another attractive candidate for the functional response observed here.

References

- Beckers, T., Burkhardt, C., Wieland, H., Gimmnich, P., Ciossek, T., Maier, T. and Sanders, K. (2007) Distinct pharmacological properties of second generation HDAC inhibitors with the benzamide or hydroxamate head group. *International journal of cancer. Journal international du cancer*, **121**, 1138-1148.
- Blasi, E., Barluzzi, R., Bocchini, V., Mazzolla, R. and Bistoni, F. (1990) Immortalization of murine microglial cells by a v-raf/v-myc carrying retrovirus. *Journal of neuroimmunology*, **27**, 229-237.
- Block, M. L., Zecca, L. and Hong, J. S. (2007) Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nature reviews. Neuroscience*, **8**, 57-69.
- Bocchini, V., Mazzolla, R., Barluzzi, R., Blasi, E., Sick, P. and Kettenmann, H. (1992) An immortalized cell line expresses properties of activated microglial cells. *Journal of neuroscience research*, **31**, 616-621.
- Boyes, J., Byfield, P., Nakatani, Y. and Ogryzko, V. (1998) Regulation of activity of the transcription factor GATA-1 by acetylation. *Nature*, **396**, 594-598.
- Camelo, S., Iglesias, A. H., Hwang, D. et al. (2005) Transcriptional therapy with the histone deacetylase inhibitor trichostatin A ameliorates experimental autoimmune encephalomyelitis. *Journal of neuroimmunology*, **164**, 10-21.
- Chen, L. F. and Greene, W. C. (2004) Shaping the nuclear action of NF-kappaB. *Nat Rev Mol Cell Biol*, **5**, 392-401.
- Chen, P. S., Wang, C. C., Bortner, C. D. et al. (2007) Valproic acid and other histone deacetylase inhibitors induce microglial apoptosis and attenuate lipopolysaccharide-induced dopaminergic neurotoxicity. *Neuroscience*, **149**, 203-212.
- Choudhary, C., Kumar, C., Gnad, F., Nielsen, M. L., Rehman, M., Walther, T. C., Olsen, J. V. and Mann, M. (2009) Lysine acetylation targets protein complexes and co-regulates major cellular functions. *Science*, **325**, 834-840.
- Dovey, O. M., Foster, C. T., Conte, N., Edwards, S. A., Edwards, J. M., Singh, R., Vassiliou, G., Bradley, A. and Cowley, S. M. (2013) Histone deacetylase 1 and 2 are essential for normal T-cell development and genomic stability in mice. *Blood*, **121**, 1335-1344.
- Eljaschewitsch, E., Witting, A., Mawrin, C. et al. (2006) The endocannabinoid anandamide protects neurons during CNS inflammation by induction of MKP-1 in microglial cells. *Neuron*, **49**, 67-79.
- Faraco, G., Pittelli, M., Cavone, L., Fossati, S., Porcu, M., Mascagni, P., Fossati, G., Moroni, F. and Chiarugi, A. (2009) Histone deacetylase (HDAC) inhibitors reduce the glial inflammatory response in vitro and in vivo. *Neurobiology of disease*, **36**, 269-279.
- Furumai, R., Ito, A., Ogawa, K., Maeda, S., Saito, A., Nishino, N., Horinouchi, S. and Yoshida, M. (2011) Histone deacetylase inhibitors block nuclear factor-kappaB-dependent transcription by interfering with RNA polymerase II recruitment. *Cancer Sci*, **102**, 1081-1087.
- Glass, C. K., Saijo, K., Winner, B., Marchetto, M. C. and Gage, F. H. (2010) Mechanisms underlying inflammation in neurodegeneration. *Cell*, **140**, 918-934.
- Glozak, M. A., Sengupta, N., Zhang, X. and Seto, E. (2005) Acetylation and deacetylation of non-histone proteins. *Gene*, **363**, 15-23.
- Greene, W. C. and Chen, L. F. (2004) Regulation of NF-kappaB action by reversible acetylation. *Novartis Found Symp*, **259**, 208-217; discussion 218-225.
- Gresa-Arribas, N., Vieitez, C., Dentesano, G., Serratosa, J., Saura, J. and Sola, C. (2012) Modelling neuroinflammation in vitro: a tool to test the potential neuroprotective effect of anti-inflammatory agents. *PloS one*, **7**, e45227.

