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Novel bioglasses for bone tissue repair and regeneration: Effect of glass design on sintering ability, ion release and biocompatibility

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ABSTRACT

Eight novel silicate, phosphate and borate glass compositions (coded as NCLx, where x = 1 to 8), containing different oxides (*i.e.* MgO, MnO₂, Al₂O₃, CaF₂, Fe₂O₃, ZnO, CuO, Cr₂O₃) were designed and evaluated alongside apatite-wollastonite (used as comparison material), as potential biomaterials for bone tissue repair and regeneration. Glass frits of all the formulations were processed to have particle sizes under 53 µm, with their morphology and dimensions subsequently investigated by scanning electron microscopy (SEM). In order to establish the nature of the raw glass powders, X-ray diffraction (XRD) analysis was also performed. The sintering ability of the novel materials was determined by using hot stage microscopy (HSM). Ionic release potential was assessed by inductively coupled plasma optical emission spectroscopy (ICP-OES). Finally, the cytotoxic effect of the novel glass powders was evaluated for different glass concentrations *via* a colorimetric assay, on which basis three formulations are considered promising biomaterials.

G R A P H I C A L A B S T R A C T



1. Introduction

The first reported use of a glass intended for bone tissue repair dates back to 1969, when Professor L. Hench proposed a composition in the Na₂O-CaO-SiO₂-P₂O₅ system, designated as bioglass 45S5 [1,2], commercially known as Bioglass[®].

Although Bioglass[®] proved to be an excellent material, considered for long time the gold standard bone substitute, it suffers from several

drawbacks. Specifically, the main difficulties are related to the material processing in form of 3D porous scaffolds, due to the limited ability of this glass in sintering [3]. Additionally, other weaknesses include: its slow degradation kinetic with the consequent difficulties to match the formation rate of new tissue, and the abrupt pH variations of the biological microenvironment, due to the increase in the concentration of ions such as Na⁺ and Ca²⁺, especially in the short term when the release is faster [4–6].

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 Table 1

 Rationale of the novel glass compositions.

| CODE | MAIN NETWORK FORMER | AIM |
|------|---------------------|--|
| NCL1 | SiO_2 | To develop a material with osteogenic properties, mainly determined by the presence of a high amount of silica. |
| NCL2 | SiO ₂ | To develop a load-bearing material with osteogenic properties and tailored degradation rate. |
| NCL3 | B_2O_3 | To develop a material with improved degradation rate and appropriate level of bioactivity as well as mechanical properties |
| NCL4 | B_2O_3 | To develop a material with tailored degradation rate and osteogenic effects. |
| NCL5 | P_2O_5 | To develop a resorbable glass with controlled degradation rate. |
| NCL6 | P_2O_5 | To develop a resorbable glass with controlled degradation rate, and improved mechanical strength |
| NCL7 | SiO ₂ | To develop a material with antibacterial properties, mainly determined by the presence of silver oxide, and a good level of bioactivity. |
| NCL8 | SiO ₂ | To develop a material with osteogenic properties and tailored degradation rate for non-load bearing applications. |

Worldwide many researchers have used the $SiO_2-Na_2O-CaO-P_2O_5$ system as a template for developing new silica-based compositions [7]. Subsequently, many formulations in the phosphate and borate-based system have been also designed to overcome the Bioglass[®] and silicatebased glass limitations [8–11], and thus to meet the set of requirements that are both crucial and necessary for optimised tissue-engineered substitutes [12].

The possibility to tailor glass properties by doping the main composition with network modifiers and/or intermediate oxides [13–20] offers significant potential for this class of biomaterials. In addition to promoting bone bonding, the release of soluble ions (*i.e.* Si, Ca, P and Na) from these glasses have been demonstrated to promote cell proliferation, differentiation and activate gene expression [20–24]. Furthermore, it has been also revealed that even slight changes in the glass main formulation can substantially affect the material behaviour, particularly the physico-chemical and mechanical properties, dissolution rate, bioactivity and bioresorbability [5,16,18,25–27].

However, there are still several criticisms related to the clinical use of this class of biomaterials in bone repair [12]. Firstly, whether or not glass dissolution products have a positive effect on adult stem cells is still an open debate [28]. Secondly, they have often proved inadequate when used in load-bearing bone defects, due to their low tensile strength and fracture toughness [29]. Ultimately, there are no large-scale porous bioactive glasses on the market, thus their commercial success as bone scaffolds is limited [6,12].

The aim of this work was the development and characterisation of eight novel silicate, phosphate and borate glass formulations (coded as NCLx, where x = 1 to 8), containing different oxides and in diverse molar percentages as promising biomaterials for the repair and regeneration of bone tissue.

