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Supplementary Content

Fabrication, structure and properties of three-dimensional biodegradable poly(glycerol sebacate urethane) scaffolds

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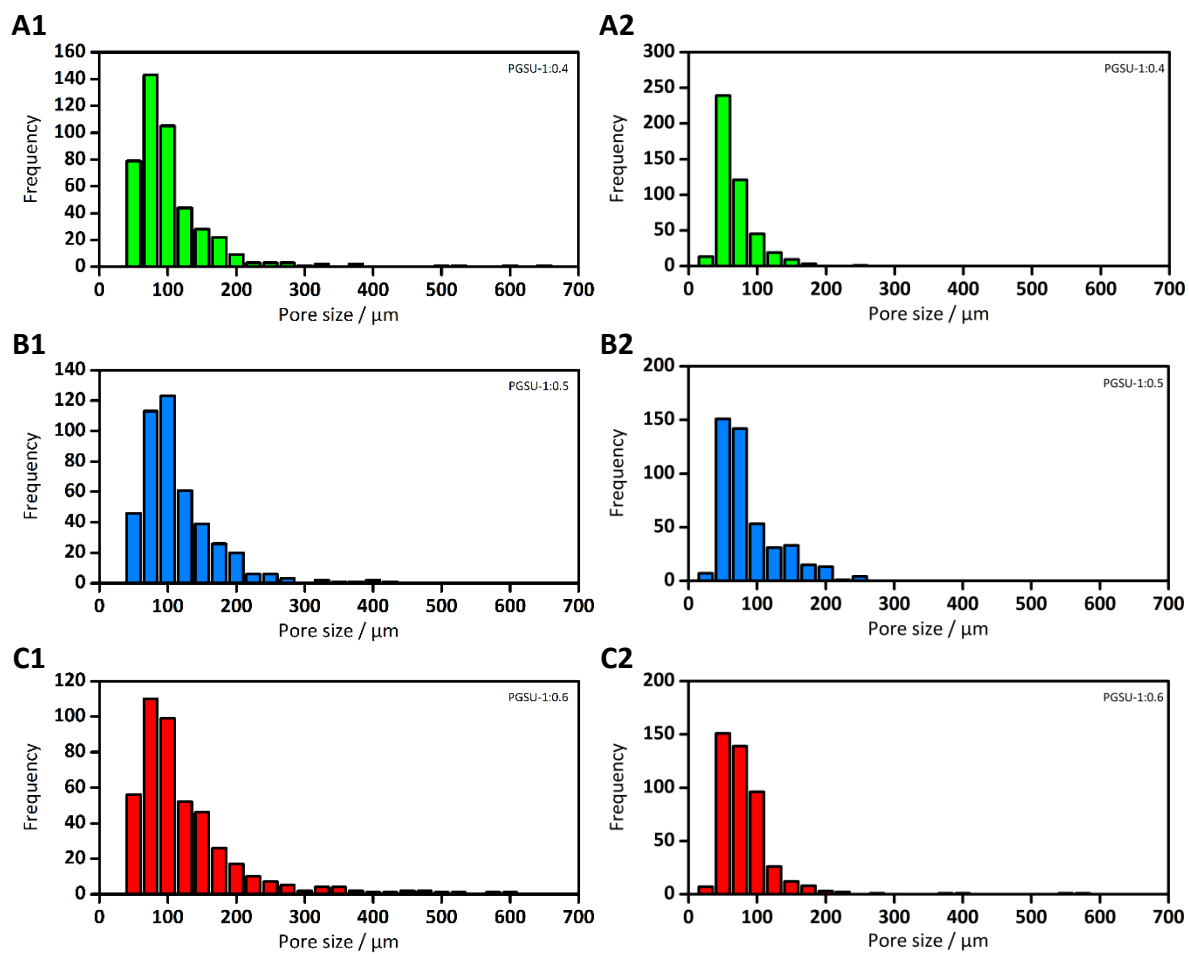


Figure S1. Histograms of the pore size distribution of as-prepared ((A1): PGSU-1:0.4, (B1): PGSU-1:0.5, (C1): PGSU-1:0.6) and cleaned and dry ((A2): PGSU-1:0.4, (B2): PGSU-1:0.5, (C2): PGSU-1:0.6) scaffold samples (n = 450).