- Gu, W. and Roeder, R. G. (1997) Activation of p53 Sequence-Specific DNA Binding by Acetylation of the p53 C-Terminal Domain. *Cell*, **90**, 595-606.
- He, F. Q., Qiu, B. Y., Li, T. K., Xie, Q., Cui, D. J., Huang, X. L. and Gan, H. T. (2011) Tetrandrine suppresses amyloid-beta-induced inflammatory cytokines by inhibiting NF-kappaB pathway in murine BV2 microglial cells. *Int Immunopharmacol*, **11**, 1220-1225.
- Hebbes, T. R., Thorne, A. W. and Crane-Robinson, C. (1988) A direct link between core histone acetylation and transcriptionally active chromatin. *EMBO J*, **7**, 1395-1402.
- Henn, A., Lund, S., Hedtjarn, M., Schrattenholz, A., Porzgen, P. and Leist, M. (2009) The suitability of BV2 cells as alternative model system for primary microglia cultures or for animal experiments examining brain inflammation. *Altx*, **26**, 83-94.
- Horvath, R. J., Natile-McMenemy, N., Alkaitis, M. S. and Deleo, J. A. (2008) Differential migration, LPS-induced cytokine, chemokine, and NO expression in immortalized BV-2 and HAPI cell lines and primary microglial cultures. *Journal of neurochemistry*, **107**, 557-569.
- Hu, E., Dul, E., Sung, C. M. et al. (2003) Identification of novel isoform-selective inhibitors within class I histone deacetylases. *J Pharmacol Exp Ther*, **307**, 720-728.
- Jeong, Y., Du, R., Zhu, X., Yin, S., Wang, J., Cui, H., Cao, W. and Lowenstein, C. J. (2014) Histone deacetylase isoforms regulate innate immune responses by deacetylating mitogen-activated protein kinase phosphatase-1. *Journal of leukocyte biology*, **95**, 651-659.
- Jurkin, J., Zupkovitz, G., Lagger, S., Grausenburger, R., Hagelkruys, A., Kenner, L. and Seiser, C. (2011) Distinct and redundant functions of histone deacetylases HDAC1 and HDAC2 in proliferation and tumorigenesis. *Cell Cycle*, **10**, 406-412.
- Kannan, V., Brouwer, N., Hanisch, U. K., Regen, T., Eggen, B. J. and Boddeke, H. W. (2013) Histone deacetylase inhibitors suppress immune activation in primary mouse microglia. *Journal of neuroscience research*.
- Kelly, R. D. and Cowley, S. M. (2013) The physiological roles of histone deacetylase (HDAC) 1 and 2: complex co-stars with multiple leading parts. *Biochem Soc Trans*, **41**, 741-749.
- Khan, N., Jeffers, M., Kumar, S. et al. (2008) Determination of the class and isoform selectivity of small-molecule histone deacetylase inhibitors. *Biochem J*, **409**, 581-589.
- Kim, H. J. and Chuang, D. M. (2014) HDAC inhibitors mitigate ischemia-induced oligodendrocyte damage: potential roles of oligodendrogenesis, VEGF, and anti-inflammation. *American journal of translational research*, **6**, 206-223.
- Kim, H. J., Rowe, M., Ren, M., Hong, J. S., Chen, P. S. and Chuang, D. M. (2007) Histone deacetylase inhibitors exhibit anti-inflammatory and neuroprotective effects in a rat permanent ischemic model of stroke: multiple mechanisms of action. *J Pharmacol Exp Ther*, **321**, 892-901.
- Lauffer, B. E., Mintzer, R., Fong, R. et al. (2013) Histone deacetylase (HDAC) inhibitor kinetic rate constants correlate with cellular histone acetylation but not transcription and cell viability. *The Journal of biological chemistry*.
- Lew, M. (2007) Good statistical practice in pharmacology. Problem 2. *British journal of pharmacology*, **152**, 299-303.
- Liu, F., Yang, M., Wang, X., Yang, S., Gu, J., Zhou, J., Zhang, X. E., Deng, J. and Ge, F. (2014) Acetylome analysis reveals diverse functions of lysine acetylation in *Mycobacterium tuberculosis*. *Mol Cell Proteomics*, **13**, 3352-3366.
- Lundby, A., Lage, K., Weinert, B. T. et al. (2012) Proteomic analysis of lysine acetylation sites in rat tissues reveals organ specificity and subcellular patterns. *Cell Rep*, **2**, 419-431.
- Peart, M. J., Smyth, G. K., van Laar, R. K., Bowtell, D. D., Richon, V. M., Marks, P. A., Holloway, A. J. and Johnstone, R. W. (2005) Identification and functional significance