2. Materials and methods

2.1. Development of novel glass formulations

Based on the current state of the art, the eight bioceramic formulations were developed using: silicon dioxide, phosphorous pentoxide and boron trioxide as network formers due to their promising bioactive potential [1,30,31], distinctive resorbable properties [32,33], and customable degradation rate [5,34], along with a range of different doping agents (*i.e.* MgO, MnO₂, Al₂O₃, CaF₂, Fe₂O₃, ZnO, CuO, Cr₂O₃), which were used to tailor the properties of the main composition [35–41]. The rationale and innovative characteristics of the novel materials are reported in Table 1. Additionally, considering the excellent biocompatibility either *in vitro* and *in vivo* of apatite wollastonite (AW) [42–44], and the fact that it has been adopted for a broad range of medical applications, either in the form of powder, porous structures or bulk material [45,46], AW glass-ceramic was used as comparison material in this study.

2.2. Glass production and processing

The novel glasses were produced and supplied by Glass Technology

Service (GTS) Ltd. (Sheffield, UK) along with AW. Briefly, the individual components (see Table 2) of each formulation were weighed out and then mixed together to obtain a uniform blend, which was subsequently melted in platinum crucibles at temperatures up to 1500 °C. The individual melts of glass were cast as solid blocks and then thermally shocked in de-ionised water to produce the precursor materials, known as frits.

Glass frits of all the compositions were ground in a one-bowl zirconia ball milling machine (Planetary Mono Mill Pulverisette 6, Fritsch GmbH, Germany) using a rotational speed of 400 rpm for 30 min, and then sieved using a mechanical sieve shaker (Impact Test Equipment Ltd., UK) to have a final particle size about 20 μ m and below 53 μ m.

Powders were prepared for pressing through mixing with an isopropanol solution (Sigma Aldrich, UK) in the proportion 1:3 (w/w). Powders were then pressed using an automatic hydraulic press (Specac-AtlasTM 8T, Specac Ltd., UK) to make 10 mm diameter and 2.5 mm high pellets. The pressed pellets were then sintered in a furnace (Carbolite 1200 CWF, Carbolite GmbH, Germany), with the sintering times and temperatures defined by the results of the hot stage microscopy analysis, reported in Section 2.3.2.

2.3. Physico-chemical characterisation

2.3.1. Microstructural characterisation

Powder glasses and dense pellets were sputtered with a thin layer of gold (approximately 10 nm, sputter time 40 s at 40 mA), and afterward analysed using a Philips XL30 Field-Emission Environmental Scanning Electron Microscope (ESEM FEG), which is fitted with a Rontec Quantax system for the Energy-Dispersive Spectroscopy (EDS) analysis. All the images were taken at an operation voltage of 20 kV, and working distance between 5 and 10 mm.

2.3.2. Hot stage microscopy (HSM)

The sintering ability of the novel glass powders was determined using hot stage microscopy (Misura[®], Expert System Solutions, Italy). Tests were performed in air using a heating rate of 10 °C/min up to 1200 °C. Glass powders were manually pressed into a small cylindrical die (2×3 mm) and placed on a $10 \times 15 \times 1$ mm alumina support. During the process the specimens were observed by a video camera and images of the changing sample profile were acquired up to 1450 °C. Afterwards, the sample shrinkage at different temperatures was calculated from the variation of the sample area, using the following formula:

shrinkage (%) =
$$\frac{A_T}{A_0} \times 100$$

where A_0 (mm²) was the initial area of the specimen at room temperature and A_T (mm²) was the area of the specimen at the temperature *T*.

2.3.3. XRD Analysis

To investigate the nature of the novel materials. XRD analysis was

| CODE | GLASS COMPOSITION (wt%) |
|------|--|
| NCL1 | 30.44SiO ₂ -9P ₂ O ₅ -6.29Na ₂ O-7.10CaO-9.54K ₂ O-5.55MgO-5.52MnO ₂ -5.16ZnO-13.14SrO-1.01CuO-5.91Bi ₂ O ₃ -2.02TeO ₂ -2.31V ₂ O ₅ |
| NCL2 | 36.90510 ₂ -9.70P ₂ 05-1.90B ₂ 03-3.39Na ₂ O-11.48CaO-3.85K ₂ O-4.41MgO-2.38MnO ₂ -6.97Al ₂ O ₃ -2.13CaF ₂ -10.92Fe ₂ O ₃ -0.41Li ₂ O-1.97MoO ₃ -1.52SeO ₂ -2.07Cr ₂ O ₃ |
| NCL3 | $20.03810_{2^{-3}}.79P_{8}0_{5^{-3}}.252B_{2}0_{3^{-4}}.97Na_{5}0-5.23CaO-6.27K_{3}O-2.69MgO-2.72M_{5}O_{3^{-3}}.20T10_{2^{-1}}0.67Fe_{2}O_{3^{-0}}.0.401i_{3}O-2.05BaO-1CoO-2.43V_{2}O_{3^{-2}}O_{$ |
| NCL4 | $16.28510_{2}-9.63P_{2}0_{5}-37.77B_{2}0_{3}-4.21Na_{5}0-3.80CaO-6.38K_{2}0-2.73MgO-5.52ZnO-7.03SrO-2.12CaF_{2}-1.08CuO-1.95M00_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95M0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CuO-1.95W0_{3}-1.51SeO_{2}-1.08CUO-1.95W0_{3}-1.51SeO_{2}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.51SeO_{2}-1.08CUO-1.95W0_{3}-1.08CUO-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.95W0_{3}-1.08CUO-1.08CUO-1.08CUU-$ |
| NCL5 | 60.28P_05_4 09Na20-5.32K20-5.32K20-3.80Mg0-3.84Zn0-4.10Mn02-9.77Sr0-0.75Ti02-2.75Sb203 |
| NCL6 | 4.61SiO ₂ -68.19P ₂ O ₅ -6.68B ₂ O ₃ -4.76Na ₂ O-5.38CaO-2.71K ₂ O-1.55MgO-1.50CaF ₂ -1.67MnO ₂ -0.76CuO-0.72CoO-1.46Cr ₂ O ₃ |
| NCL7 | 39.96SiO ₂ -9.46P ₂ O ₅ -12.39Na ₂ O-11.19CaO-2.50K ₂ O-1.61MgO-15.44AgO-2.13TiO ₂ -4.26Fe ₂ O ₃ -1.06CuO |
| NCL8 | 38.93510z-10.24P205-2.01B203-8.94Na50-12.11CaO-6.78K20-2.91MgO-6.27MnOz-1.17ZnO-2.30Fe203-1.495rO-2.81CaFz-0.57CuO-0.54CoO-1.04MoO3-0.80SeOz-1.10Cr203 |
| AW | 4.6 MgO-44.7-CaO-34SiO ₂ -16.2P ₂ O ₅ -0.5 CaF ₂ |
| | |