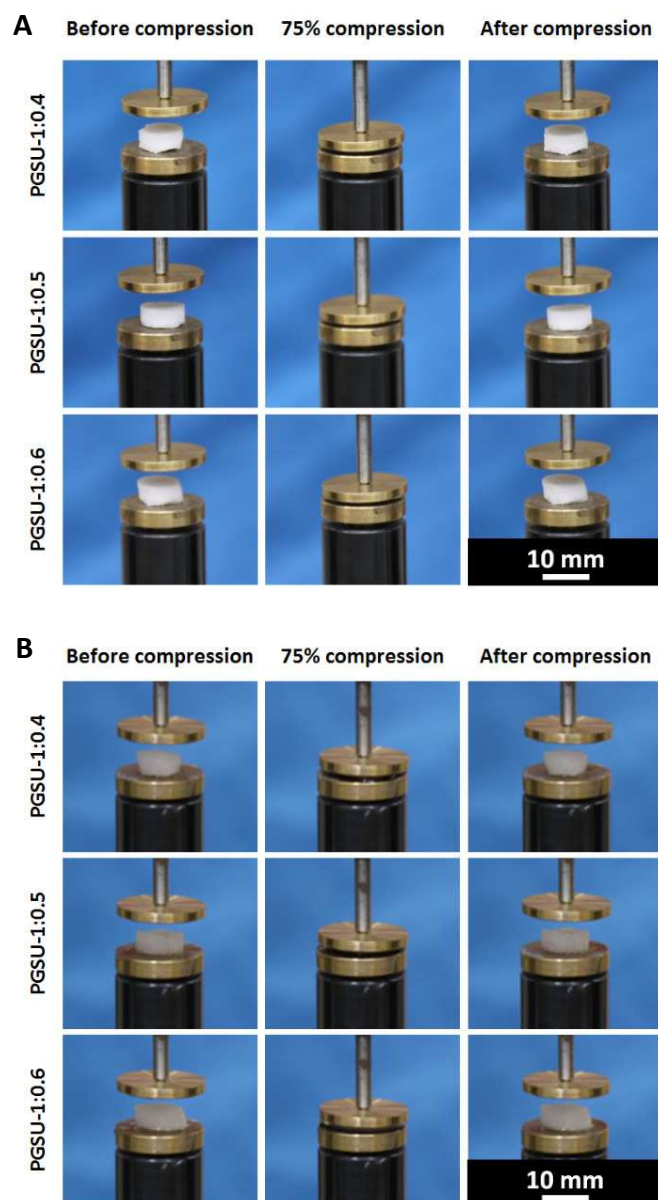


Figure S2. Images of the compressive behaviour of (A) dry and (B) hydrated PGSU scaffolds (24 h immersion in PBS solution at 37 °C and pH 7.4), illustrating the shape restorability to the original shape after releasing compression load.