- of genes regulated by structurally different histone deacetylase inhibitors. *Proc Natl Acad Sci U S A*, **102**, 3697-3702.
- Peng, G. S., Li, G., Tzeng, N. S., Chen, P. S., Chuang, D. M., Hsu, Y. D., Yang, S. and Hong, J. S. (2005) Valproate pretreatment protects dopaminergic neurons from LPS-induced neurotoxicity in rat primary midbrain cultures: role of microglia. *Brain research. Molecular brain research*, **134**, 162-169.
- Rothgiesser, K. M., Fey, M. and Hottiger, M. O. (2010) Acetylation of p65 at lysine 314 is important for late NF-kappaB-dependent gene expression. *BMC Genomics*, **11**, 22.
- Schuettengruber, B., Simboeck, E., Khier, H. and Seiser, C. (2003) Autoregulation of mouse histone deacetylase 1 expression. *Mol Cell Biol*, **23**, 6993-7004.
- Shein, N. A. and Shohami, E. (2011) Histone deacetylase inhibitors as therapeutic agents for acute central nervous system injuries. *Molecular medicine*, **17**, 448-456.
- Simonini, M. V., Camargo, L. M., Dong, E., Maloku, E., Veldic, M., Costa, E. and Guidotti, A. (2006) The benzamide MS-275 is a potent, long-lasting brain region-selective inhibitor of histone deacetylases. *Proceedings of the National Academy of Sciences of the United States of America*, **103**, 1587-1592.
- Sinn, D. I., Kim, S. J., Chu, K. et al. (2007) Valproic acid-mediated neuroprotection in intracerebral hemorrhage via histone deacetylase inhibition and transcriptional activation. *Neurobiology of disease*, **26**, 464-472.
- Stansley, B., Post, J. and Hensley, K. (2012) A comparative review of cell culture systems for the study of microglial biology in Alzheimer's disease. *Journal of neuroinflammation*, **9**, 115.
- Strahl, B. D. and Allis, C. D. (2000) The language of covalent histone modifications. *Nature*, **403**, 41-45.
- Suh, H. S., Choi, S., Khattar, P., Choi, N. and Lee, S. C. (2010) Histone deacetylase inhibitors suppress the expression of inflammatory and innate immune response genes in human microglia and astrocytes. *J Neuroimmune Pharmacol*, **5**, 521-532.
- Suuronen, T., Huuskonen, J., Pihlaja, R., Kyrlylenko, S. and Salminen, A. (2003) Regulation of microglial inflammatory response by histone deacetylase inhibitors. *Journal of neurochemistry*, **87**, 407-416.
- Xuan, A., Long, D., Li, J., Ji, W., Hong, L., Zhang, M. and Zhang, W. (2012) Neuroprotective effects of valproic acid following transient global ischemia in rats. *Life sciences*, **90**, 463-468.
- Yao, Y. L. and Yang, W. M. (2011) Beyond histone and deacetylase: an overview of cytoplasmic histone deacetylases and their nonhistone substrates. *J Biomed Biotechnol*, **2011**, 146493.
- Zhang, B., West, E. J., Van, K. C., Gurkoff, G. G., Zhou, J., Zhang, X. M., Kozikowski, A. P. and Lyeth, B. G. (2008) HDAC inhibitor increases histone H3 acetylation and reduces microglia inflammatory response following traumatic brain injury in rats. *Brain research*, **1226**, 181-191.
- Zhang, Z. Y. and Schluesener, H. J. (2013) Oral Administration of Histone Deacetylase Inhibitor MS-275 Ameliorates Neuroinflammation and Cerebral Amyloidosis and Improves Behavior in a Mouse Model. *Journal of neuropathology and experimental neurology*, **72**, 178-185.
- Zhang, Z. Y., Zhang, Z. and Schluesener, H. J. (2010) MS-275, an histone deacetylase inhibitor, reduces the inflammatory reaction in rat experimental autoimmune neuritis. *Neuroscience*, **169**, 370-377.