Composition of the novel glass formulations

Table 2

performed using a PANalytical X'Pert Pro MPD, powered by a Philips PW3040/60 X-ray generator, and fitted with an X'Celerator detector. Diffraction data was acquired by exposing powder samples to Cu-K_{α} X-ray radiation, at 40 kV and 40 mA. The data were collected over a 2 θ range between 5 and 80°, with a step size equal to 0.0334°, a counting time per step of 200 s using the scanning X'Celerator detector. Phase identification was carried out using the PANalytical X'Pert HighScore Plus© software.

2.3.4. Ion leaching evaluation

Un-sintered glass powders with a concentration of 10 mg/ml were immersed in deionised water (Veolia Water Technologies, UK) and incubated under an atmosphere of 5% CO_2 and 95% air at 37 °C. After each storage period (1, 3, 7, 14 and 28 days), the specimens were removed *via* filtration, and filtrates retained to analyse the ion release potential of each compositions. An inductively coupled plasma optical emission spectroscopy (ICP-OES) (Specto-Ciros-Vision, Sheffield University, UK), which allows simultaneous multi-element analysis following the calibration of the instrument by standards of known concentrations of the elements of interest, was employed.

2.4. Biological characterisation

The *in vitro* cytotoxicity of the novel glass formulations was evaluated according to ISO 10993–5 [47] using rat calvaria osteoblast cells in indirect contact with glass powders up to 7 days. Each powder sample was firstly sterilised using 100% ethanol solution, and after added to Dulbecco's modified Eagle medium (DMEM, Gibco[®] UK) at three different concentrations (0.1, 1 and 10 mg/ml) and incubated for 24 h at 37 °C. After incubation, the glass-conditioned medium was filtered through a 0.22 μ m microbiological filter, and used for *in vitro* tests.

Cells at early passages were provided by Institute of Cellular Medicine (Medical School, Newcastle University, UK), and then were cultured in T75 flasks at 37 °C in a humidified incubator with 5% CO₂, using DMEM supplemented with 10% fetal bovine serum (FBS), 1% penicillin-streptomycin and 1% glutamine (Gibco[®], UK). Cells were seeded at a density of 1×10^4 cells/well in 96-well plates and incubated at 37 °C. After 24 h culture period the culture media was discarded and replaced with the filtered solution for indirect cytotoxicity testing. Rat OBs cultured in the absence of glass powders were used as negative control.

The culture plates were then incubated for 1 and 7 days. The cytotoxic effect was measured exposing each well to 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT, Sigma–Aldrich, UK) solubilisation at a concentration of 0.5 mg/ml. 100 μ l of MTT solution was added to each well and incubated at 37 °C for another 4 h. MTT was taken up only by active cells and reduced in their mitochondria to insoluble purple formazan granules. The medium was then removed and 100 μ l of dimethyl sulfoxide (DMSO) was added to dissolve the precipitated formazan. The absorbance of the solution was evaluated spectrophotometrically at a wavelength of 570 nm, and acquired using a Sunrise microplate reader (Tecan Group Ltd., Switzerland).

The absorbance values from three replicates were averaged and statistically analysed using two–way analysis of variance (ANOVA) followed by Bonferroni *post hoc* analysis. *P*-values < 0.05 were considered statistically significant.