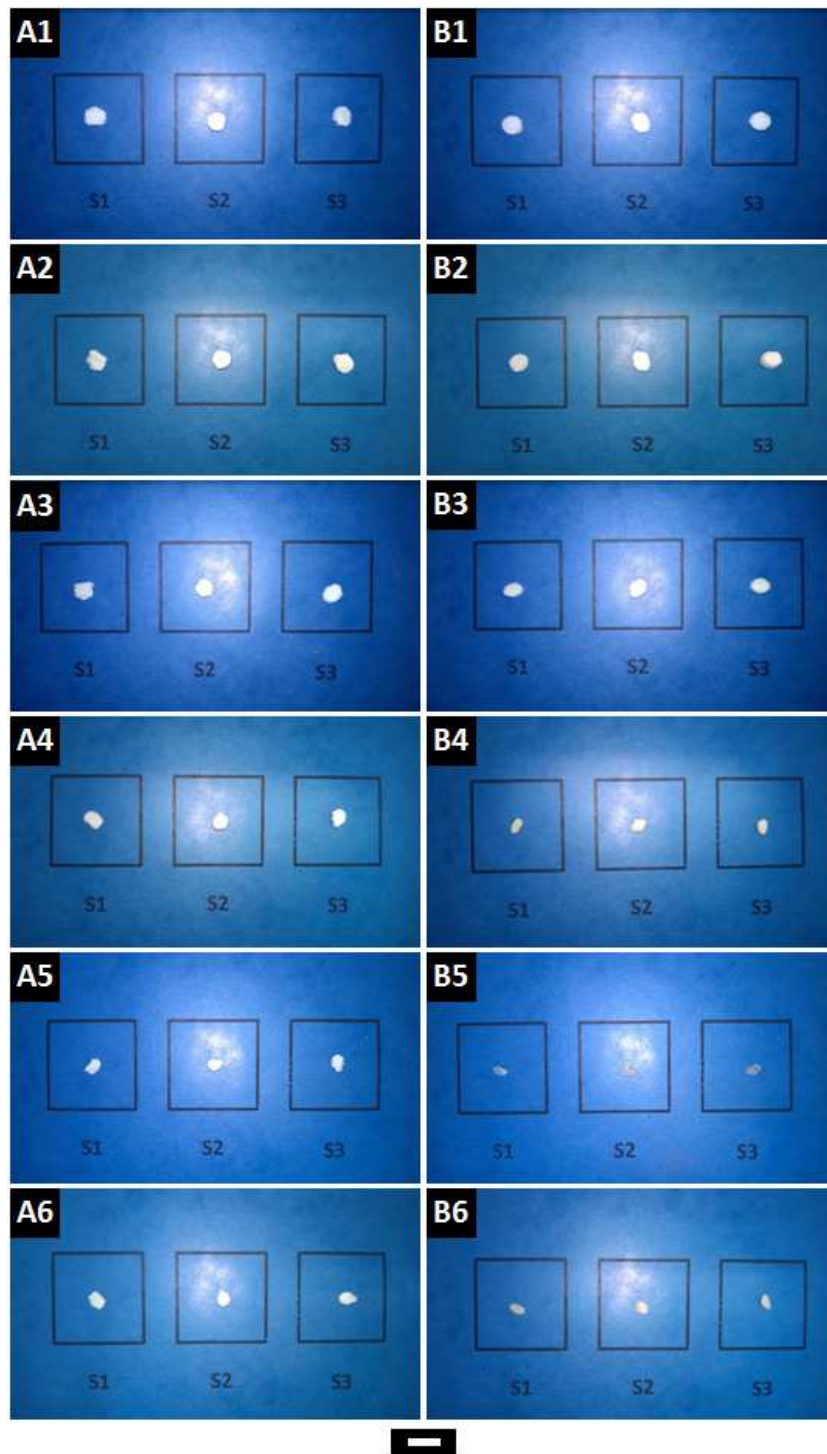


Figure S3. Pictures of PGSU-1:0.4 scaffold samples during incubation in PBS solution ((A1): Day 0; (A2): Day 7; (A3): Day 28; (A4): Day 56; (A5): Day 84; (A6): Day 112), and in PBS solution with the addition of lipase enzyme ((B1): Day 0; (B2): Day 7; (B3): Day 28; (B4): Day 56; (B5): Day 84; (B6): Day 112) under dynamic conditions for up to 112 days at 37 °C (scale bar: 1 cm), showing the degradation behaviour of the PGSU-1:0.4 scaffolds samples over time.