Figure 1. Histone deacetylase (HDAC) inhibition suppress activation of cytokine expression in murine microglia. A) BV2 microglia were stimulated with 500 ng/mL lipopolysaccharide (LPS), 1 μ M oligomeric Beta-Amyloid 1-42 (Abeta), 100 ng/ml IFN- γ or vehicle (Con) in the presence or absence of 1 μ M SAHA and mRNA expression of interleukin-6 (IL-6) was determined using qPCR. Relative transcript levels were normalized to the U6 gene and expressed as a percentage of the expression in LPS treated cells. Shown are mean \pm SEM, n=3. B) Quantitative RT-PCR of RNA extracted from BV2 cells. Expression levels are shown as a percentage of the U6 gene. Shown are mean \pm SEM, n=3 C) Western blot analysis of protein extracted from BV2 cells and analysed using anti-HDAC1, HDAC2, HDAC3 and β -actin. Representative blots are shown and positions of molecular weight markers are identified on the left. D) Histone proteins were extracted from control cells and cells treated with 1 μ M SAHA or 10 mM VPA for 1, 2, 4, 6, or 24hr. Proteins underwent western blotting for acetylated histone H4 (Ac-H4) and total histone H3. Representative blots are shown. E) BV2 microglia were treated with vehicle or 500 ng/mL LPS \pm 1 mM SAHA or 5 mM valproic acid (VPA) for 6 hours and the mRNA expression of IL-6 and TNF-a was determined using qPCR, normalized to the U6 house-keeping gene and expressed as a percentage of the expression in control cells treated LPS, shown are mean \pm SEM, n=3. F) BV2 microglia were treated with 500 ng/mL LPS \pm 1 mM SAHA for 24 hours and the changes in IL-6 protein secretion was determined by ELISA. Shown are mean protein concentration \pm SEM, n=3.). G) Quantitative RT-PCR of RNA extracted from vehicle and LPS treated BV2 cells. Shown are HDAC mRNA levels in LPS treated cells expressed as a percentage of the HDAC levels in control cells, mean \pm SEM, n=3 H) Quantitative western blot analysis of proteins extracted from vehicle and LPS treated BV2 cells. Shown are HDAC protein levels in LPS treated cells

expressed as a percentage of the HDAC levels in control cells, mean \pm SEM, n=3 *P < 0.05, **P < 0.01, ***P < 0.001.

Figure 2. Selective Histone deacetylase (HDAC) inhibitors suppress activation of cytokine expression in murine microglia. A) Histone proteins were extracted from control cells and cells treated with 500nM mM Apicidin, 1 μ M MI-192 or 5 μ M MS-275 for 1, 2, 4, 6, or 24hr. Proteins underwent western blotting for acetylated histone H4 (Ac-H4) and total histone H3. Representative blots are shown. B) Quantification of acetylated histone H4 levels, normalised to Histone H3 and expressed relative to acetylation levels at 24hr. C) BV2 microglia were pretreated with HDACi for 24 hours and then stimulated with vehicle or 500 ng/mL lipopolysaccharide (LPS) for 6 hours. mRNA expression of interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α) was determined using qPCR. Relative transcript levels were normalized to the U6 house-keeping gene and expressed as a percentage of the expression in control cells treated with vehicle. Shown are mean \pm SEM, n=3. *P < 0.05, **P < 0.01, ***P < 0.001 compared to LPS stimulated cells.