3. Results

3.1. Glass production and powder processing

During synthesis, composition NCL5 could not form a glass at any temperature up to 1500 $^{\circ}$ C, which was the highest temperature the available furnaces could reach. Glass frits from the seven compositions



Fig. 1. SEM analysis (magnification 1500 ×) showing the glass powders morphology: a) NLC1, b) NCL2, c) NCL3, d) NCL4, e) NCL6, f) NCL7, g) NCL8, and h) AW.

which did form a glass, were ground using a zirconia ball mill, and sieved to obtain fine powder in the range 0-53 $\mu m.$

3.2. Glass powder microstructural analysis

Fig. 1(a–h) shows the morphology of as-synthesised glass powders. Non-spherical and irregular shape particles with sharp edges can be observed for all the compositions. Furthermore, it can be seen that for all the glasses most of the particles were very fine (ranging from 20 μ m to 53 μ m), with the presence also of grains smaller than 10 μ m, which

tended to compact producing aggregates.

3.3. HSM

The experiments started at room temperature (T_R) with heating rate of 10 °C/min up to 1450 °C. All the specimens maintained their initial rectangular shape before the first shrinkage temperature (T_{FS}), which varied between 550 °C and 1225 °C (see Fig. 2(a–h)). For temperatures higher than the corresponding T_{FS} , the samples started to shrink until the temperature of maximum shrinkage (T_{MS}). The variation of sample



Fig. 2. Shrinkage profile derived from hot stage microscopy as function of temperature for: a) NCL1, b) NCL2, c) NCL3, d) NCL4, e) NCL6, f) NCL7, g) NCL8 and h) AW.

dimensions during the sintering process are reported in Fig. 2(a–h). Fig. 2(a–b, f–g) reveal also how the silicate-based specimens (NCL1, NCL2, NCL7 and NCL8), before reaching the melting status at complete melting temperature (T_{CM}), started to expand up to their temperature of maximum volume (T_{MV}). Furthermore, these silicate-based glasses

showed a similar thermal profile, whereas the phosphate-based (NCL6) had a thermal curve (Fig. 2(e)) more comparable to the borate-based glasses (NCL3 and NCL4) (Fig. 2(c-d)).



Fig. 3. XRD patterns of as-synthesised glass powders (\blacktriangle hydroxylapatite, $\blacksquare \beta$ -wollastonite, \blacklozenge silver \bullet calcium sodium phosphate).

3.4. XRD

The XRD patterns of the raw materials are reported in Fig. 3. For NCL1, NCL2, NCL3, NCL4, and NCL8 the presence of a broad peak, common for glass samples, indicated the completely amorphous nature of these compositions. The amorphous peak was detected at 20 values between 25° and 30°, and confirmed that NCL1, NCL2, NCL3, NCL4 and NCL8 formulations were free from any detectable crystalline phase. Different patterns were detected for NCL6 formulation, which showed a glass-ceramic nature with a crystalline phase identified as calcium sodium phosphate (ICDD ref. code 01-074-1950). Moreover, for the NCL7 composition a silver crystalline phase (ICDD ref. code 04-003-1425) was detected. Regarding the AW composition, the presence of hydroxylapatite phase (ICDD ref. code 01-080-6260) and a less intense β -wollastonite phase (ICDD ref. code 04-010-0710) were observed, proving the glass-ceramic structure of this material.

3.5. Ion leaching evaluation

The ionic release of the common elements (Si, P, B, Ca and Mg) of each glass composition is reported in Fig. 4. It is interesting to observe that, except for NCL2 composition, the amount of silicon released in solution after 28 days was proportional to the molar content present in the main formulation. As network former, boron was released very quickly in comparison to silicon and phosphorous, and it displayed a progressive increase over the time period for both NCL3 and NCL4 compositions. Moreover, it was found that the amount of phosphorous, as main network former, in the NCL6 composition was released more slowly than boron from NCL3 and NCL4 formulations.

3.6. Bioceramic pellets sintering and characterisation

Sintering was performed with a heating rate of 10 $^{\circ}$ C/min up to a maximum temperature in the range 550–850 $^{\circ}$ C, which was held for 1 h; after this the samples were left to cool down at room temperature. Sintering temperatures were selected on the basis of the HSM results. SEM investigations were conducted to evaluate the effect of different maximum temperatures on sample morphology. Fig. 5 reports the pellet surface of each composition for two heat treatments per material. Micrographs (a) of Fig. 5 show a poor sintering level for all the

formulations, where glass particles started to aggregate, but were not properly sintered. The microstructures reported in micrographs (b) of Fig. 5 demonstrate how an increase in the heating temperatures led to an appropriate densification status, as a result of increased liquid phase. Particularly for NCL1, NCL3, NCL6, NCL7, NCL8 and AW compositions the formation of sintering necks (red arrows in the figure) became evident.

In light of the SEM microstructural observations, and after a thorough qualitative analysis, a summary of the heating temperatures that qualitatively were considered to provide the best densification degree are shown in Table 3.

3.7. Biocompatibility assay

The MTT results are summarised in Fig. 6. For NCL1 the cells displayed good metabolic activity after 1 day in culture for all concentrations tested, however the effect of the glass became detrimental after 7 days in culture at a concentration of 10 mg/ml. NCL2 shows not meaningful positive effects on metabolic activity after day 1, with no significant negative effect compared to control at day 7. The NCL3, NCL6 and NCL8 formulations all show cytotoxic effects on cell mitochondrial activity after 7 days in culture, which increases with increasing quantities of glass powder. NCL4 shows a broadly uniform response over the time period, with no significant differences in response across concentrations or time points. The NCL7 composition showed a positive effect on cell viability and proliferation at both time points, being this statistically significant for the lowest concentration after 7 days in culture. For AW glass-ceramic, the findings of this study confirmed the results of previous work [48], according to which the material initially has a slightly cytotoxic effect on cell mitochondrial activity, followed by enhanced cell proliferation.