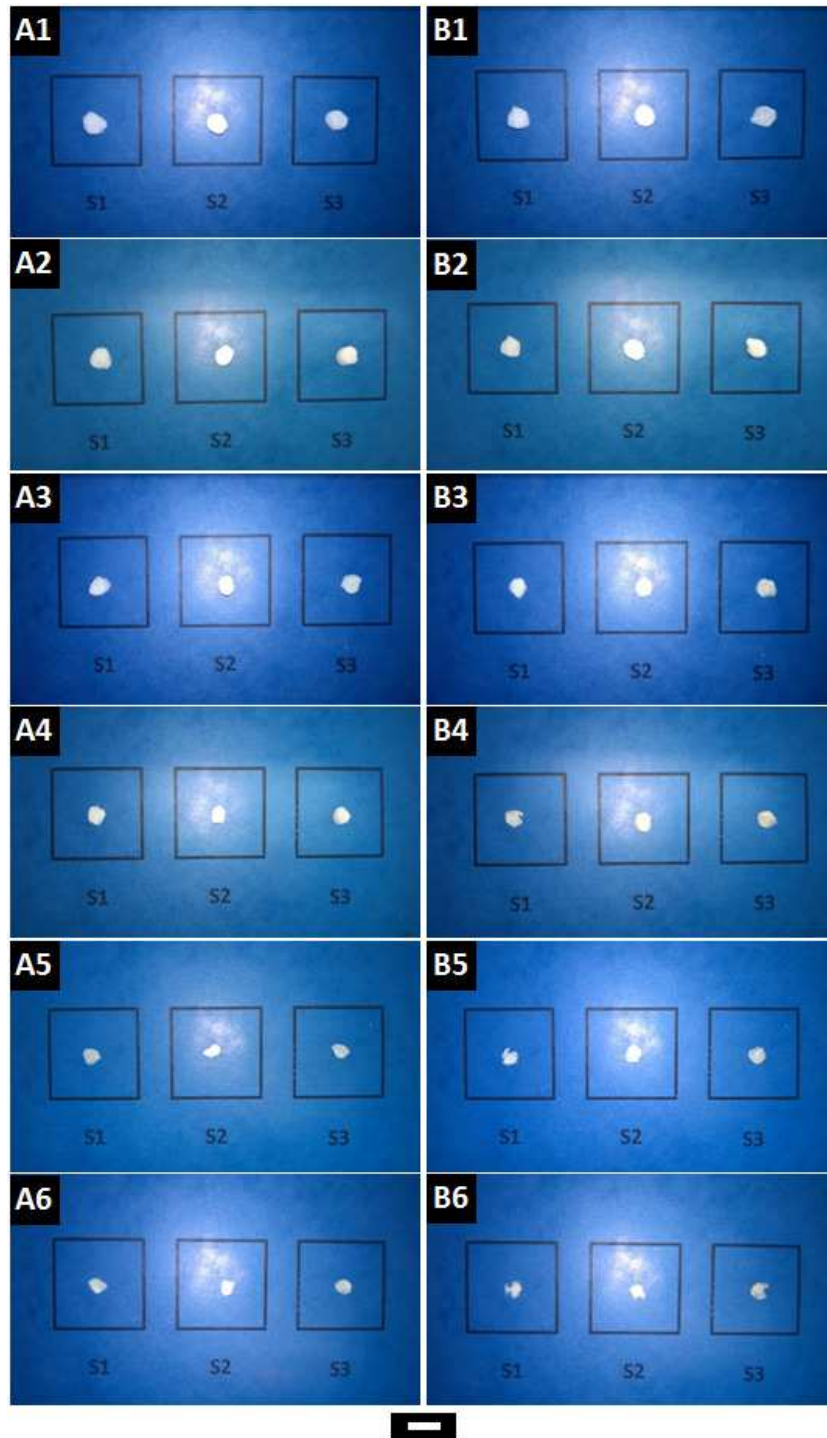


Figure S4. Pictures of PGSU-1:0.5 scaffold samples during incubation in PBS solution ((A1): Day 0; (A2): Day 7; (A3): Day 28; (A4): Day 56; (A5): Day 84; (A6): Day 112), and in PBS solution with the addition of lipase enzyme ((B1): Day 0; (B2): Day 7; (B3): Day 28; (B4): Day 56; (B5): Day 84; (B6): Day 112) under dynamic conditions for up to 112 days at 37 °C (scale bar: 1 cm), showing the degradation behaviour of the PGSU-1:0.5 scaffolds samples over time.