Figure 3. HDAC1 and HDAC2 are involved in the inflammatory response in microglia. A) BV2 microglia were transfected with either scrambled (Scr) siRNA or HDAC1, HDAC2 or HDACs1 and 2 siRNAs. Levels of HDAC1, 2 and 3 and Beta-actin were quantified by immunoblotting. Shown are a representative blots (left) and quantification of protein levels (right) expressed relative to levels in cells treated with Scrambled siRNA. Shown are mean \pm SEM, n=3 *p<0.05 compared to scrambled siRNA. B) BV2 microglia transfected with Scr, HDAC1, HDAC2 or HDAC1 and 2 siRNAs were stimulated with LPS and changes in IL-6 and TNF- α mRNA expression was determined using qPCR. Relative transcript levels in each treatment were normalized to the U6 gene and expressed relative to the expression in the Scr siRNA +LPS. Shown are mean \pm SEM, n=3. *P < 0.05

Figure 4: Protein synthesis is not required for HDAC inhibitor efficacy. A) BV2 microglia were treated for 1 or 3 hours with or without cycloheximide and incorporation of O-propargyl-puromycin (green, New protein) identified new protein synthesis. Bottom shows phase contrast images, scale bar 50 μ m. B) Expression of the IL-6 was measured by quantitative PCR in cells exposed to LPS and treated with vehicle, SAHA or apicidin in the presence of cycloheximide. Transcript levels for each treatment were normalised to U6 and data shown are mean mRNA expression levels expressed as a percentage of the expression in LPS +vehicle \pm SEM, n=3, ***P<0.001 vs. LPS +vehicle