4. Discussion

For this study eight novel bioceramic formulations were designed in order to develop biomaterials with beneficial performance towards bone tissue repair and regeneration, by using *i*) silicon dioxide, *ii*) phosphorous pentoxide and *iii*) boron trioxide as network formers.

The glasses were synthetized *via* a melting-quenching route; however, during the glass production their forming ability was composition dependant. Even though both NCL5 and NCL6 are phosphate-based glasses, the NCL5 formulation did not form a liquid at 1500 °C. This contradicts the commonly held view that phosphate-based glasses can be prepared at relatively low temperatures [32]. Based on the findings reported by Abou Neel et al., the incorporation of high density oxides like SrO₂ in the phosphate glass structure might be associated with an increase of the glass melting temperature; and this is considered the most likely reason for the poor processability of the NCL5 formulation [49].

HSM analysis revealed that the silicate-based glasses (NCL1, NCL2, NCL7 and NCL8) displayed similar thermal profiles, characterised by an increase in sample dimensions after the maximum shrinkage temperature and before the melting onset occurred. These results are comparable with the findings reported by Baino et al., who found that a less complex silicate-based glass (CEL2) exhibited a significant volumetric expansion after the first densification step [14]. Within the scope of the formulations reported in Table 2, we conclude that the silicate glass sintering profiles are relatively insensitive to the presence of different network modifiers in the glass structure.

HSM analysis performed on NCL6 phosphate-based glass showed a thermal profile with densification that takes place in two steps (elbow-shape profile). Furthermore, similarly to ICEL2 phosphate glass (45% P_2O_5 , 3% SiO₂, 26% CaO, 7% MgO, 15% Na₂O and 4% K₂O) [50], NCL6 formulation displayed an HSM thermograph with a volume increase at temperatures higher than T_{MS} . This observation is consistent with the findings reported by Arstila et al., according to which the elbow shape



Fig. 4. Ionic concentrations of Si, P, B, Ca and Mg released into deionised water from all the formulations, without refreshing the solutions and at different time points (1, 7, 14 and 28 days).

curve could indicate the possible sintering interval of the glass [51,52]. Moreover, the presence of network modifiers such as MnO_2 , CaF_2 , CuO, CoO, Cr_2O_3 , and in particular B_2O_3 could have affected the thermal behaviour of this formulation, leading to a shrinkage profile more comparable to the one developed by the borate-based compositions.

The sintering behaviour of borate-based bioglasses has to date been explored in less depth than silicate or phosphate based glasses. In this study, the two borate-based glasses (NCL3 and NCL4) exhibited similar one-step densification behaviour, with a small sintering interval in comparison to the silicate and phosphate-based glasses [53].

The selection of sintering temperatures is a key step during the manufacturing process to consolidate ceramic-based structures [54,55]. Although a further optimisation process (based on thermal treatment and morphological analysis evaluation) was required, the outcomes from the HSM were a very useful guide in predicting the optimal

sintering temperatures of bioceramic pellets: the temperatures that led to optimal consolidated structures, as demonstrated by SEM investigation, were all in the range of sintering temperatures provided by HSM [56].

According to the ICP-OES results, and by comparing the release of the main network formers, boron was released very quickly in comparison to silicon and phosphorus, therefore proving the highly reactive nature of borate-based glasses [5]. These findings reflect the study design (Table 1), which aimed to develop compositions with tailored degradation rates. It is also notable that the presence of boron in the glass structure, even as intermediate oxide, enhanced the release of the other elements (*i.e.* phosphorous release from NCL3 glass with respect to NCL1 composition) in the formulation.

For all the novel glasses, apart from the NCL2 composition, silicon leaching proved to be proportional to its molar content in the parental



Fig. 5. SEM micrographs of bioceramic pellet surface at (a) low and (b) appropriate sintering level (temperature = °C, magnification = 2500 ×); red arrows indicate necking formation phenomena. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

Table 3

Sintering intervals of the novel glass formulations (derived from HSM) and optimal sintering temperatures of dense bioceramic pellets.

| Code | Composition | HSM sintering interval (°C) | Sintering temperature (°C) |
|------|-----------------|--------------------------------|-------------------------------|
| NCL1 | Silicate-based | 575–785 | 625 |
| NCL2 | Silicate-based | 600–730 | 700 |
| NCL3 | Borate-based | 555-625 | 625 |
| NCL4 | Borate-based | 550-650 | 625 |
| NCL6 | Phosphate-based | 580–775 | 725 |
| NCL7 | Silicate-based | 575–785 | 625 |
| NCL8 | Silicate-based | 500–730 | 625 |
| AW | Silicate-based | 800–1225 | 850 |

glass. Additionally, it was found that the release of phosphorus increased with soaking time only for the NCL6 formulation, in which it was used as network former, most likely as a result of the higher levels of phosphorous in NCL6 [32,57].