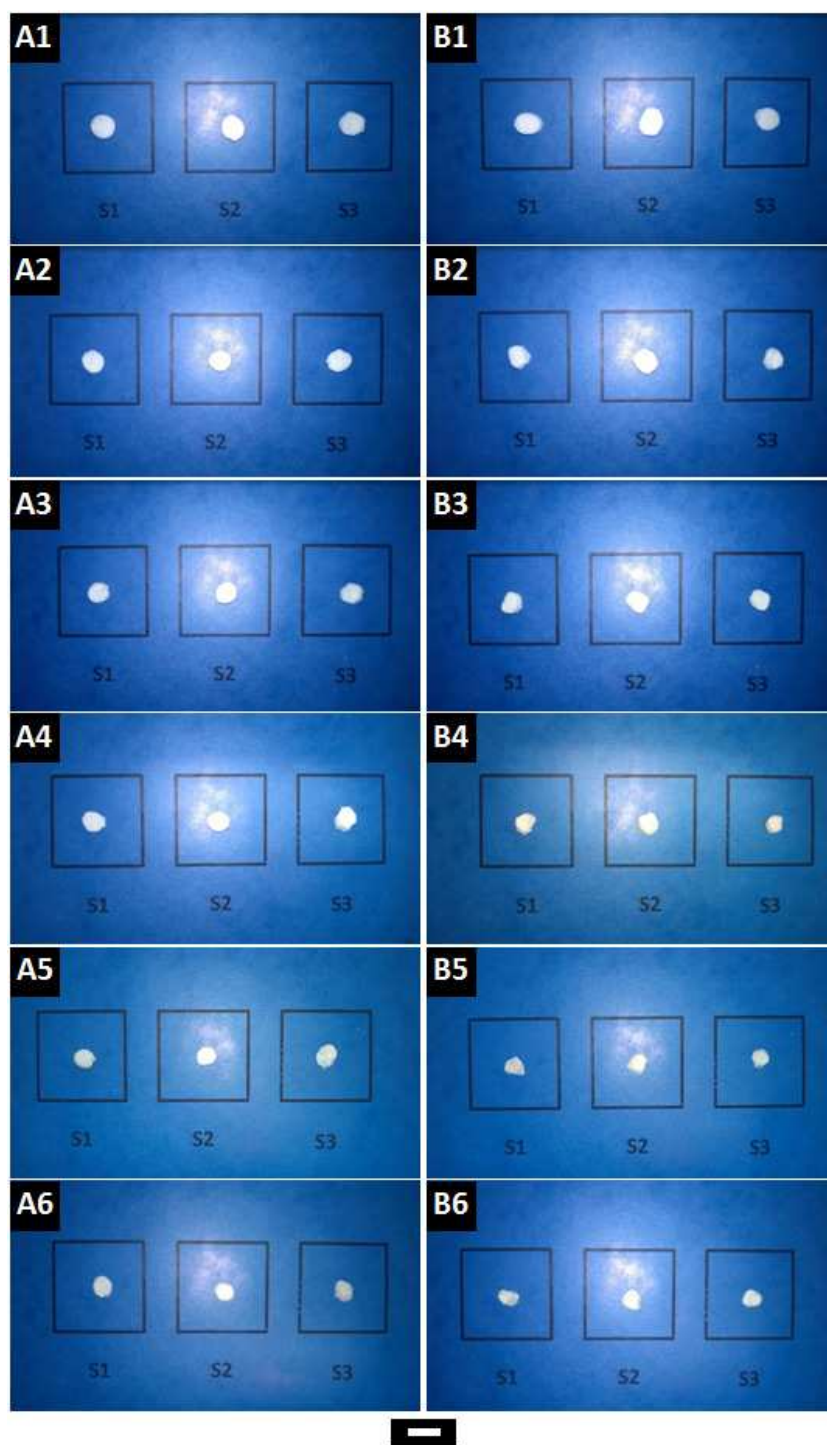


Figure S5. Pictures of PGSU-1:0.6 scaffold samples during incubation in PBS solution ((A1): Day 0; (A2): Day 7; (A3): Day 28; (A4): Day 56; (A5): Day 84; (A6): Day 112), and in PBS solution with the addition of lipase enzyme ((B1): Day 0; (B2): Day 7; (B3): Day 28; (B4): Day 56; (B5): Day 84; (B6): Day 112) under dynamic