Figure 1

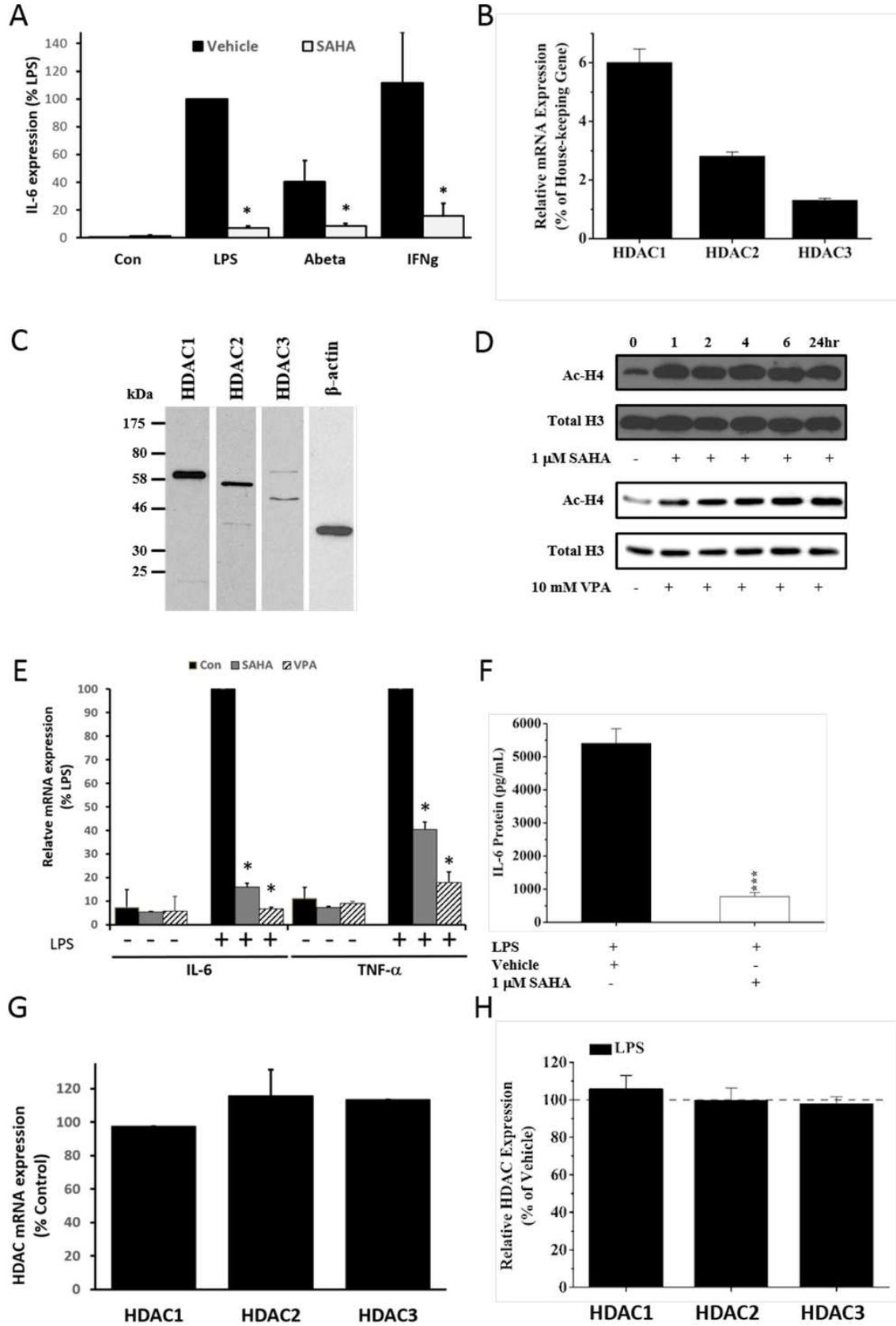


Figure 2

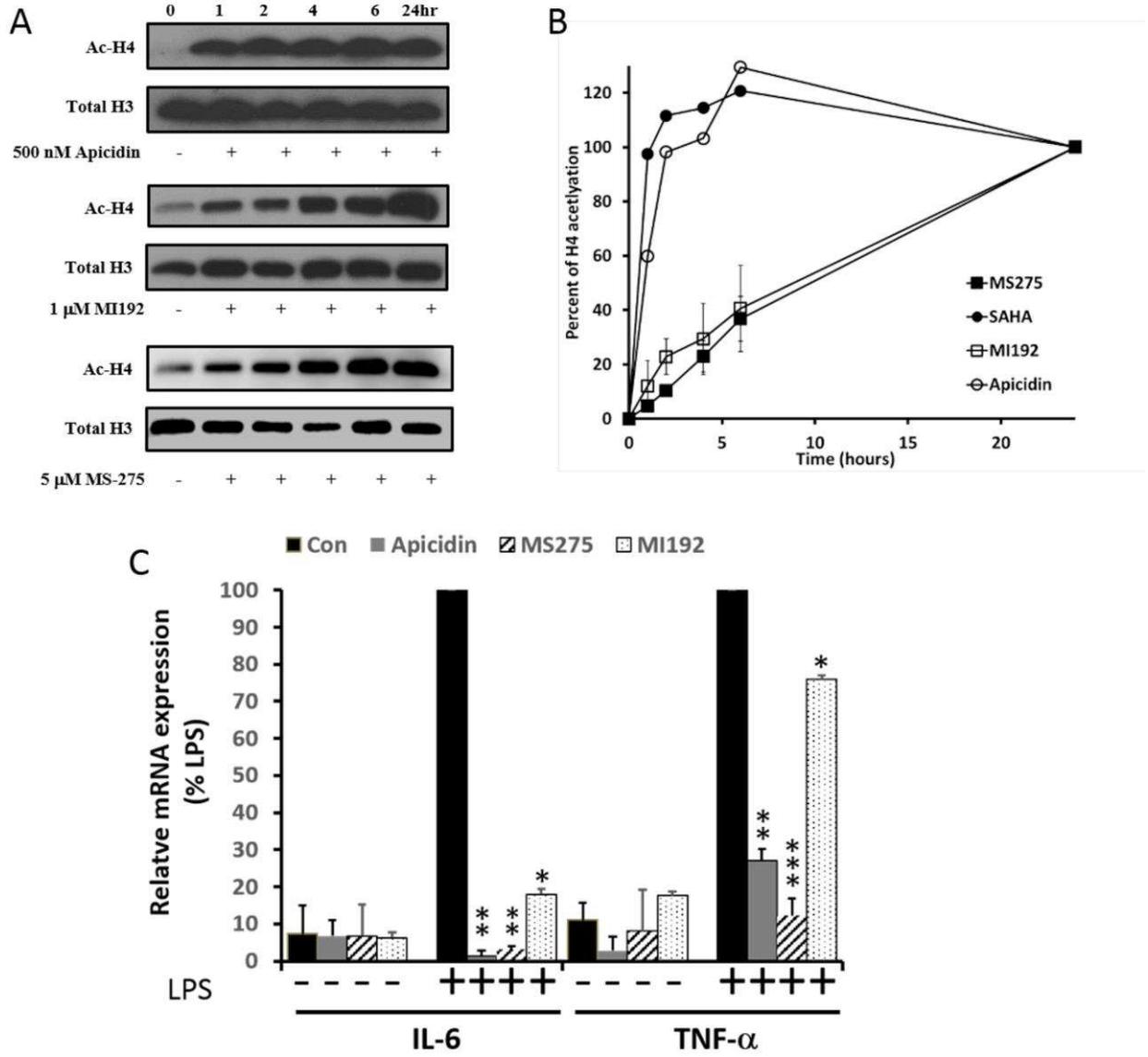