The results of the *in vitro* cytotoxicity tests revealed that for high concentrations of glass powder (10 mg/l), cell mitochondrial activity was significantly reduced for NCL1, NCL3, NCL6 and NCL8 compositions [58], meaning that the NCL2, NCL4 and NCL7 formulations are suitable for further study. No single effect provided a direct correlation between the cytotoxicity and ionic release potential, but a number of observations can be made:

• For both the NCL1 and NCL3 formulations, it is most likely that the

release of vanadium had a negative effect on cell mitochondrial activity [59,60].

- The NCL6 composition did not contain many dopants, and because the release of Co, Cu and Cr after 28 days in immersion was quite low (data not shown) [61], it is most likely that the high levels of phosphorus and boron were the cause of the detrimental effect of this glass.
- No clear reasons for the toxicity of NCL8 were found; hence, the negative effect is most likely due to the combination of the different oxides.
- The highly reactive nature of the NCL4 borate-based glass enabled the release of strontium at a higher rate than NCL1 composition, which has been demonstrated to increase proliferation as well as differentiation of osteoblast cells [62,63].
- The positive effect of the NCL7 formulation might derive from the combined effect of titanium, iron and copper, which enhance osteoblasts proliferation and activity [64].

5. Conclusion

This paper has presented a systematic process for the evaluation of novel bioceramic materials, considering basic glass processability, hot stage microscopy, ICP analysis and a biocompatibility assay. The main conclusions which can be drawn are:

(i) It is possible for phosphate-based glasses to have high melting temperatures, as the NCL5 formulation did not form a liquid at



Fig. 6. Effect of a) NCL1, b) NCL2, c) NCL3, d) NCL4, e) NCL6, f) NCL7, g) NCL8 and h) AW glass powders (measured in triplicate) on formazan formation after indirect contact with rat osteoblast cells, evaluated through MTT assay after 1 day and 7 days in culture. Error bars represent the standard deviation (* = P < 0.05).

1500 °C.

- (ii) The sintering range identified by HSM consistently included the optimum sintering temperature, indicating the usefulness of this technique in defining sintering routines.
- (iii) No direct correlation between ion release and cytotoxicity could be observed across all of the glasses studied, although silicon release seems to be mostly dependent on silicon content; and vanadium

does not seem to have enhanced biocompatibility at the concentrations studied in this paper.

(iv) Of the eight proposed glasses the NCL2, NCL4 and NCL7 formulations have shown sufficient biocompatability to merit further study as new biomaterials.