conditions for up to 112 days at 37 °C (scale bar: 1 cm), showing the degradation behaviour of the PGSU-1:0.6 scaffolds samples over time.

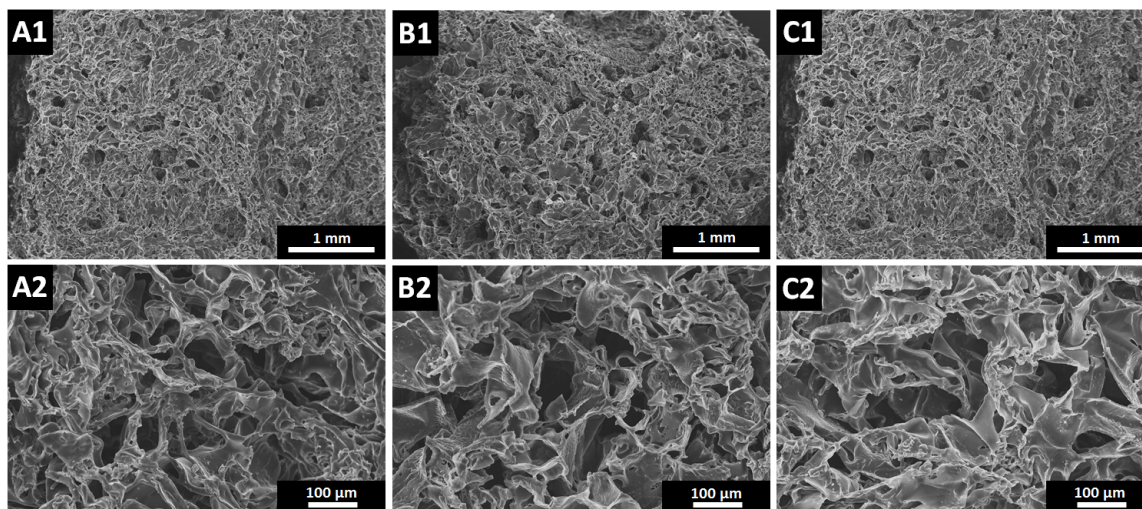


Figure S6. SEM micrographs of (A1-2) PGSU-1:0.4, (B1-2) PGSU-1:0.5 and (C1-2) PGSU-1:0.6 scaffolds after 34 days *in vitro* degradation tests in enzyme-free PBS solution at 37 °C.