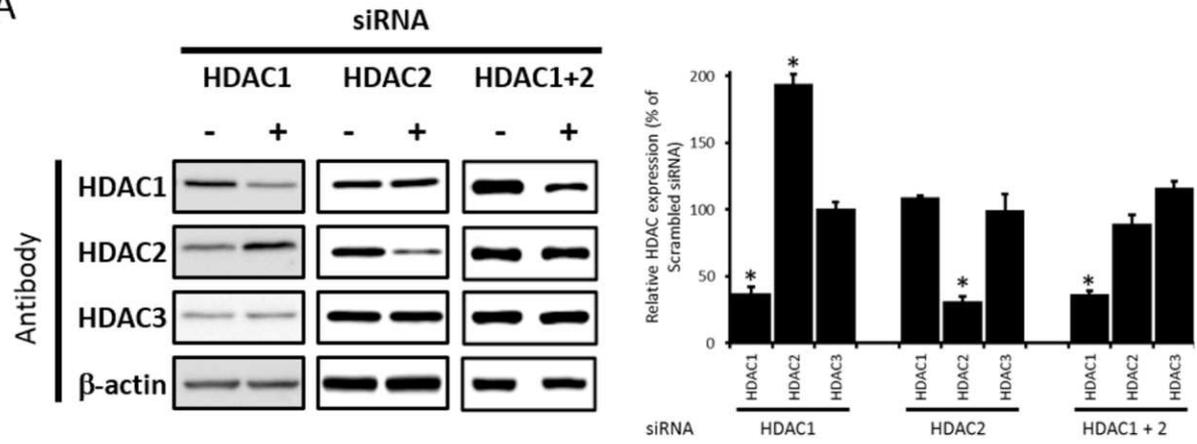


Figure 3

A



B

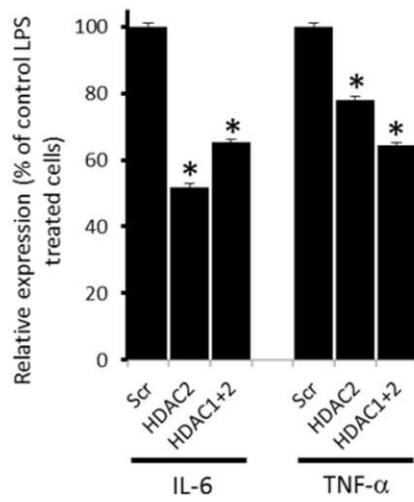


Figure 4