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References

- [1] L.L. Hench, Bioceramics, J. Am. Ceram. Soc. 81 (1998) 1705–1728.
- [2] L.L. Hench, J. Wilson, An Introduction to Bioceramics, World Scientific, Singapore, 1993.
- [3] L.C. Gerhardt, A.R. Boccaccini, Bioactive glass and glass-ceramic scaffolds for bone tissue engineering, Materials 3 (7) (2010) 3867–3910.
- [4] H. Fu, et al., Long-term conversion of 45S5 bioactive glass-ceramic microspheres in aqueous phosphate solution, J. Mater. Sci. Mater. Med. 23 (2012) 1181–1191.
- [5] W. Huang, et al., Mechanisms for converting bioactive silicate, borate, and borosilicate glasses to hydroxyapatite in dilute phosphate solution, Phys. Chem. Glasses Eur. J. Glass Sci. Technol. B 47 (6) (2006) 647–658.
- [6] M.N. Rahaman, et al., Bioactive glass in tissue engineering, Acta Biomater. 7 (6) (2011) 2355–2373.
- [7] V. Krishnan, T. Lakshmi, Bioglass: A novel biocompatible innovation, J. Adv. Pharm. Technol. Res. 4 (2) (2013) 78–83.
- [8] M.T. Souza, et al., New highly bioactive crystallization-resistant glass for tissue engineering applications, Trans. Mater. Res. 4 (1) (2017) 014002.
- [9] W.C. Lepry, S.N. Nazhat, Highly bioactive sol-gel-derived borate glasses, Chem. Mater. 27 (13) (2015) 4821-4831.
- [10] C. Tan, et al., Structural, thermal and dissolution properties of MgO- and CaOcontaining borophosphate glasses: effect of Fe₂O₃ addition, J. Mater. Sci. 52 (12) (2017) 7489–7502.
- [11] F. Baino, et al., Design, selection and characterization of novel glasses and glassceramics for use in prosthetic applications, Ceram. Int. 42 (1) (2016) 1482–1491.
- [12] L.L. Hench, J.R. Jones, Bioactive glasses: frontiers and challenges, Front. Bioeng. Biotechnol. 3 (2015).
- [13] S. Murphy, et al., The effect of composition on ion release from Ca-Sr-Na-Zn-Si glass bone grafts, J. Mater. Sci. Mater. Med. 20 (11) (2009) 2207–2214.
- [14] F. Baino, et al., Optimization of composition, structure and mechanical strength of bioactive 3-D glass-ceramic scaffolds for bone substitution, J. Biomater. Appl. 27 (7) (2013) 872–890.
- [15] G. Kaur, et al., A review of bioactive glasses: their structure, properties, fabrication, and apatite formation, J. Biomed. Mater. Res. A 102 (1) (2013) 254–274.
- [16] Q. Fu, et al., Silicate, borosilicate, and borate bioactive glass scaffolds with controllable degradation rate for bone tissue engineering applications. II. In vitro and in vivo biological evaluation, J. Biomed. Mater. Res. A 95 (1) (2010) 172–179.
- [17] Q. Fu, et al., Silicate, borosilicate, and borate bioactive glass scaffolds with controllable degradation rate for bone tissue engineering applications. I. Preparation and in vitro degradation, J. Biomed. Mater. Res. A 95 (1) (2010) 164-171.
- [18] S. Murphy, et al., The effect of ionic dissolution products of Ca-Sr-Na-Zn-Si bioactive glass on in vitro cytocompatibility, J. Mater. Sci. Mater. Med. 21 (10) (2010) 2827–2834.
- [19] H.B. Pan, et al., Strontium borate glass: potential biomaterial for bone regeneration, J. R. Soc. Interface 7 (48) (2010) 1025–1031.
- [20] A. Hoppe, N.S. Guldal, A.R. Boccaccini, A review of the biological response to ionic dissolution products from bioactive glasses and glass-ceramics, Biomaterials 32 (2011) 2757–2774.
- [21] J.M. Kanczler, R.O. Oreffo, Osteogenesis and angiogenesis: the potential for engineering bone, Eur. Cell. Mater. 15 (2008) 100–114.
- [22] I.D. Xynos, et al., Ionic products of bioactive glass dissolution increase proliferation of human osteoblasts and induce insulin-like growth factor II mRNA expression and protein synthesis, Biochem. Biophys. Res. Commun. 276 (2000) 461–465.
- [23] I.D. Xynos, et al., Gene-expression profiling of human osteoblasts following treatment with the ionic products of Bioglass(R) 45S5 dissolution, J. Biomed. Mater. Res. 55 (2001) 151–157.
- [24] A. Hoppe, A.R. Boccaccini, Biological impact of bioactive glasses and their dissolution products, Front. Oral Biol. (2015) 22–32.
- [25] C. Wu, et al., Copper-containing mesoporous bioactive glass scaffolds with multifunctional properties of angiogenesis capacity, osteostimulation and antibacterial activity, Biomaterials 34 (2) (2013) 422–433.
- [26] K. Franks, I. Abrahams, J.C. Knowles, Development of soluble glasses for biomedical use Part I: in vitro solubility measurement, J. Mater. Sci. Mater. Med. 11 (10) (2000) 609–614.
- [27] K. Sharma, M.N. Deo, G.P. Kothiyal, Effect of iron oxide addition on structural properties of calcium silico phosphate glass/glass-ceramics, J. Non-Cryst. Solids 358 (16) (2012) 1886–1891.
- [28] F. Baino, G. Novajra, C. Vitale-Brovarone, Bioceramics and scaffolds: a winning

combination for tissue engineering, Front. Bioeng. Biotechnol. 3 (2015) 202.