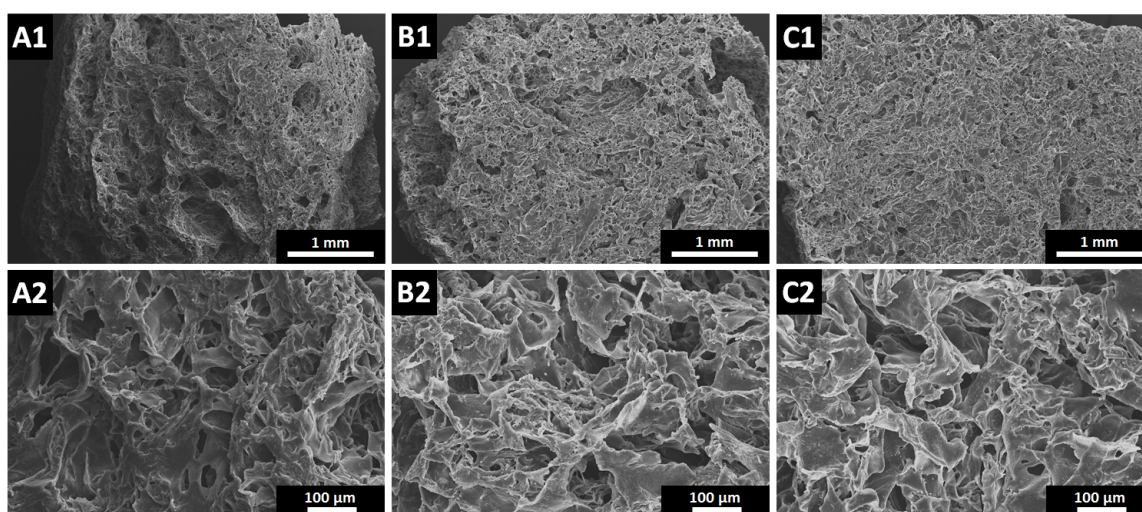


Figure S7. SEM micrographs of (A1-2) PGSU-1:0.4, (B1-2) PGSU-1:0.5 and (C1-2) PGSU-1:0.6 scaffolds after 34 days *in vitro* degradation tests in enzyme-containing PBS solution at 37 °C.

Table S1. Physical properties of selected polyester-based biopolymer scaffolds reported in the literature.

Polymer	Fabrication Method	Porosity, $P_f / \%$	Relative density, $\rho_r / (1-P_f)$	Compressive modulus, E_c / MPa	Reference
PGS/PLLA	FD ^a	92	0.08	0.014	[1]
PGS/PLLA	FD ^a	91	0.09	0.006	[1]
PLLA	FD ^a	93.0	0.070	4.7	[1]
PLLA	TIPS ^b	82.4	0.176	4.4	[2]
PLLA	TIPS ^b	81.3	0.187	7.5	[2]
PLLA	TIPS ^b	92.7	0.073	6.0	[3]
PLLA	SC/PL ^c	93.5	0.065	3.6	[4]
PLLA	SC/PL ^c	95.5	0.045	3.1	[4]
PLLA	SC/PL ^c	96.4	0.036	2.3	[4]
PLLA	SC/PL ^c	98.5	0.015	2.1	[4]
PLLA	TIPS ^b	87.0	0.130	1.8	[5]
PLLA	TIPS ^b	93.0	0.070	4.3	[6]
PLLA	SC/PL ^c	94.5	0.055	0.30	[7]
PLLA	SC/PL ^c	96.8	0.032	0.02	[7]
PLLA	SC/PL ^c	95.2	0.048	0.05	[7]
PLLA	SC/PL ^c	95.8	0.042	0.05	[7]
PLLA	SC/PL ^c	96.1	0.039	0.08	[7]
PDLLA	RM/PL	90.0	0.100	5.2	[8]
PDLLA	RM/PL	92.6	0.074	1.7	[8]
PDLLA	TIPS ^b	94.0	0.06	0.89	[9]
PDLLA	SC/PL ^c	93.0	0.070	2.4	[10]
PLGA	SC/PL	90.0	0.100	0.16	[11]
PLGA	GF/PL	90.0	0.100	0.29	[11]
PLGA	SC/PL ^c	97.0	0.030	0.25	[12]
PLGA	SC/PL ^c	93.0	0.070	2.0	[12]
PLGA	SC/PL ^c	92.0	0.080	3.0	[12]
PLGA	SC/PL ^c	91.5	0.085	3.5	[12]
PLGA	SC/PL ^c	87.0	0.130	7.5	[12]
PLGA	SC/PL ^c	80.0	0.200	12	[12]
PLGA	RM/PL	88.0	0.120	15	[13]
PLGA	RM/PL	88.0	0.120	7.5	[13]
PLGA	RM/PL	88.0	0.120	7.0	[13]
PLGA	RM/PL	88.0	0.120	5.5	[13]
PLGA	RM/PL	88.0	0.120	4.5	[13]
PCL	SC/PL ^c	74.0	0.260	0.40	[14]
PCL	SC/PL ^c	88.1	0.119	0.22	[15]
PCL	TIPS ^b	80.0	0.200	0.38	[16]
PCL	SC/PL ^c	76.0	0.240	4.3	[17]
PCL	SC/PL ^c	93.0	0.070	3.1	[18]
PCL	SC/PL ^c	65.0	0.350	1.2	[19]
PCL	TIPS ^b	89.0	0.110	0.08	[20]
PCL	TIPS ^b	88.0	0.120	0.19	[20]