- [29] A. Entezari, et al., Fracture behaviors of ceramic tissue scaffolds for load bearing applications, Sci. Report. 6 (2016) 28816.
- [30] L.L. Hench, Bioactive materials: the potential for tissue regeneration, J. Biomed. Mater. Res. 41 (4) (1998) 511–518.
- [31] J.S. Fernandes, et al., Design and properties of novel substituted borosilicate bioactive glasses and their glass-ceramic derivatives, Cryst. Growth Des. 16 (7) (2016) 3731–3740.
- [32] E.A. Abou Neel, et al., Bioactive functional materials: a perspective on phosphatebased glasses, J. Mater. Chem. 19 (6) (2009) 690–701.
- [33] J.C. Knowles, Phosphate based glasses for biomedical applications, J. Mater. Chem. 13 (10) (2003) 2395–2401.
- [34] A. Yao, et al., In vitro bioactive characteristics of borate-based glasses with controllable degradation behavior, J. Am. Ceram. Soc. 90 (1) (2007) 303–306.
- [35] E. Dietrich, et al., In vitro bioactivity of melt-derived glass 4656 doped with magnesium, J. Biomed. Mater. Res. A 88 (4) (2009) 1087–1096.
- [36] G. Fielding, S. Bose, SiO(2) and ZnO dopants in 3D printed TCP scaffolds enhances osteogenesis and angiogenesis in vivo, Acta Biomater. (2013) 9(11).
- [37] M. Miola, et al., In vitro study of manganese-doped bioactive glasses for bone regeneration, Mater. Sci. Eng. C 38 (2014) 107–118.
- [38] C. Stähli, et al., Effect of ion release from cu-doped 45S5 Bioglass & reg; on 3D endothelial cell morphogenesis, Acta Biomater. 19 (2015) 15–22.
- [39] S. Tarafder, et al., SrO- and MgO-doped microwave sintered 3D printed tricalcium phosphate scaffolds: mechanical properties and in vivo osteogenesis in a rabbit model, J. Biomed. Mater. Res. B Appl. Biomater. 103 (3) (2015) 679–690.
- [40] V. Vyas, et al., Bioactivity and mechanical behaviour of cobalt oxide-doped bioactive glass, Bull. Mater. Sci. 38 (4) (2015) 957–964.
- [41] S. Zhao, et al., Copper-doped borosilicate bioactive glass scaffolds with improved angiogenic and osteogenic capacity for repairing osseous defects, Acta Biomater. 14 (2015) 185–196.
- [42] J.A. Lee, et al., In vivo biocompatibility of custom-fabricated apatite-wollastonitemesenchymal stromal cell constructs, J. Biomed. Mater. Res. A 103 (10) (2015) 3188–3200.
- [43] J.A. Dyson, et al., Development of custom-built bone scaffolds using mesenchymal stem cells and apatite-wollastonite glass-ceramics, Tissue Eng. 13 (12) (2007) 2891–2901.
- [44] K. Ohsawa, et al., In vivo absorption of porous apatite-and wollastonite-containing glass-ceramic, J. Mater. Sci. Mater. Med. 15 (8) (2004) 859–864.
- [45] T. Kokubo, A/W Glass-ceramic: Processing and Properties. An Introduction to Bioceramics, (1999), pp. 75–88.
- [46] T. Kokubo, H.M. Kim, M. Kawashita, Novel bioactive materials with different mechanical properties, Biomaterials 24 (13) (2003) 2161–2175.
- [47] Standard, ISO 10993-5:2009, Biological Evaluation of Medical Devices. Tests for in Vitro Cytotoxicity, (2009).
- [48] M. Magallanes-Perdomo, et al., In vitro study of the proliferation and growth of human bone marrow cells on apatite-wollastonite-2M glass ceramics, Acta Biomater. 6 (6) (2010) 2254–2263.
- [49] E.A. Abou Neel, et al., Structure and properties of strontium-doped phosphate-based glasses, J. R. Soc. Interface 6 (34) (2009) 435–446.
- [50] O. Bretcanu, et al., Novel resorbable glass-ceramic scaffolds for hard tissue engineering: from the parent phosphate glass to its bone-like macroporous derivatives, J. Biomater. Appl. 28 (9) (2014) 1287–1303.
- [51] H. Arstila, et al., The sintering range of porous bioactive glasses, Glass Technol. 46 (2) (2005) 138–141.
- [52] H. Arstila, et al., Liquidus temperatures of bioactive glasses, Adv. Mater. Res. (2008) 287–292.
- [53] D. Bellucci, A. Sola, V. Cannillo, Hydroxyapatite and tricalcium phosphate composites with bioactive glass as second phase: state of the art and current applications, J. Biomed. Mater. Res. A 104 (4) (2016) 1030–1056.
- [54] S.V. Dorozhkin, Biocomposites and hybrid biomaterials based on calcium orthophosphates, Biomatter 1 (1) (2011) 3–56.
- [55] H. Palmour, R.M. Spriggs, D.P. Uskokovic, Science of Sintering: New Directions for Materials Processing and Microstructural Control, Springer, US, 2013.
- [56] W. Panna, P. Wyszomirski, P. Kohut, Application of hot-stage microscopy to evaluating sample morphology changes on heating, J. Therm. Anal. Calorim. 125 (3) (2016) 1053–1059.
- [57] J. Jones, A. Clare, Bio-glasses : An Introduction, Wiley, 2012.
- [58] G. Thrivikraman, G. Madras, B. Basu, In vitro/in vivo assessment and mechanisms of toxicity of bioceramic materials and its wear particulates, RSC Adv. 4 (25) (2014) 12763–12781.
- [59] T. Sakai, S. Takeda, M. Nakamura, The effects of particulate metals on cell viability of osteoblast-like cells in vitro, Dent. Mater. 21 (2) (2002) 133–146.
- [60] Garc, et al., In vivo effects of vanadium pentoxide and antioxidants (ascorbic acid and alpha-tocopherol) on apoptotic, cytotoxic, and genotoxic damage in peripheral blood of mice, Oxidative Med. Cell. Longev. 2016 (2016) 11.
- [61] E. Mancuso, Processing and Characterisation of Novel Bioceramics for Load Bearing Applications, in School of Mechanical and Systems Engineering, Newcastle University, 2016.
- [62] E. Gentleman, et al., The effects of strontium-substituted bioactive glasses on osteoblasts and osteoclasts in vitro, Biomaterials 31 (14) (2010) 3949–3956.
- [63] S. Hesaraki, et al., Physico-chemical and in vitro biological evaluation of strontium/ calcium silicophosphate glass, J. Mater. Sci. Mater. Med. 21 (2) (2010) 695–705.
- [64] V. Mourino, J.P. Cattalini, A.R. Boccaccini, Metallic ions as therapeutic agents in tissue engineering scaffolds: an overview of their biological applications and strategies for new developments, J. R. Soc. Interface 9 (68) (2012) 401–419.