^aFreeze-drying; ^bThermally induced phase separation; ^cSolvent casting/particulate leaching; RM/PL = Room temperature compression moulding/particulate leaching; GF/PL = Gas foamed/particulate leaching.

References

- [1] M. Frydrych, S. Román, S. MacNeil and B. Chen, *Acta Biomater.*, 2015, **18**, 40–49.
- [2] L. Budyanto, Y. Q. Goh and C. P. Ooi, *J. Mater. Sci. Mater. Med.*, 2009, **20**, 105–111.
- [3] R. Zhang and P. X. Ma, *J. Biomed. Mater. Res.*, 1999, **44**, 446–455.
- [4] C. Tu, Q. Cai, J. Yang, Y. Wan, J. Bei and S. Wang, *Polym. Advan. Technol.*, 2003, **14**, 565–573.
- [5] E. Nejati, V. Firouzdor, M. B. Eslaminejad and F. Bagheri, *Mater. Sci. Eng. C*, 2009, **29**, 942–949.
- [6] G. Wei and P. X. Ma, *Biomaterials*, 2004, **25**, 4749–4757.
- [7] P. X. Ma and J. W. Choi, *Tissue Eng.*, 2001, **7**, 23–33.
- [8] D. Jing, L. Wu and J. Ding, *Macromol. Biosci.*, 2006, **6**, 747–757.
- [9] J. J. Blaker, V. Maquet, R. Jérôme, A. R. Boccaccini and S. N. Nazhat, *Acta Biomater.*, 2005, **1**, 643–652.
- [10] L. Wu, H. Zhang, J. Zhang and J. Ding, *Tissue Eng.*, 2005, **11**, 1105–1114.
- [11] L. D. Harris, B. S. Kim and D. J. Mooney, *J. Biomed. Mater. Res.*, 1998, **42**, 396–402.
- [12] J. Zhang, H. Zhang, L. Wu and J. Ding, *J. Mater. Sci.*, 2006, **41**, 1725–1731.
- [13] L. Wu, J. Zhang, D. Jing and J. Ding, *J. Biomed. Mater. Res. A*, 2006, **76**, 264–271.
- [14] H. M. Wong, P. K. Chu, F. K. L. Leung, K. M. C. Cheung, K. D. K. Luk and K. W. K. Yeung, *Prog. Nat. Sci.*, 2014, **24**, 561–567.
- [15] M. J. Chern, L. Y. Yang, Y. K. Shen and J. H. Hung, *Int. J. Precis. Eng. Man.*, 2013, **14**, 2201–2207.
- [16] F. Naghizadeh, N. Sultana, M. R. A. Kadir, T. Muzaffar T. M. Shihabudin, R. Hussain and T. Kamarul, *J. Nanomater.*, 2014, Article ID 253185, 9 pages.
- [17] F. Wu, C. Liu, B. O'Neill, J. Wei and Y. Ngothai, *Appl. Surf. Sci.*, 2012, **258**, 7589–7595.
- [18] M. Diba, M. Kharaziha, M.H. Fathi, M. Gholipourmalekabadi and A. Samadikuchaksaraei, *Compos. Sci. Technol.*, 2012, **72**, 716–723.
- [19] J. J. Kim, R. K. Singh, S. J. Seo, T. H. Kim, J. H. Kim, E. J. Lee and H. W. Kim, *RSC Adv.*, 2014, **4**, 17325–17336.
- [20] P. Fabbri, V. Cannillo, A. Sola, A. Dorigato and F. Chiellini, *Compos. Sci. Technol.*, 2010, **70**, 1869–1878.